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Kinin B₁ receptors contributes to acute pain following minor surgery in humans

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Abstract

Background: Kinins play an important role in regulation of pain and hyperalgesia after tissue injury and inflammation by activating two types of G-protein-coupled receptors, the kinin B₁ and B₂ receptors. It is generally accepted that the B₂ receptor is constitutively expressed, whereas the B₁ receptor is induced in response to inflammation. However, little is known about the regulatory effects of kinin receptors on the onset of acute inflammation and inflammatory pain in humans. The present study investigated the changes in gene expression of kinin receptors and the levels of their endogenous ligands at an early time point following tissue injury and their relation to clinical pain, as well as the effect of COX-inhibition on their expression levels.

Results: Tissue injury resulted in a significant up-regulation in the gene expression of B₁ and B₂ receptors at 3 hours post-surgery, the onset of acute inflammatory pain. Interestingly, the up-regulation in the gene expression of B₁ and B₂ receptors was positively correlated to pain intensity only after ketorolac treatment, signifying an interaction between prostaglandins and kinins in the inflammatory pain process. Further, the gene expression of both B₁ and B₂ receptors were correlated. Following tissue injury, B₁ ligands des-Arg⁹-BK and des-Arg¹⁰-KD were significantly lower at the third hour compared to the first 2 hours in both the placebo and the ketorolac treatment groups but did not differ significantly between groups. Tissue injury also resulted in the down-regulation of TRPV1 gene expression at 3 hours post-surgery with no significant effect by ketorolac treatment. Interestingly, the change in gene expression of TRPV1 was correlated to the change in gene expression of B₁ receptor but not B₂ receptor.

Conclusions: These results provide evidence at the transcriptional level in a clinical model of tissue injury that up-regulation of kinin receptors are involved in the development of the early phase of inflammation and inflammatory pain. The up-regulation of B₁ receptors may contribute to acute inflammatory pain through TRPV1 activation.

Background

Tissue injury results in the liberation of various pain and inflammatory mediators; we have reported earlier in the oral surgery model of acute inflammatory pain the production or up-regulation of a number of prostanoids, cytokines and chemokines [1-4] following tissue injury. Hargreaves and colleagues have also shown an increase in bradykinin (BK) concentration following third molar tooth extraction [5]. Further, they showed that the NSAID flurbiprofen prevented this increase in BK levels.

Bradykinin-related peptides, collectively known as kinins, are proinflammatory mediators that mediate vascular responses and pain following tissue injury. Kinins

bind to two types of G protein-coupled receptors, the B₁ and B₂ receptors, both of which have been cloned [6,7]. B₁ receptors are activated by the endogenous kinins lacking the carboxy-terminal Arg residue, namely des-Arg⁹-BK and Lys-des-Arg⁹-BK, also known as des-Arg¹⁰-KD. B₂ receptors are activated by the full sequence of the endogenous kinins BK, and Lys-BK, also known as kallidin (KD) [8]. B₂ receptors are constitutively expressed but undergo extensive desensitization by their agonists. They are widely distributed and mediate most of the biological actions of BK. On the other hand, B₁ receptors are induced during the inflammatory processes or at least strongly regulated, except in the spinal cord, where they are constitutively expressed in both rat and man [9]. Further, B₁ receptors are only subjected to limited desensitization, which make them a

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better target for analgesics [10,11]. In experimental studies, the expression of B₁ receptors has been reported to occur in response to the B₁ ligand Lys-des-Arg⁹-BK [12] and inflammatory cytokines such as IL-1 β and TNF- α [13-15]. Regulation of B₁ expression by B₂ receptor through activation of NF κ B and MAP kinases has also been observed [12,15]. However, to our knowledge this has not been shown in man.

Kinin receptors are expressed in neuronal tissues, are upregulated in response to painful stimuli and their antagonists produce an antinociceptive effect in different pain models [16,17]. Furthermore, B₁ receptor knockout mice are refractory to chemical and thermal nociceptive stimuli [18]. Transient receptor potential vanilloid 1 (TRPV1) is suggested to mediate ionic mechanisms coupling BK receptors to the excitation and sensitization of nociceptors [19].

The interaction between prostaglandins (PG) and BK in the process of inflammatory pain is well established [20,21] as BK induces prostaglandin release in various tissues [20,22,23]. It is suggested that BK-induced sensitization is in part secondary to prostaglandin synthesis, since NSAIDs inhibit BK-mediated sensitization of heat responses, while prostaglandin E₂/I₂ reverse this inhibition [24,25].

The aim of the present study was to investigate the role of kinin receptors in acute inflammatory pain in humans by assessing the gene expression of B₁ and B₂ receptors following oral surgery and its correlation to self-reported pain intensity, as well as to evaluate the levels of their immunoreactive ligands at the site of tissue injury using the microdialysis technique. The interaction between the kinin system and COX-PG pathway was also studied.

Results

1. Effect of tissue injury and ketorolac treatment on BK and des-Arg⁹-BK

Both BK and des-Arg⁹-BK levels in microdialysate were detectable at all time points and decreased gradually over the 3 h collection period with the third hour being significantly lower than the first and second hours in both the placebo and ketorolac treatment groups ($p < 0.05$). However, there was no significant difference between both treatment groups and Fig. 1 (a&b) shows the levels measured at the left side. The right side showed the same results (data not shown).

HPLC analysis performed on microdialysate samples collected at the site of tissue injury from both placebo and ketorolac treatment groups, showed that kinins detected by chemiluminescent enzyme immunoassays were mainly detected at the retention time corresponding to the reference peptides des-Arg⁹-BK (18.6 min), BK (24.1 min), des-Arg¹⁰-KD (24.7 min) and KD (29.8 min).

2. Effect of tissue injury and ketorolac treatment on *BDKRB1* and *BDKRB2* gene expression at the site of tissue injury

Following the identification of four different immunoreactive kinins and their increased levels following tissue injury, it becomes of interest to test for the expression of kinin receptors in the same model. Both *BDKRB1* and *BDKRB2* were significantly upregulated 3 hours following tissue injury in both the placebo and ketorolac treatment group (Table 1). *BDKRB1* was 2.5-fold upregulated in the placebo treated group ($p < 0.0001$; paired t-test; $n = 15$) and 2.4-fold upregulated in the ketorolac treatment group ($p < 0.0001$; paired t-test; $n = 15$). Similarly, *BDKRB2* was 2-fold upregulated in the placebo treated group ($p < 0.0001$; paired t-test; $n = 15$) and 1.9-fold upregulated in the ketorolac treatment group ($p < 0.0001$; paired t-test; $n = 15$). Ketorolac treatment did not have a significant effect on the gene expression of either *BDKRB1* ($p = 0.71$; unpaired t-test) or *BDKRB2* ($p = 0.73$; unpaired t-test).

3. Correlation between pain intensity and the gene expression of BK receptors

The contribution of kinins to pain is well established. Further, the correlation between BK and KD and change in pain intensity was recently reported in man [26]. We assessed here whether the gene expression of kinin receptors was also correlated to pain intensity.

Pain intensity has increased throughout the three hours observation period (time: $p < 0.001$), and as expected ketorolac treatment induced a significant analgesic effect ($p = 0.023$; two-way repeated measure ANOVA) over the three hours observation period (Fig. 2a).

The correlation between the sum of pain intensity scores as measured by VAS and the up-regulation of gene expression of *BDKRB1* and *BDKRB2* following tissue injury was examined using Pearson's correlation coefficients (Fig. 2 b-e). Both *BDKRB1* ($r = 0.68$; $p = 0.005$; $n = 15$) and *BDKRB2* ($r = 0.58$; $p = 0.02$; $n = 15$) were positively correlated to pain intensity in the ketorolac treatment group. On the other hand, *BDKRB1* ($r = 0.22$; $p = 0.425$; $n = 15$) and *BDKRB2* ($r = 0.50$; $p = 0.058$; $n = 15$) did not significantly correlate to pain intensity in the placebo treatment group.

4. Effect of tissue injury and ketorolac treatment on TRPV1 and NF κ B gene expression at the site of tissue injury

To investigate the mechanism of B₁ receptor up-regulation and the contribution of TRPV1 receptor to the algesic mechanisms of both kinin receptors, we further evaluated the gene expression of both TRPV1 and NF κ B in the same biopsies and examined the correlation among the changes in their gene expression levels.

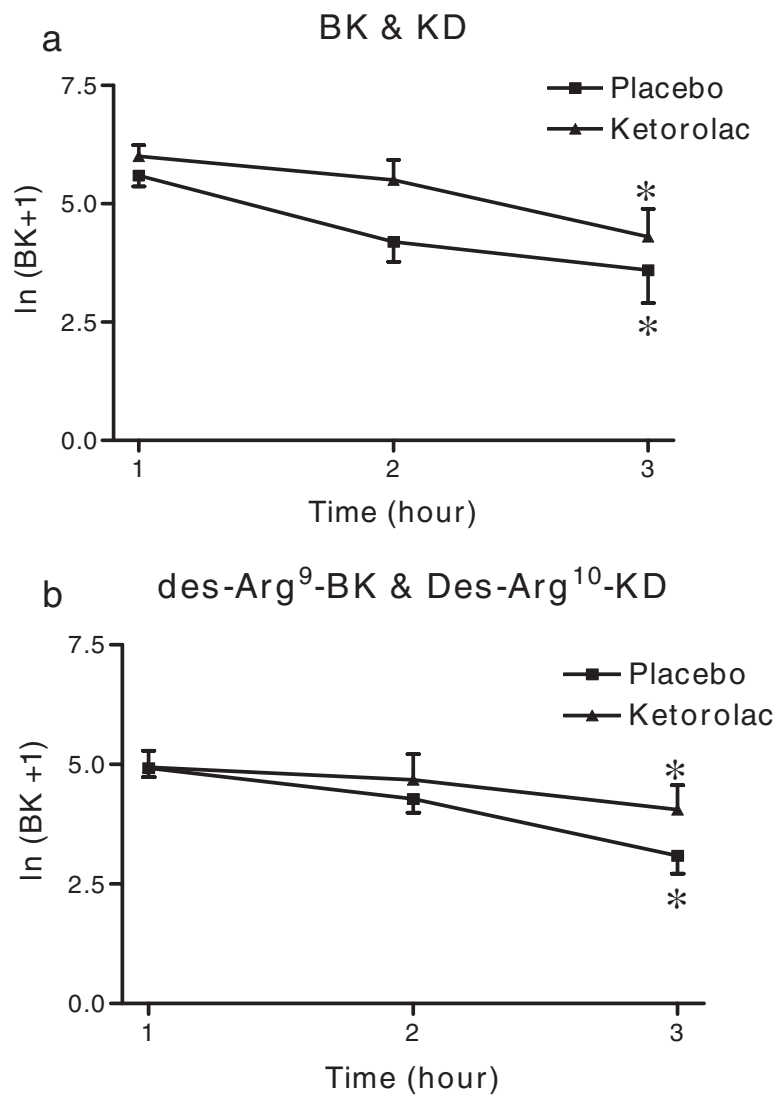


Figure 1 The levels of immunoreactive kinins were significantly lower at the third hour compared to the first 2 hours. There was no significant difference between the placebo and ketorolac treatment groups. Fig. (1a) shows the levels of B₂ receptor ligands (BK and KD) and Fig. (1b) shows the levels of B₁ receptor ligands (des-Arg⁹-BK and des-Arg¹⁰-KD). Concentrations of immunoreactive kinins (pg/ml) were transformed into Ln (X+1), which is shown here. For details, please refer to the methods section. Figures show the levels measured at the left side. The right side showed similar results. Data are presented as mean ± SEM; *indicated p < 0.05, 3 way ANOVA.

NFκB was slightly, though significantly, up-regulated following tissue injury in both the placebo treated group (1.3-fold, p = 0.027; paired t-test; n = 15) and the ketorolac treatment group (1.4-fold, p = 0.042; paired t-test; n = 13; Table 1). There was no significant difference between the two treatment groups (p = 0.56; unpaired t-test).

On the other hand, TRPV1 was significantly down regulated following tissue injury in both the placebo treated group (2.2-fold, p < 0.0001; paired t-test; n = 15) and the ketorolac treatment group (2.6-fold, p < 0.0001; paired t-test; n = 15). However, there was no significant

difference between the two treatment groups (p = 0.33; unpaired t-test).

5. Correlation between the gene expression of *BDKRB1*, *BDKRB2*, *TRPV1* and *NFκB*

Since there was no significant difference between the two treatment groups (placebo and ketorolac) in the gene expression of either of the four genes studied, the two treatment groups were pooled to calculate the correlation coefficient for both receptors [4]. The gene expression of *BDKRB1* and *BDKRB2* were significantly correlated (r = 0.52; p = 0.004; n = 30; Fig. 3a). Further,

Table 1 Gene expression of kinin receptors, TRPV1 and NFκB:

	Placebo			Ketorolac		
	Pre-surgery (Average ΔCt)	Post-surgery (Average ΔCt)	Fold change* (P value)	Pre-surgery (Average ΔCt)	Post-surgery (Average ΔCt)	Fold change* (P value)
B ₁ receptor (<i>BDKRB1</i>)	17.4 ± 3.6	16.2 ± 3.6	2.5 ± 0.9 (p < 0.0001)	18.6 ± 3.4	17.4 ± 3.5	2.4 ± 0.8 (p < 0.0001)
B ₂ receptor (<i>BDKRB2</i>)	14.8 ± 3.0	13.8 ± 2.9	2.0 ± 0.6 (p < 0.0001)	15.6 ± 2.7	14.7 ± 2.7	1.9 ± 0.5 (p < 0.0001)
NFκB	17.3 ± 0.6	17.0 ± 0.4	1.3 ± 1.2 (p = 0.027)	17.3 ± 0.5	17.0 ± 0.7	1.4 ± 0.8 (p = 0.042)
TRPV1	18.8 ± 2.9	19.8 ± 2.7	-2.2 ± 0.9 (p < 0.0001)	19.5 ± 2.7	20.7 ± 2.9	-2.6 ± 1.1 (p < 0.0001)

Data are presented as mean ± S.D.; n = 13-15 per group.

*Fold change is calculated as $2^{-\Delta\Delta C_t}$, Average ΔC_t is the normalized cycle threshold in reference to the endogenous control (18S rRNA). Data were analyzed using paired t test.

the gene expression of TRPV1 was significantly correlated to *BDKRB1* ($r = 0.49$; $p = 0.006$; $n = 30$; Fig. 3b) but not *BDKRB2* ($r = 0.33$; $p = 0.077$; $n = 30$; data not shown). On the other hand, the gene expression of NFκB was not correlated to either *BDKRB1* ($r = -0.106$; $p = 0.61$; $n = 26$) or *BDKRB2* ($r = 0.19$; $p = 0.35$).

Discussion

Kinin levels are very difficult to measure accurately in biological samples, particularly in blood, due to the presence of metabolic enzymes that have the capacity to synthesize or metabolize kinins in vitro. Further, kinins act locally close to their site of formation before they are rapidly hydrolyzed [27]. The microdialysis technique overcomes these difficulties by collecting the kinins at the site of inflammation and excluding the large molecular weight enzymes that may change kinin levels.

In the present study, and in accordance with previous studies [5,26], we show an elevation in the tissue levels of BK and KD following tissue injury. While the chemiluminoenzyme immunoassay does not distinguish between the two peptides, the use of immuno-HPLC approach showed clearly that both peptides were present in the microdialysis samples. Since microdialysis sampling in this study started at the completion of third molar extraction (approximately 30 minutes duration), and the intervals of sampling was relatively long (1 hour) to allow for sufficient volume of each sample, the rise of immunoreactive peptide concentration was not detected, but rather the fall of its concentration 3 hours following surgery was detected. This is expected since the concentration of both kinins are rapidly elevated within 15 min following injury [26].

Further, we show for the first time in an in-vivo human study, a similar pattern for B₁ receptor ligands (des-Arg⁹-BK, des-Arg¹⁰-KD), being elevated following tissue injury before their levels decrease at the latter time points. The immuno-HPLC approach provides

evidence for the presence of des-Arg¹⁰-KD, which is the most potent known human B₁ receptor agonist [28]. Most of the earlier studies of acute inflammatory pain focused on BK and KD due to the impression that the B₁ receptors are only expressed in chronic inflammation [29]. However, we show here that the B₁ receptor is up-regulated as early as 3 hours following tissue injury in humans. Further, pre-surgery biopsies showed expression of both B₁ and B₂ mRNA before any tissue injury. This early expression of B₁ receptor [30] or even its constitutive expression [31] has been shown earlier in some experimental studies. The importance of B₁ receptor lies in the role that it plays in maintaining inflammation in chronic pain states, in contrast to the B₂ receptor that is quickly desensitized. Several mechanisms have been suggested to induce B₁ receptor up-regulation in response to inflammation. First, it may be cross up-regulated by activation of B₂ receptors [12]. The correlation we show here between the gene expression of B₂ and B₁ receptor supports this mechanism in the present clinical model of tissue injury. Second, our earlier study showed marked up-regulation of IL-1β [1] and TNF-α [2] following surgery, and TNF-α was densely expressed on infiltrated inflammatory cells, particularly, on macrophages. Given both TNF-α and IL-1β are known to upregulate B₁ receptor both in-vitro [32] and in-vivo [15], we hypothesize that the up-regulation of both IL-1β and TNF-α may contribute to the up-regulation of B₁ shown in this study. Finally, our recent study demonstrates that an inflammatory cascade mediated by IL-6, IL-8, CCL2, CXCL1 and CXCL2 contributes to the development of acute inflammation and inflammatory pain [4]. This cascade may also contribute to the B₁ receptor up-regulation since neutrophil influx has been suggested to contribute to the up-regulation of B₁ receptors [15].

The activation of the transcription factor NFκB is a critical step in up-regulation of B₁ receptors that are

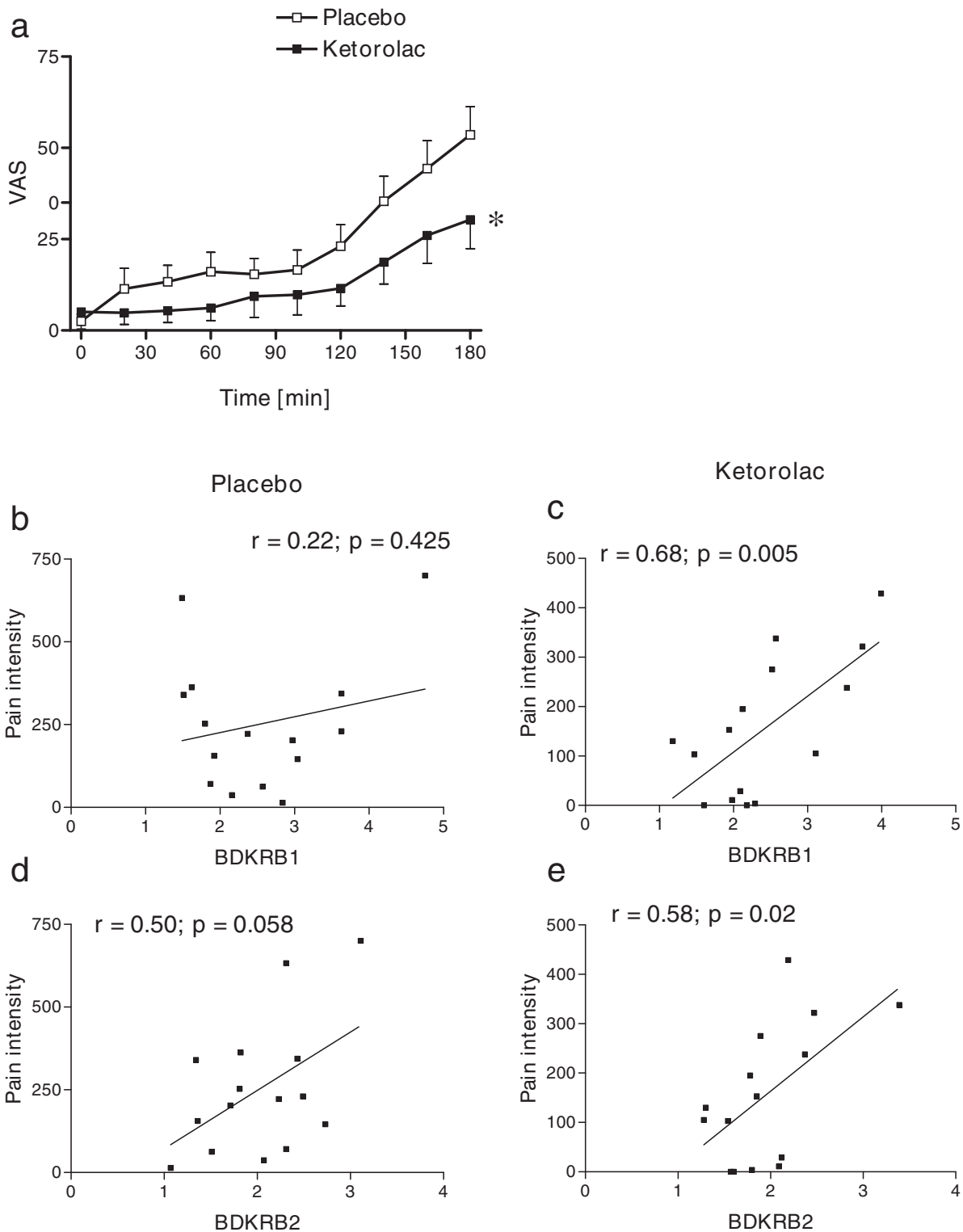
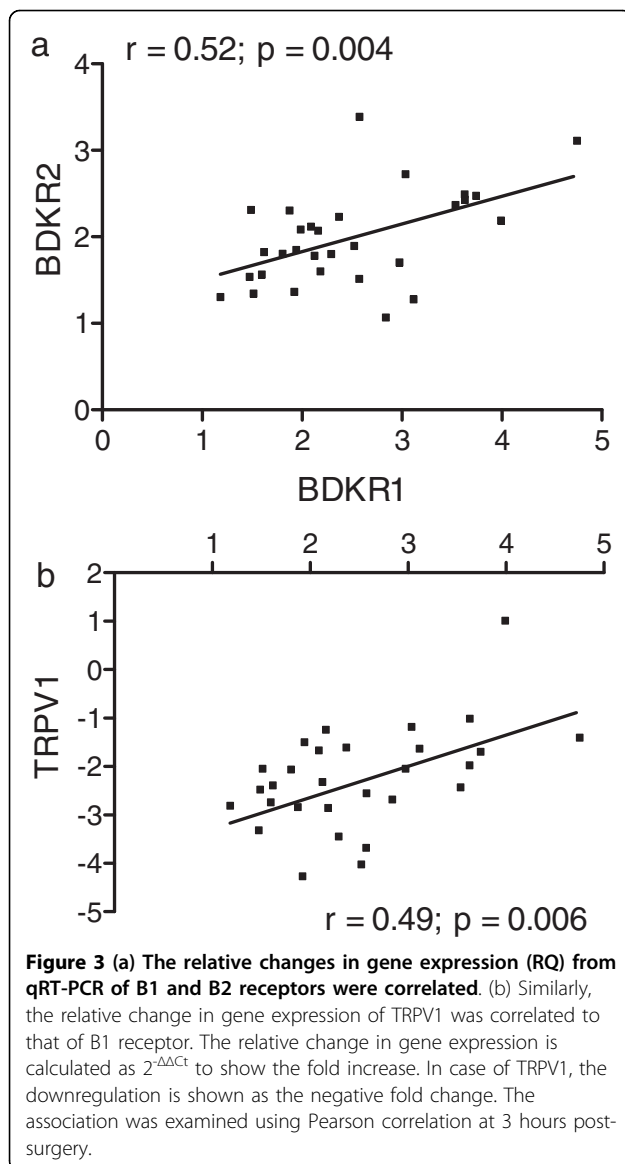
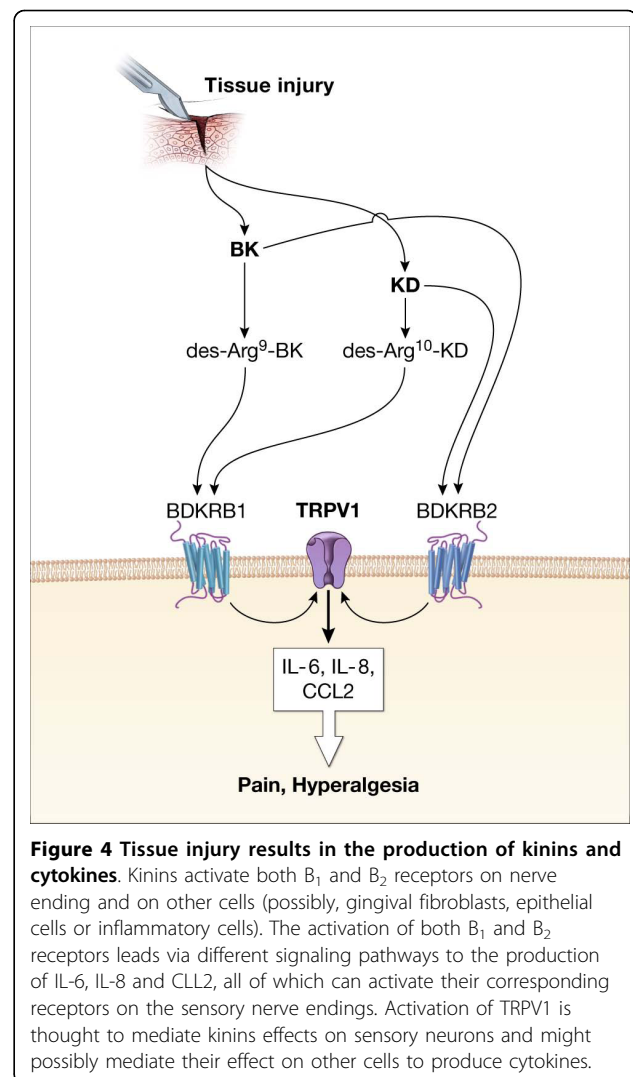


Figure 2 (a) Pain intensity assessed by 100 mm VAS was lower in the ketorolac treatment group ($p = 0.023$, two way ANOVA); Data are presented as mean \pm SEM. (b-e) The change in gene expression level (RQ) of both B1 and B2 receptors was correlated to pain intensity (VAS) in the ketorolac treatment group but not in the placebo treatment group. The association between the gene expression and pain scale was examined using Pearson correlation. The y-axis represents the sum of the pain intensity over the first 3 hours post-surgery. The x-axis represents the relative changes in gene expression (RQ) from qRT-PCR.



induced not only by IL-1 β and TNF- α [15,33], but also by its own agonists [34] or by activation of B₂ receptors [35]. The small change in NF κ B gene expression suggests that this may be mediated primarily at the protein level (i.e. the activation of NF κ B) and the role of NF κ B gene expression is relatively small at the early stage of this clinical model of tissue injury. However, to confirm the role of activation of NF κ B in this model further analysis at the protein level is warranted. Due to the limited size of the biopsy, we were unable to conduct the study at both the protein and the gene expression levels.

The role of kinin receptors and their endogenous ligands in inflammation and pain is well recognized (Fig. 4). Not only does activation of B₂ receptors on primary sensory neurons lead, via second messenger pathways, to the activation of polymodal nociceptors and



hyperalgesia [29], but bradykinin can also sensitize nociceptors following the release of prostaglandins [22,25]. This might explain the correlation between pain intensity and B₂ receptors expression in the ketorolac treatment group but not in the placebo treatment group. The presence of prostaglandins affects pain intensity, therefore, confounding the correlation between B₂ receptor and pain intensity shown in the absence of prostaglandins. Prostanoids released in response to tissue injury [1] would contribute to the inflammatory pain response by activating their receptors on nerve endings as well as by interacting with mediators released to sensitize nociceptors. The similar correlation between B₁ expression and pain intensity points to the participation of B₁ receptor in pain processes at this early time point (3 hours) in contrast to earlier reports [16] confining the contribution of B₁ receptors to nociception to later time points. While the present data shows only the gene expression of kinin receptors due to limited biopsy

size, a correlation between B₁ gene expression level and functional up-regulation of the receptor has been reported earlier in LPS treated rats [15]. Further, the correlation between pain intensity and gene expression level reported here, argues in favor of a functional relationship between tissue injury and increased BK receptor activation leading to an increase in pain.

Kinin receptors are known to be expressed in different tissues besides primary sensory neurons. Immunohistochemical studies showed immunoreactivity for both B₁ and B₂ receptors in epithelial cells, submucosal glands, fibroblast, vascular smooth muscle, vascular endothelial cells, and macrophages of human nasal turbinates [36]. Further, B₁ and B₂ receptor genes are expressed in gingival fibroblasts [37]. Inflammatory cells such as mast cells also, shows expression of B₁ and B₂ receptor [38]. Thus, cellular localization of B₁ and B₂ receptor in the present study could be gingival fibroblasts, epithelial cells in oral mucosa or the inflammatory cellular infiltrate. However, since we did not carry out immunohistochemical studies, we cannot be certain of the cell types of the kinin receptors in oral mucosa, as qRT-PCR cannot distinguish the cellular origin of mRNA expressed.

In gingival fibroblasts, BK up-regulates IL-1 β and TNF α -stimulated IL-6 [39] and IL-8 [40] production. Several other studies showed an induction of IL-6, IL-8 and CCL2 via activation of B₁ or B₂ receptors in a variety of cells [41-47]. Protein kinase C (PKC) signaling pathway seems to be responsible for BK induced IL-8 production in gingival fibroblasts [40], and PI-PLC and NF- κ B signaling pathways are thought to be responsible for BK increased IL-6 production, in synovial fibroblasts [47]. As mentioned earlier, we have recently shown a correlation between the gene expression of IL-6, IL-8 and CCL2 and pain intensity, in the oral surgery model [4]. Therefore, it is possible that kinins besides activating B₁ and B₂ receptors on sensory nerve terminals, also activate B₁ and B₂ receptors on gingival fibroblasts or inflammatory cells inducing the expression of these cytokines, which contributes to the development of inflammatory pain.

TRPV1, a non-selective cation channel expressed in a group of sensory neurons [48], is considered an important target of BK signaling pathways to activate sensory neurons [19]. However, several other tissues express TRPV1 in humans, including keratinocytes [49], mast cells [50], synovial cells [51] and dental pulp fibroblasts [52]. Activation of the TRPV1 results in an increased release of IL-8 and PGE₂ by epidermal keratinocytes [49], IL-6 [53] and IL-8 [54] by human bronchial epithelial cells and IL-6 by dental pulp fibroblast [52]. Engler et al. showed an increased gene and protein expression of IL-6 in cultured synovial fibroblasts from

osteoarthritis (OA) and rheumatoid arthritis (RA) patients in response to the TRPV1 agonist capsaicin, suggesting a possible non-neuronal role for TRPV1 in OA and RA pain [51]. Therefore, it is possible that TRPV1 represent a target of BK signaling pathways both to induce cytokines by non-neuronal cells and to activate sensory neurons. It is suggested that B₂ receptor via PKC signaling pathway activates TRPV1 in neuronal cells [55]. The activation of TRPV1 by B₁ receptors was also suggested earlier [56,57]. This would pose a plausible explanation to the correlation between the change in gene expression of both receptors even with the up-regulation of B₁ and down-regulation of TRPV1.

In the present study, we did not see a correlation between B₂ receptor and TRPV1 gene expression. The absence of significant correlation between B₂ and TRPV1 gene expression could possibly reflect the dissociation between gene expression levels and the functionality of B₂ receptors due to the constitutive nature of B₂ receptors and their rapid desensitization. It could also be due to the contribution of other TRP receptors to the signaling mechanism of B₂ receptors. Recently, using TRPV1-KO mice, TRPV1-independent mechanisms were shown to contribute to BK-evoked pain responses [58], which might include TRPA1 and other ion channels [59]. The down-regulation of TRPV1 in response to tissue injury is however, an unexpected finding, given its pronociceptive role [60]. Yet, the same finding was seen following spinal cord injury in rat [61], where TRPV-1 receptor gene expression levels were decreased at the level of spinal cord injury but increased at rostral spinal cord areas. The up-regulation of TRPV1 is frequently reported at the spinal level following neuropathic pain models [61,62]). It is possible that TRPV1 would play different roles in the periphery or at the spinal level.

In conclusion, we propose here that B₁ receptor is upregulated early following tissue injury in man, possibly in response to kinin activation, and cytokines and chemokines secretion. B₁ receptor activation contributes to the development of acute inflammatory pain possibly via the activation of TRPV1.

Methods

Subjects, timeline of clinical procedures and biopsy collection

Healthy volunteers (n = 59) between 16 to 30 years old who required extraction of impacted third molars were included in the present study (Table 2). The protocol was approved by the Institutional Review Board of the National Institute of Dental and Craniofacial Research, National Institutes of Health (NIH). Written informed consent was obtained from all participants before treatment. Pregnant or lactating females or patients with the presence of clinical signs of infection or inflammation at

Table 2 Demographic data of participants:

Treatment	Microdialysis		Biopsies	
	Placebo	Ketorolac	Placebo	Ketorolac
Number	15	14	15	15
Age	22.3 ± 3.5	19.2 ± 2.9	19 ± 3.6	18.8 ± 2.6
Gender M/F	5/10	4/10	8/7	10/5
Race				
White	5	11	12	11
African	3	1	1	2
Others	7	2	2	2
Difficulty*	7.1 ± 2.1	7.1 ± 1.0	7.5 ± 0.9	7.2 ± 0.9
Rescue Medicine	11 (73%)	7 (50%)	8 (53%)	4 (27%)

* Extraction difficulty is the sum calculated by assigning a score of (2) for soft tissue impactions, (3) for partial bony impactions and (4) for full bony impactions. Data are presented as mean ± S.D.

the extraction sites were not included in the study. All subjects received intravenous midazolam (4.8 ± 0.5 mg) and mandibular nerve block with 2% lidocaine (164.6 ± 26.9 mg) with epinephrine 1:100,000 prior to surgery.

For 29 patients, following satisfactory local anesthesia, two mandibular third molars were extracted and a surgical difficulty score was assigned for each tooth as follows: a score of (2) for soft tissue impactions, (3) for partial bony impactions and (4) for full bony impactions. After extraction, a microdialysis probe (CMA/20 Microdialysis Probe; CMA/Microdialysis, North Chelmsford, MA) was placed bilaterally along the buccal aspect of the mandible, beneath the mucogingival flap elevated for the surgical procedure. The probe fiber consists of a 10-mm flexible, nonmetallic, semipermeable dialysis membrane with a molecular cutoff ranging from 3000 to 20,000 Da. The probes were secured to an adjacent tooth with silk suture and the flap closed in the usual fashion using 3-0 chromic gut suture. Sterile lactated Ringer's solution was pumped at 10 µL/min and samples collected at 60-min intervals after the completion of surgery, before pain onset. Subjects randomly received either placebo or ketorolac (30 mg) intravenously 30 min before biopsy or microdialysis sampling. They remained under observation for the first 3 h after surgery to evaluate pain and adverse events and collect samples by microdialysis. During the immediate post-operative period, subjects were allowed one dose of tramadol (100 mg) as a rescue medication, if requested, after 1 h of evaluation. Patients rated their pain intensity every 20 min for the first 3 postoperative hours using a 100-mm visual analog scale. At the conclusion of the observation period, the microdialysis probes were removed. Microdialysis samples were placed on dry ice after each collection period, and stored at -70°C. For the other 30 patients, a preoperative 3 mm punch biopsy was taken from the oral mucosa overlying the impacted

third molar and a second biopsy was taken from a different surgical site 3 hours post-surgery. All biopsies were immediately frozen in liquid nitrogen and stored at -70°C until ready for RNA extraction.

Quantitative real-time PCR

Oral mucosal biopsies ($n = 60$) were used to detect gene expression using ABI Prism 7900 HT Sequence Detection System (Applied Biosystems, Foster City, CA) as described previously [2,3,63]. All reagents were purchased from Applied Biosystems and 2 µg of DNase-treated RNA was used to synthesize cDNA using random primers from the High-Capacity cDNA Archive Kit according to the manufacturer's instruction. Polymerase chain reaction was performed with cDNA template using the PCR Master Mix with AmpErase UNG. Sequence-specific primers and TaqMan MGB probes were purchased from Assays-on-Demand Gene expression product. Quantification of gene expression was performed in a 20-µL reaction (384-well plate) and each sample was run in triplicate. The housekeeping gene 18S rRNA was used as endogenous control and negative controls were processed under the same conditions without a cDNA template. Data acquisition was conducted using User Bulletin #2 software (v1.6, Applied Biosystems). The threshold cycle (Ct) of 18S rRNA was used to normalize target gene expression (ΔCt) to correct for experimental variations and average ΔCt was used for comparison of the gene expression in post-surgery tissue versus that in pre-surgery tissue. The relative change in gene expression calculated as $2^{-\Delta\Delta Ct}$ was used to compare different treatment groups.

Quantification of BK and des-Arg⁹-BK

Microdialysis samples were diluted in cold absolute ethanol (1:8 v/v), incubated overnight at 4°C then centrifuged. The supernatant was evaporated to dryness and the residue of evaporation was stored at -20°C until the quantification and characterization of immunoreactive kinins. Residues of evaporated ethanol extracts of microdialysates were resuspended in 50 mM Tris-HCl buffer pH 7.4, containing 100 mM NaCl and 0.05% Tween-20. After resuspension, B₂ receptor ligands (BK and KD) and B₁ receptor ligands (des-Arg⁹-BK and des-Arg¹⁰-KD) were quantified by competitive chemiluminescent enzyme immunoassays as previously described [64,65]. These methods have been validated and their analytical performances were reported. As described earlier, these immunoassays shows cross reactivity between BK and KD and between des-Arg⁹-BK and des-Arg¹⁰-KD.

Identification of immunoreactive kinins (BK, KD, des-Arg⁹-BK, des-Arg¹⁰-KD) was carried out using HPLC (Agilent 1100 Series system, Agilent Technologies Canada, Mississauga, ON, Canada) as described earlier

[66]. Briefly, 40 microdialysis samples were pooled, and extracted with cold ethanol as described above. The residue of samples was dissolved in 200 μ l of 5 mM KH_2PO_4 , pH 3.0, 25% acetonitrile with 1% H_3PO_4 . BK, KD, des-Arg⁹-BK, des-Arg¹⁰-KD were separated on a 2-sulfoethyl aspartamide column (PolySULFOETHYL A, The Nest Group Inc., Southborough, MA, USA) using a linear gradient of KCl (0-300 mM) in 5 mM KH_2PO_4 and 25% acetonitrile (v/v) pH 3 for 30 min. Fractions of 1 ml were collected from minutes 15 to 35, evaporated to dryness before quantification of immunoreactive BK and des-Arg⁹-BK. Retention time of each immunoreactive fraction was compared to that of standard peptides: des-Arg⁹-BK (18.6 min), BK (24.1 min), des-Arg¹⁰-KD (24.7 min) and KD (29.8 min).

Statistical Analysis

All statistical analyses were conducted using SPSS (v. 16.0, SPSS, Chicago, IL). Paired t-test was used to compare the gene expression in post-surgery tissue versus that in pre-surgery tissue. The relative change in gene expression was expressed as $2^{-\Delta\Delta\text{Ct}}$ and was used to compare the change in gene expression in different treatment groups by unpaired t-test. Linear regression analysis was used to examine the association between the fold changes in gene expression ($2^{-\Delta\Delta\text{Ct}}$) and the sum of patient-reported pain scores over the first 3 h post-surgery as measured by VAS. In the case of down-regulation, the negative fold change was used. The association among these gene expressed was examined by Pearson's correlation coefficients.

The effect of ketorolac treatment on VAS was assessed using repeated measures two-way ANOVA and the effect of ketorolac treatment on immunoreactive kinins was assessed using three-way ANOVA with two within subject factors (side and time) and one between subjects factor (treatment). A contrast analysis using t-tests with Bonferroni adjustment was used to measure the differences, which were statistically significant. Data did not follow normal distribution, so was transformed to $\ln(x+1)$. Results were considered significant at $p < 0.05$.

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Authors' contributions

MH participated in the laboratory experimental design, conducted qRT-PCR experiments, data analysis and drafted manuscript. XMW participated in the laboratory experimental design, conducted qRT-PCR experiments, data analysis and partially participated in the manuscript writing. AA conducted

the HPLC quantification of bradykinin and des-Arg⁹-BK analysis and data collection. GNC contributed to patient biopsy collection and data collection. JSB, JSR and RAD participated in the patient enrollment, surgical procedures, patient care and biopsy collection. RAD was entirely responsible for the overall study design, overseeing data collection, analysis and interpretation as well as manuscript version. All authors have read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

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References

1. Lee Y-S, Kim H, Brahim JS, Rowan J, Lee G, Dionne RA: Acetaminophen selectively suppresses peripheral prostaglandin E2 release and increases COX-2 gene expression in a clinical model of acute inflammation. *Pain* 2007, **129**:279-286.
2. Wang X, Hamza M, Gordon SM, Wahl SM, Dionne RA: COX inhibitors down-regulate PDE4D expression in a clinical model of inflammatory pain. *Clin Pharmacol Ther* 2008, **84**:39-42.
3. Wang XM, Wu TX, Hamza M, Ramsay ES, Wahl SM, Dionne RA: Rofecoxib modulates multiple gene expression pathways in a clinical model of acute inflammatory pain. *Pain* 2007, **128**:136-147.
4. Wang X-M, Hamza M, Wu T-X, Dionne RA: Upregulation of IL-6, IL-8 and CCL2 gene expression after acute inflammation: Correlation to clinical pain. *Pain* 2009, **142**:275-283.
5. Swift JQ, Garry MG, Roszkowski MT, Hargreaves KM: Effect of flurbiprofen on tissue levels of immunoreactive bradykinin and acute postoperative pain. *J Oral Maxillofac Surg* 1993, **51**:112-117.
6. Menke JG, Borkowski JA, Bierilo KK, MacNeil T, Derrick AW, Schneck KA, Ransom RW, Strader CD, Linemeyer DL, Hess JF: Expression cloning of a human B1 bradykinin receptor. *J Biol Chem* 1994, **269**:21583-21586.
7. Hess JF, Borkowski JA, Young GS, Strader CD, Ransom RW: Cloning and pharmacological characterization of a human bradykinin (BK-2) receptor. *Biochem Biophys Res Commun* 1992, **184**:260-268.
8. Moreau ME, Garbacki N, Molinaro G, Brown NJ, Marceau F, Adam A: The kallikrein-kinin system: current and future pharmacological targets. *J Pharmacol Sci* 2005, **99**:6-38.
9. Ma QP, Heavens R: Basal expression of bradykinin B(1) receptor in the spinal cord in humans and rats. *Neuroreport* 2001, **12**:2311-2314.
10. Blais C, Marceau F, Rouleau J-L, Adam A: The kallikrein-kininogen-kinin system: lessons from the quantification of endogenous kinins. *Peptides* 2000, **21**:1903-1940.
11. Leeb-Lundberg LMF, Marceau F, Muller-Esterl W, Pettibone DJ, Zuraw BL: International union of pharmacology. XLV. Classification of the kinin receptor family: from molecular mechanisms to pathophysiological consequences. *Pharmacol Rev* 2005, **57**:27-77.
12. Phagoo SB, Poole S, Leeb-Lundberg LMF: Autoregulation of bradykinin receptors: agonists in the presence of interleukin-1beta shift the repertoire of receptor subtypes from B2 to B1 in human lung fibroblasts. *Mol Pharmacol* 1999, **56**:325-333.
13. Campos MM, Souza GEP, Calixto JB: Modulation of kinin B1 but not B2 receptors-mediated rat paw edema by IL-1[beta] and TNF[alpha]. *Peptides* 1998, **19**:1269-1276.
14. Fernandes ES, Passos GF, Campos MM, de Souza GEP, Fittipaldi JF, Pesquero JL, Teixeira MM, Calixto JB: Cytokines and neutrophils as important mediators of platelet-activating factor-induced kinin B1 receptor expression. *Br J Pharmacol* 2005, **146**:209-216.
15. Passos GF, Fernandes ES, Campos MM, Araujo JGVC, Pesquero JL, Souza GEP, Avellar MCW, Teixeira MM, Calixto JB: Kinin B1 receptor up-regulation after lipopolysaccharide administration: role of proinflammatory cytokines and neutrophil influx. *J Immunol* 2004, **172**:1839-1847.
16. Levy D, Zochodne DW: Increased mRNA expression of the B1 and B2 bradykinin receptors and antinociceptive effects of their antagonists in an animal model of neuropathic pain. *Pain* 2000, **86**:265-271.
17. Sufka KJ, Roach JT: Stimulus properties and antinociceptive effects of selective bradykinin B1 and B2 receptor antagonists in rats. *Pain* 1996, **66**:99-103.

18. Pesquero JB, Araujo RC, Heppenstall PA, Stucky CL, Silva JA, Walther T, Oliveira SM, Pesquero JL, Paiva ACM, Calixto JoB, et al: **Hypoalgesia and altered inflammatory responses in mice lacking kinin B1 receptors.** *Proc Natl Acad Sci USA* 2000, **97**:8140-8145.
19. Shin J, Cho H, Hwang SW, Jung J, Shin CY, Lee S-Y, Kim SH, Lee MG, Choi YH, Kim J, et al: **Bradykinin-12-lipoxygenase-VR1 signaling pathway for inflammatory hyperalgesia.** *Proc Natl Acad Sci USA* 2002, **99**:10150-10155.
20. Lembeck F, Popper H, Juan H: **Release of prostaglandins by bradykinin as an intrinsic mechanism of its analgesic effect.** *Naunyn Schmiedebergs Arch Pharmacol* 1976, **294**:69-73.
21. Kumazawa T, Mizumura K: **Chemical responses of polymodal receptors of the scrotal contents in dogs.** *J Physiol* 1980, **299**:219-231.
22. Sauer SK, Schäfer D, Kress M, Reeh PW: **Stimulated prostaglandin E2 release from rat skin, in vitro.** *Life Sci* 1998, **62**:2045-2055.
23. Averbach B, Peisler M, Izydorczyk I, Reeh PW: **Inflammatory mediators do not stimulate CGRP release if prostaglandin synthesis is blocked by S (+)-flurbiprofen in isolated rat skin.** *Inflamm Res* 2003, **52**:519-523.
24. Pethö G, Derow A, Reeh PW: **Bradykinin-induced nociceptor sensitization to heat is mediated by cyclooxygenase products in isolated rat skin.** *Eur J Neurosci* 2001, **14**:210-218.
25. Mayer S, Izydorczyk I, Reeh PW, Grubb BD: **Bradykinin-induced nociceptor sensitization to heat depends on COX-1 and COX-2 in isolated rat skin.** *Pain* 2007, **130**:14-24.
26. Gerde B, Hilgenfeldt U, Larsson B, Kristiansen J, Sjøgaard K, Rosendal L: **Bradykinin and kallidin levels in the trapezius muscle in patients with work-related trapezius myalgia, in patients with whiplash associated pain, and in healthy controls - A microdialysis study of women.** *Pain* 2008, **139**:578-587.
27. Marceau F, Regoli D: **Bradykinin receptor ligands: therapeutic perspectives.** *Nat Rev Drug Discov* 2004, **3**:845-852.
28. Bastian S, Loillier B, Paquet JL, Pruneau D: **Stable expression of human kinin B₁ receptor in 293 cells: pharmacological and functional characterization.** *Br J Pharmacol* 1997, **122**:393-399.
29. Couture R, Harrison M, Vianna RM, Cloutier F: **Kinin receptors in pain and inflammation.** *Eur J Pharmacol* 2001, **429**:161-176.
30. Regoli D, Marceau F, Barabe J: **De novo formation of vascular receptors for bradykinin.** *Can J Physiol Pharmacol* 1978, **56**:674-677.
31. Wotherspoon G, Winter J: **Bradykinin B1 receptor is constitutively expressed in the rat sensory nervous system.** *Neurosci Lett* 2000, **294**:175-178.
32. Phagoo SB, Yaqoob M, Herrera-Martinez E, McIntyre P, Jones C, Burgess GM: **Regulation of bradykinin receptor gene expression in human lung fibroblasts.** *Eur J Pharmacol* 2000, **397**:237-246.
33. Larrivee J-F, Bachvarov DR, Houle F, Landry J, Huot J, Marceau F: **Role of the mitogen-activated protein kinases in the expression of the kinin B1 receptors induced by tissue injury.** *J Immunol* 1998, **160**:1419-1426.
34. Schanstra JP, Bataille E, Marin Castano ME, Barascud Y, Hirtz C, Pesquero JB, Pecher C, Gauthier F, Girolami JP, Bascands JL: **The B1-agonist [des-Arg10]-kallidin activates transcription factor NF-kappaB and induces homologous upregulation of the bradykinin B1-receptor in cultured human lung fibroblasts.** *J Clin Invest* 1998, **101**:2080-2091.
35. Pan ZK, Zuraw BL, Lung CC, Prossnitz ER, Browning DD, Ye RD: **Bradykinin stimulates NF-kappaB activation and interleukin 1beta gene expression in cultured human fibroblasts.** *J Clin Invest* 1996, **98**:2042-2049.
36. Shirasaki H, Kanaizumi E, Himi T: **Immunohistochemical localization of the bradykinin B1 and B2 receptors in human nasal mucosa.** *Mediators Inflamm* 2009, **2009**:102406.
37. Brechter AB, Persson E, Lundgren I, Lerner UH: **Kinin B1 and B2 receptor expression in osteoblasts and fibroblasts is enhanced by interleukin-1 and tumour necrosis factor-[alpha]. Effects dependent on activation of NF-[kappa]B and MAP kinases.** *Bone* 2008, **43**:72-83.
38. Dlamini Z, Bhoola KD: **Upregulation of tissue kallikrein, kinin B1 receptor, and kinin B2 receptor in mast and giant cells infiltrating oesophageal squamous cell carcinoma.** *J Clin Pathol* 2005, **58**:915-922.
39. Modéer T, Andurén I, Yucel-Lindberg T: **Bradykinin Synergistically Stimulates Interleukin 6 Production in Human Gingival Fibroblasts Challenged with Interleukin 1 or Tumour Necrosis Factor [alpha].** *Cytokine* 1998, **10**:26-31.
40. Brunius G, Domeij H, Gustavsson A, Yucel-Lindberg T: **Bradykinin upregulates IL-8 production in human gingival fibroblasts stimulated by interleukin-1beta and tumor necrosis factor alpha.** *Regul Pept* 2005, **126**:183-188.
41. Koyama S, Sato E, Numanami H, Kubo K, Nagai S, Izumi T: **Bradykinin Stimulates Lung Fibroblasts to Release Neutrophil and Monocyte Chemotactic Activity.** *Am J Respir Cell Mol Biol* 2000, **22**:75-84.
42. Huang CD, Tliba O, Panettieri RA Jr, Amrani Y: **Bradykinin induces interleukin-6 production in human airway smooth muscle cells: modulation by Th2 cytokines and dexamethasone.** *Am J Respir Cell Mol Biol* 2003, **28**:330-338.
43. Wang PH, Cenedeze MA, Campanholle G, Malheiros DM, Torres HA, Pesquero JB, Pacheco-Silva A, Camara NO: **Deletion of bradykinin B1 receptor reduces renal fibrosis.** *Int Immunopharmacol* 2009, **9**:653-657.
44. Klein J, Gonzalez J, Duchene J, Esposito L, Pradere JP, Neau E, Delage C, Calise D, Ahluwalia A, Carayon P, et al: **Delayed blockade of the kinin B1 receptor reduces renal inflammation and fibrosis in obstructive nephropathy.** *Faseb J* 2009, **23**:134-142.
45. Marney AM, Ma J, Luther JM, Ikizler TA, Brown NJ: **Endogenous bradykinin contributes to increased plasminogen activator inhibitor 1 antigen following hemodialysis.** *J Am Soc Nephrol* 2009, **20**:2246-2252.
46. Dos Santos AC, Roffe E, Arantes RM, Juliano L, Pesquero JL, Pesquero JB, Bader M, Teixeira MM, Carvalho-Tavares J: **Kinin B2 receptor regulates chemokines CCL2 and CCL5 expression and modulates leukocyte recruitment and pathology in experimental autoimmune encephalomyelitis (EAE) in mice.** *J Neuroinflammation* 2008, **5**:49.
47. Lee CH, Shieh DC, Tzeng CY, Chen CP, Wang SP, Chiu YC, Huang CY, Hsu CJ, Fong YC, Tang CH: **Bradykinin-induced IL-6 expression through bradykinin B2 receptor, phospholipase C, protein kinase Cdelta and NF-kappaB pathway in human synovial fibroblasts.** *Mol Immunol* 2008, **45**:3693-3702.
48. Caterina MJ, Schumacher MA, Tominaga M, Rosen TA, Levine JD, Julius D: **The capsaicin receptor: A heat-activated ion channel in the pain pathway.** *Nature* 1997, **389**:816-824.
49. Southall MD, Li T, Gharibova LS, Pei Y, Nicol GD, Travers JB: **Activation of Epidermal Vanilloid Receptor-1 Induces Release of Proinflammatory Mediators in Human Keratinocytes.** *Journal of Pharmacology and Experimental Therapeutics* 2003, **304**:217-222.
50. Stander S, Moormann C, Schumacher M, Buddenkotte J, Artuc M, Shpacovitch V, Brzoska T, Lippert U, Henz BM, Luger TA, et al: **Expression of vanilloid receptor subtype 1 in cutaneous sensory nerve fibers, mast cells, and epithelial cells of appendage structures.** *Exp Dermatol* 2004, **13**:129-139.
51. Engler A, Aeschlimann A, Simmen BR, Michel BA, Gay RE, Gay S, Sprott H: **Expression of transient receptor potential vanilloid 1 (TRPV1) in synovial fibroblasts from patients with osteoarthritis and rheumatoid arthritis.** *Biochem Biophys Res Commun* 2007, **359**:884-888.
52. Miyamoto R, Tokuda M, Sakuta T, Nagaoka S, Torii M: **Expression and Characterization of Vanilloid Receptor Subtype 1 in Human Dental Pulp Cell Cultures.** *Journal of Endodontics* 2005, **31**:652-658.
53. Reilly CA, Taylor JL, Lanza DL, Carr BA, Crouch DJ, Yost GS: **Capsaicinoids Cause Inflammation and Epithelial Cell Death through Activation of Vanilloid Receptors.** *Toxicol Sci* 2003, **73**:170-181.
54. Veronesi B, Carter JD, Devlin RB, Simon SA, Oortgiesen M: **Neuropeptides and capsaicin stimulate the release of inflammatory cytokines in a human bronchial epithelial cell line.** *Neuropeptides* 1999, **33**:447-456.
55. Cesare P, Dekker LV, Sardini A, Parker PJ, McNaughton PA: **Specific involvement of PKC-epsilon in sensitization of the neuronal response to painful heat.** *Neuron* 1999, **23**:617-624.
56. Rajpal S, Gerovac TA, Turner NA, Tilghman JI, Allcock BK, McChesney SL, Miranpuri GS, Park SW, Resnick DK: **Antihyperalgesic effects of vanilloid-1 and bradykinin-1 receptor antagonists following spinal cord injury in rats.** *J Neurosurg Spine* 2007, **6**:420-424.
57. Ueda H: **Molecular mechanisms of neuropathic pain-phenotypic switch and initiation mechanisms.** *Pharmacol Ther* 2006, **109**:57-77.
58. Katanosaka K, Banik RK, Giron R, Higashi T, Tominaga M, Mizumura K: **Contribution of TRPV1 to the bradykinin-evoked nociceptive behavior and excitation of cutaneous sensory neurons.** *Neurosci Res* 2008, **62**:168-175.
59. Bandell M, Story GM, Hwang SW, Viswanath V, Eid SR, Petrus MJ, Earley TJ, Patapoutian A: **Noxious cold ion channel TRPA1 is activated by pungent compounds and bradykinin.** *Neuron* 2004, **41**:849-857.

60. Szallasi A, Cortright DN, Blum CA, Eid SR: **The vanilloid receptor TRPV1: 10 years from channel cloning to antagonist proof-of-concept.** *Nat Rev Drug Discov* 2007, **6**:357-372.
61. DomBourian MG, Turner NA, Gerovac TA, Vemuganti R, Miranpuri GS, Tureyen K, Satriotomo I, Miletic V, Resnick DK: **B1 and TRPV-1 receptor genes and their relationship to hyperalgesia following spinal cord injury.** *Spine* 2006, **31**:2778-2782.
62. Kanai Y, Nakazato E, Fujjuchi A, Hara T, Imai A: **Involvement of an increased spinal TRPV1 sensitization through its up-regulation in mechanical allodynia of CCI rats.** *Neuropharmacology* 2005, **49**:977-984.
63. Wang XM, Wu TX, Lee YS, Dionne RA: **Rofecoxib regulates the expression of genes related to the matrix metalloproteinase pathway in humans: implication for the adverse effects of cyclooxygenase-2 inhibitors.** *Clin Pharmacol Ther* 2006, **79**:303-315.
64. Raymond P, Drapeau G, Raut R, Audet R, Marceau F, Ong H, Adam A: **Quantification of des-Arg9-bradykinin using a chemiluminescence enzyme immunoassay: application to its kinetic profile during plasma activation.** *J Immunol Methods* 1995, **180**:247-257.
65. Décarie A, Drapeau G, Closset J, Couture R, Adam A: **Development of digoxigenin-labeled peptide: Application to chemiluminoenzyme immunoassay of bradykinin in inflamed tissues.** *Peptides* 1994, **15**:511-518.
66. Désormeaux A, Moreau ME, Lepage Y, Chanard J, Adam A: **The effect of electronegativity and angiotensin-converting enzyme inhibition on the kinin-forming capacity of polyacrylonitrile dialysis membranes.** *Biomaterials* 2008, **29**:1139-1146.

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