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RESEARCH ARTICLE

Race and virulence characterization of *Puccinia graminis* f. sp. *tritici* in China

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Abstract

Wheat stem rust was once the most destructive plant disease, but it has been largely controlled. However, to prevent future problems, the ongoing development of resistant wheat varieties requires knowledge of the changing virulence patterns for Pgt virulence of the fungus that causes wheat stem rust and the detection of new races. Surveys were conducted from 2013–2014 to determine the races of the Pgt present in China. Low levels of stem rust infections were found in China during this investigation and 11 Puccinia graminis f. sp. tritici (Pgt) samples were obtained. In addition, 22 Pgt samples collected from the alternate host (Berberis) were obtained and have been reported for the first time. Fifty-three isolates were obtained from all samples. Four race groups, including 13 physiological races, were identified. They included the most prevalent races, 34C3MTGQM and 34C6MRGQM, with 13.2% predominance, followed by 34C0MRGQM at 11.3%. Six new races that were virulent against the resistance genes, Sr5 + Sr11, were found for the first time in China, namely 34C0MRGQM, 34C3MTGQM, 34C3MKGQM, 34C3MKGSM, 34C6MTGSM, and 34C6MRGQM, with a predominance of 11.3, 13.2, 9.4, 9.4, 1.9, and 13.2%, respectively. Most of the genes studied were ineffective against one or more of the tested isolates, except Sr9e, Sr21, Sr26, Sr31, Sr33, Sr38, Sr47, and SrTt3. Genes Sr35, SrTmp, Sr30, Sr37, Sr17, and Sr36 were effective in 92.5, 86.8, 84.9, 84.9, 79.3, and 77.4% of the tested isolates, respectively. In contrast, all of the isolates were virulent against Sr6, Sr7b, Sr9a, Sr9b, Sr9d, Sr9g, and SrMcN. Our results indicate that remarkable differences exist among the categories of the races in this study (i.e., their known virulence gene spectra) and the Pgt races reported previously. In addition, the sexual cycle of Pgt may contribute to its diversity in China.

Introduction

Wheat stem rust caused by *Puccinia graminis* Pers. f. sp. *tritici* Eriks. and E. Henn. (*Pgt*) is a major disease of wheat. Heavy epidemics occurred frequently from the 1920s until the 1960s, causing huge yield losses. In recent years, because of the cultivation and utilization of resistant cultivars of wheat, and changes in the cultivation system in China after 1980, wheat stem rust now only occurs sporadically under natural conditions in the Yangtze River area [1]. However, it is undeniable that this disease had devastated wheat production in China and other parts of

the world. Exploring resistance genes and growing resistant cultivars are effective measures for controlling this disease because of low cost, effectiveness, and environmental friendliness [2]. However, the changing virulence patterns and the evolution of fungal races constitute a threat to most wheat germplasm resources. For example, the new races, TTKSK (known as Ug99), TKTTF, and TTTTF, and their variants, threaten wheat production worldwide [3–5].

Maintaining dynamic information on pathogens is the basis for breeding disease resistance. Although wheat stem rust has not been common in recent years, understanding the population structure, virulence diversity, and distribution of *Pgt* at various times and in different locations is essential for selecting effective resistance genes to control this disease [6]. Furthermore, work on detecting and identifying races of *Pgt* has not stopped. Particularly, the emergence and rapid propagation of Ug99 and its variants have attracted increasing attention worldwide, and 28 trapping plots or nurseries have been established yearly in China under the unified arrangement of the wheat rust and powdery mildew cooperative group [7]. Simultaneously, wheat breeding and disease researchers have focused on the investigation and collection of *Pgt* to identify and analyze wheat stem rust. Particularly, aecia from the wheat stem rust alternate host (*Berberis*) were collected from 2013–2014. The objective of the present study was to analyze the pathotypes and characterization of *Pgt* races.

Materials and methods

Ethic statement

Samples in this study were obtained from public areas that no specific permissions were required for these locations. Fields sites are public access and *P. graminis* f. sp. *tritici* is not an endangered or protected species.

Collection of wheat stem rust samples

In 2013–2014, eleven samples of rust-infected stems were collected from wheat fields in Hubei, Heilongjiang, Liaoning, Guizhou, Sichuan, Yunnan, and Gansu provinces (Fig 1), and 22 samples previously collected from the alternate host, *Berberis*, alternate host for many *Puccinia* spp. including *graminis* and *striiformis*, were isolated and identified.

Isolation and multiplication of spores

Seedlings of the universally rust-susceptible wheat variety, Little Club (LC), were raised in 12-cm diameter porcelain pots for the propagation of strains. Infected samples were removed from the refrigerator (4°C), and single pustules from the wheat stem rust samples were isolated in order to inoculate leaves of LC collected from seven-day-old seedlings or seedlings with fully expanded primary and secondary leaves. We followed the procedure of Huang [8]. LC leaves were cut into small pieces (6-7 cm long) and were placed between two pieces of filter paper kept in 15-cm petri dishes. The two ends of each leaf segment were pressed with glass (length 120 mm, width 10 mm, thickness 3 mm) and 40 mg·L⁻¹ 6-benzylaminopurine (BA) was added as a preservative solution to soak the filters. The uredia were scraped from the rustinfected stem samples using flat toothpicks and were then sprayed on the abaxial surface of the lamina. Each leaf was inoculated at two spots by daubing gently, preventing scratching of the foliage. Thereafter, the inoculated leaves were moistened with 0.05% Tween 20 using an atomizer, and were placed in an incubation chamber for 20 h in the dark at 16–20°C. Subsequently, they were transferred from the dew chamber to glass compartments in an artificial culture room, where the conditions were controlled with a 14 h:10 h (light:dark) photoperiod and temperature of $18-22 \pm 1$ °C. When the uredinia (diameter of 0.5–0.7 mm) appeared after 5–6



Fig 1. Regions for the detection and investigation of Pgt in 2013–2014. Note: ▲: HLJ: Heilongjiang, LN: Liaoning, HB: Hebei, HN: Henan, HB: Hubei, CQ: Chongqing, GS: Gansu, SC: Sichuan, YN: Yunnan. ●: KS: Keshan, SY: Shenyang, BD: Baoding, WD: Wendeng, HZ: Hanzhong, YL: Yangling, GG: Gangu, QY: Qingyang, WX: Wenxian, HS: Heishui, YA: Yaan, YB: Yibing, JG: Jiange, KX: Kaixian, FD: Fengdu, TN: Tongnan, YX: Yuxi, MD: Midu, KM: Kunming, ST: Shaotong, DH: Dehong, XY: Xingyi, BJ: Bijie, XF: Xiangfan, XZ: Xuzhou. ■: 1: Southeastern Gansu Province 2: Southwestern Shannxi Province 3: Northeastern Sichuan Province 4: Northwestern Yunnan Province.

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days, single pustules were isolated and used for re-inoculation. The spore multiplication procedure was repeated 2–3 times until sufficient numbers of spores were produced for experimentation. Twenty-three and 30 single pustules were identified from 11 samples obtained from wheat and 22 samples obtained from *Berberis*, respectively. All materials used in the experimental set-up had been sterilized.

Inoculation of differential lines and designation of races

Differential wheat were planted in 12-cm diameter porcelain pots [9]. Each pot contained four genotypes (5–6 seeds per genotype) separated from each other. The susceptible wheat variety, LC, was used as the control to ascertain the viability of the spores. The leaves of seven-day-old seedlings were moistened with 0.05% Tween 20 using an atomizer. Thereafter, purified single-pustule isolates were sprayed/inoculated evenly onto the back of the leaves with flat toothpicks and then moistened again with 0.05% Tween 20. After inoculation, the plants were placed in a dew chamber with 16–20 h of darkness at 18-22°C. Then, they were transferred to a greenhouse at a temperature of $18-22 \pm 1$ °C and relative humidity (RH) of 65–75%. The infections were assessed 14 days after inoculation using the 0–4 Infection Types (ITs) scale, as described by Stakman et al. [10]. The infections were grouped into two types, where values of 0, 1, 1+, 2, and 2+ were considered low infection types (indicating resistance) and values of 3-, 3, 3+, and 4 were considered high infection types (indicating susceptibility). Race names were designated according to the methods described by Li et al. [9]. In this study, the isolates were purified and assessed in three replicates to obtain uniform data regarding infection type.

Determination of virulence frequency of races

Forty-six single *Sr* gene lines were used to test the virulence frequency of the races, including 20 North American single genes (*Sr5*, *Sr21*, *Sr9e*, *Sr7b*, *Sr11*, *Sr6*, *Sr8a*, *Sr9g*, *Sr36*, *Sr9b*, *Sr30*,

Sr17, *Sr9a*, *Sr9d*, *Sr10*, *SrTmp*, *Sr24*, *Sr31*, *Sr38*, and *SrMcN*) and 26 additional single genes provided by the Minnesota Cereal Disease Laboratory (*Sr9f*, *Sr12*, *Sr13*, *Sr14*, *Sr15*, *Sr16*, *Sr18*, *Sr19*, *Sr20*, *Sr22*, *Sr23*, *Sr25*, *Sr26*, *Sr27*, *Sr28*, *Sr29*, *Sr32*, *Sr33*, *Sr34*, *Sr35*, *Sr37*, *Sr39*, *Sr47*, *SrTt3*, *SrGT*, and *SrWld-1*). The inoculation and assessment methods followed the procedures mentioned earlier.

Results

Occurrence and frequency of Pgt races

From the twenty-three single pustules obtained from the 11 samples, we identified six races, 21C3CTHTM, 21C3CTQSC, 21C3CTTSC, 21C3CTTTM, 34C0MKGQM, and 34C0MRGSM that can be included in the race groups, 21C3 and 34C0. Races 21C3CTQSC, 21C3CTTSC, 21C3CTTTM, and 34C0MRGSM are new physiological races. To the best of our know-ledge, this is the first report of the virulence of 34C0MRGSM to *Sr5* + *Sr11* in China. Race 21C3CTTTM was virulent to the 20 North American single-gene differentials, except *Sr5*, *Sr9e*, *Sr31*, and *Sr38* (Table 1). Thirty single pustules were identified from the 22 samples obtained from *Berberis*. Three race groups, 34C0, 34C6, and 34C3, including seven physiological races, were identified. Six of the seven races are novel and virulent against *Sr5* + *Sr11*, namely 34C0MRGQM, 34C6MTGSM, 34C6MRGQM, 34C3MTGQM, 34C3MKGQM, and 34C3MKGSM, whereas the other race (34C0MTGSM) has been previously identified.

Among the 13 races identified from the 53 isolates, the most predominant races were 34C3MTGQM and 34C6MRGQM, each with a frequency of 13.2%. The next predominant race was 34C0MRGQM, with a frequency of 11.3%. (Table 2).

Virulence of Pgt towards 46 single-gene differentials

The virulence frequencies of the 53 isolates against *Sr6*, *Sr7b*, *Sr9a*, *Sr9b*, *Sr9d*, *Sr9g*, and *SrMcN* were 100%, and those against *Sr5*, *Sr8a*, *Sr9f*, *Sr12*, *Sr13*, *Sr15*, *Sr16*, *Sr18*, *Sr20*, *Sr27*, *Sr28*, *Sr29*, *Sr32*, and *Sr39* were also high, with frequencies of 58.5–96.2%. However, the virulence frequencies toward *Sr10*, *Sr14*, *SrGT*, *Sr25*, *Sr19*, *Sr22*, *Sr23*, and *SrWld-1* were relatively low, with frequencies of 49.1, 47.2, 45.3, 43.4, 41.5, 37.7, 35.8, and 33.9%, respectively (Table 3).

However, the resistance genes, *Sr9e*, *Sr21*, *Sr26*, *Sr31*, *Sr33*, *Sr38*, *Sr47*, and *SrTt3* were effective against all of the 53 isolates tested. Genes *Sr35*, *SrTmp*, *Sr30*, *Sr37*, *Sr17*, and *Sr36* were effective against most of the isolates tested, with a resistance frequency of 92.5, 86.8, 84.9, 84.9, 79.3, and 77.4%, respectively.

The virulence information of 13 physiological races against 46 genes revealed that the races, 21C3CTTTM, 21C3CTTSC, and 21C3CTQSM, included in the 21C3 race group and the race, MKGSM, in the 34C3 race group have broader spectra than do the other races. Race 21C3CTTTM and 34C3MKGSM have the broadest spectra and rendered 34 of 46 stem rust resistant genes ineffective. Race 21C3CTTSC and 21C3CTQSM have second broadest spectra, which demonstrated virulence against 31 of 46 stem rust resistance genes (Table 4).

Discussion

Due to the potential threat of new emerging races of *Pgt* such as Ug99 and their potential threat to global wheat production, this study aimed to identifying and characterising *Pgt* races in China from 2013–2014. The results revealed race groups that had been documented earlier, such as 34C1 and 34C2, and rare race groups (e.g., 34C4 and 116), all of which were not detected in the present study. In the 2013–2014 study, race groups 21C3 and 34C3 were predominant in China [11, 12]. Previous studies suggested that this observation was related to the

Differentials	Sr genes	R1	R2	R3	R4	R5	R6	R 7	R8	R9	R10	R11	R12	R13 ^a
ISr5-Ra	5	1 ^b	0	1	0	4	4	4	4	4	3	4	3+	4
CnS_T_mono_deriv	21	2	1+	1-	1+	2	;1 =	1+	2	2	1	2	1+	1+
Vernstine	9e	1-	;	;1 =	1-	1	;	;1-	1	;1	;1	1+	1	1
ISr7b-Ra	7b	4	4	4	4	4	4	4	4	4	3	4	4	4
ISr11-Ra	11	3	4	3	4	0	4	4	4	4	3	1	;	4
ISr6-Ra	6	3	4	4	4	4	4	4	4	4	3	3	4	3
ISr8a-Ra	8a	4	3	4	4	4	1	1+	3	2	3	3	4	4
CnSr9g	9g	4	4	4	4	4	4	4	4	4	4	4	4	4
W2691SrTt-1	36	0	4	4	4	0	0	0	1	1	0	0	0	1
W2691Sr9b	9b	4	4	3	4	3	3	4	4	4	4	4	4	4
BtSr30Wst	30	1	2	4	4	1	;	1	1	2	1	1	;	1
Combination VII	17+13	4	2	3	4	0	0	0	;	0	;	0	;	;
ISr9a-Ra	9a	4	4	3	3	4	4	4	4	4	4	4	4	4
ISr9d-Ra	9d	4	4	4	4	4	4	4	4	4	4	4	4	4
W2691Sr10	10	4	4	4	4	1+N	1+N	3	4	1+N	3	;	3C	;
CnsSrTmp	Ттр	3	1C	1-	3	0	1N	1+	;	1N	0	0	;	;
LcSr24Ag	24	3C	3	;1 =	3	3	3	4	4	4	4	4	3	4
Sr31/6*LMPG	31	1C	;1-	1-	1	1	;	1	1	1	1	1	1	1
Trident	38	;	0	1N	;	1	;	;1-	;	1N	;1-	;	;	;1-
McNair 701	McN	4	4	4	4	4	4	4	4	4	4	4	4	4
Reliance	5, 16, 18,20	0	0	0	0	4	4	4	4	4	4	4	4	3
Vernal	9e	;1 =	;1 =	;	;1	;1-	;	;1	1	1	1	1	;1	1
Einkorn	21	1	;	;1 =	1	;1 =	;	;1-	1	;1	1	;1-	;1-	;1
Mianzi 52	17, +? ^C	4	4	4	4	;	0	0	;	0	0	0	0	;
Mini 2761	5, +?	0	0	0	0	;	0	0	0	0	0	0	0	;
Orofen	5, 6, +?	0	0	0	;	0	;	1	;	2	2	3	4	4
Rulofen	6, 30, +?	0	;	0	0	0	0	0	0	0	0	0	0	0
Little Club		4	4	4	4	4	4	4	4	4	4	4	4	4

Table 1. Infection types of identified P. graminis f. sp. tritici races collected from 2013-2014 in China toward differential wheat cultivars.

^a R1: 21C3CTHTM; R2: 21C3CTQSM; R3:21C3CTTSC; R4:21C3CTTTM; R5: 34C0MKGQM; R6: 34C0MRGQM; R7:34C0MRGSM

R8: 34C0MTGSM; R9: 34C6MRGQM; R10: 34C6MTGSM; R11: 34C3MKGQM; R12:34C3MKGSM; R13: 34C3MTGQM

^b Infection types (ITs): are based on a 0-to-4 scale where ITs of 0, 1, or 2 are indicative of a resistant (low) response and ITs of 3 or 4 are indicative of a susceptible (high) response; Symbols + and-indicate slightly larger and smaller pustule sizes, respectively; N was used for cases with significant necrosis [10]. ^C Symbol '+?' means the cultivar carry other unknown resistance gene.

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strong competitiveness of this pathogen [13, 14]. Its wide spectrum of virulence may be another reason [11]. This was confirmed in our study by testing the virulence of 53 different strains against 46 resistance genes. For instance, race CTTTM, which has the widest virulence spectrum in group 21C3, was virulent to 33 of the 46 resistance genes, including *Sr17*, *Sr24*, *SrTmp*, and *Sr36*. Only 15 resistance genes, specifically *Sr5*, *Sr9e*, *Sr14*, *Sr19*, *Sr22*, *Sr23*, *Sr26*, *Sr27*, *Sr31*, *Sr35*, *Sr37*, *Sr38*, *Sr47*, *SrGT*, and *SrTt3*, were found to be effective against CTTTM. Race group 21C3 was dominant, occurring with high frequency, and significant changes in its population structure have occurred. For example, races CTHTM, CTQSM, CTTSC, and CTTTM are new to the 21C3 group.

Berberis is the alternate host of *Pgt* and plays a vital role throughout the sexual life cycle of *Pgt* [15]. Sexual reproduction of *Pgt* is completed on the leaves of *Berberis*, resulting in the diversity of *Pgt* races [16, 17]. Since the 1950s, wheat stem rust has been effectively controlled



Race	Host	Province (No. of isolates)	Frequency (%)
21C3CTHTM	Wheat	Liaoning (3)	5.7
21C3CTQSM	Wheat	Guizhou (1); Yunnan (3)	7.5
21C3CTTSC	Wheat	Guizhou (4)	7.5
21C3CTTTM	Wheat	Guizhou (2); Hubei (2)	7.5
34C0MRGSM	Wheat	Sichuan (1); Hubei (1)	3.8
34C0MKGQM	Wheat, Barberry	Sichuan (1); Heilongjiang (1)	3.8
34C0MRGQM	Wheat, Barberry	Heilongjiang (5); Gansu (1)	11.3
34C0MTGSM	Barberry	Gansu (3)	5.7
34C6MRGQM	Barberry	Shaanxi (1); Gansu (6)	13.2
34C6MTGSM	Barberry	Gansu (1)	1.9
34C3MKGQM	Barberry	Shaanxi (2); Gansu (4)	9.4
34C3MKGSM Barberry		Shaanxi (1); Gansu (3)	9.4
34C3MTGQM Barberry		Shaanxi (2); Gansu (5)	13.2

Table 2. Races of P. gramins f. sp. tritici in China from 2013-2014.

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by using highly resistant winter/spring wheat cultivars and eradicating the common barberry in the United States [18, 19], whereas in China, insufficient research was carried out to determine whether sexual reproduction could lead to diversity of the *Pgt* population. However, after *Berberis* was shown to be the alternate host of *P. striiformis* f. sp. *tritici* (*Pst*) in 2010 [16], this plant Genus was found to be distributed widely in China, and it is infected by *Pst* under natural conditions. Researchers believed that the diversity of *Pst* would be contributed through the

Table 3.	Virulence fre	quencies of r	aces of P.	graminis f. sp	. tritici collected	from China	during 2013-2014.
							0

Sr gene	No. of virulent isolates	Virulence frequency (%)	Sr gene	No. of virulent isolates	Virulence frequency (%)
5	38	71.7	23	19	35.8
6	53	100	24	45	84.9
7b	53	100	25	23	43.4
8a	38	71.7	26	0	0
9a	53	100	27	41	77.4
9b	53	100	28	43	81.1
9d	53	100	29	39	73.6
9f	49	92.5	30	8	15.1
9g	53	100	31	0	0
9е	0	0	32	43	81.1
10	26	49.1	33	0	0
11	41	77.4	34	31	58.5
12	44	84.9	35	4	7.5
13	33	62.3	36	12	22.6
14	24	47.2	37	8	15.1
15	43	81.1	38	0	0
16	51	96.2	39	45	84.9
17	11 ^a	20.7	47	0	0
18	37	69.8	Tt3	0	0
19	22	41.5	Ттр	7	13.2
20	41	77.3	GT	24	45.3
21	0	0	Wld-1	18	33.9
22	20	37.7	McN	53	100

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Race	Effective Sr genes	Ineffective Sr genes
21C3CTHTM	5, 9e, 14, 19, 21, 22, 23, 25, 26, 28, 29, 30, 31, 33, 35, 36, 37, 38, 47, Tt3	6, 7b, 8a, 9g, 9a, 9b, 9d, 9f, 9g, 10, 11, 12, 13, 15, 16, 17, 18, 20, 24, 27, 32, 34, 39, Tmp, GT, Wld-1, McN
21C3CTQSM	5, 9e, 17, 19, 20, 21, 26, 27, 30, 31, 33, 34, 35, 37, 38, 39, 47, Tmp, Tt3	6, 7b, 8a, 9a, 9b, 9d, 9f, 9g, 10, 11, 12, 13, 14, 15, 16, 18, 22, 23, 24, 25, 28, 29, 32, 36, GT, Wld-1, McN
21C3CTTSC	5, 9e, 19, 20, 21, 22, 24, 26, 27, 29, 31, 32, 33, 37, 38, 47, Tmp, Tt3,	6, 7b, 8a, 9a, 9b, 9d, 9f, 9g, 10, 11, 12, 13, 14, 15, 16, 17, 18, 23, 25, 28, 30, 34, 35, 36, 39, GT, Wld-1, McN
21C3CTTTM	5, 9e, 14, 19, 21, 22, 23, 26, 27, 31, 33, 35, 37, 38, 39, 47, GT, Tt3	6, 7b, 8a, 9a, 9b, 9d, 9f, 9g, 10, 11, 12, 13, 15, 16, 17, 18, 20, 24, 25, 28, 29, 30, 32, 34, 36, Tmp, Wld-1, McN
34C0MKGQM	9e, 10, 11, 13, 14, 17, 18, 19, 20, 21, 23, 25, 26, 30, 31, 32, 33, 34, 35, 36, 37, 38, 47, Tmp, Tt3, Wld-1	5, 6, 7b, 8a, 9a, 9b, 9d, 9f, 9g, 12, 15, 16, 22, 24, 27, 28, 29, 39, GT, McN
34C0MRGQM	8a, 9e, 10, 13, 14, 17, 19, 21, 22, 23, 26, 27, 29, 30, 31, 33, 35, 36, 37, 38, 47, Tmp, Tt3, GT, Wld-1	5, 6, 7b, 9a, 9b, 9d, 9f, 9g, 11, 12, 15, 16, 18, 20, 24, 25, 28, 32, 34, 39, McN
34C0MRGSM	8a, 9e, 17, 19, 21, 22, 23, 26, 30, 31, 32, 33, 35, 36, 37, 38, 47, Tmp, Tt3	5, 6, 7b, 9a, 9b, 9d, 9f, 9g, 10, 11, 12, 13, 14, 15, 16, 18, 20, 24, 25, 27, 28, 29, 34, 39, GT, Wld-1, McN
34C0MTGSM	9e, 17, 21, 26, 30, 31, 33, 35, 36, 37, 38, 47, GT, Tmp, Tt3, Wld-1	5, 6, 7b, 8a, 9a, 9b, 9d, 9f, 9g, 10, 11, 12, 13, 14, 15, 16, 18, 19, 20, 22, 23, 24, 25, 27, 28, 29, 32, 34, 39, McN
34C6MTGSM	9e, 12, 17, 19, 21, 22, 23, 26, 27, 30, 31, 33, 34, 35, 36, 38, 47, GT, Tmp, Tt3, Wld-1	5, 6, 7b, 8a, 9a, 9b, 9d, 9f, 9g, 10, 11, 13, 14, 15, 16, 18, 20, 24, 25, 28, 29, 32, 37, 39, McN
34C6MRGQM	8a, 9e, 10, 14, 17, 21, 23, 26, 30, 31, 32, 33, 35, 36, 38, 47, Tmp, Tt3, Wld-1	5, 6, 7b, 9a, 9b, 9d, 9f, 9g, 11, 12, 13, 15, 16, 18, 19, 20, 22, 24, 25, 27, 28, 29, 34, 37, 39, GT, McN
34C3MKGQM	9e, 10, 11, 13, 14, 15, 17, 18, 20, 21, 22, 25, 26, 30, 31, 33, 34, 35, 36, 37, 38, 47, GT, Tmp, Tt3, Wld-1	5, 6, 7b, 8a, 9a, 9b, 9d, 9f, 9g, 12, 16, 19, 23, 24, 27, 28, 29, 32, 39, McN
34C3MKGSM	9e, 11, 17, 21, 26, 30, 31, 33, 35, 36, 37, 38, Tmp, Tt3, 47,	5, 6, 7b, 8a, 9a, 9b, 9d, 9f, 9g, 10, 12, 13, 14, 15, 16, 18, 19, 20, 22, 23, 24, 25, 27, 28, 29, 32, 34, 39, GT, Wld-1, McN
34C3MTGQM	9e, 10, 15, 17, 19, 21, 25, 26, 27, 29, 30, 31, 33, 35, 36, 37, 38, 47, Tmp, Tt3, GT, dp-2, Wld-1	5, 6, 7b, 8a, 9a, 9b, 9d, 9f, 9g, 11, 12, 13, 14, 16, 18, 20, 22, 23, 24, 28, 32, 34, 39, McN

Table 4. Effective/ ineffective Sr genes against 13 races of P. graminis f. sp. tritici.

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sexual cycle of the fungus on Berberis [20]. Our results are consistent with this hypothesis; for example, the races identified from isolates that were collected from aecia are all new races, except 34C0MKGQM. Race group 34C3 was first discovered in 1976 and threatened the Orofen wheat cultivar and its progeny planted in more than million hectares. This caused great fear, but fortunately, owing to a series of timely emergency measures taken by researchers, the occurrence of race group 34C3 remained low from 1983 to 1987, and it has not been detected since 1987 [8]. In 1990, three races, 34C3MKGQM, 34C3MKGSM, and 34C3MTGQM, were identified for the first time in China by using the North American stem rust nomenclature system of *Pgt* [21]. Race MKGSM, having the widest virulence spectrum, possessed virulence to the remaining 30 resistance genes, except *Sr9e*, *Sr11*, *Sr17*, *Sr26*, *Sr30*, *Sr31*, *Sr35*, *Sr36*, *Sr37*, *Sr38*, *Sr47*, *SrTmp*, and *SrTt3*. In addition, MKGSM and MTGQM had wide virulence spectra.

In our study, it was evident that the majority of the resistance genes were ineffective against most of the isolates according to the information and frequency of virulence. Among the 46 single-genes, *Sr9e*, *Sr21*, *Sr26*, *Sr31*, *Sr38*, *Sr47*, and *SrTt3* were effective against all of the isolates and these 7 currently effective resistance genes should be used in breeding programs to ensure that Chinese wheat cultivars are resistant to the new races, while *Sr17*, *Sr30*, *Sr33*, *Sr35*, *Sr36*, *Sr37*, and *SrTmp* were effective against most of the isolates. Similar results were reported by

Han et al. and Yao et al. [11, 22]. However, except for the 14 resistance genes mentioned directly above, the resistance genes tested was ineffective against one or more isolates. More than 58.5% of the isolates identified were virulent to *Sr5*, *Sr6*, *Sr7b*, *Sr13*, *Sr24*, *Sr32*, and *Sr34*. However, these resistance genes exist widely in wheat varieties of China [23, 24]. Notably, the MR races (e.g., 34C0MRGSM) and MT races (e.g., 34C6MTGSM) were first discovered in China. They are virulent against Sr5 + Sr11, which are two of the most commonly used resistance genes. This is a threat to wheat production [25]; therefore, urgent attention is necessary to address this situation.

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