



A Cold Plasma Technology for Ensuring the Microbiological Safety and Quality of Foods

Ozioma Forstinus Nwabor¹ · Helen Onyeaka² · Taghi Miri² · Kechrist Obileke³ · Christian Anumudu² · Abarasi Hart⁴

Received: 20 January 2022 / Accepted: 7 May 2022
© The Author(s) 2022

Abstract

Changing consumers' taste for chemical and thermally processed food and preference for perceived healthier minimally processed alternatives is a challenge to food industry. At present, several technologies have found usefulness as choice methods for ensuring that processed food remains unaltered while guaranteeing maximum safety and protection of consumers. However, the effectiveness of most green technology is limited due to the formation of resistant spores by certain foodborne microorganisms and the production of toxins. Cold plasma, a recent technology, has shown commendable superiority at both spore inactivation and enzymes and toxin deactivation. However, the exact mechanism behind the efficiency of cold plasma has remained unclear. In order to further optimize and apply cold plasma treatment in food processing, it is crucial to understand these mechanisms and possible factors that might limit or enhance their effectiveness and outcomes. As a novel non-thermal technology, cold plasma has emerged as a means to ensure the microbiological safety of food. Furthermore, this review presents the different design configurations for cold plasma applications, analysis the mechanisms of microbial spore and biofilm inactivation, and examines the impact of cold plasma on food compositional, organoleptic, and nutritional quality.

Keywords Cold plasma · Food safety · Shelf-life · Microbial spores · Food quality

Introduction

Food preservation is an age-long science that has continuously evolved. Ancient people employed physical means, including sun drying, roasting, smoking, fermentation, and salting, to preserve farm produce. With the advent of science and technology, chemicals with antimicrobial and antioxidant properties were adopted as ideal food preservatives. However, contemporary consumers' taste, guided by scientific revelations on the adverse health impact of most

food preservatives, has resulted in a craving for minimal preserved food, considered healthy and safe. Synthetic preservatives including butylated hydroxytoluene, butylated hydroxyanisole, sorbic acid, propyl gallate, and sodium nitrate have been correlated with cytotoxicity, suppression of immune response, and genotoxicity [1–3]. Thus, to accommodate the trending consumer's taste, and ensure consumers' safety, several advances in food science and technology have emerged as an alternative, effective, and safe approach for preserving food products and agricultural produce [4–6]. Also, the use of multiple combinations of food preservation techniques is suggested to enhance the safety of food products while ensuring extended product shelf-life [7, 8]. Unfortunately, high demands are placed on the ideal preservation technique, with several lofty expectations. The acceptance of preservative technique depends on factors ranging from health effects, cost and energy demands, duration of the shelf-life extension, ability to inactivate foodborne pathogenic and spoilage vegetative microorganisms and spores, effects on food nutritional and organoleptic properties, timing, and adaptability. Food preservation is a sacrosanct component of the food production chain that promotes food

✉ Helen Onyeaka
h.onyeaka@bham.ac.uk

¹ Division of Infectious Diseases, Department of Internal Medicine, Faculty of Medicine, Prince of Songkla University, Hat Yai, Songkhla 90112, Thailand

² School of Chemical Engineering, University of Birmingham, Edgbaston B15 2TT, UK

³ Renewable and Sustainable Energy, University of Fort Hare, Alice 5700, Eastern Cape, South Africa

⁴ Department of Chemical and Biological Engineering, The University of Sheffield, Sheffield S1 3JD, UK

security. Current estimates suggest that one-third of agricultural produce are lost to food spoilage, with 25% of the total food loss attributed to microbial spoilage [9]. Microbial-mediated food deterioration is likely the most frequent cause of food loss and wastage of agricultural produce. The lack of a proper preservative/storage facility contributes significantly to the global food security crisis. Hence, proper storage and food preservation are key agricultural goals to ensure food security and availability. Although gaint progressive strides have been made in the science of food preservation and processing, the goal of achieving a natural, unaltered food with maximum safety remains elusive. Microbial hazards including food spoilage and foodborne diseases and food contamination demand greater investments towards ensuring consumers' safety.

Food safety is a critical global issue that requires the concerted efforts of both food producers and consumers. The spread of pathogenic microorganisms, microbial toxins, and other contaminants through food and drinks is a major disease and infection source. Mortality and morbidity due to foodborne diseases and intoxication are the major sources of economic loss. Although the primary aim of preservation is the extension of food shelf-life, by extension, preservation ensures the safety of food products through the controls of pathogenic foodborne microorganisms. Various chemical food preservatives possess antimicrobial properties that inhibit the proliferation of spoilage and pathogenic microorganisms.

Similarly, most evolving green technologies employed to preserve food inhibits microbial growth and, at increased intensity, inactivates microbial cells. Although most preservation and storage techniques such as pasteurization and freezing can effectively inhibit or inactivate foodborne microorganisms, the presence of microbial spores in food constitutes a significant risk to food safety. Under preservation treatments and unfavorable storage conditions, certain pathogenic foodborne bacteria belonging to the genus *Clostridium* and *Bacillus* adopt the inactive spore forms to survive. The subsequent germination of spores into vegetative cells leads to food contamination, especially of finished products. Microbial spores are recalcitrant to food processing and preservation treatments and are frequently associated with contaminated of finished processed products. Effective inactivation of microbial spores is a fundamental requirement for an ideal food preservation regimen. Several emerging technologies, although capable of effectively inactivating microbial vegetative cells, fail to eliminate spores. Microbial spores survive treatments such as thawing, heating [10], freezing, and UV radiations [11–13]. The presence of microbial spores in food products is a major cause of foodborne disease and intoxication. Thus, several novel technologies have been proposed to address challenges posed by the spore forming pathogenic foodborne microorganisms. Some

of these methods have been reported to inactivate microbial spores in food, food processing facilities, and contact surfaces. Treatment of food with pulse electric field [14, 15], plasma technology [16, 17], ultrasonication [18–20], and high hydrostatic pressure [21] alone or in combinations demonstrated promising results for spore inactivation in food products.

Moreover, cold plasma must be evaluated critically as an emerging food processing alternative before being accepted by the food industry. In 2018, Pankaj et al. [22] published a review of cold plasma technology that discussed the negative effects and limitations. The overview did not explain how spore-forming microbes and spores are inactivated. Recently, Laroque et al. [23] published another review that describes in detail the most promising cold plasma sources and operation modes, the influence of operating conditions on plasma properties, and the efficacy of this technology in food processing. This review looked at the design of cold plasma processes that inactivate target microorganisms with minimal impact on food quality. The previous review papers examined the impact of cold plasma species on food components at molecular level and design methods to minimize the impact of the interactions. A review on the effects of cold plasma on the chemical structure of different food components and their impact on food attributes has been reported by Saremnezhad et al. [24]. But this review aims to highlight cold plasma technique as a recent ideal green preservation technique by comprehensively elucidating its benefits/advantages, superior attributes at spore inactivation, mechanism of microbial inactivation, and factors that should be considered to ensure optimal outcomes. Conversely, the formation of biofilm on food contact surfaces is a critical safety concern in the food industry, since it is a potential source of contamination; for this reason, the most distinctive contribution of this review focuses on exploring how biofilm can be disrupted through plasma treatment. The objective is to provide food researchers, technologist/engineers, and industrialist with additional knowledge that could help promote and ensure effective understanding and application of cold plasma technique in food preservation and for enhancement of food safety.

Plasma Technology

In the food industry, several green technologies have been adopted to rid production lines and products of microorganisms that could compromise product quality, resulting in recalls and foodborne illness outbreaks. This has led to an upsurge on the topic of applications of green technologies in preservation and shelf-life extension. Over the last decade, the use of plasma has gained extensive applicability in the food industry as a relatively new and promising

non-thermal decontamination technology. The technology is not limited to the food industry alone but extends to other sectors, including the surface decontamination of medical devices and environments [25–28], sanitization of heat-sensitive biological and chemical agents [29], environmental degradation of waste and toxic residues [30], and in the textile industry [31–33]. Its technique involves the application of plasma such as ionized or partially ionized gases to inactivate food contaminants, including microbial cells, enzymes, and toxins. The plasma is created either by sufficiently heating gas in an enclosed chamber under deep vacuum or by using radiofrequency or microwave energy to excite the gas molecules in order to produce free radicals, which are the main constituents of plasma.

Plasma has been described as the fourth state of matter (i.e., solid, liquid, gas, and plasma). It is a collection of both neutral and charged particles referred to as reactive species. In a plasma state, molecules dissociate into their atomic elements, losing electrons and acquiring a higher energy state [34]. The efficacy of plasma technology depends on factors such as energy defined as pressure, temperature, the thermodynamic equilibrium between the particles [34], and the type of gas used [35]. Though plasma induction requires sufficiently high energy, recent advancements in plasma physics have made it possible to generate “cold plasma” at ambient temperatures and atmospheric pressures. This generated cold plasma can be applied in medicine for sterilization [26], wound healing [36], and disease treatment [37, 38]; in agriculture to enhance seed germination [39–41]; and in the food industry for inactivation of vegetative and spores of foodborne pathogenic and spoilage microorganisms, enzyme inactivation, and toxin inactivation. Consequently, cold plasma has also been employed in environmental management for the degradation of contaminants such as pesticides [42] and dyes [43], and for the decontamination and treatment of wastewater [44–47]. Other areas of applicability include catalysis and material sciences for surface modification and functionalization [48–50] as well as sterilization. However, the application of cold plasma in food sterilization has revolutionized preservation technology, offering a healthy alternative with negligible effect on the food nutritional and organoleptic properties. The cold plasma is produced through partial ionization of gaseous molecules [51]. For food processing, plasma generation methods are classified as dielectric barrier discharge (DBD), plasma jet (PJ), corona discharge (CD), radiofrequency (RF), and microwave (MW). A detailed review of the specifics and applicability for each has been reported by Laroque et al. [23]. Researchers are most interested in DBD plasma generation methods because they are a safe and low-cost alternative for processing applications. Applied electric fields and spatial charge distribution are responsible for electron energy, which drives plasmas’ reactivity, which is influenced

by the chemical composition of the gas. However, plasma properties are influenced by the size, time scale, temperature, and density of the charged and reactive species [52]. In a recent review published by Laroque et al. [23], factors such as gas composition, relative humidity, and electrode and dielectric barrier including electrode and dielectric materials, electrode geometry, and shape have been discussed in relation to plasma reactive species formation and ionization, efficiency, and action.

Processing food is an important step in extending the shelf life and preserving the nutritional quality of food while reducing post-harvest losses. With cold plasma, a gaseous matter is converted into an ionized gas known as plasma (comprising electrons, ions, neutral molecules, and atoms as well as charged reactive species) when sufficient energy is applied. The application of cold plasma technology for food safety and preservation can be categorized into three types as follows: (i) direct plasma treatment which is generated at the site of application, (ii) indirect plasma treatment in which it is created at a remote location and delivered to the target, and (iii) solution treated with plasma such as plasma-activated water used as a disinfectant [53]. The configuration of different cold plasma designs food safety applications is shown in Fig. 1. As shown in Fig. 1a, plasma can be generated via dielectric barrier discharge (DBD) by dispersing current flow through dielectric materials between electrodes. In packaging where reactive oxygen and nitrogen species can be generated directly within sealed packages, the DBD is highly appropriate for inactivating microorganisms on fresh produce. Typical operation conditions are gas pressure in the range of 10^4 – 10^6 Pa and frequency 10–50 MHz. Due to its cost-effectiveness, configuration flexibility enables food treatment in-package (such as meats, poultry, fruits, and vegetables) and prevents post-contamination, and consequently, the most convenient method of plasma generation, the DBD method, is the most commonly used [23, 54]. Figure 1b illustrates the design configuration of plasma jet in which an inlet gas flows between electrodes while the outer electrode is grounded; the central electrode is driven by high-voltage power supply to produce free electrons that collide with gas molecules to produce various reactive species. The discharge plasma is utilized for the treatment of the food product. However, the application is area-limited. Corona plasma discharge is classified as atmospheric pressure cold plasma. It occurs when current flows from an electrode with a high potential into a space filled with air or other gases, ionizing it in the process and creating a region of plasma around the electrode as shown in Fig. 1c. It can be powered with high-pulsed DC or AC voltage. Unlike the other electrode configurations, the electrodes here are highly asymmetrical, designed with a thin wire or needle electrode facing flat

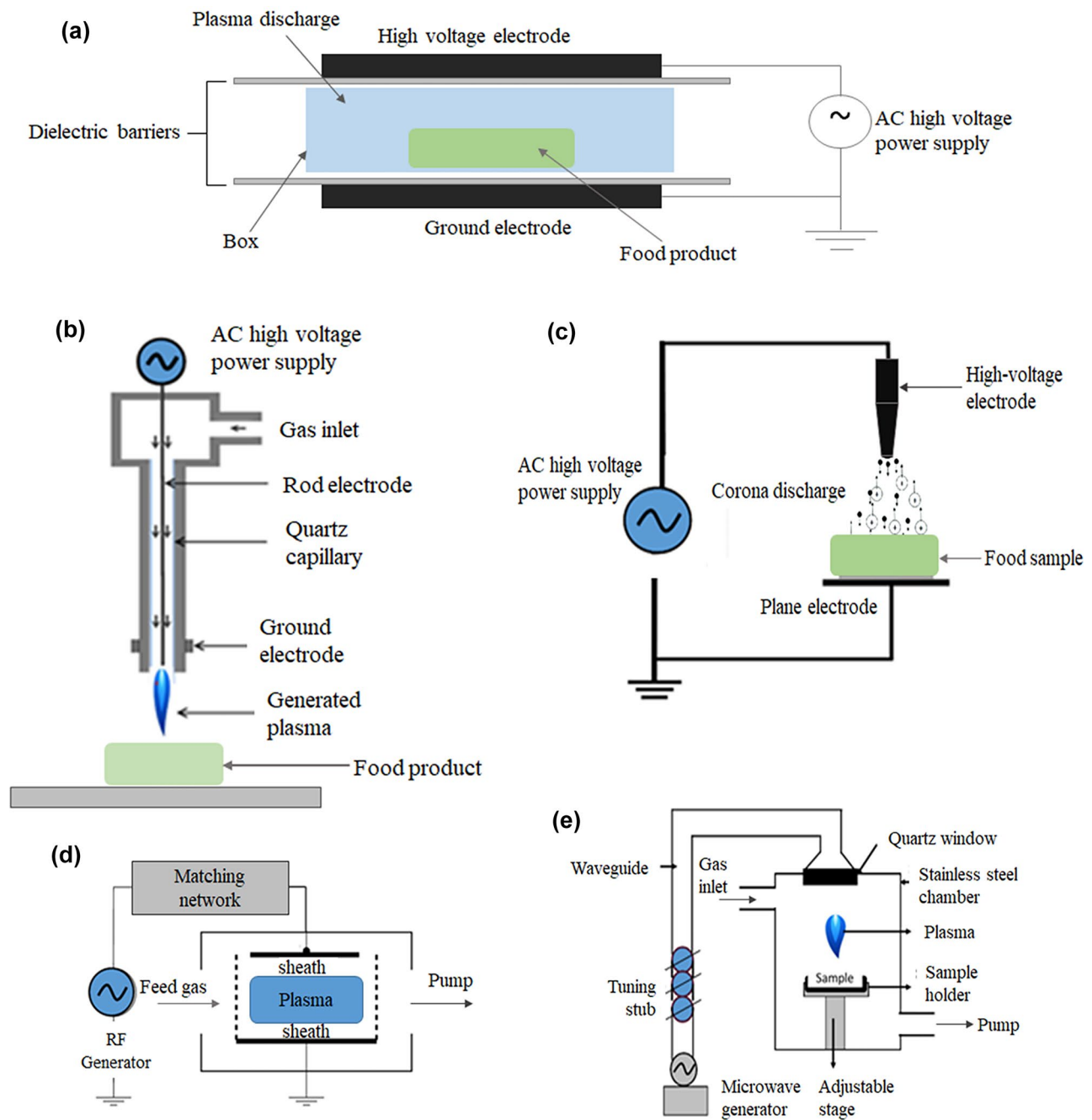


Fig. 1 Different configuration of cold plasma designs for food safety applications: **a** dielectric barrier discharge, **b** plasma jet, **c** corona discharge, **d** radiofrequency, and **e** microwave plasma system

plane or large diameter cylinder electrode. Furthermore, corona discharge arcs are commonly created by strong electric fields generated by small diameter wires, needles, or sharp-edge electrode (see Fig. 1c). Notably, the corona plasma region occurs in the immediate locality of the of the point electrode. Unlike plasma jet, the corona plasma discharge has a more extensive coverage of food sample area and also produces denser and more energetic plasma

compared to DBD [55]. However, the produced plasma is characteristically inhomogeneous, limiting its application for uniform treatment.

On the other hand, radiofrequency (RF) plasma is formed by applying a radio frequency field to a gas flow as shown in Fig. 1d. RF plasmas can be classified into capacitively coupled plasma, inductively coupled plasma and helicon wave sources. The design configuration consists of two parallel

electrodes separated by a gap of a few centimeters in a vacuum chamber [23, 54]. Methodologically, it can be operated in the range of 1–100 MHz [56]. In contrast, microwave discharge is generated by a magnetron with a typical electromagnetic wave frequency of about 2.45 GHz (Fig. 1e). By directing the electromagnetic waves into the treatment chamber, the microwaves are absorbed by the gas which heats and ionizes, causing the release of electrons. This results in ionization reactions because of inelastic collisions, releasing energy as photons of visible light and UV light. Unlike other plasma source that uses electrodes, it uses antenna switches as microwave gas discharge plasma devices which uses the high-power microwave pulse for plasma generation [57]. The purpose of this pulse is to prevent damage to the receivers of low-signal microwaves through the circuit. The microwave electromagnetic field accelerates the electrons of gas molecules to create plasma. The technique has the potential to generate both quasi-equilibrium and non-equilibrium plasma for different applications [57]. Microwave plasma configuration comprises of the following components: a power source such as magnetron, circulator, standing wave ratio meter, matching circuit/network and microwave-to-plasma applicator.

Cold Plasma Design

Three major designs of cold plasma technology for food sterilization are currently employed. These include remote treatment, direct treatment, and close proximity with an electrode. These designs are configured according to the positioning of the source that generates the plasma and the surface of the target [53]. In the remote treatment design, the target product is not placed directly in the plasma chamber. The ionized gas used for remote treatment might be air, nitrogen, or a mixture of noble gases [53]. The limitation of the remote treatment design is the generation of reactive species which might interact with other plasma species such as charged particles or photon species. However, in terms of design, the remote treatment is preferred over others because of its design simplicity, size flexibility, and target physical shape. Unlike the remote treatment, in the direct treatment, the target product is placed in direct contact with the generated plasma and often employs the plasma needle and microwave plasma tube [53]. The direct treatment results in higher exposure to UV radiation because of the closeness of the target product. This results in high moisture content due to the heat generation by conduction in the product [58]. This process alters the appearance and texture of food products such as meats and deteriorates vitamins and nutrients. In close proximity design, the food product is placed in close proximity to one of the electrodes. This type of design ensures that the target product is exposed to a higher combination of reactive charged particles, electrons that are

negatively charged, and UV radiation [59]. For effective operation of this design, products to be treated or sterilized must fit between the electrode. The system can be best used for smaller food products such as seeds, berries, nuts, and flatter objects like chicken breast [53].

Cold Plasma as an Ideal Preservation Method

Plasma processing uses cold plasma to extend the shelf-life of products and is a green, contemporary preservation method that is compatible with a range of food types. The efficacy of the technology has been demonstrated in several reported articles on a wide variety of food, including natural and processed products. Also, the application of the method to surface sanitization of in-package fruit and other food products, including vegetables, meat, cereal grains, has demonstrated promising results, with effective eradication of microorganisms, prolonging of products shelf life, reducing spoilage losses, and improving nutritional, functional, and sensory properties of food products [60]. In addition, cold plasma technology offers numerous advantages, including rapid processes, enhanced process efficiency, elimination of process steps, better quality product, and retention of product characteristics (e.g., texture, nutrition value, organoleptic properties), and improved shelf life. Table 1 highlights reported applications of cold plasma technology in different food types. It is clear that significant log reduction of spore-formers microbes as well as inactivation spores itself can be achieved with cold plasma technology. Also, the technique can be applied for a wide variety of purpose such as decontamination, inactivation of spores and spore-formers, food quality preservation, in-packaging treatment, and food self-life extension (Table 1). Consequently, it can be applied to food products in various forms: juice, vegetables and fruits, and meats. One of the major advantages of cold plasma processing over other non-thermal technologies is the evidenced effectiveness at spore inactivation [17, 61], and it also inactivates microbial toxins [17, 62, 63]. More also, co-usage of cold plasma processing with other technologies or as an adjunctive technique ensures effective deactivation of vegetative cells and microbial spores by efficiently ensuring the inactivation of microbial proteins.

Mechanism of Microbial Inactivation

The inactivation of bacteria, spores, and other microorganisms in foods and on non-food surfaces has been demonstrated through a variety of studies using atmospheric cold plasma. It has also been reported that cold plasma processing effectively inactivates a broad range of microorganisms by generating reactive species lethal to cells [87]. The process of generating plasma through the oxidation of gases produces reactive species such as reactive oxygen species

Table 1 Applications of cold plasma technology in food preservation and shelf-life extension

Food	Operation parameters	Organisms targeted	Microbial Log reduction	Technologies	Gas	Purpose	Reference
Raw chicken breast meat	60, 70, or 80 kV for 60, 180, or 300 s	Psychrophiles	> 1.0 log	Dielectric discharge	O ₂ and CO ₂	In-packaging treatment	[64]
Fresh and frozen pork	20 kV, 58 kHz, for 0, 30, 60, 90, and 120 s	<i>Escherichia coli</i> O157:H7 and <i>Listeria monocytogenes</i>	1.5 log and > 1.0 log	Corona discharge plasma jet	Air	Quality Preservation	[65]
Sprout of rapeseed (<i>Brassica napus</i> L.) seeds	20 kV, 1.5 A, and 58 kHz	<i>Bacillus cereus</i> , <i>Escherichia coli</i> , <i>Salmonella</i> spp.	1.2–2.2 log	Corona discharge plasma jet	Air	Decontamination	[66]
Ready-to-eat ham	30 kV at 3.5 kHz	<i>Listeria monocytogenes</i>	4 log CFU/cm ²	Dielectric barrier discharge	O ₂ + N ₂ + CO ₂ OR CO ₂ + N	Microbial inactivation	[67]
Cherry	(40, 60, 80 kV) at (60, 80, 100, 140 s)	TBC	3 log	Dielectric barrier discharge	Dry air	Preservation	[68]
Blueberries	-	Aerobic mesophilic total viable count, yeasts and molds	0.68–1.25 log CFU/g	Diffuse coplanar surface barrier discharge	Nitrogen gas	Microbial inhibition	[69]
Asian sea bass	230 V at 50 Hz for 5 min	<i>Psychrophilic bacteria</i> , <i>lactic acid bacteria</i> , <i>Pseudomonas</i> , <i>Enterobacteriaceae</i> , <i>Clostridium perfringens</i>	3.4 log CFU/g	Dielectric barrier discharge	Ar ₂ /O ₂	Shelf-life extension	[70]
Black peppercorns	10.3 kV and 22.1 min	<i>Bacillus tequilensis</i>	3.4 log CFU/g	Dielectric barrier discharge	Helium	Decontamination	[71]
Red pepper flakes	50–1000 W	<i>Bacillus cereus</i> and <i>Aspergillus flavus</i>	0.7 ± 0.1 and 1.4 ± 0.3 log spores cm ⁻² 1.5 ± 0.3 and 1.5 ± 0.2 log spores cm ⁻²	Microwave plasma	Helium/oxygen (99.8:0.2 v/v)	Preservation	[72]
Fresh-cut carrot	100 kV for 5 min (~250 W power)	Total aerobic mesophiles, and yeast and mold	2 log10 CFU/g	Dielectric barrier discharge	Atmospheric air	Decontamination	[73]
Ready-to-eat rocket leafy salad	6 kV, 45 kHz	Total Viable Count (TVC), <i>Pseudomonas</i> spp., lactic acid bacteria (LAB), yeasts and molds	0.57 to 1.02 log CFU/g	Dielectric barrier discharge	Atmospheric air	shelf-life extension	[74]
Powdered Spirulina algae	10–20 W	<i>Bacillus subtilis</i> spore	2 logs	Dielectric barrier discharge	Nitrogen gas	Spore inactivation and preservation	[16]
Blueberry	45 kV, 50 s	<i>Botrytis cinerea</i> spore	-	Dielectric barrier discharge	-	Preservation	[75]
Tomato juice	50 Hz, 3.8 kV, and 40 W	<i>C. albicans</i> and <i>S. cerevisiae</i> ,	2 to 7 log10 CFU/m	AC powered GlidArc reactor	Air	Shelf-life extension	[60]

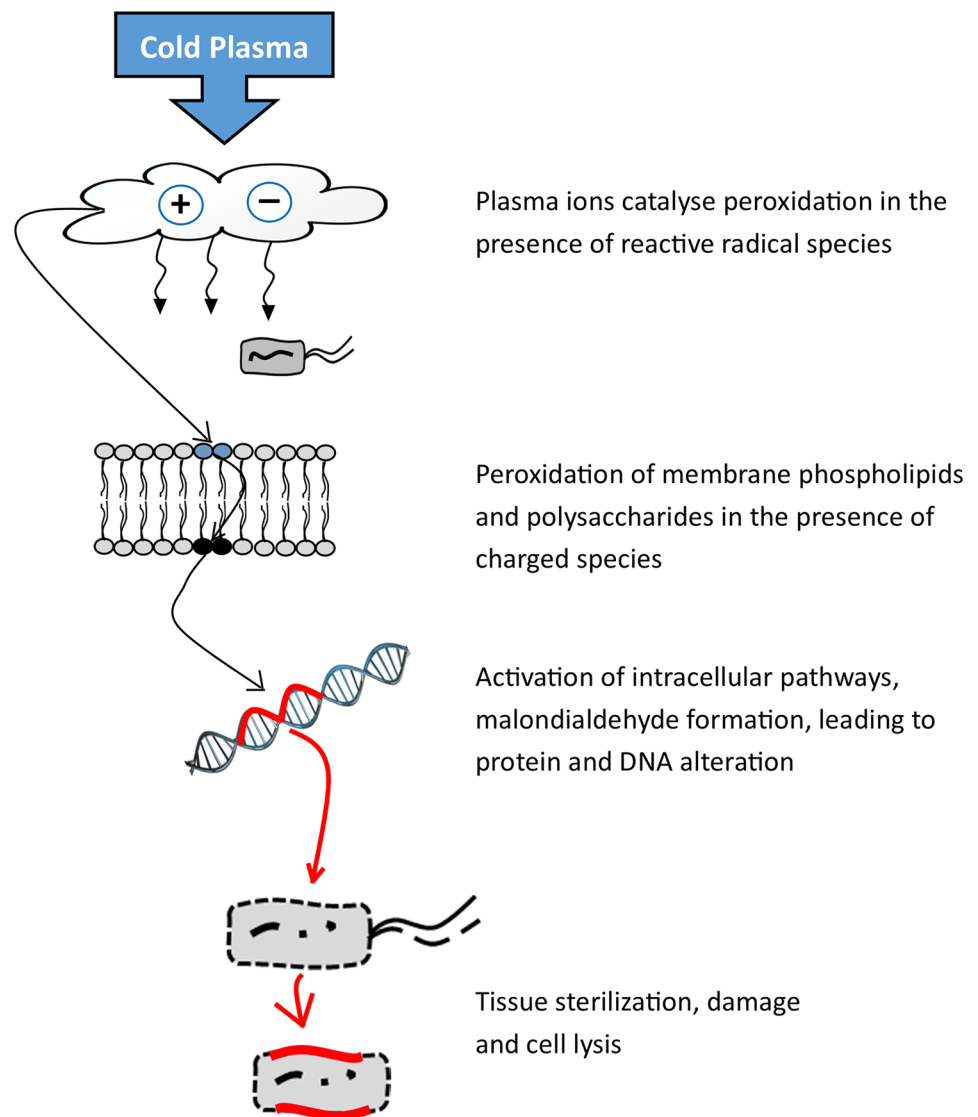
Table 1 (continued)

Food	Operation parameters	Organisms targeted	Microbial Log reduction	Technologies	Gas	Purpose	Reference
Fresh leafy vegetable	26 kV, 2500 Hz	<i>Escherichia coli</i>	5.5×103 CFU/mL	Dielectric barrier discharge	Air	Microbial inhibition	[76]
White shrimp	10 min, 500 Hz, 40 kV	<i>Staphylococcus spp.</i> , and <i>Salmonella sp.</i>	1–2 log	Dielectric barrier discharge	Atmospheric air	Preservation	[77]
Chub mackerel	60 kV and 60 s	Total viable count	1.47 and 2.7 log CFU	Dielectric barrier discharge	Atmospheric air	Preservation	[78]
Ready to eat meat (Ham)	10 kV, 2 kHz	<i>Salmonella Typhimurium</i> and <i>Listeria monocytogenes</i>	1.14 log And 1.02 log	Dielectric barrier discharge	N ₂ , CO ₂	Decontamination	[79]
Tomato	260 V at 50 Hz	<i>Escherichia coli</i>	6 log CFU mL ⁻¹	Dielectric barrier discharge	Atmospheric air	Sanitization	[80]
Fresh meat	80 kV for 60, 120 or 300 s	<i>Brochothrix thermosphacta</i>	2 Log	Dielectric barrier discharge	CO ₂ + O ₂	Shelf-life extension	[81]
Radish sprouts	2.45 GHz, 50–1000 W,	<i>Salmonella typhimurium</i>	2.6 ± 0.4 log CFU/g	Microwave generated	Nitrogen gas	Preservation	[82]
Onion powder	170 mW m ⁻² , 250 mW m ⁻²	<i>Bacillus cereus</i> , <i>Aspergillus brasiliensis</i> and <i>Escherichia coli</i> O157:H7	2.1 log spores/cm ² , 1.6 log spores/cm ² , and 1.9 CFU/cm ²	Microwave generated	Helium	Decontamination	[83]
Beef loin	9 kHz, 8.16, 8.88, 9.44 kV	<i>Staphylococcus aureus</i> , <i>Listeria monocytogenes</i> and <i>Escherichia coli</i>	> 2 log	Dielectric barrier discharge	Air	Preservation	[84]
Raw chicken breast	50 W	<i>Escherichia coli</i>	1.44 log CFU/g	Plasma jet based on cold arc plasma	N ₂ + O ₂	Decontamination	[85]
Apple juice	65 V, 1.1 MHz	<i>Citrobacter freundii</i>	5 log	Plasma jet	Argon and oxygen	Decontamination	[86]

(ROS), reactive nitrogen species (RNS), UV radiation, energetic ions, and charged particles [88, 89]. However, the antimicrobial effects of cold plasma have been reported to be primarily due to the activities of reactive species [90]. During plasma discharge, the radical species exert antimicrobial effects principally through the induction of oxidative stress, resulting in loss of cellular function and lysis of cell. The induced oxidative stress enhances cell damaged through membrane poration, lipid peroxidation, enzyme inactivation, and DNA cleavage. At the same time, the cold plasma species act on multiple sites of both bacterial and fungal cell, resulting in structural and functional alterations, and ultimately cell death. Likewise, the species cause the chemical breakdown of toxins such as aflatoxin to produce degradation products that are less toxic [51]. An investigation into

the roles of plasma species on microbial inactivation found that NO and ions were contributed minimally to the cellular effects, whereas ROS caused rapid bacterial inactivation and induced eukaryotic and prokaryotic oxidative stress [91]. Figure 2 describes the mechanism of free radical-induced oxidative stress resulting in damage and cell lysis. Other plasma byproducts such as UV radiation, H₂O₂, and electromagnetic fields acting together enhance microbial inactivation. Treatment of *C. difficile* spores with cold plasma led to ~3 log reduction in viable spore counts after 5 min [92]. The detection of reactive species responsible for cell death using optical emission spectroscopy demonstrated the presence of atomic oxygen, atomic nitrogen, hydroxyl radicals, nitrite oxide, and nitrate in light emitted from the plasma. In addition, cell membrane damage was observed using scanning

Fig. 2 Mechanism of microbial inactivation by cold plasma treatment. Adopted from Dobrynin et al. [88]



electron microscopy, SEM [93]. The SEM showed that CP caused the mycelium fold and collapse depression on *Botrytis cinerea* [75].

Effect of Cold Plasma on Microbial Spores and Toxins

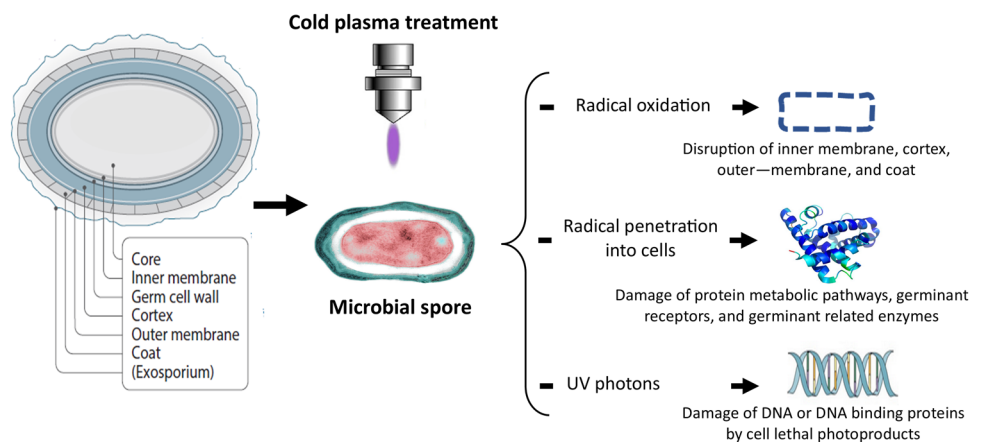
The effectiveness of cold plasma processing has been extensively investigated on both vegetative microbial cells and inactive spores. Several researchers have demonstrated the efficacy of the technology with arrays of food spoilage and foodborne pathogenic microorganisms including bacteria, and fungi. The effect of cold plasma on resistant spores of *Bacillus* spp., *Geobacillus* spp., and *Penicillium* spp., investigated on food matrix, demonstrated a 3-log₁₀ inactivation of *B. coagulans* spores after 10 s [94]. Sporocidal efficacy of cold plasma on *C. difficile* displayed similar results with ~3 log reduction in viable spore counts after 5 min of treatment [92]. An electron microscopic study demonstrated complete disintegration of fungal spore membrane due to electroporation and etching caused by the reactive plasma species [95]. *B. cereus* and *A. flavus* spores on red pepper flakes were significantly reduced by plasma treatment [72]; whereas, vegetative cells of *G. stearothermophilus* and *B. cereus* spores were effectively eliminated by exposure to cold plasma [96]. Besides the menace of food spoilage and foodborne diseases, toxins produced by certain foodborne microorganisms pose a serious health threat to consumers, resulting to foodborne intoxication. Microbial contamination of food is among the leading cause of hospitalization and death annually. It is estimated that 600 million (about 1 in 10 people in the world) fall ill after eating contaminated food. Microbial toxins mediate wide health consequences, ranging from mild enteric upset to severe and lethal outcomes. Among bacteria involved in foodborne diseases, *Salmonella* spp., *Vibrio parahaemolyticus*, *Vibrio cholerae*, *Staphylococcus aureus*, *Clostridium botulinum*, *Clostridium perfringens*, *Bacillus cereus*, and *Listeria monocytogenes* are considered toxigenic. Similarly, several mycotoxins have been

identified in contaminated food and food products, including aflatoxins, ochratoxin A, patulin, fumonisins, zearalenone, and nivalenol/deoxynivalenol. Thus, effective denaturing of microbial toxins ensures food safety. Cold plasma treatment has been evidenced to neutralize microbial toxins and inhibit the synthesis of toxins. High-performance liquid chromatography revealed inactivation of aflatoxin production by *A. parasiticus* and *A. flavus* following plasma treatment [95]. Similarly, a 50% reduction in ochratoxin A content was observed in roasted coffee samples artificially inoculated with the mycotoxigenic fungi (*Aspergillus* and *Penicillium* spp.), after 30 min of cold plasma treatment [97]. To date, the mechanisms of cold plasma-mediated spore inactivation remains unclear. However, spore shell have been suggested as the primary and main target for a plasma-induced inactivation [94]; neutral reactive oxygen species and UV radiation were also reported to play dominant role in the inactivation of spores [98]. Furthermore, an increases in hydrogen peroxide (H₂O₂) concentrations in plasma-treated cells and the increased nitrate (NO₃⁻) concentrations indicated the role of plasma generated radical ions [61]. The proposed site of action and mechanism of cold plasma inactivation of spores are presented in Fig. 3.

Effects on Nutritional and Organoleptic Qualities of Food

With the increasing demand for a fresh and healthy product and unaltered nutritional/sensory properties, food preservation entails more than just shelf-life extension. Ensuring that treatment and processes employed for preservation do not adversely affect the properties of the food is of equal significance as a shelf-life extension. Thus, emerging preservation techniques are required to meet the prerequisite of preserving or enhancing the product's organoleptic properties. Unlike thermal technologies known for altering food products' innate properties, including nutritional, physical, and textural, cold plasma technology is commended for

Fig. 3 Proposed sites and mechanism of plasma spore inactivation



protecting and preserving food properties. An investigation of the effects of cold plasma technology on the total phenolic content of black pepper grains uncovered that there were no significant differences in the total phenolic content between treated and non-treated samples ($p > 0.05$) [93]. The amino acid species of wine pickled *Bullacta exarata* did not change before and after ACP treatment; however, the contents of serine, glutamine, aspartate, glycine, threonine, and leucine were significantly increased [99]. A similar study reported the absence of significant effects on lipid oxidation, fatty acid composition, and nutritional quality of commercially packaged mackerel fillets treated at 80 kV for extended treatment times of 5 min when compared with untreated control samples [100]. Moreover, high-voltage atmospheric cold plasma treatment increased the yield of phenolic extracts from grape pomace, with high content of anthocyanins and improved antioxidant capacity [101]. In addition, microwave-combined cold plasma treatment had no effect on the antioxidant activity, color, sensory properties, and concentrations of capsaicin and dihydrocapsaicin of red pepper flakes during storage [72]. Furthermore, physiological and metabolomic analysis of cold plasma treated fresh-cut strawberries suggested that plasma treatment improved the biosynthesis of the metabolites in the flavones and flavonol biosynthesis pathway and biosynthesis of phenylpropanoid pathway without altering the textural properties of the fresh-cut strawberries. The study further revealed that plasma treatment enhanced enzyme activities and activated critical gene expression in phenylpropanoid as well as reactive oxygen species metabolism, leading to enhancement of antioxidant capacity and the accumulation of total phenolics, total flavonoid, and anthocyanin [102].

Effects of Cold Plasma Technology on Biofilm Formation on Food Contact Surfaces

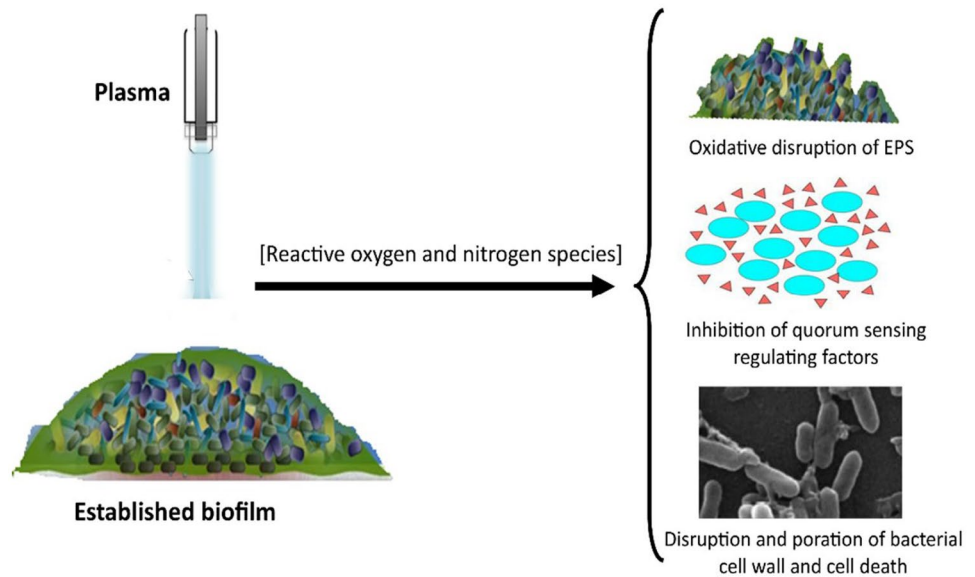
The formation of biofilm on food contact surfaces is a critical safety concern, due to the cross contamination of products that might arise from dislodged biofilm cells. Biofilm formation on contact surfaces is a major microbial survival mechanism and is often associated with resistance to food preservative treatments, sanitizers, and processing. The exopolymeric matrix of biofilms serves as a protective layer that shields the cells from the antimicrobial effects of food-grade sanitizers, antimicrobial chemicals and agents, and food treatment processes. In addition, cells within the biofilm community have been noted to synergistically resist the effects of antimicrobial compounds through the process of quorum sensing. Quorum sensing mechanism enables the community of cells to respond concertedly to changes within their environment, often resulting in the production of neutralizing enzymes, and well as toxins. Several chemical and technological methods have demonstrated excellent activity

against planktonic cells but are incapable of inactivating the sessile cells of the biofilm community. This often is associated with the additional protective layer of exopolymeric matrix which inhibits the effective penetration of the antimicrobial agent and hence alters the effective dose or concentration required to inactivate the organism. The biofilm architecture and water channels are often seen as contributors to the slowdown of convective transport and limit diffusion. The use of plasma technology have demonstrated promising effects for the inactivation of biofilms. Exposure of *Staphylococcus aureus* and *Escherichia coli* biofilms to an air-based atmospheric-pressure dielectric-barrier discharge plasma for up to 4 min caused approximately 70% and 85% disruption for *S. aureus* and *E. coli* biofilms, respectively [103]. A similar result was reported when atmospheric air plasma technology was applied to inactivate *E. coli* and *Listeria innocua* biofilms. Atmospheric air plasma damaged both the bacterial biofilm cells and its structural integrity. Scanning electron microscopy evinced the disruption of biofilms and pore formation in bacterial cells after exposure to plasma treatment [104]. The elevated reactive oxygen and nitrogen species in bacterial cells treated with atmospheric air plasma demonstrated their primary role in the observed bacterial inactivation process. Penetration of plasma species into samples depends on several factors, including the type of plasma, delivery mode, and the gas composition that makes up the plasma. Active plasma species penetrates into bacterial biofilms, reaching cells through the water channels. Figure 4 presents the proposed mechanisms of biofilm disruption by plasma treatment. A study aimed at investigating the capacity and extent of biofilm penetration by plasma demonstrated that plasma could penetrate a *Porphyromonas gingivalis* 10 days biofilms, of about 30 layers of cells and a thickness of about 15 μm , and effectively deactivate all the bacteria in the 15- μm -thick biofilms [105]. The penetration of cold plasma depends on factors such as the generating gas, intensity, voltage, and the free radical species generated. Liu et al. [106] noted a higher penetration capacity for H_2O_2 compared to nitrous/nitric acid and $\text{O}_3(\text{aq})$, even though O_3 is chemically stable with a much lower concentration than H_2O_2 in the gas phase [106]. In addition, oxidative disruption of the exopolymeric matrix by activated plasma species breakdown the protective shield, exposing the encased cells to the direct effects of plasma. Furthermore, atmospheric cold plasma technology significantly reduced *Pseudomonas aeruginosa* quorum sensing-regulated factors [107] and thus might be a relevant mechanism of biofilm disruption.

Factors Influencing the Efficacy of Cold Plasma

The efficacy of cold plasma treatment is influenced by several factors, including microbial factors, food factors, and

Fig. 4 Mechanisms of biofilm disruption by plasma treatment



plasma operational parameters. It has been demonstrated that tweaking of instrument setting parameter such as voltage, frequency, treatment time, and working gas composition alters the treatment outcome. A study aiming to investigate the effects of a range of dielectric barrier discharge high voltage atmospheric cold plasma parameters on the inactivation of *Bacillus atrophaeus* spores in a sealed package [108] observed a strong effect of process parameters on the inactivation. Direct plasma exposure for 60 s resulted in $\geq 6 \log_{10}$ cycle reduction of spores in all gas types tested. However, indirect exposure for 60 s resulted in either 2.1 or 6.3 \log_{10} cycle reduction of spores depending on gas types used. The authors noted that relative humidity was a critical factor in bacterial spore inactivation by high-voltage atmospheric cold plasma. Several factors regulate the antimicrobial effects of cold plasma treatments [109, 110]. Figure 5 highlights the factors that might affect the outcome of cold plasma treatment. The working parameters and instrumental setting of the cold plasma system are crucial to the effectiveness. An increase in microbial inactivation rates has been reported with variations in process parameters, including power, frequency, and voltage. At higher power 50 W, a higher reduction in the microbial count was achieved at a short time interval; reduction in power required an increased treatment time to achieve an equivalent reduction in microbial count [111]. Similarly, cold plasma treatment on almonds demonstrated an increased reduction in the microbial count after 30-s treatment with increasing voltage [112]. Since plasma can be generated by the excitation of various gases, the type of gas used for plasma generation is crucial to the effectiveness of plasma-mediated microbial inactivation. A comparison of air and nitrogen gas in dielectric barrier discharge plasma for the

inactivation of *Campylobacter jejuni* reported that nitrogen gas for 20 s did not yield any reduction ($p > 0.05$) in viable cell count. However, a 0.8-log reduction ($p < 0.05$) in colony-forming units (CFU) was observed when the nitrogen gas was supplemented with 2% (vol/vol) air [113]. Shi et al. [114] noted that relative humidity, gas type, and treatment time are all significant on generated ozone concentration and aflatoxin degradation. The study further reported the generation of a higher ozone concentration in MA65 than in air. In addition, high ozone concentration resulted in the effective degrading of aflatoxin in corn compared with air. Furthermore, working gas mixture and treatment time were observed to exert a strong effect on the atmospheric cold plasma inactivation of *E. coli* and *L. monocytogenes*. Working gas ratios were associated with different bactericidal efficacies. The effectiveness of atmospheric air (gas mix 1), 90% N_2 + 10% O_2 (gas mix 2) and 65% O_2 + 30% CO_2 + 5% N_2 (gas mix 3) was found to vary for individual microbial species and strain [115]. The presence of oxygen in plasma generation is critically important for increased microbial inactivation [113]. The enhanced effect demonstrated by oxygen is thought to be due to the generation of ozone (O_3), a strong oxidizing agent that is used in water treatment as a disinfectant. The dissociation of oxygen molecules at high voltages results in the production of ozone [116]. It has also been observed that treatment time plays a vital role in cold plasma-mediated microbial inhibition [117]. Food intrinsic and extrinsic factors primarily influence the shelf-life and microbial composition of the products. Similarly, factors including nature, type, moisture content, and composition are important to the effectiveness of a food preservation regimen [118].

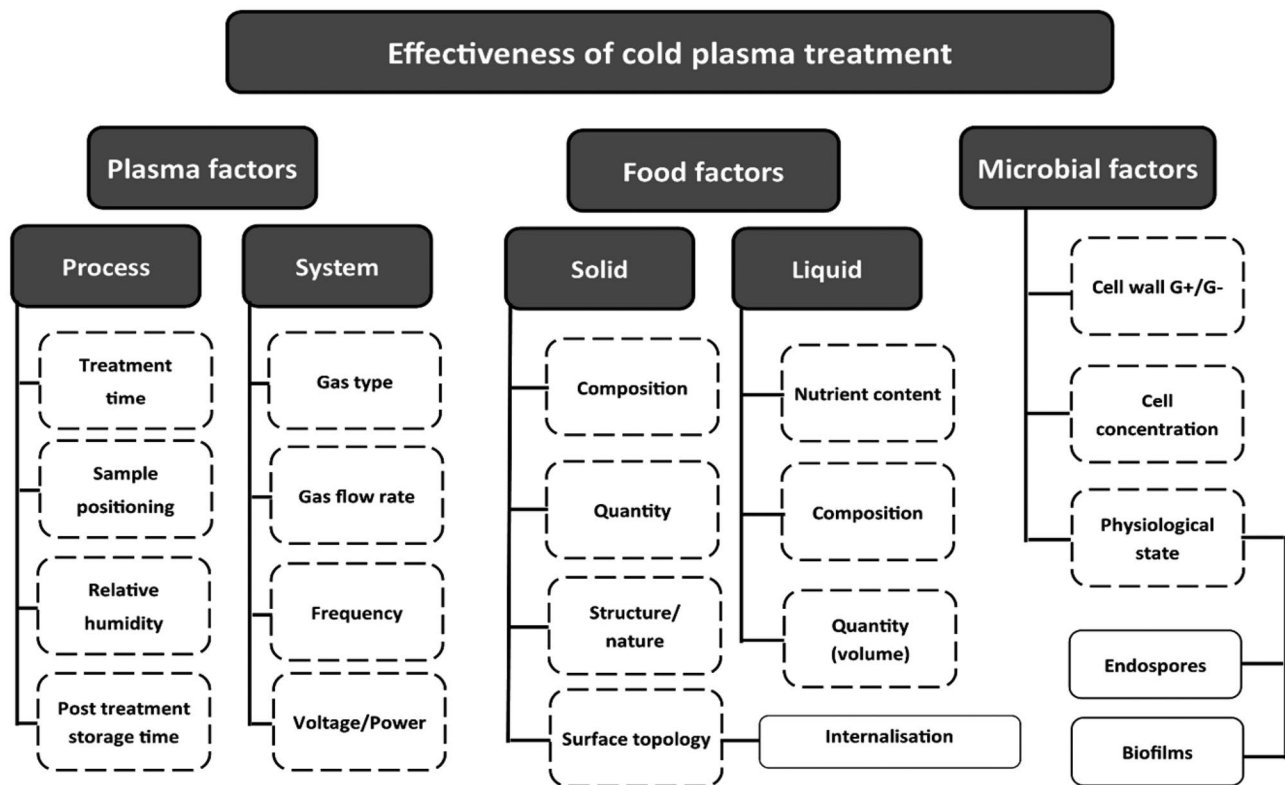


Fig. 5 Factors influencing cold plasma treatment. Adopted from Bourke et al. [119]

The antimicrobial efficacy of cold plasma treatment might be influenced by the nature of food constituents (e.g., fat, protein, carbohydrate content). Although several reviews have highlighted the possible effects of food composition and nature on the effectiveness of cold plasma treatment, only a few research has been dedicated to providing clarity on the subject.

The importance of osmotic stress and suboptimal pH on the efficacy of cold plasma inactivation of *Salmonella* Typhimurium and *Listeria monocytogenes* studied on various food structures demonstrated that growth under osmotic stress or at sub-optimal pH promotes microbial cell adaptation and resistance to cold plasma treatment [118]. In addition, a study that investigated the influence of surface roughness on the efficacy of cold atmospheric pressure plasma on microbial inactivation showed that an increase in surface roughness resulted in a decreased microbial inactivation efficacy [120]. The water content of food product affects the effectiveness of cold plasma treatment for inactivation of food contaminating microorganisms. The high hydroxyl radicals generated during treatment in liquid water phase promotes effective and increased inactivation of microbial cells [121]. Microbial characteristics, including type, strain, physiological state, growth phase, and mode of existence (planktonic or sessile). Foodborne microorganisms such as

Clostridium and *Bacillus* species is notable for spore formation and resistance to various food preservation treatments. Spore resistance to preservation treatments can be induced by a number of factors, including structure of the spores, formed with cortex, coat, and exosporangium of the spores; water content in the central region of core of the spore; the saturation of spore DNA by a group of acid-soluble proteins; and low permeability of the inner spore membrane to hydrophilic molecules [122–124]. Microbial exosporium layer protects the interior components of the cell from degradative effects of preservation treatment. Thus, several studies have demonstrated that preservation treatments including cold plasma are usually more effective against vegetative cells. Cold plasma inactivation of *G. stearothermophilus* and *B. cereus* vegetative cells and spores demonstrated a statistically significant difference in the inactivation of *G. stearothermophilus* vegetative cells receiving indirect and direct, as well as for *B. cereus* vegetative cells and spores. However, no statistically significant difference in the inactivation of *G. stearothermophilus* spores receiving indirect or direct exposure [96]. Los et al. [117] reported that the efficacy of cold plasma treatment was strongly affected by the type of microorganism studied. The authors observed high resistance for *B. atrophaeus* endospores, after direct and indirect plasma treatment for 20 min, and concluded that

the endospores of *B. atrophaeus* were considerably more resistant against ACP treatment than the vegetative cells. Furthermore, antimicrobial application in the food industry and processing facilities extend beyond preservation and shelf-life extension. Attachment of microorganisms on food contact surfaces and processing lines is a major cause of food contamination, especially of finished products. Persistent microorganisms in the food-processing environment and resistance to food-grade antimicrobial sanitizers and preservatives have emerged a food safety threat, resulting in finished product recall and foodborne disease outbreak. In addition, the formation of biofilm on food contact surfaces is a leading cause of food contamination, foodborne disease outbreaks, and recall of finished food products. The dense exopolymer matrix of biofilm communities is associated with various undesirable effects in food processing, including impaired heat flow, corrosion of contact surfaces, hampered diffusion of food-grade antimicrobial sanitizers, and resistance to antimicrobials [125, 126]. In recent years, the use of modern green technologies for the decontamination of food processing lines and facilities as well as hospital premises is replacing the conventional use of antimicrobial chemical sanitizers [127–129]. Studies have demonstrated that cold plasma treatment can effectively disrupt and inactivate microbial biofilm. The antimicrobial efficacy of atmospheric cold plasma against *Pseudomonas aeruginosa* biofilms revealed that treatment for 60 s by either the direct or the indirect exposure reduced bacterial populations by an average of 5.4 log cycles from an initial 6.6 log₁₀ CFU/mL. The extension of the treatment time from 60 to 120 s and 300 s reduced biofilms to undetectable levels [130]. Plasma treatment is also indicated for effective disruption of fungi biofilm. The in vitro and in vivo inactivation effects of cold plasma treatment on *C. albicans* biofilm indicated significant inactivation effects [131]. A similar study demonstrated that both gas plasma and plasma activated water treatment decreases of *A. flavus* metabolic activity and spore counts, with maximal reductions of 2.2 and 0.6 log₁₀ units for gas plasma and plasma-activated water, respectively. Biofilm study revealed detrimental effects of gas plasmas on biofilm structure [61]. Atmospheric cold plasma treatment on *E. coli* spp., *B. subtilis*, and *Lactobacillus* spp. biofilms resulted in > 3 log₁₀ after 5 min but was ineffective against *B. atrophaeus* [132]. Spores have generally been shown to require extended treatment in order to achieve enhanced inactivation compared with vegetative cells. Ziuzina et al. [133] reported that plasma treatment for 30 s reduced planktonic populations of *Salmonella*, *L. monocytogenes* and *E. coli* in lettuce broth to undetectable levels. However, depending on storage conditions, bacterial type, and age of biofilm, 300 s of treatment was required to reduce biofilm populations on lettuce by a maximum of 5 log₁₀ CFU/sample. An investigation of the effects of atmospheric cold plasma

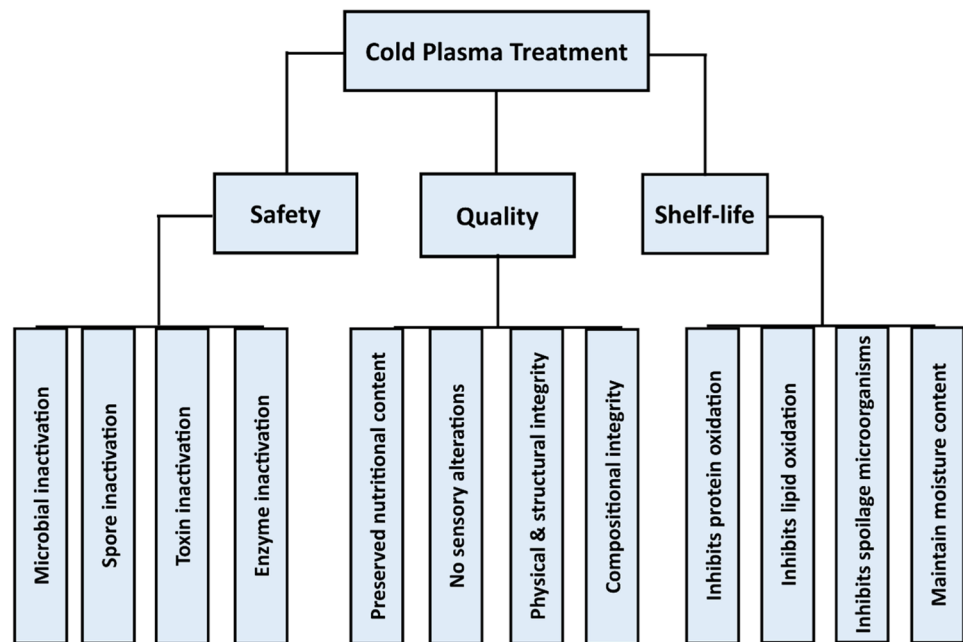
against microbial biofilms of foodborne pathogenic bacteria demonstrated that plasma treatment for 60 s reduced populations of *E. coli* to undetectable levels, whereas 300 s was necessary to significantly reduce populations of *L. monocytogenes* and *S. aureus* biofilms. The authors, however, suspected possible induction of viable but non-culturable state of bacteria following plasma treatment [107]. A 4-day-old single-species biofilms of *Chromobacterium violaceum* was effectively inactivated after 10-min plasma treatment [134]. Moreover, the effectiveness of plasma treatment varies from cell to cell due to variations in cellular properties. Besides spore formation and the formation of biofilms, the thickness of the bacterial cell wall has been correlated to the effectiveness of plasma treatments. Cells with thicker cell walls are more resistant to plasma treatment compared with cells with thin cell walls. Also, the effectiveness of cold plasma treatment is reported to vary between gram-negative and gram-positive bacteria, due to the difference in the cell wall composition [135].

Advantages and Disadvantages of Cold Plasma Treatment Over Other Technologies

The transition from a chemical preservative-based food processing to a green technology mediated food processing orchestrated by changing consumers taste, safety concerns, and changing food regulations has prompted a proliferation of several technologies seeking application in the food processing industry. Unfortunately, most of the proposed green technology is either limited due to expensive cost of machinery, effect on product quality, not suited for all food type or are inadequate for maximum protection of food product. Cold plasma technique has shown numerous advantages and suitability for the treatment of numerous food types and can ensure adequate inactivation of food contaminating microorganism, including recalcitrant microbial spores, as well as enzymes and toxins. Figure 6 highlights some of the benefits of cold plasma technique in the food industry. In addition, cold atmospheric plasma is versatile with applications in biomedical and food processing and relatively safe. However, in the absence of standardization in terms of treatment parameters and resulting biological effects [136].

Decontamination of food products with uneven surfaces might also pose safety concerns due to the internalization of microorganisms and subsequent compromise of food quality. Furthermore, although cold plasma treatment is assumed to preserve food quality, mild alterations might lead to unpleasant physical changes that might affect the consumer acceptability and result in economic loss. Chemical changes, such as lipid oxidation, may lead to both economic loss and prompt health concerns [137]. An increase in lipid oxidation was reported in cold plasma-treated mackerel. Similarly, the oleic acid and eicosapentaenoic acid contents were lower in

Fig. 6 Benefits of cold plasma processing in food preservation



mackerel slices [138]. Thiobarbituric acid reactive substance value showed a time-dependent manner increase in Asian sea bass slices after cold plasma treatment [70].

Adjunctive Application of Cold Plasma

As new technologies emerge and gain relevance in the food industry, it has become evident that few weaknesses might limit the spectrum of applicability irrespective of the excellent qualities in certain areas. The use of multiple food treatment technologies is perhaps a strategic approach for enhancing food safety. Complimentary usage of food processing technologies at mild or reduced process parameter (for example low temperature and low voltage) augments the effect of individual methods by introducing several hurdles while preserving the sensory, nutritional, and compositional qualities of the food product. This section reviews reported applications of cold plasma technique as an adjunctive or complementary method to promote product safety and shelf-life extension. Cold plasma-based hurdles can be employed to overcome limitations of individual cold plasma treatment, improve the inactivation efficacy, and retain the maximum food quality attributes [139]. The combined effects of ultrasound and cold plasma significantly improved carrot juice quality and safety, with a reduction in total plate count and yeast and mold count compared with products treated with individual technique. Ultrasound and cold plasma treatment enhance carrot juice's stability with increase, chlorogenic acid, sugar contents, and mineral profile [140]. Microwave-combined cold plasma treatment was effective for inactivating *Bacillus cereus* spores on contaminated red pepper without altering the physical properties. However,

process parameters influenced the degree of effectiveness [141]. A similar study reported that microwave-combined cold plasma treatment numbers of *B. cereus* and *A. flavus* spores on red pepper flakes. The antioxidant activity and color of the flakes were unaltered during storage following treatment with either low microwave cold plasma or high microwave cold plasma treatment. Furthermore, low microwave cold plasma also did not affect the sensory properties and the concentrations of capsaicin and dihydrocapsaicin of the flakes [72]. Shiekh et al. [142] employed a combination of pulsed electric field (PEF) pre-treatment, immersion in chamuang leaf extract (CLE) followed by high voltage cold atmospheric plasma for quality preservation and shelf-life extension of *Litopenaeus vannamei*, and reported the lowest microbial load and spoilage bacteria count, and higher likeness scores in PEF treated sample with 2% CLE followed by high voltage cold plasma treatment. They reported synergy between phenolic compounds in CLE and active species generated from high voltage cold atmospheric plasma on the effective inhibition of microbial growth of *L. vannamei* during refrigerated storage. A reduction melanosis scores, lipid oxidation, total volatile base, protein carbonyl contents, and microbial load were attained in CLE and PEF pre-treated shrimp samples exposed to cold plasma compared with control and other treated samples [143]. Application of cold plasma as an adjunctive treatment to nisin pre-treated was effectively employed for the inactivation of *Listeria monocytogenes* of the surface of apples [144]. Simultaneous ultraviolet and cold plasma treatments were used to inactivate indigenous mesophilic aerobic bacteria and *Bacillus tequilensis* spores on black peppercorns without altering the color of the black peppercorns [71]. The versatility of cold

plasma and complementarity with other technologies is a principal benefit that can be strategically harnessed for better product quality and shelf-life preservation.

Cold Plasma for Industrial Application

Part of the challenges associated with cold plasma is the presence of reactive oxygen species (ROS), which triggers lipid oxidation mostly in meat tissue and fish, fatty acids, aldehydes, hydroxyl acids, and keto acid which results in off-flavors and odors during storage [145]. These compounds negatively affect the product's acceptability and shelf life. In juices, cold plasma technology leads to the degradation of extremely polymerized oligosaccharides [146], which is caused by ozonolysis. The high consumption of gas is also cited as a limitation or drawback of the technology [147], especially for large-scale applications. Furthermore, the high cost of setting up the cold plasma technology tends to be a major challenge or drawback. Moreover, cold plasma treatment is associated with the undesirable alteration of textural properties, acidity, and decolorization of treated foods. Products of animal origin when treated with cold plasma exhibit acceleration of lipid oxidation which has a negative impact on the sensory characteristics. Several opportunities are associated with the use of cold plasma technology. Studies have demonstrated the potential and application of cold plasma technology in the decontamination of wastewater. Cold plasma decontamination lowers the amount of pollutants in wastewater [147]. Furthermore, treatment with cold plasma reduced several water-borne pathogenic microorganisms [46, 148–151] revealed the substantial inactivation of spike protein in plasma inactivated water to inhibit coronavirus transmission. Hence, the combination of CP with other suitable treatments approach possesses viable potential for wastewater remediation in the future.

Conclusion and Future Directives

In recent past, cold plasma treatment has received increasing popularity in the food industry due to the reported efficiency in microbial inactivation, toxin, enzyme degradation, and mild to absolutely no impact on food properties. Several research and reviews have attempted to provide clarity on the mechanism of microbial spore inactivation by cold plasma technique. This notwithstanding, the detailed mechanism of cold plasma is still not fully elucidated. Herein, the various suggested mechanisms as reported by research articles were explored, which provide direction for further studies. However, as the food production and processing sectors continuously adapt to the changing consumer's demands and regulations, more technological transition is expected. Non-thermal technologies with less impact on food sensory,

flavor, and textural attributes have so far proved beneficial to the food industry, with cold plasma as one of the most recent advances. The effectiveness of plasma treatment in food processing has been demonstrated by several studies on varying types of food products. However, it is relevant to fully understand the mechanism of action of cold plasma treatment, the role of the radical species generated, and to investigate its efficacy across the diverse physiological microbial state. Optimization of process parameters in line with individual food type and product should be a major scientific focus to harmonize parameters and promote usage ease. In addition, there is a need for research to focus on the effects of cold plasma treatment on the compositional, nutritional, and organoleptic properties of food products. Moreover, data on the chemical residual effects and potential toxicity of the various gases used in the generation of plasma would be necessary in guiding decision and regulation. The effects of combined usage of cold plasma with other technologies would be another interesting area for future outlook, and in addition to the evaluation of the residual chemical generation and chemical modification in antimicrobial compounds following plasma exposure. It is also important to verify the claims that treatment of microorganisms with cold plasma could induce a viable but not culturable state.

Author Contribution Conceptualization: HO, OFN; writing—original draft: HO, OFN; writing review and editing: OFN, AH, KO, CA, TM; data curation: OFN, KO; supervision: HO. All authors were involved in the preparation of the final draft.

Declarations

Conflict of Interest The authors declare no competing interests.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

References

1. Pandey H, Kumar V, Roy B (2014) Assessment of genotoxicity of some common food preservatives using *Allium cepa* L. as a test plant. *Toxicol Rep* 1:300–308
2. Sarıkaya R, Çakır Ş (2005) Genotoxicity testing of four food preservatives and their combinations in the *Drosophila* wing spot test. *Environ Toxicol Pharmacol* 20:424–430

3. Winkler C, Frick B, Schroecksnadel K, Schennach H, Fuchs D (2006) Food preservatives sodium sulfite and sorbic acid suppress mitogen-stimulated peripheral blood mononuclear cells. *Food Chem Toxicol* 44:2003–2007
4. Hernández-Hernández H, Moreno-Vilet L, Villanueva-Rodríguez S (2019) Current status of emerging food processing technologies in Latin America: novel non-thermal processing. *Innov Food Sci Emerg Technol* 58:102233
5. Khouryieh HA (2021) Novel and emerging technologies used by the US food processing industry. *Innov Food Sci Emerg Technol* 67:102559
6. Rifna E, Singh SK, Chakraborty S, Dwivedi M (2019) Effect of thermal and non-thermal techniques for microbial safety in food powder: recent advances. *Food Res Int* 126:108654
7. Leistner L, Gould GW (2012) Hurdle technologies: combination treatments for food stability, safety and quality. Springer Science & Business Media
8. Singh S, Shalini R (2016) Effect of hurdle technology in food preservation: a review. *Crit Rev Food Sci Nutr* 56:641–649
9. Gustavsson J, Cederberg C, Sonesson U, Van Otterdijk R, Meybeck A (2011) Global food losses and food waste. FAO, Rome
10. Silva FV (2018) Differences in the resistance of microbial spores to thermosonication, high pressure thermal processing and thermal treatment alone. *J Food Eng* 222:292–297
11. Cortesão M, De Haas A, Unterbusch R, Fujimori A, Schütze T, Meyer V, Moeller R (2020) *Aspergillus niger* spores are highly resistant to space radiation. *Front Microbiol* 11:560
12. Nicholson WL, Galeano B (2003) UV resistance of *Bacillus anthracis* spores revisited: validation of *Bacillus subtilis* spores as UV surrogates for spores of *B. anthracis* Sterne. *Appl Environ Microbiol* 69:1327–1330
13. Setlow P (2001) Resistance of spores of *Bacillus* species to ultraviolet light. *Environ Mol Mutagen* 38:97–104
14. Li J, Lu W, Wang H, Fan Y, Chang Z (2020) Groundwater contamination source identification based on a hybrid particle swarm optimization-extreme learning machine. *J Hydrol* 584
15. Soni A, Oey I, Silcock P, Ross IK, Bremer PJ (2020) Effect of pulsed electric field with moderate heat (80 °C) on inactivation, thermal resistance and differential gene expression in *B. cereus* spores. *J Food Process Preserv* 44:e14503
16. Beyrer M, Pina-Perez MC, Martinet D, Andlauer W (2020) Cold plasma processing of powdered *Spirulina* algae for spore inactivation and preservation of bioactive compounds. *Food Control* 118:107378
17. Ott LC, Appleton HJ, Shi H, Keener K, Mellata M (2021) High voltage atmospheric cold plasma treatment inactivates *Aspergillus flavus* spores and deoxynivalenol toxin. *Food Microbiol* 95:103669
18. Fan L, Hou F, Muhammad AI, Ruiling L, Watharkar RB, Guo M, Ding T, Liu D (2019) Synergistic inactivation and mechanism of thermal and ultrasound treatments against *Bacillus subtilis* spores. *Food Res Int* 116:1094–1102
19. Lv R, Muhammad AI, Zou M, Yu Y, Fan L, Zhou J, Ding T, Ye X, Guo M, Liu D (2020) Hurdle enhancement of acidic electrolyzed water antimicrobial efficacy on *Bacillus cereus* spores using ultrasonication. *Appl Microbiol Biotechnol* 1–9
20. Lv R, Zou M, Chantapakul T, Chen W, Muhammad AI, Zhou J, Ding T, Ye X, Liu D (2019) Effect of ultrasonication and thermal and pressure treatments, individually and combined, on inactivation of *Bacillus cereus* spores. *Appl Microbiol Biotechnol* 103:2329–2338
21. Sourri P, Argyri AA, Panagou EZ, Nychas G-JE, Tassou CC (2020) *Alicyclobacillus acidoterrestris* strain variability in the inactivation kinetics of spores in orange juice by temperature-assisted high hydrostatic pressure. *Appl Sci* 10:7542
22. Pankaj SK, Wan Z, Keener KM (2018) Effects of cold plasma on food quality: a review. *Foods* 7:4
23. Laroque DA, Seó ST, Valencia GA, Laurindo JB, Carciofi BAM (2022) Cold plasma in food processing: design, mechanisms, and application. *J Food Eng* 312:110748
24. Saremnezhad S, Soltani M, Faraji A, Hayaloglu AA (2021) Chemical changes of food constituents during cold plasma processing: a review. *Food Res Int* 147:110552
25. Isbary G, Shimizu T, Li Y-F, Stolz W, Thomas HM, Morfill GE, Zimmermann JL (2013) Cold atmospheric plasma devices for medical issues. *Expert Rev Med Devices* 10:367–377
26. Klämpfl TG, Isbary G, Shimizu T, Li Y-F, Zimmermann JL, Stolz W, Schlegel J, Morfill GE, Schmidt H-U (2012) Cold atmospheric air plasma sterilization against spores and other microorganisms of clinical interest. *Appl Environ Microbiol* 78:5077–5082
27. Konesky G (2013) Cold plasma decontamination device. Google Patents
28. O'Connor N, Cahill O, Daniels S, Galvin S, Humphreys H (2014) Cold atmospheric pressure plasma and decontamination. Can it contribute to preventing hospital-acquired infections? *J Hosp Infect* 88:59–65
29. Güçeri S, Fridman A, Gibson K, Haas C (2008) Plasma assisted decontamination of biological and chemical agents. Springer
30. Moutiq R, Pankaj S, Wan Z, Mendonca A, Keener K, Misra N (2020) Atmospheric pressure cold plasma as a potential technology to degrade carbamate residues in water. *Plasma Chem Plasma Process* 40:1291–1309
31. Shahidi S, Ghoranneviss M, Moazzenchi B (2014) New advances in plasma technology for textile. *J Fusion Energy* 33:97–102
32. Shishoo R (2007) Plasma technologies for textiles. Elsevier
33. Sparavigna A (2008) Plasma treatment advantages for textiles. arXiv preprint. <https://arxiv.org/abs/0801.3727>. Accessed Nov 2021
34. Yopez XV, Misra N, Keener KM (2020) Nonthermal plasma technology. In: Demirci A, Feng H, Krishnamurthy K (eds) *Food Safety Engineering*. Springer, Cham
35. Feizollahi E, Misra N, Roopesh M (2020). Factors influencing the antimicrobial efficacy of dielectric barrier discharge (DBD) atmospheric cold plasma (ACP) in food processing applications. *Crit Rev Food Sci Nutr* 1–24
36. Isbary G, Stolz W, Shimizu T, Monetti R, Bunk W, Schmidt H-U, Morfill GE, Klämpfl T, Steffes B, Thomas H (2013) Cold atmospheric argon plasma treatment may accelerate wound healing in chronic wounds: Results of an open retrospective randomized controlled study in vivo. *Clin Plasma Med* 1:25–30
37. Isbary G, Morfill G, Zimmermann J, Shimizu T, Stolz W (2011) Cold atmospheric plasma: a successful treatment of lesions in Hailey-Hailey disease. *Arch Dermatol* 147:388–390
38. Keidar M, Shashurin A, Volotskova O, Ann Stepp M, Srinivasan P, Sandler A, Trink B (2013) Cold atmospheric plasma in cancer therapy. *Phys Plasmas* 20:057101
39. De Groot GJ, Hundt A, Murphy AB, Bange MP, Mai-Prochnow A (2018) Cold plasma treatment for cotton seed germination improvement. *Sci Rep* 8:1–10
40. Ling L, Jiangang L, Minchong S, Chunlei Z, Yuanhua D (2015) Cold plasma treatment enhances oilseed rape seed germination under drought stress. *Sci Rep* 5:1–10
41. Sivachandiran L, Khacef A (2017) Enhanced seed germination and plant growth by atmospheric pressure cold air plasma: combined effect of seed and water treatment. *RSC Adv* 7:1822–1832
42. Sarangapani C, Misra N, Milosavljevic V, Bourke P, O'regan F, Cullen P (2016) Pesticide degradation in water using atmospheric air cold plasma. *J Water Process Eng* 9:225–232
43. Pankaj S, Wan Z, Colonna W, Keener KM (2017) Degradation kinetics of organic dyes in water by high voltage atmospheric air and modified air cold plasma. *Water Sci Technol* 76:567–574

44. Kim H-J, Won C-H, Kim H-W (2018) Pathogen deactivation of glow discharge cold plasma while treating organic and inorganic pollutants of slaughterhouse wastewater. *Water Air Soil Pollut* 229:1–10
45. Nguyen DV, Ho PQ, Pham TV, Nguyen TV, Kim L (2019) Treatment of surface water using cold plasma for domestic water supply. *Environ Eng Res* 24:412–417
46. Patange A, Boehm D, Giltrap M, Lu P, Cullen P, Bourke P (2018) Assessment of the disinfection capacity and eco-toxicological impact of atmospheric cold plasma for treatment of food industry effluents. *Sci Total Environ* 631:298–307
47. van Nguyen D, Ho NM, Hoang KD, Le TV, Le VH (2020) An investigation on treatment of groundwater with cold plasma for domestic water supply. *Groundw Sustain Dev* 10:100309
48. Ibrahim NA, Eid BM (2020) Plasma treatment technology for surface modification and functionalization of cellulosic fabrics. In: Shahid M, Adivarekar R (eds) *Advances in Functional Finishing of Textiles*. Springer, Singapore
49. Wang M-J, Chang Y-I, Poncin-Epaillard F (2003) Effects of the addition of hydrogen in the nitrogen cold plasma: the surface modification of polystyrene. *Langmuir* 19:8325–8330
50. Yoshinari M, Matsuzaka K, Inoue T (2011) Surface modification by cold-plasma technique for dental implants–bio-functionalization with binding pharmaceuticals. *Jpn Dent Sci Rev* 47:89–101
51. Misra NN, Yadav B, Roopesh MS, Jo C (2019) Cold plasma for effective fungal and mycotoxin control in foods: mechanisms, inactivation effects, and applications. *Compr Rev Food Sci Food Saf* 18:106–120
52. Bruggeman P, Walsh JL, Schram DC, Leys C, Kong MG (2009) Time dependent optical emission spectroscopy of sub-microsecond pulsed plasmas in air with water cathode. *Plasma Sources Sci Technol* 18:045023
53. Varilla C, Marcone M, Annor GA (2020) Potential of cold plasma technology in ensuring the safety of foods and agricultural produce: a review. *Foods* 9:1435
54. Ganesan AR, Tiwari U, Ezhilarasi P, Rajauria G (2021) Application of cold plasma on food matrices: a review on current and future prospects. *J Food Process Preserv* 45:e15070
55. Wu T-Y, Sun N-N, Chau C-F (2018) Application of corona electrical discharge plasma on modifying the physicochemical properties of banana starch indigenous to Taiwan. *J Food Drug Anal* 26:244–251
56. Takahashi K, Nakano Y, Ando A (2017) Frequency-tuning radio-frequency plasma source operated in inductively-coupled mode under a low magnetic field. *J Phys D Appl Phys* 50:265201
57. Lebedev YA (2010) Microwave discharges: generation and diagnostics. *Journal of physics: conference series*. IOP Publishing
58. Niemira BA, Gutsol A (2011) Nonthermal plasma as a novel food processing technology. *Nonthermal processing technologies for food*. pp 272–288
59. Sharma A, Josephson G, Camaioni D, Goheen S (2000) Destruction of pentachlorophenol using glow discharge plasma process. *Environ Sci Technol* 34:2267–2272
60. Starek A, Pawlat J, Chudzik B, Kwiatkowski M, Terebun P, Sagan A, Andrejko D (2019) Evaluation of selected microbial and physicochemical parameters of fresh tomato juice after cold atmospheric pressure plasma treatment during refrigerated storage. *Sci Rep* 9:1–11
61. Los A, Ziuzina D, Boehm D, Cullen PJ, Bourke P (2020) Inactivation efficacies and mechanisms of gas plasma and plasma-activated water against *Aspergillus flavus* spores and biofilms: A comparative study. *Appl Environ Microbiol* 86:e02619–e2719
62. Thomas-Popo E, Mendonça A, Misra N, Little A, Wan Z, Moutiq R, Coleman S, Keener K (2019) Inactivation of Shiga-toxin-producing *Escherichia coli*, *Salmonella enterica* and natural microflora on tempered wheat grains by atmospheric cold plasma. *Food Control* 104:231–239
63. Timmons C, Pai K, Jacob J, Zhang G, Ma LM (2018) Inactivation of *Salmonella enterica*, Shiga toxin-producing *Escherichia coli*, and *Listeria monocytogenes* by a novel surface discharge cold plasma design. *Food Control* 84:455–462
64. Zhuang H, Rothrock MJ Jr, Line JE, Lawrence KC, Gamble GR, Bowker BC, Keener KM (2020) Optimization of in-package cold plasma treatment conditions for raw chicken breast meat with response surface methodology. *Innov Food Sci Emerg Technol* 66:102477
65. Choi S, Puligundla P, Mok C (2016) Corona discharge plasma jet for inactivation of *Escherichia coli* O157: H7 and *Listeria monocytogenes* on inoculated pork and its impact on meat quality attributes. *Annals of Microbiology* 66:685–694
66. Puligundla P, Kim J-W, Mok C (2017) Effect of corona discharge plasma jet treatment on decontamination and sprouting of rapeseed (*Brassica napus* L.) seeds. *Food Control* 71:376–382
67. Yadav B, Spinelli AC, Misra NN, Tsui YY, McMullen LM, Roopesh MS (2020) Effect of in-package atmospheric cold plasma discharge on microbial safety and quality of ready-to-eat ham in modified atmospheric packaging during storage. *J Food Sci* 85:1203–1212
68. Wu X, Zhao W, Zeng X, Zhang QA, Gao G, Song S (2020) Effects of cold plasma treatment on cherry quality during storage. *Food Sci Technol Int* 27:441–455
69. Pathak N, Grossi Bovi G, Limnaios A, Fröhling A, Brincat JP, Taoukis P, Valdramidis VP, Schlüter O (2020) Impact of cold atmospheric pressure plasma processing on storage of blueberries. *J Food Process Preserv* 44:e14581
70. Olatunde OO, Benjakul S, Vongkamjan K (2020) Shelf-life of refrigerated Asian sea bass slices treated with cold plasma as affected by gas composition in packaging. *Int J Food Microbiol* 324:108612
71. Bang IH, Kim YE, Lee SY, Min SC (2020) Microbial decontamination of black peppercorns by simultaneous treatment with cold plasma and ultraviolet C. *Innov Food Sci Emerg Technol* 63:102392
72. Kim JE, Oh YJ, Song AY, Min SC (2019) Preservation of red pepper flakes using microwave-combined cold plasma treatment. *J Sci Food Agric* 99:1577–1585
73. Mahnot NK, Siyu LP, Wan Z, Keener KM, Misra NN (2020) In-package cold plasma decontamination of fresh-cut carrots: microbial and quality aspects. *J Phys D Appl Phys* 53
74. Giannoglou M, Stergiou P, Dimitrakellis P, Gogolides E, Stoforos NG, Katsaros G (2020) Effect of cold atmospheric plasma processing on quality and shelf-life of ready-to-eat rocket leafy salad. *Innov Food Sci Emerg Technol* 66
75. Zhou D, Wang Z, Tu S, Chen S, Peng J, Tu K (2019) Effects of cold plasma, UV-C or aqueous ozone treatment on *Botrytis cinerea* and their potential application in preserving blueberry. *J Appl Microbiol* 127:175–185
76. Shah U, Ranieri P, Zhou Y, Schauer CL, Miller V, Fridman G, Sekhon JK (2019) Effects of cold plasma treatments on spot-inoculated *Escherichia coli* O157:H7 and quality of baby kale (*Brassica oleracea*) leaves. *Innov Food Sci Emerg Technol* 57:102104
77. da Silva Campelo MC, de Oliveira Soares Rebouças LU, de Oliveira Vitoriano J, Alves C, da Silva JBA, de Oliveira Lima P (2019) Use of cold atmospheric plasma to preserve the quality of white shrimp (*Litopenaeus vannamei*). *J Food Prot* 82:1217–1223
78. Chen J, Wang SZ, Chen JY, Chen DZ, Deng SG, Xu B (2019) Effect of cold plasma on maintaining the quality of chub mackerel (*Scomber japonicus*): biochemical and sensory attributes. *J Sci Food Agric* 99:39–46

79. Lis KA, Boulaaba A, Binder S, Li Y, Kehrenberg C, Zimmermann JL, Klein G, Ahlfeld B (2018) Inactivation of *Salmonella* Typhimurium and *Listeria monocytogenes* on ham with nonthermal atmospheric pressure plasma. *PLoS ONE* 13
80. Prasad P, Mehta D, Bansal V, Sangwan RS (2017) Effect of atmospheric cold plasma (ACP) with its extended storage on the inactivation of *Escherichia coli* inoculated on tomato. *Food Res Int* 102:402–408
81. Patange A, Boehm D, Bueno-Ferrer C, Cullen PJ, Bourke P (2017) Controlling *Brochothrix thermosphacta* as a spoilage risk using in-package atmospheric cold plasma. *Food Microbiol* 66:48–54
82. Oh YJ, Song AY, Min SC (2017) Inhibition of *Salmonella typhimurium* on radish sprouts using nitrogen-cold plasma. *Int J Food Microbiol* 249:66–71
83. Kim JE, Oh YJ, Won MY, Lee KS, Min SC (2017) Microbial decontamination of onion powder using microwave-powered cold plasma treatments. *Food Microbiol* 62:112–123
84. Bauer A, Ni Y, Bauer S, Paulsen P, Modic M, Walsh JL, Smulders FJM (2017) The effects of atmospheric pressure cold plasma treatment on microbiological, physical-chemical and sensory characteristics of vacuum packaged beef loin. *Meat Sci* 128:77–87
85. Yong HI, Kim HJ, Park S, Choe W, Oh MW, Jo C (2014) Evaluation of the treatment of both sides of raw chicken breasts with an atmospheric pressure plasma jet for the inactivation of *Escherichia coli*. *Foodborne Pathog Dis* 11:652–657
86. Surowsky B, Fröhling A, Gottschalk N, Schlüter O, Knorr D (2014) Impact of cold plasma on *Citrobacter freundii* in apple juice: Inactivation kinetics and mechanisms. *Int J Food Microbiol* 174:63–71
87. Nicol MJ, Brubaker TR, Honish BJ, Simmons AN, Kazemi A, Geissel MA, Whalen CT, Siedlecki CA, Bilén SG, Knecht SD (2020) Antibacterial effects of low-temperature plasma generated by atmospheric-pressure plasma jet are mediated by reactive oxygen species. *Sci Rep* 10:1–11
88. Dobrynin D, Fridman G, Friedman G, Fridman A (2009) Physical and biological mechanisms of direct plasma interaction with living tissue. *New J Phys* 11:115020
89. Han L, Patil S, Boehm D, Milosavljević V, Cullen P, Bourke P (2016) Mechanisms of inactivation by high-voltage atmospheric cold plasma differ for *Escherichia coli* and *Staphylococcus aureus*. *Appl Environ Microbiol* 82:450–458
90. Laroussi M, Leipold F (2004) Evaluation of the roles of reactive species, heat, and UV radiation in the inactivation of bacterial cells by air plasmas at atmospheric pressure. *Int J Mass Spectrom* 233:81–86
91. Pai K, Timmons C, Roehm KD, Ngo A, Narayanan SS, Ramachandran A, Jacob JD, Ma LM, Madhally SV (2018) Investigation of the roles of plasma species generated by surface dielectric barrier discharge. *Sci Rep* 8:1–13
92. Connor M, Flynn PB, Fairley DJ, Marks N, Manesiotis P, Graham WG, Gilmore BF, Mcgrath JW (2017) Evolutionary clade affects resistance of *Clostridium difficile* spores to Cold atmospheric plasma. *Sci Rep* 7
93. Charoux CMG, Free L, Hinds LM, Vijayaraghavan RK, Daniels S, O'Donnell CP, Tiwari BK (2020) Effect of non-thermal plasma technology on microbial inactivation and total phenolic content of a model liquid food system and black pepper grains. *LWT* 118
94. Beyrer M, Smeu I, Martinet D, Howling A, Pina-Pérez MC, Ellert C (2020) Cold atmospheric plasma inactivation of microbial spores compared on reference surfaces and powder particles. *Food Bioprocess Technol* 13:827–837
95. Devi Y, Thirumdas R, Sarangapani C, Deshmukh RR, Annapure US (2017) Influence of cold plasma on fungal growth and aflatoxins production on groundnuts. *Food Control* 77:187–191
96. Morris AD, McCombs GB, Akan T, Hynes W, Laroussi M, Tolle SL (2009) Cold plasma technology: bactericidal effects on *Geobacillus stearothermophilus* and *Bacillus cereus* microorganisms. *Am Dent Hyg Assoc* 83:55–61
97. Casas-Junco PP, Solís-Pacheco JR, Ragazzo-Sánchez JA, Aguilar-Uscanga BR, Bautista-Rosales PU, Calderón-Santoyo M (2019) Cold plasma treatment as an alternative for ochratoxin a detoxification and inhibition of mycotoxigenic fungi in roasted coffee. *Toxins* 11:337
98. Dobrynin D, Fridman G, Mukhin YV, Wynosky-Dolfi MA, Rieger J, Rest RF, Gutsol AF, Fridman A (2010) Cold plasma inactivation of *Bacillus cereus* and *Bacillus anthracis* (anthrax) spores. *IEEE Trans Plasma Sci* 38:1878–1884
99. Lin H-M, Zhang S, Zheng R-S, Miao J-Y, Deng S-G (2020) Effect of atmospheric cold plasma treatment on ready-to-eat wine-pickled *Bullacta exarata*. *LWT* 120:108953
100. Pérez-Andrés JM, De Alba M, Harrison SM, Brunton NP, Cullen P, Tiwari BK (2020) Effects of cold atmospheric plasma on mackerel lipid and protein oxidation during storage. *LWT* 118:108697
101. Bao Y, Reddivari L, Huang J-Y (2020) Enhancement of phenolic compounds extraction from grape pomace by high voltage atmospheric cold plasma. *LWT* 133:109970
102. Li M, Li X, Han C, Ji N, Jin P, Zheng Y (2019) Physiological and Metabolomic analysis of cold plasma treated fresh-cut strawberries. *J Agric Food Chem* 67:4043–4053
103. Khosravi S, Jafari S, Zamani H, Nilkar M (2021) Inactivation of *Staphylococcus aureus* and *Escherichia coli* biofilms by air-based atmospheric-pressure DBD plasma. *Appl Biochem Biotechnol* 193:3641–3650
104. Patange AD, Simpson JC, Curtin JF, Burgess CM, Cullen P, Tiwari BK (2021) Inactivation efficacy of atmospheric air plasma and airborne acoustic ultrasound against bacterial biofilms. *Sci Rep* 11:1–14
105. Xiong Z, Du T, Lu X, Cao Y, Pan Y (2011) How deep can plasma penetrate into a biofilm? *Appl Phys Lett* 98:221503
106. Liu D, Liu Z, Chen C, Yang A, Li D, Rong M, Chen H, Kong M (2016) Aqueous reactive species induced by a surface air discharge: heterogeneous mass transfer and liquid chemistry pathways. *Sci Rep* 6:1–11
107. Ziuzina D, Boehm D, Patil S, Cullen P, Bourke P (2015) Cold plasma inactivation of bacterial biofilms and reduction of quorum sensing regulated virulence factors. *PLoS ONE* 10:e0138209
108. Patil S, Moiseev T, Misra N, Cullen P, Mosnier J, Keener K, Bourke P (2014) Influence of high voltage atmospheric cold plasma process parameters and role of relative humidity on inactivation of *Bacillus atrophaeus* spores inside a sealed package. *J Hosp Infect* 88:162–169
109. Olatunde OO, Benjakul S, Vongkamjan K (2019) Dielectric barrier discharge cold atmospheric plasma: bacterial inactivation mechanism. *J Food Saf* 39:e12705
110. Olatunde OO, Benjakul S, Vongkamjan K (2019) High voltage cold atmospheric plasma: antibacterial properties and its effect on quality of Asian sea bass slices. *Innov Food Sci Emerg Technol* 52:305–312
111. Liao X, Li J, Muhammad AI, Suo Y, Chen S, Ye X, Liu D, Ding T (2018) Application of a dielectric barrier discharge atmospheric cold plasma (Dbd-Acp) for *Escherichia coli* inactivation in apple juice. *J Food Sci* 83:401–408
112. Labarrere CA, Woods J, Hardin J, Campana G, Ortiz M, Jaeger B, Reichart B, Bonnin J, Currin A, Cosgrove S (2011) Early prediction of cardiac allograft vasculopathy and heart transplant failure. *Am J Transplant* 11:528–535
113. Kim J-S, Lee E-J, Kim Y-J (2014) Inactivation of *Campylobacter jejuni* with dielectric barrier discharge plasma using air and nitrogen gases. *Foodborne Pathog Dis* 11:645–651

114. Shi H, Ileeleji K, Stroshine RL, Keener K, Jensen JL (2017) Reduction of aflatoxin in corn by high voltage atmospheric cold plasma. *Food Bioprocess Technol* 10:1042–1052
115. Lu H, Patil S, Keener KM, Cullen P, Bourke P (2014) Bacterial inactivation by high-voltage atmospheric cold plasma: influence of process parameters and effects on cell leakage and DNA. *J Appl Microbiol* 116:784–794
116. Morgan N (2009) Atmospheric pressure dielectric barrier discharge chemical and biological applications. *Int J Phys Sci* 4:885–892
117. Los A, Ziuzina D, Akkermans S, Boehm D, Cullen PJ, Van Impe J, Bourke P (2018) Improving microbiological safety and quality characteristics of wheat and barley by high voltage atmospheric cold plasma closed processing. *Food Res Int* 106:509–521
118. Smet C, Noriega E, Rosier F, Walsh J, Valdramidis V, van Impe J (2016) Influence of food intrinsic factors on the inactivation efficacy of cold atmospheric plasma: impact of osmotic stress, suboptimal pH and food structure. *Innov Food Sci Emerg Technol* 38:393–406
119. Bourke P, Ziuzina D, Han L, Cullen P, Gilmore B (2017) Microbiological interactions with cold plasma. *J Appl Microbiol* 123:308–324
120. Bhide S, Salvi D, Schaffner DW, Karwe MV (2017) Effect of surface roughness in model and fresh fruit systems on microbial inactivation efficacy of cold atmospheric pressure plasma. *J Food Prot* 80:1337–1346
121. Shen J, Sun Q, Zhang Z, Cheng C, Lan Y, Zhang H, Xu Z, Zhao Y, Xia W, Chu PK (2015) Characteristics of DC gas-liquid phase atmospheric-pressure plasma and bacteria inactivation mechanism. *Plasma Processes Polym* 12:252–259
122. Akbas MY, Ozdemir M (2008) Effect of gaseous ozone on microbial inactivation and sensory of flaked red peppers. *Int J Food Sci Technol* 43:1657–1662
123. Kim JE, Lee D-U, Min SC (2014) Microbial decontamination of red pepper powder by cold plasma. *Food Microbiol* 38:128–136
124. Young S, Setlow P (2003) Mechanisms of killing of *Bacillus subtilis* spores by hypochlorite and chlorine dioxide. *J Appl Microbiol* 95:54–67
125. Alvarez-Ordóñez A, Coughlan LM, Briandet R, Cotter PD (2019) Biofilms in food processing environments: challenges and opportunities. *Annu Rev Food Sci Technol* 10:173–195
126. Galie S, García-Gutiérrez C, Miguélez EM, Villar CJ, Lombó F (2018) Biofilms in the food industry: health aspects and control methods. *Front Microbiol* 9:898
127. Cassar JR, Ouyang B, Krishnamurthy K, Demirci A (2020) Microbial decontamination of food by light-based technologies: ultraviolet (UV) Light, pulsed UV light (PUV), and UV light-emitting diodes (UV-LED). Springer, Food Safety Engineering
128. Dancer SJ (2014) Controlling hospital-acquired infection: focus on the role of the environment and new technologies for decontamination. *Clin Microbiol Rev* 27:665–690
129. Oliveira M, Tiwari BK, Duffy G (2020) Emerging technologies for aerial decontamination of food storage environments to eliminate microbial cross-contamination. *Foods* 9:1779
130. Ziuzina D, Patil S, Cullen P, Boehm D, Bourke P (2014) Dielectric barrier discharge atmospheric cold plasma for inactivation of *Pseudomonas aeruginosa* biofilms. *Plasma Med* 4:137–152
131. He M, Duan J, Xu J, Ma M, Chai B, He G, Gan L, Zhang S, Duan X, Lu X (2020) *Candida albicans* biofilm inactivated by cold plasma treatment in vitro and in vivo. *Plasma Processes Polym* 17:1900068
132. Los A, Ziuzina D, Boehm D, Cullen PJ, Bourke P (2017) The potential of atmospheric air cold plasma for control of bacterial contaminants relevant to cereal grain production. *Innov Food Sci Emerg Technol* 44:36–45
133. Ziuzina D, Han L, Cullen PJ, Bourke P (2015) Cold plasma inactivation of internalised bacteria and biofilms for *Salmonella enterica* serovar Typhimurium, *Listeria monocytogenes* and *Escherichia coli*. *Int J Food Microbiol* 210:53–61
134. Abramzon N, Joaquin JC, Bray J, Brelles-Mariño G (2006) Biofilm destruction by RF high-pressure cold plasma jet. *IEEE Trans Plasma Sci* 34:1304–1309
135. Mai-Prochnow A, Clauson M, Hong J, Murphy AB (2016) Gram positive and gram negative bacteria differ in their sensitivity to cold plasma. *Sci Rep* 6:1–11
136. Boehm D, Bourke P (2019) Safety implications of plasma-induced effects in living cells—a review of in vitro and in vivo findings. *Biol Chem* 400:3–17
137. Gavahian M, Chu YH, Jo C (2019) Prospective applications of cold plasma for processing poultry products: benefits, effects on quality attributes, and limitations. *Compr Rev Food Sci Food Saf* 18:1292–1309
138. Albertos I, Martín-Diana A, Cullen P, Tiwari BK, Ojha S, Bourke P, Álvarez C, Rico D (2017) Effects of dielectric barrier discharge (DBD) generated plasma on microbial reduction and quality parameters of fresh mackerel (*Scomber scombrus*) fillets. *Innov Food Sci Emerg Technol* 44:117–122
139. Liao X, Cullen P, Muhammad AI, Jiang Z, Ye X, Liu D, Ding T (2020) Cold plasma-based hurdle interventions: new strategies for improving food safety. *Food Eng Rev* 12:321–332
140. Umair M, Jabbar S, Senan AM, Sultana T, Nasiru MM, Shah AA, Zhuang H, Jianhao Z (2019) Influence of combined effect of ultra-sonication and high-voltage cold plasma treatment on quality parameters of carrot juice. *Foods* 8:593
141. Kim JE, Choi H-S, Lee D-U, Min SC (2017) Effects of processing parameters on the inactivation of *Bacillus cereus* spores on red pepper (*Capsicum annuum* L.) flakes by microwave-combined cold plasma treatment. *Int J Food Microbiol* 263:61–66
142. Shiekh KA, Zhou P, Benjakul S (2021) Combined effects of pulsed electric field, Chamuang leaf extract and cold plasma on quality and shelf-life of *Litopenaeus vannamei*. *Food Biosci* 41:100975
143. Shiekh KA, Benjakul S, Qi H, Zhang B, Deng S (2021) Combined hurdle effects of pulsed electric field and vacuum impregnation of Chamuang leaf extract on quality and shelf-life of Pacific white shrimp subjected to high voltage cold atmospheric plasma. *Food Packag Shelf Life* 28:100660
144. Ukuku DO, Niemira BA, Ukanalis J (2019) Nisin-based antimicrobial combination with cold plasma treatment inactivate *Listeria monocytogenes* on Granny Smith apples. *LWT* 104:120–127
145. Ekezie FGC, Sun DW, Cheng JH (2017) A review on recent advances in cold plasma technology for the food industry: Current applications and future trends. *Trends Food Sci Technol* 69:46–58
146. Almeida FDL, Cavalcante RS, Cullen PJ, Frias J, Bourke P, Fernandes FAN, Rodrigues S (2015) Effects of atmospheric cold plasma and ozone on prebiotic orange juice. *Innov Food Sci Emerg Technol* 32:127–135
147. Gururani P, Bhatnagar P, Bisht B, Kumar V, Joshi NC, Tomar MS, Pathak B (2021) Cold plasma technology: advanced and sustainable approach for wastewater treatment. *Env Sci Poll Res*
148. Guo L, Xu R, Gou L, Liu Z, Zhao Y, Liu D, Kong MG (2018) Mechanism of virus inactivation by cold atmospheric-pressure plasma and plasma-activated water. *Appl Environ Microbiol* 84(17)
149. Ma S, Kim K, Chun S, Moon SY, Hong Y (2020) Plasma-assisted advancedoxidation process by a multi-hole dielectric barrier discharge in water and its application to wastewater treatment. *Chemosphere* 243:125377

150. Wang N, Chen DZ, Zou LS (2015) Influence of nonthermal plasma pretreatment on the scaling characteristics of viscous oil wastewater during evaporation. *Appl Therm Eng* 75:779–788
151. Guo L, Yao Z, Yang L, Zhang H, Qi Y, Gou L, Kong MG (2020) Plasmaactivated water: An alternative disinfectant for S protein inactivation to prevent SARS-CoV-2 infection. *Chem Eng J*. 421(Part 2):127742

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.