





## Complete Genome Sequence of a New Strain of *Rice yellow mottle virus* from Malawi, Characterized by a Recombinant VPg Protein

Innocent Ndikumana, a Agnès Pinel-Galzi, b Denis Fargette, b © Eugénie Hébrardb

Research Department, Rwanda Agriculture Board, Kigali, Rwanda<sup>a</sup>; Interactions Plantes Microorganismes Environnement, Institut de Recherche Pour le Développement, Cirad, Université Montpellier, Montpellier, France<sup>b</sup>

**ABSTRACT** The complete sequence of the isolate Mw10 of *Rice yellow mottle virus* was determined. Sequence comparisons revealed 8.4% to 10.7% nucleotide divergence from the published sequences, resulting in the definition of the strain S7. Importantly, a putative recombination event was identified encompassing the viral genome-linked protein involved in host adaptation.

ice yellow mottle virus (RYMV) is endemic in Africa and Madagascar. The virus was detected first in Kenya in 1966 (1) and since then in most rice-growing countries (2). The infection induces symptoms of yellowing and mottling on leaves. The infected plants show a reduction of height and fertility. RYMV causes high losses ranging from 25% to 100% depending on the virus strain, plant genotype, and growth stage of the plants (3, 4). The transmission is mediated mechanically through agricultural practices, insects, or animals (5-7). The host range of RYMV is narrow, restricted to rice and a few wild Poaceae (1). RYMV is a member of the genus Sobemovirus (8), and its genome consists of a single-stranded positive-sense RNA with five open reading frames (9). Based on phylogenetic analyses, RYMV has been classified in six major strains with a strong, spatially based diversity (10). The highest diversity is located in East Africa, the putative center of origin and diversification of RYMV (11). Here, we present the whole-genome sequence of a new strain collected in 2014 from symptomatic rice leaves in paddy fields in the Zomba District, in the south of Malawi. The infection was confirmed after serological detection by a double-antibody sandwich (DAS) enzymelinked immunosorbent assay (ELISA) test as previously described (12). The total RNA was extracted using the RNeasy plant minikit (Qiagen). Specific retrotranscription of the viral genome was performed using the 5' CTCCCCCACCCATCCGAGAATT 3' primer (11). Two overlapping fragments covering the complete RYMV genome were amplified using primers 5' CAATTGAAGCTAGGAAAGGAG 3' and 5' ACTTCGCCGGTTT CGCAGAGGATT 3' for a first fragment and primers 5' CATGCTGGGAAAAGTGTCTG 3' and 5' CTCCCCACCCATCCGAGAATT 3' for a second fragment (11). The sequence of the isolate Mw10 was compared to the 33 published full-length sequences using MEGA6.06 (13). The isolate Mw10 shared only 89.3% to 91.4% nucleotide identity with the other East African sequences. This isolate belongs to a new strain distinct from the lineage S4lm found in Malawi (14) and from the other strains, S4 to S6, found in neighboring Tanzania (15), and has been named strain S7. The sequence of the isolate Mw10 was analyzed using the Recombination Detection Program version 4.0 (RDP4) (16). The strain S7 is mainly related to the strain S6, and shared its characteristic insertion-deletion polymorphisms in the coat protein gene. However, a putative recombinant event was identified in the ORF2a inside the VPg domain from an unknown

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Address correspondence to Eugénie Hébrard, eugenie.hebrard@ird.fr.

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parent. Unexpectedly, the sequence of the central domain of the VPg protein accumulates single polymorphisms found only in resistance-breaking genotypes of RYMV (17, 18). Also surprising, the amino acid at position 49 involved in host adaptation is neither a glutamic acid nor a threonine, but a glutamine. This unique feature is under investigation through experimental and phylogeographic studies, and the recombination breakpoints will be assessed from a larger corpus of full sequences.

**Accession number(s).** This genomic sequence has been deposited in GenBank under the accession no. MF989228.

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## **REFERENCES**

- Bakker W. 1974. Characterisation and ecological aspects of rice yellow mottle virus in Kenya. Agricultural University, Wageningen, the Netherlands.
- Séré Y, Fargette D, Abo ME, Wydra K, Bimerew M, Onasanya A, Akator SK. 2013. Managing the major diseases of rice in Africa, p 213–228. In Wopereis MCS, Johnson DE, Ahmadi N, Tollens E, Jalloh A (ed), Realizing Africa's rice promise. CAB International, Wallingford, United Kingdom.
- Kouassi NK, N'Guessan P, Albar L, Fauquet CM, Brugidou C. 2005. Distribution and characterization of *Rice yellow mottle virus*: a threat to African farmers. Plant Dis 89:124–133. https://doi.org/10.1094/PD-89-0124.
- E. Traoré VS, Néya BJ, Camara M, Gracen V, Offei SK, Traoré O. 2015. Farmers' perception and impact of rice yellow mottle disease on rice yields in Burkina Faso. Agric Sci 6:943–952. https://doi.org/10.4236/as.2015.69091.
- Sarra S, Peters D. 2003. Rice yellow mottle virus is transmitted by cows, donkeys, and grass rats in irrigated rice crops. Plant Dis 87:804–808. https://doi.org/10.1094/PDIS.2003.87.7.804.
- Peters D, Engels C, Sarra S. 2012. Natural spread of plant viruses by birds. J Phytopathol 160:591–594. https://doi.org/10.1111/j.1439-0434 .2012.01937.x.
- Traoré O, Traoré MD, Fargette D, Konaté G. 2006. Rice seedbeds as a source of primary infection by *Rice yellow mottle virus*. Eur J Plant Pathol 115:181–186. https://doi.org/10.1007/s10658-006-9004-9.
- Fargette D, Truve E. 2011. Sobemovirus, p 1185–1190. In King AMQ, Adams MJ, Carstens EB, Lefkowitz EJ (ed), Virus taxonomy. Elsevier, Oxford, United Kingdom.
- Ling R, Pate AE, Carr JP, Firth AE. 2013. An essential fifth coding ORF in the sobemoviruses. Virology 446:397–408. https://doi.org/10.1016/j.virol .2013.05.033.
- Fargette D, Pinel A, Abubakar Z, Traoré O, Brugidou C, Fatogoma S, Hébrard E, Choisy M, Séré Y, Fauquet C, Konaté G. 2004. Inferring the evolutionary history of *Rice yellow mottle virus* from genomic, phyloge-

- netic, and phylogeographic studies. J Virol 78:3252–3261. https://doi.org/10.1128/JVI.78.7.3252-3261.2004.
- 11. Pinel-Galzi A, Traoré O, Séré Y, Hébrard E, Fargette D. 2015. The biogeography of viral emergence: *Rice yellow mottle virus* as a case study. Curr Opin Virol 10:7–13. https://doi.org/10.1016/j.coviro.2014.12.002.
- Fargette D, Pinel A, Halimi H, Brugidou C, Fauquet C, Van RM. 2002. Comparison of molecular and immunological typing of isolates of *Rice yellow mottle virus*. Arch Virol 147:583–596. https://doi.org/10.1007/s007050200008
- Tamura K, Stecher G, Peterson D, Filipski A, Kumar S. 2013. MEGA6: molecular evolutionary genetics analysis version 6.0. Mol Biol Evol 30: 2725–2729. https://doi.org/10.1093/molbev/mst197.
- Ndikumana I, Galzi-Pinel A, Mzengeza T, N'chimbi Msolla S, Njau P, Choi IR, Murori R, Bigirimana J, Fargette D, Hébrard E. 2015. First report of *Rice* yellow mottle virus in rice in Malawi. Plant Dis 99:899. https://doi.org/10 .1094/PDIS-12-14-1298-PDN.
- Traoré O, Sorho F, Pinel A, Abubakar Z, Banwo O, Maley J, Hébrard E, Winter S, Séré Y, Konaté G, Fargette D. 2005. Processes of diversification and dispersion of *Rice yellow mottle virus* inferred from large-scale and high-resolution phylogeographical studies. Mol Ecol 14:2097–2110. https://doi.org/10.1111/j.1365-294X.2005.02578.x.
- Martin DP, Murrell B, Golden M, Khoosal A, Muhire B. 2015. RDP4: detection and analysis of recombination patterns in virus genomes. Virus Evol 1:vev003. https://doi.org/10.1093/ve/vev003.
- Pinel-Galzi A, Rakotomalala M, Sangu E, Sorho F, Kanyeka Z, Traoré O, Sérémé D, Poulicard N, Rabenantoandro Y, Séré Y, Konaté G, Ghesquière A, Hébrard E, Fargette D. 2007. Theme and variations in the evolutionary pathways to virulence of an RNA plant virus species. PLoS Pathog 3:e180. https://doi.org/10.1371/journal.ppat.0030180.
- Traoré O, Pinel-Galzi A, Issaka S, Poulicard N, Aribi J, Aké S, Ghesquière A, Séré Y, Konaté G, Hébrard E, Fargette D. 2010. The adaptation of *Rice yellow mottle virus* to the elF(iso)4G-mediated rice resistance. Virology 408:103–108. https://doi.org/10.1016/j.virol.2010.09.007.

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