

# Differentiating Cannabis Products: Drugs, Food, and Supplements

Arash Salehi<sup>1†</sup>, Keely Puchalski<sup>2†</sup>, Yalda Shokoohinia<sup>2</sup>, Behzad Zolfaghari<sup>1</sup> and Sedigheh Asgary<sup>3</sup>\*

<sup>1</sup>Department of Pharmacognosy, School of Pharmacy and Pharmaceutical Sciences, Isfahan University of Medical Sciences, Isfahan, Iran, <sup>2</sup>Ric Scalzo Institute for Botanical Research, Southwest College of Naturopathic Medicine, Tempe, AZ, United States, <sup>3</sup>Isfahan Cardiovascular Research Center, Cardiovascular Research Institute, Isfahan University of Medical Sciences, Isfahan, Iran

"Hemp" refers to non-intoxicating, low delta-9 tetrahydrocannabinol ( $\Delta$ 9-THC) cultivars of Cannabis sativa L. "Marijuana" refers to cultivars with high levels of  $\Delta$ 9-THC, the primary psychoactive cannabinoid found in the plant and a federally controlled substance used for both recreational and therapeutic purposes. Although marijuana and hemp belong to the same genus and species, they differ in terms of chemical and genetic composition, production practices, product uses, and regulatory status. Hemp seed and hemp seed oil have been shown to have valuable nutritional capacity. Cannabidiol (CBD), a nonintoxicating phytocannabinoid with a wide therapeutic index and acceptable side effect profile, has demonstrated high medicinal potential in some conditions. Several countries and states have facilitated the use of THC-dominant medical cannabis for certain conditions, while other countries continue to ban all forms of cannabis regardless of cannabinoid profile or low psychoactive potential. Today, differentiating between hemp and marijuana in the laboratory is no longer a difficult process. Certain thin layer chromatography (TLC) methods can rapidly screen for cannabinoids, and several gas and liquid chromatography techniques have been developed for precise guantification of phytocannabinoids in plant extracts and biological samples. Geographic regulations and testing guidelines for cannabis continue to evolve. As they are improved and clarified, we can better employ the appropriate applications of this uniquely versatile plant from an informed scientific perspective.

#### Keywords: cannabis, cbd, HEMP, marijuana, thc

# INTRODUCTION

*Cannabis sativa* L. is a member of the Cannabaceae family, a small family of annual herbaceous plants that includes the widely cultivated genus, *Humulus* (hops), and eight other genera (Jin et al., 2019). The terms "hemp" and "marijuana" both refer to plants derived from the *Cannabis sativa L.* species, but from different cultivars or chemotypes having either low or high  $\Delta$ 9-THC content, respectively. The general term "cannabis" includes both hemp and marijuana types of plants. (Johnson, 2019). Cannabis has been utilized by several populations for millennia. Central and southeast Asia are considered plausible origins (Steven et al., 2016). It has been claimed that the application of the herb for various purposes dates to 12,000 years ago based on Neolithic evidence found in Taiwan (Li, 1974). Hemp is reputed to be the oldest cultivated fiber plant (Cherney and Small, 2016), and hemp seed and seed oil have historically been used as food (Farinon et al., 2020).

#### **OPEN ACCESS**

#### Edited by:

Carl R. Lupica, National Institute on Drug Abuse (NIH), United States

#### Reviewed by:

Andrea Mastinu, University of Brescia, Italy Ana Paula Duarte, University of Beira Interior, Portugal

> \*Correspondence: Sedigheh Asgary

sedighehasgary@gmail.com

equally to this work

#### Specialty section:

This article was submitted to Ethnopharmacology, a section of the journal Frontiers in Pharmacology

Received: 28 March 2022 Accepted: 01 June 2022 Published: 27 June 2022

#### Citation:

Salehi A, Puchalski K, Shokoohinia Y, Zolfaghari B and Asgary S (2022) Differentiating Cannabis Products: Drugs, Food, and Supplements. Front. Pharmacol. 13:906038. doi: 10.3389/fphar.2022.906038

1

Marijuana has long been utilized for recreational as well as medical purposes (Piluzza et al., 2013). The oldest report of the medical use of cannabis dates to 5,000 years ago when it was used for the treatment of fatigue, rheumatism, and malaria (Abel, 2013). The Assyrians, Egyptians, Indians, Persians, Greeks, and Romans are all reported to have used cannabis for medical purposes (Leghissa et al., 2018a).

Classifications for different cannabis species continues to be a highly debated topic amongst taxonomists and botanists (Sawler 2015). Although some experts recognize three different species of Cannabis: *C. sativa*, *C. indica*, and *C. ruderalis* (Pollio, 2016), and others one monospecific species [*C. sativa* L.] with two subspecies [subsp. *sativa* and subsp. *indica* (Lam)] (Small and Cronquist, 1976, UNODC.org), neither of these nomenclature systems accurately reflects the diversity and complexity of modern cannabis plants. Because cannabis species have been cultivated globally over many years to exhibit nearly indistinguishable phenotypes with overlapping genotypes, *Cannabis sativa* L. varieties are now most accurately identified by cultivar or chemotype, including specific cannabinoid profiling and even subtyping (Sarma et al., 2020). Recently proposed suggestions for chemotyping will be discussed.

Although it has valuable nutritional capacity, hemp material was initially cultivated mainly to produce textiles and ropes. Due to expanded applications of synthetic fibers over the past two centuries, its cultivation decreased precipitously. In recent years, hemp regained popularity due to the rediscovery of its nutritional benefits, economic value, and variety of medicinal uses (Farinon et al., 2020) Despite the resurgence of hemp, in ongoing efforts to curtail recreational drug use, several countries and states have restricted the cultivation of cannabis varieties with high concentrations of  $\Delta$ 9-THC, the primary psychoactive metabolite of the plant (Aguilar et al., 2018).

For the past several decades, a lack of understanding and proper differentiation between hemp- and marijuana-type plants has slowed the development of cannabis research on the potential health benefits of the plant (Farinon et al., 2020). And despite having clearer definitions, improper use of these terms is still regularly seen in the literature. It is pertinent to clarify terminology and provide useful tools and legal definitions to differentiate between food, drug, and supplement derivatives of cannabis.

In this review we focus mainly on the question, "What is essential for defining *Cannabis* as a food, supplement, or drug?" We also investigate the nutritional potential of hempseed and seed oil, the medicinal benefits of certain phytocannabinoids (one of the predominant phytochemical groups in Cannabis) as drugs and supplements, and the regulatory status of cannabis and cannabis products across the globe. Finally, we review several analytical techniques for the detection and quantification of cannabinoids in plants extracts, biological samples, and cannabis products.

## **Definitions, Production Practices, and Uses**

Cannabis terminology has been a significant source of confusion for many. In public opinion at large, marijuana is often viewed as a plant purposed solely for recreational use, which has overshadowed the high value of medical cannabis, hemp-type plants, and hemp products in nutrition, the health and wellness industry, scientific and medical communities, and the global economy (Cadena, 2018). Because of this, it is increasingly important to implement clear nomenclature based on the literature, legal definitions, and the recommendations of agencies specializing in this controversial plant. Here we explore some of the current definitions and nomenclature:

Fundamentally, "marijuana" refers to cultivars of cannabis with psychoactive potential used for both recreational and therapeutic purposes, and "hemp" refers to non-intoxicating cultivars of cannabis with various end uses including fabrics and textiles, nutraceuticals, pharmaceuticals, food products, beverages, oral and topical self-care products, veterinary products, and other manufactured and industrial goods. The term "industrial hemp," in some cases, may be used interchangeably with "hemp" (Johnson, 2019; Crini et al., 2020).

Officially, based on the U.S. 2018 Farm bill, hemp is now legally defined as follows:

"The plant *Cannabis sativa* L. and any part of that plant, including the seeds thereof and all derivatives, extracts, cannabinoids, isomers, acids, salts, and salts of isomers, whether growing or not, with a  $\Delta$ 9-tetrahydrocannabinol ( $\Delta$ 9-THC) concentration of not more than 0.3 percent on a dry weight basis" ((7 U.S.C. \$5940(b)(2)).

Marijuana is still broadly described in the 1970 Controlled Substances Act (CSA) without any limits for  $\Delta$ 9-THC or any other cannabinoid:

"The term marijuana means all parts of the plant *Cannabis* sativa L., whether growing or not; the seeds thereof; the resin extracted from any part of such plant; and every compound, manufacture, salt, derivative, mixture, or preparation of such plant, its seeds or resin." (21 U.S.C. §802(16)).

This definition of marijuana excludes the mature stalks of *Cannabis sativa* L. and products derived from the stalks (e.g. fiber), as well as sterilized seeds incapable of germination, and any certain preparations of viable seeds, such as oil or cake, provided those preparations do not contain resin.

The institution of the above 2018 Farm bill effectively removed hemp from the legal definition of marijuana originally set forth in the CSA in 1970. The CSA had classified cannabis, including hemp, marijuana, and all associated cannabinoids, as a Schedule I controlled substance. The removal of hemp from controlled substance status allowed for agricultural cultivation of hemp plants on U.S. soil as well as greater federal oversight of hemp in commerce, including commodities like food and supplements. Although the separation of hemp from marijuana allowed significantly more freedom for hemp growers and product manufacturers, as well as researchers, clinicians, and consumers, the change also lead to some new aspects of confusion, specifically surrounding use of the isolated compound, cannabidiol (CBD). We will later explore the nuances of CBD applications in food, pharmaceuticals, and supplements.

The terms hemp and marijuana are both affiliated with lengthy cultural and political histories, and in the case of marijuana, polarizing ones. Today, these terms are no longer fully adequate to describe the numerous varieties and phytochemical complexities found in modern day cannabis, nor accurately describe their potential applications. In 2020, in attempts to improve existing cannabis nomenclature, the United States Pharmacopeia (USP) Cannabis Expert Panel (CEP) suggested, due to cannabis's complex secondary metabolome and highly variable distribution of chemical constituents, that the following three broad categories be adopted for classifying cannabis based on phytocannabinoid chemotype: 1) tetrahydrocannabinol (THC)-dominant chemotype; 2) intermediate chemotype with both THC and cannabidiol (CBD); and 3) CBD-dominant chemotype (Sarma et al., 2020). They also proposed that these categories could be further subcategorized by other cannabinoids and/or mono- and sesquiterpene profiles as we continue to learn more about the therapeutic potential of these other compounds in humans (Sarma et al., 2020).

Hemp and marijuana plants generally share a common genetic trait pool with some predictable functional genetic variations; however, distinct genetic variations have also been observed amongst cannabis cultivars (particularly hemp cultivars) spanning the entire genome, not just genes related to THC or cannabinoid synthesis (Sawler et al., 2015). Single nucleotide variant analysis demonstrates separation between CBD and THC (Van Bakel et al., 2011). In most cases, the quantitative ratio of phytochemicals tetrahydrocannabinolic acid (THCA) and cannabidiolic acid (CBDA) (THCA:CBDA) may effectively be used for differentiating cannabis cultivars as either CBD- or THC-dominant (or intermediate) as THCA: CBDA ratios strongly reflect underlying genetic nuances in cannabinoid synthase enzymes (THCA and CBDA synthases) and their sequencing, loci distribution, patterns of expression (functional vs. non-functional), and affinities for CBGA (cannabigerolic acid), their shared substrate (Weiblen et al., 2015). Weiblen et al. (2015) also proposed that all three types of cannabis chemotypes (CBD- or THC-dominant, or intermediate) likely contain multiple linked loci with both THCA and CBDA synthase enzymes; however, identifying the presence of a non-functional CBDA synthase may directly indicate THC-dominance in a plant, as this non-functional enzyme has been selected for over many years where marijuana cultivation has thrived. This novel finding, if proven reliable as an identification method, would allow for rapid screening of THC vs. CBD-dominant plant material at the genetic level prior to any plant cultivation.

Based on the genetic biosynthetic pathways of CBD and THC, Fernandez et al., 2020 were able to report a systematic genetic prediction of *Cannabis* chemotypes of 62 European agricultural hemp cultivars. They suggested three main chemotype groupings for *Cannabis sativa* L, similar to the chemotypes proposed by the USP CEP: Chemotype I, or THC-*pre*dominant varieties, in which the ratio of CBD/THC is low (0.00–0.05) and characterized by a THC level >0.3% (in the U.S.; dry weight of reproductive parts in the female plant at flowering time), Chemotype III, or CBD*pre*dominant varieties, in which the ratio of CBD/THC is 15–25 and, in the U.S., contains <0.3% THC concentration (dry weight of reproductive part in female plants at flowering time), and Chemotype II varieties, or THC-intermediate types, which have both THC and CBD in a content ratio (CBD/THC) of about 0.5–3.0 (Fernandez et al., 2020).

These novel nomenclature systems eliminate much of the confusion surrounding hemp and marijuana, and provide a destigmatized, more objective way of discussing Cannabis for certain applications. These classifications are particularly helpful when discussing the female inflorescence material of Cannabis plants which is cultivated for many different chemotypes and cannabinoid concentrations (both THC and CBD-dominant plants) for therapeutic use. For clarity, we will continue to use the three USP categories suggested above (THC-dominant-, CBD-dominant-, and intermediate), where applicable, for the rest of this article.

In addition to the many chemotypes of plant material, there are also differences in production practices applied to cannabis. Based on different applications, hemp is generally cultivated for three different crops: fiber, seed, and CBD-dominant inflorescence. In contrast, THC-dominant plants are grown solely for their inflorescence for both medicinal and recreational use, as the inflorescence contains the most phytocannabinoids (i.e. Δ9-THC) (Johnson, 2019). Generally, all phytocannabinoids in cannabis exist in resin from the secretory glandular trichomes on the flowering tops of female plants. The numbers of these trichomes are fewer in male plants (Hill et al., 2012). For this reason, female flowers are more valuable for phytocannabinoid production, while male plants are more appropriate for fiber production. Cannabis growers cultivating female plants for their phytocannabinoid production (generally CBD-, THC-, or CBD/THC co-dominant chemotypes) usually remove all male plants to prevent pollinating and seed yielding. Conversely, only male plants are grown to produce hemp fiber and seeds. Some hemp farmers producing fiber and seeds try to prevent flowering to encourage taller stalk growth and less branching (Johnson, 2019), whereas those cultivating hemp for CBD, will promote growth of the plant's phytocannabinoidrich flowering tops.

Historically, there were significant visual differences between hemp and marijuana plants. Modern cultivated plants appear more phenotypically similar when grown for cannabinoid production, but some distinctions can still be made. Cultivated hemp for fiber and seed usually has the following characteristics: a single tall stalk, few leaves and branches due to high-density cultivation in order to prevent branching, and very tall, up to 4.5 m tall. In contrast, the flowering, phytocannabinoid-rich Cannabis plants (i.e. THC-dominant, CBD-dominant, or intermediate chemotypes) are usually characterized by short, tightly clustered, bushy, low-density cultivation with many leaves and branches (in order to develop buds and flowers), and are only 1.2 m tall (Johnson, 2019).

## **Chemical and Genetic Composition**

To date, more than 500 compounds have been identified from *C. sativa* L., out of which 125 compounds are phytocannabinoids. Structurally these compounds have a C-21 terpenophenolic backbone consisting of an alkyl resorcinol with a monoterpene moiety. Eleven types are described for these cannabinoids based on their structure. They are as follows: cannabigerol (CBG),



 $(-)-\Delta 9$ -trans-tetrahydrocannabinol ( $\Delta 9$ -THC),  $(-)-\Delta 8$ -transtetrahydrocannabinol  $(\Delta 8-THC),$ cannabinol (CBN), cannabidiol (CBD), cannabichromene (CBC), cannabielsoin cannabicyclol (CBL), cannabinodiol (CBE), (CBND), cannabitriol (CBT), and miscellaneous-type cannabinoids (Figure 1). Among the phytocannabinoids, 25 of them are reported to be classified as  $\Delta$ 9-THC types and 10 have been elucidated as CBD types. (Hanus et al., 2016; Radwan et al., 2021). Apart from the cannabinoids, four major categories of noncannabinoid-type compounds are present, including alkaloids, flavonoids, non-cannabinoid phenols, and terpenes (Radwan 2021).

Although marijuana is technically defined as having greater than just 0.3%  $\Delta$ 9-THC concentration by dry weight in the U.S., today these plants are often bred to have  $\Delta$ 9-THC levels averaging as high as 10–30%. (Crini et al., 2020). Modern hemp plants

cultivated for CBD production range anywhere from 5–15% total CBD ((Stack G.M. 2021). The number of cannabis plants cultivated for high CBD:THC ratios has grown exponentially in the past decade. Some researchers are finding that specific CBD-dominant chemovars have a CBDAS gene (CBDA synthase) that synthesizes not only CBDA, the precursor of CBD, but also synthesizes some THCA as a side product (Stack G.M. 2021). With accumulation of CBD, where CBDAS is present, there may be concomitant accumulation of THC that exceeds most legal thresholds prior to the time that the plant accumulates the desired 10% or more total CBD (Stack G.M. 2021). This illustrates the importance of chemovar selection and genetic testing prior to large scale cultivation.

Phytocannabinoids are the most abundant constituents in cannabis. Terpenes or isprenoids are the second most abundant constituents, numbering over 200 total terpenes



(Russo 2011; Radwan 2021). Typically found in cannabis trichromes at about 10%, and in flowers at about 1%, some chemotypes have been bred to contain as much as 3.5% terpene content or higher in the flowers (Potter, 2009; Fischedick et al., 2010). Although relatively modest in concentration to the cannabinoids, terpenes (mainly mono and sesquiterpenes) are another important class of compounds. Terpenes are of increasing interest to researchers, healthcare practitioners, and the natural products industry as both isolated therapeutic agents and as enhancing compounds in holistic cannabis formulas designed to engage what has recently been labeled "the entourage effect" (Sommano et al., 2020). Variation in terpene content is likely responsible for the more sedative vs. stimulating effects of different cannabis cultivars, a notion that had historically been attributed to "indica" vs. "sativa" varieties. Terpenes are a family of organic compounds which biosynthetically produced by isoprene units found mainly in plants. Certain cannabis terpenes have been identified and studied in plants such as tea tree, lavender, thyme, basil, Pinus sp., frankincense and citrus fruits like lemon and mandarin (Russo & Marcu 2017; Del Prado-Audelo ML et al., 2021). Monoterpenes and sesquiterpenes of cannabis (the volatile compounds) mainly exist in the essential oil which is isolated by hydrodistillation (Radwan et al., 2021). Among terpenes, the

monoterpene;  $\beta$ -myrcene is mentioned to have the highest prevalence in modern cannabis chemovars in the US (Russo & Marcu 2017). The Sixteen commonly encountered monoterpenes from three classes (acyclic, monocyclic and bicyclic monoterpenes) are shown (**Figure 2**). The most common terpene that exists in cannabis extracts is the sesquiterpene  $\beta$ caryophyllene. Sesquiterpenes have a 15-carbon skeleton. The most abundant sesquiterpenes of cannabis are drawn (**Figure 3**) (Russo & Marcu 2017). Other sources cite additional terpenes as most predominant based on samples from more specific geographic locations (Fischedick, 2017; Radwan 2021).

## **Pharmacological Effects**

Cannabis is known to affect nearly every system in the human body, including but not limited to the central and peripheral nervous systems, as well as the endocrine, immune, gastrointestinal, and musculoskeletal systems. Thousands of articles and over 100 clinical studies have been published on the pharmacodynamics and bioactive effects of cannabis on human psychology, appetite, cognition, sleep, and pain (Russo & Marcu 2017). Only in this century are we are beginning to understand the complexities of these pharmacological actions and interactions, which are largely due to the actions of cannabis on the human endocannabinoid system (ECS) and the "entourage



effect" or bioactive synergisms between phytocannabinoids and other compounds within the plant.

The largest body of cannabis research prior to the past two or three decades focused primarily on the  $\Delta 9$ -THC as a medicinal and recreational drug and emphasized its psychoactive attributes and safety profile. In recent years, CBD has garnered more substantial research interest as a non-intoxicating but equally potent therapeutic compound. Structurally,  $\Delta$ 9-THC (THC) was elucidated in 1964 as the main psychoactive compound in cannabis. In 1988, Huestis et al. discovered that THC exerted its psychoactive effects through agonizing cannabinoid receptor 1 (CBR1). CBR1 is a G protein coupled receptor (GPCR) distributed widely throughout the central nervous system (CNS). A few years later, Murano et al. identified a second cannabinoid receptor, CBR2, that also exists in the CNS (in lower concentrations than CB1R), but in greater concentrations in the peripheral nervous system (PNS) and immune system. Two endogenous ligands (endocannabinoids), were later discovered to bind to these cannabinoid receptors - N-arachidonoyl ethanolamide (AEA) and 2-arachidonoyl glycerol (2-AG) (Lader et al., 2009). AEA and 2-AG are physiologic lipid-based retrograde neurotransmitters that interact with the ECS receptors and proteins in a similar manner to exogenous phytocannabinoids CBD and THC.

The pathways by which exogenous cannabinoids such as  $\Delta 9$ -THC and CBD interact with the human endocannabinoid system (ECS) are complex and only partially understood. Definitively, THC has been shown to directly bind CB1 and CB2 receptors, exerting psychoactive and other neurologically-mediated effects. CBD does not directly bind CB1 or CB2 receptors, exhibiting a low affinity for both, but can antagonize synthetic agonists of cannabinoid receptors suggesting a negative allosteric modulatory effect (Freeman et al., 2019; Premoli et al., 2019). Although CBD does not agonize CBR1 or CBR2, it exhibits several cannabinoid receptor-independent activities including direct agonism of TRPV1 and 5-HT1A receptors, enhancement of adenosine receptor signaling, upregulation of peroxisome proliferator activated receptor gamma (PPARy) protein expression, and suppression of GPR55 receptor activity (Russo & Marcu 2017). CBD is non-intoxicating with a wide therapeutic index and acceptable side effect profile, and has preclinically exerted antipsychotic, intestinal anti-prokinetic, neuroprotective, anti-proliferative, anti-ischemic, vasorelaxant, analgesic, anxiolytic, anti-inflammatory, and antiepileptic effects (Noreen et al., 2018; Cerino et al., 2021). CBD also has been shown to negate the unpleasant side effects of THC, including anxiety, psychosis, tachycardia, and drowsiness through negative allosteric modulation of CB1 receptors. (Laprairie et al., 2015).

Other prominent cannabaninoids of pharmaceutical interest include the following: cannabigerol (CBG), cannabichromene (CBC), cannabinol (CBN), tetrahydrocannabivarin (THCV), cannabidivarin (CBDV), and the acidic cannabinoid precursors cannabidiolic acid (CBDA) and tetrahydrocannabinolic acid (THCA). Most of these cannabinoids are still being investigated in preliminary in vitro and animal model research studies; however some mechanisms of action have been elucidated and reported in the literature. CBG, for example, has weak affinity for CB1 and CB2 receptors, but is a potent  $\alpha$ 2-adrenoreceptor agonist, an efficient serotonin 5-HT<sub>1A</sub> receptor antagonist, and might activate a number of other dominant receptors involved in heat/cold sensitization, pain, and inflammation via antagonism of TRPV8 receptors and stimulation of TRPV1, TRPV2, TRPA1, TRPV3, and TRPV4 (Cascio et al., 2010; De Petrocellis & Di Marzo, 2010; De Petrocellis et al., 2011; Morales et al., 2017)). CBC can have profound therapeutic effects on inflammation and pain through CB2 receptor activity and stimulation and desensitization of TRP ankyrin-type 1 (TRPA1) cation channels, interactions with TRPV3 and TRPV4 cation channels, and desensitization of TRPV2 and TRPV4 channels (De Petrocellis et al., 2012; Cascio & Pertwee, 2014). CBC has also been shown to relieve pain in mice by augmenting the analgesic effects of THC when the two are co-administered (Davis & Hatoum, 1983; Maione et al., 2011; Cascio & Pertwee, 2014), and can increase concentrations and prolong the duration of endocannabinoids like anandamide through its interactions with TRP channels (Shinjyo & Di Marzo, 2013; De Petrocellis et al., 2011, 2012). CBN is being investigated for several topical therapeutic targets for skin conditions like psoriasis, burns, and MRSA infections, via inhibition of keratinocyte proliferation, TRPV2 agonism, and direct antimicrobial activity, respectively (Wilkinson & Williamson, 2007; Appendio et al., 2008; Qin et al., 2008; Russo, 2014). In addition, CBN is an important marker compound of THC degradation in plant material, an artifact that occurs naturally with storage and may be expedited by high heat exposure as the oxidation byproduct of THC (Upton et al., 2013). THCV, a propyl analogue of THC, exhibits concentration-dependent activity at CB1 receptors and may perform as an agonist or act as an antagonist, accordingly (Pertwee, 2008). THCV demonstrates anticonvulsant attributes in mouse pyriform cortex and cerebellum (Hill et al., 2010), and may promote weight loss and other cardiometabolic benefits (Riedel et al., 2009). CBDV has been reported to have significant anticonvulsant activity and is potentially of equal therapeutic value to CBD in treating epilepsy, especially focal/partial onset seizures (Williams, Jones, & Whalley, 2014). CBDV also activates or blocks a wide range of cation channels, depending on the concentration and experimental conditions, including TRPA1, TRPM8, and TRPV channels 1-4. Of additional importance, CBDV inhibits the cellular uptake of anandamide and inhibits endocannabinoid degradation through modulating enzymes diacylglycerol lipase and N-acylethanolamine-hydrolyzing acid amidase (NAAA) (Pertwee & Cascio, 2014). CBDA, the acidic precursor to CBD, shares the same enhancing activity of CBD at 5-HT<sub>1A</sub> receptors, even up to ten times higher than the activity of

CBD, but does not show agonism or antagonism at CB1 receptors (Bolognini et al., 2013; McPartland et al., 2015). CBDA can also inhibit COX1 and COX2, and, at low concentrations (between 1 and 10 µM), targets GPR55, TRPA1, TRPV1, and TRPM8 (Takeda et al., 2008). THCA, like CBDA, is the acidic precursor of THC, representing up to 90% of total THC in the plant prior to prolonged storage, UV exposure, or heat (Moreno-Sanz, 2016). THCA, like THC, is an agonist of CB1 and CB2 receptors, but there is discrepancy as to which (THC or THCA) has greater affinity (Verhoeckx et al., 2006; Rock et al., 2013; Rosenthaler et al., 2014). Although several cannabinoids have been shown to deposit in brain tissue (Alozie, et al., 1980; Deiana et al., 2012), THCA seems to have limited access to the CNS or ability to cross the blood brain barrier (BBB), likely due to its carboxylic acid group (Moreno-Sanz et al., 2016). This means it also does not elicit any psychoactivity. Preliminary research suggests THCA exhibits antineoplastic, neuroprotective, and anti-inflammatory activities via modulation of COX pathways, TRP cation channels, and other immunomodulatory and cell signaling pathways.

In terms of pharmacological effects of cannabis, where there is strong support for the cannabinoids, there is an equal amount of hesitancy in embracing the therapeutic effects of the terpenoids. Although low in concentration in plant material and final cannabis preparations, several aromatic terpenes, especially the monoterpenoids, have been shown to have remarkably potent physiologic effects when administered in humans and animals via inhalation in small amounts ((Falk et al., 1990; Falk et al. 1991; Falk et al. 1993). Terpenes are lipophilic, allowing them to penetrate the skin and the blood brain barrier (Buchbauer et al., 1993). The subsequent activity of terpenes in the CNS seems to be part pharmacological and part psychological, where the psychological effects actually predominate when fragrances are inhaled and the sense of smell is stimulated via G-protein coupled (odorant) receptors (Buchbauer, 2010). In addition, when scent processing of odor information is prevented, pharmacological effects that might be overridden when fragrances are inhaled become more evident (Buchbauer, 2010). Though most often studied for inducing stimulant or sedative psychoactive effects on the nervous system, terpenes also exhibit marked pharmacological versatility outside of the nervous system. Some additional biological effects are related to their antioxidant, antiviral, antinociceptive, antiphlogistic (antiinflammatory), and drug penetration enhancing effects (Buchbauer, 2010). These effects are mediated by neuronal and muscle cell ion channels, interactions with cell membranes, second messenger systems and enzymes (Bowles, 2003; Buchbauer, 2010). B-myrcene, one of the most prevalent terpenes found in all nearly all cannabis plants, increases CB1 receptor saturation, thereby enhancing the psychoactive effects of THC (Wedman-St.Louis 2020). Linalool, a terpene also found in mint, lavender, and cinnamon, may increase serotonin and dopamine levels in the brain (Wedman-St.Louis). a-Pinene binds GABA<sub>A</sub>-Benzodiazepine receptors (GABA<sub>A</sub>-BZD) in the brains of rat models, acting as a partial modulator, and may also display acetylcholinesterase inhibitor activity, either alone or in combination with other terpenes commonly found in the same

plants (Salehi B, 2019)  $\beta$ -Caryophyllene is a potent selective agonist for CB1 receptors which, synergistically, could potentiate the antipruritic and gastro-protective activities of THC and the anti-inflammatory effects of CBD (Gertsch et al., 2008; Russo & Marcu 2017).

Finally, hemp food products can also induce some important pharmacological effects. Phytol, a major diterpene from hemp seed oil has been shown to have anti-cancer as well as antioxidant activities (Vetter et al., 2012). Several studies suggest antihypertensive effects of hydrolyzed hemp seed proteins due to their inhibitory effects on angiotensin-converting enzyme and renin (Aluko et al., 2017). There is also a patented hemp seed flour which was claimed to have an HDL boosting effect and may be beneficial in preventing cardiovascular disease (Guang et al., 2010).

#### **Food Derivatives**

Many food products can be derived from hemp, most of which are made from hemp seed. Common hemp seed food products include bulk or packaged raw hulled hemp seeds, hemp seed oil, hemp protein "concentrates" or powders, and hemp milk, among others. The use of hemp in food products has continued to increase in popularity since the 2018 Federal Farm Bill removed hemp from the Controlled Substances Act. The bill also separated hemp from marijuana and allowed for the interstate transportation of seeds, plants, and processed hemp products. As previously stated, hemp refers to C. sativa with less than 0.3% THC by dry weight. For legal and safety reasons, the level of THC must also be determined in final hemp products and included in the label (Ruphasinghe et al., 2020). Oral intake of as low as 2.5 mg THC per day can cause intoxicating effects in adults (Ross et al., 2000). BgVV recommended a maximum limit of 1-2 µg/kg THC per day (BgVV, 1977). Consumption of higher doses in food products should be avoided due to health threatening consequences. To be precautionary, it is important to consider variations in an individual's sensitivity, drug metabolism, and food-drug interactions. It should also be noted that hemp ingredients from plant parts other than the seeds (including the flower) are not allowed as food ingredients. Non-seed ingredients are prohibited regardless of whether they are added to the food item by the grower/producer, manufacturer, retailer, or by the consumer.

Hemp seeds are a highly nutritious food. Due to the high nutritional value of hemp seed, it could be a complete food source for mankind. Each 100 g hemp seed provides 500–600 Kcal energy (Rodriguez-Leyva et al., 2010). Hemp seed usually contains 25–35% lipids, 20–25% proteins, and 20–30% carbohydrates as well as vitamins (thiamine, riboflavin, pyridoxine, vitamin E and C), minerals (mainly magnesium and iron), flavonoids, tocopherols (alpha, beta, gamma, and delta tocopherols), terpenes, phytosterols and bioactive peptides (Irakli et al., 2019).

A very low but detectable amount of THC is present in hemp seeds, which mainly exists on the seed surface that seems to be a contaminant from glandular trichomes at harvest time. Cultivating and handling processes should be done in such a way that prevents contamination with high levels of THC (Ross et al., 2000). Like THC, CBD is also found in relatively low amounts in the seed oil (Leizer et al., 2000).

Hemp seed oil includes a large percentage of polyunsaturated fatty acids (PUFA) (72-83%) primarily consisting of linoleic (omega-6) and linolenic (omega-3) acids. The ratio of omega-6: omega-3 in the oil is 3:1 which explains its protective potential against cardiovascular disease, osteoporosis, and eczema (Dunford et al., 2015; Ruphasinghe et al., 2020). From a nutritive point of view, the hemp seed oil is much richer than soybean oil (Crini et al., 2020). Human daily nutritional needs of fats can be provided by only three tablespoons of oil (Leizer et al., 2000). Furthermore, hemp seed oil shows a great antioxidant potential. Tocopherol isomers which exist to a large extent in the oil, as well as terpenes and polyphenols (mainly flavonoids), are responsible for this activity (Kriese et al., 2004; Andre et al., 2016; Ruphasinghe et al., 2020). The hemp seed oil also contains a noteworthy amount of phytosterols. It has been noted that plant sterols and stanols exert low-density lipoprotein (LDL) lowering effects in humans (Klingberg et al., 2013).

It is important not to heat the seeds during oil extraction. Unrefined and cold pressed oil is preferred. Cooking the oil produces toxic trans-fatty acids and should be avoided. In addition, unsaturated oil is vulnerable to oxidation, therefore the oil has a very short shelf life and must be protected from air, heat, and light (Crini et al., 2020).

Hemp seed protein is abundant in edestin and albumin, which makes it easily digestible, and provides a beneficial supply of essential amino acids, particularly arginine and glutamic acid (Odani and Odani, 1998; Callaway, 2004). Hemp seed flour is free from gluten and is not known to have any known allergens, hence it does not cause any problem in individuals with celiac disease (Dunford et al., 2015).

Of the many nutritious hemp-derived food products, consumption of hemp seed oil is the most popular. In addition to the more common hemp seed oil, flour, and protein powder, the market of hemp food products continues to expand and now includes a range of novel products from sauces, butter, pasta, and cheese, to energy bars, chocolate, ice cream, and sweets (Klein 2017; Crini et al., 2020).

It should be noted that CBD, the cannabinoid typically found in highest concentration in the inflorescence of hemp plants cultivated for medicinal use, is naturally present, along with other cannabinoids and phytochemicals, in hemp products made from Cannabis inflorescence. There is, however, little to no CBD naturally found in hemp food products like seeds or seed oil, flour, or protein. With rising interest in the beneficial therapeutic effects of CBD and the passage of the 2018 Farm Bill, many retailers began selling CBD-infused foods, beverages, and personal care items, presuming CBD also fell under "hemp" food and supplement regulations; However, CBD was approved as the active ingredient in a seizure medication called Epidiolex in June of 2018. As the active ingredient in a pharmaceutical drug, CBD cannot legally be used as an additive in food or supplements in the U.S. The European Commission, however, has put CBD-containing supplements in the novel foods category (European Commission, 2019). CBD-dominant fullspectrum or broad-spectrum hemp products (<0.3% THC)



with naturally occurring cannabinoids, including CBD, are still able to be consumed as food or supplements, but using CBD in any other context is not allowed. CBD and CBD-infused products have remained in the market despite FDA claims of the illegality of CBD supplements and consumers should be aware of these distinctions. Daily intake of 1–2 mg/kg CBD is implausible to be unsafe (Cerino et al., 2021); however, after identifying concerns of hepatotoxicity, this limitation was done to reduce potential lifethreatening adverse effects and prevent serious drug or supplement interactions (Cohen et al., 2019).

## **Pharmaceutical Drug Derivatives**

The United States FDA has approved four Cannabis-related pharmaceutical products to date: one natural, Cannabisderived drug product-Epidiolex-which contains highly purified CBD, and three synthetic THC-related products: Marinol (dronabinol), Syndros (dronabinol), and Cesamet (nabilone) (Figure 4). Although both drugs containing dronabinol (Marinol and Syndros) are synthetic, dronabinol is also a naturally occurring compound found in Cannabis. In contrast, nabilone is not naturally occurring; it is a man-made drug with structural similarities to THC. In addition to the four cannabis-related products available in the U.S., a product called nabiximols (Sativex), a naturally sourced full spectrum botanical extract with ~1:1 mixture of THC:CBD in an oromucosal spray, is currently marketed legally in over 25 countries including Mexico, Canada, Australia, the United Kingdom and parts of Europe (Namdar et al., 2020; Pearlson 2020). A Dutch-based company, Bedrocan, also offers five different GMP certified, pharmaceutical grade cannabis products with varying THC:CBD ratios (e.g. THC-dominant, CBD-dominant, and intermediate) with complete terpene profiles. Bedrocan's products are available in Australia, South Africa, most of Europe, parts of the United Kingdom and Sweden (Pearlson 2020). Finally, a CBD based transdermal gel called Zygel (Zynerba), is being tested in various stages of clinical trials for the treatment of Fragile X Syndrome and other neuropsychiatric conditions including Autism Spectrum Disorder (ASD) and 22q (ClinicalTrials.gov, Urits et al., 2019).

Cannabinoid drugs each fall under a different level of controlled substance regulation (regulations initiated by the CSA and enforced by the DEA), based on each drug's risk for abuse and physical and/or psychological dependence (Joy et al., 1999). A drug will occasionally change in status with new legislation. Marinol (dronabinol) was originally a schedule II controlled substance, and is now listed under schedule III, indicating it has lower risk for abuse than schedule I or II substances, and has risk of low-to-moderate physical dependency or high psychological dependency if abused (Joy et al., 1999). Syndros (dronabinol) and Cesamet (nabilone) controlled substances are both under schedule II. Epidiolex, a drug containing <0.1% THC, was originally classified in schedule I, but after the hemp farm bill was passed in 2018, it was reduced to schedule V, the least restrictive class of controlled substances (Verma et al., 2021). In April of 2020, Epidiolex was entirely descheduled and is no longer subject to the tracking and monitoring requirements of the CSA. The term "medical marijuana" does not refer to any pharmaceutical drug, but rather use of the whole cannabis plant (with >0.3% THC) as medicine in any product (dried flower, resin, tincture, capsule, etc.) or delivery system (inhalation, oral, sublingual, topical, etc.) (Ebbert 2018). Despite medical and recreational marijuana becoming increasingly legal in several states, medical marijuana is still federally illegal and is classified as a schedule I controlled substance. Physicians may write prescriptions for medical marijuana for patients with certain state-approved medical conditions, which vary widely by state (Ebbert 2018). Hemp, referring to whole plant cannabis with <0.3% THC, was officially removed from the CSA in 2018, so it is no longer a controlled substance and is now regulated under the category of products "generally regarded as safe" (GRAS).

In the U.S., dronabinol (Marinol) was the first FDA approved cannabinoid drug in 1985. By 1999, annual sales of Marinol had reached over \$20 billion, and it was primarily being used (>80% of patients) in the treatment of HIV and AIDS as an appetite stimulant (Joy et al., 1999). Marinol is an oil-based capsule taken orally and, since 1992, has been approved to treat two different conditions: chemotherapy-induced nausea and vomiting (CINV) in cancer patients unresponsive to traditional antiemetics and cachexia in AIDS wasting syndrome (Joy et al, 1 1999). Preliminary clinical trials suggest dronabinol may also be beneficial for patients with obstructive sleep apnea (OSA) and chronic pain, but further evidence is needed (Fisher 2019). For some patients suffering with chemo-induced nausea and vomiting, oral capsules are an impractical delivery system due to swallowing difficulties, long absorption times (approximately two hours), and high variability in intraindividual absorption and bioavailability (Badowski, 2017; Musty & Rossi 2001). Syndros, a

liquid delivery form of dronabinol, was approved in July 2016, as an easier-to-swallow administration option with lower intraindividual variability, faster absorption time (nine minutes *vs.* two hours), and the option to titrate the dosage to clinical effect (Badowski, 2017). Albeit more consistent and tolerable than capsules, because of its liquid delivery system, Syndros may be more easily abused as a drug than Marinol, as it can be converted for inhalation (thus, the schedule II status).

Although many systematic reviews and meta analyses have verified dronabinol's appetite stimulatory and potent antiemetic effects, both alone and in combination with traditional antiemetics like Odensatron, (Ben Amar, 2006; Mücke 2018; Fisher 2019; Namdar et al., 2020), others suggest dronabinol may be no more effective in treating CINV than modern conventional antiemetics and may also have more side effects (Whiting & Wolff, 2015). In addition, many patients reportedly prefer smoking or vaping naturally derived THC-containing products to using synthetic THC medications (Gieringer 2004; Turgeman 2018; Musty & Rossi 2001; Verma et al., 2021). Some patients also seem to benefit more from CBD and THC combination products (Corroon 2018; Verma et al., 2021) and/or whole plant medical marijuana with its synergistic cannabinoids, terpenes, and flavonoids than using THC alone. It is important to note that FDA approved cannabinoid medications have better dosage regulation and quality control than most natural cannabis products that are state regulated. These approved medications are backed by in depth multi-stage clinical trials evaluating their safety and efficacy, which is not true for most medical marijuana products. Due to historical difficulties in studying cannabis as a schedule I controlled substance, a lot more research is needed in this area (Turgeman 2018; Verma et al., 2021).

The same year that dronabinol was approved (1985), a second oral THC-related drug, nabilone (Cesamet) (Figure 4) was also approved for the treatment of CINV unresponsive to conventional antiemetics and anorexia in AIDS patients (Balter 2017; Taylor 2022). As a synthetic analog of THC (Figure 4), nabilone has enough chemical structural differences from THC that it cannot be converted to THC after ingestion and is not detected as a positive for THC in urinary toxicology screenings (Balter 2017). As a potent cannabinoid receptor agonist, nabilone is reported to have a absorption faster time, stronger potency, increased bioavailability, and longer duration of action compared to dronabinol, thus requiring lower and less frequent dosing (Balter 2017). In addition to its approved uses, early research suggests nabilone may also be useful in treating chronic and neuropathic pain, insomnia, cannabis withdrawal and relapse, posttraumatic stress disorder (PTSD), chronic anxiety, and any combination of these frequently comorbid conditions (Lader., 2009; Balter 2017; Cameron 2014).

In June of 2018, Epidiolex (an oral solution containing 100 mg/ml CBD and <0.1% THC) was approved in the US, initially for seizures associated with two rare and drug-resistant conditions—Lennox-Gastaut Syndrome and Dravet Syndrome—in patients  $\geq 2$  ears of age (updated to  $\geq 1$  year of

age in 2020). In August of 2020, Epidiolex was also approved for use in tuberous sclerosis complex (TSC) related seizures in patients  $\geq 1$  year of age (Sekar et al., 2019; Namdar et al., 2020). Cannabidiol (CBD), the primary active constituent in Epidiolex, is metabolized largely through cytochrome P450 (CYP450) enzymes, is a substrate of CYP3A4 and CYP2C19, and also affects the CYP2C family (Abu-Sawwa 2020, Epidiolex [package insert]). Several studies raise concerns regarding drug-drug interactions with CBD and the potential hepatotoxicity of CBD with substantial doses, such as those given with Epidiolex (Abu-Sawwa 2020). Hepatotoxicity has been demonstrated through liver transaminase elevation and increased liver weights of rhesus monkeys and histopathological evidence (Rosenkrantz et al., 1981; Ewing et al., 2019; Huestis et al., 2019; Abu-Sawwa 2020). The application of therapeutic doses in human trials have suggested that hepatotoxicity doesn't occur in all cases and heavily relies on setting, dose, and timing (Walker et al., 2020). The benefits, in nearly all cases, are determined to outweigh the risks (Abu-Sawwa 2020).

The same manufacturer of Epidiolex-UK-based GW Pharmaceuticals-also manufactures a medicinal cannabis product called Sativex (nabiximols) for the treatment of moderate to severe spasticity in multiple sclerosis patients who have not responded adequately to other MS medications (Russo & Guy 2007; Keating 2017). Unlike any of the above cannabinoid drugs with a single purified or synthesized cannabinoid, Sativex comprises a standardized whole plant Cannabis extract with controlled amounts of not just  $\Delta^9$ -THC and CBD (2.7 and 2.5 mg per 100 uL single spray, respectively), but also minor cannabinoids, terpenoids, flavonoids, fatty acids, and sterols (Russo and Guy, 2007, GWPharm.com). Patients generally self-titrate the medication, starting with a single spray and increasing up to twelve sprays daily over the course of two weeks. Several systematic reviews of Sativex in uncontrolled MS-related spasticity suggest it has long term benefit in reducing spasticity, especially as adjunctive treatment, and is also generally well tolerated and safe, does not typically result in dependency or abuse, and has low risk for negative psychoactive effects (Russo and Guy 2007; Keating 2017; Rice and Cameron, 2018; Fragoso 2020).

In 2017, the committee on health effects of marijuana from the National Academies of Sciences, Engineering, and Medicine (U.S.) by investigating more than 10.700 abstracts, published an evidence-based review of the health effects of cannabis and cannabinoids. Regarding the efficacy of cannabis and cannabinoids (either natural or synthetic) weight of evidence was categorized mainly as "conclusive evidence", "substantial evidence", "moderate evidence", "limited evidence" and "no or insufficient evidence" based on the clinical trials (**Table 1**) (National Academies of Sciences, Engineering, and Medicine, 2017).

## **Regulatory Status**

Before the late 1950s, the production of hemp was encouraged by the U.S Department of Agriculture (USDA) (Johnson, 2019).

#### TABLE 1 | Health effects of cannabis and cannabinoids.

Plant Material	Effects of Cannabis and Cannabinoids (Natural or Synthetic)	Effective/In- effective	Weight of Evidence
Cannabis	Treatment of chronic pain in adults	Effective	Conclusive or substantial evidence
Oral cannabinoids	Treatment of chemotherapy-induced nausea and vomiting (antiemetic)	Effective	Conclusive or substantial evidence
Oral cannabinoids	Improving patient-reported multiple sclerosis spasticity symptoms	Effective	Conclusive or substantial evidence
Cannabinoids, primarily nabiximols	Improving short-term sleep outcomes in individuals with sleep disturbance associated with obstructive sleep apnea syndrome, fibromyalgia, chronic pain, and multiple sclerosis	Effective	Moderate evidence
Cannabis and oral cannabinoids	Increasing appetite and decreasing weight loss associated with HIV/AIDS	Effective	Limited evidence
Oral cannabinoids	Improving clinician-measured multiple sclerosis spasticity symptoms	Effective	Limited evidence
Cannabis	Improving symptoms of Tourette syndrome	Effective	Limited evidence
Cannabidiol	Improving anxiety symptoms, as evaluated by a public speaking test, in individuals with social anxiety disorders	Effective	Limited evidence
Nabilone	Improving symptoms of posttraumatic stress disorder	Effective	Limited evidence
Cannabis	Better outcomes (i.e., mortality, disability) after a traumatic brain injury or intracranial hemorrhage	Effective	Limited evidence
Cannabinoids	Improving symptoms associated with dementia	In-effective	Limited evidence
Cannabinoids	Improving intraocular pressure associated with glaucoma	In-effective	Limited evidence
Nabiximols, dronabinol, and nabilone	Reducing depressive symptoms in individuals with chronic pain or multiple sclerosis	In-effective	Limited evidence
Cannabinoids	Cancers, including glioma	Effective	No or insufficient evidence
Cannabinoids	Cancer-associated anorexia cachexia syndrome and anorexia nervosa	Effective	No or insufficient evidence
Dronabinol	Symptoms of irritable bowel syndrome	Effective	No or insufficient evidence
Cannabinoids	Epilepsy	Effective	No or insufficient evidence
Cannabinoids	Spasticity in patients with paralysis due to spinal cord injury	Effective	No or insufficient evidence
Cannabinoids	Symptoms associated with amyotrophic lateral sclerosis	Effective	No or insufficient evidence
Oral cannabinoids	Chorea and certain neuropsychiatric symptoms associated with Huntington's disease	Effective	No or insufficient evidence
Cannabinoids	Motor system symptoms associated with Parkinson's disease or the levodopa-induced dyskinesia	Effective	No or insufficient evidence
Nabilone and dronabinol	Dystonia	Effective	No or insufficient evidence
Cannabinoids	Achieving abstinence in the use of addictive substances	Effective	No or insufficient evidence
Cannabidiol	Mental health outcomes in individuals with schizophrenia or schizophreniform psychosis	Effective	No or insufficient evidence

After introducing THC as the psychotropic constituent of Cannabis sativa L., authorities started to make policies against the expansion of cultivating all types of the cannabis plant without distinguishing between low and high THC varieties (Farinon et al., 2020). Cannabis was internationally banned by the Single Convention on Narcotic Drugs in 1961 and classified in schedule I, along with substances like heroin. Schedule I controlled substances are concluded to be highly addictive, have great potential for abuse, and/or have capacity for use as precursors for other illegal drugs. In addition, the Convention on Psychotropic Substances in 1971, listed THC in schedule I due to its potential for abuse, threat to public health, and low therapeutic potential. In 1988, the United Nations Convention against Illicit Traffic in Narcotic Drugs and Psychotropic Substances prohibited "the production, manufacture, supply, distribution, sale, transportation, import or export of any narcotic drug or psychotropic substance." It also prohibited possession of these substances and cultivation of plants under the legislation of states parties (UNODC, 2013). As a result, the importance of scientific and medical research regarding cannabis was marginalized (Aguilar et al., 2018).

Today, the regulatory aspect of cannabis that is of greatest concern in most jurisdictions is the employment of strict THC limits to determine which plants fall under "hemp" status and can be legally cultivated, sold, and marketed as such. A limit of 0.3% THC (dry weight basis) in "young, vigorous leaves of relatively mature plants" was a guideline arbitrarily adopted by Small et al. (1976) for this purpose, and many countries, including the U.S., have established this same criterion of a 0.3% THC limit. The range for THC limits across the globe currently spans between 0.2 and 1.0%, but most countries have chosen 0.2% as the upper limit. Relatively few jurisdictions, including the Swiss government, have increased the legal limit for hemp cultivars to as high as 1.0% THC, a much higher percent than the rest of the European Union (EU), which currently limits THC content to just 0.2%. The European Union (EU) legalized the cultivation of C. sativa L. plants for industrial applications in 2013 limiting trade to varieties with less than 0.2% THC dry weight of leaves and flowering parts (EMCDDA, 2018). The EU had initially set the upper limit for industrial hemp to 0.5% THC in 1984, then reduced it to 0.3% in 1987, and finally to 0.2% in 1999. In Canada, cultivating hemp varieties with THC content below 0.3% dry

weight of leaves and flowering parts was allowed in 1998. Today, Canada is the largest producer and exporter of hemp products, in particular, hemp-based food and ingredients, around the world (Johnson, 2018). The hemp-producing market of EU is the second only to Canada. Countries including France, Netherlands, Lithuania, and Romania are among the largest producers in the EU (Johnson, 2018). The EU also certifies specific hemp varieties of seeds in the "Common Catalogue of Varieties of Agriculatural Plant Species," and grants subsidies to farmers using these catalogued seeds as long as they remain <0.2% THC (Glivar et al., 2020).

In each of the above cases, measurements of THC concentrations are usually required to include not just THC, but THC's carboxylated precursor, THCA, and sometimes its non-enzymatic oxidation byproduct, CBN (Stack G.M. 2021, Glivar T, et al., 2020, Sarma 2020). THCA is more prevalent than THC in fresh cannabis plant material, as well as in samples dried at low temperature that have not been in storage for long (Glivar T, et al., 2020). When THCA is naturally decarboxylated to THC, there is a loss of molecular mass, so calculations for %w/ w [%mg of cannabinoid/100 mg herbal drug] for total THC concentration should include the following ratio of 0.877% THCA:

% of Total  $\Delta$ 9 -THC = (%  $\Delta$ 9-THC-A x 0.877) + %  $\Delta$ 9-THC + % CBN.

Similarly, when calculating total CBD concentration, the following should be performed to account for the decarboxylation reaction of CBDA to CBD:

% of Total CBD = (% CBD-A x 0.877) + % CBD.

The preceding paragraphs specifically discuss hemp regulations, which likely define the most important aspect of Cannabis governance today; however, this unique plant presents other regulatory challenges, such as the legalization of medical and/or recreational THC-dominant cannabis, determining which conditions are eligible for medical THC use, discrepancies between federal and state cannabis regulations, and confusion around the use of isolated cannabinoids, like CBD, which are present in full- and broad-spectrum hemp extracts.

In terms of recreational cannabis legalization, Latin America has been the world's pioneer in facilitating regulations for recreational THC-dominant cannabis. Uruguay was the first country in the world which legalized production, distribution, sale, and consumption of marijuana and its derivatives for purposes other than medical and scientific uses (MacKay & Philips, 2016). Cannabis markets in Uruguay were allowed to sell marijuana for medicinal, scientific, industrial, and recreational purposes under several considerations (Aguilar et al., 2018). After regulatory changes in 2017, Germany was one of the first countries to pronounce that both public health services and private medical insurances should cover medical cannabis treatments (Aguilar et al., 2018). Select countries like Portugal have decriminalized ("eliminating criminal penalties for the unauthorized consumption and possession of small amounts for personal use only of a controlled substance") the use of marijuana. In Spain, possession of marijuana was decriminalized too, but not its sale. In the Netherlands, use of marijuana has been *de facto* decriminalized ("not applying

statutes that penalize the production, distribution, or consumption of a substance to the fullest extent") (MacKay & Philips, 2016). Also in the Netherlands, Cannabis manufacturer, Bedrocan, has expanded to procure vast ownership over medicinal cannabis production in the country and produces, standardizes and exports large amounts of cannabis products with different THC and CBD concentrations. More strict policies still apply in parts of Asia, as medical cannabis has remained prohibited in Iran, Japan, Vietnam, and Pakistan; but in India, several legal provisions for use of medical cannabis have been permitted (Aguilar et al., 2018; Ghiabi et al., 2018).

In the U.S. although marijuana remains a schedule I controlled substance under federal law which bans the manufacture, distribution, dispensation, and possession of marijuana, several individual states have elected to legalize marijuana for recreational use apart from federal guidelines (Johnson, 2018). In the U.S., marketing of all cannabisderived products and CBD was banned until a few years ago based on their schedule I status and concerns about their side effects. The 2018 Farm Bill modified the definition of hemp so that hemp was no longer a controlled substance. Hemp cultivation in the US is now under USDAapproved license while growing marijuana is generally prohibited because it is considered to have drug-abuse potential (Johnson, 2019). Regardless of federal restrictions, select states allow for the legal cultivation of cannabis plants both for medicinal and recreational purposes. Dietary supplements of CBD remain federally illegal, but FDA has primarily taken action only against marketers making disease treatment claims. After approving Epidiolex, which contains the active ingredient, CBD, for certain epilepsy conditions, CBD was inevitably assigned pharmaceutical drug status and could no longer be treated like a dietary supplement (Walker et al., 2020).

## **Analytical Methods**

Based on International Narcotic Control Board (INCB) reports, total globally production of cannabis increased from 1.3 tons in 2000 to 100.2 tons in 2015 (INCB, 2016). Third party cannabis testing laboratories are responsible for analyzing and quality control of this massive amount of raw material as well as any final product containing cannabis, whether THC-dominant, CBD-dominant, or intermediate. In addition to phytocannabinoid potency testing, which is typically done via HPLC (high performance liquid chromatography), many cannabis labs also now commonly test for terpene content (typically done via GCMS: gas chromatography with mass spectrometry), as well as several contaminants-pesticides, heavy metals (cannabis is a hyperaccumulator), mycotoxins, and residual solvents on derivatives and edibles. For any final cannabis products, in addition to the above analyses, many labs also test for water activity, moisture content, and homogeneity to determine potential for microbial growth and an equal distribution of cannabinoids within the product, respectively. Testing requirements are overseen by specific regulating bodies

where cannabis is being grown, traded, or consumed. Labs regularly consult with growers, insurers, producers, distributors, and retailers to ensure final products are labeled and marketed accurately and are safe for consumers. In addition to the above tests, some other tests labs occasionally perform include product stability, flavonoid and lipid concentrations, gender testing, nutrient testing, and leachability studies. The breadth of all things cannabis lab testing is beyond the scope of this article. Our focus will be on phytocannbinoid profiling to determine CBD-vs. THCdominant chemovars and pros and cons of the most common analysis platforms.

Though different analytical methods and guidelines exist, differentiating between CBD- and THC-dominant cannabis chemotypes is no longer a difficult process, due to increased knowledge of biochemical and biomolecular characteristics of C. sativa L. and the development of instruments and analytical methods that facilitate detection and exact quantification of cannabinoid content (Farinon et al., 2020; Stefkov 2022). The United Nations suggested identifying cannabinoids fast and easily with color tests-e.g. the Fast Corinth V salt test, the Fast Blue B salt test, and the Rapid Duquenois test. Thin layer chromatography (TLC) in both normal and reverse phase systems, besides visualizing with Fast Blue B salt as a reagent, may be used to detect cannabinoids generally. TLC is typically used for fast screening of cannabinoids and does not provide further information; in addition it has low specificity and sensitivity and false-positive results may be seen (UNODC, 2009; Leghissa et al., 2018a; Citti et al., 2018). An accurate and reproducible rapid high-performance TLC (HPTLC) method was developed for quantitative analysis of total THC quantification (decarboxylated THCA + THC + CBN) along with a qualitative fingerprint of the other primary neutral cannabinoids (CBD, THCV, CBG and CBC), with results comparable to that of an HPLC fingerprint (Fischedick et al., 2009; Stefkov 2022). HPTLC may be helpful in screening for high THC content in hemp and hemp products.

Gas chromatography (GC) is one of the oldest and most common methods used for quantifying cannabinoids. The method is accomplished in a short period (under 20 min) at 300°C with low polarity stationary phases (e.g. 5% diphenyl and 95% dimethyl polysiloxan). Cannabinoids are partly present in the plant in their acidic forms. Quantification of acidic cannabinoids with GC is impossible because they undergo decarboxylation during passage through the hot injection port; hence it is necessary for cannabinoids to be derivatized prior to GC. Derivatizing also results in better peak shape and more volatility (Dussy et al., 2005; Leghissa et al., 2018a). FID and MS commonly are used as detectors for the recognition and quantification of cannabinoids coupled with GC. GC-MS is the most researched and reviewed platform for analytical cannabinoid profiling to date (Citti 2018; Kowalska et al, 2020; Kenezvic et al, 2021). GC-FID provides a more accurate and cheaper quantification than GC-MS because GC-MS requires expensive and not always available equivalent deuterated standards (Citti et al.,

2018). Due to its higher efficacy, helium gas generally is preferred over nitrogen and hydrogen in studies as a carrier gas in GC-MS (Lazarjani et al., 2020). Several other GC methods have been reported. Some of these include, GC technology and starch gel electrophoresis (Hillig & Mahlberg, 2004), multiple reaction monitoring (MRM) analysis GC coupled with triple quadrupole mass spectroscopy (Leghissa et al., 2018b), vacuum ultraviolet spectroscopy (GC-VUV) (Leghissa et al., 2018c) and two-dimensional gas chromatography (Omar et al., 2014).

High-performance liquid chromatography (HPLC) is gaining popularity as the botanical chemical fingerprint method of choice, including for phytocannabinoid profiling (Stefkov et al., 2022). The most commonly analyzed cannabinoids include  $\Delta 9$  -THC, THCA, CBD, CBDA, CBN, CBG, and CBC. Using C18 columns for the stationary phase allows for high resolution and separation of cannabinoids. The addition of 0.1% formic acid to methanol or water for the mobile phase improves peak shapes and enhances resolution (Leghissa et al., 2018a; Citti et al., 2018). It is possible to quantify both neutral and acidic cannabinoids with HPLC (i.e. THC and THCA), due to low-temperature operation. In contrast, HPLC is not appropriate for the analysis of volatile compounds such as terpenes, which are better analyzed via GCMS (Lazarjani et al., 2020). Mass spectroscopy (MS) and ultraviolet (UV) are commonly coupled with HPLC as a detector (Lazarjani et al., 2020). UV is cheaper and more straightforward than MS (Leghissa et al., 2018a). Neutral cannabinoids show a UV absorption peak around 220 nm and acidic cannabinoids peak in the range of 270-310 nm (Hazekamp et al., 2005; Citti et al., 2016). MS may provide higher specificity than UV, which makes it possible to analyze complex matrices of cannabis (Citti et al., 2018). To yield further information and improve specificity, the diode array detector (DAD) has been suggested by some authors (Leghissa et al., 2018a). HPLC-DAD differentiates between Cannabis sativa L. chemotypes based on different absorption spectra (Peschel & Polit, 2015). In order to differentiate between CBD and CBG, using HPLC-MS is preferred over HPLC-UV because UV does not provide enough resolution in this case, with certainty, in above 10% concentration of extract (Giese et al., 2015; Lazarjani et al., 2020). Some other methods have also been shown to have different advantages in the quantification of cannabinoids including LC-MS/MS and APCI (Grauwiler et al., 2007), HPLC-MS/MS (Aizpurua-Olaizola et al., 2014), and UPLCqTOF (Lazarjani et al., 2020).

Many labs prefer either GC- or LC- based methods for cannabinoid testing. According to a recent comprehensive literature review of the past four decades of analytical methods reported for cannabinoid profiling in over 220 scientific papers, Stefkov et al., 2022 states that GC- and LC-based methods, at this point in time, exhibit comparable accuracy, selectivity, linearity, sensitivity and precision. Both are used routinely and in investigational analysis of cannabis and cannabis products. HPTLC may soon prove to be superior to either one of these (GC techniques or HPLC) due to the possibilities of lower running costs, quicker analysis run time, automation of sample application and spotting, and ability to analyze multiple samples simultaneously.

In addition to developing a highly sensitive, precise, and affordable instrumental analysis, identification of plant materials, sample selection, and preparation should be taken into account.

Macroscopic and microscopic features are primarily used to identify the plant material. Differential identification is accomplished to figure out the presence of closely related species or other adulterations. HPTLC fingerprint could complete the macroscopic and microscopic techniques, as it provides important data about the major cannabinoids. The USP general chapters <563>, <203>, and <1064> describe general procedures which could be useful in the identification of botanicals (Sarma et al., 2020). A proper sample of plant material should represent the characteristics of the whole batch to make sure of reproducibility in the results of quantification analyses. A non-homogenized or stratified batch may lead to under- or overestimation of cannabinoid content (due to higher levels of cannabinoids in glandular hair trichomes) (Sarma et al., 2020). The USP general chapter <561> explains a valid sampling procedure for botanical drugs. The procedure is based on the weight of the holding container. Contents should be mixed for less than 1 kg containers. For 1-5 kg containers, sampling should be performed by withdrawing an equal amount of plant material from the upper, middle, and lower parts of the container. For above 5 kg containers, three samples must be taken from each of the upper, middle and lower parts (USP <561>, 2019).

Sample preparation varies and depends mainly on the aim of analysis and sample form (solid, liquid, or gas) (Chen et al., 2021). For non-biological samples which range from raw materials and cannabis extracts to semi- (mixture of fully-purified cannabinoids) or (single cannabinoid) materials and formulated cannabis preparations (e.g, edible oil or capsule), analysis of cannabinoids is done to determine the chemical profile of cannabinoids. Cannabis extraction is the process of isolating cannabinoids or terpenes from cannabis trichomes with the purpose of producing medical or recreational supplements and is necessary before any quantification or purification. This process involves several consecutive procedures as follows: Harvesting, trimming the flowers, drying in the absence of sunlight to avoid chemical degradation, milling to decrease particle size, immersion in a solvent (constituents release into the solvent), winterizing (a deep-freezing procedure with the aim of the sedimentation of a viscous black residue called wax), filtration to remove wax from the extract and finally, removal of the solvent using a rotary evaporator. Further purification methods, might be followed according to the final goal, such as crystallization or chromatography. Polar solvents are routinely used for the solvent extraction of semi-polar cannabinoids or terpenes from the flower trichomes e.g, ethanol, methanol, isopropanol, dimethyl ether (Valizadehderakhshan et al., 2021).

Biological specimens (blood, plasma, serum, urine, oral fluid, breath, and hair) typically are analyzed for research purposes and

forensic toxicological and drug abuse investigations (Karschner et al., 2020). Several solvent-based extraction approaches are used for solid samples to extract the marker analytes (cannabinoids in this case) and remove any byproducts. Liquid-phase extraction (LPE), solid-phase extraction (SPE), and solvent extraction assisted with microwave, ultrasonic, or pressure methods have been proposed for solid sample preparation. Solid-phase microextraction (SPME) is widely employed for hair samples due to increased automation and potential for high throughput. Liquid samples are mostly involved in the detection of drug abuse cases. Similar to solid samples, liquid samples must be processed for isolation of cannabinoids and elimination of interferences. Liquid-Liquid extraction (LLE) and solid-phase extraction (SPE), in addition to recent microextraction methods including solid-phase microextraction, packed sorbent microextraction, dispersive solid-phase dispersive liquid-liquid extraction, microextraction. and hollow fiber-based liquid-phase microextraction, are used for this purpose. As far as gas samples like a sample of air is typically prepared through capillary microextraction. For aerosols, liquid-phase extraction is utilized after collecting airborne particles (Chen et al., 2021).

## DISCUSSION

Confusion and inappropriate use of the terms "marijuana" and "hemp" is a great limitation in regards to broader acceptance and widespread use of the cannabis plant. Distinctions between hemp and marijuana extend beyond definitions and concentrations of THC, as they differ in chemical and genetic composition, production practices, and especially in their use as medicine, food, and in industrial applications. Hundreds of preclinical and clinical studies have been done to evaluate the efficacy and safety of phytocannabinoids like CBD in different conditions, but the body of evidence from high-quality clinical studies still lacks robust conclusions about CBD's safety and efficacy in treating diseases other than epilepsy.

Due to the complex nature of the endocannabinoid system (ECS) and its role on several physiologic functions in the body such as mood, memory, inflammation, and appetite, there are still many uncertainties about the way exogenous phytocannabinoids interact with the human ECS, and many areas that still need further research.

In addition, there is an ongoing need for legislative and regulatory intervention in some countries in order to provide a platform for developing research about cannabis and related industries. Making policies that promote the advantages of cannabis and prevent its misuse is necessary, and policymakers should attempt to increase public knowledge about cannabis safety based on current research. Most of the primary issues with cannabis could likely be resolved through identifying and quantifying intoxicating compounds in both original plant material and cannabis products, agreement on universal nomenclature, reporting and labeling guidelines, and continued education for all parties involved—farmers, insurers, policy makers, scientists, physicians, and consumers. Instrumental analytical methods and genetic testing will play a crucial role in achieving this goal. .

Despite all of the recent advances, several cannabis topics remain to be addressed. For example, exploring advantages of natural plant extracts over synthetic drugs, clarifying legal use of full spectrum hemp products *vs.* low dose CBD-containing products *vs.* high dose, purified CBD in prescription medication, and exploring the many benefits of lesser known terpenes and minor cannabinoids. Nevertheless, due to expanding knowledge about cannabis's value in medicine and industry, and recent changes in legislation allowing for easier access to this plant, novel research findings are continuing to grow and spread around the world.

## REFERENCES

- Abel, E. L. (2013). *Marihuana: The First Twelve Thousand Years*. Germany: Springer Science & Business Media.
- Abu-Sawwa, R., Scutt, B., and Park, Y. (2020). Emerging Use of Epidiolex (Cannabidiol) in Epilepsy. J. Pediatr. Pharmacol. Ther. 25 (6), 485–499. doi:10.5863/1551-6776-25.6.485
- Aguilar, S., Gutiérrez, V., Sánchez, L., and Nougier, M. (2018). Medicinal Cannabis Policies and Practices Around the World. *Int. Drug Policy Consort. Brief. Pap.*, 1–32.
- Aizpurua-Olaizola, O., Omar, J., Navarro, P., Olivares, M., Etxebarria, N., and Usobiaga, A. (2014). Identification and Quantification of Cannabinoids in Cannabis Sativa L. Plants by High Performance Liquid Chromatography-Mass Spectrometry. *Anal. Bioanal. Chem.* 406 (29), 7549–7560. doi:10.1007/s00216-014-8177-x
- Alozie, S. O., Martin, B. R., Harris, L. S., and Dewey, W. L. (1980). 3H-delta 9-Tetrahydrocannabinol, 3H-Cannabinol and 3H-Cannabidiol: Penetration and Regional Distribution in Rat Brain. *Pharmacol. Biochem. Behav.* 12 (2), 217–221. doi:10.1016/0091-3057(80)90359-7
- Aluko, R. E. (2017). Hemp Seed (Cannabis Sativa L.) Proteins: Composition, Structure, Enzymatic Modification, and Functional or Bioactive Properties. Sustain. protein sources, 121–132. doi:10.1016/b978-0-12-802778-3.00007-x
- Andre, C. M., Hausman, J. F., and Guerriero, G. (2016). Cannabis Sativa: The Plant of the Thousand and One Molecules. *Front. Plant Sci.* 7, 19. doi:10.3389/fpls. 2016.00019
- Appendino, G., Gibbons, S., Giana, A., Pagani, A., Grassi, G., Stavri, M., et al. (2008). Antibacterial Cannabinoids from Cannabis Sativa: a Structure-Activity Study. J. Nat. Prod. 71 (8), 1427–1430. doi:10.1021/np8002673
- Badowski, M. E. (2017). A Review of Oral Cannabinoids and Medical Marijuana for the Treatment of Chemotherapy-Induced Nausea and Vomiting: a Focus on Pharmacokinetic Variability and Pharmacodynamics. *Cancer Chemother. Pharmacol.* 80 (3), 441–449. doi:10.1007/s00280-017-3387-5
- Balter, R. E., and Haney, M. (2017). The Synthetic Analog of ∆ 9 -Tetrahydrocannabinol (THC): Nabilone. Pharmacology and Clinical Application. *Handb. Cannabis Relat. Pathologies*, 821–827. doi:10.1016/b978-0-12-800756-3.00099-5

Ben Amar, M. (2006). Cannabinoids in Medicine: a Review of Their Therapeutic Potential. J. Ethnopharmacol. 105 (1–2), 1–25. doi:10.1016/j.jep.2006.02.001

BgVV, E. (1997). Von Hanf in Lebensmitteln kann gesundheitlich problematischsein. Berlin, Germany: BgVV Pressedienst.

Bolognini, D., Rock, E. M., Cluny, N. L., Cascio, M. G., Limebeer, C. L., Duncan, M., et al. (2013). Cannabidiolic Acid Prevents Vomiting in Suncus Murinus and Nausea-Induced Behaviour in Rats by Enhancing 5-HT1A Receptor Activation. *Br. J. Pharmacol.* 168 (6), 1456–1470. doi:10.1111/bph.12043

Bowles, E. J. (2003). The Chemistry of Aromatherapeutic Oils. 3rd edn. Crow's Nest, NSW: Allen & Unwin.

- Buchbauer, G., Jirovetz, L., Jäger, W., Plank, C., and Dietrich, H. (1993). Fragrance Compounds and Essential Oils with Sedative Effects upon Inhalation. J. Pharm. Sci. 82 (6), 660–664. doi:10.1002/jps.2600820623
- Buchbauer, G. (2010). "Biological Activities of Essential Oils," in Handbook of Essential Oils: Science, Technology, and Applications. Editors K. H. C. Baser and

## **AUTHOR CONTRIBUTIONS**

SA conceived the presented idea and managed the team. AS and KP wrote the article with Co-operation. YS, SA, and BZ reviewed the article critically and they substantially contribute to the final manuscripts.

## ACKNOWLEDGMENTS

The authors thank Roy Upton (the founder, executive director, and editor of the American Herbal Pharmacopoeia (AHP)) for technical assistance and critical review.

- G. Buchbauer (Boca Raton, FL: CRC Press), 235–280. doi:10.1201/ 9781420063165-c9
- Cadena, A. (2018). Hemp vs Marijuana: the Difference Explained (2019 Update). Available at: https://cbdorigin.com/hemp-vs-marijuana/(Accessed Sep 02, 2019).
- Callaway, J. C. (2004). Hempseed as a Nutritional Resource: An Overview. *Euphytica* 140, 65–72. doi:10.1007/s10681-004-4811-6
- Cameron, C., Watson, D., and Robinson, J. (2014). Use of a Synthetic Cannabinoid in a Correctional Population for Posttraumatic Stress Disorder-Related Insomnia and Nightmares, Chronic Pain, Harm Reduction, and Other Indications: a Retrospective Evaluation. J. Clin. Psychopharmacol. 34 (5), 559–564. doi:10.1097/JCP.00000000000180
- Cascio, M. G., Gauson, L. A., Stevenson, L. A., Ross, R. A., and Pertwee, R. G. (2010). Evidence that the Plant Cannabinoid Cannabigerol Is a Highly Potent Alpha2-Adrenoceptor Agonist and Moderately Potent 5HT1A Receptor Antagonist. Br. J. Pharmacol. 159 (1), 129–141. doi:10.1111/j.1476-5381. 2009.00515.x
- Cascio, M. G., and Pertwee, R. G. (2014). "Known Pharmacological Actions of Nine Nonpsychotropic Phytocannabinoids," in *Handbook of Cannabis*. Editor R. G. Pertwee (Oxford, UK: Oxford University Press), 137–156. doi:10.1093/ acprof:oso/9780199662685.003.0007
- Cerino, P., Buonerba, C., Cannazza, G., D'Auria, J., Ottoni, E., Fulgione, A., et al. (2021). A Review of Hemp as Food and Nutritional Supplement. *Cannabis Cannabinoid Res.* 6 (1), 19–27. doi:10.1089/can.2020.0001
- Chen, X., Wu, X., Luan, T., Jiang, R., and Ouyang, G. (2021). "Sample Preparation and Instrumental Methods for Illicit Drugs in Environmental and Biological Samples: A Review." J. Chromatogr. A 1640, 461961.b doi:10.1016/j.chroma. 2021.461961
- Cherney, J., and Small, E. (2016). Industrial Hemp in North America: Production, Politics and Potential. *Agronomy* 6 (4), 58. doi:10.3390/agronomy6040058
- Citti, C., Braghiroli, D., Vandelli, M. A., and Cannazza, G. (2018). Pharmaceutical and Biomedical Analysis of Cannabinoids: A Critical Review. J. Pharm. Biomed. Anal. 147, 565–579. doi:10.1016/j.jpba.2017.06.003
- Citti, C., Ciccarella, G., Braghiroli, D., Parenti, C., Vandelli, M. A., and Cannazza, G. (2016). Medicinal Cannabis: Principal Cannabinoids Concentration and Their Stability Evaluated by a High Performance Liquid Chromatography Coupled to Diode Array and Quadrupole Time of Flight Mass Spectrometry Method. J. Pharm. Biomed. Anal. 128, 201–209. doi:10.1016/j.jpba.2016.05.033
- Cohen, P. A., and Sharfstein, J. (2019). The Opportunity of CBD Reforming the Law. N. Engl. J. Med. 381 (4), 297-299. doi:10.1056/NEJMp1906409
- Corroon, J., and Kight, R. (2018). Regulatory Status of Cannabidiol in the United States: A Perspective. *Cannabis Cannabinoid Res.* 3 (1), 190–194. doi:10.1089/can.2018.0030
- Crini, G., Lichtfouse, E., Chanet, G., and Morin-Crini, N. (2020). Applications of Hemp in Textiles, Paper Industry, Insulation and Building Materials, Horticulture, Animal Nutrition, Food and Beverages, Nutraceuticals, Cosmetics and Hygiene, Medicine, Agrochemistry, Energy Production and Environment: a Review. *Environ. Chem. Lett.* 18 (5), 1451–1476. doi:10.1007/ s10311-020-01029-2

- Davis, W. M., and Hatoum, N. S. (1983). Neurobehavioral Actions of Cannabichromene and Interactions with Delta 9-tetrahydrocannabinol. *Gen. Pharmacol.* 14 (2), 247–252. doi:10.1016/0306-3623(83)90004-6
- De Petrocellis, L., and Di Marzo, V. (2010). Non-CB1, Non-CB2 Receptors for Endocannabinoids, Plant Cannabinoids, and Synthetic Cannabimimetics: Focus on G-Protein-Coupled Receptors and Transient Receptor Potential Channels. J. Neuroimmune Pharmacol. 5 (1), 103–121. doi:10.1007/s11481-009-9177-z
- De Petrocellis, L., Ligresti, A., Moriello, A. S., Allarà, M., Bisogno, T., Petrosino, S., et al. (2011). Effects of Cannabinoids and Cannabinoid-Enriched Cannabis Extracts on TRP Channels and Endocannabinoid Metabolic Enzymes. *Br. J. Pharmacol.* 163 (7), 1479–1494. doi:10.1111/j.1476-5381. 2010.01166.x
- De Petrocellis, L., Orlando, P., Moriello, A. S., Aviello, G., Stott, C., Izzo, A. A., et al. (2012). Cannabinoid Actions at TRPV Channels: Effects on TRPV3 and TRPV4 and Their Potential Relevance to Gastrointestinal Inflammation. *Acta Physiol. (Oxf)* 204 (2), 255–266. doi:10.1111/j.1748-1716.2011.02338.x
- Deiana, S., Watanabe, A., Yamasaki, Y., Amada, N., Arthur, M., Fleming, S., et al. (2012). Plasma and Brain Pharmacokinetic Profile of Cannabidiol (CBD), Cannabidivarine (CBDV),  $\Delta^9$ -tetrahydrocannabivarin (THCV) and Cannabigerol (CBG) in Rats and Mice Following Oral and Intraperitoneal Administration and CBD Action on Obsessive-Compulsive Behaviour. *Psychopharmacol. Berl.* 219 (3), 859–873. doi:10.1007/s00213-011-2415-0
- Del Prado-Audelo, M. L., Cortés, H., Caballero-Florán, I. H., González-Torres, M., Escutia-Guadarrama, L., Bernal-Chávez, S. A., et al. (2021). Therapeutic Applications of Terpenes on Inflammatory Diseases. *Front. Pharmacol.* 12, 704197. doi:10.3389/fphar.2021.704197
- Dunford, N. T. (2015). Hemp and Flaxseed Oil: Properties and Applications for Use in Food. Specialty Oils and Fats in Food and Nutrition: Properties. *Process. Appl.*, 39–63. doi:10.1016/b978-1-78242-376-8.00002-8
- Dussy, F. E., Hamberg, C., Luginbühl, M., Schwerzmann, T., and Briellmann, T. A. (2005). Isolation of Delta9-THCA-A from Hemp and Analytical Aspects Concerning the Determination of Delta9-THC in Cannabis Products. *Forensic Sci. Int.* 149 (1), 3–10. doi:10.1016/j.forsciint.2004.05.015
- Ebbert, J. O., Scharf, E. L., and Hurt, R. T. (2018). Medical Cannabis. *Mayo Clin. Proc.* 93 (12), 1842–1847. doi:10.1016/j.mayocp.2018.09.005
- European Commission (2019). EU Novel Food Catalogue (v.1.1). Available at: http://ec.europa.eu/food/safety/novel\_food/catalogue/search/public/index.cfm. (Accessed November 20, 2019).
- European Monitoring Centre for Drugs and Drug Addiction (EMCDDA) (2018). Cannabis Legislation in Europe: An Overview. Luxembourg: Publications Office of the European Union.
- Ewing, L. E., Skinner, C. M., Quick, C. M., Kennon-McGill, S., McGill, M. R., Walker, L. A., et al. (2019). Hepatotoxicity of a Cannabidiol-Rich Cannabis Extract in the Mouse Model. *Molecules* 24 (9). doi:10.3390/molecules24091694
- Falk, A., Löf, A., Hagberg, M., Hjelm, E. W., and Wang, Z. (1991). Human Exposure to 3-carene by Inhalation: Toxicokinetics, Effects on Pulmonary Function and Occurrence of Irritative and CNS Symptoms. *Toxicol. Appl. Pharmacol.* 110 (2), 198–205. doi:10.1016/s0041-008x(05)80002-x
- Falk, A. A., Hagberg, M. T., Löf, A. E., Wigaeus-Hjelm, E. M., and Wang, Z. P. (1990). Uptake, Distribution and Elimination of Alpha-Pinene in Man after Exposure by Inhalation. *Scand. J. Work Environ. Health* 16 (5), 372–378. doi:10. 5271/sjweh.1771
- Falk-Filipsson, A., Löf, A., Hagberg, M., Hjelm, E. W., and Wang, Z. (1993). D-Limonene Exposure to Humans by Inhalation: Uptake, Distribution, Elimination, and Effects on the Pulmonary Function. J. Toxicol. Environ. Health 38 (1), 77–88. doi:10.1080/15287399309531702
- Farinon, B., Molinari, R., Costantini, L., and Merendino, N. (2020). The Seed of Industrial Hemp (Cannabis Sativa L.): Nutritional Quality and Potential Functionality for Human Health and Nutrition. *Nutrients* 12 (7). doi:10. 3390/nu12071935
- Fischedick, J. T., Glas, R., Hazekamp, A., and Verpoorte, R. (2009). A Qualitative and Quantitative HPTLC Densitometry Method for the Analysis of Cannabinoids in Cannabis Sativa L. *Phytochem. Anal.* 20 (5), 421–426. doi:10.1002/pca.1143
- Fischedick, J. T., Hazekamp, A., Erkelens, T., Choi, Y. H., and Verpoorte, R. (2010). Metabolic Fingerprinting of Cannabis Sativa L., Cannabinoids and Terpenoids

for Chemotaxonomic and Drug Standardization Purposes. *Phytochemistry* 71 (17–18), 2058–2073. doi:10.1016/j.phytochem.2010.10.001

- Fischedick, J. T. (2017). Identification of Terpenoid Chemotypes Among High (–)-Trans-Δ9- Tetrahydrocannabinol-ProducingCannabis sativaL. Cultivars. *Cannabis Cannabinoid Res.* 2, 34–47. doi:10.1089/can.2016.0040
- Fisher, E., Eccleston, C., Degenhardt, L., Finn, D. P., Finnerup, N. B., Gilron, I., et al. (2019). Cannabinoids, Cannabis, and Cannabis-Based Medicine for Pain Management: a Protocol for an Overview of Systematic Reviews and a Systematic Review of Randomised Controlled Trials. *Pain Rep.* 4 (3), e741. doi:10.1097/PR9.00000000000741
- Fragoso, Y. D., Carra, A., and Macias, M. A. (2020). Cannabis and Multiple Sclerosis. *Expert Rev. Neurother.* 20 (8), 849–854. doi:10.1080/14737175.2020. 1776610
- Freeman, A. M., Petrilli, K., Lees, R., Hindocha, C., Mokrysz, C., Curran, H. V., et al. (2019). How Does Cannabidiol (CBD) Influence the Acute Effects of Delta-9-Tetrahydrocannabinol (THC) in Humans? A Systematic Review. *Neurosci. Biobehav Rev.* 107, 696–712. doi:10.1016/j.neubiorev. 2019.09.036
- Gertsch, J. (2008). Anti-inflammatory Cannabinoids in Diet: Towards a Better Understanding of CB(2) Receptor Action? *Commun. Integr. Biol.* 1, 26–28. doi:10.4161/cib.1.1.6568
- Ghiabi, M., Maarefvand, M., Bahari, H., and Alavi, Z. (2018). Islam and Cannabis: Legalisation and Religious Debate in Iran. *Int. J. Drug Policy* 56, 121–127. doi:10.1016/j.drugpo.2018.03.009
- Gieringer, D., St, L. J., and Goodrich, S. (2004). Cannabis Vaporizer Combines Efficient Delivery of THC with Effective Suppression of Pyrolytic Compounds. *J. Cannabis Ther.* 4 (1), 7–27. doi:10.1300/j175v04n01\_02
- Giese, M. W., Lewis, M. A., Giese, L., and Smith, K. M. (2015). Development and Validation of a Reliable and Robust Method for the Analysis of Cannabinoids and Terpenes in Cannabis. J. AOAC Int. 98 (6), 1503–1522. doi:10.5740/ jaoacint.15-116
- Glivar, T., Eržen, J., Kreft, S., Zagožen, M., Čerenak, A., Čeh, B., et al. (2020). Cannabinoid Content in Industrial Hemp (Cannabis Sativa L.) Varieties Grown in Slovenia. *Industrial crops Prod.* 145, 112082. doi:10.1016/j.indcrop.2019. 112082
- Grauwiler, S. B., Scholer, A., and Drewe, J. (2007). Development of a LC/MS/ MS Method for the Analysis of Cannabinoids in Human EDTA-Plasma and Urine after Small Doses of Cannabis Sativa Extracts. J. Chromatogr. B Anal. Technol. Biomed. Life Sci. 850 (1-2), 515–522. doi:10.1016/j.jchromb.2006. 12.045
- Guang, H., and Wenwei, C. (2010). Application of Powder of Whole Cannabis Sativa Seeds for Preparing Functional Food with Adjuvant Therapy of Lowering Blood Fat. *China Pat.*
- Hanuš, L. O., Meyer, S. M., Muñoz, E., Taglialatela-Scafati, O., and Appendino, G. (2016). Phytocannabinoids: a Unified Critical Inventory. *Nat. Prod. Rep.* 33 (12), 1357–1392. doi:10.1039/c6np00074f
- Hazekamp, A., Peltenburg, A., Verpoorte, R., and Giroud, C. (2005). Chromatographic and Spectroscopic Data of Cannabinoids fromCannabis sativaL. J. Liq. Chromatogr. Relat. Technol. 28 (15), 2361–2382. doi:10.1080/ 10826070500187558
- Hill, A. J., Weston, S. E., Jones, N. A., Smith, I., Bevan, S. A., Williamson, E. M., et al. (2010). Δ<sup>9</sup>-Tetrahydrocannabivarin Suppresses *In Vitro* Epileptiform and *In Vivo* Seizure Activity in Adult Rats. *Epilepsia* 51 (8), 1522–1532. doi:10.1111/j. 15281167.2010.02523
- Hill, A. J., Williams, C. M., Whalley, B. J., and Stephens, G. J. (2012). Phytocannabinoids as Novel Therapeutic Agents in CNS Disorders. *Pharmacol. Ther.* 133 (1), 79–97. doi:10.1016/j.pharmthera.2011.09.002
- Hillig, K. W., and Mahlberg, P. G. (2004). A Chemotaxonomic Analysis of Cannabinoid Variation in. *Am J Bot.* 91, 966-75. doi:10.3732/ajb.91.6.966
- Huestis, M. A., Solimini, R., Pichini, S., Pacifici, R., Carlier, J., and Busardò, F. P. (2019). Cannabidiol Adverse Effects and Toxicity. *Curr. Neuropharmacol.* 17 (10), 974–989. doi:10.2174/1570159X17666190603171901
- International Narcotics Control Board Narcotic Drugs: Estimated World Requirements for 2017: Statistics for 2015. Available online: https://www. incb.org/documents/Narcotic-Drugs/Technical-Publications/2016/Narcotic\_ Drugs\_Publication\_2016.pdf (Accessed March 2, 2022).
- Irakli, M., Tsaliki, E., Kalivas, A., Kleisiaris, F., Sarrou, E., and Cook, C. M. (2019). Effect of Genotype and Growing Year on the Nutritional, Phytochemical, and

Antioxidant Properties of Industrial Hemp (Cannabis Sativa L.) Seeds. *Antioxidants (Basel)* 8 (10). doi:10.3390/antiox8100491

- Jin, J., Yang, M., Fritsch, P. W., Velzen, R. V., Li, D., and Yi, T. (2019). Born Migrators: Historical Biogeography of the Cosmopolitan Family Cannabaceae. J. Syst. Evol. 58, 461–473. doi:10.1111/jse.12552
- Johnson, R. (2019). Defining Hemp: A Fact Sheet. Washington, D.C. Congressional Research Service, 44742. Available online: https://www.everycrsreport.com/ files/20190322\_R44742\_1b0195c6aa7e2cad29256c85a8574347c1ee833d.pdf.
- Johnson, R. (2018). Hemp as an Agricultural Commodity; CRS Report RL32725. Washington, DC, USA: Congressional Research Service. Available online: https://fas.org/sgp/crs/misc/RL32725.pdf (Accessed on March 2, 2022).
- Karschner, E. L., Swortwood-Gates, M. J., and Huestis, M. A. (2020). Identifying and Quantifying Cannabinoids in Biological Matrices in the Medical and Legal Cannabis Era. *Clin. Chem.* 66 (7), 888–914. doi:10.1093/clinchem/hvaa113
- Klein, R. F. X. (2017). Analysis of "Marijuana Edibles"—Food Products Containing Marijuana or Marijuana Extracts—An Overview, Review, and Literature Survey. *Microgram J.* 14, 24.
- Klingberg, S., Ellegård, L., Johansson, I., Jansson, J. H., Hallmans, G., and Winkvist, A. (2013). Dietary Intake of Naturally Occurring Plant Sterols Is Related to a Lower Risk of a First Myocardial Infarction in Men but Not in Women in Northern Sweden. J. Nutr. 143 (10), 1630–1635. doi:10.3945/jn.113.178707
- Knezevic, F., Nikolai, A., Marchart, R., Sosa, S., Tubaro, A., and Novak, J. (2021). Residues of Herbal Hemp Leaf Teas - How Much of the Cannabinoids Remain? *Food control.* 127, 108146. doi:10.1016/j.foodcont.2021.108146
- Kowalska, T., Sajewicz, M., and Sherma, J. (2020). PlanarChromatography—MassSpectrometry;CRCPress. USA: BocaRaton,FL.
- Kriese, U., Schumann, E., Weber, W. E., Beyer, M., Brühl, L., and Matthäus, B. (20042004). Oil Content, Tocopherol Composition and Fatty Acid Patterns of the Seeds of 51 Cannabis Sativa L. Genotypes. *Euphytica* 137 (3), 339–351. doi:10.1023/b:euph.0000040473.23941.76;
- Lader, M. (2009). Addiction and the Pharmacology of Cannabis: Implications for Medicine and the Law. Med. Sci. Law 49 (1), 1–17. doi:10.1258/rsmmsl.49.1.1
- Laprairie, R. B., Bagher, A. M., Kelly, M. E., and Denovan-Wright, E. M. (2015). Cannabidiol Is a Negative Allosteric Modulator of the Cannabinoid CB1 Receptor. Br. J. Pharmacol. 172 (20), 4790–4805. doi:10.1111/bph.13250
- Lazarjani, M. P., Torres, S., Hooker, T., Fowlie, C., Young, O., and Seyfoldin, A. (2020). Methods for Quantification of Cannabinoids: a Narrative Review. J. Cannabis Res. 2 (1), 1–10. doi:10.1186/s42238-020-00040-2
- Leghissa, A., Hildenbrand, Z. L., Foss, F. W., and Schug, K. A. (2018b). Determination of Cannabinoids from a Surrogate Hops Matrix Using Multiple Reaction Monitoring Gas Chromatography with Triple Quadrupole Mass Spectrometry. J. Sep. Sci. 41 (2), 459–468. doi:10.1002/jssc.201700946
- Leghissa, A., Hildenbrand, Z. L., and Schug, K. A. (2018a). A Review of Methods for the Chemical Characterization of Cannabis Natural Products. J. Sep. Sci. 41 (1), 398–415. doi:10.1002/jssc.201701003
- Leghissa, A., Smuts, J., Qiu, C., Hildenbrand, Z. L., and Schug, K. A. (2018c). Detection of Cannabinoids and Cannabinoid Metabolites Using Gas Chromatography with Vacuum Ultraviolet Spectroscopy. Sep. Sci. plus 1 (1), 37–42. doi:10.1002/sscp.201700005
- Leizer, C., Ribnicky, D., Poulev, A., Dushenkov, S., and Raskin, I. (2000). The Composition of Hemp Seed Oil and its Potential as an Important Source of Nutrition. J. Nutraceuticals, Funct. Med. Foods 2 (4), 35–53. doi:10.1300/ j133v02n04\_04
- Li, H-L. (1974). An Archaeological and Historical Account of Cannabis in China. *Econ. Bot.* 28 (4), 437–448. doi:10.1007/bf02861426
- MacKay, R., and Phillips, K. (2016). *The Legal Regulation of Marijuana in Canada and Selected Other Countries*. Delhi: Library of Parliament.
- Maione, S., Piscitelli, F., Gatta, L., Vita, D., De Petrocellis, L., Palazzo, E., et al. (2011). Non-psychoactive Cannabinoids Modulate the Descending Pathway of Antinociception in Anaesthetized Rats through Several Mechanisms of Action. *Br. J. Pharmacol.* 162 (3), 584–596. doi:10.1111/j.1476-5381.2010.01063.x
- McPartland, J. M., Duncan, M., Di Marzo, V., and Pertwee, R. G. (2015). Are Cannabidiol and  $\Delta(9)$  -tetrahydrocannabivarin Negative Modulators of the Endocannabinoid System? A Systematic Review. *Br. J. Pharmacol.* 172 (3), 737–753. doi:10.1111/bph.12944
- Morales, P., Hurst, D. P., and Reggio, P. H. (2017). "Molecular Targets of the Phytocannabinoids: A Complex Picture,". Editors A. D. Kinghorn, H. Falk, S. Gibbons, and J. Kobayashi (Cham, Switzerland: Springer International

Publishing), 103, 103–131. doi:10.1007/978-3-319-45541-9\_4Phytocannabinoids. Unraveling Complex Chem. Pharmacol. Cannabis sativa

- Moreno-Sanz, G. (2016). Can You Pass the Acid Test? Critical Review and Novel Therapeutic Perspectives of Δ9-Tetrahydrocannabinolic Acid A. *Cannabis Cannabinoid Res.* 1 (1), 124–130. doi:10.1089/can.2016.0008
- Mücke, M., Weier, M., Carter, C., Copeland, J., Degenhardt, L., Cuhls, H., et al. (2018). Systematic Review and Meta-Analysis of Cannabinoids in Palliative Medicine. J. Cachexia Sarcopenia Muscle 9 (2), 220–234. doi:10.1002/jcsm. 12273
- Musty, R. E., and Rossi, R. (2001). Effects of Smoked Cannabis and Oral δ9-Tetrahydrocannabinol on Nausea and Emesis After Cancer Chemotherapy. J. Cannabis Ther. 1 (1), 29–56. doi:10.1300/J175V01N01\_03
- Namdar, D., Anis, O., Poulin, P., and Koltai, H. (2020). Chronological Review and Rational and Future Prospects of Cannabis-Based Drug Development. *Molecules* 25 (20), 4821. doi:10.3390/molecules25204821
- National Academies of Sciences (2017). Engineering, and Medicine; Health and Medicine Division; Board on Population Health and Public Health Practice; Committee on the Health Effects of Marijuana: An Evidence Review and Research Agenda. The Health Effects of Cannabis and Cannabinoids: The Current State of Evidence and Recommendations for Research. Washington (DC): National Academies Press US. COMMITTEE ON THE HEALTH EFFECTS OF MARIJUANA: AN EVIDENCE REVIEW AND RESEARCH AGENDA. Available from https://www.ncbi.nlm.nih.gov/sites/books/ NBK425770/.
- Noreen, N., Muhammad, F., Akhtar, B., Azam, F., and Anwar, M. I. (2018). Is Cannabidiol a Promising Substance for New Drug Development? A Review of its Potential Therapeutic Applications. *Crit. Rev. Eukaryot. Gene Expr.* 28 (1), 73–86. doi:10.1615/CritRevEukaryotGeneExpr.2018021528
- Odani, S., and Odani, S. (1998). Isolation and Primary Structure of a Methionineand Cystine-Rich Seed Protein of Cannabis Sativa. *Biosci. Biotechnol. Biochem.* 62 (4), 650–654. doi:10.1271/bbb.62.650
- Omar, J., Olivares, M., Amigo, J. M., and Etxebarria, N. (2014). Resolution of Coeluting Compounds of Cannabis Sativa in Comprehensive Two-Dimensional Gas Chromatography/mass Spectrometry Detection with Multivariate Curve Resolution-Alternating Least Squares. *Talanta* 121, 273–280. doi:10.1016/j. talanta.2013.12.044
- Pearlson, G. (2020). Medical Marijuana and Clinical Trials. Weed Sci., 243–260. doi:10.1016/b978-0-12-818174-4.00010-0
- Pertwee, R. G. (2008). The Diverse CB1 and CB2 Receptor Pharmacology of Three Plant Cannabinoids: Delta9-Tetrahydrocannabinol, Cannabidiol and Delta9-Tetrahydrocannabivarin. Br. J. Pharmacol. 153 (2), 199–215. doi:10.1038/sj.bjp. 0707442
- Pertwee, R. G., and Cascio, M. G. (2014). "Known Pharmacological Actions of Delta-9-Tetrahydrocannabinol and of Four Other Chemical Constituents of Cannabis that Activate Cannabinoid Receptors," in *Handbook of Cannabis*. Editor R. G. Pertwee (Oxford, UK: Oxford University Press), 115–136. doi:10. 1093/acprof:0s0/9780199662685.003.0006
- Peschel, W., and Politi, M. (2015). <sup>1</sup>H NMR and HPLC/DAD for Cannabis Sativa L. Chemotype Distinction, Extract Profiling and Specification. *Talanta* 140, 150–165. doi:10.1016/j.talanta.2015.02.040
- Piluzza, G., Delogu, G., Cabras, A., Marceddu, S., and Bullitta, S. (2013).
  Differentiation between Fiber and Drug Types of Hemp (Cannabis Sativa L.) from a Collection of Wild and Domesticated Accessions. *Genet. Resour. Crop Evol.* 60 (8), 2331–2342. doi:10.1007/s10722-013-0001-5
- Pollio, A. (2016). The Name of Cannabis: A Short Guide for Nonbotanists. Cannabis Cannabinoid Res. 1 (1), 234–238. doi:10.1089/can.2016.0027
- Potter, D. J. (2009). The Propagation, Characterisation and Optimisation of Cannabis Sativa L. As a Phytopharmaceutical. London: Pharmaceutical SciencesKing's College, 224.
- Premoli, M., Aria, F., Bonini, S. A., Maccarinelli, G., Gianoncelli, A., Pina, S. D., et al. (2019). Cannabidiol: Recent Advances and New Insights for Neuropsychiatric Disorders Treatment. *Life Sci.* 224, 120–127. doi:10.1016/j. lfs.2019.03.053
- Qin, N., Neeper, M. P., Liu, Y., Hutchinson, T. L., Lubin, M. L., and Flores, C. M. (2008). TRPV2 Is Activated by Cannabidiol and Mediates CGRP Release in Cultured Rat Dorsal Root Ganglion Neurons. J. Neurosci. 28 (24), 6231–6238. doi:10.1523/JNEUROSCI.050408.2008.28/24/6231

- Radwan, M. M., Chandra, S., Gul, S., and ElSohly, M. A. (2021). Cannabinoids, Phenolics, Terpenes and Alkaloids of Cannabis. *Molecules* 26 (9), 2774. doi:10. 3390/molecules26092774
- Rice, J., and Cameron, M. (2018). Cannabinoids for Treatment of MS Symptoms: State of the Evidence. *Curr. Neurol. Neurosci. Rep.* 18 (850), 50. doi:10.1007/ s11910-018-0859-x
- Riedel, G., Fadda, P., Susan, M-S., Pertwee, R. G., Platt, B., and Robinson, L. (2009). Synthetic and Plant-Derived Cannabinoid Receptor Antagonists Show Hypophagic Proper- Ties in Fasted and Non-fasted Mice. Br. J. Pharmacol. 156 (7), 1154–1166. doi:10.1111/j.1476-5381.2008.00107.x
- Rock, E. M., Kopstick, R. L., Limebeer, C. L., and Parker, L. A. (2013). Tetrahydrocannabinolic Acid Reduces Nausea-Induced Conditioned Gaping in Rats and Vomiting in Suncus Murinus. *Br. J. Pharmacol.* 170 (3), 641–648. doi:10.1111/bph.12316
- Rodriguez-Leyva, D., and Pierce, G. N. (2010). The Cardiac and Haemostatic Effects of Dietary Hempseed. Nutr. Metab. (Lond) 7, 32. doi:10.1186/1743-7075-7-32
- Rosenkrantz, H., Fleischman, R. W., and Grant, R. J. (1981). Toxicity of Short-Term Administration of Cannabinoids to Rhesus Monkeys. *Toxicol. Appl. Pharmacol.* 58 (1), 118–131. doi:10.1016/0041-008x(81)90122-8
- Rosenthaler, S., Pöhn, B., Kolmanz, C., Huu, C. N., Krewenka, C., Huber, A., et al. (2014). Differences in Receptor Binding Affinity of Several Phytocannabinoids Do Not Explain Their Effects on Neural Cell Cultures. *Neurotoxicol Teratol.* 46, 49–56. doi:10.1016/j.ntt.2014.09.003
- Ross, S. A., Mehmedic, Z., Murphy, T. P., and Elsohly, M. A. (2000). GC-MS Analysis of the Total Delta9-THC Content of Both Drug- and Fiber-type Cannabis Seeds. J. Anal. Toxicol. 24 (8), 715–717. doi:10.1093/jat/24. 8.715
- Rupasinghe, H. P. V., Davis, A., Kumar, S. K., Murray, B., and Zheljazkov, V. D. (2020). Industrial Hemp (Cannabis Sativa Subsp. Sativa) as an Emerging Source for Value-Added Functional Food Ingredients and Nutraceuticals. *Molecules* 25 (18). doi:10.3390/molecules25184078
- Russo, E. B., Guy, G. W., and Robson, P. J. (2007). Cannabis, Pain, and Sleep: Lessons from Therapeutic Clinical Trials of Sativex, a Cannabis-Based Medicine. *Chem. Biodivers.* 4 (8), 1729–1743. doi:10.1002/cbdv.200790150
- Russo, E. B., and Marcu, J. (2017). Cannabis Pharmacology: The Usual Suspects and a Few Promising Leads. Adv. Pharmacol. 80, 67–134. doi:10.1016/bs.apha. 2017.03.004
- Russo, E. B. (2011). Taming THC: Potential Cannabis Synergy and Phytocannabinoid-Terpenoid Entourage Effects. Br. J. Pharmacol. 163 (7), 1344–1364. doi:10.1111/j.1476-5381.2011.01238.x
- Russo, E. B. (2014). "The Pharmacological History of Cannabis," in *Hand- Book of Cannabinoids*. Editor R. Pertwee (Oxford, United Kingdom: Oxford University Press), 23–43. doi:10.1093/acprof:oso/9780199662685.003.0002
- Sarma, N. D., Waye, A., ElSohly, M. A., Brown, P. N., Elzinga, S., Johnson, H. E., et al. (2020). Cannabis Inflorescence for Medical Purposes: USP Considerations for Quality Attributes. *J. Nat. Prod.* 83 (4), 1334–1351. doi:10.1021/acs. jnatprod.9b01200
- Sarma, N. D., Waye, A., ElSohly, M. A., Brown, P. N., Elzinga, S., Johnson, H. E., et al. (2020). Cannabis Inflorescence for Medical Purposes: USP Considerations for Quality Attributes. *J. Nat. Prod.* 83 (4), 1334–1351. doi:10.1021/acs. jnatprod.9b01200
- Sawler, J., Stout, J. M., Gardner, K. M., Hudson, D., Vidmar, J., Butler, L., et al. (2015). The Genetic Structure of Marijuana and Hemp. *PloS one* 10 (8), e0133292. doi:10.1371/journal.pone.0133292
- Sekar, K., and Pack, A. (2019). Epidiolex as Adjunct Therapy for Treatment of Refractory Epilepsy: a Comprehensive Review with a Focus on Adverse Effects. *F1000Re* 8. doi:10.12688/f1000research.16515.1
- Shinjyo, N., and Di Marzo, V. (2013). The Effect of Cannabichromene on Adult Neural Stem/progenitor Cells. *Neurochem. Int.* 63 (5), 432–437. doi:10.1016/j. neuint.2013.08.002
- Small, E., and Cronquist, A. (1976). A Practical and Natural Taxonomy for Cannabis. *Taxon* 25 (4), 405–435. doi:10.2307/1220524
- Sommano, S. R., Chittasupho, C., Ruksiriwanich, W., and Jantrawut, P. (2020). The Cannabis Terpenes. *Molecules* 25 (24), 5792. doi:10.3390/ molecules25245792
- Stefkov, G., Cvetkovikj Karanfilova, I., Stoilkovska Gjorgievska, V., Trajkovska, A., Geskovski, N., Karapandzova, M., et al. (2022). Analytical Techniques for

Phytocannabinoid Profiling of Cannabis and Cannabis-Based Products—A Comprehensive Review. *Molecules2022* 27, 975. doi:10.3390/molecules27030975

- Stevens, C. J., Murphy, C., Roberts, R., Lucas, L., Silva, F., and Fuller, D. Q. (2016). Between China and South Asia: A Middle Asian Corridor of Crop Dispersal and Agricultural Innovation in the Bronze Age. *Holocene* 26 (10), 1541–1555. doi:10.1177/0959683616650268
- Takeda, S., Misawa, K., Yamamoto, I., and Watanabe, K. (2008). Cannabidiolic Acid as a Selective Cyclooxygenase-2 Inhibitory Component in Cannabis. Drug Metab. Dispos. 36 (9), 1917–1921. doi:10.1124/dmd.108.020909
- Taylor, B. N., Mueller, M., and Sauls, R. S. (2022). Cannaboinoid Antiemetic Therapy. [Updated 2021 Aug 25]. In: StatPearls [Internet]. Treasure Island (Florida): StatPearls Publishing. Available from: https://www.ncbi.nlm.nih.gov/ books/NBK535430/.
- Turgeman, I., and Bar-Sela, G. (2018). Cannabis for Cancer Illusion or the Tip of an Iceberg: a Review of the Evidence for the Use of Cannabis and Synthetic Cannabinoids in Oncology. *Expert Opin. Investigational Drugs* 28, 285–296. doi:10.1080/13543784.2019.1561859
- United Nations Office on Drugs and Crime (2009). *Recommended Methods for the Identification and Analysis of Cannabis and Cannabis Products Manual for Use by National Drug Analysis Laboratories*. New York, NY, USA: United Nations. Available online: https://www.unodc.org/documents/scientific/ST-NAR-40-Ebook\_1.pdf ((accessed on March 5, 2022).
- UNODC, The International Drug Control Conventions, United Nations (2013). The International Drug Control Conventions, United Nations. Available online: https://www.unodc.org/documents/commissions/CND/Int\_Drug\_ Control\_Conventions/Ebook/The\_International\_Drug\_Control\_Conventions\_ E.pdf.
- Upton, R., Craker, L., ElSohly, M., Romm, A., Russo, E., and Sexton, M. (2013). "Cannabis Inflorescence: Cannabis spp.: Standards of Identity, Analysis and Quality Control," in *Scotts Valley*. Editor R. Upton (, CA, USA: American Herbal Pharmacopoeia).
- Urits, I., Borchart, M., Hasegawa, M., Kochanski, J., Orhurhu, V., and Viswanath, O. (2019). An Update of Current Cannabis-Based Pharmaceuticals in Pain Medicine. *Pain Ther.* 8 (1), 41–51. doi:10.1007/ s40122-019-0114-4
- Valizadehderakhshan, M., Shahbazi, A., Kazem-Rostami, M., Todd, M. S., Bhowmik, A., and Wang, L. (2021). Extraction of Cannabinoids from Cannabis Sativa L. (Hemp)-Review. Agriculture 11 (5), 384. doi:10.3390/ agriculture11050384
- Van Bakel, H., Stout, J. M., Cote, A. G., Tallon, C. M., Sharpe, A. G., Hughes, T. R., et al. (2011). The Draft Genome and Transcriptome of Cannabis Sativa. *Genome Biol.* 12 (10), R102–R118. doi:10.1186/gb-2011-12-10r102
- Verhoeckx, K. C., Korthout, H. A., van Meeteren-Kreikamp, A. P., Ehlert, K. A., Wang, M., van der Greef, J., et al. (2006). Unheated Cannabis Sativa Extracts and its Major Compound THC-Acid Have Potential Immuno-Modulating Properties Not Mediated by CB1 and CB2 Receptor Coupled Pathways. Int. Immunopharmacol. 6 (4), 656–665. doi:10.1016/j.intimp. 2005.10.002
- Verma, R., Hoda, F., Arshad, M., Iqubal, A., Siddiqui, A. N., Khan, M. A., et al. (2021). Cannabis, a Miracle Drug with Polyvalent Therapeutic Utility: Preclinical and Clinical-Based Evidence. *Med. Cannabis Cannabinoids* 4 (1), 43–60. doi:10.1159/000515042
- Vetter, W., Schröder, M., and Lehnert, K. (2012). Differentiation of Refined and Virgin Edible Oils by Means of the Trans- and Cis-Phytol Isomer Distribution. J. Agric. Food Chem. 60 (24), 6103–6107. doi:10.1021/ jf301373k
- Walker, L. A., Koturbash, I., Kingston, R., ElSohly, M. A., Yates, C. R., Gurley, B. J., et al. (2020). Cannabidiol (CBD) in Dietary Supplements: Perspectives on Science, Safety, and Potential Regulatory Approaches. J. Diet. Suppl. 17 (5), 493–502. doi:10.1080/19390211.2020.1777244
- Wedman, -St., and Louis, B. (2020). Cannabis as Medicine.
- Weiblen, G. D., Wenger, J. P., Craft, K. J., ElSohly, M. A., Mehmedic, Z., Treiber, E. L., et al. (2015). Gene Duplication and Divergence Affecting Drug Content in Cannabis Sativa. *New Phytol.* 208 (4), 1241–1250. doi:10.1111/nph.13562
- Whiting, P. F., Wolff, R. F., Deshpande, S., Di Nisio, M., Duffy, S., Hernandez, A. V., et al. (2015). Cannabinoids for Medical Use: A Systematic Review

and Meta-Analysis. Jama 313 (24), 2456-2473. doi:10.1001/jama.2015. 6358

- Wilkinson, J. D., and Williamson, E. M. (2007). Cannabinoids Inhibit Human Keratinocyte Proliferation through a Non-cb1/cb2 Mechanism and Have a Potential Therapeutic Value in the Treatment of Psoriasis. J. Dermatol Sci. 45 (2), 87–92. doi:10.1016/j.jdermsci.2006.10.009
- Williams, C. M., Jones, N. A., and Whalley, B. J. (2014). "Cannabis and Epilepsy," in *Handbook of Cannabis* (Oxford, UK: Oxford University Press), 547–563. doi:10.1093/acprofoso/9780199662685.003.0030

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

**Publisher's Note:** All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Salehi, Puchalski, Shokoohinia, Zolfaghari and Asgary. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.