



Breaking bad family ties: Pan-ERBB blockers inhibit KRAS driven lung tumorigenesis

Herwig P. Moll ^a and Emilio Casanova ^{a,b}

^aDepartment of Physiology, Center of Physiology and Pharmacology & Comprehensive Cancer Center (CCC), Medical University of Vienna, Vienna, Austria; ^bLudwig Boltzmann Institute for Cancer Research (LBI-CR), Vienna, Austria

ABSTRACT

Oncogenic K-RAS mutations were believed to lock the molecular switch in the ON state, independent of upstream activation. However, we demonstrate in preclinical models that activity of mutated K-RAS depends on upstream signaling events involving EGF receptor family members. This finding reveals a potential therapeutic vulnerability using pan-ERBB inhibitors to fight K-RAS mutated lung tumors.

ARTICLE HISTORY

Received 27 July 2018
Revised 13 August 2018
Accepted 16 August 2018

KEYWORDS

NSCLC; KRAS mutations;
ERBB signaling; tyrosine
kinase inhibitors; afatinib

Oncogenic kirsten rat sarcoma oncogene homolog (KRAS) mutations were believed to lock the molecular switch in the ON state, independent of upstream activation. However, we demonstrate in preclinical models that activity of mutated KRAS depends on upstream signaling events involving epidermal growth factor (EGF) receptor family members. This finding reveals a potential therapeutic vulnerability using pan-ERBB inhibitors to fight KRAS mutated lung tumors.

Despite the advent of targeted therapy and immunotherapy, lung cancer still remains the deadliest form of cancer.¹ In general, lung cancers are sub-classified into several subtypes based on morphology, and further subdivided according to the tumor driving oncogene. Overall, kirsten rat sarcoma oncogene homolog (KRAS) mutated lung adenocarcinomas (LUAD) represent the most common group of all lung cancers. Unfortunately, and in contrast to other frequently mutated oncogenes as for instance the epidermal growth factor receptor (EGFR) there are no direct inhibitors targeting mutant KRAS available in the clinics. Accordingly, research focused to identify up- or downstream signal mediators to reduce activity of mutated KRAS. In this context, initial clinical studies using EGFR targeting first generation tyrosine kinase inhibitors (TKI) erlotinib or gefitinib demonstrated that patients harboring oncogenic mutations in EGFR responded to these inhibitors. However, most of these trials came to the same devastating results indicating that KRAS mutated patients barely responded to erlotinib or gefitinib and it became literally a dogma that KRAS mutations mediate resistance to EGFR inhibitors.^{2–4} Oncogenic KRAS was thought to be locked in a constitutive active form being independent of upstream activators, such as EGFR, thus explaining the tumor resistance to TKI. However, recent reports has challenged this view, opening the possibility to use TKI to treat KRAS mutated tumors.^{5,6} Based in this new findings, we re-examined the use of TKI in KRAS mutated LUAD.

Immunohistochemical analysis of KRAS mutated LUAD patient biopsies showed elevated phosphorylation of various activating sites of the EGFR compared to healthy tissue, suggesting the implication of EGFR signaling in KRAS driven tumorigenesis.⁷ Based on that finding, we initiated studies using genetically engineered mouse models (GEMM) of Kras mutated LUAD. In clear contrast to the clinical data, we found that genetic deletion of Egfr in *Kras*^{G12D} driven lung tumors significantly reduced tumor growth and hence tremendously extended life of tumor bearing mice. We extended our study to human cell line derived xenografts, where we deleted EGFR in the LUAD cell line A549 using CRISPR-Cas9 technology. Indeed, EGFR deficient A549 cells exhibited significantly reduced growth *in vitro* as well as following transplantation into immunodeficient mice. Furthermore, we observed that EGFR knockout reduced the activity of mutated KRAS both in human and in mouse cell lines in agreement with previous reports.^{5,6}

Based on the above, we re-evaluated the potential of EGFR targeting TKI *in vitro*. Confirming the data obtained from patients, different human and mouse LUAD cell lines harboring oncogenic KRAS mutations hardly responded to erlotinib or gefitinib treatment. Unexpectedly, KRAS mutated LUAD cell lines showed 10 – 20 fold increased sensibility towards afatinib treatment compared to erlotinib/gefitinib. Notably, afatinib is a second generation EGFR TKI which covalently binds to EGFR and also exhibits pronounced activity against all other members of the family of EGFRs, namely ERBB2, ERBB3, and ERBB4.⁸ Next, we tested the potential of afatinib based therapies for KRAS mutated LUAD patients *in vivo* by using cell line derived as well as patient derived xenografts. In all these different models we observed a remarkable inhibition of tumor growth in response to afatinib treatment. Hence, we also examined the efficacy of EGFR TKI in already established autochthonous tumors. Therefore we started to treated mice suffering from *Kras*^{G12D} mutated LUAD 10 weeks after tumor induction for a period of another 10 weeks. Strikingly, following afatinib administration

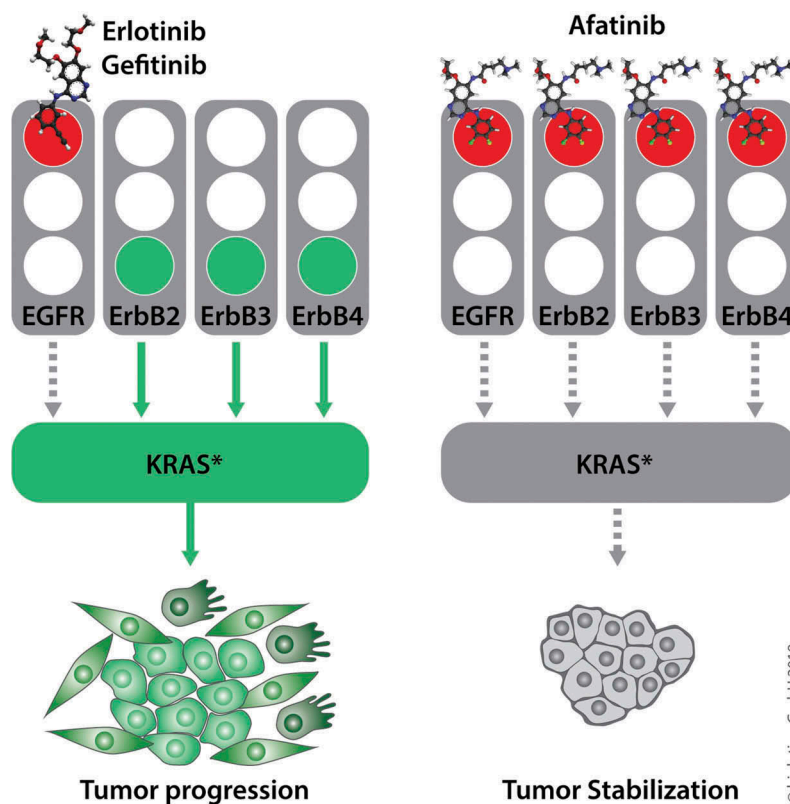


Figure 1. Afatinib restrains kirsten rat sarcoma oncogene homolog (KRAS) driven lung tumorigenesis. Afatinib, but not erlotinib neither gefitinib inhibits activation of all ERBB family members, thereby abrogating mutant KRAS activity and inhibiting tumor progression.

the tumor burden remained unchanged compared to the 10 weeks' time point (i.e. before treatment). This was in clear contrast to erlotinib or gefitinib treated mice, whose tumors remained growing comparable to the vehicle treated control mice. While the latter results confirmed the data observed in clinical trials using first generation TKI, these experiments also highlighted the clinical potential of afatinib based therapies for KRAS mutated LUAD patients.

Nevertheless, it remained still unclear why erlotinib or gefitinib treatment was ineffective, while the second generation TKI afatinib proved to be a promising drug to block KRAS driven LUAD. When culturing the EGFR deficient cells for different follow up experiments, we noticed that these cells quickly regained proliferative capacity, and after passaging these cells for up to 50 times they completely recovered from the burden of EGFR deletion. Analysis of these cells indicated increased expression and activation ERBB2 and ERBB3 most likely to compensate for the loss of EGFR. Similarly, autochthonous *Kras* driven LUAD lacking *Egfr* regained their proliferative capacity at late tumor stages in vivo. This was concomitant with increased ErbB2, ErbB3 and ErbB4 expression and (re)activation of downstream effectors in these tumors. Importantly, also erlotinib or gefitinib induced expression of ERBB2, 3 and 4 and activation of ERBB3 in KRAS mutated LUAD cells, thereby feeding the tumor escape mechanism. However, and in contrast to first generation TKIs, the pan-ERBB TKI afatinib completely blocked activation of all ERBB receptors and thereby prevented the tumor compensatory mechanism (Figure 1).⁷

While in the process of preparing our manuscript for submission, we became aware of a study conducted by Kruspig et al. In this study, the authors demonstrate increased expression of ligands of the ErbB receptors in advanced *Kras* mutated LUAD, and proof that the pan-ERBB inhibitor neratinib was highly efficient in combination with MEK inhibition.⁹ Hence, their works confirms our data that KRAS driven LUAD are sensitive to pan-ERBB TKI. Altogether, both studies provide a solid rationale to revisit the potential of these inhibitors in clinical trials.

ORCID

Herwig P. Moll  <http://orcid.org/0000-0001-6438-9068>

Emilio Casanova  <http://orcid.org/0000-0001-7992-5361>

References

1. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2017. *CA*. 2017;67:7–30. doi:10.3322/caac.21387.
2. Linardou H, Dahabreh IJ, Kanaloupiti D, Siannis F, Bafaloukos D, Kosmidis P, Papadimitriou CA, Murray S. Assessment of somatic k-RAS mutations as a mechanism associated with resistance to EGFR-targeted agents: a systematic review and meta-analysis of studies in advanced non-small-cell lung cancer and metastatic colorectal cancer. *Lancet Oncol*. 2008;9:962–972. doi:10.1016/S1470-2045(08)70206-7.
3. Mao C, Qiu LX, Liao RY, Du FB, Ding H, Yang WC, Li J, Chen Q. KRAS mutations and resistance to EGFR-TKIs treatment in patients with non-small cell lung cancer: a meta-analysis of 22 studies. *Lung Cancer*. 2010;69:272–278. doi:10.1016/j.lungcan.2009.11.020.

4. Rulli E, Marabese M, Torri V, Farina G, Veronese S, Bettini A, Longo F, Moscetti L, Ganzinelli M, Lauricella C, et al. Value of KRAS as prognostic or predictive marker in NSCLC: results from the TAILOR trial. *Ann oncol*. 2015;26:2079–2084. doi:10.1093/annonc/mdv318.
5. Patricelli MP, Janes MR, Li LS, Hansen R, Peters U, Kessler LV, Chen Y, Kucharski JM, Feng J, Ely T, et al. Selective inhibition of oncogenic KRAS output with small molecules targeting the inactive state. *Cancer Discov*. 2016;6:316–329. doi:10.1158/2159-8290.CD-15-1105.
6. Lito P, Solomon M, Li LS, Hansen R, Rosen N. Allele-specific inhibitors inactivate mutant KRAS G12C by a trapping mechanism. *Science*. 2016;351:604–608. doi:10.1126/science.aad6204.
7. Moll HP, Pranz K, Musteanu M, Grabner B, Hruschka N, Mohrherr J, Aigner P, Stiedl P, Brcic L, Laszlo V, et al. Afatinib restrains K-RAS-driven lung tumorigenesis. *Sci Transl Med*. 2018;10. doi:10.1126/scitranslmed.aao4496.
8. Solca F, Dahl G, Zoepfel A, Bader G, Sanderson M, Klein C, Kraemer O, Himmelsbach F, Haaksma E, Adolf GR. Target binding properties and cellular activity of afatinib (BIBW 2992), an irreversible ErbB family blocker. *J Pharmacol Exp Ther*. 2012;343:342–350. doi:10.1124/jpet.112.197756.
9. Kruspig B, Monteverde T, Neidler S, Hock A, Kerr E, Nixon C, Clark W, Hedley A, Laing S, Coffelt SB, et al. The ERBB network facilitates KRAS-driven lung tumorigenesis. *Sci Transl Med*. 2018;10. doi:10.1126/scitranslmed.aao4496.