OPEN ACCESS International Journal of Molecular Sciences ISSN 1422-0067 www.mdpi.com/journal/ijms

Review

Targeting the Redox Balance in Inflammatory Skin Conditions

Frank A. D. T. G. Wagener *, Carine E. Carels and Ditte M. S. Lundvig *

Department of Orthodontics and Craniofacial Biology, Nijmegen Centre for Molecular Life Sciences, Radboud University Nijmegen Medical Centre, PO Box 9101, 6500 HB Nijmegen, The Netherlands; E-Mail: c.carels@dent.umcn.nl

* Authors to whom correspondence should be addressed;
E-Mails: f.wagener@dent.umcn.nl (F.A.D.T.G.W.); d.lundvig@dent.umcn.nl (D.M.S.L.);
Tel.: +31-24-3614082 (F.A.D.T.G.W.); Fax: +31-24-3540631 (F.A.D.T.G.W. & D.M.S.L.).

Received: 1 March 2013; in revised form: 10 April 2013 / Accepted: 16 April 2013 / Published: 26 April 2013

Abstract: Reactive oxygen species (ROS) can be both beneficial and deleterious. Under normal physiological conditions, ROS production is tightly regulated, and ROS participate in both pathogen defense and cellular signaling. However, insufficient ROS detoxification or ROS overproduction generates oxidative stress, resulting in cellular damage. Oxidative stress has been linked to various inflammatory diseases. Inflammation is an essential response in the protection against injurious insults and thus important at the onset of wound healing. However, hampered resolution of inflammation can result in a chronic, exaggerated response with additional tissue damage. In the pathogenesis of several inflammatory skin conditions, e.g., sunburn and psoriasis, inflammatory-mediated tissue damage is central. The prolonged release of excess ROS in the skin can aggravate inflammatory injury and promote chronic inflammation. The cellular redox balance is therefore tightly regulated by several (enzymatic) antioxidants and pro-oxidants; however, in case of chronic inflammation, the antioxidant system may be depleted, and prolonged oxidative stress occurs. Due to the central role of ROS in inflammatory pathologies, restoring the redox balance forms an innovative therapeutic target in the development of new strategies for treating inflammatory skin conditions. Nevertheless, the clinical use of antioxidant-related therapies is still in its infancy.

Keywords: ROS; oxidative stress; inflammation; antioxidant; skin

Abbreviations: APE/Ref-1, apurinic/apyrimidinic endonuclease/redox effector factor-1; ARE, antioxidant responsive element; CO, carbon monoxide; CoQ10, coenzyme Q10; ERK, extracellular signal-regulated protein kinases; G6PD, glucose-6-phosphate dehydrogenase; GPx, glutathione peroxidase; GSH; reduced glutathione; H_2O_2 , hydrogen peroxide; HO•, hydroxyl radical; HIF-1, hypoxia inducible factor-1; HO, heme oxygenase; HRE, hypoxia response element; JNK, c-Jun *N*-terminal kinases; KO, knockout; LDL, low density lipoprotein; MAPK, mitogen-activated protein kinase; Nn, manganese; NAC, *N*-acetyl cysteine; NF- κ B, nuclear factor- κ B; Nox, NADPH oxidase; Nrf2, NF-E2-related factor 2; $O_2^{-\bullet}$, superoxide anion; Prdx, peroxiredoxin; PTP, protein tyrosine phosphatase; ROS, reactive oxygen species; SOD, superoxide dismutase.

1. Introduction

The primary function of healthy skin is to form a physical and chemical barrier between the external environment and the organism's internal milieu to defend against injurious insults. Harmful stimuli such as trauma, pathogens or irritants evoke a complex response known as inflammation (1). Inflammation protects organisms against pathogenic invaders and cleans up damaged cells after injury to prevent further tissue damage. Hereto the injurious agents are identified and eliminated and wound healing is initiated for re-establishment of tissue homeostasis. The strength and duration of the inflammatory response depends on stimulus and context, however, and the initial steps are stereotyped as part of the innate immune response [1]. The five classical signs of acute inflammation are: pain, heat, redness, swelling, and functional loss. These signs can be explained by the different phases that the inflammatory response generally follows: (1) dilation of capillaries to increase blood flow; (2) vasopermeabilization; (3) leukocyte recruitment; (4) elimination can also become chronic and destructive, and accumulating evidence demonstrate a contribution of chronic inflammation to the development of diseases like Alzheimer's disease, atherosclerosis, and type 2 diabetes [2].

In most cases, inflammation of skin represents a beneficial and protective process after injury or infection [3]. However, the skin can also be subjected to excessive inflammatory responses resulting in chronic inflammation, auto-inflammation and auto-immunity [4]. The epidermal layer of the skin is composed of predominantly keratinocytes, a few Langerhans cells—which are specialized dendritic cells—and some pigment-producing melanocytes [3,4]. Keratinocytes produce different keratins that generate the toughness of the epidermis [5]. Embedded in the connective tissue of the underlying dermis different types of immune cells can be found, such as macrophages, T cells and mast cells [4]. The connective tissue is composed of an extracellular matrix produced and secreted by fibroblasts, the principal cell type of the dermis [6].

Under homeostatic conditions, the skin surface is colonized by a diversity of microorganisms. However, a dynamic, healthy equilibrium between the epidermis and the microorganismal population is regulated by production of antibiotic and antifungal compounds by dermal sebocytes as well as the microorganisms themselves. Additionally, keratinocytes also produce antibacterial substances constitutively and after infection or injury [7]. Some of these keratinocyte-derived antimicrobics and cytokines influence the immunological properties of dendritic cells and T cells [3,4]. Thus, the skin

balances between ensuring an efficient pathogen defense and immunosurveillance and to reduce excessive immune responses that can lead to disease [1].

At the molecular level, inflammation is activated by the inflammasome that is a cytosolic multiprotein complex regulating caspase-1 activation. Caspase-1 subsequently activates the pro-inflammatory cytokines IL-1 β and IL-18 by proteolytic cleavage. The inflammasome is composed of the danger sensor protein NALP and the caspase-1 recruiter protein ASC [8]. Dependent on the activation stimuli, different inflammasomes are formed as a response to danger signals as each inflammasome has unique roles in pathogen recognition [8].

Signal transduction by IL-1 β and IL-18 results in a series of phosphorylation and ubiquitination events that ultimately leads to activation of nuclear factor (NF)- κ B and p38 mitogen-activated protein kinase (MAPK) pathways, which cooperatively induce the expression of IL-1 target genes, including IL-6 [9]. Pathogen-mediated activation of the inflammasome also induces a specific form of caspase-1-dependent cell death, pyroptosis, that results in osmotic swelling and plasma membrane rupture [10,11]. This induces a strong inflammatory response via the release of pro-inflammatory cytokines and spreading of pro-inflammatory molecules [12].

In contrast to immune cells, human keratinocytes constitutively express inflammasome proteins and are a potent source of the pro-inflammatory cytokines pro-IL1 α and pro-IL1 β [13–15], which are activated and released upon UV exposure [13,16]. This implies important roles for keratinocytes as non-professional immune cells following sunburn, and in innate immune responses dependent on the inflammasome and IL1 β . Keratinocytes are thus important in the inflammatory response under both physiological and pathological conditions [13,17].

Notably, it has been reported that the NLRP3 inflammasome assembly is stimulated in a ROS-sensitive manner by ROS generation [18,19]. The predominant source of ROS as response to danger stimuli are mitochondria, which also control inflammation via release of mitochondrial DNA [20,21]. Excessive release of ROS by mitochondria, activated leukocytes and endothelial cells in chronic inflammatory conditions can ultimately result in severe cell and tissue damage and can further promote and aggravate inflammatory injury. In many inflammatory diseases, currently available intervention strategies fail or are of limited success, warranting the need for novel strategies to treat chronic inflammatory conditions. The prolonged presence of oxidative stress is postulated to promote and fuel these deleterious inflammatory processes and may form a novel target for treatment of chronic inflammatory conditions.

2. Reactive Species Mediate Cellular Signaling

ROS are free radicals generated from molecular oxygen, such as superoxide anion (O_2^{-}) , hydroxyl radical (HO•), and non-radical species including hydrogen peroxide (H₂O₂). Today, it is well accepted that ROS mediate and modulate signaling processes [22,23]. In moderate concentrations, ROS induce a cascade of cell signaling networks, triggering a ROS wave that propagates throughout tissues carrying signals across large distances [24]. They thereby act as important regulatory mediators in different signaling pathways and processes, including cell proliferation, differentiation, and apoptosis [25,26].

Today H₂O₂ is recognized as a major ROS signaling molecule [27] that can mediate oxidation of protein thiols [28], and it is widely accepted that ROS can elicit cellular effects by covalently modifying amino acids and subsequently affecting protein activity. Redox-regulated proteins sense the changes in cellular redox state by different mechanisms. Several transcription factors, including nuclear factor κ B (NF- κ B), hypoxia inducible factor-1 (HIF-1), and p53, contain redox-sensitive cysteine residues in their DNA binding sites [29]. Oxidative modifications of these residues affect DNA binding and subsequently regulate gene transcription of redox sensitive genes [26,30,31]. Additionally, the DNA binding properties of these transcriptional regulators are further indirectly affected by redox-sensitive proteins, like apurinic/apyrimidinic endonuclease/redox effector factor-1 (APE/Ref-1) protein and histone deacetylases [32,33].

Oxidative posttranslational modifications form a major redox-regulated mechanism of protein function [34], targeting multiple types of amino acids with various susceptibilities and subsequent structural and functional consequences [35]. For instance, reversible oxidation of cysteines and oxidative nitration of tyrosines regulate the activity of various kinases involved in signaling pathways, including c-Jun N-terminal kinases (JNK), mitogen-activated protein kinase (MAPK), and protein kinase C [26]. Altered kinase activities due to oxidative modifications affect downstream signaling pathways and consequently transcription factor activation and contribute to additional redox-regulated gene transcription [36]. Effects of some of these redox-mediated alterations of signaling are exemplified below.

Based on the chemical characteristics, only a few amino acids can undergo oxidative modification, namely cysteine, methionine, and tyrosine [37]. Selective oxidation, reduction or chemical modification of these sensor thiols results in a change in protein activity and signal transduction in response to redox fluctuations [38]. However, the reactivity and modification of a particular cysteine residue is highly dependent on the microenvironment [28,39].

The side chain of a cysteine residue contains a terminal thiol (–SH) as a functional group allowing multiple oxidation states and is a common event in redox signaling [27,40–43]. Oxidation of these residues result in reactive sulfenic acid (–SOH) that can form disulfide bonds (RS–SR') with nearby cysteines or undergo further oxidation to sulfinic acid (–SO₂H) and sulfonic acid (–SO₃H) [44]. Most of these modifications are generated by diffusible small molecules like H_2O_2 and are reversible by reducing systems like thioredoxin and peroxiredoxin with the exception of the sulfinic and sulfonic states [27]. For instance, reversible inhibition of protein tyrosine phosphatase (PTP) activity and subsequently regulation of cellular tyrosine phosphorylation and downstream signaling is mediated by H_2O_2 -mediated oxidation of the catalytic cysteine to the sulfenic form [45,46].

Disulfide bond formation is important for protein structure and function [47], and recently its role as signaling event has been demonstrated [48,49]. Some cysteine-containing proteins also form intra- and intermolecular disulfide bonds, resulting in conformational changes and altered function due to ROS-mediated oxidation. For instance, ROS-mediated cysteine oxidation of PTPs is accompanied by intramolecular disulfide bond formation to protect against further oxidation [50–53]. Moreover, the accompanying conformational change might ease access for reducing enzymes and subsequent reactivation of the enzyme [54]. Cysteine redox switches in enzymes have been extensively discussed elsewhere [55].

S-glutathionylation is one of the most common S-thionylation reactions inside the cell and is generated from the reaction between cysteine sulfenic acid and a reduced thiol like GSH. This forms a mixed glutathione disulfide (GSSG) that prevents further irreversible oxidation [56]. Hereby, cells can effectively and reversibly respond to redox input, and glutathionylation indeed regulates most cellular pathways [30]. For instance, activation of the master regulator of several antioxidant genes, NF-E2-related factor 2 (Nrf2), is physically confined to binding partner Keap1 in the cytosol [57]. Keap1 glutathionylation leads to dissociation of the Nrf2-Keap1 complex, allowing Nrf2 to translocate to the nucleus and activate expression of target genes [58]. S-glutathionylation has also been found to control NF-kB pathway activation on different levels [30]. Several members of this pathway are inhibited by S-glutathionylation [59], and NF-KB itself is subjected to redox-regulation [36,60]. Oxidative modifications may also modulate protein-protein interactions and thereby affect the stability of protein complexes and activity of the protein partners involved [61]. For instance, NF-kB is sequestered by IKB in the cytosol. ROS activate IKB kinases that phosphorylate IKB, leading to its degradation by the proteasome and exposure of the nuclear localization sequence of NF-KB. Next, NF-KB translocates to the nucleus, and activates transcription of various target genes [62]. Moderate ROS levels promote NF-κB activation and cell survival, whereas high levels of ROS inactivate NF-kB, leading to cell death [26,36].

S-nitrosylation is the nonenzymatic adduction of NO to a thiol group (-SNO), and is reversible through the action of the protein denitrosylation system mediated by reduced glutathione (GSH) or the thioredoxin system [63,64]. Redox effects of NO are mediated by S-nitrosylation [65] that is thought to exert its regulatory effects through direct modification of protein function via conformational changes or through the protection of thiol groups against further oxidation [66]. However, the exact mechanisms underlying S-nitrosylation-mediated regulation is still not fully explored. S-nitrosylation plays a dual role as it can act as a protective modification and as an intermediate to further oxidation [67] as well as intermediate for formation of secondary posttranslational modifications such as ubiquitination [68]. Also, S-nitrosylation mediates the anti-inflammatory effects of NO in the cardiovascular system, as S-nitrosylation of N-ethylmaleimide factor suppresses vascular inflammation as well as inhibits NF- κ B-dependent expression of pro-inflammatory cytokines and adhesion molecules [65].

Mitochondrial ROS production is also a participant in redox signaling networks [69,70]. The respiratory chain generates O_2^{\bullet} that is converted to H_2O_2 , and both species can act as redox signals, but with different properties and interactions [27,71]. Membrane-impermeable O_2^{\bullet} is restricted to the mitochondrial matrix where it is thought to act as a redox signal within the mitochondria [72,73]. O_2^{\bullet} itself is not particularly reactive with most biomolecules except for iron–sulfur proteins, NO, and quinones [74–76]. Mitochondrial aconitase is an Fe-S cluster containing protein that reacts with O_2^{\bullet} to release H_2O_2 and ferrous iron [77]. ROS-mediated damage to the Fe–S cluster leads to aconitase inactivation and a cellular metabolic shift from ATP production to fat storage due to coenzyme A buildup [78]. This feedback loop has been suggested to be an antioxidant defense mechanism by reducing respiration and subsequently mitochondrial ROS formation [72,78]. Indeed, mitochondrial oxidative stress has been suspected to contribute to metabolic syndrome [78].

Also, mitochondrial ROS production is thought to play a role in oxygen sensing via HIF-1 [79,80] under both hypoxic [81] and non-hypoxic conditions [82]. Additionally, NADPH oxidases contribute for ROS formation and redox-dependent HIF-1 stabilization [83]. HIF-1 is a heterodimeric transcription factor that activates numerous genes involved in hypoxic adaptation via its binding to a

hypoxia response element (HRE) in gene promoter regions [84]. Under normoxic conditions, the HIF1 α subunit is tagged at the oxygen-dependent degradation domain for ubiquitination and subsequent proteasomal degradation by oxygen- and iron-dependent prolyl hydroxylases. Under hypoxia, the hydroxylation of HIF-1 α subunit is inhibited, thereby enabling the formation of the active HIF transcription factor and subsequent induction of target gene transcription [79]. This is achieved through ROS-mediated oxidation of ferrous iron that is a co-activator of a prolyl hydroxylase function [85]. Moreover, a recent study describes perinuclear accumulation of mitochondria due to an altered microtubule-dependent transport under hypoxia [86]. As a result, increased levels of ROS in the nucleus caused oxidative modifications of DNA bases in the HRE of the VEGF promoter that is important for facilitated assembly of the HIF-1 transcriptional complex and thus increased VEGF gene expression [80,86].

Together, these examples illustrate that ROS-mediated posttranslational modifications act as key regulatory mediators in different signaling pathways and processes, including cell proliferation, differentiation, and apoptosis [25,26]. Paradoxically, ROS-signaling also promotes pathways protecting against oxidative stress to restore an imbalanced ratio between cellular oxidants and antioxidants [87]. Also, from these examples it becomes clear that the redox system affects protein function and biological activity either indirectly at the level of protein expression or stability or directly through posttranslational modifications [26]. An exhaustive collection of literature further points towards the importance of well-regulated ROS-mediated signaling as ROS has been linked to diverse groups of diseases, ranging from cardiovascular problems to neurodegeneration, see e.g., [88]. However, despite the ever-increasing number of studies, it is evident that ROS-mediated signaling and importantly, the regulation and specificity hereof, is still poorly defined and warrants for more investigation [27].

3. ROS and (Chronic) Inflammation of the Skin

Excess levels of ROS due to overproduction or because of insufficient scavenging generate oxidative stress, leading to injurious effects via: (1) oxidative modification and damage of biomolecules, altering lipid/protein/DNA structure and function; (2) further irreversible oxidation of reactive protein thiol groups which is hallmark of oxidative stress [89]; and (3) dysregulation of cell signaling pathways [90], triggering downstream signaling cascades leading to altered cytokine release and exacerbation of inflammation [91]. Combined, excess ROS lead to pathological changes in cells and tissues, as exemplified by inflammatory skin conditions like psoriasis.

Inflammatory skin diseases range from acute rashes with itching and redness to chronic conditions like dermatitis (eczema) and psoriasis. Acute skin inflammation may develop following exposure to, e.g., UV radiation (sunburns), allergens, physical wounding, or contact with chemical irritants, and is resolved within two weeks with only minor tissue destruction. In contrast, chronic skin inflammation results from a sustained, exaggerated inflammatory response, negatively affecting skin health.

3.1. Sunburn

Acute dermal overexposure to UV radiation causes sunburn and is an inflammatory response with increased prostaglandin and pro-inflammatory cytokine production, causing erythema, vasodilation, and leukocyte infiltration as predominant features [92,93].

Oxidative stress is thought to play a central role in the cellular response following UV exposure. Solar UV radiation is classified as UV-A (320–400 nm), UV-B (290–320 nm) and UV-C (100–290 nm) [94]. However, skin will only be exposed to UV-A and UV-B, as UV-C and partly UV-B radiation is absorbed by the ozone layer. UV-A photons are absorbed by endogenous UV-absorbing chromophores (e.g., riboflavin, quinines, tryptophan, and porphyrins) that subsequently transfer this energy to molecular oxygen, resulting in the formation of O_2^{-1} . Superoxide anions are then converted to H_2O_2 which in the presence of redox-active transition metals can be converted into HO• [95]. UV-B radiation is mainly absorbed by the epidermis and is primarily responsible for sun burn, whereas UV-A penetrates deeply into the skin. UV-A and UV-B radiation have different targets and outcomes. DNA is a prominent target of UV-B radiation, resulting in the formation pyrimidine and purine photoproducts and DNA strand breaks [96]. Additionally, UV-B-derived HO• can also damage the DNA, inhibiting normal cell function [97].

The effects of UV-A radiation exposure are a result of both direct and indirect damage to biomolecules and the subsequent physiological consequences hereof. Firstly, UV radiation damages skin lipids [98] and lipid peroxidation leads to increased production of prostaglandins, promoting inflammation in the skin [99]. Additionally, after UV exposure, keratinocytes and other skin-related cells upregulate pro-inflammatory cytokine production, e.g., IL-1, IL-6 and TNF α , and induce the expression of vascular adhesion molecules. TNF α is considered central to mediating UV-induced inflammation [100].

Furthermore, UV radiation of sunlight has been demonstrated to generate ROS leading to oxidative stress in skin due to depletion of endogenous antioxidant enzymes [101–103]. Chronically sun-exposed skin demonstrated no difference to sun-protected skin with respect to expression levels of Cu/Zn-dependent superoxide dehydrogenase (SOD)1, Mn-dependent SOD2, and catalase; however, sun exposure induced a marked increase in heme oxygenase expression [98]. ROS generated by UV radiation primarily cause damage to DNA through oxidative modifications and mutations, but also by inducing expression of different genes, such as matrix metalloproteinases and collagenases, thereby affecting collagen integrity and skin aging [104,105]. Additionally, UV-mediated ROS generation also indirectly affects cellular function and survival via its effect on cell signaling pathways [106]. For instance, activation of MAPK proteins occurs after UV exposure, suggesting that it may be responsible for executing the effects of UV-induced oxidative stress [93,98].

Thus, UV-induced oxidative damage is due to either immediate damage from UV radiation or indirectly from activated immune cells and dysregulated cellular signaling. An important aspect of UV damage is the depletion of the antioxidant defenses, which leaves the skin vulnerable to additional ROS damage [107]. This redox imbalance may also have systemic effects due to the wide-ranging effects of ROS, thereby rendering the body more prone to other ROS-mediated pathologies. This is further supported by several studies demonstrating an association between psoriasis and the prevalence and incidence of diabetes [108–110], a condition with ROS-mediated pathology due to increased ROS production and impaired antioxidant defense [111].

3.2. Psoriasis

Psoriasis is a chronic inflammatory skin disease manifested by red, thickened skin and skin scales due to keratinocyte hyperproliferation and is caused by a multitude of factors, including genetic, immunological, and environmental factors [112]. The pathogenesis of psoriasis includes complex interactions between skin and immune cells in concert with growth factors and pro-inflammatory chemo- and cytokines [113,114]; however, the exact underlying mechanisms still remain to be elucidated.

Oxidative stress is believed to be a key factor in the pathogenesis of psoriasis [115], as studies have suggested the involvement of increased ROS levels in psoriasis pathogenesis [101,116]. Increased ROS generation by infiltrated leukocytes into psoriatic lesions [117] is accompanied by substantial biomolecular damage, like psoriatic skin lesions containing oxidized LDL [118,119]. Indeed, a relationship between psoriasis severity, lipoprotein levels and oxidative damage has been proposed [120]. Notably, decreased antioxidant levels have been found together with increased levels of lipid peroxidation markers in blood of psoriasis patients [121,122]. Also, serum levels of catalase were elevated in psoriatic patients [123] and increased activity of superoxide dismutase (SOD) [124], and expression levels of peroxiredoxin (Prdx)2 and glutathione peroxidase (GPx)6 have been found in psoriatic skin lesions [125,126]. It is tempting to speculate that increased compensatory antioxidant levels counteract the skewed redox balance.

Besides direct damaging effects of unregulated ROS production, dysregulation of several pro-inflammatory pathways, like MAPK, NF-κB, and JAK-STAT, has been considered to contribute to psoriasis etiology [127]. Several members of the MAPK signaling pathways, like ERK1/2, JNK, and MAPK, were activated in psoriatic skin, further supporting this notion [128–130].

NF-κB, another redox-sensitive transcription involved in cellular processes like inflammation, cell proliferation and survival, has recently been demonstrated to be upregulated and active in psoriatic skin [129,131,132]. Dysregulation of NF-κB-mediated signaling may further exaggerate disease severity, as NF-κB-mediated upregulation of pro-inflammatory cytokines activates NF-κB via a positive feedback loop [133]. Importantly, inhibition of NF-κB nuclear translocation and DNA binding activity dampens the inflammatory component of psoriasis [134–136]. Many of the cytokines involved in psoriasis pathology induce keratinocyte proliferation and these signals are via ROS-mediated action transmitted to transcription factors and associated proliferation pathways [137,138]. Together, these observations strongly support a misbalanced oxidant/antioxidant balance as the major culprit in sustaining the inflammatory component in the pathology of psoriasis.

3.3. Burn Injury

A skin burn is a posttraumatic inflammatory condition accompanied by local as well as distant effects, leading to exaggerated inflammation, tissue damage, and infection. After skin burn, molecular signals, including inflammatory mediators and oxidants, are released at the injury site, further contributing to tissue damage and ischemic tissue necrosis [139,140]. Liquid from burn injury blisters contains a substantial amount of keratinocyte-derived pro-inflammatory cytokine IL-1 β [141]. Moreover, induction of the inflammatory phase and attraction of immune cells leads to ROS production, exacerbating tissue damage [142]. Also, burns are often accompanied by secondary tissue

damage distant to the site of injury [143–145] that is thought to be mediated by ROS and activated immune cells/neutrophils [146,147]. Besides local effects, burns also initiate systemic inflammatory reactions via ROS production, leading to distant oxidative damage to lipids and proteins [144,148]. Additionally, this systemic inflammatory response contributes to secondary damage as neutrophils have been demonstrated in various organs distant from the burn site within hours after injury and may lead to exaggerated oxidative stress and damage [145]. Notably, the intensity of the systemic inflammatory response and subsequently ROS generation correlates with the severity of the burn injury [149,150]. This pathological ROS production may affect cell signaling networks at the site of injury, thereby damaging cells and biomolecules. Also, ROS-mediated lipid peroxidation in skin is an important cause of cellular membrane dysfunction and subsequently cell death after burn injury [151,152]. A correlation has been described between the lipid peroxidation load and the degree of complications [153–156].

Importantly, these pathological effects seem to be caused by an overwhelming of the antioxidant systems, as decreased antioxidant scavenging capacity and reduced levels of SOD, GSH, and bilirubin have been reported after burns [148,156–159], likely due to massive consumption of these antioxidants as an attempt to counteract the oxidative stress. Thus, a tight control of the cellular redox balance is crucial, as excessive ROS production or decreased activity of ROS detoxifying enzymes results in aberrant wound healing [160] caused by exacerbated tissue damage due to macromolecular damage or by responses via stress-induced pathways.

4. Maintaining the Redox Balance in the Skin

Free radicals are formed in the skin following exposure to environmental stimuli and immune reactions. In addition, ROS generation in skin occurs naturally as part of normal cellular metabolism, like mitochondrial respiration. These ROS are normally rapidly neutralized by non-enzymatic and enzymatic antioxidants, thereby maintaining the oxidant/antioxidant balance and thus tissue homeostasis [101] (Figure 1).

Figure 1. Redox balance maintenance in skin. ROS in the skin originate from normal cellular metabolism, e.g., mitochondrial respiration, and enzymatic activity. Besides, exogenous ROS are generated following physical insults, like UV light or persistent presence of leukocytes, facilitating chronic inflammatory skin conditions. To regulate ROS levels, the skin is rich in enzymatic and non-enzymatic antioxidant defense systems, thereby maintaining physiological homeostasis. In addition to the classical antioxidant defense, the cytoprotective enzyme heme oxygenase exhibits antioxidant properties via its degradation of pro-oxidant heme and generation of its antioxidant effector molecule bilirubin.



However, if the cytoprotective antioxidant factors get overwhelmed or are depleted, as may occur in hyperglycemic patients, the redox balance gets skewed towards oxidative stress, and may aggravate inflammatory injury.

4.1. Enzymatic Sources of Oxidants in the Skin

A major ROS source is the mitochondrial electron transport chain, leaking electrons during respiration. These electrons contribute to $O_2^{\bullet-}$ generation that is released into both the mitochondrial intermembranal space and matrix [69]. Matrix $O_2^{\bullet-}$ is converted to H_2O_2 by mitochondrial Mn-dependent SOD2 and can easily reach the cytosol by diffusion, whereas $O_2^{\bullet-}$ from the intermembranal space exits the mitochondria via voltage-dependent anion channels and is converted to H_2O_2 by Cu/Zn-dependent SOD1 in the cytosol [161].

Various enzyme systems produce ROS secondary to their main enzymatic function, including xanthine oxidoreductase [162], lipid peroxidases [163], cytochrome P450 enzymes [164], and nitric oxide synthase [165]. Nitric oxide (NO) synthase produces the free radical NO, which, under normal conditions acts protective, but which under oxidative circumstances gets converted into peroxynitrite, which is highly damaging [166]. Moreover, under certain conditions, like loss of cofactor tetrahydrobiopterine, NO synthase uncouples and produces O_2^{-} instead of NO [167] leading to oxidative stress affecting cardiovascular performance [168].

The most important enzyme responsible for ROS production is the membrane-bound enzyme complex NAPDH oxidase (Nox). The Nox family consists of 7 members, Nox1-5 and Duox1-2, bearing a catalytic Nox or Duox domain, respectively. Furthermore, Nox isoforms contain a stabilizing domain (p22phox) and several regulatory subunits [169]. They display differential expression, regulation, and subcellular localization, and produce different ROS products [170]. Nox1, 2 and 5 are the key sources of O_2^{--} , whereas Nox4 mainly produces H_2O_2 from molecular oxygen using NADPH as electron donor [171]. Nox-dependent ROS generation is activated by a wide range of chemical, physical, environmental, and biological factors [172]. The functions of the different Nox isoforms depend on their cellular localization and mode of activation [173] and have been thoroughly reviewed [174]. In keratinocytes, Nox1 is the only Nox expressed despite the detection of mRNAs encoding Nox1, Nox2, and Nox4 [175]. Keratinocyte Nox activity has been suggested to induce VEGF expression [176], MAPK activation [177], and cell growth [178]. Upon inflammation, infiltration of activated macrophages and neutrophils and their Nox2 and myeloperoxidase activities will exacerbate oxidative stress and prolong the inflammatory state [179].

4.2. Non-Enzymatic Sources of Oxidants in the Skin

Heme is crucial as the functional group of various hemoproteins, such as hemoglobin, cytochromes, peroxidases, and catalases [180]. In addition, heme can act as signaling molecule [181]. However, large amounts of free heme may be injurious to cells since heme is an iron chelate with the potential to catalyze iron-dependent reactions leading to ROS generation and membrane peroxidation [182]. Heme has indeed been demonstrated to catalyze ROS formation via the Fenton reaction [183] through its iron-dependent reaction with H_2O_2 [184,185].

After injury, large amounts of free heme are released from hemoproteins and aggravate tissue damage [182,186]. High local accumulation of heme can overwhelm the cellular ROS detoxification systems and prolongs oxidative and inflammatory stress [186–188]. Additionally, several studies have indicated that free heme also possesses pro-inflammatory properties [189–192], whereas low concentrations of free heme contribute to resolution of inflammation by downregulating inflammatory mediators, probably via induction of heme oxygenase (HO) activity [193–195]. Heme has therefore been suggested to act as a molecular switch due to its opposite, concentration-dependent effects [196].

Furthermore, free redox active metals, e.g., copper, zinc, and iron provide catalytic function to diverse (anti)oxidant enzymes [197], and redox cycling between Cu^+/Cu^{2+} and Fe^{2+}/Fe^{3+} is an integral part of the mitochondrial electron transport chain and ATP generation [198]. Transition metal homeostasis is regulated on both cellular and systemic levels and metal overload due to defective metal transporters is a central feature of several human diseases, like neurodegeneration [199]. Moreover, unregulated interaction of these metals with molecular oxygen also facilitates excessive ROS generation, predominantly via Fenton chemistry [200].

4.3. Antioxidant Systems in the Skin

Normally, the skin is constantly exposed to free radicals from the internal and external environment, challenging the functionality of the skin. Is our skin well enough prepared to withstand exogenous and endogenous ROS, and could targeting the redox status of the skin be a strategy to combat inflammatory skin conditions?

Under physiological conditions, ROS buildup in the skin is limited by numerous antioxidant defense systems, including both non-enzymatic and enzymatic mechanisms that either scavenge generated ROS before it can cause damage or prevent its formation (Figure 1, Table 1). In contrast to non-enzymatic antioxidants the enzymatic counterparts are not consumed and have a high affinity and reaction rate when scavenging ROS. Furthermore, the efficiency of the dietary non-enzymatic antioxidants depends on bioavailability as well as conversion into the active form upon ingestion [201] and antioxidant enzymes may therefore confer more efficient protection against acute oxidative and inflammatory stress [201].

Antioxidants	Examples	Target				
	Superoxide dismutase	Superoxide				
	Catalase	Hydrogen peroxide				
Enzymatic	Glutathione peroxidase	Hydrogen peroxide, lipid peroxides				
	Peroxiredoxin	Hydrogen peroxide				
	Heme oxygenase	Heme				
	Bilirubin	Lipid peroxides				
Non anzumatia	Vitamin C	Superoxide, hydroxyl radical, reactive				
Inon-enzymatic		nitrogen species, trace metals				
	Vitamin E	Lipid peroxides				
Metal-binding proteins	Ferritin	Free iron				

T٤	able	1.	Enzyme	s and	factor	s invo	lved	in	antioxi	dant	defense	in the	e skin.

The antioxidant defense system in skin is mainly comprised of the abundantly expressed antioxidant enzymes catalase, SOD, GPx, and Prdx [202].

SOD exists as three isoforms: cytosolic Cu/Zn-dependent SOD1, mitochondrial Mn-dependent SOD2, and extracellular SOD3 and catalyzes the conversion of O_2^{-} radicals into H_2O_2 using NAPDH as cofactor [203]. Excessive amounts of H_2O_2 are harmful to cells and rapid scavenging hereof is thus important [204]. This task is performed by either catalase, Prdx, or by GPx and the glutathione system that reduces the H_2O_2 to oxygen and water.

Mammalian cells may express 5 GPx isoforms [205], with GPx1 being the most prominent isoform in skin cells, reducing H_2O_2 and a range of organic peroxides [206]. GPx1 expression and activity is upregulated by ROS [207], and GPx1 and SOD display similar expression patterns [208], thereby securing an effective detoxification of H_2O_2 generated by SODs and avoiding the generation of HO• radicals. Moreover, GPx1 has been linked to the regulation of acute oxidative stress [209], as GPx1 knockout (KO) mice are highly susceptible to oxidative injury induced by paraquat and H_2O_2 [207] and GPx1 KO fibroblasts show increased sensitivity to oxidant-induced apoptosis [209].

Besides catalase and GPx, the six members of the Prdx family all catalyze the reduction of H₂O₂ and a wide spectrum of organic peroxides and peroxynitrite using GSH together with thioredoxin (Prdx1-5) or ascorbate (Prdx6), respectively [210]. Prdx6 is important in the cellular stress response, as Prdx6 overexpressing cells are protected from ROS-induced toxicity [211,212] and Prdx6 overexpressing keratinocytes are less sensitive towards photo-damage by UV radiation [213]. On the contrary, Prdx6 knockdown increases sensitivity towards oxidative stress [214–216].

Additionally, the NADPH/NADP+ ratio is an important index reflecting the cellular redox status. NADPH, the principal intracellular reductant, is generated from NADP+ by different groups of enzymes: (1) cytosolic glucose-6-phosphate dehydrogenase (G6PD) and 6-gluconate phosphate dehydrogenase; (2) cytosolic and mitochondrial isocitrate dehydrogenases; (3) cytosolic and mitochondrial malic enzymes; and (4) mitochondrial transhydrogenase [217]. NADPH is a central component in the cellular antioxidant system, as NADPH is required by glutathione reductase to reduce glutathione disulfide to GSH, an obligate co-substrate for GPxs [218]. Also, NAPDH is required by several pro-oxidant and antioxidant enzymes, including Nox and catalase [219]. G6PD is a major contributor to NADPH generation that despite its status as housekeeping enzyme is subjected to tissue-specific regulation by various factors, including oxidative stress, nutrients and hormones [220]. Notably, studies have shown that abrogation of G6PD activity dramatically increases cellular sensitivity to oxidative stress in vitro [221,222]. However, G6PD plays a dual role in the (anti)oxidative system as G6PD activity also provides Nox with NADPH. Obese and hyperglycemic rats demonstrated significantly higher Nox-derived O_2^{-} production due to increased G6PD activity and subsequently elevated NADPH levels to fuel this overproduction [223]. The resulting oxidative stress induced pathological changes of the heart and aorta and reduced cardiovascular function [223]. Together, this places G6PD and NAPDH centrally in the maintenance of a balanced ratio between pro- and antioxidants.

Besides the classical ROS-detoxifying enzymes discussed earlier, the HO system also exhibits potent antioxidant functions [224]. HO enzymes comprise the inducible HO-1 and the predominantly constitutively expressed HO-2 isoforms [225] and catalyze the degradation of heme into carbon monoxide (CO), iron, and biliverdin. Biliverdin is rapidly converted to bilirubin by biliverdin

reductase [226,227]. Because heme is a redox active molecule that is cytotoxic in high concentrations [228], a direct beneficial effect of HO activity on the cellular redox balance can be ascribed to its active heme detoxification.

Numerous studies have linked the HO system to the regulation of various (patho)physiological processes, including cellular adaptation to oxidative stress and promotion of inflammatory resolution [229]. The expression of HO-1 is induced by various ROS-producing stresses [230], including heme and heavy metals [231], UV light [98,232], and H₂O₂ [233]. Also, oxidative stress-mediated induction of HIF-1 stimulates HO-1 activity in many tissues [234]. Actually, HO-1 induction is considered a marker of cellular oxidative stress and is involved in the protective response against oxidative damage [235].

Moreover, HO-1 overexpression counteracts the cytotoxic effects caused by high concentrations of free heme [181,190,236,237], whereas inhibition of HO activity intensifies oxidative cellular and tissue damage [238–241]. Preclinical and epidemiological evidence indicate that the cytoprotective and oxidative effects of the HO system are mediated via the generated effector molecules CO, bilirubin/biliverdin and by co-induced ferritin [For a recent review, see 230].

Both biliverdin and bilirubin generated from HO-mediated heme degradation are strong antioxidants [242,243], and bilirubin ameliorates oxidative stress in different diseases, including atherosclerosis, diabetes mellitus, and inflammatory and autoimmune diseases (Recently reviewed in [244]). For instance, bilirubin protects cells against high H₂O₂ concentrations [242,245,246] and nanomolar concentrations of bilirubin suppress Nox activity *in vitro*, thereby reducing ROS levels and subsequent tissue damage [247–249]. Thus, the involvement of biliverdin and bilirubin in antioxidant defense is evident [250].

Ferrous iron (Fe²⁺) is released during heme degradation by HO enzymes and can participate in ROS-generating Fenton chemistry, leading to cellular damage [251]. Ferritin is a ubiquitous iron-binding protein that can accommodate up to 4,500 iron atoms per molecule [252,253]. Notably, ferritin expression is not only regulated by cellular iron levels but also by oxidative stress via Nrf2 binding to an antioxidant responsive element (ARE) in the ferritin promoter [254], leading to co-induction with HO-1 [196]. Thus, the HO system contributes significantly to the cellular antioxidant defense by degrading redox active heme, thereby generating ROS-targeting effector molecules that further contributing to counteracting oxidative stress.

Numerous studies using cellular and animal models have demonstrated protective effects of vitamin C, a water-soluble compound, and vitamin E, a fat-soluble compound, against cytoplasmic oxidative damage and lipid peroxidation, respectively, in the etiology of atherosclerosis. Disappointingly, clinical studies have turned out less promising [255].

Together, under normal physiological conditions, ROS-mediated reactions in the skin are well balanced and protect the cells against oxidative stress. However, under pathological conditions like inflammation excessive damaging ROS formation may occur due to overwhelming of the antioxidant systems, contributing to a worsened clinical outcome.

The skin is constantly subjected to ROS formation via UV irradiation, environmental exposure, and cellular metabolism and is rich in enzymatic and non-enzymatic antioxidant systems keeping the ROS levels at homeostatic levels. However, inflammatory conditions in combination with an overwhelmed antioxidant system leads to pathological levels of ROS and oxidative stress, further exacerbating disease state. Attenuation of ROS levels and restoring the redox balance could then normalize the inflammatory response.

A potential therapeutic approach to restore antioxidant levels in the skin could therefore be achieved through (1) reducing the ROS production; (2) increasing endogenous antioxidant enzymatic defenses; or (3) enhancing the non-enzymatic antioxidant defenses via dietary or pharmacological approaches.

5.1. Reduction of ROS Production

5.1.1. Nox Inhibition

Increased ROS production and oxidative stress contribute to cellular and tissue injury during the progress to chronic inflammatory skin diseases. Recent studies suggest that Nox-generated ROS contribute to cellular and tissue damage by fuelling the acute inflammatory response [174,256–259]. For instance, studies indicate that Nox-generated ROS contribute to TNF α -mediated activation of NF- κ B and vascular adhesion molecule expression [256,260–262]. Indeed, the central role of Nox in a multitude of diseases suggests it to be a putative therapeutic target, and several Nox-targeting inhibitors have been developed [263,264] with the main focus on the macrophage-specific Nox2 due to its involvement in several inflammatory conditions [173]. However, a recent study demonstrated that genetic Nox deficiency enhanced inflammatory responses after LPS challenge *in vivo*, suggesting that Nox-generated ROS in certain settings also have anti-inflammatory functions [265,266]. These contradictive and highly condition-specific outcomes of Nox-generated ROS may in the future challenge the use of Nox as therapeutic target to counteract redox imbalances.

5.1.2. Metal Scavenging Proteins

Redox active metals obstruct cellular signaling pathways by ROS-dependent and independent mechanisms. Both enzymatic and non-enzymatic antioxidants protect against metal-mediated ROS generation by (i) chelating redox-active metals; (ii) maintaining the metal redox state and preventing Fenton chemistry; and (iii) scavenging of metal-mediated ROS [200].

Excess free iron released after insults can catalyze ROS formation via Fenton reaction and cause tissue damage [228]. Furthermore, under *in vivo* stress conditions, an excess of superoxide releases iron from iron-binding molecules, including ferritin [267], contributing to further iron overload that can have deleterious effects [268,269]. Thus, the application of suitable metal chelators may contribute to a reduction in metal-induced ROS formation.

Chelation therapy is medical treatment for heavy metal poisoning and scavenging of redox active metals. Normally, labile cellular iron not contained by ferroproteins is scavenged by ferritin, thereby neutralizing the pro-inflammatory and pro-oxidative potential of free iron and preventing immune-mediated inflammatory conditions [270]. The bacterial siderophore Deferoxamine is an iron chelator frequently used to treat acute iron overload and related complications [271]. Also, plant-derived polyphenolic compounds (so-called botanicals) can be effective metal chelators [272]. For instance, the antioxidative effect of the plant-derived flavanoid quercetin is predominantly due to its chelating of redox-active iron [273].

5.2. Increasing Enzymatic Antioxidant Defenses

5.2.1. Antioxidant Upregulation

As mentioned earlier, superoxide scavenging is performed by the three mammalian SOD enzymes. The primary location of SOD3 is the extracellular matrix and on cell surfaces, whereas SOD1 and SOD2 are intracellularly located [274]. SOD3 is expressed in the epidermis and dermis of skin [275], but the role of SOD3 in this tissue is less clear. SODs have been suggested to be involved in the defense against UV-mediated ROS, as SOD enzyme expression and activity are affected by UV exposure [276–278]; however, SOD3 needs significantly higher UV doses before being activated than SOD1 and SOD2.

Numerous studies have investigated the antioxidative effects of elevated SOD levels in tissues by different means. For instance, intramuscular injections of SOD1 were successfully applied as anti-fibrotic therapy in treating cutaneous radiation-induced fibrosis in humans [279] and similar promising results were obtained with SOD2 in a porcine model of radiation-induced fibrosis [280].

Additionally, cutaneous SOD2 gene therapy reduced superoxide levels and normalized wound healing in mice with chemically-induced diabetes [281]. Similarly, diabetic transgenic SOD2 mice also demonstrated reduced superoxide levels and improved wound healing after ischemic stress compared to wild type controls [282].

Notably, chemically-induced contact dermatitis was alleviated in transgenic mice overexpressing SOD3 under the control of the keratin14 promoter, including a reduction in the levels of ROS and pro-inflammatory cytokines as well a reduced immune cell infiltration [283,284]. By contrast, SOD3 KO mice display exaggerated IL23-mediated psoriasis-like skin inflammation, including increased immune cell infiltration and higher levels of pro-inflammatory cytokines compared to WT controls [285,286], suggesting a role for SOD3 in cutaneous inflammation. Also, SOD3 expression is reduced in psoriasis patients compared to healthy subjects [284], further supporting this.

Moreover, SOD3 is suggested to play a role in pulmonary, arthritic, and neurological conditions [287]. Importantly, increasing SOD3 levels in various experimental disease models, e.g., chemically induced diabetes, hypertension, and inflammatory arthritis, reduced oxidative stress and improved disease state [288], thereby placing SOD3 as a central therapeutic target. Many studies have focused on SOD3 therapy in targeting ROS-mediated cardiovascular effects; however, the outcome has been variable [288]. Importantly, this discrepancy may relate to the use of predominantly rats in these studies despite that fact that rat SOD3 differs structurally and chemically from other mammalian SOD3 forms, resulting in a lower SOD3 concentration in rat vascular tissues [289], probably creating a therapeutic window that would not be present in other mammals, e.g., dogs, that did not show a therapeutic benefit from SOD3 gene therapy [290].

Thus, it is clear that increasing the enzymatic antioxidant defense by exogenous applications like injection or gene transfer may be a future therapeutic approach for multiple disorders with an inflammatory component; however, more research in terms of tissue- and condition-specific effects is warranted about the role of the SOD enzymes in skin inflammation.

5.2.2. Synthetic Antioxidants

Low-molecular mass synthetic compounds exhibiting catalytic activity, thus operating as enzyme mimics, have been developed as putative antioxidant therapy. The first mimetics were SOD-like because SOD is the first line of antioxidant defense, and today different classes of SOD mimetics based on metalloporphyrins, manganese (Mn) cyclic polyamines, and salen Mn derivatives have been developed [291]. Fortunately, their chemical and biophysical properties not only make them potent SOD mimics but also allow them to neutralize other types of ROS, including peroxynitrite and H₂O₂, and can thus also be considered catalase/peroxidase mimetics [292]. This broad specificity allows for modulation of the cellular redox environment and thus confers advantages over non-enzymatic antioxidants [293].

Moreover, these compounds have been shown to be effective in reducing oxidative stress in different *in vitro* cytotoxicity models involving ROS production [293,294] as well as in numerous *in vivo* models [292]. Notably, many of these compounds are not only functionally protective but also reduce oxidative damage to biomolecules [295–300] and the salen Mn compounds confer protection against mitochondrial damage [299].

Importantly, these mimetics display anti-inflammatory potential as H_2O_2 mediates activation of inflammatory genes as mentioned earlier, and therefore conditions with an inflammatory component may be the main target for such strategy. Also, reduced levels of activated macrophages were reported following treatment with mimetics in a radiation-induced lung injury model, however, no effects on pro-inflammatory cytokine levels were detected [300]. Also, a recent study demonstrated beneficial effects of systemic mimetic treatment on wound healing after skin irradiation by reducing oxidative damage [301].

Thus, these compounds do show promising prospects for therapeutic strategies in treating ROS-mediated complications in a wide range of conditions. Unfortunately, comparative studies on the different synthetic antioxidant classes in *in vitro* and *in vivo* settings are still very sparse and so far none of the antioxidant mimetics has been approved for clinical use.

5.2.3. Induction of the HO System

HO-1 is a stress-responsive enzyme with both antioxidant and cytoprotective effects mediated by the generated effector molecules biliverdin/bilirubin and CO, placing HO-1 as a central player in protection and homeostatic re-establishment after a wide range of pathological insults [302]. Numerous studies have demonstrated significant therapeutic effects of upregulation of HO-1 expression and/or activity as well as effector molecule administration in multiple pathological inflammatory conditions [230,302]. Also, ferritin is co-induced with HO-1 induction, which may provide a beneficial side effect, as ferritin scavenges free iron and thereby contributes to a restoration of the redox balance.

Recently, strategies to employ HO-1 as therapeutic target have been considered. This is further substantiated by HO-1 gene deficiency cases [303,304] and by the fact that promoter polymorphisms of the HMOX1 gene affect HO-1 protein expression levels and, subsequently, the severity of pathological human conditions [305]. Being an inducible enzyme, several synthetic molecules, including porphyrins [306] and heme arginate [307] have been developed and identified as possible HO-1 inducers. Heme arginate has for years been used as a clinical approach to treat porphyria, a disorder caused by non-functional heme metabolism [308]. Lately, a focus has been shed on the expanding number of plant-derived dietary compounds, so-called phytochemicals, with HO-1 inducing effects [309], like curcumin [310–312] or quercetin [313,314]. However, the effects of these pharmacological non-toxic, low-cost compounds still need to be studied in more detail before clinical use.

5.3. Increasing Non-Enzymatic Antioxidant Defenses

5.3.1. Oral and Topical Administration

Epidemiological studies have suggested that consumption of antioxidant-rich food is associated with lower disease rates and preventive protection of cardiovascular disease [315]. Thus, the supplementation of non-enzymatic, dietary antioxidants could be a feasible way of restoring redox homeostasis and reduce ROS-associated diseases.

However, clinical studies employing antioxidant supplement therapy have been inconclusive. For instance, the antioxidant compound *N*-acetyl cysteine (NAC) has been successfully used in the treatment of idiopathic pulmonary fibrosis [316–319], an inflammatory condition with etiology linked to Nox4-mediated ROS generation [320]; however, this therapeutic effect may not purely be ascribed to direct antioxidant effects of NAC itself but to its link to cellular glutathione replenishment [321]. Also, the combined use of NAC together with other commonly used treatment protocols should be carefully dissected and considered for each patient situation, as a recent study demonstrated increased mortality and severe treatment-related adverse effects when employing NAC in a three-drug regimen [322].

Frequently studied dietary and naturally occurring antioxidants such as carotenoids, flavonoids, and several vitamins have been implicated as promoters of skin health and rejuvenation [323]. External factors like chronic sun exposure, smoking, and pollution are significant contributors to skin aging, and both vitamins C and E have been demonstrated to have differential UVB photoprotective effects when applied both topically and orally [324,325]. Moreover, oral combination therapies of vitamins C and E resulted in a dramatically increased photoprotective effect compared to monotherapies [326]. Also oral intake of β -carotene or provitamin A reduces UV-induced erythema formation in different clinical studies; however, this effect is highly dose- and time-dependent [327–329]. Notably, β -carotene has been demonstrated *in vitro* to quench UV-induced radical formation and lipid peroxidation [330,331] and to reduce mitochondrial mutagenesis after UV exposure of skin fibroblasts [332].

Polyphenols are plant-derived micronutrients such as green tea polyphenols and curcumin that also have gained more attention in skin research during the last decade due to their antioxidant properties and their potential beneficial effects on cancer, neurodegenerative and cardiovascular diseases that all have been linked to oxidative stress [333]. Various polyphenols have been reported to be photoprotectors [334]. Oral as well as topical application of green tea polyphenols to mice protected in a time-dependent manner against UV-induced cutaneous edema, depletion of the epidermal antioxidant defense, and cyclooxygenase induction [335,336]. Studies using animal models have also demonstrated anti-inflammatory activities of administration of green tea polyphenols, predominantly mediated by the major green tea polyphenols dose-dependently reduced erythema formation and sunburn cells [337]. Sunburn cells are keratinocytes undergoing apoptosis due to irreversible DNA damage [339]. However, the underlying mechanism of these compounds is still not well understood but has been suggested to be mediated via effects on signal transduction pathways [340].

Another powerful antioxidant involved in skin antioxidant defense is coenzyme Q10 (CoQ10) [341]. However, being part of the respiratory chain, CoQ10 also contributes to ROS formation, as discussed earlier. In skin, the CoQ10 level is 10 times higher in the epidermis compared to the dermis [341]. Being the outermost skin layer, the epidermis is directly exposed to UV irradiation and it is known that UV irradiation depletes antioxidants in the skin [107]. The epidermis may thus be an optimal target for CoQ10 administration. Dietary CoQ10 supplementation to rats increased the levels of CoQ10 and its homologs in tissues and mitochondria therein [342]. This was accompanied by a reductive shift in plasma aminothiol status and decreased oxidative damage to mitochondrial proteins in skeletal muscle [342]. In contrast, mice on CoQ10 enriched diets did not show any effect on the systemic redox balance, nor the lifespan, despite a buildup of CoQ10 in tissues [343]. Notably, human epidermal keratinocytes isolated from topically CoQ10-treated skin demonstrated improved mitochondrial function and protection against UV-induced mitochondrial damage compared to non-treated controls [344]. Other studies demonstrated that CoQ10 stabilizes mitochondrial function, improves cell viability, and attenuates oxidative effects in human skin [345,346].

In summary, despite their great availability and use, the effects of dietary supplements on skin and general health still remain controversial. However, several things that could be crucial for treatment outcome must be considered. Firstly, these bioactive compounds have to be prepared and taken up by the target organ. The stability of these compounds will ultimately determine efficiency, and the use of vitamin C in creams has proven difficult due to a low stability in the presence of oxygen [347]. To account for this and to facilitate uptake, more stable derivates are often used though several of these compounds are not efficiently converted into the active form of the antioxidant [348]. Another issue is the bioavailability of the oral supplements. Dietary supplements have to pass through the gastrointestinal tract, enter circulation, and reach the target tissue and selected cell types and/or cellular compartments, which may be important for the effective dose to be given. Also, toxic effects due to e.g., cross-reactions and organ-specific reactions to certain compounds should be taken into account [349]. Additionally, route of administration is important, as e.g., curcumin is effective to skin when applied topically, but only to the colon when applied orally [340]. Importantly, antioxidant supplementation may interfere with the endogenous antioxidant response normally initiated after exercise and subsequently interferes with ROS signaling [350]. Other clinical studies on vitamins A, C and E, coenzyme Q10, carotene, and plant-derived flavanoids have turned out rather disappointing, as no significant effects these dietary supplements on general health have been detected [351,352]. More

importantly, the long-term effects of many of these supplements are yet to be assessed, but one study linked increased mortality to long-term intake of antioxidant supplements [353].

Interestingly, the oxidated targeted site may determine the success of antioxidant therapy. Although both GSH and bilirubin are potent endogenous antioxidants, they protect against distinct targets. GSH mainly protects against hydrophilic proteins, whereas bilirubin protects against lipid peroxidation [354]. Recently, we demonstrated that iatrogenic induction of mild hyperbilirubinemia ameliorated the serum antioxidant status and vascular function in diabetic patients [355]. Thus, more in-depth studies are still needed to gain more knowledge about *in vivo* and *in vitro* obtained data on the diverse group of potential beneficial antioxidants. Knowledge concerning dose-response, optimal administration, cellular and compartmental targeting and translational studies are still urgently needed.

5.3.2. Targeted Antioxidant Delivery

Mitochondria are central organelles in cell survival and function [356] and increasing attention has been paid to the role of mitochondrial dysfunction in aging, apoptosis, neurodegeneration, and cancer [357]. Despite already being equipped with antioxidant systems, great interest has been paid to developing supplementary approaches to further protect the mitochondria from ROS-mediated damage. For instance, CoQ10 and related ubiquinones have been used as therapy to decrease mitochondrial damage in Parkinson's disease patients [358,359]. However, delivery issues limited the therapeutic effect as oral administration only resulted in a limited mitochondrial uptake of ubiquinones [360–362]. Conjugation of a lipophilic triphenylphosphonium cation to ubiquinones led to the development of MitoQ, which selectively is taken up by and accumulated within mitochondria [363]. Moreover, MitoQ efficiently prevented oxidative stress in isolated mitochondria, as well as in the ischemic heart [367–366]. Later, other effective compounds such as SkQ1 and Trolex were developed and characterized [367–369]. Lately, nanotechnology has been employed in targeted mitochondrial delivery. Different drugs employed in cancer, Alzheimer's disease, and obesity demonstrated improvement in the drug therapeutic index after nanoparticle-mediated delivery compared to a non-targeted carrier or to the free form of the therapeutics [370].

Notably, a recent report described induction and subsequently mitochondrial translocation of HO-1 following gastric mucosal injury, resulting in prevention of mitochondrial oxidative stress and pathology [371], suggesting that HO-1 induction may represent another mitochondrial targeting strategy. Together, this targeted approach thus holds big potential for future treatment strategies.

4. Concluding Remarks

ROS are crucial for cellular functions and provide essential protective mechanisms. However, ROS and oxidative stress have also been linked to various disease states, including inflammation, diabetes mellitus, cardiovascular diseases, and aging [88]. A well-balanced redox homeostasis is important, as oxidative stress either by increased levels of ROS and/or by depleted antioxidant systems may dysregulate protein function due to oxidative modifications and further aggravate inflammatory conditions. Targeting oxidative stress in inflammatory skin conditions may ameliorate disease outcome by dampening inflammation and improving recovery. However, although promising, efficacy and clinical studies employing antioxidant therapy are still in its infancy.

Acknowledgments

This work was supported by grants from the Dutch Burns Foundation (#09.110) and TASENE (NWO-WOTRO; W02.29.101).

Conflict of Interest

The authors declare no conflict of interest.

References

- 1. Contassot, E.; Beer, H.D.; French, L.E. Interleukin-1, inflammasomes, autoinflammation and the skin. *Swiss Med. Wkly.* **2012**, *142*, doi:10.4414/smw.2012.13590.
- 2. Martinon, F.; Mayor, A.; Tschopp, J. The inflammasomes: Guardians of the body. *Annu. Rev. Immunol.* **2009**, *27*, 229–265.
- 3. Feldmeyer, L.; Werner, S.; French, L.E.; Beer, H.D. Interleukin-1, inflammasomes and the skin. *Eur. J. Cell Biol.* **2010**, *89*, 638–644.
- 4. Nestle, F.O.; Di Meglio, P.; Qin, J.Z.; Nickoloff, B.J. Skin immune sentinels in health and disease. *Nat. Rev. Immunol.* **2009**, *9*, 679–691.
- 5. Fuchs, E.; Raghavan, S. Getting under the skin of epidermal morphogenesis. *Nat. Rev. Genet.* **2002**, *3*, 199–209.
- 6. Sorrell, J.M.; Caplan, A.I. Fibroblast heterogeneity: More than skin deep. J. Cell Sci. 2004, 117, 667–675.
- Glaser, R.; Harder, J.; Lange, H.; Bartels, J.; Christophers, E.; Schroder, J.M. Antimicrobial psoriasin (S100A7) protects human skin from Escherichia coli infection. *Nat. Immunol.* 2005, *6*, 57–64.
- 8. Masters, S.L. Specific inflammasomes in complex diseases. *Clin. Immunol.* **2012**, doi:10.1016/j.clim.2012.12.006.
- 9. Weber, A.; Wasiliew, P.; Kracht, M. Interleukin-1 (IL-1) pathway. *Sci. Signal.* 2010, *3*, doi:10.1126/scisignal.3105cm1.
- 10. Bergsbaken, T.; Fink, S.L.; Cookson, B.T. Pyroptosis: Host cell death and inflammation. *Nat. Rev. Microbiol.* **2009**, *7*, 99–109.
- 11. Yeretssian, G.; Labbe, K.; Saleh, M. Molecular regulation of inflammation and cell death. *Cytokine* **2008**, *43*, 380–390.
- Miao, E.A.; Leaf, I.A.; Treuting, P.M.; Mao, D.P.; Dors, M.; Sarkar, A.; Warren, S.E.; Wewers, M.D.; Aderem, A. Caspase-1-induced pyroptosis is an innate immune effector mechanism against intracellular bacteria. *Nat. Immunol.* 2010, *11*, 1136–1142.
- Feldmeyer, L.; Keller, M.; Niklaus, G.; Hohl, D.; Werner, S.; Beer, H.D. The inflammasome mediates UVB-induced activation and secretion of interleukin-1beta by keratinocytes. *Curr. Biol.* 2007, *17*, 1140–1145.
- Watanabe, H.; Gaide, O.; Petrilli, V.; Martinon, F.; Contassot, E.; Roques, S.; Kummer, J.A.; Tschopp, J.; French, L.E. Activation of the IL-1beta-processing inflammasome is involved in contact hypersensitivity. *J. Investig. Dermatol.* 2007, *127*, 1956–1963.

- Kupper, T.S.; Ballard, D.W.; Chua, A.O.; McGuire, J.S.; Flood, P.M.; Horowitz, M.C.; Langdon, R.; Lightfoot, L.; Gubler, U. Human keratinocytes contain mRNA indistinguishable from monocyte interleukin 1 alpha and beta mRNA. Keratinocyte epidermal cell-derived thymocyte-activating factor is identical to interleukin 1. *J. Exp. Med.* **1986**, *164*, 2095–2100.
- Kondo, S.; Sauder, D.N.; Kono, T.; Galley, K.A.; McKenzie, R.C. Differential modulation of interleukin-1 alpha (IL-1 alpha) and interleukin-1 beta (IL-1 beta) in human epidermal keratinocytes by UVB. *Exp. Dermatol.* 1994, *3*, 29–39.
- 17. Barker, J.N.; Mitra, R.S.; Griffiths, C.E.; Dixit, V.M.; Nickoloff, B.J. Keratinocytes as initiators of inflammation. *Lancet* **1991**, *337*, 211–214.
- 18. Zhou, R.; Tardivel, A.; Thorens, B.; Choi, I.; Tschopp, J. Thioredoxin-interacting protein links oxidative stress to inflammasome activation. *Nat. Immunol.* **2010**, *11*, 136–140.
- 19. Tschopp, J.; Schroder, K. NLRP3 inflammasome activation: The convergence of multiple signalling pathways on ROS production? *Nat. Rev. Immunol.* **2010**, *10*, 210–215.
- Nakahira, K.; Haspel, J.A.; Rathinam, V.A.; Lee, S.J.; Dolinay, T.; Lam, H.C.; Englert, J.A.; Rabinovitch, M.; Cernadas, M.; Kim, H.P.; *et al.* Autophagy proteins regulate innate immune responses by inhibiting the release of mitochondrial DNA mediated by the NALP3 inflammasome. *Nat. Immunol.* 2011, *12*, 222–230.
- Shimada, K.; Crother, T.R.; Karlin, J.; Dagvadorj, J.; Chiba, N.; Chen, S.; Ramanujan, V.K.; Wolf, A.J.; Vergnes, L.; Ojcius, D.M.; *et al.* Oxidized mitochondrial DNA activates the NLRP3 inflammasome during apoptosis. *Immunity* 2012, *36*, 401–414.
- 22. Droge, W. Free radicals in the physiological control of cell function. *Physiol. Rev.* 2002, 82, 47–95.
- Forman, H.J.; Fukuto, J.M.; Miller, T.; Zhang, H.; Rinna, A.; Levy, S. The chemistry of cell signaling by reactive oxygen and nitrogen species and 4-hydroxynonenal. *Arch. Biochem. Biophys.* 2008, 477, 183–195.
- 24. Niethammer, P.; Grabher, C.; Look, A.T.; Mitchison, T.J. A tissue-scale gradient of hydrogen peroxide mediates rapid wound detection in zebrafish. *Nature* **2009**, *459*, 996–999.
- 25. Kamata, H.; Hirata, H. Redox regulation of cellular signalling. Cell. Signal. 1999, 11, 1-14.
- 26. Trachootham, D.; Lu, W.; Ogasawara, M.A.; Nilsa, R.D.; Huang, P. Redox regulation of cell survival. *Antioxid. Redox Signal.* **2008**, *10*, 1343–1374.
- 27. Finkel, T. Signal transduction by reactive oxygen species. J. Cell. Biol. 2011, 194, 7–15.
- 28. Rhee, S.G.; Chang, T.S.; Bae, Y.S.; Lee, S.R.; Kang, S.W. Cellular regulation by hydrogen peroxide. *J. Am. Soc. Nephrol.* **2003**, *14*, S211–S215.
- 29. Haddad, J.J. Antioxidant and prooxidant mechanisms in the regulation of redox(y)-sensitive transcription factors. *Cell. Signal.* **2002**, *14*, 879–897.
- Pastore, A.; Piemonte, F. S-Glutathionylation signaling in cell biology: Progress and prospects. *Eur. J. Pharm. Sci.* 2012, 46, 279–292.
- 31. Jones, D.P. Radical-free biology of oxidative stress. Am. J. Physiol. Cell Physiol. 2008, 295, C849-C868.
- 32. Fritz, G.; Grosch, S.; Tomicic, M.; Kaina, B. APE/Ref-1 and the mammalian response to genotoxic stress. *Toxicology* **2003**, *193*, 67–78.

- Rahman, I.; Marwick, J.; Kirkham, P. Redox modulation of chromatin remodeling: Impact on histone acetylation and deacetylation, NF-kappaB and pro-inflammatory gene expression. *Biochem. Pharm.* 2004, 68, 1255–1267.
- 34. England, K.; Cotter, T.G. Direct oxidative modifications of signalling proteins in mammalian cells and their effects on apoptosis. *Redox Rep.* **2005**, *10*, 237–245.
- 35. Bourdon, E.; Blache, D. The importance of proteins in defense against oxidation. *Antioxid. Redox Signal.* **2001**, *3*, 293–311.
- 36. Pantano, C.; Reynaert, N.L.; van der Vliet, A.; Janssen-Heininger, Y.M. Redox-sensitive kinases of the nuclear factor-kappaB signaling pathway. *Antioxid. Redox Signal.* **2006**, *8*, 1791–1806.
- Shao, D.; Oka, S.; Brady, C.D.; Haendeler, J.; Eaton, P.; Sadoshima, J. Redox modification of cell signaling in the cardiovascular system. *J. Mol. Cell. Cardiol.* 2012, *52*, 550–558.
- 38. Jacob, C.; Giles, G.I.; Giles, N.M.; Sies, H. Sulfur and selenium: The role of oxidation state in protein structure and function. *Angew. Chem.* **2003**, *42*, 4742–4758.
- Martyniuk, C.J.; Fang, B.; Koomen, J.M.; Gavin, T.; Zhang, L.; Barber, D.S.; Lopachin, R.M. Molecular mechanism of glyceraldehyde-3-phosphate dehydrogenase inactivation by alpha,beta-unsaturated carbonyl derivatives. *Chem. Res. Toxicol.* 2011, 24, 2302–2311.
- 40. Go, Y.M.; Duong, D.M.; Peng, J.; Jones, D.P. Protein cysteines map to functional networks according to steady-state level of oxidation. *J. Proteomics Bioinform.* **2011**, *4*, 196–209.
- 41. Forman, H.J.; Fukuto, J.M.; Torres, M. Redox signaling: Thiol chemistry defines which reactive oxygen and nitrogen species can act as second messengers. *Am. J. Physiol. Cell Physiol.* **2004**, 287, C246–C256.
- 42. Winterbourn, C.C.; Hampton, M.B. Thiol chemistry and specificity in redox signaling. *Free Radic. Biol. Med.* **2008**, *45*, 549–561.
- 43. Paulsen, C.E.; Carroll, K.S. Orchestrating redox signaling networks through regulatory cysteine switches. *ACS Chem. Biol.* **2010**, *5*, 47–62.
- Chung, H.S.; Wang, S.B.; Venkatraman, V.; Murray, C.I.; Van Eyk, J.E. Cysteine oxidative posttranslational modifications: Emerging regulation in the cardiovascular system. *Circ. Res.* 2013, *112*, 382–392.
- 45. Denu, J.M.; Dixon, J.E. Protein tyrosine phosphatases: Mechanisms of catalysis and regulation. *Curr. Opin. Chem. Biol.* **1998**, *2*, 633–641.
- 46. Ostman, A.; Frijhoff, J.; Sandin, A.; Bohmer, F.D. Regulation of protein tyrosine phosphatases by reversible oxidation. *J. Biochem.* **2011**, *150*, 345–356.
- 47. Tu, B.P.; Weissman, J.S. Oxidative protein folding in eukaryotes: Mechanisms and consequences. *J. Cell. Biol.* **2004**, *164*, 341–346.
- 48. Frand, A.R.; Cuozzo, J.W.; Kaiser, C.A. Pathways for protein disulphide bond formation. *Trends Cell. Biol.* **2000**, *10*, 203–210.
- 49. Jones, D.P.; Go, Y.M.; Anderson, C.L.; Ziegler, T.R.; Kinkade, J.M., Jr.; Kirlin, W.G. Cysteine/cystine couple is a newly recognized node in the circuitry for biologic redox signaling and control. *FASEB J.* **2004**, *18*, 1246–1248.
- 50. Chen, C.Y.; Willard, D.; Rudolph, J. Redox regulation of SH2-domain-containing protein tyrosine phosphatases by two backdoor cysteines. *Biochemistry* **2009**, *48*, 1399–1409.

- 52. Lee, S.R.; Yang, K.S.; Kwon, J.; Lee, C.; Jeong, W.; Rhee, S.G. Reversible inactivation of the tumor suppressor PTEN by H₂O₂. *J. Biol. Chem.* **2002**, *277*, 20336–20342.
- 53. Sohn, J.; Rudolph, J. Catalytic and chemical competence of regulation of cdc25 phosphatase by oxidation/reduction. *Biochemistry* **2003**, *42*, 10060–10070.
- 54. Tonks, N.K. Redox redux: Revisiting PTPs and the control of cell signaling. *Cell* **2005**, *121*, 667–670.
- 55. Klomsiri, C.; Karplus, P.A.; Poole, L.B. Cysteine-based redox switches in enzymes. *Antioxid. Redox Signal.* 2011, 14, 1065–1077.
- Shackelford, R.E.; Heinloth, A.N.; Heard, S.C.; Paules, R.S. Cellular and molecular targets of protein S-glutathiolation. *Antioxid. Redox Signal.* 2005, 7, 940–950.
- 57. Shlomai, J. Redox control of protein-DNA interactions: From molecular mechanisms to significance in signal transduction, gene expression, and DNA replication. *Antioxid. Redox Signal.* **2010**, *13*, 1429–1476.
- Dinkova-Kostova, A.T.; Holtzclaw, W.D.; Cole, R.N.; Itoh, K.; Wakabayashi, N.; Katoh, Y.; Yamamoto, M.; Talalay, P. Direct evidence that sulfhydryl groups of Keap1 are the sensors regulating induction of phase 2 enzymes that protect against carcinogens and oxidants. *Proc. Natl. Acad. Sci. USA* 2002, 99, 11908–11913.
- 59. Shelton, M.D.; Mieyal, J.J. Regulation by reversible *S*-glutathionylation: Molecular targets implicated in inflammatory diseases. *Mol. Cells* **2008**, *25*, 332–346.
- Qanungo, S.; Starke, D.W.; Pai, H.V.; Mieyal, J.J.; Nieminen, A.L. Glutathione supplementation potentiates hypoxic apoptosis by S-glutathionylation of p65-NFkappaB. J. Biol. Chem. 2007, 282, 18427–18436.
- 61. Cross, J.V.; Templeton, D.J. Regulation of signal transduction through protein cysteine oxidation. *Antioxid. Redox Signal.* **2006**, *8*, 1819–1827.
- 62. Gilmore, T.D. Introduction to NF-kappaB: Players, pathways, perspectives. *Oncogene* **2006**, *25*, 6680–6684.
- 63. Benhar, M.; Forrester, M.T.; Stamler, J.S. Protein denitrosylation: Enzymatic mechanisms and cellular functions. *Nat. Rev. Mol. Cell Biol.* **2009**, *10*, 721–732.
- 64. Benhar, M.; Forrester, M.T.; Hess, D.T.; Stamler, J.S. Regulated protein denitrosylation by cytosolic and mitochondrial thioredoxins. *Science* **2008**, *320*, 1050–1054.
- 65. Lima, B.; Forrester, M.T.; Hess, D.T.; Stamler, J.S. *S*-nitrosylation in cardiovascular signaling. *Circ. Res.* **2010**, *106*, 633–646.
- Evangelista, A.M.; Kohr, M.J.; Murphy, E. S-Nitrosylation: Specificity, occupancy, and interaction with other post-translational modifications. *Antioxid. Redox Signal.* 2013, doi:10.1089/ars.2012.5056.
- 67. Becker, K.; Savvides, S.N.; Keese, M.; Schirmer, R.H.; Karplus, P.A. Enzyme inactivation through sulfhydryl oxidation by physiologic NO-carriers. *Nat. Struct. Biol.* **1998**, *5*, 267–271.
- 68. Kim, S.; Wing, S.S.; Ponka, P. S-nitrosylation of IRP2 regulates its stability via the ubiquitin-proteasome pathway. *Mol. Cell. Biol.* **2004**, *24*, 330–337.

- 69. Murphy, M.P. How mitochondria produce reactive oxygen species. *Biochem. J.* 2009, 417, 1–13.
- Collins, Y.; Chouchani, E.T.; James, A.M.; Menger, K.E.; Cocheme, H.M.; Murphy, M.P. Mitochondrial redox signalling at a glance. J. Cell Sci. 2012, 125, 801–806.
- Murphy, M.P.; Holmgren, A.; Larsson, N.G.; Halliwell, B.; Chang, C.J.; Kalyanaraman, B.; Rhee, S.G.; Thornalley, P.J.; Partridge, L.; Gems, D.; *et al.* Unraveling the biological roles of reactive oxygen species. *Cell Metab.* 2011, *13*, 361–366.
- Hoehn, K.L.; Salmon, A.B.; Hohnen-Behrens, C.; Turner, N.; Hoy, A.J.; Maghzal, G.J.; Stocker, R.; van Remmen, H.; Kraegen, E.W.; Cooney, G.J.; *et al.* Insulin resistance is a cellular antioxidant defense mechanism. *Proc. Natl. Acad. Sci. USA* 2009, *106*, 17787–17792.
- 73. Zhou, L.; Aon, M.A.; Almas, T.; Cortassa, S.; Winslow, R.L.; O'Rourke, B. A reaction-diffusion model of ROS-induced ROS release in a mitochondrial network. *PLoS Comput. Biol.* **2010**, *6*, e1000657.
- 74. Winterbourn, C.C. Reconciling the chemistry and biology of reactive oxygen species. *Nat. Chem. Biol.* **2008**, *4*, 278–286.
- 75. Hausladen, A.; Fridovich, I. Superoxide and peroxynitrite inactivate aconitases, but nitric oxide does not. *J. Biol. Chem.* **1994**, *269*, 29405–29408.
- Fridovich, I. Superoxide anion radical (O₂⁻⁻), superoxide dismutases, and related matters. J. Biol. Chem. 1997, 272, 18515–18517.
- 77. Vasquez-Vivar, J.; Kalyanaraman, B.; Kennedy, M.C. Mitochondrial aconitase is a source of hydroxyl radical. An electron spin resonance investigation. *J. Biol. Chem.* **2000**, *275*, 14064–14069.
- 78. James, A.M.; Collins, Y.; Logan, A.; Murphy, M.P. Mitochondrial oxidative stress and the metabolic syndrome. *Trends Endocrinol. Metab.* **2012**, *23*, 429–434.
- 79. Kietzmann, T.; Gorlach, A. Reactive oxygen species in the control of hypoxia-inducible factor-mediated gene expression. *Semin. Cell Dev. Biol.* **2005**, *16*, 474–486.
- Murphy, M.P. Modulating mitochondrial intracellular location as a redox signal. *Sci. Signal.* 2012, 5, doi:10.1126/scisignal.2003386.
- Chandel, N.S.; Maltepe, E.; Goldwasser, E.; Mathieu, C.E.; Simon, M.C.; Schumacker, P.T. Mitochondrial reactive oxygen species trigger hypoxia-induced transcription. *Proc. Natl. Acad. Sci. USA* 1998, 95, 11715–11720.
- Patten, D.A.; Lafleur, V.N.; Robitaille, G.A.; Chan, D.A.; Giaccia, A.J.; Richard, D.E. Hypoxia-inducible factor-1 activation in nonhypoxic conditions: The essential role of mitochondrial-derived reactive oxygen species. *Mol. Biol. Cell* 2010, *21*, 3247–3257.
- 83. Ushio-Fukai, M.; Nakamura, Y. Reactive oxygen species and angiogenesis: NADPH oxidase as target for cancer therapy. *Cancer Lett.* **2008**, *266*, 37–52.
- Tsai, Y.P.; Wu, K.J. Hypoxia-regulated target genes implicated in tumor metastasis. *J. Biomed. Sci.* 2012, 19, doi:10.1186/1423-0127-19-102.
- 85. Acker, T.; Fandrey, J.; Acker, H. The good, the bad and the ugly in oxygen-sensing: ROS, cytochromes and prolyl-hydroxylases. *Cardiovasc. Res.* **2006**, *71*, 195–207.
- Al-Mehdi, A.B.; Pastukh, V.M.; Swiger, B.M.; Reed, D.J.; Patel, M.R.; Bardwell, G.C.; Pastukh, V.V.; Alexeyev, M.F.; Gillespie, M.N. Perinuclear mitochondrial clustering creates an oxidant-rich nuclear domain required for hypoxia-induced transcription. *Sci. Signal.* 2012, *5*, doi:10.1126/scisignal.2002712.

- Valko, M.; Leibfritz, D.; Moncol, J.; Cronin, M.T.; Mazur, M.; Telser, J. Free radicals and antioxidants in normal physiological functions and human disease. *Int. J. Biochem. Cell Biol.* 2007, 39, 44–84.
- 88. Alfadda, A.A.; Sallam, R.M. Reactive oxygen species in health and disease. J. Biomed. Biotechnol. 2012, 2012, doi:10.1155/2012/936486.
- 89. Mailloux, R.J.; Harper, M.E. Mitochondrial proticity and ROS signaling: Lessons from the uncoupling proteins. *Trends Endocrinol. Metab.* **2012**, *23*, 451–458.
- 90. Lenaz, G. Mitochondria and reactive oxygen species. Which role in physiology and pathology? *Adv. Exp. Med. Biol.* **2012**, *942*, 93–136.
- Garcia-Bailo, B.; El-Sohemy, A.; Haddad, P.S.; Arora, P.; Benzaied, F.; Karmali, M.; Badawi, A. Vitamins D, C, and E in the prevention of type 2 diabetes mellitus: Modulation of inflammation and oxidative stress. *Biologics* 2011, *5*, 7–19.
- 92. Nasti, T.H.; Timares, L. Inflammasome activation of IL-1 family mediators in response to cutaneous photodamage. *Photochem. Photobiol.* **2012**, *88*, 1111–1125.
- 93. Muthusamy, V.; Piva, T.J. The UV response of the skin: A review of the MAPK, NFkappaB and TNFalpha signal transduction pathways. *Arch. Dermatol. Res.* **2010**, *302*, 5–17.
- 94. Diaz, J.H.; Nesbitt, L.T., Jr. Sun exposure behavior and protection: Recommendations for travelers. *J. Travel Med.* **2013**, *20*, 108–118.
- 95. Aroun, A.; Zhong, J.L.; Tyrrell, R.M.; Pourzand, C. Iron, oxidative stress and the example of solar ultraviolet A radiation. *Photochem. Photobiol. Sci.* **2012**, *11*, 118–134.
- 96. Rastogi, R.P.; Richa; Kumar, A.; Tyagi, M.B.; Sinha, R.P. Molecular mechanisms of ultraviolet radiation-induced DNA damage and repair. *J. Nucleic Acids* **2010**, *2010*, doi:10.4061/2010/592980.
- 97. Valko, M.; Rhodes, C.J.; Moncol, J.; Izakovic, M.; Mazur, M. Free radicals, metals and antioxidants in oxidative stress-induced cancer. *Chem. Biol. Interact.* **2006**, *160*, 1–40.
- Akasaka, E.; Takekoshi, S.; Horikoshi, Y.; Toriumi, K.; Ikoma, N.; Mabuchi, T.; Tamiya, S.; Matsuyama, T.; Ozawa, A. Protein oxidative damage and heme oxygenase in sunlight-exposed human skin: Roles of MAPK responses to oxidative stress. *Tokai J. Exp. Clin. Med.* 2010, 35, 152–164.
- Hruza, L.L.; Pentland, A.P. Mechanisms of UV-induced inflammation. J. Investig. Dermatol. 1993, 100, 35S–41S.
- 100. Clydesdale, G.J.; Dandie, G.W.; Muller, H.K. Ultraviolet light induced injury: Immunological and inflammatory effects. *Immunol. Cell Biol.* **2001**, *79*, 547–568.
- 101. Bickers, D.R.; Athar, M. Oxidative stress in the pathogenesis of skin disease. J. Investig. Dermatol. 2006, 126, 2565–2575.
- 102. Vile, G.F.; Tanew-Ilitschew, A.; Tyrrell, R.M. Activation of NF-kappa B in human skin fibroblasts by the oxidative stress generated by UVA radiation. *Photochem. Photobiol.* 1995, 62, 463–468.
- Vile, G.F.; Tyrrell, R.M. UVA radiation-induced oxidative damage to lipids and proteins *in vitro* and in human skin fibroblasts is dependent on iron and singlet oxygen. *Free Radic. Biol. Med.* 1995, *18*, 721–730.

- 104. Scharffetter, K.; Wlaschek, M.; Hogg, A.; Bolsen, K.; Schothorst, A.; Goerz, G.; Krieg, T.; Plewig, G. UVA irradiation induces collagenase in human dermal fibroblasts *in vitro* and *in vivo*. *Arch. Dermatol. Res.* **1991**, *283*, 506–511.
- Wlaschek, M.; Briviba, K.; Stricklin, G.P.; Sies, H.; Scharffetter-Kochanek, K. Singlet oxygen may mediate the ultraviolet A-induced synthesis of interstitial collagenase. *J. Investig. Dermatol.* 1995, 104, 194–198.
- 106. Ryter, S.W.; Tyrrell, R.M. Singlet molecular oxygen ((1)O2): A possible effector of eukaryotic gene expression. *Free Radic. Biol. Med.* **1998**, *24*, 1520–1534.
- 107. Podda, M.; Traber, M.G.; Weber, C.; Yan, L.J.; Packer, L. UV-irradiation depletes antioxidants and causes oxidative damage in a model of human skin. *Free Radic. Biol. Med.* **1998**, *24*, 55–65.
- 108. Armstrong, A.W.; Harskamp, C.T.; Armstrong, E.J. Psoriasis and the risk of diabetes mellitus: A systematic review and meta-analysis. *JAMA Dermatol.* **2013**, *149*, 84–91.
- 109. Azfar, R.S.; Seminara, N.M.; Shin, D.B.; Troxel, A.B.; Margolis, D.J.; Gelfand, J.M. Increased risk of diabetes mellitus and likelihood of receiving diabetes mellitus treatment in patients with psoriasis. *Arch. Dermatol.* **2012**, *148*, 995–1000.
- 110. Cheng, J.; Kuai, D.; Zhang, L.; Yang, X.; Qiu, B. Psoriasis increased the risk of diabetes: A meta-analysis. *Arch. Dermatol. Res.* **2012**, *304*, 119–125.
- 111. Maritim, A.C.; Sanders, R.A.; Watkins, J.B. 3rd Diabetes, oxidative stress, and antioxidants: A review. *J. Biochem. Mol. Toxicol.* **2003**, *17*, 24–38.
- 112. Enamandram, M.; Kimball, A.B. Psoriasis epidemiology: The interplay of genes and the environment. *J. Investig. Dermatol.* **2013**, *133*, 287–289.
- 113. Nickoloff, B.J.; Xin, H.; Nestle, F.O.; Qin, J.Z. The cytokine and chemokine network in psoriasis. *Clin. Dermatol.* **2007**, *25*, 568–573.
- 114. Lowes, M.A.; Bowcock, A.M.; Krueger, J.G. Pathogenesis and therapy of psoriasis. *Nature* **2007**, *445*, 866–873.
- 115. Zhou, Q.; Mrowietz, U.; Rostami-Yazdi, M. Oxidative stress in the pathogenesis of psoriasis. *Free Radic. Biol. Med.* **2009**, *47*, 891–905.
- Briganti, S.; Picardo, M. Antioxidant activity, lipid peroxidation and skin diseases. What's new. J. Eur. Acad. Dermatol. Venereol. 2003, 17, 663–669.
- 117. Racz, E.; Prens, E.P. Molecular pathophysiology of psoriasis and molecular targets of antipsoriatic therapy. *Expert Rev. Mol. Med.* **2009**, *11*, e38.
- 118. Rocha-Pereira, P.; Santos-Silva, A.; Rebelo, I.; Figueiredo, A.; Quintanilha, A.; Teixeira, F. Dislipidemia and oxidative stress in mild and in severe psoriasis as a risk for cardiovascular disease. *Clin. Chim. Acta* 2001, *303*, 33–39.
- 119. Tekin, N.S.; Tekin, I.O.; Barut, F.; Sipahi, E.Y. Accumulation of oxidized low-density lipoprotein in psoriatic skin and changes of plasma lipid levels in psoriatic patients. *Mediators Inflamm.* 2007, 2007, 78454.
- 120. Simonetti, O.; Ferretti, G.; Salvi, A.; Offidani, A.M.; Bossi, G. Plasma lipid changes in psoriatic children. *Dermatology* **1992**, *185*, 96–100.
- 121. Kokcam, I.; Naziroglu, M. Antioxidants and lipid peroxidation status in the blood of patients with psoriasis. *Clin. Chim. Acta* **1999**, *289*, 23–31.

- 122. Drewa, G.; Krzyzynska-Malinowska, E.; Wozniak, A.; Protas-Drozd, F.; Mila-Kierzenkowska, C.; Rozwodowska, M.; Kowaliszyn, B.; Czajkowski, R. Activity of superoxide dismutase and catalase and the level of lipid peroxidation products reactive with TBA in patients with psoriasis. *Med. Sci. Monit.* 2002, *8*, BR338–BR343.
- 123. Yildirim, M.; Inaloz, H.S.; Baysal, V.; Delibas, N. The role of oxidants and antioxidants in psoriasis. *J. Eur. Acad. Dermatol. Venereol.* **2003**, *17*, 34–36.
- 124. Therond, P.; Gerbaud, P.; Dimon, S.; Anderson, W.B.; Evain-Broin, D.; Raynaud, F. Antioxidant enzymes in psoriatic fibroblasts and erythrocytes. *J. Investig. Dermatol.* **1996**, *106*, 1325–1328.
- 125. Frank, S.; Munz, B.; Werner, S. The human homologue of a bovine non-selenium glutathione peroxidase is a novel keratinocyte growth factor-regulated gene. *Oncogene* **1997**, *14*, 915–921.
- 126. Ryu, J.; Park, S.G.; Park, B.C.; Choe, M.; Lee, K.S.; Cho, J.W. Proteomic analysis of psoriatic skin tissue for identification of differentially expressed proteins: Up-regulation of GSTP1, SFN and PRDX2 in psoriatic skin. *Int. J. Mol. Med.* 2011, 28, 785–792.
- Armstrong, A.W.; Voyles, S.V.; Armstrong, E.J.; Fuller, E.N.; Rutledge, J.C. Angiogenesis and oxidative stress: Common mechanisms linking psoriasis with atherosclerosis. *J. Dermatol. Sci.* 2011, 63, 1–9.
- 128. Takahashi, H.; Ibe, M.; Nakamura, S.; Ishida-Yamamoto, A.; Hashimoto, Y.; Iizuka, H. Extracellular regulated kinase and c-Jun *N*-terminal kinase are activated in psoriatic involved epidermis. *J. Dermatol. Sci.* **2002**, *30*, 94–99.
- 129. Johansen, C.; Flindt, E.; Kragballe, K.; Henningsen, J.; Westergaard, M.; Kristiansen, K.; Iversen, L. Inverse regulation of the nuclear factor-kappaB binding to the p53 and interleukin-8 kappaB response elements in lesional psoriatic skin. J. Investig. Dermatol. 2005, 124, 1284–1292.
- 130. Yu, X.J.; Li, C.Y.; Dai, H.Y.; Cai, D.X.; Wang, K.Y.; Xu, Y.H.; Chen, L.M.; Zhou, C.L. Expression and localization of the activated mitogen-activated protein kinase in lesional psoriatic skin. *Exp. Mol. Pathol.* 2007, *83*, 413–418.
- 131. Abdou, A.G.; Hanout, H.M. Evaluation of survivin and NF-kappaB in psoriasis, an immunohistochemical study. J. Cutan. Pathol. 2008, 35, 445–451.
- Lizzul, P.F.; Aphale, A.; Malaviya, R.; Sun, Y.; Masud, S.; Dombrovskiy, V.; Gottlieb, A.B. Differential expression of phosphorylated NF-kappaB/RelA in normal and psoriatic epidermis and downregulation of NF-kappaB in response to treatment with etanercept. *J. Investig. Dermatol.* 2005, *124*, 1275–1283.
- 133. Lawrence, T. The nuclear factor NF-kappaB pathway in inflammation. *Cold Spring Harb. Perspect. Biol.* **2009**, *1*, doi:10.1101/cshperspect.a001651.
- 134. Treumer, F.; Zhu, K.; Glaser, R.; Mrowietz, U. Dimethylfumarate is a potent inducer of apoptosis in human T cells. *J. Investig. Dermatol.* **2003**, *121*, 1383–1388.
- 135. Amigo, M.; Paya, M.; De Rosa, S.; Terencio, M.C. Antipsoriatic effects of avarol-3'-thiosalicylate are mediated by inhibition of TNF-alpha generation and NF-kappaB activation in mouse skin. *Br. J. Pharmacol.* 2007, *152*, 353–365.
- 136. Amigo, M.; Schalkwijk, J.; Olthuis, D.; De Rosa, S.; Paya, M.; Terencio, M.C.; Lamme, E. Identification of avarol derivatives as potential antipsoriatic drugs using an *in vitro* model for keratinocyte growth and differentiation. *Life Sci.* 2006, 79, 2395–2404.

- 137. Teunissen, M.B.; Koomen, C.W.; de Waal Malefyt, R.; Wierenga, E.A.; Bos, J.D. Interleukin-17 and interferon-gamma synergize in the enhancement of proinflammatory cytokine production by human keratinocytes. *J. Investig. Dermatol.* **1998**, *111*, 645–649.
- 138. Bito, T.; Nishigori, C. Impact of reactive oxygen species on keratinocyte signaling pathways. *J. Dermatol. Sci.* **2012**, *68*, 3–8.
- 139. Kao, C.C.; Garner, W.L. Acute burns. Plast. Reconstr. Surg. 2000, 101, 2482-2493.
- 140. Latha, B.; Babu, M. The involvement of free radicals in burn injury: A review. *Burns* 2001, 27, 309–317.
- 141. Kupper, T.S.; Deitch, E.A.; Baker, C.C.; Wong, W.C. The human burn wound as a primary source of interleukin-1 activity. *Surgery* **1986**, *100*, 409–415.
- 142. Ward, P.A.; Till, G.O. Pathophysiologic events related to thermal injury of skin. *J. Trauma* **1990**, *30*, S75–S79.
- 143. Till, G.O.; Beauchamp, C.; Menapace, D.; Tourtellotte, W., Jr.; Kunkel, R.; Johnson, K.J.; Ward, P.A. Oxygen radical dependent lung damage following thermal injury of rat skin. *J. Trauma* 1983, 23, 269–277.
- Till, G.O.; Hatherill, J.R.; Tourtellotte, W.W.; Lutz, M.J.; Ward, P.A. Lipid peroxidation and acute lung injury after thermal trauma to skin. Evidence of a role for hydroxyl radical. *Am. J. Pathol.* 1985, *119*, 376–384.
- 145. Gurbuz, V.; Corak, A.; Yegen, B.C.; Kurtel, H.; Alican, I. Oxidative organ damage in a rat model of thermal injury: The effect of cyclosporin A. *Burns* **1997**, *23*, 37–42.
- 146. Jahovic, N.; Guzel, E.; Arbak, S.; Yegen, B.C. The healing-promoting effect of saliva on skin burn is mediated by epidermal growth factor (EGF): Role of the neutrophils. *Burns* 2004, *30*, 531–538.
- Cetinkale, O.; Konukoglu, D.; Senel, O.; Kemerli, G.D.; Yazar, S. Modulating the functions of neutrophils and lipid peroxidation by FK506 in a rat model of thermal injury. *Burns* 1999, 25, 105–112.
- 148. Haycock, J.W.; Ralston, D.R.; Morris, B.; Freedlander, E.; MacNeil, S. Oxidative damage to protein and alterations to antioxidant levels in human cutaneous thermal injury. *Burns* **1997**, *23*, 533–540.
- 149. Tredget, E.E.; Yu, Y.M. The metabolic effects of thermal injury. World J. Surg. 1992, 16, 68–79.
- Bertin-Maghit, M.; Goudable, J.; Dalmas, E.; Steghens, J.P.; Bouchard, C.; Gueugniaud, P.Y.; Petit, P.; Delafosse, B. Time course of oxidative stress after major burns. *Intensive Care Med.* 2000, 26, 800–803.
- 151. Horton, J.W. Free radicals and lipid peroxidation mediated injury in burn trauma: The role of antioxidant therapy. *Toxicology* **2003**, *189*, 75–88.
- 152. Demling, R.; Ikegami, K.; Lalonde, C. Increased lipid peroxidation and decreased antioxidant activity correspond with death after smoke exposure in the rat. *J. Burn Care Rehabil.* **1995**, *16*, 104–110.
- 153. Hosnuter, M.; Gurel, A.; Babuccu, O.; Armutcu, F.; Kargi, E.; Isikdemir, A. The effect of CAPE on lipid peroxidation and nitric oxide levels in the plasma of rats following thermal injury. *Burns* 2004, *30*, 121–125.

- 154. Saez, J.C.; Ward, P.H.; Gunther, B.; Vivaldi, E. Superoxide radical involvement in the pathogenesis of burn shock. *Circ. Shock* **1984**, *12*, 229–239.
- 155. Oldham, K.T.; Guice, K.S.; Till, G.O.; Ward, P.A. Activation of complement by hydroxyl radical in thermal injury. *Surgery* **1988**, *104*, 272–279.
- 156. Sener, G.; Sehirli, A.O.; Satiroglu, H.; Keyer-Uysal, M.; Yegen, B.C. Melatonin improves oxidative organ damage in a rat model of thermal injury. *Burns* **2002**, *28*, 419–425.
- 157. Saitoh, D.; Ookawara, T.; Fukuzuka, K.; Kawakami, M.; Sakamoto, T.; Ohno, H.; Okada, Y. Characteristics of plasma extracellular SOD in burned patients. *Burns* **2001**, *27*, 577–581.
- 158. Saitoh, D.; Shirani, K.Z.; Cioffi, W.G.; Kizaki, T.; Ohno, H.; Okada, Y.; Mason, A.D., Jr.; Pruitt, B.A., Jr. Changes in the tissue and plasma superoxide dismutase (SOD) levels in a burned rat model. *Tohoku J. Exp. Med.* 2001, 193, 27–36.
- Cetinkale, O.; Belce, A.; Konukoglu, D.; Senyuva, C.; Gumustas, M.K.; Tas, T. Evaluation of lipid peroxidation and total antioxidant status in plasma of rats following thermal injury. *Burns* 1997, 23, 114–116.
- 160. Steiling, H.; Munz, B.; Werner, S.; Brauchle, M. Different types of ROS-scavenging enzymes are expressed during cutaneous wound repair. *Exp. Cell Res.* **1999**, *247*, 484–494.
- Sena, L.A.; Chandel, N.S. Physiological roles of mitochondrial reactive oxygen species. *Mol. Cell* 2012, 48, 158–167.
- Agarwal, A.; Banerjee, A.; Banerjee, U.C. Xanthine oxidoreductase: A journey from purine metabolism to cardiovascular excitation-contraction coupling. *Crit. Rev. Biotechnol.* 2011, 31, 264–280.
- 163. Zhang, R.; Brennan, M.L.; Shen, Z.; MacPherson, J.C.; Schmitt, D.; Molenda, C.E.; Hazen, S.L. Myeloperoxidase functions as a major enzymatic catalyst for initiation of lipid peroxidation at sites of inflammation. *J. Biol. Chem.* 2002, 277, 46116–46122.
- 164. Fleming, I.; Michaelis, U.R.; Bredenkotter, D.; Fisslthaler, B.; Dehghani, F.; Brandes, R.P.; Busse, R. Endothelium-derived hyperpolarizing factor synthase (Cytochrome P450 2C9) is a functionally significant source of reactive oxygen species in coronary arteries. *Circ. Res.* 2001, 88, 44–51.
- 165. Tabima, D.M.; Frizzell, S.; Gladwin, M.T. Reactive oxygen and nitrogen species in pulmonary hypertension. *Free Radic. Biol. Med.* **2012**, *52*, 1970–1986.
- 166. Andrew, P.J.; Mayer, B. Enzymatic function of nitric oxide synthases. *Cardiovasc. Res.* 1999, 43, 521–531.
- Xia, Y.; Tsai, A.L.; Berka, V.; Zweier, J.L. Superoxide generation from endothelial nitric-oxide synthase. A Ca2+/calmodulin-dependent and tetrahydrobiopterin regulatory process. *J. Biol. Chem.* 1998, 273, 25804–25808.
- 168. Roe, N.D.; Ren, J. Nitric oxide synthase uncoupling: A therapeutic target in cardiovascular diseases. *Vasc. Pharmacol.* **2012**, *57*, 168–172.
- 169. Altenhofer, S.; Kleikers, P.W.; Radermacher, K.A.; Scheurer, P.; Rob Hermans, J.J.; Schiffers, P.; Ho, H.; Wingler, K.; Schmidt, H.H. The NOX toolbox: Validating the role of NADPH oxidases in physiology and disease. *Cell. Mol. Life Sci.* **2012**, *69*, 2327–2343.
- 170. Brown, D.I.; Griendling, K.K. Nox proteins in signal transduction. *Free Radic. Biol. Med.* **2009**, 47, 1239–1253.

- 171. Takac, I.; Schroder, K.; Zhang, L.; Lardy, B.; Anilkumar, N.; Lambeth, J.D.; Shah, A.M.; Morel, F.; Brandes, R.P. The E-loop is involved in hydrogen peroxide formation by the NADPH oxidase Nox4. J. Biol. Chem. 2011, 286, 13304–13313.
- 172. Jiang, F.; Zhang, Y.; Dusting, G.J. NADPH oxidase-mediated redox signaling: Roles in cellular stress response, stress tolerance, and tissue repair. *Pharmacol. Rev.* **2011**, *63*, 218–242.
- 173. Lambeth, J.D.; Kawahara, T.; Diebold, B. Regulation of Nox and Duox enzymatic activity and expression. *Free Radic. Biol. Med.* **2007**, *43*, 319–331.
- 174. Bedard, K.; Krause, K.H. The NOX family of ROS-generating NADPH oxidases: Physiology and pathophysiology. *Physiol. Rev.* **2007**, *87*, 245–313.
- 175. Chamulitrat, W.; Stremmel, W.; Kawahara, T.; Rokutan, K.; Fujii, H.; Wingler, K.; Schmidt, H.H.; Schmidt, R. A constitutive NADPH oxidase-like system containing gp91phox homologs in human keratinocytes. J. Investig. Dermatol. 2004, 122, 1000–1009.
- 176. Sen, C.K.; Khanna, S.; Babior, B.M.; Hunt, T.K.; Ellison, E.C.; Roy, S. Oxidant-induced vascular endothelial growth factor expression in human keratinocytes and cutaneous wound healing. *J. Biol. Chem.* **2002**, *277*, 33284–33290.
- 177. Chiu, C.; Maddock, D.A.; Zhang, Q.; Souza, K.P.; Townsend, A.R.; Wan, Y. TGF-beta-induced p38 activation is mediated by Rac1-regulated generation of reactive oxygen species in cultured human keratinocytes. *Int. J. Mol. Med.* **2001**, *8*, 251–255.
- 178. Chamulitrat, W.; Schmidt, R.; Tomakidi, P.; Stremmel, W.; Chunglok, W.; Kawahara, T.; Rokutan, K. Association of gp91phox homolog Nox1 with anchorage-independent growth and MAP kinase-activation of transformed human keratinocytes. *Oncogene* **2003**, *22*, 6045–6053.
- 179. Steinbeck, M.J.; Khan, A.U.; Karnovsky, M.J. Extracellular production of singlet oxygen by stimulated macrophages quantified using 9,10-diphenylanthracene and perylene in a polystyrene film. *J. Biol. Chem.* **1993**, *268*, 15649–15654.
- Tsiftsoglou, A.S.; Tsamadou, A.I.; Papadopoulou, L.C. Heme as key regulator of major mammalian cellular functions: Molecular, cellular, and pharmacological aspects. *Pharmacol. Ther.* 2006, 111, 327–345.
- 181. Wagener, F.A.; Volk, H.D.; Willis, D.; Abraham, N.G.; Soares, M.P.; Adema, G.J.; Figdor, C.G. Different faces of the heme-heme oxygenase system in inflammation. *Pharmacol. Rev.* 2003, 55, 551–571.
- Ryter, S.W.; Tyrrell, R.M. The heme synthesis and degradation pathways: Role in oxidant sensitivity. Heme oxygenase has both pro- and antioxidant properties. *Free Radic. Biol. Med.* 2000, 28, 289–309.
- Halliwell, B.; Gutteridge, J.M. Oxygen toxicity, oxygen radicals, transition metals and disease. *Biochem. J.* 1984, 219, 1–14.
- 184. Kumar, S.; Bandyopadhyay, U. Free heme toxicity and its detoxification systems in human. *Toxicol. Lett.* **2005**, *157*, 175–188.
- 185. Thomas, D.D.; Espey, M.G.; Vitek, M.P.; Miranda, K.M.; Wink, D.A. Protein nitration is mediated by heme and free metals through Fenton-type chemistry: An alternative to the NO/O2– reaction. *Proc. Natl. Acad. Sci. USA* 2002, *99*, 12691–12696.
- 186. Jeney, V.; Balla, J.; Yachie, A.; Varga, Z.; Vercellotti, G.M.; Eaton, J.W.; Balla, G. Pro-oxidant and cytotoxic effects of circulating heme. *Blood* **2002**, *100*, 879–887.

- 187. Balla, G.; Jacob, H.S.; Eaton, J.W.; Belcher, J.D.; Vercellotti, G.M. Hemin: A possible physiological mediator of low density lipoprotein oxidation and endothelial injury. *Arterioscler. Thromb.* 1991, 11, 1700–1711.
- 188. Balla, J.; Jacob, H.S.; Balla, G.; Nath, K.; Eaton, J.W.; Vercellotti, G.M. Endothelial-cell heme uptake from heme proteins: Induction of sensitization and desensitization to oxidant damage. *Proc. Natl. Acad. Sci. USA* **1993**, *90*, 9285–9289.
- 189. Wagener, F.A.; Abraham, N.G.; van, K.Y.; de, W.T.; Figdor, C.G. Heme-induced cell adhesion in the pathogenesis of sickle-cell disease and inflammation. *Trends Pharmacol. Sci.* **2001**, *22*, 52–54.
- 190. Wagener, F.A.; da Silva, J.L.; Farley, T.; de, W.T.; Kappas, A.; Abraham, N.G. Differential effects of heme oxygenase isoforms on heme mediation of endothelial intracellular adhesion molecule 1 expression. *J. Pharmacol. Exp. Ther.* **1999**, *291*, 416–423.
- 191. Wagener, F.A.; Feldman, E.; de, W.T.; Abraham, N.G. Heme induces the expression of adhesion molecules ICAM-1, VCAM-1, and E selectin in vascular endothelial cells. *Proc. Soc. Exp. Biol. Med.* 1997, 216, 456–463.
- 192. Tolosano, E.; Fagoonee, S.; Hirsch, E.; Berger, F.G.; Baumann, H.; Silengo, L.; Altruda, F. Enhanced splenomegaly and severe liver inflammation in haptoglobin/hemopexin double-null mice after acute hemolysis. *Blood* **2002**, *100*, 4201–4208.
- 193. Ma, J.L.; Yang, P.Y.; Rui, Y.C.; Lu, L.; Kang, H.; Zhang, J. Hemin modulates cytokine expressions in macrophage-derived foam cells via heme oxygenase-1 induction. *J. Pharmacol. Sci.* 2007, *103*, 261–266.
- 194. Cambos, M.; Bazinet, S.; Abed, E.; Sanchez-Dardon, J.; Bernard, C.; Moreau, R.; Olivier, M.; Scorza, T. The IL-12p70/IL-10 interplay is differentially regulated by free heme and hemozoin in murine bone-marrow-derived macrophages. *Int J. Parasitol.* 2010, 40, 1003–1012.
- 195. Cambos, M.; Scorza, T. Robust erythrophagocytosis leads to macrophage apoptosis via a hemin-mediated redox imbalance: Role in hemolytic disorders. *J. Leukoc. Biol.* **2011**, *89*, 159–171.
- 196. Wagener, F.A.; van Beurden, H.E.; von den Hoff, J.W.; Adema, G.J.; Figdor, C.G. The heme-heme oxygenase system: A molecular switch in wound healing. *Blood* **2003**, *102*, 521–528.
- 197. Harris, E.D. Regulation of antioxidant enzymes. FASEB J. 1992, 6, 2675-2683.
- 198. Yoshikawa, S.; Muramoto, K.; Shinzawa-Itoh, K. The O(2) reduction and proton pumping gate mechanism of bovine heart cytochrome c oxidase. *Biochim. Biophys. Acta* **2011**, *1807*, 1279–1286.
- 199. Martinez-Finley, E.J.; Chakraborty, S.; Fretham, S.J.; Aschner, M. Cellular transport and homeostasis of essential and nonessential metals. *Metallomics* **2012**, *4*, 593–605.
- 200. Jomova, K.; Valko, M. Advances in metal-induced oxidative stress and human disease. *Toxicology* **2011**, *283*, 65–87.
- 201. Ratnam, D.V.; Ankola, D.D.; Bhardwaj, V.; Sahana, D.K.; Kumar, M.N. Role of antioxidants in prophylaxis and therapy: A pharmaceutical perspective. J. Control. Release 2006, 113, 189–207.
- 202. Packer, L.; Valacchi, G. Antioxidants and the response of skin to oxidative stress: Vitamin E as a key indicator. *Skin Pharmacol. Appl. Skin Physiol.* 2002, 15, 282–290.
- 203. Zelko, I.N.; Mariani, T.J.; Folz, R.J. Superoxide dismutase multigene family: A comparison of the CuZn-SOD (SOD1), Mn-SOD (SOD2), and EC-SOD (SOD3) gene structures, evolution, and expression. *Free Radic. Biol. Med.* 2002, *33*, 337–349.

- 204. Zamocky, M.; Jakopitsch, C.; Furtmuller, P.G.; Dunand, C.; Obinger, C. The peroxidase-cyclooxygenase superfamily: Reconstructed evolution of critical enzymes of the innate immune system. *Proteins* **2008**, *72*, 589–605.
- Brigelius-Flohe, R. Glutathione peroxidases and redox-regulated transcription factors. *Biol. Chem.* 2006, 387, 1329–1335.
- 206. Arthur, J.R. The glutathione peroxidases. Cell. Mol. Life Sci. 2000, 57, 1825-1835.
- 207. De Haan, J.B.; Bladier, C.; Griffiths, P.; Kelner, M.; O'Shea, R.D.; Cheung, N.S.; Bronson, R.T.; Silvestro, M.J.; Wild, S.; Zheng, S.S.; *et al.* Mice with a homozygous null mutation for the most abundant glutathione peroxidase, Gpx1, show increased susceptibility to the oxidative stress-inducing agents paraquat and hydrogen peroxide. *J. Biol. Chem.* **1998**, *273*, 22528–22536.
- 208. Auf dem Keller, U.; Kumin, A.; Braun, S.; Werner, S. Reactive oxygen species and their detoxification in healing skin wounds. J. Investig. Dermatol. Symp. Proc. 2006, 11, 106–111.
- 209. De Haan, J.B.; Crack, P.J.; Flentjar, N.; Iannello, R.C.; Hertzog, P.J.; Kola, I. An imbalance in antioxidant defense affects cellular function: The pathophysiological consequences of a reduction in antioxidant defense in the glutathione peroxidase-1 (Gpx1) knockout mouse. *Redox Rep.* 2003, 8, 69–79.
- Hanschmann, E.M.; Godoy, J.R.; Berndt, C.; Hudemann, C.; Lillig, C.H. Thioredoxins, glutaredoxins, and peroxiredoxins-molecular mechanisms and health significance: From cofactors to antioxidants to redox signaling. *Antioxid. Redox Signal.* 2013, doi:10.1089/ars.2012.4599.
- 211. Manevich, Y.; Sweitzer, T.; Pak, J.H.; Feinstein, S.I.; Muzykantov, V.; Fisher, A.B. 1-Cys peroxiredoxin overexpression protects cells against phospholipid peroxidation-mediated membrane damage. *Proc. Natl. Acad. Sci. USA* 2002, *99*, 11599–11604.
- Wang, Y.; Manevich, Y.; Feinstein, S.I.; Fisher, A.B. Adenovirus-mediated transfer of the 1-cys peroxiredoxin gene to mouse lung protects against hyperoxic injury. *Am. J. Physiol. Lung Cell. Mol. Physiol.* 2004, 286, L1188–L1193.
- 213. Kumin, A.; Huber, C.; Rulicke, T.; Wolf, E.; Werner, S. Peroxiredoxin 6 is a potent cytoprotective enzyme in the epidermis. *Am. J. Pathol.* **2006**, *169*, 1194–1205.
- Pak, J.H.; Manevich, Y.; Kim, H.S.; Feinstein, S.I.; Fisher, A.B. An antisense oligonucleotide to 1-cys peroxiredoxin causes lipid peroxidation and apoptosis in lung epithelial cells. *J. Biol. Chem.* 2002, 277, 49927–49934.
- 215. Mo, Y.; Feinstein, S.I.; Manevich, Y.; Zhang, Q.; Lu, L.; Ho, Y.S.; Fisher, A.B. 1-Cys peroxiredoxin knock-out mice express mRNA but not protein for a highly related intronless gene. *FEBS Lett.* 2003, 555, 192–198.
- 216. Wang, X.; Phelan, S.A.; Forsman-Semb, K.; Taylor, E.F.; Petros, C.; Brown, A.; Lerner, C.P.; Paigen, B. Mice with targeted mutation of peroxiredoxin 6 develop normally but are susceptible to oxidative stress. *J. Biol. Chem.* **2003**, *278*, 25179–25190.
- 217. Ying, W. NAD+/NADH and NADP+/NADPH in cellular functions and cell death: Regulation and biological consequences. *Antioxid. Redox Signal.* **2008**, *10*, 179–206.
- 218. Wu, G.; Fang, Y.Z.; Yang, S.; Lupton, J.R.; Turner, N.D. Glutathione metabolism and its implications for health. *J. Nutr.* **2004**, *134*, 489–492.
- 219. Stanton, R.C. Glucose-6-phosphate dehydrogenase, NADPH, and cell survival. *IUBMB Life* 2012, 64, 362–369.

- Kletzien, R.F.; Harris, P.K.; Foellmi, L.A. Glucose-6-phosphate dehydrogenase: A "housekeeping" enzyme subject to tissue-specific regulation by hormones, nutrients, and oxidant stress. *FASEB J.* 1994, *8*, 174–181.
- 221. Pandolfi, P.P.; Sonati, F.; Rivi, R.; Mason, P.; Grosveld, F.; Luzzatto, L. Targeted disruption of the housekeeping gene encoding glucose 6-phosphate dehydrogenase (G6PD): G6PD is dispensable for pentose synthesis but essential for defense against oxidative stress. *EMBO J.* 1995, 14, 5209–5215.
- 222. Rosenstraus, M.; Chasin, L.A. Isolation of mammalian cell mutants deficient in glucose-6-phosphate dehydrogenase activity: Linkage to hypoxanthine phosphoribosyl transferase. *Proc. Natl. Acad. Sci. USA* 1975, 72, 493–497.
- 223. Serpillon, S.; Floyd, B.C.; Gupte, R.S.; George, S.; Kozicky, M.; Neito, V.; Recchia, F.; Stanley, W.; Wolin, M.S.; Gupte, S.A. Superoxide production by NAD(P)H oxidase and mitochondria is increased in genetically obese and hyperglycemic rat heart and aorta before the development of cardiac dysfunction. The role of glucose-6-phosphate dehydrogenase-derived NADPH. *Am. J. Physiol. Heart .Circ. Physiol.* 2009, 297, H153–H162.
- 224. Haines, D.D.; Lekli, I.; Teissier, P.; Bak, I.; Tosaki, A. Role of haeme oxygenase-1 in resolution of oxidative stress-related pathologies: Focus on cardiovascular, lung, neurological and kidney disorders. *Acta Physiol.* **2012**, *204*, 487–501.
- 225. Abraham, N.G.; Kappas, A. Pharmacological and clinical aspects of heme oxygenase. *Pharmacol. Rev.* 2008, *60*, 79–127.
- 226. Maines, M.D. Heme oxygenase: Function, multiplicity, regulatory mechanisms, and clinical applications. *FASEB J.* **1988**, *2*, 2557–2568.
- 227. Tenhunen, R.; Marver, H.S.; Schmid, R. The enzymatic conversion of heme to bilirubin by microsomal heme oxygenase. *Proc. Natl. Acad. Sci. USA* **1968**, *61*, 748–755.
- 228. Wagener, F.A.; Scharstuhl, A.; Tyrrell, R.M.; Von den Hoff, J.W.; Jozkowicz, A.; Dulak, J.; Russel, F.G.; Kuijpers-Jagtman, A.M. The heme-heme oxygenase system in wound healing; Implications for scar formation. *Curr. Drug Targets* 2010, *11*, 1571–1585.
- 229. Lundvig, D.M.; Immenschuh, S.; Wagener, F.A. Heme oxygenase, inflammation, and fibrosis: The good, the bad, and the ugly? *Front. Pharmacol.* **2012**, *3*, 81.
- 230. Grochot-Przeczek, A.; Dulak, J.; Jozkowicz, A. Haem oxygenase-1: Non-canonical roles in physiology and pathology. *Clin. Sci.* **2012**, *122*, 93–103.
- 231. Alam, J.; Shibahara, S.; Smith, A. Transcriptional activation of the heme oxygenase gene by heme and cadmium in mouse hepatoma cells. *J. Biol. Chem.* **1989**, *264*, 6371–6375.
- Applegate, L.A.; Noel, A.; Vile, G.; Frenk, E.; Tyrrell, R.M. Two genes contribute to different extents to the heme oxygenase enzyme activity measured in cultured human skin fibroblasts and keratinocytes: Implications for protection against oxidant stress. *Photochem. Photobiol.* 1995, *61*, 285–291.
- 233. Cisowski, J.; Loboda, A.; Jozkowicz, A.; Chen, S.; Agarwal, A.; Dulak, J. Role of heme oxygenase-1 in hydrogen peroxide-induced VEGF synthesis: Effect of HO-1 knockout. *Biochem. Biophys. Res. Commun.* 2005, 326, 670–676.
- 234. Tosaki, A.; Das, D.K. The role of heme oxygenase signaling in various disorders. *Mol. Cell. Biochem.* **2002**, *232*, 149–157.

- 235. Tyrrell, R.M. Solar ultraviolet A radiation: An oxidizing skin carcinogen that activates heme oxygenase-1. *Antioxid. Redox Signal.* **2004**, *6*, 835–840.
- 236. Wagener, F.A.; Eggert, A.; Boerman, O.C.; Oyen, W.J.; Verhofstad, A.; Abraham, N.G.; Adema, G.; van Kooyk, Y.; de Witte, T.; Figdor, C.G. Heme is a potent inducer of inflammation in mice and is counteracted by heme oxygenase. *Blood* 2001, *98*, 1802–1811.
- 237. Wagener, F.A.; Dankers, A.C.; van Summeren, F.; Scharstuhl, A.; van den Heuvel, J.J.; Koenderink, J.B.; Pennings, S.W.; Russel, F.G.; Masereeuw, R. Heme oxygenase-1 and breast cancer resistance protein protect against heme-induced toxicity. *Curr. Pharm. Des.* 2013, 19, 2698–2707.
- 238. Hayashi, S.; Takamiya, R.; Yamaguchi, T.; Matsumoto, K.; Tojo, S.J.; Tamatani, T.; Kitajima, M.; Makino, N.; Ishimura, Y.; Suematsu, M. Induction of heme oxygenase-1 suppresses venular leukocyte adhesion elicited by oxidative stress: Role of bilirubin generated by the enzyme. *Circ. Res.* **1999**, *85*, 663–671.
- 239. Vachharajani, T.J.; Work, J.; Issekutz, A.C.; Granger, D.N. Heme oxygenase modulates selectin expression in different regional vascular beds. *Am. J. Physiol.* **2000**, *278*, H1613–H1617.
- 240. Takahashi, T.; Shimizu, H.; Morimatsu, H.; Inoue, K.; Akagi, R.; Morita, K.; Sassa, S. Heme oxygenase-1: A fundamental guardian against oxidative tissue injuries in acute inflammation. *Mini Rev. Med. Chem.* 2007, 7, 745–753.
- 241. Takahashi, T.; Shimizu, H.; Morimatsu, H.; Maeshima, K.; Inoue, K.; Akagi, R.; Matsumi, M.; Katayama, H.; Morita, K. Heme oxygenase-1 is an essential cytoprotective component in oxidative tissue injury induced by hemorrhagic shock. J. Clin. Biochem. Nutr. 2009, 44, 28–40.
- 242. Baranano, D.E.; Rao, M.; Ferris, C.D.; Snyder, S.H. Biliverdin reductase: A major physiologic cytoprotectant. *Proc. Natl. Acad. Sci. USA* **2002**, *99*, 16093–16098.
- 243. Stocker, R.; Yamamoto, Y.; McDonagh, A.F.; Glazer, A.N.; Ames, B.N. Bilirubin is an antioxidant of possible physiological importance. *Science* **1987**, *235*, 1043–1046.
- 244. Kim, S.Y.; Park, S.C. Physiological antioxidative network of the bilirubin system in aging and age-related diseases. *Front. Pharmacol.* **2012**, *3*, 45.
- 245. Dore, S.; Snyder, S.H. Neuroprotective action of bilirubin against oxidative stress in primary hippocampal cultures. *Ann. N. Y. Acad. Sci.* **1999**, *890*, 167–172.
- 246. Dore, S.; Takahashi, M.; Ferris, C.D.; Zakhary, R.; Hester, L.D.; Guastella, D.; Snyder, S.H. Bilirubin, formed by activation of heme oxygenase-2, protects neurons against oxidative stress injury. *Proc. Natl. Acad. Sci. USA* **1999**, *96*, 2445–2450.
- 247. Lanone, S.; Bloc, S.; Foresti, R.; Almolki, A.; Taille, C.; Callebert, J.; Conti, M.; Goven, D.; Aubier, M.; Dureuil, B.; *et al.* Bilirubin decreases nos2 expression via inhibition of NAD(P)H oxidase: Implications for protection against endotoxic shock in rats. *FASEB J.* **2005**, *19*, 1890–1892.
- 248. Jiang, F.; Roberts, S.J.; Datla, S.; Dusting, G.J. NO modulates NADPH oxidase function via heme oxygenase-1 in human endothelial cells. *Hypertension* **2006**, *48*, 950–957.
- 249. Matsumoto, H.; Ishikawa, K.; Itabe, H.; Maruyama, Y. Carbon monoxide and bilirubin from heme oxygenase-1 suppresses reactive oxygen species generation and plasminogen activator inhibitor-1 induction. *Mol. Cell. Biochem.* 2006, 291, 21–28.

- 250. McDonagh, A.F. The biliverdin-bilirubin antioxidant cycle of cellular protection: Missing a wheel? *Free Radic. Biol. Med.* **2010**, *49*, 814–820.
- 251. Jomova, K.; Valko, M. Importance of iron chelation in free radical-induced oxidative stress and human disease. *Curr. Pharm. Des.* **2011**, *17*, 3460–3473.
- 252. Harrison, P.M.; Arosio, P. The ferritins: Molecular properties, iron storage function and cellular regulation. *Biochim. Biophys. Acta* **1996**, *1275*, 161–203.
- 253. Torti, F.M.; Torti, S.V. Regulation of ferritin genes and protein. Blood 2002, 99, 3505-3516.
- 254. Orino, K.; Watanabe, K. Molecular, physiological and clinical aspects of the iron storage protein ferritin. *Vet. J.* **2008**, *178*, 191–201.
- 255. Villacorta, L.; Azzi, A.; Zingg, J.M. Regulatory role of vitamins E and C on extracellular matrix components of the vascular system. *Mol. Asp. Med.* **2007**, *28*, 507–537.
- 256. Fan, J.; Frey, R.S.; Rahman, A.; Malik, A.B. Role of neutrophil NADPH oxidase in the mechanism of tumor necrosis factor-alpha -induced NF-kappa B activation and intercellular adhesion molecule-1 expression in endothelial cells. *J. Biol. Chem.* **2002**, *277*, 3404–3411.
- 257. Kono, H.; Rusyn, I.; Yin, M.; Gabele, E.; Yamashina, S.; Dikalova, A.; Kadiiska, M.B.; Connor, H.D.; Mason, R.P.; Segal, B.H.; *et al.* NADPH oxidase-derived free radicals are key oxidants in alcohol-induced liver disease. *J. Clin. Investig.* **2000**, *106*, 867–872.
- 258. Sadikot, R.T.; Zeng, H.; Yull, F.E.; Li, B.; Cheng, D.S.; Kernodle, D.S.; Jansen, E.D.; Contag, C.H.; Segal, B.H.; Holland, S.M.; *et al.* p47phox deficiency impairs NF-kappa B activation and host defense in Pseudomonas pneumonia. *J. Immunol.* 2004, *172*, 1801–1808.
- 259. Sato, K.; Kadiiska, M.B.; Ghio, A.J.; Corbett, J.; Fann, Y.C.; Holland, S.M.; Thurman, R.G.; Mason, R.P. *In vivo* lipid-derived free radical formation by NADPH oxidase in acute lung injury induced by lipopolysaccharide: A model for ARDS. *FASEB J.* 2002, *16*, 1713–1720.
- Rahman, A.; Kefer, J.; Bando, M.; Niles, W.D.; Malik, A.B. E-selectin expression in human endothelial cells by TNF-alpha-induced oxidant generation and NF-kappaB activation. *Am. J. Physiol.* 1998, 275, L533–L544.
- 261. Li, J.M.; Fan, L.M.; Christie, M.R.; Shah, A.M. Acute tumor necrosis factor alpha signaling via NADPH oxidase in microvascular endothelial cells: Role of p47phox phosphorylation and binding to TRAF4. *Mol. Cell. Biol.* 2005, *25*, 2320–2330.
- 262. Gu, Y.; Xu, Y.C.; Wu, R.F.; Souza, R.F.; Nwariaku, F.E.; Terada, L.S. TNFalpha activates c-Jun amino terminal kinase through p47(phox). *Exp. Cell Res.* **2002**, *272*, 62–74.
- 263. Lambeth, J.D.; Krause, K.H.; Clark, R.A. NOX enzymes as novel targets for drug development. *Semin. Immunopathol.* **2008**, *30*, 339–363.
- Jaquet, V.; Scapozza, L.; Clark, R.A.; Krause, K.H.; Lambeth, J.D. Small-molecule NOX inhibitors: ROS-generating NADPH oxidases as therapeutic targets. *Antioxid. Redox Signal.* 2009, 11, 2535–2552.
- Zhang, W.J.; Wei, H.; Frei, B. Genetic deficiency of NADPH oxidase does not diminish, but rather enhances, LPS-induced acute inflammatory responses *in vivo*. *Free Radic. Biol. Med.* 2009, *46*, 791–798.

- Marriott, H.M.; Jackson, L.E.; Wilkinson, T.S.; Simpson, A.J.; Mitchell, T.J.; Buttle, D.J.; Cross, S.S.; Ince, P.G.; Hellewell, P.G.; Whyte, M.K.; *et al.* Reactive oxygen species regulate neutrophil recruitment and survival in pneumococcal pneumonia. *Am. J. Respir. Crit. Care Med.* 2008, 177, 887–895.
- 267. Liochev, S.I.; Fridovich, I. The role of O2.- in the production of HO.: In vitro and in vivo. Free Radic. Biol. Med. 1994, 16, 29-33.
- 268. Kakhlon, O.; Cabantchik, Z.I. The labile iron pool: Characterization, measurement, and participation in cellular processes(1). *Free Radic. Biol. Med.* **2002**, *33*, 1037–1046.
- 269. Fleming, R.E.; Ponka, P. Iron overload in human disease. N. Engl. J. Med. 2012, 366, 348-359.
- 270. Larsen, R.; Gouveia, Z.; Soares, M.P.; Gozzelino, R. Heme cytotoxicity and the pathogenesis of immune-mediated inflammatory diseases. *Front. Pharmacol.* **2012**, *3*, 77.
- 271. Kell, D.B. Towards a unifying, systems biology understanding of large-scale cellular death and destruction caused by poorly liganded iron: Parkinson's, Huntington's, Alzheimer's, prions, bactericides, chemical toxicology and others as examples. *Arch. Toxicol.* **2010**, *84*, 825–889.
- 272. Perron, N.R.; Brumaghim, J.L. A review of the antioxidant mechanisms of polyphenol compounds related to iron binding. *Cell Biochem. Biophys.* **2009**, *53*, 75–100.
- 273. Sestili, P.; Guidarelli, A.; Dacha, M.; Cantoni, O. Quercetin prevents DNA single strand breakage and cytotoxicity caused by tert-butylhydroperoxide: Free radical scavenging *versus* iron chelating mechanism. *Free Radic. Biol. Med.* **1998**, *25*, 196–200.
- 274. Fukai, T.; Ushio-Fukai, M. Superoxide dismutases: Role in redox signaling, vascular function, and diseases. *Antioxid. Redox Signal.* **2011**, *15*, 1583–1606.
- 275. Marklund, S.L. Extracellular superoxide dismutase and other superoxide dismutase isoenzymes in tissues from nine mammalian species. *Biochem. J.* **1984**, *222*, 649–655.
- 276. Moysan, A.; Marquis, I.; Gaboriau, F.; Santus, R.; Dubertret, L.; Morliere, P. Ultraviolet A-induced lipid peroxidation and antioxidant defense systems in cultured human skin fibroblasts. *J. Investig. Dermatol.* **1993**, *100*, 692–698.
- 277. Poswig, A.; Wenk, J.; Brenneisen, P.; Wlaschek, M.; Hommel, C.; Quel, G.; Faisst, K.; Dissemond, J.; Briviba, K.; Krieg, T.; *et al.* Adaptive antioxidant response of manganese-superoxide dismutase following repetitive UVA irradiation. *J. Investig. Dermatol.* **1999**, *112*, 13–18.
- 278. Choung, B.Y.; Byun, S.J.; Suh, J.G.; Kim, T.Y. Extracellular superoxide dismutase tissue distribution and the patterns of superoxide dismutase mRNA expression following ultraviolet irradiation on mouse skin. *Exp. Dermatol.* **2004**, *13*, 691–699.
- Delanian, S.; Baillet, F.; Huart, J.; Lefaix, J.L.; Maulard, C.; Housset, M. Successful treatment of radiation-induced fibrosis using liposomal Cu/Zn superoxide dismutase: Clinical trial. *Radiother*. *Oncol.* 1994, *32*, 12–20.
- 280. Lefaix, J.L.; Delanian, S.; Leplat, J.J.; Tricaud, Y.; Martin, M.; Nimrod, A.; Baillet, F.; Daburon, F. Successful treatment of radiation-induced fibrosis using Cu/Zn-SOD and Mn-SOD: An experimental study. *Int. J. Radiat. Oncol. Biol. Phys.* **1996**, *35*, 305–312.
- 281. Luo, J.D.; Wang, Y.Y.; Fu, W.L.; Wu, J.; Chen, A.F. Gene therapy of endothelial nitric oxide synthase and manganese superoxide dismutase restores delayed wound healing in type 1 diabetic mice. *Circulation* 2004, *110*, 2484–2493.

- 282. Ceradini, D.J.; Yao, D.; Grogan, R.H.; Callaghan, M.J.; Edelstein, D.; Brownlee, M.; Gurtner, G.C. Decreasing intracellular superoxide corrects defective ischemia-induced new vessel formation in diabetic mice. J. Biol. Chem. 2008, 283, 10930–10938.
- 283. Ha, H.Y.; Kim, Y.; Ryoo, Z.Y.; Kim, T.Y. Inhibition of the TPA-induced cutaneous inflammation and hyperplasia by EC-SOD. *Biochem. Biophys. Res. Commun.* **2006**, *348*, 450–458.
- 284. Kim, Y.; Kim, B.H.; Lee, H.; Jeon, B.; Lee, Y.S.; Kwon, M.J.; Kim, T.Y. Regulation of skin inflammation and angiogenesis by EC-SOD via HIF-1alpha and NF-kappaB pathways. *Free Radic. Biol. Med.* 2011, 51, 1985–1995.
- 285. Lee, Y.S.; Cheon, I.S.; Kim, B.H.; Kwon, M.J.; Lee, H.W.; Kim, T.Y. Loss of extracellular superoxide dismutase induces severe IL-23-mediated skin inflammation in mice. *J. Investig. Dermatol.* 2013, 133, 732–741.
- 286. Kwon, M.J.; Jeon, Y.J.; Lee, K.Y.; Kim, T.Y. Superoxide dismutase 3 controls adaptive immune responses and contributes to the inhibition of ovalbumin-induced allergic airway inflammation in mice. *Antioxid. Redox Signal.* 2012, 17, 1376–1392.
- 287. Nozik-Grayck, E.; Suliman, H.B.; Piantadosi, C.A. Extracellular superoxide dismutase. *Int. J. Biochem. Cell Biol.* 2005, *37*, 2466–2471.
- 288. Qin, Z.; Reszka, K.J.; Fukai, T.; Weintraub, N.L. Extracellular superoxide dismutase (ecSOD) in vascular biology: An update on exogenous gene transfer and endogenous regulators of ecSOD. *Transl. Res.* 2008, 151, 68–78.
- Carlsson, L.M.; Marklund, S.L.; Edlund, T. The rat extracellular superoxide dismutase dimer is converted to a tetramer by the exchange of a single amino acid. *Proc. Natl. Acad. Sci. USA* 1996, 93, 5219–5222.
- 290. Yamaguchi, M.; Zhou, C.; Heistad, D.D.; Watanabe, Y.; Zhang, J.H. Gene transfer of extracellular superoxide dismutase failed to prevent cerebral vasospasm after experimental subarachnoid hemorrhage. *Stroke* 2004, *35*, 2512–2517.
- 291. Batinic-Haberle, I.; Reboucas, J.S.; Spasojevic, I. Superoxide dismutase mimics: Chemistry, pharmacology, and therapeutic potential. *Antioxid. Redox Signal.* **2010**, *13*, 877–918.
- 292. Day, B.J. Catalase and glutathione peroxidase mimics. Biochem. Pharmacol. 2009, 77, 285-296.
- 293. Doctrow, S.R.; Huffman, K.; Marcus, C.B.; Tocco, G.; Malfroy, E.; Adinolfi, C.A.; Kruk, H.; Baker, K.; Lazarowych, N.; Mascarenhas, J.; *et al.* Salen-manganese complexes as catalytic scavengers of hydrogen peroxide and cytoprotective agents: Structure-activity relationship studies. *J. Med. Chem.* **2002**, *45*, 4549–4558.
- 294. Day, B.J. Catalytic antioxidants: A radical approach to new therapeutics. *Drug Discov. Today* **2004**, *9*, 557–566.
- 295. Rong, Y.; Doctrow, S.R.; Tocco, G.; Baudry, M. EUK-134, a synthetic superoxide dismutase and catalase mimetic, prevents oxidative stress and attenuates kainate-induced neuropathology. *Proc. Natl. Acad. Sci. USA* **1999**, *96*, 9897–9902.
- 296. Liu, R.; Liu, I.Y.; Bi, X.; Thompson, R.F.; Doctrow, S.R.; Malfroy, B.; Baudry, M. Reversal of age-related learning deficits and brain oxidative stress in mice with superoxide dismutase/catalase mimetics. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 8526–8531.

- 297. Zhang, H.J.; Doctrow, S.R.; Xu, L.; Oberley, L.W.; Beecher, B.; Morrison, J.; Oberley, T.D.; Kregel, K.C. Redox modulation of the liver with chronic antioxidant enzyme mimetic treatment prevents age-related oxidative damage associated with environmental stress. *FASEB J.* 2004, *18*, 1547–1549.
- 298. Clausen, A.; Doctrow, S.; Baudry, M. Prevention of cognitive deficits and brain oxidative stress with superoxide dismutase/catalase mimetics in aged mice. *Neurobiol. Aging* **2010**, *31*, 425–433.
- 299. Liesa, M.; Luptak, I.; Qin, F.; Hyde, B.B.; Sahin, E.; Siwik, D.A.; Zhu, Z.; Pimentel, D.R.; Xu, X.J.; Ruderman, N.B.; *et al.* Mitochondrial transporter ATP binding cassette mitochondrial erythroid is a novel gene required for cardiac recovery after ischemia/reperfusion. *Circulation* 2011, *124*, 806–813.
- 300. Mahmood, J.; Jelveh, S.; Calveley, V.; Zaidi, A.; Doctrow, S.R.; Hill, R.P. Mitigation of radiation-induced lung injury by genistein and EUK-207. *Int. J. Radiat. Biol.* **2011**, *87*, 889–901.
- 301. Doctrow, S.R.; Lopez, A.; Schock, A.M.; Duncan, N.E.; Jourdan, M.M.; Olasz, E.B.; Moulder, J.E.; Fish, B.L.; Mader, M.; Lazar, J.; *et al.* A synthetic superoxide dismutase/catalase mimetic EUK-207 mitigates radiation dermatitis and promotes wound healing in irradiated rat skin. *J. Investig. Dermatol.* **2013**, *133*, 1088–1096.
- Soares, M.P.; Bach, F.H. Heme oxygenase-1: From biology to therapeutic potential. *Trends Mol. Med.* 2009, 15, 50–58.
- 303. Saikawa, Y.; Kaneda, H.; Yue, L.; Shimura, S.; Toma, T.; Kasahara, Y.; Yachie, A.; Koizumi, S. Structural evidence of genomic exon-deletion mediated by Alu-Alu recombination in a human case with heme oxygenase-1 deficiency. *Hum. Mutat.* 2000, *16*, 178–179.
- 304. Radhakrishnan, N.; Yadav, S.P.; Sachdeva, A.; Pruthi, P.K.; Sawhney, S.; Piplani, T.; Wada, T.; Yachie, A. Human heme oxygenase-1 deficiency presenting with hemolysis, nephritis, and asplenia. J. Pediatr. Hematol./Oncol. 2011, 33, 74–78.
- 305. Exner, M.; Minar, E.; Wagner, O.; Schillinger, M. The role of heme oxygenase-1 promoter polymorphisms in human disease. *Free Radic. Biol. Med.* **2004**, *37*, 1097–1104.
- 306. Kappas, A.; Drummond, G.S. Control of heme and cytochrome P-450 metabolism by inorganic metals, organometals and synthetic metalloporphyrins. *Environ. Health Perspect.* **1984**, *57*, 301–306.
- 307. Sasaki, T.; Takahashi, T.; Maeshima, K.; Shimizu, H.; Toda, Y.; Morimatsu, H.; Takeuchi, M.; Yokoyama, M.; Akagi, R.; Morita, K. Heme arginate pretreatment attenuates pulmonary NF-kappaB and AP-1 activation induced by hemorrhagic shock via heme oxygenase-1 induction. *Med. Chem.* 2006, 2, 271–274.
- 308. Tenhunen, R.; Mustajoki, P. Acute porphyria: Treatment with heme. *Semin. Liver Dis.* **1998**, *18*, 53–55.
- 309. Barbagallo, I.; Galvano, F.; Frigiola, A.; Cappello, F.; Riccioni, G.; Murabito, P.; D'Orazio, N.; Torella, M.; Gazzolo, D.; Li Volti, G. Potential therapeutic effects of natural heme oxygenase-1 inducers in cardiovascular diseases. *Antioxid. Redox Signal.* 2013, 18, 507–521.
- 310. Motterlini, R.; Foresti, R.; Bassi, R.; Green, C.J. Curcumin, an antioxidant and anti-inflammatory agent, induces heme oxygenase-1 and protects endothelial cells against oxidative stress. *Free Radic. Biol. Med.* 2000, 28, 1303–1312.

- 311. Balogun, E.; Hoque, M.; Gong, P.; Killeen, E.; Green, C.J.; Foresti, R.; Alam, J.; Motterlini, R. Curcumin activates the haem oxygenase-1 gene via regulation of Nrf2 and the antioxidant-responsive element. *Biochem. J.* 2003, 371, 887–895.
- 312. McNally, S.J.; Harrison, E.M.; Ross, J.A.; Garden, O.J.; Wigmore, S.J. Curcumin induces heme oxygenase 1 through generation of reactive oxygen species, p38 activation and phosphatase inhibition. *Int. J. Mol. Med.* 2007, 19, 165–172.
- 313. Lin, H.C.; Cheng, T.H.; Chen, Y.C.; Juan, S.H. Mechanism of heme oxygenase-1 gene induction by quercetin in rat aortic smooth muscle cells. *Pharmacology* **2004**, *71*, 107–112.
- 314. Yao, P.; Nussler, A.; Liu, L.; Hao, L.; Song, F.; Schirmeier, A.; Nussler, N. Quercetin protects human hepatocytes from ethanol-derived oxidative stress by inducing heme oxygenase-1 via the MAPK/Nrf2 pathways. J. Hepatol. 2007, 47, 253–261.
- 315. Wannamethee, S.G.; Lowe, G.D.; Rumley, A.; Bruckdorfer, K.R.; Whincup, P.H. Associations of vitamin C status, fruit and vegetable intakes, and markers of inflammation and hemostasis. *Am. J. Clin. Nutr.* 2006, *83*, 567–574; quiz 726–567.
- 316. Homma, S.; Azuma, A.; Taniguchi, H.; Ogura, T.; Mochiduki, Y.; Sugiyama, Y.; Nakata, K.; Yoshimura, K.; Takeuchi, M.; Kudoh, S.; *et al.* Efficacy of inhaled *N*-acetylcysteine monotherapy in patients with early stage idiopathic pulmonary fibrosis. *Respirology* **2012**, *17*, 467–477.
- 317. Demedts, M.; Behr, J.; Buhl, R.; Costabel, U.; Dekhuijzen, R.; Jansen, H.M.; MacNee, W.; Thomeer, M.; Wallaert, B.; Laurent, F.; *et al.* High-dose acetylcysteine in idiopathic pulmonary fibrosis. *N. Engl. J. Med.* **2005**, *353*, 2229–2242.
- 318. Tomioka, H.; Kuwata, Y.; Imanaka, K.; Hashimoto, K.; Ohnishi, H.; Tada, K.; Sakamoto, H.; Iwasaki, H. A pilot study of aerosolized *N*-acetylcysteine for idiopathic pulmonary fibrosis. *Respirology* 2005, 10, 449–455.
- 319. Behr, J.; Demedts, M.; Buhl, R.; Costabel, U.; Dekhuijzen, R.P.; Jansen, H.M.; MacNee, W.; Thomeer, M.; Wallaert, B.; Laurent, F.; *et al.* Lung function in idiopathic pulmonary fibrosis—Extended analyses of the IFIGENIA trial. *Respir. Res.* 2009, 10, 101.
- 320. Crestani, B.; Besnard, V.; Boczkowski, J. Signalling pathways from NADPH oxidase-4 to idiopathic pulmonary fibrosis. *Int. J. Biochem. Cell Biol.* **2011**, *43*, 1086–1089.
- 321. Atkuri, K.R.; Mantovani, J.J.; Herzenberg, L.A.; Herzenberg, L.A. *N*-Acetylcysteine—A safe antidote for cysteine/glutathione deficiency. *Curr. Opin. Pharmacol.* **2007**, *7*, 355–359.
- 322. Idiopathic Pulmonary Fibrosis Clinical Research, N.; Raghu, G.; Anstrom, K.J.; King, T.E., Jr.; Lasky, J.A.; Martinez, F.J. Prednisone, azathioprine, and *N*-acetylcysteine for pulmonary fibrosis. *N. Engl. J. Med.* 2012, *366*, 1968–1977.
- 323. Schagen, S.K.; Zampeli, V.A.; Makrantonaki, E.; Zouboulis, C.C. Discovering the link between nutrition and skin aging. *Dermato-Endocrinol.* **2012**, *4*, 298–307.
- 324. Placzek, M.; Gaube, S.; Kerkmann, U.; Gilbertz, K.P.; Herzinger, T.; Haen, E.; Przybilla, B. Ultraviolet B-induced DNA damage in human epidermis is modified by the antioxidants ascorbic acid and D-alpha-tocopherol. *J. Investig. Dermatol.* 2005, *124*, 304–307.
- 325. Morganti, P.; Bruno, C.; Guarneri, F.; Cardillo, A.; Del Ciotto, P.; Valenzano, F. Role of topical and nutritional supplement to modify the oxidative stress. *Int. J. Cosmet. Sci.* **2002**, *24*, 331–339.
- 326. Eberlein-Konig, B.; Ring, J. Relevance of vitamins C and E in cutaneous photoprotection. *J. Cosmet. Dermatol.* **2005**, *4*, 4–9.

- 327. Stahl, W.; Heinrich, U.; Jungmann, H.; Sies, H.; Tronnier, H. Carotenoids and carotenoids plus vitamin E protect against ultraviolet light-induced erythema in humans. Am. J. Clin. Nutr. 2000, 71, 795–798.
- 328. Lee, J.; Jiang, S.; Levine, N.; Watson, R.R. Carotenoid supplementation reduces erythema in human skin after simulated solar radiation exposure. *Proc. Soc. Exp. Biol. Med.* **2000**, *223*, 170–174.
- 329. Heinrich, U.; Gartner, C.; Wiebusch, M.; Eichler, O.; Sies, H.; Tronnier, H.; Stahl, W. Supplementation with beta-carotene or a similar amount of mixed carotenoids protects humans from UV-induced erythema. *J. Nutr.* **2003**, *133*, 98–101.
- 330. Telfer, A.; Dhami, S.; Bishop, S.M.; Phillips, D.; Barber, J. beta-Carotene quenches singlet oxygen formed by isolated photosystem II reaction centers. *Biochemistry* **1994**, *33*, 14469–14474.
- 331. Bose, B.; Chatterjee, S.N. UVA-induced peroxidation of lipid in the dried film state. *J. Photochem. Photobiol. B* 1994, 23, 119–123.
- 332. Eicker, J.; Kurten, V.; Wild, S.; Riss, G.; Goralczyk, R.; Krutmann, J.; Berneburg, M. Betacarotene supplementation protects from photoaging-associated mitochondrial DNA mutation. *Photochem. Photobiol. Sci.* 2003, *2*, 655–659.
- 333. Manach, C.; Scalbert, A.; Morand, C.; Remesy, C.; Jimenez, L. Polyphenols: Food sources and bioavailability. *Am. J. Clin. Nutr.* **2004**, *79*, 727–747.
- 334. Nichols, J.A.; Katiyar, S.K. Skin photoprotection by natural polyphenols: Anti-inflammatory, antioxidant and DNA repair mechanisms. *Arch. Dermatol. Res.* **2010**, *302*, 71–83.
- 335. Agarwal, R.; Katiyar, S.K.; Khan, S.G.; Mukhtar, H. Protection against ultraviolet B radiation-induced effects in the skin of SKH-1 hairless mice by a polyphenolic fraction isolated from green tea. *Photochem. Photobiol.* **1993**, *58*, 695–700.
- 336. Katiyar, S.K.; Elmets, C.A.; Agarwal, R.; Mukhtar, H. Protection against ultraviolet-B radiation-induced local and systemic suppression of contact hypersensitivity and edema responses in C3H/HeN mice by green tea polyphenols. *Photochem. Photobiol.* **1995**, *62*, 855–861.
- 337. Elmets, C.A.; Singh, D.; Tubesing, K.; Matsui, M.; Katiyar, S.; Mukhtar, H. Cutaneous photoprotection from ultraviolet injury by green tea polyphenols. J. Am. Acad. Dermatol. 2001, 44, 425–432.
- 338. Katiyar, S.K.; Elmets, C.A. Green tea polyphenolic antioxidants and skin photoprotection (Review). *Int. J. Oncol.* 2001, *18*, 1307–1313.
- 339. Van Laethem, A.; Claerhout, S.; Garmyn, M.; Agostinis, P. The sunburn cell: Regulation of death and survival of the keratinocyte. *Int. J. Biochem. Cell Biol.* **2005**, *37*, 1547–1553.
- 340. Lambert, J.D.; Hong, J.; Yang, G.Y.; Liao, J.; Yang, C.S. Inhibition of carcinogenesis by polyphenols: Evidence from laboratory investigations. *Am. J. Clin. Nutr.* **2005**, *81*, 284S–291S.
- 341. Shindo, Y.; Witt, E.; Han, D.; Epstein, W.; Packer, L. Enzymic and non-enzymic antioxidants in epidermis and dermis of human skin. *J. Investig. Dermatol.* **1994**, *102*, 122–124.
- 342. Kwong, L.K.; Kamzalov, S.; Rebrin, I.; Bayne, A.C.; Jana, C.K.; Morris, P.; Forster, M.J.; Sohal, R.S. Effects of coenzyme Q(10) administration on its tissue concentrations, mitochondrial oxidant generation, and oxidative stress in the rat. *Free Radic. Biol. Med.* 2002, *33*, 627–638.

- 343. Sohal, R.S.; Kamzalov, S.; Sumien, N.; Ferguson, M.; Rebrin, I.; Heinrich, K.R.; Forster, M.J. Effect of coenzyme Q10 intake on endogenous coenzyme Q content, mitochondrial electron
- transport chain, antioxidative defenses, and life span of mice. *Free Radic. Biol. Med.* 2006, 40, 480–487.
 344. Prahl, S.; Kueper, T.; Biernoth, T.; Wohrmann, Y.; Munster, A.; Furstenau, M.; Schmidt, M.;
- Schulze, C.; Wittern, K.P.; Wenck, H.; *et al.* Aging skin is functionally anaerobic: Importance of coenzyme Q10 for anti aging skin care. *BioFactors* **2008**, *32*, 245–255.
- 345. Blatt, T.; Lenz, H.; Koop, U.; Jaspers, S.; Weber, T.; Mummert, C.; Wittern, K.P.; Stab, F.; Wenck, H. Stimulation of skin's energy metabolism provides multiple benefits for mature human skin. *BioFactors* 2005, 25, 179–185.
- 346. Blatt, T.; Littarru, G.P. Biochemical rationale and experimental data on the antiaging properties of CoQ(10) at skin level. *BioFactors* **2011**, *37*, 381–385.
- Makrantonaki, E.; Zouboulis, C.C. Skin alterations and diseases in advanced age. *Drug Discov. Today Dis. Mech.* 2008, 5, 153–162.
- 348. Han, R.; Liu, L.; Li, J.; Du, G.; Chen, J. Functions, applications and production of 2-O-D-glucopyranosyl-L-ascorbic acid. *Appl. Microbiol. Biotech.* **2012**, *95*, 313–320.
- 349. Pawar, R.S.; Tamta, H.; Ma, J.; Krynitsky, A.J.; Grundel, E.; Wamer, W.G.; Rader, J.I. Updates on chemical and biological research on botanical ingredients in dietary supplements. *Anal. Bioanal. Chem.* 2013, doi:10.1007/s00216-012-6691-2.
- 350. Ristow, M.; Zarse, K.; Oberbach, A.; Kloting, N.; Birringer, M.; Kiehntopf, M.; Stumvoll, M.; Kahn, C.R.; Bluher, M. Antioxidants prevent health-promoting effects of physical exercise in humans. *Proc. Natl. Acad. Sci. USA* 2009, *106*, 8665–8670.
- 351. Stanner, S.A.; Hughes, J.; Kelly, C.N.; Buttriss, J. A review of the epidemiological evidence for the 'antioxidant hypothesis'. *Public Health Nutr.* **2004**, *7*, 407–422.
- 352. Marik, P.E.; Flemmer, M. Do dietary supplements have beneficial health effects in industrialized nations: What is the evidence? *J. Parenteral Enteral Nutr.* **2012**, *36*, 159–168.
- 353. Mursu, J.; Robien, K.; Harnack, L.J.; Park, K.; Jacobs, D.R., Jr. Dietary supplements and mortality rate in older women: The Iowa Women's Health Study. Arch. Intern. Med. 2011, 171, 1625–1633.
- Sedlak, T.W.; Saleh, M.; Higginson, D.S.; Paul, B.D.; Juluri, K.R.; Snyder, S.H. Bilirubin and glutathione have complementary antioxidant and cytoprotective roles. *Proc. Natl. Acad. Sci. USA* 2009, *106*, 5171–5176.
- 355. Dekker, D.; Dorresteijn, M.J.; Pijnenburg, M.; Heemskerk, S.; Rasing-Hoogveld, A.; Burger, D.M.; Wagener, F.A.; Smits, P. The bilirubin-increasing drug atazanavir improves endothelial function in patients with type 2 diabetes mellitus. *Arterioscler. Thromb. Vasc. Biol.* 2011, 31, 458–463.
- 356. Scheffler, I.E. Mitochondria make a come back. *Advan. Drug Delivery Rev.* 2001, 49, 3–26.
- 357. Davis, R.E.; Williams, M. Mitochondrial function and dysfunction: An update. J. Pharmacol. Exp. Ther. 2012, 342, 598–607.
- 358. Shults, C.W.; Oakes, D.; Kieburtz, K.; Beal, M.F.; Haas, R.; Plumb, S.; Juncos, J.L.; Nutt, J.; Shoulson, I.; Carter, J.; *et al.* Effects of coenzyme Q10 in early Parkinson disease: Evidence of slowing of the functional decline. *Arch. Neurol.* 2002, *59*, 1541–1550.

- 359. Shults, C.W.; Flint Beal, M.; Song, D.; Fontaine, D. Pilot trial of high dosages of coenzyme Q10 in patients with Parkinson's disease. *Exp. Neurol.* **2004**, *188*, 491–494.
- 360. Ernster, L.; Dallner, G. Biochemical, physiological and medical aspects of ubiquinone function. *Biochim. Biophys.* **1995**, *1271*, 195–204.
- 361. Zhang, Y.; Aberg, F.; Appelkvist, E.L.; Dallner, G.; Ernster, L. Uptake of dietary coenzyme Q supplement is limited in rats. *J. Nutr.* **1995**, *125*, 446–453.
- 362. Matthews, R.T.; Yang, L.; Browne, S.; Baik, M.; Beal, M.F. Coenzyme Q10 administration increases brain mitochondrial concentrations and exerts neuroprotective effects. *Proc. Natl. Acad. Sci. USA* 1998, 95, 8892–8897.
- 363. Cocheme, H.M.; Kelso, G.F.; James, A.M.; Ross, M.F.; Trnka, J.; Mahendiran, T.; Asin-Cayuela, J.; Blaikie, F.H.; Manas, A.R.; Porteous, C.M.; *et al.* Mitochondrial targeting of quinones: Therapeutic implications. *Mitochondrion* 2007, 7, S94–S102.
- 364. Kelso, G.F.; Porteous, C.M.; Coulter, C.V.; Hughes, G.; Porteous, W.K.; Ledgerwood, E.C.; Smith, R.A.; Murphy, M.P. Selective targeting of a redox-active ubiquinone to mitochondria within cells: Antioxidant and antiapoptotic properties. J. Biol. Chem. 2001, 276, 4588–4596.
- Adlam, V.J.; Harrison, J.C.; Porteous, C.M.; James, A.M.; Smith, R.A.; Murphy, M.P.; Sammut, I.A. Targeting an antioxidant to mitochondria decreases cardiac ischemia-reperfusion injury. *FASEB J.* 2005, 19, 1088–1095.
- 366. Murphy, M.P.; Smith, R.A. Targeting antioxidants to mitochondria by conjugation to lipophilic cations. *Ann. Rev. Pharmacol. Toxicol.* **2007**, *47*, 629–656.
- 367. Skulachev, V.P. A biochemical approach to the problem of aging: "Megaproject" on membrane-penetrating ions. The first results and prospects. *Biochemistry (Mosc.)* 2007, 72, 1385–1396.
- 368. Antonenko, Y.N.; Roginsky, V.A.; Pashkovskaya, A.A.; Rokitskaya, T.I.; Kotova, E.A.; Zaspa, A.A.; Chernyak, B.V.; Skulachev, V.P. Protective effects of mitochondria-targeted antioxidant SkQ in aqueous and lipid membrane environments. *J. Membr. Biol.* 2008, 222, 141–149.
- 369. Distelmaier, F.; Valsecchi, F.; Forkink, M.; van Emst-de Vries, S.; Swarts, H.G.; Rodenburg, R.J.; Verwiel, E.T.; Smeitink, J.A.; Willems, P.H.; Koopman, W.J. Trolox-sensitive reactive oxygen species regulate mitochondrial morphology, oxidative phosphorylation and cytosolic calcium handling in healthy cells. *Antioxid. Redox Signal.* 2012, *17*, 1657–1669.
- 370. Marrache, S.; Dhar, S. Engineering of blended nanoparticle platform for delivery of mitochondria-acting therapeutics. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, 16288–16293.
- 371. Bindu, S.; Pal, C.; Dey, S.; Goyal, M.; Alam, A.; Iqbal, M.S.; Dutta, S.; Sarkar, S.; Kumar, R.; Maity, P.; *et al.* Translocation of heme oxygenase-1 to mitochondria is a novel cytoprotective mechanism against non-steroidal anti-inflammatory drug-induced mitochondrial oxidative stress, apoptosis, and gastric mucosal injury. *J. Biol. Chem.* **2011**, *286*, 39387–39402.

© 2013 by the authors; licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution license (http://creativecommons.org/licenses/by/3.0/).