

### Altered $\beta_{1-3}$ -adrenoceptor influence on $\alpha_2$ -adrenoceptor-mediated control of catecholamine release and vascular tension in hypertensive rats

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Berg T (2015) Altered  $\beta_{1-3}$ -adrenoceptor influence on  $\alpha_2$ -adrenoceptor-mediated control of catecholamine release and vascular tension in hypertensive rats. Front. Physiol. 6:120. doi: 10.3389/fphys.2015.00120  $\alpha_2$ - and  $\beta$ -adrenoceptors (AR) reciprocally control catecholamine release and vascular tension. Disorders in these functions are present in spontaneously hypertensive rats (SHR). The present study tested if  $\alpha_2AR$  dysfunctions resulted from altered  $\alpha_2$ AR/ $\beta$ AR interaction. Blood pressure (BP) was recorded through a femoral artery catheter and cardiac output by an ascending aorta flow probe. Total peripheral vascular resistance (TPR) was calculated. Norepinephrine release was stimulated by a 15-min tyramine-infusion, which allows presynaptic release-control to be reflected as differences in overflow to plasma. Surgical stress activated some secretion of epinephrine. L-659,066 (a2AR-antagonist) enhanced norepinephrine overflow in normotensive controls (WKY) but not SHR. Nadolol ( $\beta_{1+2}$ ) and ICI-118551 ( $\beta_2$ ), but not atenolol ( $\beta_1$ ) or SR59230A [ $\beta_{(3)/1L}$ ] prevented this increase. All  $\beta$ AR antagonists allowed L-659,066 to augment tyramine-induced norepinephrine overflow in SHR and epinephrine secretion in both strains. Inhibition of cAMP-degradation with milrinone and  $\beta_3$ AR agonist (BRL37344) enhanced the effect of L-659,066 on release of both catecholamines in SHR and epinephrine in WKY.  $\beta_{1/2}AR$  antagonists and BRL37344 opposed the L-659,066-dependent elimination of the TPR-response to tyramine in WKY. α<sub>2</sub>AR/βAR antagonists had little influence on the TPR-response in SHR. Milrinone potentiated the L-659,066-dependent reduction of the TPR-response to tyramine. Conclusions:  $\beta_2AR$  activity was a required substrate for  $\alpha_2AR$  auto inhibition of norepinephrine release in WKY.  $\beta_{1+2}AR$  opposed  $\alpha_2AR$  inhibition of norepinephrine release in SHR and epinephrine secretion in both strains. BAR-a2AR reciprocal control of vascular tension was absent in SHR. Selective agonist provoked  $\beta_3$ AR-G<sub>i</sub> signaling and influenced the tyramine-induced TPR-response in WKY and catecholamine release in SHR.

Keywords:  $\alpha_2$ -adrenoceptors,  $\beta$ -adrenoceptors, hypertension, total peripheral vascular resistance, sympathetic nervous system, catecholamine release, spontaneously hypertensive rats

#### Introduction

 $\alpha_2$ - and  $\beta$ -adrenoceptors (AR) comprise the  $\alpha_{2A,B,and C}$  and the  $\beta_{1,2,and 3}$  subtypes. By coupling to inhibitory (G<sub>i</sub>) and stimulatory (G<sub>s</sub>) G-proteins, respectively,  $\alpha_{2A,B,C}$ - and  $\beta_{1+2}AR$  have opposite

effect on adenylyl cyclase, and therefore have reciprocal actions on various functions involved in the control of blood pressure (BP), including the release of catecholamines and vascular tension. The β<sub>3</sub>AR has been shown to couple to G<sub>i</sub> and induce a negative inotropic effect in the heart (Gauthier et al., 1998). Transmitter release from peripheral sympathetic nerves is inhibited by presynaptic  $\alpha_2 AR$ , primarily the  $\alpha_{2A}$ -subtype (Trendelenburg et al., 2003; Berg and Jensen, 2013), and stimulated by presynaptic  $\beta_2 AR$  (Stjarne and Brundin, 1976; Westfall et al., 1979). Also the  $\beta_1 AR$  has recently been demonstrated to enhance norepinephrine release, providing a plausible rational for the antihypertensive action of  $\beta_1$ AR antagonists, the most frequently used  $\beta$ -blockers in the treatment of hypertension (Berg, 2014a).  $\alpha_2 AR$ auto-receptors in addition inhibited adrenal epinephrine secretion in in vivo experiments (Brede et al., 2003; Berg et al., 2012), whereas  $\beta_1$ - or  $\beta_2$ AR antagonists had no effect (Berg, 2014a). a2AR-mediated auto inhibition of neuronal and adrenal catecholamine release has been shown to be dysfunctional in the spontaneously hypertensive rat (SHR) (Berg and Jensen, 2013). This dysfunction may contribute to the hyper adrenergic and hypertensive state in this model of human hypertensive disease, in agreement with the high plasma norepinephrine concentration and hypertension observed in  $\alpha_{2A}$  AR-gene-deleted mice (Makaritsis et al., 1999). The failing  $\alpha_2 AR$  auto inhibition in SHR may result from an altered interaction between different presynaptic receptors, as indicated by the restored  $\alpha_2AR$  function in SHR after a2CAR stimulation or angiotensin AT1 receptor inhibition (Berg, 2013) (Figure 1). The  $\beta_3$ AR has been shown to be less sensitive to catecholamine-induced desensitization than the  $\beta_1$ and  $\beta_2 AR$  (Mallem et al., 2004; Rouget et al., 2004), and a  $\beta_3 AR$ up-regulated and B1AR down-regulated relaxation was demonstrated in SHR thoracic aortic rings (Mallem et al., 2004). It may therefore be hypothesized that alterations in  $\beta$ AR signaling may alter  $\alpha_2 AR$  auto inhibition of catecholamine release in SHR.

 $\alpha_{2B}AR$  (Philipp et al., 2002) and  $\beta AR$  are also present in vascular smooth muscle cells (VSMC), where they modulate the  $\alpha_1AR$ -mediated vasoconstrictory response to norepinephrine (**Figure 2**). VSMC tension is in addition influenced by endothelial  $\alpha_{2A}AR$  and  $\beta_2AR$ , which both stimulate nitric oxide (NO) synthesis (Shafaroudi et al., 2005; Queen et al., 2006). Also vasodilatory and vasoconstrictory  $\alpha_2AR$ -mediated control of total peripheral vascular resistance (TPR) appeared dysfunctional in SHR (Berg and Jensen, 2011, 2013).

Presynaptic receptors modulate norepinephrine release from the nerve terminal vesicles. This control is not reflected by differences in norepinephrine overflow to plasma, due to that the response is terminated by re-uptake through the norepinephrine re-uptake transporter (NET). Presynaptic control of release is therefore not easily studied *in vivo*. However, tyramine stimulates norepinephrine release by reversing the transport through NET (**Figure 1**), and consequently prevents re-uptake through NET. Presynaptic modulation of concomitant vesicular release can therefore be demonstrated as differences in overflow to plasma (Berg et al., 2012; Berg and Jensen, 2013). The presynaptic receptors will be stimulated by the released norepinephrine and/or by other agonists present in the vicinity. Secreted epinephrine is not subjected to re-uptake through NET, and was therefore not influenced by tyramine, but was stimulated to some extent by the stress induced by the experiment itself (Berg et al., 2012). Thus, the inhibitory effect of presynaptic  $\alpha_2 AR$  on catecholamine release could be demonstrated by the ability of the non-selective, peripherally restricted, α<sub>2</sub>AR antagonist L-659,066 to increase tyramine-induced norepinephrine overflow and epinephrine secretion (Berg and Jensen, 2013). The stimulating effect of presynaptic  $\beta$ AR was demonstrated by pre-treatment with  $\beta_{10r2}AR$  antagonists (Berg, 2014a). Inhibition of release could be stimulated in SHR by the β<sub>3</sub>AR agonist BRL37344, compatible with stimulation of  $\beta_3$ AR-G<sub>i</sub> activation (Berg, 2014b). The reduced release observed after the B3AR antagonist SR59230A was likely to result from its inhibitory effect on the G<sub>s</sub>-coupled low-affinity state  $\beta_1 AR$  ( $\beta_{1L}AR$ ) (Berg, 2014b). The use of tyramine also allowed a concomitant examination of the role of postsynaptic  $\alpha_2 AR$  and  $\beta AR$  in the cardiovascular response to the released norepinephrine (Figure 2).

The purpose of the present investigation was therefore to gain a better understanding of the reason underlying the failing  $\alpha_2 AR$  auto-inhibition of catecholamine release in SHR, i.e., to test if there was a difference in the interaction between the  $\alpha_2 AR$  and the three  $\beta AR$  subtypes in this strain compared to that in their normotensive controls (WKY). A second goal was to test if a difference in  $\beta_{1-3}AR$  activity was responsible for the failing  $\alpha_2 AR$  influence on the TPR-response to the tyramine-stimulated norepinephrine release in SHR.

#### **Materials and Methods**

#### **Experimental Procedure**

All experiments were approved by The Norwegian Animal Research Authority (NARA) and conducted in accordance with the Directive 2010/63/EU of the European Parliament. Male, 12-14 weeks old SHR (Okamoto, SHR/NHsd strain,  $273 \pm 2$  g body weight, n = 109) and their normotensive control, i.e., WKY (Wistar Kyoto,  $279 \pm 9$  g body weight, n = 124) on conventional rat chow diet (0.7% NaCl) were anesthetized with sodium pentobarbital (65-70 mg/kg, IP) and tracheotomized. A heparinized catheter was inserted into the femoral artery to record systolic (SBP) and diastolic (DBP) BP. The rats were subsequently connected to a positive-pressure respirator and ventilated with air throughout the experiment. Cardiac output (CO, i.e., minus cardiac flow) and heart rate (HR) were recorded by a flow probe on the ascending aorta, connected to a T206 Ultrasonic Transit-Time Flowmeter (Transonic Systems Inc., Ithaca, NY, USA). After surgery was completed, the arterial catheter was flushed with 0.15 ml heparinized (1000 U/ml) phosphate-buffered saline (PBS; 0.01 M Na-phosphate, pH 7.4, 0.14 M NaCl). Mean arterial BP [MBP = (SBP-DBP/3) + DBP] and TPR (=MBP/CO) were calculated. Body temperature was maintained at  $37 - 38^{\circ}$ C by external heating, guided by of a thermo sensor inserted inguinally into the abdominal cavity. Drugs were dissolved in PBS and administered as bolus injections (0.6-1 ml/kg) through a catheter in the femoral vein, unless otherwise indicated.

#### **Experimental Design**

After a control period of about 10 min, control rats were pre-treated with vehicle (PBS), and subsequently infused with tyramine for 15 min ( $1.26 \mu$ mol/kg/min) (Berg, 2005; Berg and



sympathetic nerve endings. lyramine stimulates norepinephrine release by reverse transport through NET. Consequently, re-uptake through NET is prevented, and presynaptic modulation of vesicular release is reflected as differences in overflow to plasma (Berg et al., 2012; Berg and Jensen, 2013). The release of norepinephrine from secretory granules is activated by adenylyl cyclase, which is stimulated by  $\beta_{1.11,2}$ AR-G<sub>s</sub> and inhibited by  $\alpha_2$ AR-G<sub>i</sub>. In WKY,  $\alpha_2$ AR auto

inhibition required  $\beta_2AR$  activity, but was independent of  $\beta_1AR$  signaling. In SHR, blocking  $\beta_{1,1L,2}AR$  activity allowed  $\alpha_2AR$  inhibition of release. The  $\beta_3AR$ -selective agonist BRL37344 reduced norepinephrine overflow in SHR but not WKY (dotted arrow). The  $\beta_3AR$  antagonist SR59230A reduced overflow apparently due to its ability to inhibit  $\beta_{1L}AR$  and not the  $\beta_3AR$ . The action of antagonists and agonist are indicated. NE, norepinephrine; Pointed arrows, postive effects; Blunted arrows, inhibitory actions.

Jensen, 2013). In the experimental groups, the rats were pretreated with drugs to modify presynaptic and postsynaptic  $\alpha_2 AR$ or  $\beta AR$  signaling, separately or combined.

#### Inhibition of βAR-Signaling

Inhibition of  $\beta$ AR-signaling was achieved by  $\beta$ AR-antagonists, the peripherally restricted, i.e., which does not cross the bloodbrain barrier, nadolol ( $\beta_{1+2}$ , 8.5 µmol/kg), or atenolol ( $\beta_1$ , 5.6 µmol/kg), the not-restricted ICI-118551 ( $\beta_2$ , 1 µmol/kg initial dose, then 0.3 µmol/kg/min throughout the experiment) or the  $\beta_3$ AR antagonist SR59230A (13.8 µmol/kg) (Berg et al., 2010). SR59230A also inhibited the putative  $\beta_4$ AR (Malinowska and Schlicker, 1997), later identified as  $\beta_{1L}$ AR (Kaumann et al., 2001).

#### Amplification of **βAR-Signaling**

Amplification of  $\beta$ AR-signaling was achieved by pre-treatment with the phosphodiesterase 3 (PDE3) inhibitor milrinone (1.4  $\mu$ mol) (Berg et al., 2009) which will prevent degradation of cyclic AMP (cAMP) (**Figure 2**). The  $\beta_3$ AR was stimulated by the agonist BRL37344 ((±)-(R\*,R\*)-[4-[2-[[2-(3-Chlorophenyl)-2-hydroxyethyl]amino]propyl]phenoxy]-acetic acid sodium hydrate) infused at a rate of 1 nmol/kg/min throughout the experiment (Malinowska and Schlicker, 1997; Berg, 2014b).

#### Inhibition of $\alpha_2$ AR-Signaling

Inhibition of  $\alpha_2$ AR-signaling was achieved by pre-treatment with the none-selective, peripherally restricted  $\alpha_2$ AR antagonist L-659,066 (Clineschmidt et al., 1988) (4.4  $\mu$ mol/kg) (Berg et al., 2012; Berg and Jensen, 2013).

#### Inhibition of G<sub>i</sub> Signaling

Since  $G_i$  represents a main signaling pathway for all  $\alpha_2 AR$ ,  $G_i$ -signaling was abolished by the  $G_i$ -inhibitor *Bordetella pertussis* toxin (PTX, 15 µg/kg, i.p., -48 h) (Anand-Srivastava et al., 1987). The latter rats were pre-treated with PBS during the experiment.

#### The Interaction between $\alpha_2$ AR and $\beta$ AR Signaling

The interaction between  $\alpha_2AR$  and  $\beta AR$  signaling was studied by combining  $\beta AR$  antagonists/agonist/milrinone with the  $\alpha_2AR$  antagonist L-659,066 as indicated in **Table 1**. Ten min was allowed between drugs, except for SR59230A which was followed by L-659,066 or tyramine after 5 min.

#### **Measurement of Plasma Catecholamines**

1.5 ml blood was collected from the femoral artery into tubes containing 45  $\mu$ l 0.2 M glutathione and 0.2 M ethylene glycol-bis(2-aminoethylether)-N,N,N',N'-tetraacetic acid (EGTA) (4°C). Plasma was stored at  $-80^{\circ}$ C until catecholamine concentrations were determined using 400  $\mu$ l plasma and the 5000 Reagent kit for HPLC analysis of catecholamines in plasma from chromsystems GmbH, Munich, Germany, as described by the manufacturer. The samples were run on a Shimadzu monoamines analyzer system, using an isocratic flow rate of 0.8 ml/min, and an electrochemical detector (Decade II) and a SenCell electrochemical flow cell (Antec Leyden, Zoeterwoude, The Netherlands).

#### Drugs

L-659,066 was a kind gift from Merck, Sharp and Dohme Labs, Rahway, NJ. ICI-118551 was obtained from ICI-Pharma,



FIGURE 2 | AR-mediated control of tension in VSMC. Inhibition of K<sub>V</sub> induces depolarization which will activate Ca<sup>2+</sup> influx through Ca<sub>V</sub> and thus precipitates vasoconstriction due to a rise in [Ca<sup>2+</sup>]<sub>i</sub>. K<sub>V</sub> is stimulated by cAMP-PKA signaling, and in pathophysiologic conditions such as that in SHR may be inhibited by PLC. The BRL37344-induced  $\beta_3$ AR-G<sub>i</sub> signaling and increased TPR was only observed in WKY during tyramine-induced norepinephrine release. Ca<sub>V</sub>, voltage sensitive Ca<sup>2+</sup> channels; K<sub>V</sub>, voltage sensitive K<sup>+</sup> channels; 4AP, the K<sub>V</sub> inhibitor 4-aminopyridine; PKA, protein kinase A; Arrow, positive action; Blunted arrow, negative action.

Cheshire, UK; SR59230A from Santa Cruz Biotechnology, Heidelberg, Germany; and pentobarbital from The Norwegian National Hospital, Oslo, Norway. The remaining drugs were from Sigma Chemical Co., St. Louis, MO, USA.

#### **Statistical Analyses**

Results are presented as mean values  $\pm$  s.e.m. Effect of pretreatment, differences in the cardiovascular baselines, (data averaged every min), and the plasma catecholamine concentrations were evaluated by overall tests (One-Way ANOVA), followed by two-tailed two-sample Student's t-tests. The cardiovascular response-curves to tyramine (data averaged every min) were analyzed using Repeated Measures Analyses of Variance and Covariance, first as over-all tests, subsequently for each group separately or between two groups. Significant responses and groups differences were subsequently located at specific times using two-tailed one- and two-sample Student's t-tests, respectively. For nonparametric data, two-sample Student's *t*-tests were substituted by Kruskal-Wallis tests. At each step, testing proceeded only when the presence of significant differences and/or interactions was indicated. For the cardiovascular data, the P-value was for all tests and each step adjusted according to Bonferroni, whereas  $P \leq 0.05$  was considered significant for the catecholamine data.

In those cases, where comparisons were made with rats included in previous publications (**Table 1**), these experiments were performed, in part or in full, intermittently with the present groups. When this was not the case, some of the previous experiments were substituted with new to ensure that all experiments overlapped in time. Control rats were included randomly throughout the study.

#### Results

# The Influence of $\beta_{1-3}AR$ Signaling on $\alpha_2AR$ -mediated Inhibition of Tyramine-induced Norepinephrine Overflow to Plasma The Effect of Modulating $\beta AR$ Signaling

As previously described (Berg and Jensen, 2013; Berg, 2014a), all  $\beta$ AR antagonists, including SR59230A, reduced the tyramineinduced overflow of norepinephrine to plasma in both strains (**Figure 3**). Since the effect of SR59230A was likely to involve inhibition of  $\beta_{IL}$ AR rather than  $\beta_3$ AR (Berg, 2014b), this result demonstrated that the overflow was enhanced by  $\beta_{1+1L+2}$ AR activation. However, the  $\beta_3$ AR agonist BRL37344 precipitated a minor reduction in overflow in SHR but not WKY (**Figure 4**), compatible with a G<sub>i</sub>-mediated inhibition of release in SHR (Berg, 2014b).

#### The Effect of inhibiting $\alpha_2 AR$ -G<sub>i</sub> Signaling

The G<sub>i</sub> inhibitor PTX had no significant effect on the tyramineinduced norepinephrine overflow in either strain (**Figure 4**). As before (Berg and Jensen, 2013), the  $\alpha_2$ AR antagonist L-659,066 increased norepinephrine overflow in WKY, but the rise was not statistically significant in SHR (**Figures 3**, **4**).

#### The Interaction between $\alpha_2AR$ and $\beta AR$ Signaling

L-659,066 eliminated the reduction in the tyramine-induced overflow induced by the  $\beta_1$ AR antagonist atenolol in WKY (P = NS compared to the controls or L-659,066 alone,)P = 0.004 compared to atenolol alone) (Figure 3). L-659,066 also totally reversed the reduction following pre-treatment with the  $\beta_{(3)+IL}AR$  antagonist SR59230A (P = 0.005 compared to SR59230A alone, P = NS compared to the controls and L-659,066 alone) (Figure 3). However, L-659,066 counter-acted only in part the reduction following the  $\beta_{1+2}AR$  antagonist nadolol (P < 0.004 compared to the WKY controls and L-659,066 alone,)P = 0.049 compared to nadolol alone), and did not at all counter-act the reduction induced by the  $\beta_2$ -selective antagonist ICI-118551 (P = 0.05 and 0.005 compared to the controls and L-659,066 alone, respectively, P = NS compared to ICI-118551 alone) (Figure 3). These results demonstrated that  $\alpha_2$ AR antagonist increased tyramine-stimulated norepinephrine release in WKY also in the presence of  $\beta_1 AR$  and  $\beta_{(3)+1L}AR$  antagonist, but not when the  $\beta_2$ AR subtype was blocked.

The plasma norepinephrine concentration in L-659,066 + milrinone-treated WKY was higher than that in the controls (P = 0.05) but not different from that after L-659,066 alone (**Figure 4**). This result showed that enhanced cAMP signaling, i.e., down-stream of adenylyl cyclase (**Figure 2**), did not influence the response to L-659,066 in this strain. BRL37344 did not alter the effect of L-659,066 on tyramine-induced norepinephrine release (**Figure 4**), demonstrating that  $\beta_3$ AR-G<sub>i</sub> signaling did not contribute to the response in WKY.

In SHR, L-659,066 eliminated the reduction in tyramineinduced norepinephrine overflow induced by all four  $\beta$ AR antagonists ( $P \leq 0.003$  compared to  $\beta$ AR-antagonist alone), and was higher than that in the controls in all groups ( $P \leq 0.018$ ), except in the ICI-118551 + L-659,066-treated group (P = NS) (**Figure 3**). These results demonstrated that  $\alpha_2$ AR-mediated

Pre-treatment	WKY				SHR			
	MBP	HR	со	TPR	MBP	HR	со	TPR
	mm Hg	beats/min	ml/min	mm Hg/ml/min	mm Hg	beats/min	ml/min	mm Hg/ml/min
PBS (control) <sup>b</sup>	66±3	$341 \pm 8$	32±2	$2.1 \pm 0.1$	92±5*	391±5*	19±1*	$4.6 \pm 0.2^{\star}$
	(-3±2)	(-6 ± 4)	(2±0)	(-0.3 ± 0.1)	(-5±5)	(-16±5)	(0±1)	(-0.3 ± 0.2)
INHIBITION OF β	AR SIGNALING							
Nadolol	65±2	338±5	$31 \pm 2$	$2.1 \pm 0.1$	$64 \pm 5^{\dagger}$	$345 \pm 10^{\dagger}$	$13 \pm 2^{\dagger}$	$4.6 \pm 0.5$
(β <sub>1+2</sub> ANT) <sup>b</sup>	(-3±1)	(-21±6)	(1 ± 1)	(-0.2 ± 0.0)	$(-25 \pm 2)^{\dagger}$	$(-80 \pm 7)^{\dagger}$	(-1 ± 1)	$(-2.0 \pm 0.8)^{\dagger}$
Atenolol	61 ± 2	$338 \pm 9$	$28 \pm 2$	$2.4 \pm 0.1$	$63 \pm 3^{\dagger}$	$338 \pm 9$	$13 \pm 1^{\dagger}$	5.1 ± 0.4
(β1ANT) <sup>b</sup>	(-12 ± 1)	$(-32 \pm 7)^{\dagger}$	(1 ± 1)	(-0.5 ± 0.1)	$(-32 \pm 5)^{\dagger}$	$(-69 \pm 10)^{\dagger}$	(-4 ± 1)	(-0.4 ± 0.5)
ICI-118551	$50 \pm 4$	$311 \pm 12 \ (-31 \pm 4)^{\dagger}$	$28 \pm 4$	$1.8 \pm 0.1$	$70 \pm 4$	$358 \pm 7^{\dagger}$	$18 \pm 1$	$4.3 \pm 0.2$
(β <sub>2</sub> ANT) <sup>b</sup>	(-6 ± 2)		(2 ± 1)	(-0.4 ± 0.1)	(-23 ± 4)	$(-59 \pm 8)^{\dagger}$	(1 ± 1)	(-1.2 ± 0.2)
SR59230A	$84 \pm 4^{\dagger}$	$390 \pm 8^{\dagger}$	$27 \pm 2$	$3.3 \pm 0.3^{\dagger}$	$90 \pm 8$	424±13	18±2	5.1 ± 0.4
(β <sub>3</sub> + β <sub>1L</sub> ANT) <sup>b</sup>	$(10 \pm 3)^{\dagger}$	$(30 \pm 7)^{\dagger}$	(4 ± 1)	(-0.2 ± 0.2)	(-6 ± 5)	(-21±5)	(0±1)	(-0.5 ± 0.3)
STIMULATION OF	THE βAR SIGN	ALING PATHWA	(					
Milrinone (PDE3	$40 \pm 2^{\dagger}$	343±7	$30 \pm 2$	$1.4 \pm 0.1^{\dagger}$	$52 \pm 3^{\dagger}$	413±14	$18 \pm 1$	$2.9 \pm 0.2^{\dagger}$
INH)	$(-35 \pm 1)^{\dagger}$	(2±8)	(-2 ± 1)	$(-1.0 \pm 0.1)^{\dagger}$	$(-59 \pm 7)^{\dagger}$	(−1±9)	(-4 ± 1)	$(-2.4 \pm 0.5)^{\dagger}$
BRL44408	51 ± 2 <sup>†</sup>	312±9	$44 \pm 4$	$\begin{array}{c} 1.2\pm 0.1^{\dagger} \\ (-0.6\pm 0.1)^{\dagger} \end{array}$	$109 \pm 7$	409±15	$23 \pm 1^{\dagger}$	$4.8 \pm 0.3$
(β <sub>3</sub> AGON) <sup>b</sup>	(3 ± 3)	(-8±9)	(9 ± 2) <sup>†</sup>		(-9 ± 7)	(0±9)	$(4 \pm 1)^{\dagger}$	(-0.4 ± 0.3)
INHIBITION OF $\alpha_2$	2AR-G <sub>i</sub> SIGNALI	ING						
PBS after PTX	52±3	338±9	$39 \pm 4$	$1.3 \pm 0.2^{\dagger}$	$65 \pm 5$	$389 \pm 12$	22±3	$3.3 \pm 0.5$
(G <sub>i</sub> INH) <sup>a</sup>	(-2±2)	(-21±4)	(3 ± 1)	(-0.1 ± 0.1)	(-8 ± 6)	(-24 ± 11)	(-1±2)	(-0.5 ± 0.2)
L-659,066	$59 \pm 7$	345±11	35±2	$1.7 \pm 0.1$	$69 \pm 6^{\dagger}$	$398 \pm 12$	17±1	4.3±0.4
(α <sub>2</sub> ANT) <sup>b</sup>	$(-17 \pm 2)^{\dagger}$	(-10±5)	(2±0)	$(-0.6 \pm 0.0)^{\dagger}$	$(-21 \pm 4)^{\dagger}$	(-16 ± 10)	(-1±0)	(-1.1±0.2)
INTERACTION BE		ND $\alpha_2$ AR SIGNAL	ING					
Nadolol +	$47 \pm 6^{\ddagger}$	$323 \pm 8$	23±3	$2.0 \pm 0.1$	$61 \pm 4^{\dagger}$	$356 \pm 8^{\dagger}$	$15 \pm 2$	4.4 ± 0.4
L-659,066	$(-6 \pm 2)^{\dashv}$	(-30 ± 10)	(3±2)	(-0.7 ± 0.2)	(-18 ± 7)	$(-69 \pm 9)^{\dagger -1}$	(-1 ± 1)	(-0.8 ± 0.6)
Atenolol +	$56 \pm 5$	$340 \pm 10$	27 ± 4	$2.2 \pm 0.2$	$67 \pm 5$	$363 \pm 8$	$14 \pm 1$	$4.7 \pm 0.3$
L-659,066	$(-18 \pm 5)^{\dagger}$	$(-52 \pm 18)^{\dagger}$	(5 ± 2)	$(-1.1 \pm 0.3)^{\dagger}$	$(-35 \pm 4)^{\dagger \dashv}$	$(-90 \pm 7)^{\dagger -1}$	(0 ± 1)	$(-2.3 \pm 0.2)^{\uparrow \ddagger \dashv}$
ICI-118551 +	$49 \pm 4$	$336 \pm 7$	$26 \pm 1$	$1.9 \pm 0.2$	$77 \pm 7$	$356 \pm 4^{\dagger}$	$19 \pm 1$	$4.0 \pm 0.1^{\dagger}$
L-659,066	$(-26 \pm 3)^{\dagger \ddagger}$	$(-39 \pm 7)^{\dagger \dashv}$	(1 ± 1)	$(-1.2 \pm 0.1)^{\dagger -1}$	(-16 ± 9)	$(-68 \pm 1)^{\dagger -1}$	(0 ± 1)	(-1.1 ± 0.2)
SR59230A +	$55 \pm 6^{\ddagger}$	364±25	28±3	$2.0 \pm 0.2^{\ddagger}$	88±9	408±11	$19\pm 2$	4.7 ± 0.4
L-659,066	$(-3 \pm 4)^{\ddagger}$	(23±9) <sup>⊣</sup>	(2±3)	(-0.3 ± 0.2)	(-3±9)	(2±9)	(-1 ± 1)	(0.2 ± 0.4)
Milrinone +	$37 \pm 1^{\dagger}$	$366 \pm 12$	28±2	$1.3 \pm 0.1^{++}$	$39 \pm 2^{\dagger \ddagger \dashv}$	$431 \pm 9^{\dagger}$	$19 \pm 1$	$2.2 \pm 0.2^{+-1}$
L-659,066	$(-42 \pm 6)^{\dagger}$	(-5 ± 5)	(1±2)	$(-1.5 \pm 0.2)^{+}$	$(-62 \pm 6)^{\dagger \dashv}$	(0 ± 15)	(-2 ± 1)	$(-3.0 \pm 0.4)^{+-1}$
BRL44408 +	$41 \pm 4^{\dagger}$	379±11 <sup>‡</sup>	45±8	$1.0 \pm 0.1^{\dagger}$	$77 \pm 8^{\ddagger}$	407±5	$25 \pm 1^{+}$	$3.1 \pm 0.2^{\ddagger \ddagger}$
L-659,066	$(-28 \pm 6)^{\dagger \dashv}$	(20±9)	(10±7)	$(-1.0 \pm 0.1)^{\dagger \ddagger \dashv}$	$(-33 \pm 9)^{\dagger \ddagger}$	(2±6)	(4 ± 2)	$(-2.4 \pm 0.5)^{\ddagger \mp \dashv}$

<sup>a</sup> Since PTX was given 4 h prior to the experiment, its effect was reflected as differences in the baselines themselves. Comparisons were made between WKY and SHR controls (\* after SHR-values) and between the PBS-controls and the experimental groups, using the response to PBS to evaluate the effect of pre-treatment (<sup>†</sup>). Comparisons were also made between groups pre-treated with  $\beta$ AR-antagonist +  $\alpha_2$ AR-antagonist and corresponding groups pre-treated with  $\beta$ AR-antagonist alone (<sup>‡</sup>) or  $\alpha_2$ AR-antagonist alone (<sup>†</sup>).

<sup>b</sup> The rats in these groups were in part or in full the same as in (Berg and Jensen, 2013; Berg, 2014a,b). Six to ten rats were included in each group, except for the WKY and SHR controls and the SHR ICI-118551 and L-659,066 groups which comprised 17, 18, 13, and 13 rats, respectively. \*, P ≤ 0.005; <sup>†</sup>, P ≤ 0.0036; <sup>‡</sup>, <sup>-1</sup>, P ≤ 0.0083.

auto inhibition of norepinephrine release was opposed by both  $\beta_{1+1L}AR$  and  $\beta_2AR$  activity in SHR, with the  $\beta_1AR$  having the greatest impact. Norepinephrine overflow was also increased after pre-treatment with L-659,066 + milrinone ( $P \leq 0.001$  compared to the control, milrinone- or L-659,066-only groups), showing that  $\alpha_2AR$  auto inhibition was enhanced by inhibition of cAMP degradation. In addition, the effect of L-659,066 on overflow was higher when combined with the  $\beta_3AR$  agonist BRL37344 ( $P \leq 0.006$  compared to the controls and after BRL37344 or L-659,066 alone) (**Figure 4**).

## The Effect of PTX, $\beta_{1-3}AR$ Antagonists, Milrinone, and $\beta_3AR$ Agonist on $\alpha_2AR$ -mediated Inhibition of Epinephrine Secretion

#### The Effect of Modulating **BAR** Signaling

The  $\beta$ AR antagonists (**Figure 3**), milrinone and  $\beta_3$ AR agonist (**Figure 4**) had by themselves no effect on the concentration of epinephrine in plasma collected at the end of the experiment in either strain, except for a minor reduction after SR59230A (**Figure 3**) and an increase after milrinone (**Figure 4**) in SHR ( $P \le 0.04$ ).



#### The Effect of Inhibiting $\alpha_2$ AR-G<sub>i</sub> Signaling (Figure 4)

PTX clearly increased the secretion of epinephrine in both strains ( $P \leq 0.032$ ), demonstrating a tonic G<sub>i</sub>-mediated inhibition of epinephrine release. Some increase in the plasma epinephrine concentration was observed in L-659,066-pre-treated WKY (P = 0.048) and, in this collection of animals, also in the L-659,066-pre-treated SHR (P = 0.004). This observation demonstrated that part of the tonic inhibition of release was due to  $\alpha_2$ AR activity.

#### The Interaction between α<sub>2</sub>AR and βAR Signaling

However, when L-659,066 was combined with  $\beta$ AR-antagonist (**Figure 3**), the plasma epinephrine concentration was for all groups in both strains higher than that in the corresponding controls or after each drug alone ( $P \leq 0.05$ ). These results demonstrated that  $\beta$ AR signaling interfered with  $\alpha_2$ AR auto inhibition of epinephrine secretion. The only exceptions were the WKY SR59230A + L-659,066-pre-treated group where the plasma concentration was not different from that in the controls or after each antagonist alone, and the SHR ICI-118551 + L-659,066 group, which was not different from that after L-659,066 alone (**Figure 3**). A potentiated effect of L-659,066 was also seen when L-659,066 was combined with milrinone or the  $\beta_3$ AR-agonist BRL37344 (**Figure 4**).



### The Role of $\beta_{1-3}AR$ and $\alpha_2AR$ in the Control of Cardiovascular Baselines (Table 1)

As previously documented (Berg et al., 2010; Berg, 2014a), nadolol, atenolol, and ICI-118551 reduced HR in WKY, and MBP, HR and TPR baselines in SHR, although the difference was not statistically significant for all. SR59230A increased MBP and HR in WKY but had no effect in SHR (Berg, 2014b). Milrinone alone had no significant effect on baseline HR or CO, but reduced MBP and TPR in both strains ( $P \leq 0.003$  compared to the controls). BRL37344 itself reduced TPR baseline in WKY and increased CO baseline in both strains (Berg, 2014b). Baselines in rats pretreated with the G<sub>i</sub> inhibitor PTX were not significantly different from that in the controls ( $P \geq 0.0036$ ), except for a reduced TPR in WKY. L-659,066 reduced baseline MBP in both strains and TPR significantly in WKY only.

After nadolol/atenolol/ICI-118551 + L-659,066, changes in the cardiovascular baselines in WKY were largely the same as the combined effect of that observed after the  $\beta$ AR and  $\alpha_2$ AR antagonists alone, except for a greater fall in TPR after ICI-118551 + L-659,066 ( $P \leq 0.005$  compared to that after each antagonist alone). In SHR, TPR baseline was reduced after atenolol + L-659,066 ( $P \leq 0.008$  compared to that after each antagonist alone), whereas the reduction following nadolol alone was not observed after nadolol + L-659,066. The TPR-response to milrinone + L-659,066 was not different from that after milrinone alone in either strain, but was greater than that after

L-659,066 alone in SHR ( $P \leq 0.005$ ). After BRL37344 + L-659,066, the changes seen after BRL37344 or L-659,066 alone remained, but in WKY there was a further reduction in TPR ( $P \leq 0.008$  compared to BRL37344 or L-659,066 alone). L-659,066 had no effect on HR or CO baselines in either strain, and did not alter the HR-response to  $\beta$ AR antagonist, milrinone or  $\beta_3$ AR agonist.

## The Influence of $\beta_{1-3}AR$ and $\alpha_2AR$ on the TPR-response to Tyramine-stimulated Norepinephrine Release

In agreement with previous studies (Berg et al., 2010; Berg and Jensen, 2013), the tyramine-induced release of norepinephrine activated a transient rise in TPR (**Figures 5**, **6**) and a sustained increase in HR (**Figures 7**, **8**), MBP and CO (not shown) in both strains. Pre-treatment with PTX or L-659,066 eliminated the TPR-response to tyramine in WKY ( $P \le 0.001$  compared to WKY controls,P = NS for single curve evaluation), and in SHR reduced the TPR-peak response ( $P \le 0.008$ ), but had no effect on the later response (**Figure 5**). Milrinone reduced the TPR-peak response in both strains (**Figure 5**). After pre-treatment with milrinone + L-659,066, the tyramine-induced vasoconstriction was reversed to a vasodilatory response in WKY ( $P \le 0.025$  compared to the control and milrinone-only groups at 3 and 15 min), and eliminated in SHR (P = NS, single curve evaluation) (**Figure 5**). When the  $\beta_3$ AR agonist BRL37344 was given

prior to L-659,066, the TPR-response to tyramine was higher than that after L-659,066 alone in WKY (P = 0.024 at 15 min) but not different in SHR (**Figure 5**).

The L-659,066-dependent elimination of the TPR-response to tyramine in WKY was in part reversed by prior administration of nadolol, atenolol or ICI-118551, with a non-additive effect of  $\beta_1$ - and  $\beta_2AR$  blockade (**Figure 6**). Significant differences were not detected between the L-659,066 and the  $\beta AR$  antagonist + L-659,066 pre-treated groups in SHR (**Figure 6**). When the  $\beta_{3+1L}AR$  antagonist SR59230A was combined with L-659,066, the TPR-response to tyramine was not altered in WKY, whereas the vasoconstriction developed more slowly than after L-659,066 alone in SHR (**Figure 6**).

## The Influence of $\beta_{1-3}AR$ and $\alpha_2AR$ on the Tyramine-induced Tachycardia

The tyramine-induced tachycardia was not influenced in either strain by inhibition of  $\alpha_2$ AR-G<sub>i</sub>-signaling by L-659,066 or PTX, or by inhibition of cAMP-degradation by milrinone or stimulation of  $\beta_3$ AR with BRL37344, either alone or combined with L-659,066 (**Figure 7**). The only exception was milrinone + L-659,066 which halved the HR-response in SHR ( $P \le 0.028$ ) (**Figure 7**). As after nadolol alone (Berg et al., 2010), nadolol + L-659,066 eliminated the tachycardia in both strains (**Figure 8**).  $\Delta$ HR was clearly reduced after atenolol + L-659,066 and to some extent also after SR59230A + L-659,066 in both strains





(Figure 8), not different from that previously observed after atenolol and SR59230A alone (Berg et al., 2010). Different from that previously documented for ICI-118551 alone, i.e., no effect on the HR-response to tyramine in WKY and a slightly reduced response in SHR (Berg et al., 2010), ICI-118551 + L-659,066 reduced the tachycardia in WKY, but slightly increased the response in SHR (Figure 8). The effect of these drugs on the HR-response to tyramine was not paralleled by similar changes in the plasma catecholamine concentrations.

#### Discussion

The main results in the present study were: (1) The  $\alpha_2AR$  antagonist L-659,066 required the presence of  $\beta_2AR$  activity to enhance tyramine-induced norepinephrine overflow in WKY but was independent of  $\beta_1AR$  signaling. (2)  $\alpha_2AR$ -mediated auto inhibition of norepinephrine release in SHR and epinephrine secretion in both strains was opposed primarily by  $\beta_{1+1L}AR$  but also by  $\beta_2AR$  activity. (3)  $\alpha_2AR$  and  $\beta_{1+2}AR$  reciprocally modulated the TPR-response to the released norepinephrine in WKY but not in SHR. (4) In the presence of L-659,066,  $\beta_3AR$  agonist stimulated vasoconstriction in WKY and  $\alpha_2AR$  auto inhibition in SHR during tyramine-induced norepinephrine release.

The reduction in tyramine-induced norepinephrine overflow to plasma after the  $\beta_1 AR$  antagonist atenolol was eliminated by

additional pre-treatment with L-659,066 in WKY. The same was observed with the  $\beta_{3+1L}AR$  antagonist SR59230A, most likely due to its ability to inhibit  $\beta_{1L}AR$  (Berg, 2014b) (Figure 1). L-659,066 therefore increased release independent of  $\beta_{1+1L}AR$ activity. However, the reduction induced by  $\beta_2AR$  blocking antagonist, i.e., nadolol or ICI-118551, was somewhat or not at all reversed by L-659,066, respectively. It therefore appeared that  $\beta_2$ AR-induced stimulation of release was a required substrate for  $\alpha_2AR$  auto inhibition in WKY. Since L-659,066 alone increased the tyramine-induced norepinephrine overflow in WKY, the  $\beta_2 AR$  were evidently active in stimulating release. However, amplifying the response to  $\beta_2 AR-G_s$  signaling by inhibiting PDE3-induced cAMP degradation with milrinone, did not enhance the effect of L-659,066 in WKY. Also the G<sub>i</sub> inhibitor PTX did not alter norepinephrine overflow. These results may suggest that  $\alpha_2$ AR-G<sub>i</sub> inhibition of norepinephrine release was activated in balance with  $\beta_2$ AR-G<sub>s</sub> stimulation of release. This balance may possibly result from a cAMP-PKA-dependent switch in  $\beta_2$ AR-signaling from G<sub>s</sub> to G<sub>i</sub> (Daaka et al., 1997) when cAMP reached a certain level.

In SHR, where L-659,066 did not increase tyramine-induced norepinephrine overflow, additional pre-treatment with  $\beta_1$ - as well as  $\beta_2AR$  blockade allowed L-659,066 to increase the plasma norepinephrine concentration. Also SR59230A + L-659,066 increased norepinephrine overflow, most likely due



to inhibition of the release-stimulating  $\beta_{1L}AR$ . Thus, both  $\beta_{1 \text{ and } 1L}AR$  and  $\beta_2AR$  counter-acted  $\alpha_2AR$ -mediated inhibition of norepinephrine release in SHR, different from that in WKY. It therefore appeared that excessive  $\beta_{1+2}AR$  activity interfered with the inhibitory effect of  $\alpha_2AR$ -G<sub>i</sub> on adenylyl cyclase in this strain.

Surprisingly, L-659,066 combined with the PDE3 inhibitor milrinone doubled norepinephrine release in SHR, even though milrinone alone had no effect. Thus, accumulation of cAMP after preventing its degradation greatly enhanced release only when the inhibitory action of  $\alpha_2$ AR-G<sub>i</sub> on adenylyl cyclase was prevented (**Figure 1**). The fact that this was observed in SHR only was compatible with the augmented  $\beta$ AR hampering of  $\alpha_2$ AR auto inhibition in this strain.

Augmented norepinephrine overflow was also observed in SHR when L-659,066 was combined with BRL37344. Since BRL37344 may also activate  $\beta_{1+2}AR$ -G<sub>s</sub> signaling (Cernecka et al., 2014), this observation may be explained by a direct stimulating effect on adenylyl cyclase and subsequent norepinephrine release. However, as will be discussed below, BRL37344 in the presence of L-659,066 induced vasoconstriction in WKY, compatible with a stimulating effect on the G<sub>i</sub>-coupled  $\beta_3AR$ . The potentiating effect of BRL37344 on release in SHR may therefore also result from  $\beta_3AR$ -G<sub>i</sub> stimulation, strengthening that activated by  $\alpha_2AR$  (**Figure 1**).

The secretion of epinephrine was tonically down-regulated by G<sub>i</sub> in both WKY and SHR, indicated by the 6-7 times increase in the plasma epinephrine concentration in PTX-treated rats. A similar increase was seen in rats without tyramine-stimulated norepinephrine release (Berg et al., 2012). Although the plasma epinephrine concentration after pre-treatment with L-659,066 was less than that after PTX, the difference was not statistically significant, suggesting  $\alpha_2$ AR auto-inhibition of epinephrine secretion to be an important although not the only regulator of G<sub>i</sub> in these cells. The secretion of epinephrine was not reduced by milrinone or  $\beta_{1/1L/2}AR$  blockade alone. It therefore appeared that  $\alpha_2 AR$  tonically inhibited the adrenal secretion of epinephrine in both strains with little  $\beta$ AR influence. In spite of this,  $\beta_{1/2}AR$  blockade in both strains and also  $\beta_{1L}AR$  antagonist in SHR potentiated the effect of L-659,066 on epinephrine secretion, apparently with a greater effect of the  $\beta_1$ - than the  $\beta_2$ AR in both strains. Thus, different from that observed for norepinephrine release in WKY but similar to that in SHR,  $\beta_1$ - and  $\beta_2AR$  in both strains and also  $\beta_{1L}AR$  in SHR opposed  $\alpha_2$ AR-mediated inhibition of the secretion of epinephrine. Like tyramine-stimulated norepinephrine release in SHR but not in WKY, the effect of L-659,066 on the secretion of epinephrine was further enhanced when combined with milrinone or BRL37344 in both strains, most likely through the same mechanisms as discussed above.



The same receptors which presynaptically control catecholamine release are also located postsynaptically and modulate vascular tension and heart rate. Tyramine induces a massive release of norepinephrine without the normal physiological termination of the response by synaptic norepinephrine re-uptake through NET. One may therefore expect the concentration of norepinephrine within the synapse to be more than sufficient to maximally stimulate the postsynaptic receptors in all groups even though norepinephrine release differed. Differences in the cardiovascular response were therefore likely to be primarily due to drug influence on the postsynaptic receptors rather than reflect differences in release due to drug effect on the presynaptic receptors. This conclusion was supported by the fact that the tyramine-induced tachycardia was clearly hampered by  $\beta_1$ - and  $\beta_2$ AR antagonists also in the presence of L-659,066 which greatly increased the level of circulating catecholamines. The effect of tyramine does not depend on neuronal action potentials and is therefore not directly influenced by differences in neuronal activity. Due to the anesthesia, the cardiovascular response to tyramine was not modified by activation of baroreflexes, demonstrated by that the HR-response to tyramine was not influenced by atropine (Berg and Jensen, 2013). Moreover, large changes in BP induced by bradykinin or phenylephrine had no effect on HR in similarly anesthetized rats of both strains (Bjørnstad-Østensen and Berg, 1994; Berg et al., 2012).

Norepinephrine release was also not much influenced by the ganglion blocker hexamethonium, but being a nicotine receptor antagonist; it clearly reduced epinephrine secretion in both strains (Berg, 2014a). The vasoconstrictory TPR-response to the tyramine-induced norepinephrine release was due to  $\alpha_1$  AR activation since it was totally abolished by prazosin (Berg et al., 2010). However, concomitant vascular  $\alpha_2$ AR- $\beta$ AR-activation will modulate this response, and this modulation differed in the two strains. In WKY, the norepinephrine-induced vasoconstriction was totally eliminated by PTX and L-659,066, showing that  $\alpha_2$ AR-G<sub>i</sub>-signaling was a major preserver of the  $\alpha_1$ AR-mediated vasoconstriction in this strain. This support was due to that  $\alpha_2$ AR-G<sub>i</sub>-signaling opposed  $\beta_1$ - and  $\beta_2$ AR-mediated vasodilatation, indicated by that  $\beta_1$ - or  $\beta_2$ AR antagonist prevented in part the L-659,066-dependent elimination of TPR-response to tyramine in WKY. Furthermore, accumulation of cAMP after pretreatment with milrinone clearly reduced the peak-response to tyramine, and, in addition, potentiated the effect of L-659,066, thus precipitating a vasodilatory response to tyramine in milrinone + L-659,066-treated WKY. SR59230A did not alter the TPR-response to tyramine in WKY (Berg, 2014b) or in L-659,066-treated WKY, showing that  $\beta_{3/1L}AR$  were not active and did not influence this response. Thus, when  $\alpha_2AR$ -mediated inhibition of adenylyl cyclase was prevented by L-659,066, an increased  $\beta_{1+2}AR$ -dependent vasodilatation in response to the released norepinephrine and/or epinephrine was allowed. When in addition the degradation of cAMP was blocked, this effect was further enhanced (**Figure 2**). Thus,  $\alpha_2$ AR- $\beta$ AR-modulation of the  $\alpha_1$ AR-mediated vasoconstriction was clearly functional in WKY.

In SHR, PTX, and L-659,066 reduced the TPR-peak-response but not the later response to tyramine, but, different from that in WKY, the TPR-response was not eliminated, and was not enhanced by additional pre-treatment with  $\beta_{1,1L,2}AR$  antagonist. The slight delay observed in the development of the TPR-response in SHR pre-treated with SR59230A + L-659,066, may possibly result from inhibition of B3AR-Gi-signaling. Thus, under the present conditions, there was little interaction between  $\alpha_2 AR$  and  $\beta AR$  in the control of vascular tension in SHR. In pathophysiological conditions, including hypertension, enhanced activation of the phospholipase C (PLC)-protein kinase C pathway may lead to inhibition of vasodilatory voltagesensitive  $K^+$  channels ( $K_V$ ) (Ko et al., 2010) (Figure 2). The presence of such inhibition in SHR was in fact confirmed by that antagonists against  $\alpha_1$ AR or angiotensin AT<sub>1</sub> and ET<sub>A</sub> receptors, which all activate PLC, enhanced the acute vasoconstrictory TPRresponse to the K<sub>V</sub> inhibitor 4-aminopyridine in SHR but not WKY (Berg, 2003). Since cAMP-induced vasodilatation may be mediated through a protein kinase A (PKA)-dependent opening of K<sub>V</sub> (Aiello et al., 1998), the PLC-dependent inhibition of  $K_V$  in SHR was therefore likely to interfere with the  $\alpha_2AR$ - $G_i/\beta AR-G_s$ -cAMP control of vascular tension (Figure 2). A PLCdependent inhibition of K<sub>V</sub> in SHR may therefore explain the absence of  $\alpha_2$ AR- and  $\beta$ AR-modulation of the TPR-response to tyramine-stimulated norepinephrine release in this strain. However, when  $\alpha_2$ AR-G<sub>i</sub>-signaling was prevented by L-659,066, and, at the same time, cAMP-signaling was amplified by milrinone, cAMP-mediated vasodilatation dominated the vascular tension response also in SHR. Thus, milrinone + L-659,066 eliminated tyramine-induced vasoconstriction in SHR. However, the effect was still less than that in WKY where milrinone + L-659,066 precipitated a tyramine-induced vasodilatation.

When the  $\beta_3AR$  were stimulated with the agonist BRL37344 in WKY, the inhibitory effect of L-659,066 on the TPR-response to tyramine was reversed, with a stronger effect as catecholamine release progressed. This effect of BRL37344 could not be explained by its weak  $\beta_{1+2}AR$  agonistic effect (Dolan et al., 1994), and BRL37344 did not interact with the putative  $\beta_4AR$ (Malinowska and Schlicker, 1997), later identified as the  $\beta_{1L}AR$ (Granneman, 2001; Kaumann et al., 2001). Since the  $\beta_3AR$  is more resistant to catecholamine-induced desensitization than  $\beta_{1/2}AR$  in human tissue (Wallukat, 2002; Rouget et al., 2004),

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this subtype may play a more prominent role during prolonged, high levels of norepinephrine such as during the late part of the tyramine-infusion period, particularly when combined with selective agonist. This vasoconstrictory component was likely to be mediated through  $\beta_3 AR$ -G<sub>i</sub> signaling.

The tyramine-induced tachycardia was reduced after  $\beta_1$ -,  $\beta_2$ -, and  $\beta_{1L(3)}AR$  antagonist in WKY also in the presence of L-659,066, apparently due to inhibition of postsynaptic  $\beta AR$ , independent of changes in norepinephrine release. In SHR, the tachycardia was reduced after L-659,066, halved after milrinone + L-659,066 and slightly increased after ICI-118551 + L-659,066. The reason for these changes was not obvious, but may result from receptor desensitization in the two former groups, and a possible switch from G<sub>s</sub> to G<sub>i</sub> for the  $\beta_2 AR$  (Daaka et al., 1997) in the latter group.

#### Conclusions

a2AR-mediated inhibition of norepinephrine release required the presence of  $\beta_2 AR$  in WKY, but was independent of  $\beta_1 AR$ activity. The balanced  $\alpha_2 AR - \beta_2 AR$  interaction in WKY may function to prevent excessive norepinephrine release during physiological conditions with increased epinephrine secretion such as hypoglycemia and exercise, since epinephrine is a better agonist for the  $\beta_2AR$  subtype than norepinephrine. In SHR, a2AR inhibition of norepinephrine release was counteracted by  $\beta_1 AR$  and  $\beta_2 AR$  activity, with an apparently stronger effect of the former. Although an a2AR-Gi tonic inhibition dominated the control of epinephrine secretion in both strains, their function was counter-acted by  $\beta_2AR$  and even more by  $\beta_1 AR$  in both strains. The more prominent role of  $\beta_1$ AR in counter-acting  $\alpha_2$ AR auto inhibition of catecholamine release in SHR may explain why B1AR blockers are useful as antihypertensive medication and protective in myocardial infarction and heart failure. The a1AR-mediated, vasoconstrictory TPR-response during tyramine-stimulated norepinephrine release was modulated by  $\alpha_2 AR$  and  $\beta_{1/2} AR$  in WKY. The latter interaction was not functional in SHR, most likely due to a PLC-dependent, reduced K<sub>V</sub> vasodilatory influence on VSMC tension, a substrate for cAMP-induced vasodilatation.

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**Conflict of Interest Statement:** The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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