

# Checklist of bees (Hymenoptera: Apoidea) from small diversified vegetable farms in south-western Montana

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## Abstract

#### Background

Over three years (2013-2015), we sampled bees using nets and bowl traps on four diversified vegetable farms in Gallatin County, Montana, USA, as part of a study evaluating the use of wildflower strips for supporting wild bees and crop pollination services on farmlands (Delphia et al. In prep). We document 202 species and morphospecies from 32 genera within five families, of which 25 species represent the first published state records for Montana. This study increases our overall understanding of the distribution of wild bee species associated with agroecosystems of the northern US Rockies, which is important for efforts aimed at conserving bee biodiversity and supporting sustainable crop pollination systems on farmlands.

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### New information

We provide a species list of wild bees associated with diversified farmlands in Montana and increase the number of published bee species records in the state from 374 to at least 399. The list includes new distributional records for 25 wild bee species, including two species that represent considerable expansions of their known ranges, *Lasioglossum (Dialictus) clematisellum* (Cockerell 1904) with previously published records from New Mexico, Arizona, California and Utah and *Melissodes (Eumelissodes) niveus* Robertson 1895 which was reported to range from New York to Minnesota and Kansas, south to North Carolina, Alabama and Mississippi.

### Keywords

wild bees, native bees, pollinators, biodiversity, range expansion, farmlands, agroecosystems, Rocky Mountains, Intermountain West, wildflower strips, bee conservation

### Introduction

Native bees are important pollinators of wild and cultivated plants in natural habitats and agricultural systems (e.g. Losey and Vaughan 2006, Klein et al. 2007, Michener 2007, Garibaldi et al. 2013). In the United States and worldwide, however, bees and other pollinators are experiencing probable declines due to factors such as diseases, pesticides and habitat loss that can reduce floral resources and nesting sites (Biesmeijer et al. 2006, Potts et al. 2010, Cameron et al. 2011, Burkle et al. 2013, Goulson et al. 2015, Koh et al. 2016). However, because we lack a baseline understanding of the bee species that occur in many parts of the US, particularly in certain western regions of the country, concerns regarding the status and trends of wild bees cannot be accurately assessed.

Montana's bee fauna is one of the least-studied amongst US states (but see Kuhlman and Burrows 2017, Dolan et al. 2017, Reese et al. 2018); of the few studies that have been conducted in Montana, even fewer have examined wild bees in agricultural systems. Documenting the diversity of bees on farmlands is important for identifying potential crop pollinators, for gauging the potential of farmland habitats to support overall bee diversity on and around farms and for guiding bee conservation measures (Burkle et al. 2017). In addition, the spine of North America, which is the Rocky Mountains, extends from British Columbia to New Mexico and runs through western Montana. This major geographic barrier separates east and west biotas and provides a habitat for alpine/boreal species. Therefore, understanding wild bee distributions in Montana is biogeographically important because its regional species pool likely includes many bee species with typically eastern, western, arctic and southern US ranges that overlap within the state (Gibbs 2010, Koch et al. 2012, Williams et al. 2014, Dolan et al. 2017). A 2017 study, for example, documented

28 bumble bee species in the state, several of which were previously considered to have purely eastern or western North American ranges (Dolan et al. 2017). Another recent survey documented 34 bee species as new state records from just one of Montana's 56 counties (Kuhlman and Burrows 2017). The results from these studies, coupled with Montana's large size (381,000 km<sup>2</sup>) and diverse ecosystems, many of which are difficult to access, suggest there is much to learn about the wild bee fauna in this region.

We report a checklist of bee species from a three-year study surveying the bee community on diversified vegetable farms in south-western Montana. This is the second study (along with Adhikari et al. In prep) in the state to survey wild bees in agroecosystems, though the habitats (Greater Yellowstone Ecosystem versus Northern Great Plains), type of farming systems (diversified versus highly-simplified) and crops (pollinator-dependent versus windpollinated) differed extensively. This work contributes to our long-term goal of creating a comprehensive bee species list for the state.

# Materials and methods

### **Study Sites**

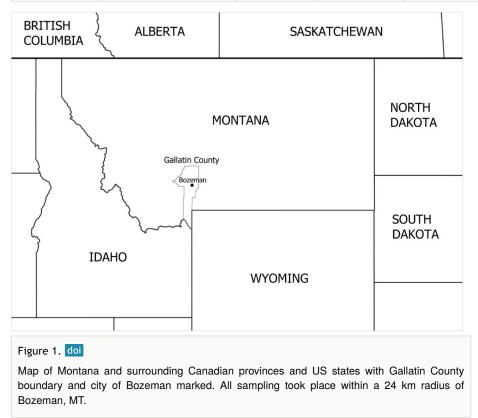
This research was conducted at four diversified farms located in south-western Montana USA within a 24 km radius of Bozeman (45.6769°N; 111.0429°W) in Gallatin County (Fig. 1, Table 1). The farms we surveyed are within the eastern end of the broad Gallatin Valley, which is surrounded by five mountain ranges, two of which are nearby: the Bridger Mountains to the northeast and the Gallatin Range to the south. Each farm had approximately 3-7 acres in cultivation each year and grew a variety of crops, including squashes and pumpkins (Cucurbita pepo L.), tomatoes (Solanum lysopersicum L.), cucumbers (*Cucumis sativus* L.) and strawberries (*Fragaria x ananassa* Duchesne), marketed locally through Community Supported Agriculture (CSA) programmes, farmers' markets, food co-ops and restaurants; two of the farms were certified organic and two followed organic or sustainable practices. Elevations of the farms ranged from 1350-1511 m above sea level. The mean annual precipitation in the area is 469 mm, the mean annual daily high temperature is 12.89°C and the mean annual low temperature is -0.44°C (Western Regional Climate Center 2018; Suppl. material 1). As we were interested in evaluating wildflower strips for supporting bees and crop pollination, our experimental design included planting wildflowers and experimental crop strips from which we sampled bees.

#### Table 1.

Site number, site name, latitude, longitude and elevation of farms sampled in 2013-2015 within a 24 km radius of Bozeman, Montana in Gallatin County.

Site Number	Study Site	Latitude	Longitude	Elevation (m)
1	Gallatin Grown	N45°46.28'	W111°17.72'	1350
2	Gallatin Valley Botanical	N45°39.63'	W110°56.85'	1511

Site Number	Study Site	Latitude	Longitude	Elevation (m)
3	Rocky Creek Farm	N45°39.82'	W110°56.95'	1508
4	Towne's Harvest Garden	N45°39.92'	W111°04.35'	1490



#### **Collection Methods**

We collected bees on each farm from May-September in 2013-2015; all sampling took place on calm, sunny days between 0900 and 1700 h MDT. We net-collected bees visiting the reproductive parts of flowers of species blooming in: 1) the established wildflower strips (*Campanula rotundifolia* L., *Erigeron speciosus* (Lindl.) DC., *Gaillardia aristata* Pursh, *Geranium viscosissimum* Fisch. & C.A. Mey. Ex C.A. Mey., *Helianthus maximiliani* Schrad., *Heterotheca villosa* (Pursh) Shinners, *Monarda fistulosa* L., *Penstemon confertus* Douglas ex Lindl. and *Phacelia hastata* Douglas ex Lehm.) during weekly timed observations in 2014 and 2015; 2) crops (acorn winter squash, *Cucurbita pepo* and sunflower, *Helianthus annuus* L.) during timed observations every other week in 2013 or weekly in 2014 and 2015; and 3) other prolific bloomers, which included primarily agricultural weeds (e.g. common tansy, *Tanacetum vulgare* L. and Canada thistle, *Cirsium arvense* (L.) Scop.), during timed observations once or twice a month, depending on the amount of surrounding

vegetation and time availability. We collected for a total of ca. 23 hours in 2013, 104 hours in 2014 and 85 hours in 2015. In all three years, the same experienced netter (15 years) was paired with a less-experienced netter (≤1 year) for bee collections with both contributing equal amounts of time to collecting; in 2015 the same netter from 2014 assisted with bee collections. Bees were freeze-killed, pinned, and labelled. We also collected bees weekly using yellow, 350-ml Solo bowls filled with soapy water. Six bowls were deployed approximately 6 m apart along each of four, 33-m linear transects (24 bowls per farm) located at different distances from the wildflower strips and left out for approximately six hours during the height of bee activity. Samples were collected into 70% EtOH and later removed from the alcohol, washed, blow-dried, pinned and labelled. We used EstimateS (Colwell 2013) to generate a Chao1 species richness predictor to estimate the "true" number of species present.

#### Species Identification

We identified bees to the lowest taxonomic level possible using published keys (Table 2) following the classifications of Michener (2007); specimens were identified to subspecies only when they could be accurately assigned. We used reference specimens from the US National Pollinating Insect Collection to identify a comprehensive subset of the bees collected in this study; these were then taken back to the O'Neill and Burkle Laboratories to use as reference specimens for species identifications and verification of the remaining material.

#### Table 2.

List of published keys used for species identification. Genus-level identifications were done using Michener et al. (1994).

Family	References		
Andrenidae	Bouseman and LaBerge 1979, Donovan 1977, LaBerge 1967, LaBerge 1969, LaBerge 1973, LaBerge 1977, LaBerge 1980, LaBerge 1985, LaBerge 1986, LaBerge 1989, LaBerge and Bouseman 1970, LaBerge and Ribble 1975, Ribble 1967, Ribble 1974, Thorp 1969, Timberlake 1956, Timberlake 1960, Timberlake 1964, Timberlake 1967		
Apidae	Brumley 1965, Daly 1973, Hurd and Linsley 1951, Koch et al. 2012, LaBerge 1956b, LaBerge 1956a, LaBerge 1961, LaBerge 1963, Rightmyer 2008, Sipes 2001, Thorp et al. 1983, Williams et al. 2014		
Colletidae	Snelling 1966b, Snelling 1966a, Snelling 1970, Stephen 1954		
Halictidae	Coelho 2004, Dumesh and Sheffield 2012, Gibbs 2010, McGinley 1986, McGinley 2003, Roberts 1972, Roberts 1973		
Megachilidae	Baker 1975, De Silva and Packer 2012, Gonzalez and Griswold 2013, Grigarick and Stange 1968, Hurd and Michener 1955, Michener 1938, Michener 1939, Michener 1947, Rightmyer et al. 2010, Sandhouse 1939, Sheffield et al. 2011		

For genera where no taxonomic literature was available for species-level identifications, we grouped bees that appeared morphologically distinct into morphospecies. We assigned each morphospecies a unique number and the letter "F" for females and "M" for males. However, because we could not reliably associate male and female morphospecies as a

single "species" and to avoid inflating species numbers, we included only female morphospecies in this checklist. Species names with *aff*. ('has affinity with') are also treated as morphospecies.

As females of *Agapostemon angelicus* Cockerell and *Agapostemon texanus* Cresson are indistinguishable from one another (Roberts 1972), they have been included with the confirmed males for species counts. Similarly, we could not distinguish amongst females of *Hylaeus mesillae* (Cockerell) and *Hylaeus rudbeckiae* (Cockerell and Casad) and they have been included with the males for species counts. For the Chao1 analysis, we have distributed the numbers of females of *A. angelicus/texanus* and *H. mesillae/rudbeckiae* according to the proportions of confirmed males of each species for each of the two genera.

Due to a paucity of regional keys and an inability to discern any distinctive characters amongst individuals, except for one morphospecies, bees in the genus *Sphecodes* were only identified to genus. Similarly, *Lasioglossum* of the subgenus *Evylaeus* were identified only to subgenus. Due to time and resource constraints, a randomly-chosen subsample of 66 of the 4,173 *Lasioglossum* (*Dialictus*) collected in this study were identified to species.

Voucher specimens will be deposited in the Montana Entomology Collection (MTEC) at Montana State University, Bozeman, MT USA.

#### Range

To determine whether a species was a new state record, we compared our checklist with other published checklists and literature focused on Montana fauna (O'Neill and Seibert 1996, Fultz 2005, Pearce et al. 2012, Kuhlman and Burrows 2017, Dolan et al. 2017, Reese et al. 2018), as well as a recent unpublished study (Adhikari et al. In prep). These comparisons revealed a subset of species unique to our study; to ensure that these were first records for the state, we reviewed each of these species in the Catalog of Hymenoptera in America north of Mexico (Hurd 1979) for additional Montana records. Where the catalogue included a record from Montana, we did not conduct further searches for specific localities since our goal was to discover new records for the state. For those not listed as present in the catalogue, we used it to guide further literature searches. We then used published primary literature (see Literature Cited below in checklist notes) to further search for Montana records and determine species ranges. Our search revealed 25 unpublished state records. We also searched DiscoverLife.org; it revealed unpublished records with specific locality information for 5 of the 25 species (Ascher and Pickering 2018; Suppl. material 2), which are indicated below in the checklist notes. For each of the 25 new state records, we also provide information on the closest records reported within the same literature examined for Montana records (see checklist notes).

# Checklist

#### Family Andrenidae

#### Subfamily Andreninae

### Tribe Andrenini

### Andrena (Andrena) thaspii Graenicher 1903

Notes: Table 1: Sites 1-3.

### Andrena (Andrena) topazana Cockerell 1906

Notes: Table 1: Sites 1, 3.

### Andrena (Callandrena) helianthi Robertson 1891

Notes: Table 1: Sites 1-4.

### Andrena (Cnemidandrena) costillensis Viereck & Cockerell 1914

**Notes:** New species record for Montana (Donovan 1977; Table 1: Sites 2, 3). Unpublished record on DiscoverLife (Suppl. material 2). The closest records reported in Donovan (1977) for this species are from neighbouring states Idaho and Wyoming.

### Andrena (Diandrena) nothocalaidis (Cockerell 1905)

Notes: Table 1: Sites 2, 3.

#### Andrena (Euandrena) astragali Viereck & Cockerell 1914

Notes: Table 1: Sites 2-4.

### Andrena (Euandrena) nigrocaerulea Cockerell 1897

Notes: Table 1: Sites 1-4.

#### Andrena (Holandrena) cressonii subsp. infasciata Robertson 1891

#### Andrena (Melandrena) nivalis Smith 1853

Notes: Table 1: Sites 2, 3.

#### Andrena (Micrandrena) microchlora Cockerell 1922

Notes: Table 1: Sites 2, 3.

#### Andrena (Plastandrena) prunorum Cockerell 1896

Notes: Table 1: Sites 1-4.

#### Andrena (Scaphandrena) scurra Viereck 1904

Notes: Table 1: Site 1.

#### Andrena (Simandrena) nasonii Robertson 1895

**Notes:** New species record for Montana (LaBerge 1989; Table 1: Sites 1, 2). The closest record reported in LaBerge (1989) for this species is from neighbouring state North Dakota.

#### Andrena (Simandrena) pallidifovea (Viereck 1904)

Notes: Table 1: Sites 2, 3.

#### Andrena (Thysandrena) candida Smith 1879

Notes: Table 1: Sites 2-4.

#### Andrena (Thysandrena) medionitens Cockerell 1902

Notes: Table 1: Sites 1-4.

#### Andrena (Thysandrena) vierecki Cockerell 1904

Notes: Table 1: Site 2.

#### Andrena (Trachandrena) hippotes Robertson 1895

Notes: Table 1: Site 2.

#### Andrena (Trachandrena) miranda Smith 1879

Notes: Table 1: Sites 2, 3.

#### Andrena (Tylandrena) aff. wilmattae Cockerell 1906

Notes: Table 1: Site 1.

#### Andrena sp. F1

Notes: Table 1: Sites 2, 3.

#### Andrena sp. F2

Notes: Table 1: Sites 2-4.

#### Andrena sp. F3

Notes: Table 1: Site 3.

### Subfamily Panurginae

#### Tribe Calliopsini

#### Calliopsis (Calliopsima) chlorops Cockerell 1899

**Notes:** New species record for Montana (Shinn 1967; Table 1: Sites 1, 3). The closest records reported in Shinn (1967) for this species are from neighbouring states Idaho and Wyoming.

#### Calliopsis (Calliopsima) coloradensis Cresson 1878

Notes: Table 1: Site 3.

#### Calliopsis (Calliopsis) and reniform is Smith 1853

Notes: Table 1: Sites 2-4.

#### Calliopsis (Nomadopsis) personata Cockerell 1897

**Notes:** New species record for Montana (Timberlake 1952, Rozen 1958; Table 1: Site 1). The closest records reported in Timberlake (1952) and Rozen (1958) for this species are from neighbouring states Idaho and Wyoming.

### Tribe Panurgini

### Panurginus atriceps (Cresson 1878)

Notes: Table 1: Sites 1-4.

### Panurginus sp. F1

Notes: Table 1: Sites 1-4.

### Tribe Perditini

### Perdita (Cockerellia) lingualis Cockerell 1896

**Notes:** New species record for Montana (Timberlake 1960; Table 1: Site 1). The closest records reported in Timberlake (1960) for this species are from Colorado, Nebraska and Utah. Mayer et al. (2000) reported this species from south-eastern Washington.

### Perdita (Perdita) fallax Cockerell 1896

Notes: Table 1: Site 3.

### Perdita (Perdita) salicis Cockerell 1896

**Notes:** New species record for Montana (Timberlake 1964; Table 1: Sites 1, 4). The closest record reported in Timberlake (1964) for this species is from neighbouring state ldaho.

### Perdita (Pygoperdita) wyomingensis subsp. segona Timberlake 1956

Notes: Table 1: Sites 2, 3.

### Tribe Protandrenini

### Protandrena (Pterosarus) innuptus (Cockerell 1896)

Notes: Table 1: Sites 1-3.

### Protandrena (Pterosarus) irregularis (Cockerell 1922)

**Notes:** New species record for Montana (Cockerell 1922; Table 1: Site 1). Unpublished record on DiscoverLife (Suppl. material 2). The closest record reported in Cockerell (1922) and Hurd (1979) for this species is from Colorado.

### Protandrena (Pterosarus) piercei (Crawford 1903)

**Notes:** New species record for Montana (Cockerell 1922; Table 1: Site 1). The closest records reported in Hurd (1979) for this species are from neighbouring Canadian province Alberta and from neighbouring US state North Dakota.

### Family Apidae

#### Subfamily Apinae

#### Tribe Anthophorini

### Anthophora (Clisodon) terminalis Cresson 1869

Notes: Table 1: Sites 1-4.

### Anthophora (Melea) bomboides Kirby 1837

Notes: Table 1: Site 1.

#### Anthophora (Mystacanthophora) urbana Cresson 1878

Notes: Table 1: Sites 1, 3, 4.

### Tribe Apini

### Apis (Apis) mellifera Linnaeus 1758

Native status: Non-native to North America.

Notes: Table 1: Sites 1-4.

### Tribe Bombini

### Bombus (Bombias) nevadensis Cresson 1874

Notes: Table 1: Sites 1-4.

### Bombus (Bombus) occidentalis Greene 1858

### Bombus (Cullumanobombus) griseocollis (De Geer 1773)

Notes: Table 1: Site 1.

#### Bombus (Cullumanobombus) rufocinctus Cresson 1863

Notes: Table 1: Sites 1-4.

#### Bombus (Psithyrus) insularis (Smith 1861)

Notes: Table 1: Sites 1-4.

### Bombus (Pyrobombus) bifarius Cresson 1878

Notes: Table 1: Sites 1-4.

### Bombus (Pyrobombus) centralis Cresson 1864

Notes: Table 1: Sites 1-4.

### Bombus (Pyrobombus) flavifrons Cresson 1863

Notes: Table 1: Sites 2, 4.

### Bombus (Pyrobombus) huntii Greene 1860

Notes: Table 1: Sites 1-4.

### Bombus (Pyrobombus) mixtus Cresson 1878

Notes: Table 1: Sites 1-4.

### Bombus (Pyrobombus) sylvicola Kirby 1837

Notes: Table 1: Site 4.

### Bombus (Subterraneobombus) appositus Cresson 1878

Notes: Table 1: Sites 1-3.

#### Bombus (Thoracobombus) fervidus (Fabricius 1798)

#### Tribe Emphorini

### Diadasia (Coquillettapis) diminuta (Cresson 1878)

Notes: Table 1: Sites 1, 3, 4.

### Tribe Eucerini

#### Melissodes (Callimelissodes) lupinus Cresson 1878

Notes: Table 1: Sites 1, 4.

#### Melissodes (Callimelissodes) metenua Cockerell 1924

**Notes:** New species record for Montana (LaBerge 1961; Table 1: Site 3). The closest records reported in LaBerge (1961) and Hurd (1979) for this species are from neighbouring states Idaho and Wyoming.

#### Melissodes (Eumelissodes) agilis Cresson 1878

Notes: Table 1: Sites 1-4.

### Melissodes (Eumelissodes) confusus Cresson 1878

Notes: Table 1: Site 4.

#### Melissodes (Eumelissodes) coreopsis Robertson 1905

Notes: Table 1: Sites 1-4.

#### Melissodes (Eumelissodes) hymenoxidis Cockerell 1906

Notes: Table 1: Site 4.

#### Melissodes (Eumelissodes) menuachus Cresson 1868

Notes: Table 1: Site 1.

#### Melissodes (Eumelissodes) microstictus Cockerell 1905

### Melissodes (Eumelissodes) niveus Robertson 1895

**Notes:** New species record for Montana (LaBerge 1961; Table 1: Site 1). The closest records reported in LaBerge (1961) for this species are from Minnesota and Nebraska.

### Melissodes (Eumelissodes) pallidisignatus Cockerell 1905

Notes: Table 1: Site 1.

### Melissodes (Eumelissodes) perlusus Cockerell 1925

**Notes:** New species record for Montana (LaBerge 1961; Table 1: Site 1). The closest records reported in LaBerge (1961) for this species are from neighbouring Canadian province Alberta and from neighbouring US states North Dakota and Wyoming.

### Melissodes (Melissodes) communis Cresson 1878

**Notes:** New species record for Montana (LaBerge 1956b; Table 1: Sites 3, 4). The closest records reported in LaBerge (1956b) for this species are from neighbouring Canadian provinces British Columbia and Alberta and from neighbouring US states Idaho, North Dakota, South Dakota and Wyoming.

#### **Tribe Melectini**

### Xeromelecta (Melectomorpha) californica (Cresson 1878)

Notes: Table 1: Site 1.

### Subfamily Nomadinae

Tribe Epeolini

#### Epeolus sp. F1

Notes: Table 1: Site 3.

### Triepeolus paenepectoralis Viereck 1905

Notes: Table 1: Sites 1, 3, 4.

### Triepeolus sp. F1

**Notes:** Table 1: Sites 1, 3, 4.

#### Triepeolus sp. F2

Notes: Table 1: Site 4.

### Tribe Nomadini

### Nomada ruficornis group sp. F1

Notes: Bidentate; Table 1: Site 2.

### Nomada ruficornis group sp. F2

Notes: Unidentate; Table 1: Sites 1, 4.

#### Nomada ruficornis group sp. F3

Notes: Unidentate; Table 1: Site 2.

### Nomada ruficornis group sp. F4

Notes: Bidentate; Table 1: Site 2.

#### Nomada ruficornis group sp. F5

Notes: Bidentate; Table 1: Sites 2, 3.

#### Nomada ruficornis group sp. F6

Notes: Bidentate; Table 1: Sites 2-4.

#### Nomada ruficornis group sp. F7

Notes: Unidentate; Table 1: Sites 1, 2, 4.

#### Nomada ruficornis group sp. F8

Notes: Unidentate; Table 1: Sites 1, 4.

#### Nomada ruficornis group sp. F9

Notes: Unidentate; Table 1: Site 1.

### Nomada ruficornis group sp. F10

Notes: Unidentate; Table 1: Sites 2, 4.

### Nomada ruficornis group sp. F11

Notes: Unidentate; Table 1: Sites 2, 3.

### Subfamily Xylocopinae

### Tribe Ceratinini

### Ceratina (Zadontomerus) acantha Provancher 1895

Notes: Table 1: Site 3.

### Ceratina (Zadontomerus) nanula Cockerell 1897

Notes: Table 1: Sites 2, 3.

### Ceratina (Zadontomerus) neomexicana Cockerell 1901

Notes: Table 1: Sites 2-4.

### Family Colletidae

### Subfamily Colletinae

### Tribe Colletini

### Colletes aff. phaceliae Cockerell 1906

Notes: Table 1: Site 4.

### Colletes consors subsp. pascoensis Cockerell 1898

Notes: Table 1: Site 4.

### Colletes fulgidus subsp. fulgidus Swenk 1904

### Colletes kincaidii Cockerell 1898

Notes: Table 1: Site 2.

### Colletes lutzi subsp. lutzi Timberlake 1943

Notes: Table 1: Site 1.

#### Colletes nigrifrons Titus 1900

Notes: Table 1: Site 2.

#### Colletes phaceliae Cockerell 1906

Notes: Table 1: Sites 2-4.

### Subfamily Hylaeinae

### Hylaeus (Cephalylaeus) basalis (Smith 1853)

Notes: Table 1: Site 3.

#### Hylaeus (Hylaeus) annulatus (Linnaeus 1758)

Notes: Table 1: Sites 2, 3.

#### Hylaeus (Hylaeus) conspicuus (Metz 1911)

Notes: Table 1: Site 3.

#### Hylaeus (Hylaeus) leptocephalus (Morawitz 1871)

Native status: Non-native to North America.

Notes: Table 1: Site 1.

#### Hylaeus (Hylaeus) mesillae (Cockerell 1896)

Notes: Table 1: Sites 2-4.

### Hylaeus (Hylaeus) rudbeckiae (Cockerell and Casad 1895)

### Hylaeus (Hylaeus) verticalis (Cresson 1869)

Notes: Table 1: Sites 2, 3.

### Hylaeus (Paraprosopis) coloradensis (Cockerell 1896)

Notes: Table 1: Sites 2-4.

#### Hylaeus (Paraprosopis) wootoni (Cockerell 1896)

Notes: Table 1: Sites 1-4.

### Hylaeus (Prosopis) affinis (Smith 1853)

**Notes:** New species record for Montana (Metz 1911, Snelling 1966a; Table 1: Sites 2-4). The closest records reported in Snelling (1966a) for this species are from neighbouring Canadian provinces British Columbia and Saskatchewan and from neighbouring US state Idaho.

### Hylaeus (Prosopis) modestus Say 1837

Notes: Table 1: Sites 1-3.

### Family Halictidae

### Subfamily Halictinae

#### Tribe Augochlorini

#### Augochlorella aurata (Smith 1853)

Notes: Table 1: Sites 2, 3.

#### Tribe Halictini

### Agapostemon (Agapostemon) angelicus Cockerell 1924

**Notes:** Table 1: Sites 1, 3, 4.

#### Agapostemon (Agapostemon) femoratus Crawford 1901

#### Agapostemon (Agapostemon) texanus Cresson 1872

Notes: Table 1: Site 1.

#### Agapostemon (Agapostemon) virescens (Fabricius 1775)

Notes: Table 1: Sites 1-4.

#### Halictus (Odontalictus) ligatus Say 1837

Notes: Table 1: Sites 1-4.

#### Halictus (Protohalictus) rubicundus (Christ 1791)

Notes: Table 1: Sites 1-4.

#### Halictus (Seladonia) confusus Smith 1853

Notes: Table 1: Sites 1-4.

#### Halictus (Seladonia) tripartitus Cockerell 1895

Notes: Table 1: Sites 1-4.

#### Lasioglossum (Dialictus) aff. admirandum (Sandhouse 1924)

Notes: Table 1: Sites 2, 4.

#### Lasioglossum (Dialictus) clematisellum (Cockerell 1904)

**Notes:** New species record for Montana (Sandhouse 1924; Table 1: Site 1). The closest record reported in Hurd (1979) for this species is from Utah.

#### Lasioglossum (Dialictus) cressonii (Robertson 1890)

**Notes:** New species record for Montana (Sandhouse 1924, Gibbs 2010; Table 1: Site 4). Unpublished record on DiscoverLife (Suppl. material 2). The closest records reported in Gibbs (2010) for this species are from neighbouring Canadian provinces British Columbia and Alberta.

#### Lasioglossum (Dialictus) laevissimum (Smith 1853)

Notes: New species record for Montana (Sandhouse 1924, Gibbs 2010; Table 1: Sites 1-4). Unpublished record on DiscoverLife Suppl. material 2). The closest records

reported in Gibbs (2010) for this species are from neighbouring Canadian provinces British Columbia, Alberta and Saskatchewan.

### Lasioglossum (Dialictus) lilliputense Gibbs 2010

Notes: Table 1: Sites 2-4.

#### Lasioglossum (Dialictus) lineatulum (Crawford 1906)

**Notes:** New species record for Montana (Gibbs 2010; Table 1: Site 2). The closest records reported in Gibbs (2010) for this species are from neighbouring Canadian provinces Alberta and Saskatchewan.

### Lasioglossum (Dialictus) occidentale (Crawford 1902)

Notes: Table 1: Sites 1-4.

#### Lasioglossum (Dialictus) semicaeruleum (Cockerell 1895)

Notes: Table 1: Sites 1, 3.

#### Lasioglossum (Dialictus) succinipenne (Ellis 1913)

Notes: Table 1: Site 2.

#### Lasioglossum (Dialictus) versans (Lovell 1905)

**Notes:** New species record for Montana (Sandhouse 1924, Gibbs 2010; Table 1: Site 3). The closest record reported in Gibbs (2010) for this species is from neighbouring Canadian province Alberta.

#### Lasioglossum (Dialictus) zephyrum (Smith 1853)

Notes: Table 1: Site 1.

### Lasioglossum (Dialictus) spp.

Notes: Table 1: Sites 1-4.

### Lasioglossum (Evylaeus) spp.

### Lasioglossum (Hemihalictus) ovaliceps (Cockerell, 1898)

Notes: Table 1: Site 3.

### Lasioglossum (Lasioglossum) egregium (Vachal 1904)

Notes: Table 1: Site 2.

#### Lasioglossum (Lasioglossum) mellipes (Crawford 1907)

**Notes:** New species record for Montana (McGinley 1986; Table 1: Site 4). The closest records reported in McGinley (1986) for this species are from neighbouring Canadian province British Columbia and from neighbouring US state Idaho.

#### Lasioglossum (Lasioglossum) paraforbesii McGinley 1986

Notes: Table 1: Site 1.

### Lasioglossum (Lasioglossum) sisymbrii (Cockerell 1895)

Notes: Table 1: Site 2.

#### Lasioglossum (Lasioglossum) titusi (Crawford 1902)

Notes: Table 1: Sites 2, 3.

### Lasioglossum (Leuchalictus) zonulum (Smith 1848)

Notes: Table 1: Sites 1, 4.

#### Lasioglossum (Sphecodogastra) lusorium (Cresson 1872)

Notes: Table 1: Site 1.

#### Sphecodes sp. F1

Notes: Table 1: Sites 2, 4.

#### Sphecodes spp.

### Subfamily Rophitinae

### Dufourea marginata (Cresson 1878)

**Notes:** New species record for Montana (Michener 1951; Table 1: Sites 1, 4). The closest records reported in Hurd (1979) for this species are from neighbouring Canadian province Alberta and from neighbouring state Wyoming.

### Dufourea maura (Cresson 1878)

Notes: Table 1: Sites 2, 3.

### Dufourea trochantera Bohart 1948

Notes: Table 1: Site 4.

### Family Megachilidae

### Subfamily Megachilinae

Tribe Anthidiini

### Anthidium (Anthidium) mormonum Cresson 1878

Notes: Table 1: Sites 1, 3.

### Anthidium (Anthidium) tenuiflorae Cockerell 1907

Notes: Table 1: Sites 1-4.

### Dianthidium (Dianthidium) heterulkei Schwarz 1940

**Notes:** New species record for Montana (Timberlake 1943, Grigarick and Stange 1968; Table 1: Site 3). The closest record reported in Hurd (1979) for this species is from neighbouring state Wyoming.

### Dianthidium (Dianthidium) subparvum Swenk 1914

Notes: Table 1: Sites 2, 3.

### Stelis (Stelis) calliphorina (Cockerell 1911)

Notes: Table 1: Site 3.

#### Stelis (Stelis) foederalis group sp. 6

Notes: Table 1: Site 3.

#### Stelis (Stelis) lateralis Cresson 1864

Notes: Table 1: Sites 2, 4.

#### Stelis (Stelis) montana Cresson 1864

Notes: Table 1: Site 3.

#### Stelis (Stelis) occidentalis Parker & Griswold 2013

**Notes:** New species record for Montana (Parker and Griswold 2013; Table 1: Site 3). The closest records reported in Parker and Griswold (2013) for this species are from neighbouring Canadian province British Columbia and from neighbouring US state Idaho.

### Stelis (Stelis) permaculata Cockerell 1898

Notes: Table 1: Site 2.

#### Stelis (Stelis) subcaerulea Cresson 1878

**Notes:** New species record for Montana (Cockerell 1911; Table 1: Site 3). The closest record reported in Hurd (1979) for this species is from neighbouring state Wyoming.

### Stelis (Stelis) subemarginata group sp. 1

Notes: Table 1: Sites 2, 4.

#### Tribe Megachilini

#### Coelioxys (Boreocoelioxys) moestus Cresson 1864

Notes: Table 1: Sites 2, 3.

#### Coelioxys (Boreocoelioxys) rufitarsis Smith 1854

### Coelioxys (Coelioxys) sodalis Cresson 1878

Notes: Table 1: Site 4.

### Coelioxys (Synocoelioxys) alternatus Say 1837

Notes: Table 1: Sites 2, 3.

### Megachile (Argyropile) parallela Smith 1853

Notes: Table 1: Sites 1, 3, 4.

### Megachile (Chelostomoides) angelarum Cockerell 1902

Notes: Table 1: Sites 2, 3.

### Megachile (Chelostomoides) campanulae (Robertson 1903)

Notes: Table 1: Site 2.

### Megachile (Eutricharaea) apicalis Spinola 1808

Native status: Non-native to North America.

Notes: Table 1: Sites 2-4.

### Megachile (Eutricharaea) rotundata (Fabricius 1787)

Native status: Non-native to North America.

Notes: Table 1: Sites 1, 3, 4.

#### Megachile (Litomegachile) brevis Say 1837

Notes: Table 1: Site 1.

### Megachile (Megachile) lapponica Thomson 1872

Notes: Table 1: Sites 2, 3.

### Megachile (Megachile) montivaga Cresson 1878

#### Megachile (Megachile) relativa Cresson 1878

Notes: Table 1: Sites 1-4.

#### Megachile (Megachiloides) anograe Cockerell 1908

Notes: Table 1: Site 4.

#### Megachile (Megachiloides) subnigra Cresson 1879

Notes: Table 1: Site 1.

#### Megachile (Sayapis) pugnata Say 1837

Notes: Table 1: Sites 1-4.

### Megachile (Xanthosarus) frigida Smith 1853

Notes: Table 1: Sites 1, 2, 4.

### Megachile (Xanthosarus) latimanus Say 1823

Notes: Table 1: Site 1.

#### Megachile (Xanthosarus) melanophaea Smith 1853

Notes: Table 1: Sites 1-3.

### Megachile (Xanthosarus) perihirta Cockerell 1898

Notes: Table 1: Sites 1-4.

### Tribe Osmiini

#### Ashmeadiella (Ashmeadiella) bucconis (Say 1837)

Notes: Table 1: Sites 1, 3, 4.

#### Heriades (Neotrypetes) carinata Cresson 1864

#### Heriades (Neotrypetes) cressoni Michener 1938

Notes: Table 1: Sites 2, 3.

#### Heriades (Neotrypetes) variolosa (Cresson 1872)

Notes: Table 1: Sites 1-4.

#### Hoplitis (Alcidamea) fulgida (Cresson 1864)

Notes: Table 1: Sites 2, 3.

#### Hoplitis (Alcidamea) grinnelli (Cockerell 1910)

Notes: Table 1: Sites 2-4.

#### Hoplitis (Alcidamea) hypocrita (Cockerell 1906)

Notes: Table 1: Site 2.

#### Hoplitis (Alcidamea) pilosifrons (Cresson 1864)

Notes: Table 1: Site 4.

#### Hoplitis (Alcidamea) producta (Cresson 1864)

Notes: Table 1: Sites 1-4.

#### Hoplitis (Alcidamea) spoliata (Provancher 1888)

**Notes:** New species record for Montana (Michener 1947, Mitchell 1960; Table 1: Sites 3, 4). Unpublished record on DiscoverLife (Suppl. material 2). The closest records reported in Michener (1947) for this species are from neighbouring Canadian provinces British Columbia and Saskatchewan and from neighbouring US state North Dakota.

### Hoplitis (Formicapis) robusta (Nylander 1848)

Notes: Table 1: Sites 2-4.

### Osmia (Cephalosmia) marginipennis Cresson 1878

Notes: Table 1: Site 2.

### Osmia (Cephalosmia) subaustralis Cockerell 1900

Notes: Table 1: Site 3.

#### Osmia (Helicosmia) coloradensis Cresson 1878

Notes: Table 1: Sites 2-4.

#### Osmia (Melanosmia) aff. paradisica Sandhouse 1924

Notes: Table 1: Sites 3, 4.

#### Osmia (Melanosmia) aff. phaceliae Cockerell 1907

Notes: Table 1: Site 3.

#### Osmia (Melanosmia) albolateralis Cockerell 1906

Notes: Table 1: Sites 2-4.

### Osmia (Melanosmia) atrocyanea Cockerell 1897

Notes: Table 1: Site 2.

#### Osmia (Melanosmia) brevis Cresson 1864

Notes: Table 1: Sites 2, 3.

### Osmia (Melanosmia) bruneri Cockerell 1897

Notes: Table 1: Site 1.

#### Osmia (Melanosmia) ednae Cockerell 1907

Notes: Table 1: Sites 2-4.

#### Osmia (Melanosmia) kincaidii Cockerell 1897

Notes: Table 1: Site 3.

#### Osmia (Melanosmia) odontogaster group sp. 1

**Notes:** Formerly, subgenus *Acanthosmioides* Ashmead (Rightmyer et al. 2013; Table 1: Site 1).

#### Osmia (Melanosmia) paradisica Sandhouse 1924

Notes: Table 1: Site 4.

#### Osmia (Melanosmia) pusilla Cresson 1864

Notes: Table 1: Sites 2-4.

#### Osmia (Melanosmia) sculleni Sandhouse 1939

Notes: Table 1: Site 4.

#### Osmia (Melanosmia) sedula Sandhouse 1924

**Notes:** New species record for Montana (Donovan 1977, Sandhouse 1939, White 1952; Table 1: Sites 2-4). The closest records reported in Sandhouse (1939) and White (1952) for this species are from neighbouring Canadian province British Columbia and from neighbouring US state Wyoming.

### Osmia (Melanosmia) simillima Smith 1853

Notes: Table 1: Sites 2, 3.

### Osmia (Melanosmia) tersula Cockerell 1912

Notes: Table 1: Site 3.

### Osmia (Melanosmia) trevoris Cockerell 1897

Notes: Table 1: Sites 1-4.

#### Osmia (Melanosmia) tristella Cockerell 1897

Notes: Table 1: Sites 2-4.

#### Osmia (Melanosmia) sp. F1

Notes: Table 1: Site 3.

# Analysis

Over 3 years (2013-2015), we collected 12,203 bees representing 202 species and morphospecies from 32 genera and five families; the list includes wild native, wild nonnative and managed non-native bee species (reviewed in Russo 2016, see 'native status' in checklist, Suppl. materials 3, 4). We found that 25 species are new published records for the state of Montana (Suppl. material 5). The total number of specimens, genera and species for each family are as follows: Andrenidae: 1,510 specimens, 5 genera, 36 species; Colletidae: 552 specimens, 2 genera, 18 species; Halictidae: 6,455 specimens, 6 genera, 35 species and morphospecies; Megachilidae: 925 specimens, 9 genera, 64 species and morphospecies. Analysis with EstimateS (Colwell 2013) yielded a Chao1 mean prediction of 251 bee species for the area (SD = 18.01; 95% confidence interval = 226.48-300.51 species), which is considerably higher than the observed number of species collected (Suppl. material 6).

# Discussion

Our study provides information on the wild bee species associated with diverse, smallscale agricultural farmlands in south-western Montana, expands the known distribution of several bee species to Montana and adds to a growing state list. The 25 new state records reported in this study brings the total number of published species records in the state to 399. Coupled with other recently published works (Dolan et al. 2017, Kuhlman and Burrows 2017, Reese et al. 2018), the high percentage (12.4%) of new state records amongst the bees at our sites further highlights the lack of published data and the need for additional bee biodiversity surveys throughout the state to develop a complete species list, especially considering that our study was conducted within an area measuring approximately 1,810 km<sup>2</sup> (< 1% the size of the state). For example, a checklist of bees recently published reported 34 new Montana state records at a site ca. 260 km from ours (Kuhlman and Burrows 2017). Additionally, two new state records were recorded the same year for bumble bees (Dolan et al. 2017), one of the most well-known and easily recognisable groups amongst the wild bees. At least six unpublished species records are reported by Reese et al. (2018). Considering that two of these lists are from just one county each (present study and Kuhlman and Burrows 2017) and one (Reese et al. 2018) included three of Montana's 56 counties and that Montana comprises a large, topographically and ecologically diverse region with disparate climatic conditions, this is only a start to the additional surveying that needs to be conducted across the state.

It is not currently possible to accurately compare the number of bee species in Montana to other US states and Canadian provinces, since we are far from a statewide inventory. In addition, few species lists have been published for western US states, though lists have been published for several mid-western and north-eastern US states. The closest, comprehensive, statewide bee list is from Colorado which has 946 species (Scott et al. 2011). Wyoming, Montana's neighbour to the south, has 487 documented bee species with additional species (>150) predicted based on distributional patterns, though faunal assessments are not complete (Lavigne and Tepedino 1976). To the north of Montana in Canada, so far, ca. 225 bee species have been recorded in Saskatchewan, ca. 325 species in Alberta and ca. 425 species in British Columbia (Canadian Endangered Species Conservation Council 2016).

Many of the bee species we document as new state records have distributions predicted to include Montana or records from states and provinces that either border Montana or are reasonably close to Montana (and with similar ecosystems). We highlight two bee species on this checklist, *L. clematisellum* and *M. niveus*, whose known distributions are considerably increased with their documentation in Montana. The south-western US distribution of *L. clematisellum* previously included New Mexico, Arizona, California and Utah (Sandhouse 1924, Hurd 1979); unpublished records exist also in Wyoming (pers. comm. Joel Gardner). The eastern US distribution of *M. niveus* ranged from New York to Minnesota and Kansas, south to North Carolina, Alabama and Mississippi (LaBerge 1961). Our findings support the importance of documenting Montana's bee fauna for understanding the full distributional ranges of the wild bees in North America and for producing more comprehensive regional keys.

In addition to wild, native bee species, we also documented four non-native species, including two economically important, commercially-managed species intentionally introduced for crop pollination and two wild, non-managed species accidentally introduced to the US (Klein et al. 2007, Pitts-Singer and Cane 2011, Russo 2016). Managed honey bee, Apis mellifera, colonies were located at one of our sites, as well as nearby sites and were captured in bowl traps and observed visiting our experimental crop strips and wildflower strips. We also captured alfalfa leafcutting bees, Megachile rotundata, in bowl traps and with nets from two non-native plant species, Lotus corniculatus L. and Melilotus officinalis (L.) Lam. Megachile rotundata are managed for alfalfa (Medicago sativa L.) seed production (Pitts-Singer and Cane 2011 in Montana, an important alfalfa seed-producing state (USDA-NASS 2017). Another leafcutting bee Megachile apicalis was recently documented in the literature to occur in Montana (Kuhlman and Burrows 2017) and was netted from E. speciosus in our wildflower strips as well as captured in bowls. Last, the yellow-faced bee Hylaeus leptocephalus was netted from E. speciosus and G. viscosissimum in our flower strips. The impacts (positive and negative) of both intentionally and accidentally introduced non-natives on other bee species (e.g. competition for floral resources and nesting sites and pathogen transmission) and plants (e.g. pollination of native plants, invasive weeds and agricultural crops) require further study (reviewed in Russo 2016).

Our results underestimate the actual bee richness from this study. Though the vast majority of bees were identified to species in our study, the absence of revisionary studies for several genera or subgenera precluded morphospecies sorts for some and, for others, the morphospecies counts may be low because male morphospecies, some of which might not be conspecific with any of the female morphospecies, were not counted. Bees in one genus, *Nomada*, could only be designated as morphospecies and bee species of *Sphecodes* and *Lasioglossum* (subgenus *Evylaeus*) were classified only to the generic or subgeneric level. In addition, only a fraction of the *Lasioglossum* in the subgenus *Dialictus*, which accounted for about 25% of the specimens collected in our study, were identified to species. Bees in the subgenus *Dialictus* are very abundant in studies using bowl traps (Droege et al. 2010) and the lack of species-level identifications for this group are similar amongst faunistic studies in Montana (e.g. over 22,000 unidentified *Dialictus* in Kuhlman and Burrows 2017 and over 2,500 unidentified *Dialictus* in Adhikari et al. In prep). All of these groups require taxonomic work in the western US before species can be fully resolved and will likely contribute dozens of species to a state list for Montana once identified.

Different habitats throughout the state, particularly farmlands versus wildland habitats, are likely to support different suites of bee species. For example, between 33-43% of species were unique to our study when compared to all species (excluding morphospecies) documented in each of two studies conducted in montane wildland habitats in Montana (Kuhlman and Burrows 2017, Reese et al. 2018). However, between 57-66% of bee species in our study were shared with each of those same two studies, indicating considerable overlap with our study and that some bee species can be supported by both types of habitats. When we compared bee species (excluding morphospecies) from all three studies, we found 58 of 195 species (30%) unique to Kuhlman and Burrows (2017), 73 of 226 species (32%) unique to Reese et al. (2018) and 40 of 170 species (24%) unique to our study. All three studies shared 81 species in common, accounting for almost half (48%) of the species in our study, 42% of the species in Kuhlman and Burrows (2017) and 36% in Reese et al. (2018). However, our comparisons did not account for differences in collection methods, sampling effort and geographic area amongst studies, all important determinants of species overlap. More standardised inventories, as well as complete statewide surveys, are needed for more accurate comparisons between studies and habitats. (Such efforts are hampered by the taxonomic impediment.)

In contrast, when we compared our study to another conducted in a highly-simplified, small grains-wheat farming system in north central (Chouteau County) Montana (i.e. the drylands of the Northern Great Plains), we found 73% of bee species were unique to our study compared to those reported by Adhikari et al. (In prep), 27% of bee species were shared (four species are amongst the new state records reported here) and 39 of 85 species (46%) of bee species were unique to Adhikari et al. (In prep). These results suggest the bee communities, which these two agricultural habitats support, are quite different and may be further driven by regional ecosystem diversity. Furthermore, if we compare all four studies, only 28 bee species (16% of the bees in our study) are shared. Again, methodological differences (e.g. sampling intensity, geographic area) between studies make comparisons difficult. Additional surveying is greatly needed in different habitats throughout the state to better understand the basic biology, ecology and distribution of Montana's wild bees, whose importance to natural- and agroecosystems is not fully understood. These types of data are valuable for directing projects aimed at supporting farmland biodiversity and for conserving wild, native bees in general throughout the state.

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# Author contributions

Delphia, Burkle and O'Neill conceived the ideas, designed methodology and secured funding. Delphia collected, processed and identified bee specimens and reviewed the published literature for new state records. Burkle and O'Neill collected specimens. Griswold and Reese identified bee specimens. All authors participated in writing the manuscript, contributed to drafts and gave final approval for publication.

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# Supplementary materials

### Suppl. material 1: Supplementary Table 1 doi

Authors: C. M. Delphia, T. Griswold, E. G. Reese, K. M. O'Neill, L. A. Burkle Data type: weather data Filename: Supp Table 1-Weather data.docx - <u>Download file</u> (27.49 kb)

### Suppl. material 2: Supplementary Table 2 doi

Authors: C. M. Delphia, T. Griswold, E. G. Reese, K. M. O'Neill, L. A. Burkle Data type: occurence

Filename: Supp Table 2-Discover Life occurrence records.xlsx - Download file (20.87 kb)

#### Suppl. material 3: Supplementary Table 3 doi

Authors: C. M. Delphia, T. Griswold, E. G. Reese, K. M. O'Neill, L. A. Burkle Data type: checklist Filename: Supp Table 3-Checklist\_Taxon\_Template for uploading.xls - Download file (393.50 kb)

#### Suppl. material 4: Supplementary Table 4 doi

Authors: C. M. Delphia, T. Griswold, E. G. Reese, K. M. O'Neill, L. A. Burkle Data type: checklist Filename: Supp Table 4-Checklist DWCA format.xlsx - Download file (42.82 kb)

#### Suppl. material 5: Supplementary Table 5 doi

Authors: C. M. Delphia, T. Griswold, E. G. Reese, K. M. O'Neill, L. A. Burkle Data type: occurence Filename: Supp Table 5-Occurence data MT records.xlsx - <u>Download file</u> (19.28 kb)

#### Suppl. material 6: Supplementary Table 6 doi

Authors: C. M. Delphia, T. Griswold, E. G. Reese, K. M. O'Neill, L. A. Burkle Data type: species Brief description: Raw data for Chao1 analysis. Filename: Supp Table 6-Raw data for Chao1 analysis.xlsx - Download file (14.16 kb)