

Received:
22 May 2018

Revised:
27 June 2018

Accepted:
04 July 2018

Cite this article as:

Flood R, Karteszi H. Incidental thoracic, hepatic and peritoneal calcifications: a case of Pentastomiasis. *BJR Case Rep* 2019; **5**: 20180058.

CASE REPORT

Incidental thoracic, hepatic and peritoneal calcifications: a case of Pentastomiasis

¹RICHARD FLOOD, BMBS and ²HEDVIG KARTESZI, FRCR

¹Department of Radiology, North Bristol NHS Trust, Bristol, UK

²Department of Radiology, University Hospitals Bristol, Bristol, UK

Address correspondence to: Dr Richard Flood
E-mail: richardflood@nhs.net

ABSTRACT

Incidental findings are not uncommon in radiology. In this case, although the incidental findings could be described as an Aunt Minnie, the patient underwent multiple investigations due to the rarity of the causative parasite. The current literature concerning Pentastomiasis suggests it may become more common in future. Our hope is that this case report will help future patients who present with the radiological pattern described to be more rapidly diagnosed and reassured.

CLINICAL PRESENTATION AND IMAGE FINDINGS

We report the case of a 49-year-old female of Nigerian origin who was referred for abdominal ultrasound as part of ongoing occupational health testing. The ultrasound was unremarkable except for multiple echogenic foci throughout both lobes of the liver (Figure 1) which were thought to be calcific. Previous imaging and clinical details were not available so a gastroenterology referral was advised.

Additional clinical information which was provided with a gastroenterology request for repeat ultrasound and a CXR (chest radiograph) included: known hepatitis B (low viral load), reported right upper quadrant discomfort and “nodules on previous CXR 20 years ago”. Repeat ultrasound appearances were unchanged. The CXR showed multiple calcific densities scattered throughout both lungs (Figure 2). Liver elastography was normal. These studies were followed by a triple phase CT liver scan which showed widespread multifocal calcification in the liver, spleen, mesenteric fat, peritoneal surfaces and lung bases. The right lobe of the liver, where the calcifications were focused, was noted to be atrophic and fibrotic. A differential of “previous inflammatory process e.g. TB or parasitic infection” was reported.

These calcifications were left unexplained and the patient underwent annual ultrasound scans for 5 years. No interval changes were seen over this period. Subsequent history obtained during this follow-up period was of a previous serious childhood illness. A pelvic radiograph was obtained following a fall (Figure 3). Multiple appendicular

radiographs over this period showed no evidence of peripheral calcific lesions.

In 2017, repeat liver elastography showed a dramatic increase in kPa and concerns about the hepatic calcifications resurfaced so a biopsy was considered. Prior to this a CT thorax, abdomen and pelvis (including triple phase liver) was requested. Again, the appearances were unchanged (Figures 4 and 5). The reporting radiologist of the second CT recognised the pathognomonic radiographic appearances¹⁻⁵ of previous *Armillifer armillatus* infestation. The clinical team were advised elastography would not be accurate due to the extensive hepatic calcifications and that the patient's hepatic fibrosis was due to the previous infestation.^{6,7}

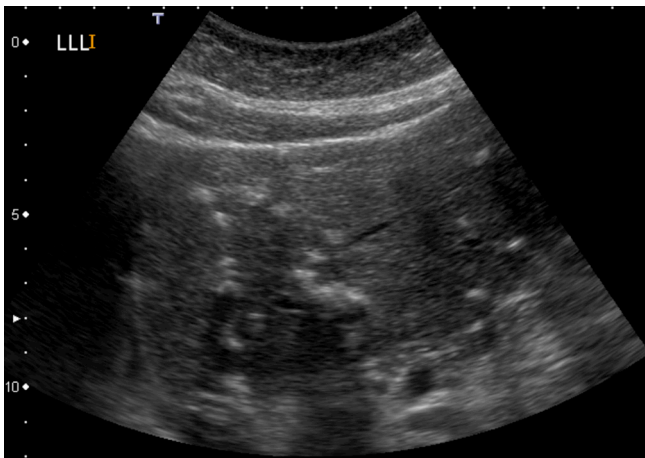
DISCUSSION

The taxonomy of Pentastomida and the terminology concerning the infections they cause is summarised in Table 1. Whilst there are many species of Pentastomida which can infect humans only two species from the *Armilliferidae* family are radiologically relevant.⁴

Armillifer armillatus and *Armillifer moniliformis* are endoparasites which feed on blood. They have a “screw like” appearance and can reach up to 20 cm in length.⁴

Adult *Armillifer* live in the trachea and bronchi of snakes (definitive hosts) such as pythons and vipers.⁴ Female *Armillifer* release eggs into the snake's intestinal and respiratory tracts. When intermediate hosts (usually rodents) come in contact with the sputum/faeces of an infected snake or

Figure 1. Initial ultrasound of the liver identified multiple presumed calcific, echogenic foci throughout both lobes, predominately in the right lobe.



ingest contaminated water, they themselves can become infected. The *Armillifer* eggs hatch in the intestinal tract of their intermediate host and develop into larvae which cross the gut wall, migrate along the peritoneum or pleura and become encysted in various tissues; most commonly liver but also spleen, mesentery, intestine, kidney, omentum, peritoneum and lung.^{1,2,4,5,8-10}

The encysted larvae grow slowly and usually die within 2 years. This leads to an inflammatory response which result in absorption or calcification of the dead larvae.⁴

The life cycle is completed if the intermediate host is ingested by a snake whilst it hosts live larvae. When the live larvae reach

Figure 2. CXR showed multiple c-shaped calcific densities in both lungs. CXR, chest radiograph.

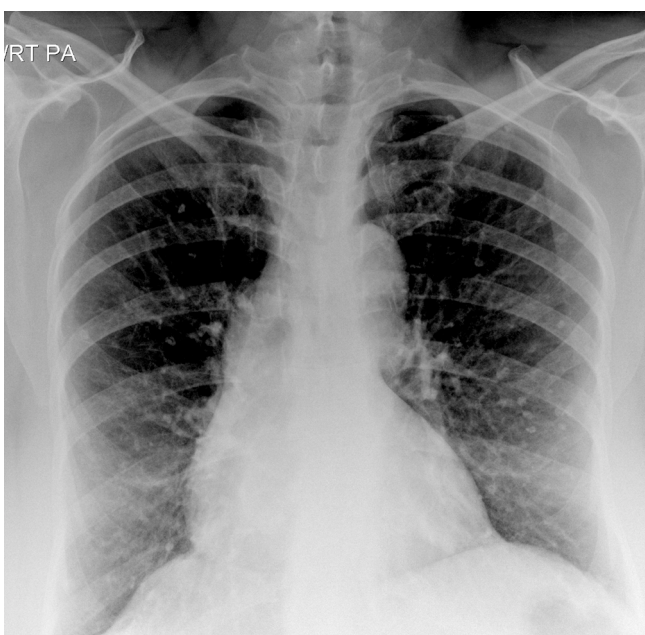


Figure 3. Pelvic radiograph following a fall. Multiple calcific densities throughout the abdominal and peritoneal cavity.



the snake's intestinal tract they migrate to the respiratory tract, develop into adult *Armillifer* and the cycle repeats.

Humans (dead end hosts) are infected by coming into contact with sputum/faeces of an infected snake, contaminated water or under cooked snake meat.^{4,5,9,11,12} It later came to light that the patient described in this case has a close family member who works with bushmeat.

Armillifer armillatus is most commonly seen in West Africa, particularly Nigeria.^{1-5,8-13} *Armillifer moniliformis* is more common in Asia.⁴

Most cases are asymptomatic.¹⁰ In cases which are symptomatic presentation varies according to the tissues affected and can range from fever and abdominal pain^{1,10,11,13} to intestinal obstruction,³ bacterial septicaemia, severe enterocolitis and death.⁸ One

Figure 4. Repeat CT demonstrated unchanged multifocal calcification in the liver and spleen.

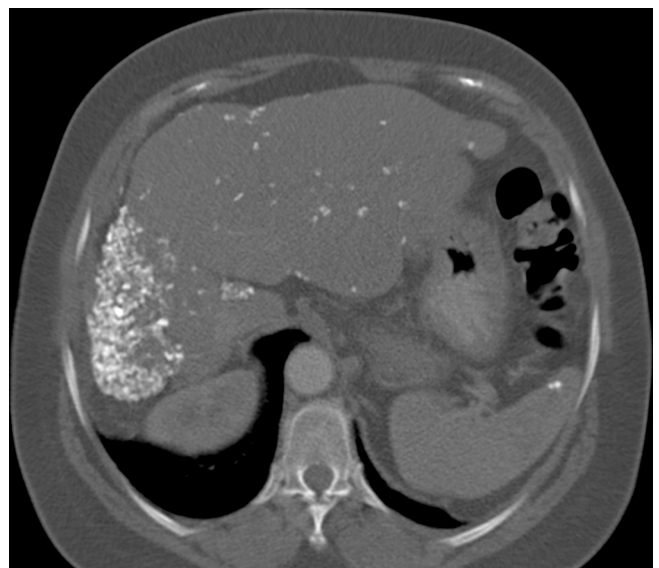


Figure 5. Unchanged calcifications in the peritoneum and mesentery on repeat CT.



study showed *Armillifer armillatus* infection to be the third most common cause of hepatic fibrosis in Nigeria, which was seen in our case and previous cases.^{5,12}

Diagnosis is usually incidental following radiological investigation, autopsy^{8,11} or laparotomy.^{3,4,10} Isolation of live larvae allows histological diagnosis. Serological and PCR tests specific to *Armillifer* are possible but not usually available.^{4,13,14} Patients may have a mild eosinophilia.^{2,4,11}

The radiographic appearances are pathognomonic; multiple crescentic, horseshoe, coiled or comma-shaped calcifications distributed mainly within the upper abdomen and thorax.¹⁻⁵ The size of these calcifications varies from 4 to 8 mm.⁴ Unlike cysticercosis (*Taenia solium* infestation), the musculature is spared in *Armillifer* infestation.⁴ Ultrasound will show multiple hyperechoic lesions in the affected tissues.^{2,5,9,11,13} It is worth noting not all cases will present in this way, a case of a Pentastomiasis granuloma with the radiographic appearances of hepatic malignancy has previously been reported.¹⁵ In this case the diagnosis was only reached after surgical resection had been carried out.¹⁵

Treatment is not required for asymptomatic patients. There is no standard treatment for symptomatic patients but surgery might be indicated in selected cases.^{4,10,13} Praziquantel, albendazole or mebendazole have reportedly been successful at eradicating Pentastomida species.^{2,4,11,13,14,16}

Snake meat is becoming more common at bushmeat markets and consumption is increasing.^{14,16} As migration increases,

Table 1. Taxonomy of Pentastomida and the terminology concerning Pentastomida infestation. *Armillifer armillatus* and *Linguatula serrata* account for > 99% of all human Pentastomida infestations

Subclass	Order	Family	Genus	Species
Pentastomida (pentastomiasis causes pentastomosis)	Porocephalidia (porocephaliasis causes porocephalosis)	Linguatulidae	<i>Linguatula</i> (linguatuliasis causes linguatulosis, Halzoun syndrome or Marrara)	<i>arctica</i>
				<i>serrata</i>
	Armilliferidae	<i>Armillifer</i>	<i>armillatus</i>	
			<i>moniliformis</i>	
<i>grandis</i>				

radiographic evidence of previous *Armillifer armillatus* infestation may become a more common finding in Western countries.

Incidental discovery of the pathognomonic appearances described has in many cases (including ours) led to multiple further investigations before a diagnosis is reached. We hope this case report will help such patients to be more rapidly diagnosed and reassured.

LEARNING POINTS

1. *Armillifer armillatus* is an endoparasite which lives in the respiratory tract of snakes, it is most commonly found in West Africa.
2. Humans can become infected, especially those from West Africa who handle or consume snake meat.
3. It is rarely symptomatic.
4. It has a pathognomonic radiological appearance: multiple crescentic, horseshoe, coiled or comma-shaped calcifications distributed mainly within the upper abdomen and thorax.
5. Treatment is not required for asymptomatic individuals, there is no standard treatment for symptomatic patients.
6. The hunting, selling and consumption of bushmeat is increasing. As migration from Africa increases, the radiographic appearances described may become a more common finding in Western countries.

CONSENT

Written informed consent for the case to be published (including images, case history and data) was obtained from the patient for publication of this case report, including accompanying images.

REFERENCES

1. Rivera O, Yakubu A, Benjamin M. Human pentastomiasis. *West Afr J Radiol* 2016; **23**: 46–8. doi: <https://doi.org/10.4103/1115-3474.162163>
2. Aiyekomogbon JO, Meseko CA, Abiodun OO. *Armillifer armillatus* infestation in Human; public health scenario of a snake parasite: a report of three cases. *Pan Afr Med J* 2016; **25**: 45. doi: <https://doi.org/10.11604/pamj.2016.25.45.10282>

3. Dakubo JC, Etwire VK, Kumoji R, Naaeder SB. Human pentastomiasis: a case report. *West Afr J Med* 2006; **25**: 166–8.
4. Magill AJ, Ryan ET, Hill DR, Solomon T. *Hunter's Tropical Medicine and Emerging Infectious Disease*. 9th ed. London: Saunders Elsevier; 2013.
5. Adeyekun AA, Ukadike I, Adetiloye VA. Severe pentastomiasis Armillifer armillatus infestation complicated by hepatic encephalopathy. *Ann Afr Med* 2011; **10**: 59–62. doi: <https://doi.org/10.4103/1596-3519.76592>
6. Gregory A, Mehrmohammadi M, Denis M, Bayat M, Stan DL, Fatemi M, et al. Effect of calcifications on breast ultrasound shear wave elastography: an investigational study. *PLoS One* 2015; **10**: e0137898. doi: <https://doi.org/10.1371/journal.pone.0137898>
7. Yi L, Qiong W, Yan W, Youben F, Bing H. Correlation between ultrasound elastography and histologic characteristics of papillary thyroid carcinoma. *Sci Rep* 2017; **7**: 45042. doi: <https://doi.org/10.1038/srep45042>
8. Yapo Ette H, Fanton L, Adou Bryn KD, Botti K, Koffi K, Malicier D. Human pentastomiasis discovered postmortem. *Forensic Sci Int* 2003; **137**: 52–4. doi: [https://doi.org/10.1016/S0379-0738\(03\)00281-0](https://doi.org/10.1016/S0379-0738(03)00281-0)
9. Tappe D, Haeupler A, Schäfer H, Racz P, Cramer JP, Poppert S. Armillifer armillatus pentastomiasis in African immigrant, Germany. *Emerg Infect Dis* 2013; **19**: 507–8. doi: <https://doi.org/10.3201/eid1903.121508>
10. Tappe D, Dijkmans AC, Brienen EA, Dijkmans BA, Ruhe IM, Netten MC, et al. Imported Armillifer pentastomiasis: report of a symptomatic infection in The Netherlands and mini-review. *Travel Med Infect Dis* 2014; **12**: 129–33. doi: <https://doi.org/10.1016/j.tmaid.2013.10.011>
11. Lai C, Wang XQ, Lin L, Gao DC, Zhang HX, Zhang YY, et al. Imaging features of pediatric pentastomiasis infection: a case report. *Korean J Radiol* 2010; **11**: 480–4. doi: <https://doi.org/10.3348/kjr.2010.11.4.480>
12. Smith JA, Oladiran B, Lagundoye SB, Lawson EAL, Francis TI. Pentastomiasis and malignancy. *Ann Trop Med Parasitol* 1975; **69**: 503–12. doi: <https://doi.org/10.1080/00034983.1975.11687039>
13. Jisieike-Onuigbo NN, Odenigbo CU, Kalu OA, Eze KC. Armillifer armillatus infection. *Niger J Clin Pract* 2011; **14**: 501–3. doi: <https://doi.org/10.4103/1119-3077.91767>
14. Tappe D, Sulyok M, Riu T, Rózsa L, Bodó I, Schoen C, et al. Co-infections in isceral Pentastomiasis, democratic Republic of the Congo. *Emerg Infect Dis* 2016; **22**: 1333–9. doi: <https://doi.org/10.3201/eid2208.151895>
15. Machado MA, Makdissi FF, Canedo LF, Martino RB, Crescentini F, Chieffi PP, et al. Unusual case of pentastomiasis mimicking liver tumor. *J Gastroenterol Hepatol* 2006; **21**: 1218–20. doi: <https://doi.org/10.1111/j.1440-1746.2006.03203.x>
16. Hardi R, Babocsay G, Tappe D, Sulyok M, Bodó I, Rózsa L. Armillifer-Infected Snakes Sold at Congolese Bushmeat Markets Represent an Emerging Zoonotic Threat. *Ecohealth* 2017; **14**: 743–9. doi: <https://doi.org/10.1007/s10393-017-1274-5>