

HDAC4 gene silencing alleviates epilepsy by inhibition of GABA in a rat model

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Objectives: Despite the availability of effective antiepileptic drugs, epileptic patients still suffer from intractable seizures and adverse events. Better control of both seizures and fewer side effects is needed in order to enhance the patient's quality of life. We performed the present study with an attempt to explore the effect that HDAC4 gene silencing would have on epilepsy simulated by model rats. Furthermore, the study made additional analysis on the relativity of the HDAC4 gene in regard to its relationship with the gamma-aminobutyric acid (GABA) signaling pathway.

Materials and methods: Tremor rats were prepared in order to establish the epilepsy model. The rats would go on to be treated with si-HDAC4 in order to identify roles of the HDAC4 in levels of GABAAR α 1, GABAAR α 4, GAD65, GAT-1, and GAT-3. Finally, both electroencephalogram behavior and cognitive function of the rats following the treatment of si-HDAC4 were observed.

Results: Levels of the GABAAR α 1 and GABAAR α 4 showed an evident increase, while GAD65, GAT-1, and GAT-3 displayed a decline in the epilepsy rats treated with the aforementioned si-HDAC4 when compared with the epilepsy rats. After injection of si-HDAC4, the epilepsy rats presented with a reduction in seizure degree, latency and duration of seizure, amount of scattered epileptic waves, and occurrence of epilepsy, with an improvement in their cognitive function.

Conclusion: The study highlighted the role that HDAC4 gene silencing played in easing the cases of epilepsy found in the model rats. This was shown to have occurred through the upregulation of both GABAAR α 1 and GABAAR α 4 levels, as well as in the downregulation of GAD65, GAT-1, and GAT-3 levels. The evidence provided shows that the HDAC4 gene is likely to present as a new objective in further experimentation in the treatment of epilepsy.

Keywords: epilepsy, HDAC4, gene silencing, GABA receptor, GABA transporter, seizures

Introduction

Epilepsy is categorized as a brain disorder characterized by an enduring predisposition, usually generating seizures by displaying emotional and cognitive dysfunction which affects ~50 million people worldwide.¹⁻³ Epilepsy is known to exhibit self-limiting abnormal discharge as well as the typical short, episodic, repetitive, and even stereotypical malfunctions relative to the central nervous system (CNS).⁴ The incidence of epilepsy in the developing nations is significantly higher than that in the developed countries.⁵ Moreover, the incidence of epilepsy present among males has been shown to be far higher than that among the female populations.⁶ To date, it has been scientifically confirmed that factors involving genetics, infection, trauma, poisoning, brain abnormalities, and interaction between both underlying genetic factors and the environment all have the potential to lead to epilepsy.⁷

Gamma-aminobutyric acid (GABA) is acknowledged as an inhibitory neurotransmitter of the CNS in mammals, widely distributed throughout the nervous

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system.⁸ Mammalian Gamma-aminobutyric acid receptors (GABARs) are composed of pentameric, ligand-gated ion channels assembled from various ($\alpha 1$ – $\alpha 6$, $\beta 1$ – $\beta 3$, $\gamma 1$ – $\gamma 3$, δ , π) polypeptide subunits. The α family is the largest of the aforementioned subunits, with six different subtypes contributing significantly to the functional characterization of the GABARs. GABAR $\alpha 1$ is the most widely expressed of all the subtypes, showing a high level of expression in most regions of the brain.⁹ Several reports have done well to highlight the decrease in the communication of GABA-mediated inhibitory synapses, resulting from an overexcitement of neurons during the process of epilepsy.¹⁰ In the synaptic cleft, only GABA transporters (GATs) are able to clear GABA.¹¹ There are four GATs in the brain of a rat: GAT-1, GAT-2, GAT-3, and BGT-1. GAT-1 and GAT-3 are commonly known subtypes and are both particularly and strongly related to incidences of epilepsy.¹² Histone deacetylase (HDAC), as a common enzyme family, has been shown to play essential roles in the stabilization of gene transcription.¹³ This is done through regulating the acetylation levels of histone as well as nonhistone.¹⁴ Presently, HDACs are known to be presented in 18 different types of mammals. Furthermore, four categories in particular, among which HDAC2, HDAC3, and HDAC4, have been proven to have a negative regulation effect on nerve growth and development, as well as on their memory cognition.^{15–17} HDAC4 has produced large expression levels in the brain tissues, while existing predominately in the cytoplasm of neurons, whose cytoplasm location is directly linked with the electrical activity of both the neurons and the phosphorylation of calcium/calmodulin-dependent protein kinase.¹⁸ HDAC4 has the ability to travel between the cytoplasm and the nucleus under instruction provided by the specific signals from HDAC4, consisting of a C-terminal binding protein region of N-terminal, myocyte enhancer factor 2 (MEF2) binding region, heterochromatin 1 binding region, and HDAC structural domain of C-terminal.¹⁹ HDAC4 gene is capable of protecting the neuron from apoptosis due to its inhibition of the cyclin-dependent kinase.²⁰ Moreover, RNA interference was discovered as a recently developed gene regulating technology, proven to be a useful tool in the further exploration of various gene functions in mammals relating to the degradation of the mRNA of a corresponding sequence.²¹ Therefore, the central aim during this study was made to explore as well as to evaluate the effects of HDAC4 gene silencing on the epilepsy in rats. Additionally, the relativity of HDAC4 gene silencing in relation to the GABA signaling pathway may act as a new approach in the treatment of epilepsy.

Materials and methods

Ethical statement

During the experiment, feeding, the intake of both material and treatment were conducted in accordance with the relative animal ethical standards of the Guide for the Care and Use of Laboratory Animals published by the Ministry of Science and Technology of the People's Republic of China in 2006. The experimental process was approved overall by The Experimental Animal Ethics Committee of Lanzhou University Second Hospital.

Subjects

Thirty healthy SD rats provided by the Experiment Animal Center of Lanzhou University Second Hospital were used during the study. Furthermore, a total of 60 tremor rats (TRM rats) found in the genetic epilepsy rat model were introduced by Kyoto University (Japan). TRM rats as well as healthy rats were both male, aged 8–12 months, and weighed between 210 and 250 g. The rats were fed in the second grade animal house at a temperature of 22°C with a contrast of 12-hour light and 12-hour darkness in alternation along with free access to food and water.

WSLP/siRNA injection synthesizing

siRNA was synthesized according to the mRNA sequence of the HDAC4 gene. The siRNA sequences were 3'-GAGACC CAAUGCCAAUGC-5' and 5'-AGGCAGCGCCAGU ACUCA-5'. Water-soluble lipid polymer (WSLP) was synthesized. Three grams of polyethylenimine (PEI) (mw 1,800, Polysciences Company) were added into the mixture containing 10 mL anhydrous dichloromethane and 100 μ L triethylamine (Aldrich Chemical), then the mixture was placed on the ice and mixed for 30 minutes. One gram of cholesterol was dissolved in 5 mL frozenly preserved anhydrous dichloromethane, while being subsequently added to the PEI solution slowly in order to dissolve the cholesterol for 30 minutes. The mixture was then stirred over the ice for a period of 12 hours, followed by drying of the product with a rotary evaporator.

The powder was dissolved in a 50 mL 1 mol/L hydrochloric acid followed by filtering with a glass fiber filter. Unreacted cholesteryl chloroformate was dissolved and removed by dichloromethane, ether, and acetone as follows: unreacted cholesteryl chloroformate was removed with a three-time extraction of aqueous solution by 100 mL dichloromethane; the extracted aqueous solution was filtered in a glass fiber filter; following the evaporation of the solvent and concentration of the product, there was a great amount of acetone and

concentrated product in the sediment, which was subsequently pumped under the vacuum. The powder was then dissolved in water, with the powder precipitated by acetone three times soon after. Following the washing process with methanol, the product was further washed with ether an additional three times. Matrix-assisted laser desorption-time-of-flight was used in order to analyze the white powder product. Voyager DE STR mass spectrometer (PE Applied Biosystems) was used with anti-4-hydroxy-3-methoxy-cinnamic acid as the auxiliary matrix, showing the cholesterol and mw 1,800 PEI using nuclear magnetic resonance at 400 MHz 1 hour with a molar ratio of 1:1. The final product of WSLP was stored at -20°C for further experimentation. Soon after, the WSLP/siRNA complex was prepared. Prior to using, both WSLP and siRNA were added into the 5% glucose solution by the mass ratio of 7:1 and incubated at room temperature for 30 minutes.

Animal grouping

The WSLP/siRNA complex was prepared with a dose of 3.5 μg of siRNA per rat along with the mass ratio of 7:1. For the si-HDAC4 group, 30 TRM rats were randomly chosen and given an intrathecal injection of WSLP/siRNA. In regard to the epilepsy control, 30 TRM rats were chosen at random and given intrathecal injections of normal saline (NS) to prepare a control group. For the normal control, 30 healthy rats were selected randomly and given intrathecal injections with NS to prepare a control group. Rats in each group were fed in the animal room and given injections twice a week (50 μL for every injection). Ten rats were observed at the 1st, 14th, and 30th day following administration regulations. The rats were observed in order to analyze the incidence of 2-hour administration. The rat seizure latency, seizure duration, and seizure grade were all subsequently recorded.

Reverse transcription quantitative PCR (RT-qPCR)

After setting the administration time, rats in each group were strongly anesthetized, receiving an intraperitoneal injection of 10% chloral hydrate. Then, the rats were swiftly beheaded with their brain carefully removed in order to expose the hippocampus, which were frozen and preserved with a Trizol solution for further use. The RT-qPCR was used to detect the mRNA level of GABAAR α 1, GABAAR α 4, GAD65, GAT-1, and GAT-3 (GATs) in the hippocampus tissues of rats in each group. TRIzol was used in order to extract the total RNA in the hippocampus tissues of each group. The OD260/280 value of each extracted RNA sample was detected using an ultraviolet spectrophotometer. The RNA concentration was then subsequently calculated, and the sample was preserved at -80°C . In the reverse transcription PCR system: 20 μL of RNA, 10 μL of diethyl pyrocarbonate-treated water, 10 μL of 5 \times buffer, 1 μL of RNasin, 3 μL of dNTP, 4 μL of Oligo (Dt) 15, and 2 μL of M-MLV with a total volume of 50 μL . Primer5.0 primer design software was used to design the primer sequences, which were synthesized by Shanghai Sangon Biotech (Shanghai, China). The primer sequences used are shown in Table 1. RT-qPCR reaction conditions were as follows: predenaturation at 95°C for 30 seconds, a cycle of 40 times of denaturation at 95°C for 5 seconds, and annealing/extension for 30 seconds with β -actin used as the internal reference. Ct value of the target gene was compared with the Ct value of β -actin. Finally, the relative mRNA expression of each target gene was calculated with the following formula: $\Delta\text{Ct}(\text{target}) = \text{Ct}(\text{target}) - \text{Ct}(\beta\text{-actin})$ and $\Delta\Delta\text{Ct} = \Delta\text{Ct}(\text{experimental group}) - \Delta\text{Ct}(\text{control group})$; $2^{-\Delta\Delta\text{Ct}}$ was used to express the ratio of the relative expression of the target gene to the internal reference gene.

Table 1 The primer sequences for reverse transcription quantitative PCR

Gene	Primer	Sequence	Tm ($^{\circ}\text{C}$)
GABAAR α 1	Upstream	AGCCGAATGCCCCATGCACT	60
	Downstream	CAACCACTGAGCGGGCTGGC	
GABAAR α 4	Upstream	5'-CCTCACGATGACCACCCTAA-3'	60
	Downstream	5'-GCGATGCGGCAGACGAAA-3'	
GAD65	Upstream	TCGAGCCGAAGCAGCTTGCCC	50
	Downstream	GCCCGTGAACTTTTGGGCCAC	
GAT-1 mRNA	Upstream	5'-TGCAAACACGTACGCACATAGAA-3'	65
	Downstream	5'-AGATGCCTCAGCCACACCAC-3'	
GAT-3 mRNA	Upstream	5'-CGGTCACTGGAACAACAAGGTG-3'	65
	Downstream	5'-AACACCACGTAAGGAATCAGGAATG-3'	
β -actin	Upstream	GTGGGGCGCCCCAGGCACCA	50
	Downstream	CTTCCTTAATGTCACGCACGATTC	

Abbreviation: Tm, temperature.

SABC immunohistochemical staining

After setting up the administration time, the rats were anesthetized and received intraperitoneal injections of 10% chloral hydrate. The chest and abdominal cavity were then cut open in order to expose the viscera. The right auricle was subsequently cut with the needle inserted from the apex all the way to the right with the use of a 45-degree angle at the beginning of the aorta to perfuse 250 mL NS rapidly. Rat perfusion was then rinsed with 250 mL 4% polyformaldehyde until the rat limbs were both rigid and its liver turned white in color, following which the puncture needle was set to be removed. The rat was then beheaded, with the brain removed to strip the hippocampal tissue. The tissue was fixed along with 4% polyformaldehyde and kept overnight at a temperature of 4°C with 30% sucrose solution until the tissue was completely deposited. Continuous coronal sections were obtained using constant cold and frozen sections in the machine with a thickness of ~20 µm and placed in the refrigerator at 4°C. The SABC method (SABC immunohistochemistry kit was purchased from BOSTER Biological Technology Co., Ltd., Wuhan, Hubei, China) was used for an immunohistochemical staining for the detection of GABAAR α 1, GABAAR α 4, GAD65, GAT-1, and GAT-3. Then, the 40-well plate was added with 0.01 mol/L PBS (~2/3 of PBS per well), supplemented with a piece of brain slice each well, and rinsed three times (5 minutes/time). The brain slice was treated with 3% H₂O₂ under conditions void of light for a period of 30 minutes. Subsequently, the brain slice was rinsed three times with 0.01 mol/L PBS (5 minutes/time) and incubated in thermostat for 30 minutes after addition of 5% serum. Then, the brain slice was added with diluted first antibody (1:400), and further incubated in a 37°C thermostat for 2 hours. PBS (0.01 mol/L) was replaced with the first antibody, acting as the negative control. The brain slices were washed three times with 0.01 mol/L PBS (5 minutes/time), added with diluted second antibody (1:400), and incubated in the 37°C thermostat for a total of 90 minutes followed by three times 0.01 mol/L PBS rinses (5 minutes/time). After the third antibody had successfully been prepared (duration of 30 minutes), the mixture was again placed in the 37°C thermostat for 90 minutes and rinsed three times with 0.01 mol/L PBS (5 minutes/time). Then, 0.05% DAB was added to brain slice for a period of 5-minute colorization, followed by the slice being rinsed three times with 0.01 mol/L PBS (5 minutes/time). The slice was bleached and dried using natural air, after which 70%, 80%, 90%, and 95% alcohol solutions were used to complete the dehydration

purposes for a period of 5 minutes. Approximately, 100% alcohol I and II were used for 10 minutes, respectively (twice each), and xylene I and II for a period of 30 minutes, respectively, to clear the slice. The slice was mounted by neutral balata, dried, and observed under the microscope.

Western blot analysis

Western blot analysis was applied to detect protein levels of GABAAR α 1, GABAAR α 4, GAD65, GAT-1, and GAT-3 in the rat hippocampal tissues. The rat in each group was beheaded, and the hippocampus was separated and ground. Next, the tissues were added with RIPA cell lysis buffer at a ratio of 150 µL/20 mg and centrifuged (4°C) at 25,764 g for 5 minutes with the supernatant collected. Then, the obtained supernatant was transferred into the new Eppendorf tube, and the protein concentration was detected based on the instructions provided by the bicinchoninic acid kit (Wuhan Boster Biological Technology Ltd., Wuhan, Hubei, China). The extracted protein was added with the loading buffer, with 25 µg of protein loaded in each lane to process with SDS-PAGE for the purposes of protein separation at an electrophoretic voltage of 80–120 v using the wet transfer method. The transmembrane voltage was set at 100 mv for 45–70 minutes, and the protein was transferred on to the polyvinylidene fluoride membrane. Then, the membrane was blocked with 5% BSA at room temperature for 1 hour. Subsequently, the membrane was incubated with addition of diluted rabbit antimouse HDAC4, GABAAR α 1, GABAAR α 4, GAD65, GAT-1, and GAT-3 antibodies (1:200, Santa Cruz Biotechnology Inc., Santa Cruz, CA, USA) overnight at 4°C. The next day, the membrane was rinsed three times with Tris-buffered saline with Tween 20 for 5 minutes, and further incubated with addition of the second antibody (1:5,000) for 1 hour. Finally, the membrane was washed three times (5 minutes/time), developed with DAB, and photographed with the Bio-Rad gel imaging system. β -actin was used as the internal reference. The images were developed using the GEL DOC EZ IMAGER (Bio-Rad Laboratories Inc., Hercules, CA, USA). The gray value of the target band was analyzed using the ImageJ software.

Epilepsy detection and behavior observation

After each injection, the epilepsy behaviors of the rats in each group were continuously monitored for 2 hours, recording the frequency, average time, and grade of each seizure. The seizure grade was decided according to the

Racine standard 17. The seizure was classified into six grades according to the degree of convulsion: grade 0, no seizure; I, rhythmic mouth or facial tic; II, nodding or tail shaking; III, single limb tic; IV, limbs tic or ankylosis; and V, generalized tonic-clonic seizure.

After the preset administration time, two rats in each group were randomly chosen to participate in an electroencephalogram (EEG). The electrode was installed using the methods as follows. The rats were anesthetized by intraperitoneal injections of 350 mg/kg of 10% chloral hydrate, and then the electrode was installed and fixed in the rat's hippocampus area and frontal cortex. The rats were fastened on the brain stereotaxic apparatus, and the collection electrodes were installed in the hippocampus and cortex acquisition area. The left ear was used as the reference electrode with a unipolar lead. A physiological signal acquisition system was used, and a 5-minute constant EEG of rats in each group was recorded using the EEG-4418K to observe the changes in EEG of the rats in each group.

Morris water maze test

After the preset administration time, six rats from each group were randomly chosen to participate in the Morris water maze test. The water maze was 150 cm in diameter, 50 cm in wall height, 40 cm in depth, and (21–23)°C in temperature. A 10 cm diameter platform 1 cm beneath the water was placed in the center of the target quadrant. Following the adaptation training, the rats were prepared to perform the positioning navigation experiment as follows. The rats were gone into the pool facing toward the wall side with the time needed to find the platform being recorded. If the rats found the platform within 2 minutes, the escape latency was recorded; if the rats did not find the platform within 2 minutes, they were led onto the platform and stayed there for an additional 20 seconds, the escape latency was recorded at 2 minutes. Then, the space search experiment was carried out as follows: after the positioning navigation experiment, the platform was removed and the rat residence time in the target quadrant was recorded.

Statistical analysis

The statistical analyses were conducted using a SPSS18.0 (IBM Corporation, Armonk, NY, USA). The measurement data were expressed using the mean \pm SD. Measurement data of normal distribution differences between groups were compared using least significant difference. Comparisons among multiple groups were analyzed using one-way ANOVA. Data of

different time points were analyzed by repeated measurement of the ANOVA. The enumeration data were expressed as both percentages and ratios. Chi-squared tests were used. $P < 0.05$ was indicative of a significant statistical difference.

Results

Increased mRNA level of GABAAR α 1 and GABAAR α 4 and reduced mRNA level of GAD65, GAT-1, and GAT-3 observed in hippocampus tissues of epilepsy rats treated with si-HDAC4

RT-qPCR and immunohistochemistry were used to detect the mRNA levels of GABAAR α 1, GABAAR α 4, GAD65, GAT-1, and GAT-3 specifically in the rat's hippocampus tissues. As the results depicted in Figure 1, the mRNA levels of GABAAR α 1 and GABAAR α 4 in the si-HDAC4 group increased, while those of GAD65, GAT-1, and GAT-3 decreased on the 1st day, 14th day, and 30th day of observation (all $P < 0.05$). The mRNA levels of GABAAR α 1 and GABAAR α 4 in the epilepsy group decreased, while those of GAD65, GAT-1, and GAT-3 all increased on the 1st day, 14th day, and 30th day of observation (all $P < 0.05$). No significant changes regarding the mRNA levels of GABAAR α 1 and GABAAR α 4 on the 1st day, 14th day, and 30th day were found in the control group (all $P > 0.05$). At the same time, in comparison with the control group, the mRNA levels of GABAAR α 1 and GABAAR α 4 in the si-HDAC4 and epilepsy groups significantly decreased, while those of GAD65, GAT-1, and GAT-3 increased (all $P < 0.05$). At the same time, in comparison with the epilepsy group, the mRNA levels of GABAAR α 1 and GABAAR α 4 in the si-HDAC4 group exhibited significant increases, while those of GAD65, GAT-1, and GAT-3 all displayed a significant decline (all $P < 0.05$). Altogether, si-HDAC4 enhanced the levels of GABAAR α 1 and GABAAR α 4, while reducing the levels of GAD65, GAT-1, and GAT-3 in epilepsy rats.

Western blot analysis was used with the intent of detecting the protein level of HDAC4 in the hippocampus tissues of rats in each group, and the results are displayed in Figure 1F and G. In comparison with the control group, the level of HDAC4 in the hippocampus tissues of the si-HDAC4 group significantly decreased, while the HDAC4 level in the hippocampus tissues of the epilepsy group markedly increased ($P < 0.05$). In comparison with the epilepsy group, the level of HDAC4 in the hippocampus tissues of the si-HDAC4 group significantly decreased ($P < 0.05$).

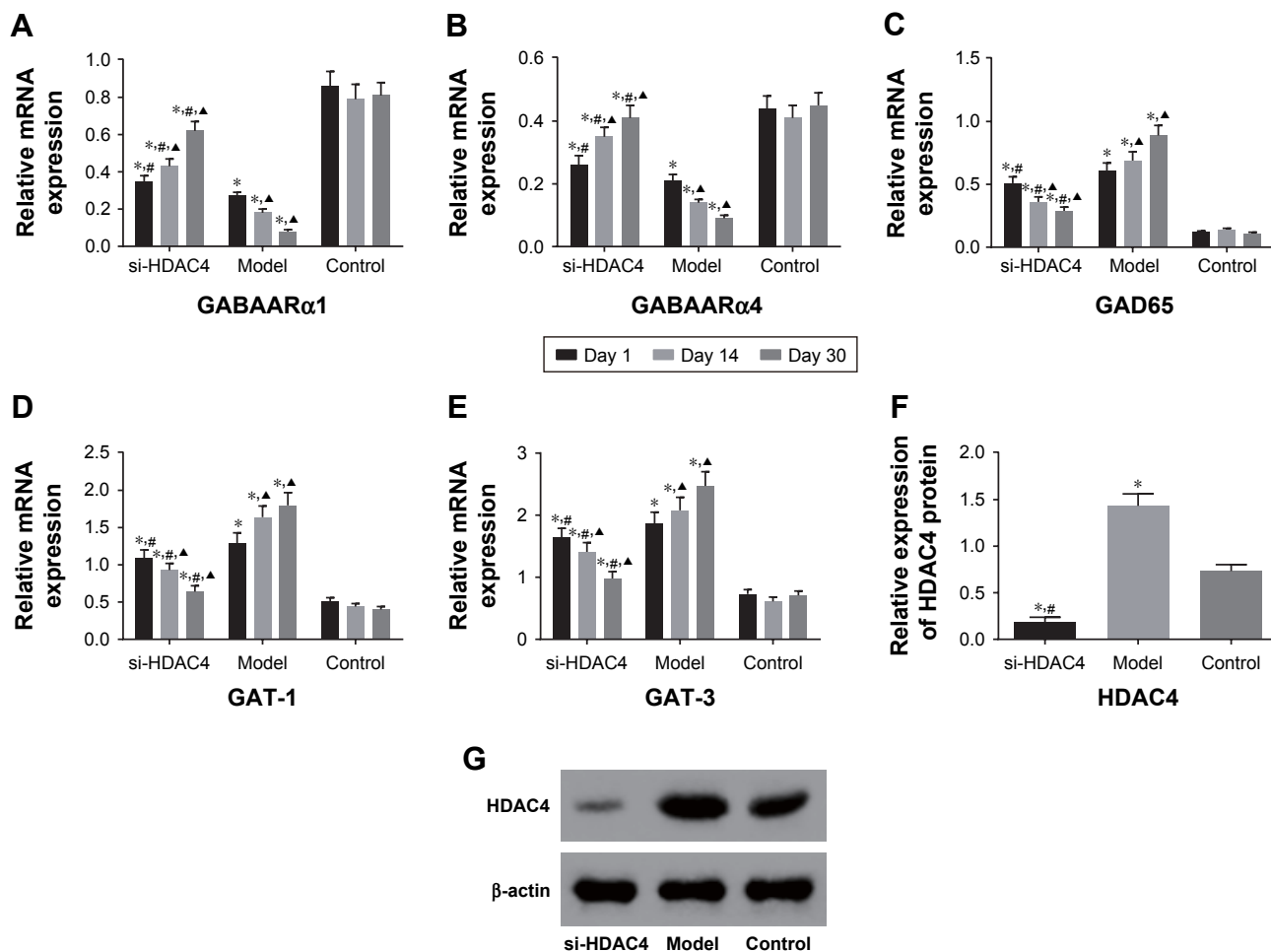


Figure 1 The relative mRNA levels of GABAAR α 1 and GABAAR α 4 increased and levels of GAD65, GAT-1, and GAT-3 decreased in epilepsy rats treated with si-HDAC4. **Notes:** (A) The mRNA level of GABAAR α 1 detected by RT-qPCR; (B) the mRNA level of GABAAR α 4 detected by RT-qPCR; (C) the mRNA level of GAD65 detected by RT-qPCR; (D) the mRNA level of GAT-1 detected by RT-qPCR; (E) the mRNA level of GAT-3 detected by RT-qPCR; (F and G) the silencing efficacy of si-HDAC4 detected by Western blot analysis. * P <0.05 vs the control group at the same time point; # P <0.05 vs the epilepsy group at the same time point; ΔP <0.05 vs the first day of administration in the same group.

Abbreviation: RT-qPCR, reverse transcription quantitative PCR.

Number of positive cells of GABAAR α 1 and GABAAR α 4 increase, while positive cells of GAD65, GAT-1, and GAT-3 decrease in the hippocampus tissues of epilepsy rats after treatment of si-HDAC4

The number of positive cells of GABAAR α 1, GABAAR α 4, GAD65, GAT-1, and GAT-3 in the rat's hippocampus tissues at different time point was counted under the microscope (Figure 2). The positive cells of GABAAR α 1 and GABAAR α 4 in the si-HDAC4 group definitely increased, while those of GAD65, GAT-1, and GAT-3 decreased on the 1st, the 14th, and the 30th day (all P <0.05). The positive cells of GABAAR α 1 and GABAAR α 4 in the epilepsy group showed a noticeable decrease, while the positive cells of the GAD65, GAT-1, and GAT-3 presented with a distinct

increase on the 1st, the 14th, and the 30th day (all P <0.05). The positive cells of GABAAR α 1 and GABAAR α 4 in the control group showed no significant difference on the 1st, the 14th, and the 30th day (all P >0.05). At the same time, in comparison with the control group, the positive cells of GABAAR α 1 and GABAAR α 4 in the epilepsy group displayed a significant decline, while those of GAD65, GAT-1, and GAT-3 showed an evident increase (all P <0.05). The number of positive cells of GABAAR α 1 and GABAAR α 4 in the si-HDAC4 group significantly decreased, while those of GAD65, GAT-1, and GAT-3 all increased on the 1st day and the 14th day compared with the control group (all P <0.05). The positive cells of GABAAR α 1, GAD65, GAT-1, and GAT-3 in the si-HDAC4 group showed no significant difference on the 30th day compared with the control group (all P >0.05). At the same time, in comparison with the epilepsy group, the positive cells of GABAAR α 1 in the si-HDAC4

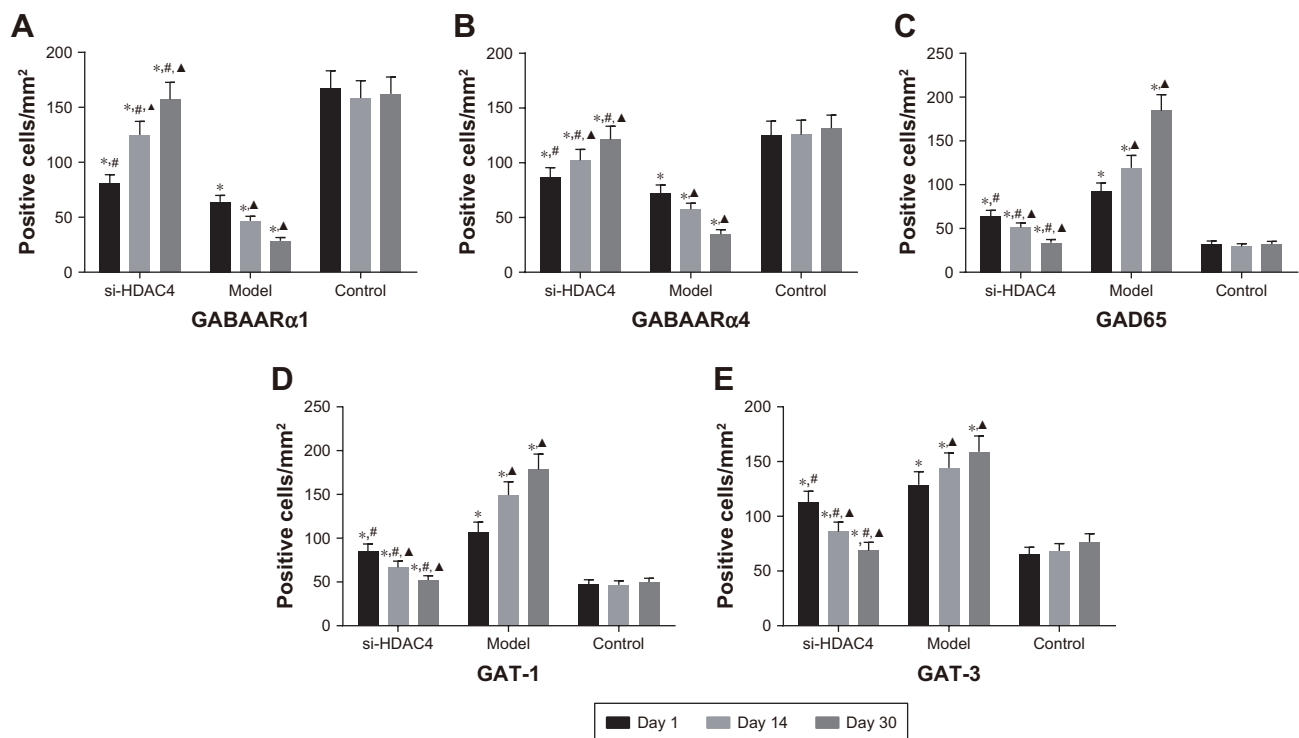


Figure 2 si-HDAC4 elevates the number of positive cells of GABAAR α 1 (A) and GABAAR α 4 (B), while reduces that of GAD65 (C), GAT-1 (D), and GAT-3 (E) of the hippocampus tissues of epilepsy rats after treatment with si-HDAC4.

Notes: * $P < 0.05$ vs the control group at the same time point; # $P < 0.05$ vs the epilepsy group at the same time point; $\Delta P < 0.05$ vs the first day of administration in the same group.

group significantly increased, while those of GAD65, GAT-1, and GAT-3 notably decreased (all $P < 0.05$). Based on the aforementioned results, the HDAC4 gene silencing was shown to increase the number of positive cells for both GABAAR α 1 and GABAAR α 4, while reducing the number of positive cells for GAD65, GAT-1, and GAT-3 in rats with epilepsy.

Protein levels of GABAAR α 1 and GABAAR α 4 are elevated, while protein levels of GAD65, GAT-1, and GAT-3 are decreased in the hippocampus tissues of epilepsy rats after treatment with si-HDAC4

Subsequently, the relative protein levels of GABAAR α 1, GABAAR α 4, GAD65, GAT-1, and GAT-3 in the rat's hippocampus tissues at different time points were all collectively detected by Western blot analysis, with the results presented in Figure 3. The relative protein levels of both GABAAR α 1 and GABAAR α 4 in the si-HDAC4 group increased, while the protein levels associated with GAD65, GAT-1, and GAT-3 all decreased on the 1st day, 14th day, and 30th day (all $P < 0.05$). The relative protein levels for GABAAR α 1 and GABAAR α 4 in the epilepsy group had decreased, while the

same of the GAD65, GAT-1, and GAT-3 increased on the 1st day, 14th day, and 30th day (all $P < 0.05$). The relative protein levels of hippocampus tissues in the control group showed no significant differences on the 1st day, the 14th day, and the 30th day (all $P > 0.05$). At the same time, in comparison with the control group, the relative protein levels of both GABAAR α 1 and GABAAR α 4 in the epilepsy group showed significant decline, while the same subject of the GAD65, GAT-1, and GAT-3 all presented with an increase (all $P < 0.05$). Relative protein levels of GABAAR α 1 and GABAAR α 4 in the si-HDAC4 group both significantly decreased, while those of GAD65, GAT-1, and GAT-3 increased on the 1st day and 14th day (all $P < 0.05$). The relative protein levels of GABAAR α 1 and GAD65, as well as the GAT-1 and GAT-3 in the si-HDAC4 group and the control group showed no significant difference on the 30th day of observation (all $P > 0.05$). At the same time, in comparison with the epilepsy group, the relative protein levels of GABAAR α 1 and GABAAR α 4 in the si-HDAC4 group significantly increased, while those of the GAD65, GAT-1, and GAT-3 all decreased (all $P < 0.05$). According to the results, we came to the conclusion that HDAC4 silencing could enhance the protein levels of both GABAAR α 1 and GABAAR α 4, while decreasing the protein level of GAD65, GAT-1, and GAT-3 in rats with epilepsy.

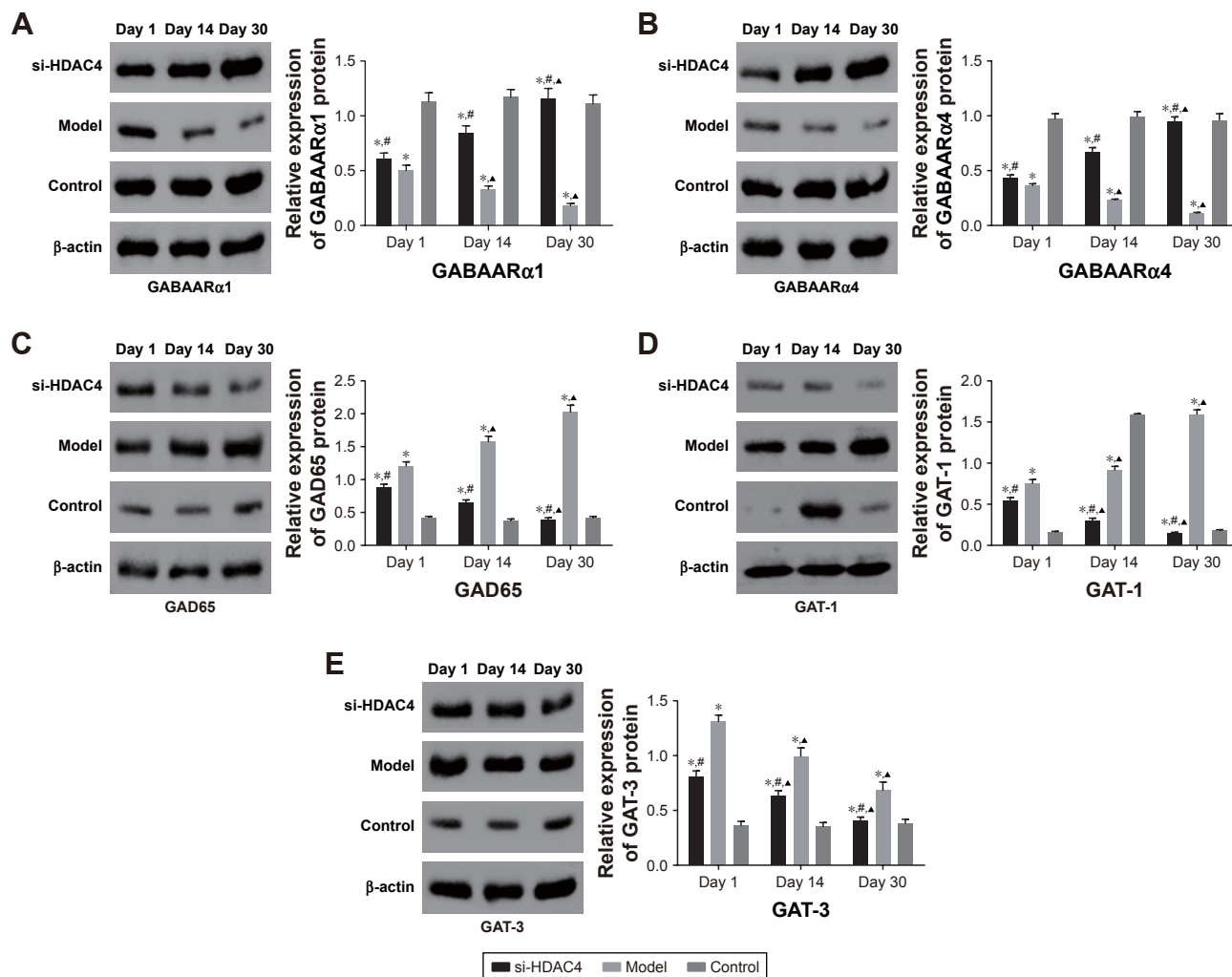


Figure 3 The protein levels of GABAAR α 1 (A) and GABAAR α 4 (B) enhanced, while that of GAD65 (C), GAT-1 (D), and GAT-3 (E) reduced in the hippocampus tissues of epilepsy rats after treatment with si-HDAC4.

Notes: * $P < 0.05$ vs the control group at the same time point; # $P < 0.05$ vs the epilepsy group at the same time point; $\Delta P < 0.05$ vs the first day of administration in the same group.

Silence of HDAC4 inhibits epilepsy

Finally, the EEG of the rats and behavior observation were conducted to identify roles of HDAC4 in the occurrence of epilepsy. The EEG of the rats in each group is shown in Figure 4. In the control group, the EEG of rats mainly presented with α wave or β wave, and the basic wave slightly scattered within θ wave with no obvious rhythm. In the si-HDAC4 group and the epilepsy group, the EEG of rats displayed a large amount of scattered epileptic waves, mainly high spike waves, spike waves, and slow spike colligation waves together with a large number of epileptic discharges. In comparison with the epilepsy group, the rats in the si-HDAC4 displayed a reduced number of scattered epileptic waves.

The behavior observation results of the epileptic rats are presented in Table 2. In the si-HDAC4 group and the epilepsy group, rats exhibited skeletal muscle tremors,

nodding, and frequent chewing; in the control group, rats showed no epilepsy. Following this, the rats were flipped to the left, displaying epileptic limb symptoms as well as a tail rigidity for ~2–5 hours. During the seizure phase, the seizure symptoms disappeared. Nonetheless, the rats appeared to be easily irritated, while displaying strong notions of aggressive behavior. At the same time, compared with the epilepsy group, on the 30th day of administration, the latency period and seizure duration of rats in the si-HDAC4 group showed a reduction in the rate, grade, and frequency of seizure ($P < 0.05$). In the same group, but at different time points, in comparison with the 1st day of administration, the rats showed no significant differences on the 14th day; on the 30th day, the latency period and seizure duration of rats had shortened. Additionally, the grade and frequency of seizure also shortened ($P < 0.05$). Collectively, these aforementioned

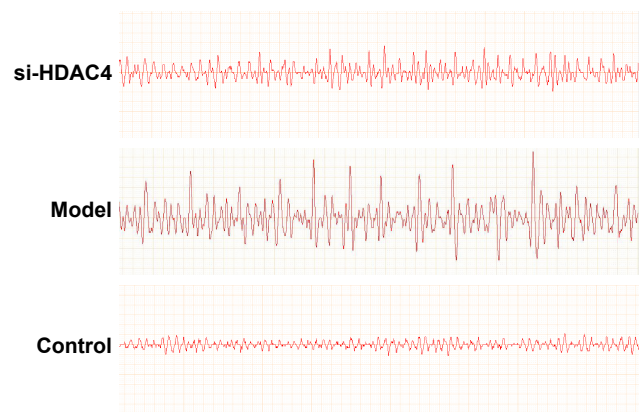


Figure 4 si-HDAC4 inhibits the occurrence of epilepsy using electroencephalogram.

findings revealed that silencing HDAC4 could inhibit the occurrence of epilepsy.

Silence of HDAC4 promotes cognitive function

Morris water maze test was used in order to detect the cognitive ability of rats in each group, with the results shown in Figure 5. In comparison with the control group, the escape latency of the epileptic rats in the si-HDAC4 group decreased, with the time spent in the target quadrant increasing successfully on the 1st day, 14th day, and 30th day ($P < 0.05$); both the model group and the control group showed no significant differences in the escape latency as well as the time spent in the target quadrant on the 1st day, 14th day, and 30th day (all $P > 0.05$). At the same time, in comparison with the control group, the escape latency of the epilepsy group significantly increased ($P < 0.05$), while the time spent in the target quadrant significantly reduced ($P < 0.05$); at the same time, in comparison with the epilepsy group, the escape latency of

the si-HDAC4 group significantly decreased ($P < 0.05$), with the time spent in the target quadrant becoming significantly enhanced ($P < 0.05$).

Discussion

In accordance with the current research, various types of diseases have been highlighted with different expression of HDAC. HDAC has been shown to play a key role in many attempts to cure disease. RNA interference has also been shown to be a powerful research tool, as well as the siRNA inhibiting endogenous gene and exogenous gene expressions in mammal cells.²² During this study, it had also been discovered that the HDAC4 gene silencing played a role in providing relief in the seizures experienced by the subject epileptic rats. Upregulation of GABAAR α 1 and GABAAR α 4, while downregulation of GAD65, GAT-1, and GAT-3 in epileptic rats allowed us to come to the idea that HDAC4 is likely to become a new therapeutic target in the treatment of epilepsy.

The GABAA receptor is known as an inhibitory neurotransmitter receptor closely related to epilepsy within the CNS; α 1 and α 4 are important among GABAAR subunits, which are widely distributed in neurons of normal brain tissues such as the hippocampus, cortex, and thalamus.^{23,24} Expressions of GABAAR, GABAAR α 1, and GABAAR α 4 were found in the brain tissues of the epileptic rats and especially decreased in the hippocampal formation.²⁵ HDAC is a chromatin modifying factor, affecting the growth of nerves by regulating the transcription factor and controlling the related expressions of its downstream function genes.²⁶ CREB is an important transcription factor regulating nerve's growth.²⁷ Evidentiary data have indicated that HDAC regulates the plasticity of memory and synapse

Table 2 Comparisons of rats' behavior observation among three groups

Time	Group	Latency (s)	Percentage of incidence (\geq grade 3) (%)	Seizure duration (h)	Percentage of incidence (\geq ten times) (%)
Day 1	si-HDAC4 group	18.32 \pm 1.88	90	1.55 \pm 0.17	90
	Epilepsy group	21.63 \pm 2.17	90	1.89 \pm 0.17	90
	Control group	–	–	–	–
Day 14	si-HDAC4 group	16.29 \pm 1.64	80	1.07 \pm 0.14	90
	Epilepsy group	19.47 \pm 1.95	90	1.46 \pm 0.15	90
	Control group	–	–	–	–
Day 30	si-HDAC4 group	7.65 \pm 0.77*#	50*#	0.21 \pm 0.07*#	40*#
	Epilepsy group	14.37 \pm 1.34	80	0.84 \pm 0.08	80
	Control group	–	–	–	–

Notes: * $P < 0.05$ vs the epilepsy group at the same time point; # $P < 0.05$ vs the first day of administration in the same group; n=10. The data were presented as mean \pm SD.

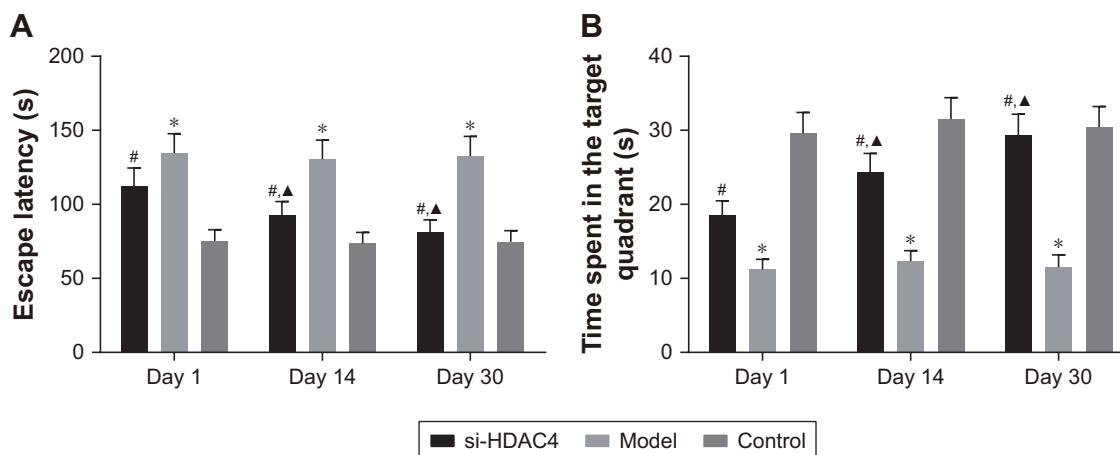


Figure 5 Silencing of HDAC4 promotes cognitive function.

Notes: (A) Escape latency of rats in each group during localized navigation experiment; (B) residence time of rats in the target quadrant of the original platform after removal of the platform; * $P < 0.05$ vs the control group at the same time point; # $P < 0.05$ vs the epilepsy group at the same time point; ▲ $P < 0.05$ vs the first day of administration in the same group.

negatively through CREB.²⁷ Through CREB's original response of luciferase reporter gene, it has been discovered that the transfer of HDAC4 into the nucleus directly inhibits transcriptional activity of CREB.²⁸ HDAC4, in the process of regulating gene expression, does not only inhibit gene expression through histone deacetylation but also controls the transfection factor's function through its binding with the transfection factors.²⁹ In muscle tissues and bones, HDAC4 can bind with MEF2 in the nucleus to inhibit the MEF2 activity, inhibiting the differentiation of muscles and bones.³⁰ Moreover, HDAC4 can also bind with MEF2 and CREB and inhibit their expressions to lead to neuronal death.³¹ In 2006, Payne et al discovered that the abnormal expression of GABA $\alpha 1$ receptor could lead to an epileptic-like abnormal discharge in mice, providing evidence that the GABA $\alpha 1$ receptor is closely linked to the discharge induced by an abnormal excitation of neurons.³² Mao et al further demonstrated the role of GABA $\alpha 1$ receptor in abnormal discharge of hippocampal neurons in 2010.³³ Therefore, it can be deduced that the reason for the increasing of GABAAR $\alpha 1$ and GABAAR $\alpha 4$ expressions lies within the idea that HDAC4 gene silencing improves the nerve electrophysiological function of cortex as well as the hippocampus by increasing collectively the GABAAR $\alpha 1$ and GABAAR $\alpha 4$ expressions, while decreasing excitability of the nervous system, and furthermore, by inhibiting high-frequency discharges of neurons to further inhibit epilepsy.

RT-qPCR results also showed us that the mRNA levels of GAD65, GAT-1, and GAT-3 in epilepsy rats treated with silenced HDAC4 significantly downregulated, which was proven once again by the Western blot analysis. GAD

as a rate-limiting enzyme synthesized by GABA can affect GABA content in the brain. The content of the GAD is similar to that of the GABA neurons, with GAD frequently used as a direct marker for the GABA neurons.³⁴ It has been displayed that epilepsy can lead to GAD changes in the brain and as a result of these changes, a decrease in the overall expression of GAD.³⁵ These findings are parallel with the results achieved throughout our study noting that HDAC4 can suppress epilepsy seizures of rats by decreasing GAD65 expressions. One study has shown that an increased GAT-1 expression after epileptic seizures is an important etiological link in the process of epileptic disease, the increase of GAT-1, so that more GABA is cleared out of the intercellular space, neural network inhibition function is further weakened, and the epileptic symptoms become aggravated.³⁶ Furthermore, a study conducted by Pirttimaki et al reported that GAT-1 and GAT-3 expressions were decreased following an episode of epilepsy, potentially due to a reactive change in the organism that decreased the intake in the transport of GABA as well as inhibited the discharge generalization of the neurons to inhibit the seizure of epilepsy.³⁷ Mattiske et al analyzed the differentially expressed genes of ARX mutant mice and normal mice by RNAseq, finding that the transcriptional regulators such as HDAC4 may be closely related to epilepsy.³⁸ It has also been reported that loss of the putative catalytic domain of HDAC4 may lead to a reduced thermal nociception and seizures.³⁹ To sum this all up, HDAC4 gene silencing can inhibit both GAT-1 and GAT-3, thereby downregulating the expressions of both GAT-1 and GAT-3 in the brain. In addition, HDAC4 silencing can cause a reduction in the intake and transport of GABA, potentially increasing the

concentration of GABA in the brain synaptic space, thus indicating its antiepileptic function.

Conclusion

In conclusion, this study has demonstrated that the HDAC4 gene silencing could reduce seizure degree, latency, duration of seizure, the amount of scattered epileptic waves, and occurrence of epilepsy. In addition, HDAC4 gene silencing provided a superior controlling mechanism in regard to the outbreak of epilepsy in rats. This controlling is essentially a product of the upregulation of GABAAR α 1 and GABAAR α 4 expressions along with the downregulation of GAD65, GAT-1, and GAT-3 expressions. However, the detailed mechanism still needs more exploration, which will be taken care of in further experiments.

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Disclosure

The authors report no conflicts of interest in this work.

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