



Mesenchymal stem cells transplantation for neuroprotection in preterm infants with severe intraventricular hemorrhage

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Severe intraventricular hemorrhaging (IVH) in premature infants and subsequent posthemorrhagic hydrocephalus (PHH) causes significant mortality and life-long neurological complications, including seizures, cerebral palsy, and developmental retardation. However, there are currently no effective therapies for neonatal IVH. The pathogenesis of PHH has been mainly explained by inflammation within the subarachnoid spaces due to the hemolysis of extravasated blood after IVH. Obliterative arachnoiditis, induced by inflammatory responses, impairs cerebrospinal fluid (CSF) resorption and subsequently leads to the development of PHH with ensuing brain damage. Increasing evidence has demonstrated potent immunomodulating abilities of mesenchymal stem cells (MSCs) in various brain injury models. Recent reports of MSC transplantation in an IVH model of newborn rats demonstrated that intraventricular transplantation of MSCs downregulated the inflammatory cytokines in CSF and attenuated progressive PHH. In addition, MSC transplantation mitigated the brain damages that ensue after IVH and PHH, including reactive gliosis, cell death, delayed myelination, and impaired behavioral functions. These findings suggest that MSCs are promising therapeutic agents for neuroprotection in preterm infants with severe IVH.

Key words: Intracranial hemorrhage, Hydrocephalus, Cell transplantation, Mesenchymal stromal cells, Premature infant

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Introduction

Intraventricular hemorrhaging (IVH) is a condition in which a germinal matrix hemorrhage ruptures through the ependyma into the lateral ventricle; it occurs mainly in premature infants. The risk of IVH increases with decreasing immaturity¹⁾. Consequently, the recent improvements toward the survival of very preterm infants through advances in neonatal intensive care medicine have resulted in an increased number of infants being at a high risk of developing IVH²⁾. A direct correlation exists between the extent of immaturity and the severity of IVH³⁾. More than 50% of preterm infants with severe IVH (grade > 3) die or develop posthemorrhagic hydrocephalus (PHH), which requires shunt surgery in up to 70% of the cases⁴⁾. Brain white-matter damage, which is induced by severe IVH, is exacerbated by PHH and results in increased mortality and long-term neurological morbidities such as seizures, cerebral palsy, and developmental retardation in the survivors⁵⁻⁷⁾. Currently, no effective therapies exist to ameliorate brain injury and prevent PHH development after severe IVH. Thus, the development of new therapeutic modalities to improve the prognosis of severe IVH in preterm infants is an urgent requirement.

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Pathogenesis of PHH after severe IVH

The germinal matrix is a highly vascularized structure located along the lateral ventricle. Its intrinsic fragility is related to the immaturity of the blood-brain barrier, and it contributes to primary bleeding into the germinal matrix. Multiple risk factors such as vaginal delivery, low Apgar score, respiratory-distress syndrome, pneumothorax, hypoxia, hypercapnia, and infection can be attributed to the disturbance in the cerebral blood flow and to the induction of IVH in preterm infants. After bleeding from the germinal matrix into the cerebral ventricles, subsequent hemolysis in the intraventricular space elevates the concentration of extracellular hemoglobin. The ensuing heme degradation increases the concentrations of bilirubin, carbon monoxide, and free iron⁸⁾ in the cerebrospinal fluid (CSF). Cell-free hemoglobin is known to initiate proinflammation, chemotaxis, and apoptosis in intracranial hemorrhaging^{9,10)}. Extravasated blood in the CSF activates an inflammatory response in the microvascular barrier and results in the dysfunction of arachnoid granulations, which reduces resorption of the CSF. This imbalance in CSF production and resorption results in the retention of CSF and progressive hydrocephalus. In newborn infants, hydrocephalus itself causes brain-parenchymal injury through various mechanisms, including increased intracranial pressure and the resultant decrease in cerebral perfusion, iron-induced free radical damage, and inflammatory cytokine levels¹¹⁻¹⁵⁾.

Neonatal animal models of IVH and hydrocephalus

Development of an appropriate animal model that simulates severe IVH in premature infants is essential for understanding its pathophysiology and for testing the efficacy of new therapeutic strategies. However, no single model is suitable for studying all the aspects of brain injury. Some animal models of neonatal IVH were induced via the direct injection of blood into the brain of newborn animals¹⁶⁻²⁰⁾. A rodent model was induced by administering a bilateral injection of 80 μ L blood into the cerebral ventricles¹⁶⁾, and another model was induced by administering a blood injection into the periventricular region¹⁷⁾. However, only 15%–65% of these models developed significant PHH^{16,17,21)}. In another model, germinal matrix hemorrhage (GMH) was developed by injecting clostridial collagenase into the germinal matrix, but no ventricular dilatation was observed²²⁾. Another GMH-IVH animal model precipitated GMH by intraperitoneal injection of glycerol into preterm rabbits delivered at 29 days gestation (full term is 32 days)²³⁾. Only 39% of this rabbit model developed severe IVH that was clinically relevant to grade 3 or 4 IVH—the main cause of PHH in preterm infants²³⁾. We recently reported a modeling method in which direct injection of dam's blood (200

μ L) into the cerebral ventricles of postnatal day (P) 4 rat pups developed IVH animal models²⁴⁾. With this method, 100% of the rats developed severe IVH (compatible with grade 3 or 4 IVH), as confirmed via brain magnetic resonance images (MRIs) on the first day after modeling. About 85% of these pups subsequently developed progressive PHH, which lasted for more than 4 weeks after the modeling²⁴⁾ (Fig. 1). In addition, as the rodent brains at birth and at P10 are compatible with those at 24- and 40-weeks' gestation, respectively, the rodent brain at P4 should represent the developmental state of the brain in preterm infants²⁵⁾. About 4 weeks after IVH induction, at P4, the rats showed significantly impaired sensorimotor functions and augmented cell death, inflammation, and delayed myelination in the brain tissue with significantly upregulated inflammatory cytokines in the CSF²⁴⁾. This observation suggested the replication of the pathogenesis of IVH in preterm infants in that hemolysis from the extravasated blood activated inflammation in the subarachnoid space, resulting in hydrocephalus and subsequent pressure injury to the brain tissue.

Attempt to downregulate inflammation and reduce PHH after severe IVH

Research on newborn IVH has shown that transforming growth factor (TGF)- β is associated with the accumulation of extracellular matrix proteins and collagen, which probably inter-

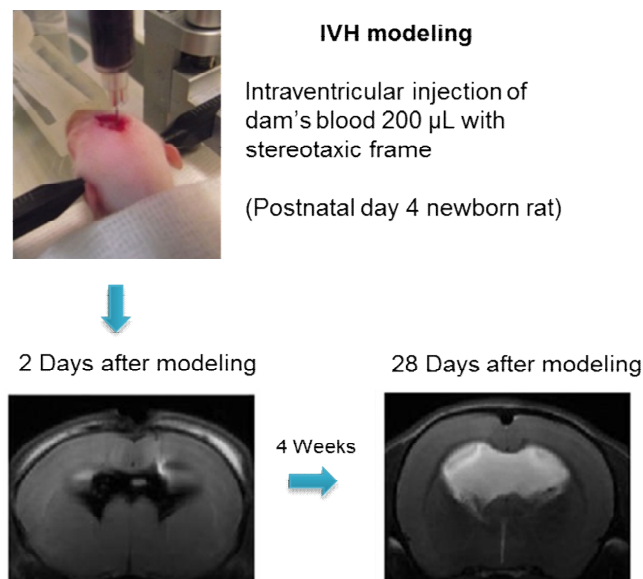


Fig. 1. Animal modeling of intraventricular hemorrhaging (IVH) in preterm infants using rat pups at postnatal day 4. The brain magnetic resonance imaging (MRI) after 2 days of direct injection of 200 μ L dam's blood showed significant enlargement of the ventricles filled with the injected blood. About 4 weeks after IVH modeling, the brain MRI showed severe hydrocephalus.

feres with the CSF flow²⁶⁻²⁸). However, several different TGF- β blocking agents failed to stop the progression of ventricular dilatation and did not improve the neuromotor performance in an IVH neonatal rat model^{19,20}. Increasing evidence suggests that inflammatory cytokines with activated macrophages are detrimental to neural cells. Inflammatory mechanisms appear to be the key factors in secondary brain injury, such as neuronal-cell death and reactive gliosis after intracerebral hemorrhaging²⁹⁻³¹. A study involving the newborn rabbit IVH model showed that the inhibition of cyclooxygenase-2 in the inflammatory cascade, which is induced by IVH, alleviated neurological impairment and delayed myelination and reactive gliosis³². Another study using this model established that the inhibition of bone morphogenic protein restored oligodendrocyte maturation, myelination, astrocyte morphology, and motor function³³. These therapeutic strategies did not reduce the size of enlarged cerebral ventricles after IVH³³; however, these reports suggest that modulating only one factor is not sufficient to improve PHH and that a multifaceted therapeutic agent is needed.

Therapeutic potential of MSCs for severe IVH

Stem-cell therapy has shown promising results in a number of brain injury or disease models³⁴⁻³⁷. Among the therapeutic sources, mesenchymal stem cells (MSCs) are well known for their anti-inflammatory property. In a mixed-lymphocyte reaction, MSCs reduced T-lymphocyte proliferation^{38,39}, probably due to

their lack of major histocompatibility-complex class II antigens, which prevent the development of a graft-versus-host response⁴⁰. In addition, MSCs have been known to shift the inflammatory response by polarizing the cytokine profile of T-cell subsets to an anti-inflammatory phenotype. This low-immunogenic property of MSCs is believed to be related to the secretion of various factors that create an immunosuppressive environment^{41,42}. MSCs decrease TNF- α , interleukin (IL) 12, and interferon- γ production and increase IL-10 production in dendritic cells⁴³. Besides the anti-inflammatory effects, various neurotropic factors secreted from MSCs promote angiogenesis, myelination, and rewiring of neurons and reduce gliosis and cell death. Thus, in a number of disease models, various paracrine factors such as numerous anti-inflammatory cytokines and tropic factors secreted from MSCs display potent anti-inflammatory efficacy by restoring brain injuries. Until date, single agents such as decorin, colchicine, TGF- β blocker, nonsteroidal anti-inflammatory drugs, and bone-morphogenic protein inhibitor have not proven effective in reducing the brain damage and PHH after severe IVH²⁴. Recently, we used various animal disease models to demonstrate the beneficial effects of MSC transplantation, including antiapoptotic, anti-inflammatory, antifibrotic, antioxidative paracrine, and regenerative effects^{24,44-48}. Considering its multiple therapeutic efficacies, MSCs, rather than single agents, are potential candidates for therapies aimed at improving the outcome of severe IVH in preterm infants.

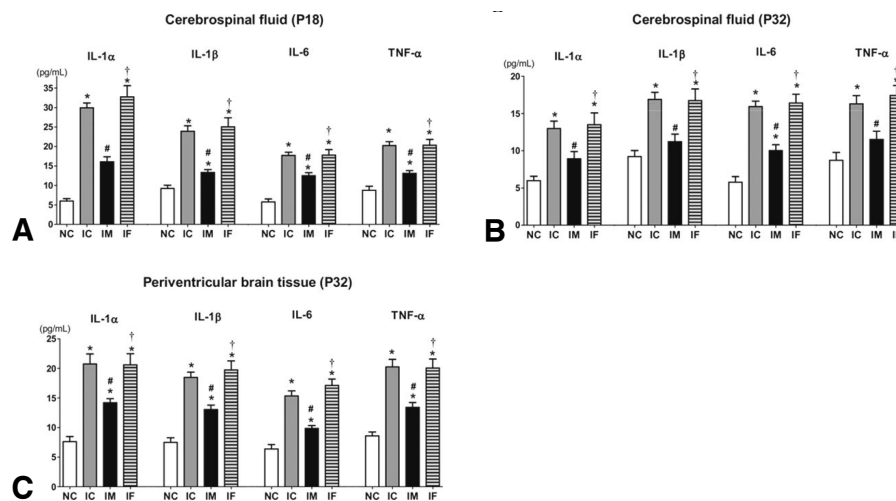


Fig. 2. Concentration of inflammatory cytokines (interleukin [IL] 1 α , IL- β , IL-6, and tumor necrosis factor [TNF]- α) in the cerebrospinal fluid obtained at postnatal day (P) 18 (A) and P32 (B) and from brain homogenate from the periventricular area at P32 (C). Data were expressed as mean \pm error of the mean. IVH, intraventricular hemorrhaging; NC, normal control rats; IC, IVH control rats; IM, IVH with human umbilical cord blood-derived mesenchymal stem cell transplantation; IF, IVH with human fibroblast transplantation. * P <0.05 vs NC. † P <0.05 vs IVH-M. # P <0.05 vs IC. Adapted from Ahn et al. Stroke 2013;44:497-504²⁴.

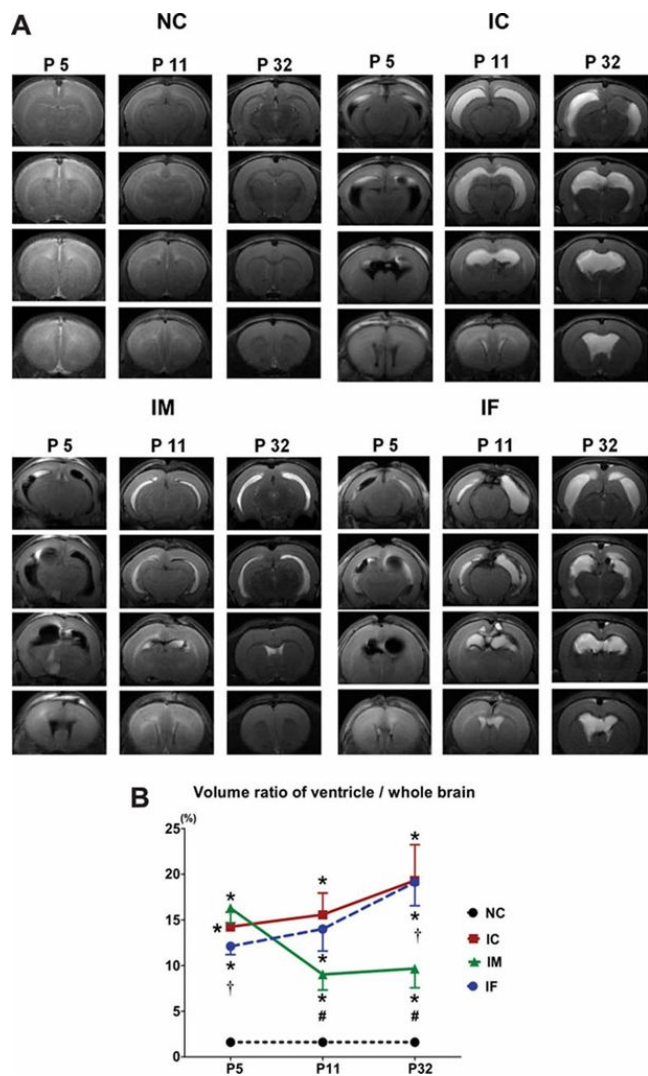


Fig. 3. Stem-cell transplantation attenuates ventricular dilatation and its progression after severe IVH. (A) Representative serial brain magnetic resonance images (MRIs) of each group on days 1, 7, and 28 after the induction of IVH (postnatal day 5, 11, and 32, respectively). (B) The ratio of ventricle volume to whole-brain volume measured by MRI. Data were expressed as mean±standard error of the mean. IVH, intraventricular hemorrhaging; NC, normal control rats; IC, IVH control rats; IM, IVH with human umbilical cord blood-derived mesenchymal stem cell transplantation; IF, IVH with human fibroblast transplantation. **P*<0.05 vs NC. †*P*<0.05 vs IVH-M. #*P*<0.05 vs IC. Adapted from Ahn et al. *Stroke* 2013;44:497-504²⁴.

Remaining challenges in MSC transplantation for severe IVH

We demonstrated that intraventricular transplantation of MSCs significantly attenuated inflammatory cytokines of the CSF and brain tissue (Fig. 2) and prevented the development of PHH after severe IVH (Fig. 3) in P4 newborn rats²⁴. MSC transplantation significantly improved brain apoptosis and reactive gliosis and

reduced myelin basic protein and abnormal sensorimotor functions in severe IVH. The neuroprotective mechanism appeared to be mediated by the paracrine anti-inflammatory effects rather than the regeneration of transplanted MSCs. Overall, the findings of the present study support the therapeutic potential of MSC transplantation in improving the prognosis of the serious complications of preterm birth, i.e., severe IVH. However, several hurdles remain, before we can achieve safe clinical transplantation of MSCs in preterm infants with severe IVH. Techniques must be developed and standardized for defining, isolating, and expanding MSCs. Although our previous study demonstrated the therapeutic efficacy of MSC transplantation, further issues remain to be addressed, such as the optimal dose, the route, and the timing of MSC transplantation. Safe clinical use of MSC transplantation to treat brain damage and prevent PHH in preterm infants with severe IVH requires further delineation of the neuroprotective mechanism of MSC and the assessment of the short- and long-term safety and efficacy of MSC transplantation.

Conclusions

Our recent observation that MSCs transplantation significantly attenuated the PHH and brain injury after severe IVH in the newborn rats holds future promise of stem cell therapy as novel therapeutic modality for treating the currently intractable severe IVH in the premature infants. However, for safe and successful clinical translation of stem cell therapy, further preclinical studies including the determination of optimal route, dose and timing, and the assessment of the short-term and long-term efficacy and safety of cell-based therapies are warranted.

Conflict of interest

No potential conflict of interest relevant to this article was reported.

Acknowledgments

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References

1. Thorp JA, Jones PG, Clark RH, Knox E, Peabody JL. Perinatal factors associated with severe intracranial hemorrhage. *Am J Obstet Gynecol* 2001;185:859-62.
2. Vohr BR, Wright LL, Dusick AM, Mele L, Verter J, Steichen JJ, et al. Neurodevelopmental and functional outcomes of extremely low birth weight infants in the National Institute of Child Health and Human Development Neonatal Research Network, 1993-1994. *Pediatrics* 2000;105:1216-26.
3. Stoll BJ, Hansen NI, Bell EF, Shankaran S, Laptook AR, Walsh MC, et al. Neonatal outcomes of extremely preterm infants from the NICHD Neonatal Research Network. *Pediatrics* 2010;126:443-56.
4. Murphy BP, Inder TE, Rooks V, Taylor GA, Anderson NJ, Mogridge N, et al. Posthaemorrhagic ventricular dilatation in the premature infant: natural history and predictors of outcome. *Arch Dis Child Fetal Neonatal Ed* 2002;87:F37-41.
5. International randomised controlled trial of acetazolamide and furosemide in posthaemorrhagic ventricular dilatation in infancy. International PHVD Drug Trial Group. *Lancet* 1998;352:433-40.
6. Randomised trial of early tapping in neonatal posthaemorrhagic ventricular dilatation. Ventriculomegaly Trial Group. *Arch Dis Child* 1990;65(1 Spec No):3-10.
7. Kennedy CR, Ayers S, Campbell MJ, Elbourne D, Hope P, Johnson A. Randomized, controlled trial of acetazolamide and furosemide in posthemorrhagic ventricular dilation in infancy: follow-up at 1 year. *Pediatrics* 2001;108:597-607.
8. Faivre B, Menu P, Labrude P, Vigneron C. Hemoglobin autooxidation/oxidation mechanisms and methemoglobin prevention or reduction processes in the bloodstream. Literature review and outline of autooxidation reaction. *Artif Cells Blood Substit Immobil Biotechnol* 1998;26:17-26.
9. Fang H, Wang PF, Zhou Y, Wang YC, Yang QW. Toll-like receptor 4 signaling in intracerebral hemorrhage-induced inflammation and injury. *J Neuroinflammation* 2013;10:27.
10. Xi G, Keep RF, Hoff JT. Mechanisms of brain injury after intracerebral haemorrhage. *Lancet Neurol* 2006;5:53-63.
11. Bhattathiri PS, Gregson B, Prasad KS, Mendelow AD; STICH Investigators. Intraventricular hemorrhage and hydrocephalus after spontaneous intracerebral hemorrhage: results from the STICH trial. *Acta Neurochir Suppl* 2006;96:65-8.
12. Halleivi H, Albright KC, Aronowski J, Barreto AD, Martin-Schild S, Khaja AM, et al. Intraventricular hemorrhage: Anatomic relationships and clinical implications. *Neurology* 2008;70:848-52.
13. Halleivi H, Walker KC, Kasam M, Bornstein N, Grotta JC, Savitz SI. Inflammatory response to intraventricular hemorrhage: time course, magnitude and effect of t-PA. *J Neurol Sci* 2012;315:93-5.
14. Del Bigio MR. Pathophysiologic consequences of hydrocephalus. *Neurosurg Clin N Am* 2001;12:639-49.
15. Whitelaw A, Thoresen M, Pople I. Posthaemorrhagic ventricular dilatation. *Arch Dis Child Fetal Neonatal Ed* 2002;86:F72-4.
16. Cherian SS, Love S, Silver IA, Porter HJ, Whitelaw AG, Thoresen M. Posthemorrhagic ventricular dilation in the neonate: development and characterization of a rat model. *J Neuropathol Exp Neurol* 2003;62:292-303.
17. Balasubramaniam J, Xue M, Buist RJ, Ivanco TL, Natuik S, Del Bigio MR. Persistent motor deficit following infusion of autologous blood into the periventricular region of neonatal rats. *Exp Neurol* 2006;197:122-32.
18. Aquilina K, Hobbs C, Cherian S, Tucker A, Porter H, Whitelaw A, et al. A neonatal piglet model of intraventricular hemorrhage and posthemorrhagic ventricular dilation. *J Neurosurg* 2007;107(2 Suppl):126-36.
19. Aquilina K, Hobbs C, Tucker A, Whitelaw A, Thoresen M. Do drugs that block transforming growth factor beta reduce posthaemorrhagic ventricular dilatation in a neonatal rat model? *Acta Paediatr* 2008;97:1181-6.
20. Hoque N, Thoresen M, Aquilina K, Hogan S, Whitelaw A. Decorin and colchicine as potential treatments for post-haemorrhagic ventricular dilatation in a neonatal rat model. *Neonatology* 2011;100:271-6.
21. Balasubramaniam J, Del Bigio MR. Animal models of germinal matrix hemorrhage. *J Child Neurol* 2006;21:365-71.
22. Lekic T, Manaenko A, Rolland W, Tang J, Zhang JH. A novel pre-clinical model of germinal matrix hemorrhage using neonatal rats. *Acta Neurochir Suppl* 2011;111:55-60.
23. Georgiadis P, Xu H, Chua C, Hu F, Collins L, Huynh C, et al. Characterization of acute brain injuries and neurobehavioral profiles in a rabbit model of germinal matrix hemorrhage. *Stroke* 2008;39:3378-88.
24. Ahn SY, Chang YS, Sung DK, Sung SI, Yoo HS, Lee JH, et al. Mesenchymal stem cells prevent hydrocephalus after severe intraventricular hemorrhage. *Stroke* 2013;44:497-504.
25. Dobbing J, Sands J. Comparative aspects of the brain growth spurt. *Early Hum Dev* 1979;3:79-83.
26. Galbreath E, Kim SJ, Park K, Brenner M, Messing A. Overexpression of TGF-beta 1 in the central nervous system of transgenic mice results in hydrocephalus. *J Neuropathol Exp Neurol* 1995;54:339-49.
27. Whitelaw A, Christie S, Pople I. Transforming growth factor-beta 1: a possible signal molecule for posthemorrhagic hydrocephalus? *Pediatr Res* 1999;46:576-80.
28. Heep A, Stoffel-Wagner B, Bartmann P, Benseler S, Schaller C, Gronck P, et al. Vascular endothelial growth factor and transforming growth factor-beta 1 are highly expressed in the cerebrospinal fluid of premature infants with posthemorrhagic hydrocephalus. *Pediatr Res* 2004;56:768-74.
29. Aronowski J, Hall CE. New horizons for primary intracerebral hemorrhage treatment: experience from preclinical studies. *Neurol Res* 2005;27:268-79.
30. Wang J, Tsirka SE. Contribution of extracellular proteolysis and microglia to intracerebral hemorrhage. *Neurocrit Care* 2005;3:77-85.
31. Barone FC, Feuerstein GZ. Inflammatory mediators and stroke: new opportunities for novel therapeutics. *J Cereb Blood Flow Metab* 1999;19:819-34.
32. Vinukonda G, Csiszar A, Hu F, Dummula K, Pandey NK, Zia MT, et al. Neuroprotection in a rabbit model of intraventricular haemorrhage by cyclooxygenase-2, prostanoid receptor-1 or tumour necrosis factor-alpha inhibition. *Brain* 2010;133(Pt 8):2264-80.
33. Dummula K, Vinukonda G, Chu P, Xing Y, Hu F, Maik S, et al. Bone morphogenetic protein inhibition promotes neurological recovery after intraventricular hemorrhage. *J Neurosci* 2011;31:12068-82.
34. Cao Q, Benton RL, Whittemore SR. Stem cell repair of central nervous system injury. *J Neurosci Res* 2002;68:501-10.
35. Conti L, Cataudella T, Cattaneo E. Neural stem cells: a pharmacological tool for brain diseases? *Pharmacol Res* 2003;47:289-97.
36. Pluchino S, Zanotti L, Rossi B, Brambilla E, Ottoboni L, Salani G, et al. Neurosphere-derived multipotent precursors promote neuroprotection by an immunomodulatory mechanism. *Nature* 2005;436:266-71.
37. Windrem MS, Schanz SJ, Guo M, Tian GF, Washco V, Stanwood N, et al. Neonatal chimerization with human glial progenitor cells can

- both remyelinate and rescue the otherwise lethally hypomyelinated shiverer mouse. *Cell Stem Cell* 2008;2:553-65.
38. Rasmuson I, Ringden O, Sundberg B, Le Blanc K. Mesenchymal stem cells inhibit the formation of cytotoxic T lymphocytes, but not activated cytotoxic T lymphocytes or natural killer cells. *Transplantation* 2003;76:1208-13.
 39. Le Blanc K, Rasmuson I, Gotherström C, Seidel C, Sundberg B, Sundin M, et al. Mesenchymal stem cells inhibit the expression of CD25 (interleukin-2 receptor) and CD38 on phytohaemagglutinin-activated lymphocytes. *Scand J Immunol* 2004;60:307-15.
 40. Titomanlio L, Kavelaars A, Dalous J, Mani S, El Ghouzzi V, Heijnen C, et al. Stem cell therapy for neonatal brain injury: perspectives and challenges. *Ann Neurol* 2011;70:698-712.
 41. Aggarwal S, Pittenger MF. Human mesenchymal stem cells modulate allogeneic immune cell responses. *Blood* 2005;105:1815-22.
 42. Kode JA, Mukherjee S, Joglekar MV, Hardikar AA. Mesenchymal stem cells: immunobiology and role in immunomodulation and tissue regeneration. *Cytotherapy* 2009;11:377-91.
 43. Jiang XX, Zhang Y, Liu B, Zhang SX, Wu Y, Yu XD, et al. Human mesenchymal stem cells inhibit differentiation and function of monocyte-derived dendritic cells. *Blood* 2005;105:4120-6.
 44. Chang YS, Oh W, Choi SJ, Sung DK, Kim SY, Choi EY, et al. Human umbilical cord blood-derived mesenchymal stem cells attenuate hyperoxia-induced lung injury in neonatal rats. *Cell Transplant* 2009;18:869-86.
 45. Chang YS, Choi SJ, Sung DK, Kim SY, Oh W, Yang YS, et al. Intratracheal transplantation of human umbilical cord blood-derived mesenchymal stem cells dose-dependently attenuates hyperoxia-induced lung injury in neonatal rats. *Cell Transplant* 2011;20:1843-54.
 46. Kim ES, Chang YS, Choi SJ, Kim JK, Yoo HS, Ahn SY, et al. Intratracheal transplantation of human umbilical cord blood-derived mesenchymal stem cells attenuates *Escherichia coli*-induced acute lung injury in mice. *Respir Res* 2011;12:108.
 47. Kim ES, Ahn SY, Im GH, Sung DK, Park YR, Choi SH, et al. Human umbilical cord blood-derived mesenchymal stem cell transplantation attenuates severe brain injury by permanent middle cerebral artery occlusion in newborn rats. *Pediatr Res* 2012;72:277-84.
 48. Ahn SY, Chang YS, Kim SY, Sung DK, Kim ES, Rime SY, et al. Long-term (postnatal day 70) outcome and safety of intratracheal transplantation of human umbilical cord blood-derived mesenchymal stem cells in neonatal hyperoxic lung injury. *Yonsei Med J* 2013;54:416-24.