



Review

The Blood-Brain Barrier and Its Intercellular Junctions in Age-Related Brain Disorders

Laura Costea ¹, Ádám Mészáros ^{2,3}, Hannelore Bauer ⁴, Hans-Christian Bauer ⁵, Andreas Traweger ⁵, Imola Wilhelm ^{1,2}, Attila E. Farkas ^{2,6,*,†} and István A. Krizbai ^{1,2,*,†}

- ¹ Institute of Life Sciences, Vasile Goldiş Western University of Arad, 310414 Arad, Romania; laura.m.costea@gmail.com (L.C.); wilhelm.imola@brc.hu (I.W.)
- Institute of Biophysics, Biological Research Centre, 6726 Szeged, Hungary; meszaros.adam@brc.hu
- Doctoral School of Biology, University of Szeged, 6726 Szeged, Hungary
- Department of Biological Sciences, University of Salzburg, 5020 Salzburg, Austria; hannelore.bauer@sbg.ac.at
- Institute of Tendon and Bone Regeneration, Paracelsus Medical University—Spinal Cord Injury and Tissue Regeneration Center Salzburg, 5020 Salzburg, Austria; hans.bauer@pmu.ac.at (H.-C.B.); andreas.traweger@pmu.ac.at (A.T.)
- Department of Physiology, Anatomy and Neuroscience, University of Szeged, 6726 Szeged, Hungary
- * Correspondence: farkas.attilae@brc.hu (A.E.F.); krizbai.istvan@brc.hu (I.A.K.); Tel.: +36-62-599601 (A.E.F. & I.A.K.)
- † These authors contributed equally to this study.

Received: 12 October 2019; Accepted: 1 November 2019; Published: 3 November 2019



Abstract: With age, our cognitive skills and abilities decline. Maybe starting as an annoyance, this decline can become a major impediment to normal daily life. Recent research shows that the neurodegenerative disorders responsible for age associated cognitive dysfunction are mechanistically linked to the state of the microvasculature in the brain. When the microvasculature does not function properly, ischemia, hypoxia, oxidative stress and related pathologic processes ensue, further damaging vascular and neural function. One of the most important and specialized functions of the brain microvasculature is the blood-brain barrier (BBB), which controls the movement of molecules between blood circulation and the brain parenchyma. In this review, we are focusing on tight junctions (TJs), the multiprotein complexes that play an important role in establishing and maintaining barrier function. After a short introduction of the cell types that modulate barrier function via intercellular communication, we examine how age, age related pathologies and the aging of the immune system affects TJs. Then, we review how the TJs are affected in age associated neurodegenerative disorders: Alzheimer's disease and Parkinson's disease. Lastly, we summarize the TJ aspects of Huntington's disease and schizophrenia. Barrier dysfunction appears to be a common denominator in neurological disorders, warranting detailed research into the molecular mechanisms behind it. Learning the commonalities and differences in the pathomechanism of the BBB injury in different neurological disorders will predictably lead to development of new therapeutics that improve our life as we age.

Keywords: aging; blood–brain barrier; tight junction

1. Introduction

Aging and age-related co-morbidities are rapidly increasing unresolved health and socio-economic problems in developed countries. Decline of cognitive brain functions represents one of the main health challenges of aging and includes vascular and neurodegenerative dementias such as Alzheimer's or Parkinson's disease.

The functional state of the central nervous system (CNS) is greatly dependent on the quality of the vasculature. As the centuries old saying goes: "A man is as old as his arteries". Today, especially for the brain, this concept should be redefined: You are as old as your microvessels and capillaries [1]. There is increasing evidence that the cerebral microvasculature and the neurovascular unit play a critical role in age-related brain dysfunctions. The multitude of brain microvascular changes accompanied by aging includes endothelial dysfunction, blood–brain barrier (BBB) breakdown, decrease in blood flow, microhemorrhages, vessel rarefication and neurovascular uncoupling. In this review, our main focus is the breakdown of the paracellular barrier and tight junctions.

2. Cells of the Neurovascular Unit (NVU)

The vasculature in the brain forms a functional unit with the surrounding neural tissue, thus the term neurovascular unit was coined [2]. A functionally intact neurovascular unit (NVU) is a prerequisite for the proper function of the CNS. The most important cellular components of the NVU are cerebral endothelial cells, pericytes, astrocytic endfeet and neurons; however, other cellular elements like microglia may also play a modulatory role. The main role of the NVU besides neurovascular coupling is the formation of the BBB.

Cerebral endothelial cells (CECs) lining brain capillaries are considered the principal barrier forming endothelial cells. They are interconnected by a continuous line of tight junctions and characterized by a high number of mitochondria and low number of caveolae [3–5]. These characteristics contribute to the formation of a paracellular and transcellular barrier.

Pericytes are localized in the duplication of the basement membrane covering the basal surface of the endothelium. The estimates of pericyte coverage show large variations in the literature [6,7]. Pericytes can secrete a large number of substances that may influence endothelial function including TGF β , angiopoetin-1 or VEGF. It seems that the differentiation stage of pericytes determines their effect on the endothelium as well [8]. The role of pericytes in the formation of the BBB is supported by the finding that absence of pericytes leads to endothelial hyperplasia, abnormal vasculogenesis and an increased BBB permeability [9,10].

Although the role of astrocytes in the formation of the physical barrier is limited, due to their influence on cerebral endothelial cells they play an important role in the maintenance of the BBB [11]. The astrocytic endfeet ensheath the brain vasculature almost completely [12] and express the water channel protein, aquaporin 4, which is suggested to play a crucial role in creating a bulk flow in the brain parenchyma from arterioles towards venules. This flow was shown to contribute to the clearance of extracellular proteins and metabolic waste products through the newly rediscovered glymphatic system [13,14]. Despite much interest in the glymphatic system, some experimental results do not support or even contradict its function [15]. Thus further refining of the glymphatic hypothesis is likely necessary [16]. Nevertheless, the aquaporin 4 in astrocytic endfeet is indispensable to BBB function as its knockout results in altered brain microvasculature and decreased water exchange through the BBB [17], furthermore its subcellular distribution shows an age dependent depolarization accompanied by decreased protein clearance [18].

The BBB is in the forefront of the defense line of the CNS and restricts the free movement of solutes and cellular elements between the systemic circulation and neuronal tissue. The BBB is involved in the pathogenesis of a large number of CNS disorders [19].

Cell types comprising the NVU are in close communication in order to maintain physiologic function and react to pathologies. Ligand-receptor type intercellular interactions and ion channels were described early as pathways that coordinate the function of the cell types constituting the NVU [20]. An example of bidirectional information exchange is the role of CD146 in coordinating the development of pericyte coverage on brain vasculature during early ontogenesis. At first CD146 is expressed by endothelial cells but as pericyte coverage increases, CD146 expression shifts to pericytes where it acts as a co-receptor of PDGFRβ. Endothelia attached pericytes down-regulate endothelial CD146 via TGFβ1 secretion, promoting BBB maturation [21]. Recently a growing body of evidence suggests that extracellular vesicle—mainly exosome—mediated bidirectional communication coordinates key functions of the NVU at the local and systemic level as well [22-25]. In a mouse model of spinal cord injury, pericyte-derived exosomes improved microcirculation and protected barrier function [26]. In response to traumatic brain injury, the loss of pericytes and consequent impairment of crosstalk among NVU cells causes barrier dysfunction, brain edema and leakage of cerebral vasculature. A recent example of pericyte to endothelial cell communication in the retina is the circular RNA cPWWP2A that is synthesized in pericytes and downregulates angiopoietin 1, occludin and sirtuin-1expression in endothelial cells by sequestering miRNA-579 [27].

3. Brain Capillaries in Aging

With aging, the density of brain vasculature is decreased and cerebrovascular dysfunction appears to precede and accompany cognitive dysfunction and neurodegeneration. Cerebrovascular angiogenesis is decreased and cerebral blood flow is inhibited by anomalous blood vessels such as tortuous arterioles and thick collagen deposits in the walls of veins and venules [28].

In most mammals, the capacity of CECs to divide is limited and endothelial cells are prone to be senescent. Aging is associated with endothelial dysfunction, arterial stiffening and remodeling, impaired angiogenesis, defective vascular repair and with an increasing prevalence of atherosclerosis [29]. In the aging brain cerebral blood flow declines and perfusion pressure either is constant or increases. In Brown-Norway and Fisher 344/Brown-Norway rats that maintain a relatively consistent cortical volume throughout life the densities of arterioles and arteriole-to-arteriole anastomoses on the cortical surface was found to be decreased with age [30]. In a spontaneously hypertensive rat model, long term hypertension was found to gradually destroy BBB, resulting in white matter lesions, one of the most important pathological changes in vascular dementia [31]. At the capillary level, increased capillary diameters and decreased capillary density paired with increased red blood cell velocities were observed [28,32]. Some capillary density measurements in humans contradict these observations, as no changes were observed in the intervascular distance on CD31 stained brain sections [33].

4. Junctional Proteins in Aging and Related Disorders

The paracellular barrier properties of CECs are determined by the tight junction (TJ), which are composed of transmembrane proteins that control the transport across the intercellular space between adjacent cells and cytoplasmic plaque. Claudin 5, member of the 27 strong claudin family, is primarily responsible for controlling the paracellular transport of water and small molecules through capillary vessel walls [34]. Other claudins may contribute to BBB function as well, though the isoforms present in human and murine brain vessels is still under investigation [35,36]. Like claudins, MARVEL (MAL and related proteins for vesicle trafficking and membrane link) proteins especially occludin play a role in restricting paracellular transport and also regulate homeostasis and TJ organization [37–40]. The third group of integral membrane proteins of the TJ belongs to the CTX (cortical thymocyte marker in *Xenopus*) family within the immunoglobulin superfamily such as junctional adhesion molecules (JAM) and coxsackie and adenovirus receptor (CAR). Integral membrane proteins of the TJ are coupled to cytoplasmic plaque proteins that provide a platform for anchoring the junction to the cytoskeleton and for interactions with signaling molecules. The main components of the cytoplasmic plaque are zonula occludens (ZO) proteins [41–43].

Junctional proteins are not restricted to the endothelial cells of the NVU. Recently, pericytic occludin was described in a new role as a NADH oxidase enzyme and an important player in BBB pericyte metabolism including the modulation of pericyte energy sharing with other NVU components via intercellular transport of mitochondria and glucose [44]. Astrocytic occludin was described two decades ago [45], and a recent report proposed a new role for it in RNA metabolism [46]. Astrocytic TJ, composed of claudin 1, 4 and JAM may also form in inflammation to control lymphocyte segregation [47].

4.1. TJ in Aging and Aging Models

Limited data is available on what changes develop in the function of the BBB and the composition and structure of endothelial TJs in the healthy aging human brain. In a meta-analysis of BBB permeability studies, the barrier function was negatively impacted by age. Though there were some discrepancies, paracellular permeability was generally increased in the aged human brain [48]. Multiple accelerated aging animal models are available to complement this data, which we will discuss mainly together with the appropriate aging associated disorders in later paragraphs. Literature data presented in this chapter are summarized in Table 1. In the review we will include large molecule (e.g., albumin or Evans blue albumin) permeability and blood component extravasation as well even though these are not markers of TJ dysfunction but rather indicate transcellular transport or disruption of the vascular wall respectively.

 Table 1. Blood-brain barrier (BBB) function and junctional proteins in aging.

Experimental System	Claudin 5	Occludin	ZO-1	Permeability	Other	Authors
BubR1 accelerated aging, cell culture	Discontinuous imp		Baker et al., 2011 [49]			
aged mouse and human brain tissue	fragmented, weak immunofluorescence, reduced protein level.		fragmented, weak immunofluorescence	increased 3kDa dextran permeability		Stamatovic et al., 2019 [50]
sirtuin-1 overexpression	rescued by sirtuin-1 overexpression			rescued by sirtuin-1 overexpression.		[2.1]
sirtuin-1 knockdown	reduced claudin 5 mRNA and protein			increased		
senescent female rats				Evans blue extravasation, hippocampus, olfactory bulb		Bake and Sohrabji 2004 [51]
senescent rats	weaker and discontinuous immunofluorescence in hippocampal vessels in senescent compared to young female rats. No change in middle aged vs. young male rats	no change in reproductive senescent vs. young female rats			age and gender dependent IgG extravasation, e.g., increased IgG extravasation in senescent female rat thalamus	Bake et al., 2009 [52]
postmenopausal women	altered distribution					
ovariectomized rats			unaffected	increased	connexin43 redistribution	Wilson et al., 2008 [53]
+estrogen replacement				rescued	rescued	2000 [50]
Pdgfrβ+/–, pericyte deficient mice	All	three decreased.				Bell et al., 2010 [54]
24-month-old rats, compared to 12-months-old 24-month-old rats,		protein decreased, mRNA unaffected protein decreased,	mRNA increased, protein unaffected			Mooradian et al., 2003 [55]
compared to 3-months-old		mRNA unaffected	protein unaffected			
mouse primary brain microvessel endothelial culture. disrupted endothelial integrin/basal lamina interaction	decreased			increased		Osada et al., 2011 [56]
integrin α5 knockout mice	increased mRNA			improved barrier	resistant to MCAO induced stroke	Roberts et al., 2017 [57]

Experimental results linking BBB dysfunction to declining cognitive function prompted the use of modern in vivo imaging such as dynamic contrast enhanced magnetic resonance imaging to study cerebrovascular permeability. A recent work revealed an age dependent BBB permeability increase in the hippocampus of individuals without any cognitive impairment, which was exacerbated in the case of mild cognitive impairment of Alzheimer's, Huntington's and multiple sclerosis patients [58].

Permeability changes are likely the result of decreased expression and disorganized localization of TJ proteins. As shown in the BubR1 hypomorphic murine model of accelerated aging, claudin 5, occludin and ZO-1 expression of cortical and striatal microvessels were decreased in aged mice, with reactive astrogliosis also apparent at the vessels. BubR1 is a key member of the mitotic checkpoint that ensures accurate chromosome segregation [49]. In an experiment using cells isolated from BubR1 hypomorphic mice, aged CECs (co-cultured with pericytes) showed a discontinuous junctional staining for claudin 5, occludin and ZO-1 and the presence of abnormal cells with smaller cytoplasm that are highly immune-reactive for TJ proteins [59]. The BubR1 hypomorphic CECs also showed increased number of spikes in the claudin 5 staining, reminiscent to those previously described as budding and docking sites for claudin bearing vesicles [60].

Aged mouse and human brains both showed fragmented and weak claudin 5 and ZO-1 staining in microvessels compared to healthy young controls, with an accompanying reduction of claudin 5 protein expression. BBB permeability to 3 kDa dextran was also increased in aged mice. Both claudin 5 levels and BBB function were rescued in transgenic mice overexpressing sirtuin-1, a NAD-dependent deacetylase, while inducible, endothelial-specific knockdown of sirtuin-1 increased barrier permeability and reduced claudin 5 mRNA and protein levels [50]. These results add a novel mode in which sirtuin-1 affects longevity and neurological disorders by maintaining BBB integrity [61,62].

Barrier function of the cerebral vasculature was shown to be under the control of estrogen [63]. Thus, a potential influence of the age-dependent decline of estrogen levels on BBB integrity in postmenopausal women or reproductive senescent female animals appeared plausible. In this context, Bake and Sohrabji [51] showed enhanced extravasation of Evans Blue-albumin in the hippocampus and olfactory bulb of reproductive senescent female rats. Moreover, the localization of the tight junction protein claudin 5 in hippocampal microvessels was affected in senescent female rats compared to young adult ones. In case of human brain microvessels, disrupted distribution and poor junctional localization of claudin 5 was altered in postmenopausal women, in contrast to premenopausal women [52]. Further confirmation to suggest that reproductive hormones regulate BBB permeability came from studies with ovariectomized rats [53]. While expression of ZO-1 was unaffected, redistribution and increased expression of connexin-43 was observed following ovariectomy. Estrogen replacement to young (4 months old) ovariectomized rats restored BBB function, while reproductive senescent animals did not benefit from estrogen treatment or even showed increased BBB permeability in the hippocampus [53].

Further evidence of age-related junctional alterations at the BBB came from a study showing that the cerebral occludin protein content in total cerebral tissue extracts from 24-month-old rats was significantly reduced compared to 12-month-old or 3-month-old male rats without a corresponding change in occludin mRNA. ZO-1 protein levels were found to be unchanged while ZO-1 mRNA was significantly increased in 24-month-old rats compared to 3-month-olds [55].

By using transmission electron microscopy, a decline in the pericytic coverage of aged capillaries has been described over thirty years ago [64]. Beyond the assumption that the age-dependent loss of pericytes decreases the ability of the BBB to compensate for transient leaks, there is evidence that pericytes regulate the expression of TJ/BBB-specific proteins in microvascular endothelial cells. A comparison of claudin 5, ZO-1 and occludin expression in 6–8 month old $Pdgfr\beta+/-$ mice has revealed significant reductions of protein expression in pericyte-deficient mice compared to controls [54]. Therefore, it is likely that aging-associated TJ dysregulation may be partly the consequence of pericyte loss.

Integrins are heterodimeric proteins that anchor cells to the extracellular matrix and are involved in ligand dependent bidirectional signaling between the cell and its environment thus playing important roles in all cell functions. Researchers have found connections between integrins and aging [65,66] and neurological disorders [67]. Disruption of cerebral endothelial integrin interaction with the basal lamina lead to decreased claudin 5 levels and BBB integrity [56]. Recently, integrin α 5 knockout mice were found resistant to transient middle cerebral artery occlusion (MCAO) induced ischemic stroke with a small initial infarct volume that disappeared within 3 days. The α 5 knockout animals showed improved barrier function and higher claudin 5 mRNA levels compared to controls [57].

CTX family proteins have not been directly linked to aging; however, some play important roles in the brain development and function or tumorigenesis and invasiveness. JAM-A in CECs undergoes subcellular relocalization in response to inflammatory stimuli [68] and its levels are reduced in response to brain injury [20]. JAM3 was found to be essential for BBB integrity and for normal lens development in humans. Homozygous mutations of JAM3 in humans led to brain hemorrhage and brain calcification [69], the latter being somewhat similar to calcification in the basal ganglia in occludin knockout mice [70]. Brain endothelial function of CAR is scarcely known. However, it was found to play key roles in neurons in synapse development in the postnatal and adult brain and its loss contributes to cognitive defects [71,72].

4.2. Brain Endothelial TJ in Ischemic Injury

In the healthy brain, practically every neuron has a capillary running next to it. With the rarefaction of capillaries in aging brain, the ability to maintain homogeneous flow during periods of localized ischemia is reduced. Hypoxia and reoxygenation are important components of many disorders that affect the central nervous system, including stroke and dementia [73–75]. The incidence of stroke is increased with age and the prognosis of elderly stroke patients is unfavorable compared to young adults [76,77]. Literature data regarding TJs presented in this chapter are summarized in Table 2.

Table 2. BBB function and junctional proteins in age related disorders, ischemic injury and immune system.

Experimental System	Claudin 5	Occludin	ZO-1	Permeability	Other	Authors
hypoxia, 8% O ₂ , mouse, brain lysate	unaffected	decreased protein levels disrupted immuno	unchanged protein levels fluorescence for both	Na-fluorescein leakage increased	MMP and VEGF dependence	Bauer et al., 2010 [78]
hypoxia in primary bovine brain microvessel endothelial culture		membrane localization perturbed	ZO2 membrane localization perturbed, ZO1 and ZO2 diffuse cytosolic staining	sucrose permeability increased	actin stress fibers, increased actin protein	Mark and Davis 2002 [79]
H_2O_2 ROS +GM6001	unaffected	phosphorylation change, degradation rescues degradation			TEER increase does not rescue	Lischper et al., 2010 [80]
hypoxia/reoxygenation modeled by DMNQ treatment of primary mouse brain microvessel endothelial culture		protein decreased			asso norrescue	Krizbai et al., 2005 [81]
hypoxia in bEND.3	altered localization, decreased protein			increased		Koto et al., 2007 [82]
reactive oxygen species	altered localization, d	ecreased protein for both			xanthine oxidase and 100 μM hypoxanthine	Schreibelt et al., 2007. [83]
MCAO, 12 months vs. 2 months old mice	decrease in both age groups, after 3 days less protein in old compared to young		decrease in both age groups, after 1 and 3 days less protein in old	larger Evans blue extravasation in old with stroke		Shen et al., 2019 [84]
young adult rats hypoxia/reoxygenation		phosphorylation	no change in protein	increased		Witt et al., 2003 [85]
young adult rats hypoxia/reoxygenation as above + free radical scavenger tempol		disrupted rescued				Lochhead et al., 2010 [73]
young adult rats hypoxia/reoxygenation	disrupted localization, slightly increase membrane fraction	disrupted localization, slightly increase membrane fraction	disrupted localization	PKC-dependent focal BBB leakage		Willis et al., 2010 [86]
EGFP-Claudin 5 transgenic mouse tMCAO after 24–30 hours	eGFP-claudin 5 no change	no change	no change		Claudin 5 transgene protein expression is double of wild type.	Knowland et al., 2014 [87]
after 48–58 hours	eGFP-claudin 5 non membrane	reduced	reduced		double of what type.	

 Table 2. Cont.

Experimental System	Claudin 5	Occludin	ZO-1	Permeability	Other	Authors
600–950 g rats compared to 200–250 g rats		degradation				Kaur et al., 2011
MCAO, both ages	slightly increased phosphorylation greatly increased					[88]
tPA after reperfusion, only in old	phosphorylation	increased degradation				
rat tMCAO +PP2 Src kinase inhibitor, brain tissue	time dependent decrease of claudin 5 protein and mRNA was rescued by PP2			brain lysate protein dec from 24, mRNA from 72 days later, fibrinogen coincided claudin 5 decr by PI	hours and persisted 7 leakage from brain ease and vas rescued	Bai et al., 2014, [89]
rat microsphere embolism, capillary lysate		decreased amount, increased phosphorylation 6–72 h after microsphere injection	decreased amount	albumin leakage starting from 48 hours		Kago et al., 2006 [90]
rat microsphere embolism, capillary lysate, +PP2		decreased amount and increased phosphorylation rescued by PP2		albumin leakage 6-24h, rescued by PP2		Takenaga et al., 2009 [91]
spontaneously hypertensive rats tMCAO, brain sections	reperfusion + MMP2 a	n in localization 3 h after activity at vessels. Protein in astrocytes.		Increased gelatinase reperfusion, rescued by		Yang et al., 2007, Yang and
brain lysate	decreased mRNA, deg	graded protein rescued by IMP inhibitor				Rosenberg 2011 [92,93]
spontaneously hypertensive rats tMCAO+GM6001 MMP inhibitor	pro	tein levels were rescued for all	three			Yang et al., 2013 [94]
Primary human brain microvascular endothelial cells+PARP inhibition	increased prot	ein levels for both				Rom et al., 2015 [95]

 Table 2. Cont.

Experimental System	Claudin 5	Occludin	ZO-1	Permeability	Other	Authors
spontaneously hypertensive rats tMCAO + mynocicline	increased protein levels for all three					
SRF, MRTF-A/-B knockdown hemorrhagic stroke model, whole brain tissue	claudin1, 3, 5 and 12 mRNA and claudin5 protein downregulated		ZO-2 and ZO-3 mRNA downregulated, ZO-1 unaffected			Weinl et al., 2015 [97]
bovine brain microvessel endothelial cells +VEGF			om membrane to cytoplasm, vels increased	permeability increase, resistance decrease		Wang et al., 2001 [98]
adenoviral expression of IL-1 in VEGFA knockout mouse		rotein expression of brain nmatory lesions for both				Argaw et al.,
EAE in VEGFA KO mouse	protein expression of brain microvessels in inflammatory lesions					2012 [99]
VEGFR2 blocking antibodies in Cortical VEGF injected wild type mice	•	expression for both				
eNOS inhibition or silencing in human brain microvessel endothelial cells	rescued protein expression					
24 months old mice compared to 3 months old, via fluorescence activated cell sorting	significantly decreased		decreased	IgG leakage	neurovascular inflammation and neuronal stress	Elahy et al., 2015 [100]
SAMP8 mice, 12 vs. 4 months old, permeability				no change in albumin permeability and insulin uptake		Banks et al., 2000 [101]

In vitro studies of hypoxia and hypoxia followed by reoxygenation have revealed much of the cellular mechanisms affected by ischemia in CECs. In oxidative stress, the interactions of occludin with claudins or proteins of the ZO family are affected, directly influencing the formation and function of the TJ. Oxidative stress downregulates occludin, reduces its specific membrane localization and regulatory contribution to barrier tightness via multiple signaling pathways [102].

Significant disruptions were seen in the distribution pattern of occludin after hypoxia [78]. These changes were less apparent in posthypoxic reoxygenation, correlating to the increased protein expression [79]. MCAO caused NADPH oxidase upregulation, ROS (reactive oxygen species) generation, matrix metalloproteinase (MMP)-9 activation, and edema formation [103]. MMP activity in oxidative stress increases paracellular permeability by occludin cleavage. However, applying the same level of MMP activity without oxidative damage, neither is occludin cleaved nor BBB permeability is increased [80]. In an opposite approach, normobaric hyperoxia protected the BBB, and the expression and distribution of occludin against MMP-9-mediated effects in cerebral ischemia [103]. Thus occludin plays a key role in the response of cellular barriers to redox changes and could be a redox sensor at the TJs [104,105]. In a brain endothelial cell culture model of hypoxia/reoxygenation, ERK1/2 was activated and the amount of occludin and barrier function decreased which was exacerbated by glucose deprivation [81]. Hypoxia (1% O₂) altered the location of claudin 5 in the plasma membrane and the level of claudin 5 protein in bEND.3 cells, and these changes were accompanied by an increase in BBB permeability. In vivo, claudin 5 was also significantly reduced under hypoxic conditions [82]. ROS-induced BBB disruption via the redistribution and decrease of claudin 5 and occludin levels was paralleled by cytoskeleton rearrangements. This rearrangement was mediated by RhoA, PI3 kinase and protein kinase B (PKB/Akt), and specific inhibitors prevented ROS-induced monocyte migration across an in vitro model of the BBB [83].

The effect of age on the outcome of ischemic stroke was studied in murine distal MCAO experiments, demonstrating an increased inflammatory reaction in 12 month old mice compared to 2 month olds based on IL-6 and IL-1 β levels in the ischemic tissue. At the same time, there was an increased number of CD68 immunoreactive cells, denoting phagocytosis and microglia activation in peri-infarct tissue and the older mice suffered more severe BBB damage as well [84]. Similar BBB permeability increase was observed in young adult rats after one hour of hypoxia (6% O_2) followed by 10 minutes of reoxygenation, which also lead to increased occludin phosphorylation without significant changes in ZO-1 expression [85]. Disrupted occludin staining of brain endothelial cells in rats was reversible by administration of ROS scavengers [73]. Another study under similar conditions showed PKC-dependent focal BBB leakage and disrupted claudin 5, occludin and ZO-1 organization at endothelial cell borders while the protein levels were slightly increased in the membrane fraction for claudin 5 and occludin [86].

Functional changes of the BBB appear shortly after a hypoxic event, but structural TJ changes develop much slower. Using an EGFP-claudin 5 transgenic mouse model, researchers demonstrated a stepwise alteration of the BBB, leading to an increase in transcytosis followed by alterations of TJ proteins, which undergo significant ultrastructural remodeling and localization changes starting at 48–58 h post tMCAO (transient middle cerebral artery occlusion) [87]. Early permeability increase elicited by tissue plasminogen activator (tPA) during reperfusion in elderly rats was accompanied by greatly increased claudin 5 phosphorylation while unphosphorylated claudin 5 decreased compared to the contralateral side. MCAO treatment itself increased claudin 5 phosphorylation slightly. Occludin protein level was decreased in elderly compared to young rats and further degradation was observed in response to MCAO and tPA. These age-dependent differences in claudin 5 phosphorylation and occludin degradation were inferred from redistribution of western blot bands [88].

In a multifocal cerebral ischemia model, Src kinase dependent tyrosine phosphorylation of occludin was found to contribute to barrier disruption. Systemic inhibition of Src resulted in decreased infarct size, decreased VEGF-A, rescuing claudin 5, occludin expression and barrier function [89–91]. Reperfusion experiments following 90-minute hypoxia in spontaneously hypertensive rats revealed

MMP dependence of BBB leakage and the disruption of claudin 5 and occludin at the TJ in vivo. Interestingly the TJ proteins from disrupted endothelial barrier appeared in adjacent astrocytes underlining intercellular communication [92,93]. In vivo use of the MMP inhibitor GM6001 in rats as an early treatment given at the time of MCAO increased the number of vessels and improved ZO-1, occludin and claudin 5 expression in the infarct region 3 weeks after the ischemic injury [94].

In a sterile inflammation model induced by TNF- α in human primary brain microvascular endothelial cells, PARP inhibition promoted BBB barrier function. Furthermore, PARP inhibition in primary endothelial cells improved TEER and increased occludin and claudin 5 protein levels [95]. In spontaneously hypertensive rats that underwent MCAO, minocycline, an inhibitor of PARP-1 and anti-inflammatory, anti-apoptotic and neuroprotective drug, significantly reduced the infarct size and prevented tissue loss, improved perfusion, reduced BBB permeability and increased ZO-1, occludin and claudin 5 protein levels. At the same time, increased MMP-2 and -3 were detected at four weeks, at which time peri-infarct microglia showed M2 activation [96].

In a recent genetic model of intracerebral hemorrhagic stroke and vascular dementia, cerebral small vessel disease symptoms were elicited by inducible knockdown of the transcription factor, serum response factor (SRF) or its cofactors Myocardin Related Transcription Factor (MRTF-A/-B). The microhemorrhagic phenotype observed in this model is attributed to loss of TJ components and is specific for brain tissue. The authors demonstrated mRNA level decrease of claudin 1, 3, 5 and 12, as well as ZO-2 and ZO-3, and significant loss of claudin 5 protein in isolated brain endothelial cells [97].

Exposing mice to normobaric hypoxia led to an increase in brain vascular permeability associated with diminished expression of occludin and inhibition of VEGF attenuated vascular leakage [78]. This confirms the results of studies in primary brain endothelial cells in which VEGF decreased occludin expression [98]. Astrocyte derived VEGF-A acts directly on brain microvasculature to downregulate occludin protein and mRNA in vitro and in mouse models of experimental autoimmune encephalomyelitis (EAE), inducing BBB permeability. In vivo inactivation of astrocytic VEGF-A expression or systemic use of the eNOS inhibitor cavtratin rescued BBB function [99].

4.3. Involvement of Brain Barriers in Aging-Associated Alterations of Immune Functions

Brain barriers are important interfaces for neuroimmune communication and any disturbance may inevitably result in CNS dysfunctions. Aging-associated alterations at the BBB and brain cerebrospinal fluid (BCSF) barrier are thus considered to be one trigger for the development of cognitive impairment(s) in the aged brain (reviewed in [106,107]). Literature data concerning TJs presented in this chapter can be found at the end of Table 2.

While it is well established that migration of immune cells from the blood into the CNS parenchyma is a process occurring in inflammatory and neurodegenerative state, accumulating evidence suggests that immune cells also infiltrate the perivascular space of non-diseased brain, though at very low levels, maintaining and contributing to the surveillance and homeostasis of the CNS (reviewed in [108–110]).

Less information exists concerning age-related immune cell trafficking across the BBB and BCSF, obviously due to the fact that normal aging of the brain is usually accompanied by at least low level inflammatory activity [111]. Leakage of IgG into the cerebral cortex and hippocampus of healthy aged (24 months old) but not young (3 months old) mice were reported [100]. This leakage was accompanied by a decrease in occludin and ZO-1 expression in cerebral endothelial cells but no signs of leukocyte transmigration into the CNS.

A progressive age-related enhancement of $TNF\alpha$ -elicited T cell infiltration in brain parenchyma and choroid plexus of mice was demonstrated by Xu et al. [112]. Further, Stichel and Luebbert [113] reported on the presence of T cells and dendritic-like cells in mouse brain parenchyma at about 12 months of age, with numbers increasing during aging. T cells from aged mice were found to exhibit increased expression of adhesion molecules, such as CD11a and CD49d, facilitating diapedesis [114] and elevated ICAM1 expression was found in brain vasculature of aged human and mouse brains [115].

The role of aging-associated T cell infiltration of the non-diseased brain is still unclear. Results from single-cell RNA-sequencing and immunofluorescence staining of the subventricular zone from young (3 month old) and old (28–29 months) mice suggest that T cells, expressing interferon- γ , were clonally expanded and markedly enriched in the subventricular zone from old mice [116]. In vitro studies have shown that T cells can inhibit the proliferation of neural stem cells in co-cultures in an interferon- γ -dependent mechanism [116]. Thus, it may be assumed that interaction of T cells with neural stem cells in neurogenic niches of old animals may contribute to the proliferative decline of neural stem cells during aging. A distinct role of immune cells was demonstrated in increasing cerebral vascular permeability [117]. Using a mouse model of CD8 T cell-mediated CNS vascular permeability it was shown that CD8 T cells are capable of initiating BBB disruption in a non-apoptotic perforin-dependent manner.

4.4. Alzheimer's Disease (AD)

The blood–brain barrier contributes to the pathology and may play a causative role in the development of the most common neurodegenerative disease afflicting the elderly population: Alzheimer's disease (AD). The involvement of the BBB happens through multiple mechanisms such as barrier disruption, transporter dysfunction and secretion of neurotoxic substances [118,119]. Amyloid- β peptide (A β), the main component of senile plaques in the AD influences the expression and localization of TJ proteins [120–122]. The extracellular matrix and TJ proteins are substrates of MMPs, whose activity is also increased in AD and after ischemic injury, which results in lower TJ proteins [123]. Data concerning TJ changes in Alzheimer's, Parkinson's and Huntington's diseases, as well as schizophrenia are summarized in Table 3.

Table 3. BBB function and junctional proteins in age related disorders, neurodegenerative diseases.

Experimental System	Claudin 5	Occludin	7	ZO-1	Permeability	Other	Authors
Triple-transgenic mice (3 \times Tg-AD), 10–11 vs. 4 month old					no change in [3H]-inulin and [14C]-sucrose uptake	Age dependent reduction in cerebrovascular volume, thickened basal membrane	Bourasset et al., 2009, Mehta et al., 2013 [124]
18–20 month old 3 × Tg-AD vs. wild type mice					no change in [3H]-inulin and [14C]-sucrose uptake decreased uptake of passively diffusing markers: [3H] diazepam and [3H] propranolol in AD mice	thickened basal membrane	
rat brain microvessel endothelial cells +Aβ1-42 for 24–72 h, immunofluorescence	disrupted localization after 24 h, increased fluorescence of Claudin 5 and 1.	no change in localization. De fluorescence at 24 hour		ZO-2 diffuse localization. ZO-2 increased fluorescence at 24 h and decreased at 72 h.			Marco and Skaper 2006 [120]
capillary cerebral amyloid angiopathy (capCAA) patient brain sections hCMEC/D3 cell culture + Aβ1-42 for 24 h	loss of protein in capillaries with amyloid β deposits dose dependent, ROS mediated decrease of mRNA for both						Carrano et al., 2011, 2012 [125,126]
ARPE-19 cell culture + oligomeric Aβ1-42 subretinal Aβ1-42 injection in C57BL/6 J mice	disrupted membrane staining, increased 40 kDa significantly decreased mRNA FITC-dextran permeability Similarly disrupted membrane staining of both in young, adult and aged mice.						Bruban et al., 2009 [127]
bEnd.3+Aβ1-42	decreased protein levels rescued by decreased protein level inhibition of either RAGE, calcineurin or MMP increased 40 kDa FITC-dextran permeability rescued by inhibition of calcineurin or MMP					Kook et al., 2012 [128]	
8 months old transgenic mice with five familial AD mutations (5XFAD) compared to wild type	TJ cross section len	gth significantly decreased, by	transmissi		RAGE expression was increased in brain capillaries		
bEnd.3	Oligomeric, but not monomeric or fibrillar $A\beta1-42$ dose dependently decreased protein levels. Protein levels were rescued by silencing RAGE with siRNA or blocking RAGE with a polyclonal antibody.						

 Table 3. Cont.

Experimental System	Claudin 5	Occludin	ZO-1	Permeability	Other	Authors		
bEnd.3 + oligomeric Aβ1–42	A β 1-42-Oligo induced expression of MMP-2 decreased protein levels and increased permeability were rescued by EGb761 in a dose dependent manner and MMP-9 was rescued by blocking RAGE via antibody.							
bEnd.3 cells+ Aβ1-42 and isolated capillaries from 5XFAD mice	exogenous ANXA1 rescued Aβ1-42 induced decrease in protein levels by blocking Rho-ROCK signaling	ANXA1 rescued Aβ1-42 induced ecrease in protein evels by blocking Rho-ROCK exogenous ANXA1 rescued Aβ1-42 induced decrease in protein levels by blocking Rho-ROCK						
APOE4 transgene expressing Apoe-/- mice	protein levels were decreased dependent on the pericytic cyclophilin A/NF-κB/MMP9 dependent pathway							
bEnd.3, CD-1 or PARP-1 KO mouse primary glial culture +Aβ1–42	endothelial protein expression reduced by activated microglia was rescued by PARP inhibition.							
MPTP mouse model +caffeine	MPTP treated animals showed decreased protein levels, which was rescued by caffeine. Caffeine rescued MPTP induced increases in MPTP-induced increases in MMP9 activity							
MPTP mouse model +trehalose	immunofluorescent signal was greatly diminished in MPTP animals, which was rescued by trehalose.							
unilateral striatal 6OHDA injection in rats				FITC-albumin or horseradish peroxidase leakage in striatum and substantia nigra in response to 6OHDA		Carvey et al., 2005 [135]		
unilateral striatal 6OHDA injection in rats	immunohistochemist signal decreased	ry protein	levels decreased for both			Huang et al., 2016 [136]		
hCMEC/D3 (co-cultured with mouse primary astrocytes) + preformed α-synuclein fibrils		decreased prote	in levels			Kuan et al., 2016 [137]		

 Table 3. Cont.

Experimental System	Claudin 5	Occludin	ZO-1	Permeability	Other	Authors
Parkinson's Disease patient tissue sections compared to age matched and deep brain stimulated patient tissue	decreased immuno	0	ompared to control, rescued by deep brain lation.	IgG extravasation		Pienaar et al., 2015 [138]
Parkinson's disease patient tissue sections				blood extravasation		Gray and Woulfe 2015 [139]
Huntington's disease patient putamen samples compared to control	de	creased protein levels		extravascular fibrin deposition., increased leakage in caudate and putamen by DCE MRI	abnormal vessel morphology	Drouin-Ouellet et al., 2015 [140]
and R6/2 mouse model, striatal samples	de	creased protein levels		increased pinocytic activity and increased albumin extravasation	morphological changes in vessel walls	
iPSC derived brain microvascular endothelial cells from HD patients	miss-localization to the cytoplasm			decreased TEER		Lim et al., 2017 [141]
schizophrenic patient brain tissue	discontinuous localization			anti-psychotic medications de claudin 5 ex	1 ,	Greene et al., 2018. [142]

Similar to data on permeability in the aging brain, the BBB permeability data of AD patients was heterogeneous. This could partly be ascribed to difficulties in separating different types of dementia and co-morbidities confounding results. In a meta analysis by Farrall and Wardlaw some results show increased BBB permeability in AD compared to controls but the permeability increase is higher and more clear cut for vascular dementia and mixed dementias compared to AD [48]. Murine models of AD allow for more precise control over experiments and make it possible to concentrate on separate aspects of the disease; however, available results are still conflicting. A number of studies showed that the paracellular route is unaffected in murine models of AD. There was a substantially decreased occludin expression in APP/PS1 mice AD model compared to wild-type controls [143]. In some cases, when using an APP/PS1 mouse model there was an unaltered BBB permeability to ¹³¹I-albumin [144] and to sodium fluorescein [145]. Even though earlier work showed small localized disruption in Senescence Accelerated Mouse-Prone 8 (SAMP8), no global BBB permeability change was observed when comparing 12 and 4 months old animals [101]. Comparing 11 and 4 months old $3 \times \text{Tg-AD}$ mice (triple transgenic mice for familial Alzheimer's disease mutations), Bourasset et al. found no changes in the distribution of vascular space markers ³H-inulin and ¹⁴C-sucrose, but found a global reduction in cerebrovascular volume that was most significant in the hippocampus [124]. In a comparison of 18–20 month old 3×Tg-AD and wild type mice researchers also did not find changes in ³H-inulin and ¹⁴C-sucrose uptake, while the uptake of passively diffusing markers ³H-diazepam and ³H-propranolol was less in AD mice [146]. These results suggest that in murine AD models global BBB breakdown is not detectable and research probably should focus on local barrier disruption, as data from multiple research groups point to the importance of cerebral amyloid angiopathy in AD and there is ample amount of research revealing the molecular mechanisms behind it.

Regarding TJ proteins in post-mortem human brain, a dramatic loss of claudin 5, occludin and ZO-1 was demonstrated in A β -laden capillaries surrounded by NADPH oxidase-2 (NOX-2)-positive activated microglia. Further results show that A β is toxic to endothelial cells in the human brain via binding to the receptor for advanced glycation end-products (RAGE) and induction of ROS production, which ultimately leads to disruption of TJs and loss of BBB integrity [125,126]. In a model of age-related macular degeneration, increased formation of ROS was found after administration of the oligomeric form of A β 1–42 to retinal pigmented epithelial cells [127], which was accompanied by a decrease in the expression of occludin. The A β 1-42 peptide accumulation around brain microvessels was found to be mediated by transport via RAGE in 5XFAD mice, increasing permeability, disrupting claudin 5, ZO-1 and occludin. This MMP and Ca²⁺-calcineurin dependent BBB disruption could be prevented in bEnd.3 cells by neutralizing antibodies against RAGE [128], small interfering RNA knockdown of RAGE [129] or *Ginkgo biloba* extract EGb761 [130].

An in vitro study on A β 1-42-treated bEnd.3 cells showed that AnnexinA1 (ANXA1), an anti-inflammatory messenger, significantly rescued the expression of claudin 5 and ZO-1 and barrier function in A β 1-42-treated bEnd.3 cells. A β 1-42 reduced ANXA1 bEnd.3 cells, and also had reduced expression in capillaries of 5XFAD mice, and the human serum of patients with AD. ANXA1 acted via the inhibition of RhoA-ROCK signaling. In co-culture experiments, pericyte secreted ANXA1 attenuated the A β 1-42-induced disruption of the tight junction [131].

Pericytes influence the BBB by promoting TJ protein expression in endothelial cells [54] and helping the alignment of TJs [147]. It has also been shown that a loss of pericytes plays a role in AD development and is followed by a decreased expression of TJ proteins [148,149].

The strongest genetic risk factor for late onset AD is Apolipoprotein E4 (ApoE4). ApoE4 and its receptors are expressed throughout the NVU and are linked to many aspects of cerebrovascular dysfunction [150,151]. The molecular mechanisms of ApoE4 mediated neurovascular injury demonstrate how the coordinated effort of multiple cell types maintains a functional NVU. Bell et al. revealed using multiple transgenic mice that the ApoE induced degradation of TJ proteins claudin 5, ZO-1 and occludin and basal membrane protein collagen IV is the result of an intercellular

communication error. ApoE4 secreted by astrocytes is unable to bind LRP1 on pericytes and thus does not block the pericytic cyclophilin A/NF- κ B/MMP9 pathway, which results in vascular dysfunction [152].

Soluble $A\beta$ is also known to induce secretion of proinflammatory cytokines (TNF and IL-6) and chemokines, which stimulate the production of MMP-2 and MMP-9 and it also activates the production of ROS [153]. Furthermore, experimental data confirms the involvement of all cells of the NVU in the effect of $A\beta$. Microglia activated by $A\beta$ treatment shows reduced expression of trophic factors that are responsible for inflammatory resolution and increased pro inflammatory NO and TNF α release. This affects both astrocytes and capillary endothelium leading to reduced BBB integrity and function [132].

AD is also accompanied by tau protein accumulation and hyperphosphorylation, which was shown to promote BBB dysfunction in AD and other tauopathies [154–156].

4.5. Parkinson's Disease (PD)

The BBB is also involved in the progression of the second most common neurodegenerative disorder: Parkinson's disease (PD). The contribution of BBB disruption to PD is not widely studied despite the implication of known BBB damaging mechanisms and agents such as oxidative stress and MMPs in the pathomechanism of the disease [157,158]. Thickened basement membrane in the cingulate cortex and degeneration of the brain microvasculature in PD was reported by Farkas et al. and Guan et al. [159,160]. Gray and Woulfe published the first report of BBB disruption evidenced by blood extravasation in striatal PD tissue in 2015 [139]. Motor function and regional blood flow can be improved in PD patients by deep brain stimulation [161], which is possibly a result of normalizing aberrant microvasculature in PD. In a study regarding the effects of deep brain stimulation of the subthalamic nucleus in PD patients, the decreased immunofluorescence signal of claudin 5, occludin and ZO-1 of PD samples could be rescued by deep brain stimulation treatments [138]. This is in line with multiple studies using experimental parkinsonism models. Significant decrease was detected in the amount of occludin in the striatum, which was associated with increased BBB leakage, in the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)-treated mouse model. Increased striatal MMP-9 activity was also detected in the MPTP model with a possible role in TJ opening [133,134]. Increased permeability of the BBB was also observed in the striatum in rats after treatment with unilateral injections with 6-Hydroxydopamine (6OHDA) [135] that leads to downregulation of occludin [136]. Similar to amyloid oligomers in AD the Lewy body component α -synuclein was found to damage BBB. An in vitro study showed that preformed α -synuclein fibrils lead to a decrease in the expression of occludin and ZO-1, but not tricellulin or marveld3, in human brain microvascular endothelial cells [137].

4.6. Huntington's Disease (HD)

Huntington's disease (HD) is a dominantly inherited autosomal neurodegenerative disorder that is not usually considered aging related as it is typically diagnosed at the age of 40, and the onset of the disease can vary from earlier than ten to over eighty years of age. However, recent research indicates that epigenetic age acceleration in specific brain regions is associated with HD [162].

Recently, morphological changes of blood vessels and BBB leakage in the caudate and putamen were observed in HD patients using magnetic resonance imaging. In putamen samples of HD patients and in striatal samples of the HD model R6/2 mice, occludin and claudin 5 protein levels were decreased and evidence of increased BBB permeability was found [140]. More recently, these results were expanded in induced pluripotent stemm cell (iPSC) derived brain microvascular endothelial cells. While in this system no significant changes in TJ protein levels were observed, claudin 5 was found to mislocalize to the cytoplasm of cells derived from HD patients; however, trans endothelial electric resistance (TEER) and MDR1 function was significantly decreased compared to control cell lines. Some of the changes in cell lines derived from HD patients could be attributed to aberrant Wnt signaling [141].

4.7. Schizophrenia

Some aspects of schizophrenia raise the possibility that accelerated aging of the brain can be responsible for some of its symptoms [163]. Individuals with the chromosomal abnormality: 22q11 deletion syndrome (22q11DS) have an increased risk of developing schizophrenia, in fact it occurs in 30% of such individuals. Interestingly, 22q11DS are haploinsufficient for claudin 5. Recent experiments on AAV mediated claudin 5 suppression in mice and inducible claudin 5 knockdown uncovered focal BBB breakdown and behavioral changes. Anti-psychotic medications were found to dose dependently increase claudin 5 expression. Furthermore post mortem analysis of schizophrenic brains revealed aberrant, discontinuous expression of claudin 5 [142].

5. Closing Remarks

Aging comes with the deterioration of all bodily functions of which the loss of cognitive function is possibly the most serious in our heavily information dependent society. With the building evidence that dysfunction of the microvasculature is not just coincident but is part of the underlying mechanisms of aging and associated neurovascular and neurological disorders, new therapeutic possibilities are opened. The significant heterogeneity of BBB disruption data in studies using aging postmortem brain tissue suggests that more data is necessary to clearly understand the role of BBB disruption and to see whether it is a symptom or a cause [48]. It is not surprising that human BBB data before the mid-1990s is mainly regarding permeability; however, even since the discovery of occludin, zonula occludens proteins and claudins, BBB TJ data in the aging brain is scarce and this deficit is just partly made up for by data from aging associated disorders. Furthermore many studies disregard that claudin 5, occludin or ZO1 does not constitute the paracellular barrier by itself and all three should be studied along with permeability to different molecular size tracers at the same time to get a picture of barrier status. Worsening the situation is that the reliability of existing TJ data has been questioned recently [35]. Thus further comprehensive BBB TJ and permeability studies are needed in the field of aging and aging associated disorders. Another focus needs to be the study of both classical and novel intercellular communication pathways between brain capillary endothelium, pericytes, astrocytes, microglia and neurons as the concerted activity of all these cell types is necessary for proper neural function. We are just beginning to understand the depth and mechanisms of intercellular information exchange that makes the NVU a functional unit. The knowledge of how the interdependent functions of the cell types constituting the NVU are affected by the process of aging can lead to the alleviation of age related impairments and a better quality of life.

Author Contributions: All authors participated in researching and discussing literature and writing parts of the manuscripts. L.C., I.A.K. and A.E.F. wrote the paper with input from all authors. All authors approved the final version.

Funding: A.E.F. is supported by the János Bolyai Research Fellowship of the Hungarian Academy of Sciences (BO/00023/17/8) and the New National Excellence Program of the Ministry of Human Capacities (UNKP-19-4-SZTE-43). Work of I.A.K. is supported by the NKFIH (grant numbers: K-116158, GINOP-2.3.2-15-2016-00020 and GINOP-2.3.2-15-2016-0034) and by the UEFISCDI PNCDI III—PCE 2016 Program (project number: PN-III-P4-ID-PCE-2016-0408, 188/2017). Work of I.W. is supported by the NKFIH FK-124114 and the UEFISCDI PN-III-P1-1.1-TE-2016-1352 projects.

Conflicts of Interest: The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

Abbreviations

BBB blood-brain barrier TJ tight-junction

CNS central nervous system

NVU neurovascular unit

CEC Cerebral endothelial cells

ZO-1 zonula occludens 1 protein

MCAO middle cerebral artery occlusion

Aβ amyloid-beta protein

Aβ1-42 amyloid-beta peptide containing amino acids 1-42

AD Alzheimer's disease

MMP matrix metalloproteinase

PD Parkinson's disease

HD Huntington's disease

References

- 1. Wilhelm, I.; Nyúl-Tóth, Á.; Kozma, M.; Farkas, A.E.; Krizbai, I.A. Role of pattern recognition receptors of the neurovascular unit in inflamm-aging. *Am. J. Physiol. Heart Circ. Physiol.* **2017**, *313*, H1000–H1012. [CrossRef] [PubMed]
- 2. Iadecola, C. The Neurovascular Unit Coming of Age: A Journey through Neurovascular Coupling in Health and Disease. *Neuron* **2017**, *96*, 17–42. [CrossRef] [PubMed]
- 3. Brightman, M.W.; Reese, T.S. Junctions between intimately apposed cell membranes in the vertebrate brain. *J. Cell Biol.* **1969**, *40*, 648–677. [CrossRef] [PubMed]
- 4. Oldendorf, W.H.; Cornford, M.E.; Brown, W.J. The large apparent work capability of the blood-brain barrier: A study of the mitochondrial content of capillary endothelial cells in brain and other tissues of the rat. *Ann. Neurol.* **1977**, *1*, 409–417. [CrossRef]
- 5. Nag, S. Morphology and molecular properties of cellular components of normal cerebral vessels. *Methods Mol. Med.* **2003**, *89*, 3–36. [CrossRef]
- 6. Hill, J.; Rom, S.; Ramirez, S.H.; Persidsky, Y. Emerging roles of pericytes in the regulation of the neurovascular unit in health and disease. *J. Neuroimmune Pharm.* **2014**, *9*, 591–605. [CrossRef]
- 7. Winkler, E.A.; Sengillo, J.D.; Bell, R.D.; Wang, J.; Zlokovic, B.V. Blood-spinal cord barrier pericyte reductions contribute to increased capillary permeability. *Br. J. Cereb. Blood Flow Metab.* **2012**, 32, 1841–1852. [CrossRef]
- 8. Thanabalasundaram, G.; Schneidewind, J.; Pieper, C.; Galla, H.J. The impact of pericytes on the blood-brain barrier integrity depends critically on the pericyte differentiation stage. *Int. J. Biochem. Cell Biol.* **2011**, *43*, 1284–1293. [CrossRef]
- 9. Hellström, M.; Gerhardt, H.; Kalén, M.; Li, X.; Eriksson, U.; Wolburg, H.; Betsholtz, C. Lack of pericytes leads to endothelial hyperplasia and abnormal vascular morphogenesis. *J. Cell Biol.* **2001**, *153*, 543–553. [CrossRef]
- 10. Armulik, A.; Genové, G.; Mäe, M.; Nisancioglu, M.H.; Wallgard, E.; Niaudet, C.; He, L.; Norlin, J.; Lindblom, P.; Strittmatter, K.; et al. Pericytes regulate the blood-brain barrier. *Nature* **2010**, *468*, 557–561. [CrossRef]
- 11. Abbott, N.J.; Rönnbäck, L.; Hansson, E. Astrocyte-endothelial interactions at the blood-brain barrier. *Nat. Rev. Neurosci.* **2006**, *7*, 41–53. [CrossRef] [PubMed]
- 12. Delaney, C.; Campbell, M. The blood brain barrier: Insights from development and ageing. *Tissue Barriers* **2017**, *5*, e1373897. [CrossRef] [PubMed]
- 13. Verheggen, I.C.M.; Van Boxtel, M.P.J.; Verhey, F.R.J.; Jansen, J.F.A.; Backes, W.H. Interaction between blood-brain barrier and glymphatic system in solute clearance. *Neurosci. Biobehav. Rev.* **2018**, *90*, 26–33. [CrossRef] [PubMed]
- 14. Iliff, J.J.; Wang, M.; Liao, Y.; Plogg, B.A.; Peng, W.; Gundersen, G.A.; Benveniste, H.; Vates, G.E.; Deane, R.; Goldman, S.A.; et al. A paravascular pathway facilitates CSF flow through the brain parenchyma and the clearance of interstitial solutes, including amyloid beta. *Sci. Transl. Med.* **2012**, *4*, 147ra111. [CrossRef] [PubMed]
- 15. Smith, A.J.; Verkman, A.S. The "glymphatic" mechanism for solute clearance in Alzheimer's disease: Game changer or unproven speculation? *FASEB J.* **2018**, 32, 543–551. [CrossRef] [PubMed]

- 16. Abbott, N.J.; Pizzo, M.E.; Preston, J.E.; Janigro, D.; Thorne, R.G. The role of brain barriers in fluid movement in the CNS: Is there a 'glymphatic' system? *Acta Neuropathol.* **2018**, *135*, 387–407. [CrossRef] [PubMed]
- 17. Zhang, Y.; Xu, K.; Liu, Y.; Erokwu, B.O.; Zhao, P.; Flask, C.A.; Ramos-Estebanez, C.; Farr, G.W.; LaManna, J.C.; Boron, W.F.; et al. Increased cerebral vascularization and decreased water exchange across the blood-brain barrier in aquaporin-4 knockout mice. *PLoS ONE* **2019**, *14*, e0218415. [CrossRef] [PubMed]
- 18. Kress, B.T.; Iliff, J.J.; Xia, M.; Wang, M.; Wei, H.S.; Zeppenfeld, D.; Xie, L.; Kang, H.; Xu, Q.; Liew, J.A.; et al. Impairment of paravascular clearance pathways in the aging brain. *Ann. Neurol.* **2014**, *76*, 845–861. [CrossRef]
- 19. Neuwelt, E.; Abbott, N.J.; Abrey, L.; Banks, W.A.; Blakley, B.; Davis, T.; Engelhardt, B.; Grammas, P.; Nedergaard, M.; Nutt, J.; et al. Strategies to advance translational research into brain barriers. *Lancet Neurol.* **2008**, *7*, 84–96. [CrossRef]
- 20. Bhowmick, S.; D'Mello, V.; Caruso, D.; Wallerstein, A.; Abdul-Muneer, P.M. Impairment of pericyte-endothelium crosstalk leads to blood-brain barrier dysfunction following traumatic brain injury. *Exp. Neurol.* **2019**, 317, 260–270. [CrossRef]
- 21. Chen, J.; Luo, Y.; Hui, H.; Cai, T.; Huang, H.; Yang, F.; Feng, J.; Zhang, J.; Yan, X. CD146 coordinates brain endothelial cell-pericyte communication for blood-brain barrier development. *Proc. Natl. Acad. Sci. USA* **2017**, *114*, E7622–E7631. [CrossRef] [PubMed]
- 22. Zagrean, A.M.; Hermann, D.M.; Opris, I.; Zagrean, L.; Popa-Wagner, A. Multicellular Crosstalk Between Exosomes and the Neurovascular Unit After Cerebral Ischemia. Therapeutic Implications. *Front. Neurosci.* **2018**, *12*, 811. [CrossRef] [PubMed]
- 23. Ramirez, S.H.; Andrews, A.M.; Paul, D.; Pachter, J.S. Extracellular vesicles: Mediators and biomarkers of pathology along CNS barriers. *Fluids Barriers CNS* **2018**, *15*, 19. [CrossRef] [PubMed]
- 24. András, I.E.; Toborek, M. Extracellular vesicles of the blood-brain barrier. *Tissue Barriers* **2016**, *4*, e1131804. [CrossRef]
- 25. Saeedi, S.; Israel, S.; Nagy, C.; Turecki, G. The emerging role of exosomes in mental disorders. *Transl. Psychiatry* **2019**, *9*, 122. [CrossRef]
- 26. Yuan, X.; Wu, Q.; Wang, P.; Jing, Y.; Yao, H.; Tang, Y.; Li, Z.; Zhang, H.; Xiu, R. Exosomes Derived from Pericytes Improve Microcirculation and Protect Blood-Spinal Cord Barrier After Spinal Cord Injury in Mice. *Front. Mol. Neurosci.* **2019**, *13*, 319. [CrossRef]
- 27. Liu, C.; Ge, H.M.; Liu, B.H.; Dong, R.; Shan, K.; Chen, X.; Yao, M.D.; Li, X.M.; Yao, J.; Zhou, R.M.; et al. Targeting pericyte-endothelial cell crosstalk by circular RNA-cPWWP2A inhibition aggravates diabetes-induced microvascular dysfunction. *Proc. Natl. Acad. Sci. USA* **2019**, *116*, 7455–7464. [CrossRef]
- 28. Brown, W.R.; Thore, C.R. Review: Cerebral microvascular pathology in ageing and neurodegeneration. *Neuropathol. Appl. Neurobiol.* **2011**, *37*, 56–74. [CrossRef]
- 29. Erusalimsky, J.D. Vascular endothelial senescence: From mechanisms to pathophysiology. *J. Appl. Physiol.* **2009**, *106*, 326–332. [CrossRef]
- 30. Sonntag, W.E.; Lynch, C.D.; Cooney, P.T.; Hutchins, P.M. Decreases in cerebral microvasculature with age are associated with the decline in growth hormone and insulin-like growth factor 1. *Endocrinology* **1997**, *138*, 3515–3520. [CrossRef]
- 31. Fan, Y.; Yang, X.; Tao, Y.; Lan, L.; Zheng, L.; Sun, J. Tight junction disruption of blood-brain barrier in white matter lesions in chronic hypertensive rats. *Neuroreport* **2015**, *26*, 1039–1043. [CrossRef] [PubMed]
- 32. Desjardins, M.; Berti, R.; Lefebvre, J.; Dubeau, S.; Lesage, F. Aging-related differences in cerebral capillary blood flow in anesthetized rats. *Neurobiol. Aging* **2014**, *35*, 1947–1955. [CrossRef] [PubMed]
- 33. Goodall, E.F.; Wang, C.; Simpson, J.E.; Baker, D.J.; Drew, D.R.; Heath, P.R.; Saffrey, M.J.; Romero, I.A.; Wharton, S.B. Age-associated changes in the blood-brain barrier: Comparative studies in human and mouse. *Neuropathol. Appl. Neurobiol.* **2018**, *44*, 328–340. [CrossRef] [PubMed]
- 34. Nitta, T.; Hata, M.; Gotoh, S.; Seo, Y.; Sasaki, H.; Hashimoto, N.; Furuse, M.; Tsukita, S. Size-selective loosening of the blood-brain barrier in claudin-5–deficient mice. *J. Cell Biol.* **2003**, *161*, 653–660. [CrossRef] [PubMed]
- 35. Castro Dias, M.; Coisne, C.; Lazarevic, I.; Baden, P.; Hata, M.; Iwamoto, N.; Francisco, D.M.F.; Vanlandewijck, M.; He, L.; Baier, F.A.; et al. Claudin-3-deficient C57BL/6J mice display intact brain barriers. *Sci. Rep.* **2019**, *9*, 203. [CrossRef]

- 36. Berndt, P.; Winkler, L.; Cording, J.; Breitkreuz-Korff, O.; Rex, A.; Dithmer, S.; Rausch, V.; Blasig, R.; Richter, M.; Sporbert, A.; et al. Tight junction proteins at the blood-brain barrier: Far more than claudin-5. *Cell Mol. Life Sci.* 2019, 76, 1987–2002. [CrossRef]
- 37. Dörfel, M.J.; Westphal, J.K.; Bellmann, C.; Krug, S.M.; Cording, J.; Mittag, S.; Tauber, R.; Fromm, M.; Blasig, I.E.; Huber, O. CK2-dependent phosphorylation of occludin regulates the interaction with ZO-proteins and tight junction integrity. *Cell Commun. Signal.* **2013**, *11*, 40. [CrossRef]
- 38. Raleigh, D.R.; Boe, D.M.; Yu, D.; Weber, C.R.; Marchiando, A.M.; Bradford, E.M.; Wang, Y.; Wu, L.; Schneeberger, E.E.; Shen, L.; et al. Occludin S408 phosphorylation regulates tight junction protein interactions and barrier function. *J. Cell Biol.* **2011**, *193*, 565–582. [CrossRef]
- 39. Runkle, E.A.; Sundstrom, J.M.; Runkle, K.B.; Liu, X.; Antonetti, D.A. Occludin localizes to centrosomes and modifies mitotic entry. *J. Biol. Chem.* **2011**, *286*, 30847–30858. [CrossRef]
- 40. Furuse, M.; Hirase, T.; Itoh, M.; Nagafuchi, A.; Yonemura, S.; Tsukita, S. Occludin: A novel integral membrane protein localizing at tight junctions. *J. Cell Biol.* **1993**, *12 Pt 2*, 1777–1788. [CrossRef]
- 41. Furuse, M.; Itoh, M.; Hirase, T.; Nagafuchi, A.; Yonemura, S.; Tsukita, S. Direct association of occludin with ZO-1 and its possible involvement in the localization of occludin at tight junctions. *J. Cell Biol.* **1994**, 127 Pt 1, 1617–1626. [CrossRef]
- 42. Haskins, J.; Gu, L.; Wittchen, E.S.; Hibbard, J.; Stevenson, B.R. ZO-3, a novel member of the MAGUK protein family found at the tight junction, interacts with ZO-1 and occludin. *J. Cell Biol.* **1998**, 141, 199–208. [CrossRef] [PubMed]
- 43. Itoh, M.; Furuse, M.; Morita, K.; Kubota, K.; Saitou, M.; Tsukita, S. Direct binding of three tight junction-associated MAGUKs, ZO-1, ZO-2, and ZO-3, with the COOH termini of claudins. *J. Cell Biol.* 1999, 147, 1351–1363. [CrossRef] [PubMed]
- 44. Castro, V.; Skowronska, M.; Lombardi, J.; He, J.; Seth, N.; Velichkovska, M.; Toborek, M. Occludin regulates glucose uptake and ATP production in pericytes by influencing AMP-activated protein kinase activity. *J. Cereb. Blood Flow Metab.* **2018**, *38*, 317–332. [CrossRef] [PubMed]
- 45. Bauer, H.; Stelzhammer, W.; Fuchs, R.; Weiger, T.M.; Danninger, C.; Probst, G.; Krizbai, I.A. Astrocytes and neurons express the tight junction-specific protein occludin in vitro. *Exp. Cell Res.* **1999**, 250, 434–438. [CrossRef] [PubMed]
- 46. Morgan, S.V.; Garwood, C.J.; Jennings, L.; Simpson, J.E.; Castelli, L.M.; Heath, P.R.; Mihaylov, S.R.; Vaquéz-Villaseñor, I.; Minshull, T.C.; Ince, P.G.; et al. Proteomic and cellular localisation studies suggest non-tight junction cytoplasmic and nuclear roles for occludin in astrocytes. *Eur. J. Neurosci.* 2018, 47, 1444–1456. [CrossRef] [PubMed]
- 47. Horng, S.; Therattil, A.; Moyon, S.; Gordon, A.; Kim, K.; Argaw, A.T.; Hara, Y.; Mariani, J.N.; Sawai, S.; Flodby, P.; et al. Astrocytic tight junctions control inflammatory CNS lesion pathogenesis. *J. Clin. Investig.* **2017**, 127, 3136–3151. [CrossRef] [PubMed]
- 48. Farrall, A.J.; Wardlaw, J.M. Blood-brain barrier: Ageing and microvascular disease-systematic review and meta-analysis. *Neurobiol. Aging* **2009**, *30*, 337–352. [CrossRef]
- 49. Baker, D.J.; Wijshake, T.; Tchkonia, T.; LeBrasseur, N.K.; Childs, B.G.; van de Sluis, B.; Kirkland, J.L.; van Deursen, J.M. Clearance of p16Ink4a-positive senescent cells delays ageing-associated disorders. *Nature* **2011**, 479, 232–236. [CrossRef]
- 50. Stamatovic, S.M.; Martinez-Revollar, G.; Hu, A.; Choi, J.; Keep, R.F.; Andjelkovic, A.V. Decline in Sirtuin-1 expression and activity plays a critical role in blood-brain barrier permeability in aging. *Neurobiol. Dis.* **2019**, *126*, 105–116. [CrossRef]
- 51. Bake, S.; Sohrabji, F. 17beta-estradiol differentially regulates blood-brain barrier permeability in young and aging female rats. *Endocrinology* **2004**, *145*, 5471–5475. [CrossRef] [PubMed]
- 52. Bake, S.; Friedman, J.A.; Sohrabji, F. Reproductive age-related changes in the blood brain barrier: Expression of IgG and tight junction proteins. *Microvasc. Res.* **2009**, *78*, 413–424. [CrossRef] [PubMed]
- 53. Wilson, A.C.; Clemente, L.; Liu, T.; Bowen, R.L.; Meethal, S.V.; Atwood, C.S. Reproductive hormones regulate the selective permeability of the blood-brain barrier. *Biochim. Biophys. Acta (BBA) Mol. Basis Dis.* **2008**, *1782*, 401–407. [CrossRef] [PubMed]
- 54. Bell, R.D.; Winkler, E.A.; Sagare, A.P.; Singh, I.; LaRue, B.; Deane, R.; Zlokovic, B.V. Pericytes control key neurovascular functions and neuronal phenotype in the adult brain and during brain aging. *Neuron* **2010**, *68*, 409–427. [CrossRef] [PubMed]

- 55. Mooradian, A.D.; Haas, M.J.; Chehade, J.M. Age-related changes in rat cerebral occludin and zonula occludens-1 (ZO-1). *Mech. Ageing Dev.* **2003**, *124*, 143–146. [CrossRef]
- 56. Osada, T.; Gu, Y.H.; Kanazawa, M.; Tsubota, Y.; Hawkins, B.T.; Spatz, M.; Milner, R.; del Zoppo, G.J. Interendothelial claudin-5 expression depends on cerebral endothelial cell-matrix adhesion by β (1)-integrins. *J. Cereb. Blood Flow Metab.* **2011**, *31*, 1972–1985. [CrossRef] [PubMed]
- 57. Roberts, J.; de Hoog, L.; Bix, G.J. Mice deficient in endothelial α5 integrin are profoundly resistant to experimental ischemic stroke. *J. Cereb. Blood Flow Metab.* **2017**, 37, 85–96. [CrossRef] [PubMed]
- 58. Montagne, A.; Barnes, S.R.; Sweeney, M.D.; Halliday, M.R.; Sagare, A.P.; Zhao, Z.; Toga, A.W.; Jacobs, R.E.; Liu, C.Y.; Amezcua, L.; et al. Blood-brain barrier breakdown in the aging human hippocampus. *Neuron* **2015**, *85*, 296–302. [CrossRef]
- 59. Yamazaki, Y.; Baker, D.J.; Tachibana, M.; Liu, C.C.; van Deursen, J.M.; Brott, T.G.; Bu, G.; Kanekiyo, T. Vascular Cell Senescence Contributes to Blood-Brain Barrier Breakdown. *Stroke* **2016**, *47*, 1068–1077. [CrossRef]
- 60. Schlingmann, B.; Overgaard, C.E.; Molina, S.A.; Lynn, K.S.; Mitchell, L.A.; Dorsainvil White, S.; Mattheyses, A.L.; Guidot, D.M.; Capaldo, C.T.; Koval, M. Regulation of claudin/zonula occludens-1 complexes by hetero-claudin interactions. *Nat. Commun.* **2016**, *7*, 12276. [CrossRef]
- 61. Lee, S.H.; Lee, J.H.; Lee, H.Y.; Min, K.J. Sirtuin signaling in cellular senescence and aging. *BMB Rep.* **2019**, 52, 24–34. [CrossRef] [PubMed]
- 62. Herskovits, A.Z.; Guarente, L. SIRT1 in neurodevelopment and brain senescence. *Neuron* **2014**, *81*, 471–483. [CrossRef] [PubMed]
- 63. Stirone, C.; Duckles, S.P.; Krause, D.N. Multiple forms of estrogen receptor-alpha in cerebral blood vessels: Regulation by estrogen. *Am. J. Physiol. Endocrinol Metab.* **2003**, *284*, E184–E192. [CrossRef] [PubMed]
- 64. Stewart, P.A.; Magliocco, M.; Hayakawa, K.; Farrell, C.L.; Del Maestro, R.F.; Girvin, J.; Kaufmann, J.C.; Vinters, H.V.; Gilbert, J. A quantitative analysis of blood-brain barrier ultrastructure in the aging human. *Microvasc. Res.* **1987**, *33*, 270–282. [CrossRef]
- 65. Borghesan, M.; O'Loghlen, A. Integrins in senescence and aging. Cell Cycle 2017, 16, 909–910. [CrossRef]
- 66. Labat-Robert, J. Cell-matrix interactions, alteration with aging and age associated diseases. A review. *Pathol. Biol.* **2001**, 49, 349–352. [CrossRef]
- 67. Wu, X.; Reddy, D.S. Integrins as receptor targets for neurological disorders. *Pharmacol. Ther.* **2012**, *134*, 68–81. [CrossRef]
- 68. Stamatovic, S.M.; Sladojevic, N.; Keep, R.F.; Andjelkovic, A.V. Relocalization of junctional adhesion molecule A during inflammatory stimulation of brain endothelial cells. *Mol. Cells Biol.* **2012**, 32, 3414–3427. [CrossRef]
- 69. Mochida, G.H.; Ganesh, V.S.; Felie, J.M.; Gleason, D.; Hill, R.S.; Clapham, K.R.; Rakiec, D.; Tan, W.H.; Akawi, N.; Al-Saffar, M.; et al. A homozygous mutation in the tight-junction protein JAM3 causes hemorrhagic destruction of the brain, subependymal calcification, and congenital cataracts. *Am. J. Hum. Genet.* **2010**, *87*, 882–889. [CrossRef]
- 70. Saitou, M.; Furuse, M.; Sasaki, H.; Schulzke, J.D.; Fromm, M.; Takano, H.; Noda, T.; Tsukita, S. Complex phenotype of mice lacking occludin, a component of tight junction strands. *Mol. Biol. Cell* **2000**, *11*, 4131–4142. [CrossRef]
- 71. Gonzalez-Lozano, M.A.; Klemmer, P.; Gebuis, T.; Hassan, C.; van Nierop, P.; van Kesteren, R.E.; Smit, A.B.; Li, K.W. Dynamics of the mouse brain cortical synaptic proteome during postnatal brain development. *Sci. Rep.* **2016**, *6*, 35456. [CrossRef] [PubMed]
- 72. Zussy, C.; Loustalot, F.; Junyent, F.; Gardoni, F.; Bories, C.; Valero, J.; Desarménien, M.G.; Bernex, F.; Henaff, D.; Bayo-Puxan, N.; et al. Coxsackievirus Adenovirus Receptor Loss Impairs Adult Neurogenesis, Synapse Content, and Hippocampus Plasticity. *J. Neurosci.* 2016, 36, 9558–9571. [CrossRef] [PubMed]
- 73. Lochhead, J.J.; McCaffrey, G.; Quigley, C.E.; Finch, J.; DeMarco, K.M.; Nametz, N.; Davis, T.P. Oxidative stress increases blood-brain barrier permeability and induces alterations in occludin during hypoxia-reoxygenation. *J. Cereb. Blood Flow Metab.* **2010**, *30*, 1625–1636. [CrossRef] [PubMed]
- 74. Raz, L.; Knoefel, J.; Bhaskar, K. The neuropathology and cerebrovascular mechanisms of dementia. *J. Cereb. Blood Flow Metab.* **2016**, *36*, 172–186. [CrossRef]
- 75. Sugawara, T.; Fujimura, M.; Noshita, N.; Kim, G.W.; Saito, A.; Hayashi, T.; Narasimhan, P.; Maier, C.M.; Chan, P.H. Neuronal death/survival signaling pathways in cerebral ischemia. *NeuroRx* **2004**, *1*, 17–25. [CrossRef]

- 76. Roy-O'Reilly, M.; McCullough, L.D. Age and Sex Are Critical Factors in Ischemic Stroke Pathology. *Endocrinology* **2018**, 159, 3120–3131. [CrossRef]
- 77. Lutski, M.; Zucker, I.; Shohat, T.; Tanne, D. Characteristics and Outcomes of Young Patients with First-Ever Ischemic Stroke Compared to Older Patients: The National Acute Stroke ISraeli Registry. *Front. Neurol.* **2017**, *8*, 421. [CrossRef]
- 78. Bauer, A.T.; Bürgers, H.F.; Rabie, T.; Marti, H.H. Matrix metalloproteinase-9 mediates hypoxia-induced vascular leakage in the brain via tight junction rearrangement. *J. Cereb. Blood Flow Metab.* **2010**, *30*, 837–848. [CrossRef]
- 79. Mark, K.S.; Davis, T.P. Cerebral microvascular changes in permeability and tight junctions induced by hypoxia-reoxygenation. *Am. J. Physiol. Heart Circ. Physiol.* **2002**, 282, H1485–H1494. [CrossRef]
- 80. Lischper, M.; Beuck, S.; Thanabalasundaram, G.; Pieper, C.; Galla, H.J. Metalloproteinase mediated occludin cleavage in the cerebral microcapillary endothelium under pathological conditions. *Brain Res.* **2010**, *1326*, 114–127. [CrossRef]
- 81. Krizbai, I.; Bauer, H.; Bresgen, N.; Eckl, P.; Farkas, A.; Szatmari, E.; Traweger, A.; Wejksza, K.; Bauer, H. Effect of oxidative stress on the junctional proteins of cultured cerebral endothelial cells. *Cell. Mol. Neurobiol.* **2005**, 25, 129–139. [CrossRef] [PubMed]
- 82. Koto, T.; Takubo, K.; Ishida, S.; Shinoda, H.; Inoue, M.; Tsubota, K.; Okada, Y.; Ikeda, E. Hypoxia disrupts the barrier function of neural blood vessels through changes in the expression of claudin-5 in endothelial cells. *Am. J. Pathol.* **2007**, *170*, 1389–1397. [CrossRef] [PubMed]
- 83. Schreibelt, G.; Kooij, G.; Reijerkerk, A.; van Doorn, R.; Gringhuis, S.I.; van der Pol, S.; Weksler, B.B.; Romero, I.A.; Couraud, P.O.; Piontek, J.; et al. Reactive oxygen species alter brain endothelial tight junction dynamics via RhoA, PI3 kinase, and PKB signaling. *Faseb. J.* **2007**, *21*, 3666–3676. [CrossRef] [PubMed]
- 84. Shen, F.; Jiang, L.; Han, F.; Degos, V.; Chen, S.; Su, H. Increased Inflammatory Response in Old Mice is Associated with More Severe Neuronal Injury at the Acute Stage of Ischemic Stroke. *Aging Dis.* **2019**, *10*, 12–22. [CrossRef] [PubMed]
- 85. Witt, K.A.; Mark, K.S.; Hom, S.; Davis, T.P. Effects of hypoxia-reoxygenation on rat blood-brain barrier permeability and tight junctional protein expression. *Am. J. Physiol. Heart Circ. Physiol.* **2003**, 285, H2820–H2831. [CrossRef] [PubMed]
- 86. Willis, C.L.; Meske, D.S.; Davis, T.P. Protein kinase C activation modulates reversible increase in cortical blood-brain barrier permeability and tight junction protein expression during hypoxia and posthypoxic reoxygenation. *J. Cereb. Blood Flow Metab.* **2010**, *30*, 1847–1859. [CrossRef]
- 87. Knowland, D.; Arac, A.; Sekiguchi, K.J.; Hsu, M.; Lutz, S.E.; Perrino, J.; Steinberg, G.K.; Barres, B.A.; Nimmerjahn, A.; Agalliu, D. Stepwise recruitment of transcellular and paracellular pathways underlies blood-brain barrier breakdown in stroke. *Neuron* **2014**, *82*, 603–617. [CrossRef]
- 88. Kaur, J.; Tuor, U.I.; Zhao, Z.; Barber, P.A. Quantitative MRI reveals the elderly ischemic brain is susceptible to increased early blood-brain barrier permeability following tissue plasminogen activator related to claudin 5 and occludin disassembly. *J. Cereb. Blood Flow Metab.* **2011**, *31*, 1874–1885. [CrossRef]
- 89. Bai, Y.; Xu, G.; Xu, M.; Li, Q.; Qin, X. Inhibition of Src phosphorylation reduces damage to the blood-brain barrier following transient focal cerebral ischemia in rats. *Int. J. Mol. Med.* **2014**, *34*, 1473–1482. [CrossRef]
- 90. Kago, T.; Takagi, N.; Date, I.; Takenaga, Y.; Takagi, K.; Takeo, S. Cerebral ischemia enhances tyrosine phosphorylation of occludin in brain capillaries. *Biochem. Biophys. Res. Commun.* **2006**, 339, 1197–1203. [CrossRef]
- 91. Takenaga, Y.; Takagi, N.; Murotomi, K.; Tanonaka, K.; Takeo, S. Inhibition of Src activity decreases tyrosine phosphorylation of occludin in brain capillaries and attenuates increase in permeability of the blood-brain barrier after transient focal cerebral ischemia. *J. Cereb. Blood Flow Metab.* **2009**, 29, 1099–1108. [CrossRef] [PubMed]
- 92. Yang, Y.; Estrada, E.Y.; Thompson, J.F.; Liu, W.; Rosenberg, G.A. Matrix metalloproteinase-mediated disruption of tight junction proteins in cerebral vessels is reversed by synthetic matrix metalloproteinase inhibitor in focal ischemia in rat. *J. Cereb. Blood Flow Metab.* **2007**, 27, 697–709. [CrossRef] [PubMed]
- 93. Yang, Y.; Rosenberg, G.A. MMP-mediated disruption of claudin-5 in the blood-brain barrier of rat brain after cerebral ischemia. *Methods Mol. Biol.* **2011**, 762, 333–345. [CrossRef]

- 94. Yang, Y.; Thompson, J.F.; Taheri, S.; Salayandia, V.M.; McAvoy, T.A.; Hill, J.W.; Estrada, E.Y.; Rosenberg, G.A. Early inhibition of MMP activity in ischemic rat brain promotes expression of tight junction proteins and angiogenesis during recovery. *J. Cereb. Blood Flow Metab.* **2013**, *33*, 1104–1114. [CrossRef] [PubMed]
- 95. Rom, S.; Zuluaga-Ramirez, V.; Dykstra, H.; Reichenbach, N.L.; Ramirez, S.H.; Persidsky, Y. Poly(ADP-ribose) polymerase-1 inhibition in brain endothelium protects the blood-brain barrier under physiologic and neuroinflammatory conditions. *J. Cereb. Blood Flow Metab.* **2015**, *35*, 28–36. [CrossRef] [PubMed]
- 96. Yang, Y.; Salayandia, V.M.; Thompson, J.F.; Yang, L.Y.; Estrada, E.Y. Attenuation of acute stroke injury in rat brain by minocycline promotes blood-brain barrier remodeling and alternative microglia/macrophage activation during recovery. *J. Neuroinflammation* **2015**, *12*, 26. [CrossRef] [PubMed]
- 97. Weinl, C.; Castaneda Vega, S.; Riehle, H.; Stritt, C.; Calaminus, C.; Wolburg, H.; Mauel, S.; Breithaupt, A.; Gruber, A.D.; Wasylyk, B.; et al. Endothelial depletion of murine SRF/MRTF provokes intracerebral hemorrhagic stroke. *Proc. Natl. Acad. Sci. USA* 2015, 112, 9914–9919. [CrossRef]
- 98. Wang, W.; Dentler, W.L.; Borchardt, R.T. VEGF increases BMEC monolayer permeability by affecting occludin expression and tight junction assembly. *Am. J. Physiol. Heart Circ. Physiol.* **2001**, 280, H434–H440. [CrossRef]
- 99. Argaw, A.T.; Asp, L.; Zhang, J.; Navrazhina, K.; Pham, T.; Mariani, J.N.; Mahase, S.; Dutta, D.J.; Seto, J.; Kramer, E.G.; et al. Astrocyte-derived VEGF-A drives blood-brain barrier disruption in CNS inflammatory disease. *J. Clin. Investig.* **2012**, 122, 2454–2468. [CrossRef]
- 100. Elahy, M.; Jackaman, C.; Mamo, J.C.; Lam, V.; Dhaliwal, S.S.; Giles, C.; Nelson, D.; Takechi, R. Blood-brain barrier dysfunction developed during normal aging is associated with inflammation and loss of tight junctions but not with leukocyte recruitment. *Immun. Ageing* **2015**, *12*, 2. [CrossRef]
- 101. Banks, W.A.; Farr, S.A.; Morley, J.E. Permeability of the blood-brain barrier to albumin and insulin in the young and aged SAMP8 mouse. *J. Gerontol. A Biol. Sci. Med. Sci.* **2000**, *55*, B601–B606. [CrossRef] [PubMed]
- 102. Engelhardt, S.; Patkar, S.; Ogunshola, O.O. Cell-specific blood-brain barrier regulation in health and disease: A focus on hypoxia. *Br. J. Pharm.* **2014**, *171*, 1210–1230. [CrossRef] [PubMed]
- 103. Liu, W.; Sood, R.; Chen, Q.; Sakoglu, U.; Hendren, J.; Cetin, O.; Miyake, M.; Liu, K.J. Normobaric hyperoxia inhibits NADPH oxidase-mediated matrix metalloproteinase-9 induction in cerebral microvessels in experimental stroke. *J. Neurochem.* 2008, 107, 1196–1205. [CrossRef] [PubMed]
- 104. Blasig, I.E.; Bellmann, C.; Cording, J.; del Vecchio, G.; Zwanziger, D.; Huber, O.; Haseloff, R.F. Occludin Protein Family: Oxidative Stress and Reducing Conditions. *Antioxid. Redox Signal.* **2011**, *15*, 1195–1219. [CrossRef] [PubMed]
- 105. Bellmann, C.; Schreivogel, S.; Günther, R.; Dabrowski, S.; Schümann, M.; Wolburg, H.; Blasig, I.E. Highly conserved cysteines are involved in the oligomerization of occludin-redox dependency of the second extracellular loop. *Antioxid. Redox Signal.* 2014, 20, 855–867. [CrossRef] [PubMed]
- 106. Erickson, M.A.; Banks, W.A. Neuroimmune Axes of the Blood-Brain Barriers and Blood-Brain Interfaces: Bases for Physiological Regulation, Disease States, and Pharmacological Interventions. *Pharm. Rev.* **2018**, *70*, 278–314. [CrossRef] [PubMed]
- 107. Erickson, M.A.; Banks, W.A. Age-Associated Changes in the Immune System and Blood-Brain Barrier Functions. *Int. J. Mol. Sci.* **2019**, *20*, 1632. [CrossRef]
- 108. Ransohoff, R.M.; Engelhardt, B. The anatomical and cellular basis of immune surveillance in the central nervous system. *Nat. Rev. Immunol* **2012**, *12*, 623–635. [CrossRef]
- 109. Ousman, S.S.; Kubes, P. Immune surveillance in the central nervous system. *Nat. Neurosci.* **2012**, *15*, 1096–1101. [CrossRef]
- 110. Varatharaj, A.; Galea, I. The blood-brain barrier in systemic inflammation. *Brain Behav. Immun.* **2017**, *60*, 1–12. [CrossRef]
- 111. Gemechu, J.M.; Bentivoglio, M. T Cell Recruitment in the Brain during Normal Aging. *Front. Cell. Neurosci.* **2012**, *6*, 38. [CrossRef] [PubMed]
- 112. Xu, Y.Z.; Nygård, M.; Kristensson, K.; Bentivoglio, M. Regulation of cytokine signaling and T-cell recruitment in the aging mouse brain in response to central inflammatory challenge. *Brain Behav. Immun.* **2010**, 24, 138–152. [CrossRef] [PubMed]
- 113. Stichel, C.C.; Luebbert, H. Inflammatory processes in the aging mouse brain: Participation of dendritic cells and T-cells. *Neurobiol. Aging* **2007**, *28*, 1507–1521. [CrossRef] [PubMed]

- 114. Ritzel, R.M.; Crapser, J.; Patel, A.R.; Verma, R.; Grenier, J.M.; Chauhan, A.; Jellison, E.R.; McCullough, L.D. Age-Associated Resident Memory CD8 T Cells in the Central Nervous System Are Primed to Potentiate Inflammation after Ischemic Brain Injury. *J. Immunol* 2016, 196, 3318–3330. [CrossRef]
- 115. Miguel-Hidalgo, J.J.; Nithuairisg, S.; Stockmeier, C.; Rajkowska, G. Distribution of ICAM-1 immunoreactivity during aging in the human orbitofrontal cortex. *Brain Behav. Immun.* **2007**, 21, 100–111. [CrossRef]
- 116. Dulken, B.W.; Buckley, M.T.; Navarro Negredo, P.; Saligrama, N.; Cayrol, R.; Leeman, D.S.; George, B.M.; Boutet, S.C.; Hebestreit, K.; Pluvinage, J.V.; et al. Single-cell analysis reveals T cell infiltration in old neurogenic niches. *Nature* **2019**, *571*, 205–210. [CrossRef]
- 117. Suidan, G.L.; Mcdole, J.R.; Chen, Y.; Pirko, I.; Johnson, A.J. Induction of blood brain barrier tight junction protein alterations by CD8 T cells. *PLoS ONE* **2008**, *3*, e3037. [CrossRef]
- 118. Erickson, M.A.; Banks, W.A. Blood-brain barrier dysfunction as a cause and consequence of Alzheimer's disease. *J. Cereb. Blood Flow Metab.* **2013**, *33*, 1500–1513. [CrossRef]
- 119. Yamazaki, Y.; Kanekiyo, T. Blood-Brain Barrier Dysfunction and the Pathogenesis of Alzheimer's Disease. *Int. J. Mol. Sci.* **2017**, *18*, 1965. [CrossRef]
- 120. Marco, S.; Skaper, S.D. Amyloid beta-peptide1–42 alters tight junction protein distribution and expression in brain microvessel endothelial cells. *Neurosci. Lett.* **2006**, *401*, 219–224. [CrossRef]
- 121. Janota, C.; Lemere, C.A.; Brito, M.A. Dissecting the Contribution of Vascular Alterations and Aging to Alzheimer's Disease. *Mol. Neurobiol.* **2016**, *53*, 3793–3811. [CrossRef] [PubMed]
- 122. Wan, W.; Chen, H.; Li, Y. The potential mechanisms of Aβ-receptor for advanced glycation end-products interaction disrupting tight junctions of the blood-brain barrier in Alzheimer's disease. *Int. J. Neurosci.* **2014**, *124*, 75–81. [CrossRef] [PubMed]
- 123. Rosenberg, G.A. Matrix metalloproteinases and their multiple roles in neurodegenerative diseases. *Lancet Neurol.* **2009**, *8*, 205–216. [CrossRef]
- 124. Bourasset, F.; Ouellet, M.; Tremblay, C.; Julien, C.; Do, T.M.; Oddo, S.; LaFerla, F.; Calon, F. Reduction of the cerebrovascular volume in a transgenic mouse model of Alzheimer's disease. *Neuropharmacology* **2009**, *56*, 808–813. [CrossRef] [PubMed]
- 125. Carrano, A.; Hoozemans, J.J.; van der Vies, S.M.; van Horssen, J.; de Vries, H.E.; Rozemuller, A.J. Neuroinflammation and blood-brain barrier changes in capillary amyloid angiopathy. *Neurodegener. Dis.* **2012**, *10*, 329–331. [CrossRef] [PubMed]
- 126. Carrano, A.; Hoozemans, J.J.; van der Vies, S.M.; Rozemuller, A.J.; van Horssen, J.; de Vries, H.E. Amyloid Beta induces oxidative stress-mediated blood-brain barrier changes in capillary amyloid angiopathy. *Antioxid. Redox Signal.* 2011, 15, 1167–1178. [CrossRef]
- 127. Bruban, J.; Glotin, A.L.; Dinet, V.; Chalour, N.; Sennlaub, F.; Jonet, L.; An, N.; Faussat, A.M.; Mascarelli, F. Amyloid-beta (1–42) alters structure and function of retinal pigmented epithelial cells. *Aging Cell* **2009**, *8*, 162–177. [CrossRef]
- 128. Kook, S.Y.; Hong, H.S.; Moon, M.; Ha, C.M.; Chang, S.; Mook-Jung, I. Aβ₁₋₄₂-RAGE interaction disrupts tight junctions of the blood-brain barrier via Ca²⁺-calcineurin signaling. *J. Neurosci.* **2012**, *32*, 8845–8854. [CrossRef]
- 129. Wan, W.; Cao, L.; Liu, L.; Zhang, C.; Kalionis, B.; Tai, X.; Li, Y.; Xia, S. Aβ (1–42) oligomer-induced leakage in an in vitro blood-brain barrier model is associated with up-regulation of RAGE and metalloproteinases, and down-regulation of tight junction scaffold proteins. *J. Neurochem.* **2015**, *134*, 382–393. [CrossRef]
- 130. Wan, W.B.; Cao, L.; Liu, L.M.; Kalionis, B.; Chen, C.; Tai, X.T.; Li, Y.M.; Xia, S.J. EGb761 provides a protective effect against Aβ1-42 oligomer-induced cell damage and blood-brain barrier disruption in an in vitro bEnd.3 endothelial model. *PLoS ONE* **2014**, *9*, e113126. [CrossRef]
- 131. Park, J.C.; Baik, S.H.; Han, S.H.; Cho, H.J.; Choi, H.; Kim, H.J.; Lee, W.; Kim, D.K.; Mook-Jung, I. Annexin A1 restores Aβ1-42-induced blood-brain barrier disruption through the inhibition of RhoA-ROCK signaling pathway. *Aging Cell* **2017**, *16*, 149–161. [CrossRef] [PubMed]
- 132. Mehrabadi, A.R.; Korolainen, M.A.; Odero, G.; Miller, D.W.; Kauppinen, T.M. Poly(ADP-ribose) polymerase-1 regulates microglia mediated decrease of endothelial tight junction integrity. *Neurochem. Int.* **2017**, *108*, 266–271. [CrossRef] [PubMed]
- 133. Chen, X.; Lan, X.; Roche, I.; Liu, R.; Geiger, J.D. Caffeine protects against MPTP-induced blood-brain barrier dysfunction in mouse striatum. *J. Neurochem.* 2008, 107, 1147–1157. [CrossRef] [PubMed]

- 134. Sarkar, S.; Chigurupati, S.; Raymick, J.; Mann, D.; Bowyer, J.F.; Schmitt, T.; Beger, R.D.; Hanig, J.P.; Schmued, L.C.; Paule, M.G. Neuroprotective effect of the chemical chaperone, trehalose in a chronic MPTP-induced Parkinson's disease mouse model. *Neurotoxicology* **2014**, *44*, 250–262. [CrossRef] [PubMed]
- 135. Carvey, P.M.; Zhao, C.H.; Hendey, B.; Lum, H.; Trachtenberg, J.; Desai, B.S.; Snyder, J.; Zhu, Y.G.; Ling, Z.D. 6-Hydroxydopamine-induced alterations in blood-brain barrier permeability. *Eur. J. Neurosci.* **2005**, 22, 1158–1168. [CrossRef]
- 136. Huang, L.; Deng, M.; He, Y.; Lu, S.; Ma, R.; Fang, Y. β-asarone and levodopa co-administration increase striatal dopamine level in 6-hydroxydopamine induced rats by modulating P-glycoprotein and tight junction proteins at the blood-brain barrier and promoting levodopa into the brain. *Clin. Exp. Pharm. Physiol.* **2016**, *43*, 634–643. [CrossRef]
- 137. Kuan, W.L.; Bennett, N.; He, X.; Skepper, J.N.; Martynyuk, N.; Wijeyekoon, R.; Moghe, P.V.; Williams-Gray, C.H.; Barker, R.A. α-Synuclein pre-formed fibrils impair tight junction protein expression without affecting cerebral endothelial cell function. *Exp. Neurol.* **2016**, *285 Pt A*, 72–81. [CrossRef]
- 138. Pienaar, I.S.; Lee, C.H.; Elson, J.L.; McGuinness, L.; Gentleman, S.M.; Kalaria, R.N.; Dexter, D.T. Deep-brain stimulation associates with improved microvascular integrity in the subthalamic nucleus in Parkinson's disease. *Neurobiol. Dis.* **2015**, *74*, 392–405. [CrossRef]
- 139. Gray, M.T.; Woulfe, J.M. Striatal blood-brain barrier permeability in Parkinson's disease. *J. Cereb. Blood Flow Metab.* **2015**, *35*, 747–750. [CrossRef]
- 140. Drouin-Ouellet, J.; Sawiak, S.J.; Cisbani, G.; Lagacé, M.; Kuan, W.L.; Saint-Pierre, M.; Dury, R.J.; Alata, W.; St-Amour, I.; Mason, S.L.; et al. Cerebrovascular and blood-brain barrier impairments in Huntington's disease: Potential implications for its pathophysiology. *Ann. Neurol.* **2015**, *78*, 160–177. [CrossRef]
- 141. Lim, R.G.; Quan, C.; Reyes-Ortiz, A.M.; Lutz, S.E.; Kedaigle, A.J.; Gipson, T.A.; Wu, J.; Vatine, G.D.; Stocksdale, J.; Casale, M.S.; et al. Huntington's Disease iPSC-Derived Brain Microvascular Endothelial Cells Reveal WNT-Mediated Angiogenic and Blood-Brain Barrier Deficits. *Cell Rep.* **2017**, *19*, 1365–1377. [CrossRef] [PubMed]
- 142. Greene, C.; Kealy, J.; Humphries, M.M.; Gong, Y.; Hou, J.; Hudson, N.; Cassidy, L.M.; Martiniano, R.; Shashi, V.; Hooper, S.R.; et al. Dose-dependent expression of claudin-5 is a modifying factor in schizophrenia. *Mol. Psychiatry* **2018**, 23, 2156–2166. [CrossRef] [PubMed]
- 143. Takechi, R.; Galloway, S.; Pallebage-Gamarallage, M.M.; Mamo, J.C. Chylomicron amyloid-beta in the aetiology of Alzheimer's disease. *Atheroscler. Suppl.* 2008, *9*, 19–25. [CrossRef]
- 144. Poduslo, J.F.; Curran, G.L.; Wengenack, T.M.; Malester, B.; Duff, K. Permeability of proteins at the blood-brain barrier in the normal adult mouse and double transgenic mouse model of Alzheimer's disease. *Neurobiol. Dis.* **2001**, *8*, 555–567. [CrossRef] [PubMed]
- 145. Cheng, Z.; Zhang, J.; Liu, H.; Li, Y.; Zhao, Y.; Yang, E. Central nervous system penetration for small molecule therapeutic agents does not increase in multiple sclerosis- and Alzheimer's disease-related animal models despite reported blood-brain barrier disruption. *Drug Metab. Dispos.* 2010, 38, 1355–1361. [CrossRef] [PubMed]
- 146. Mehta, D.C.; Short, J.L.; Nicolazzo, J.A. Altered brain uptake of therapeutics in a triple transgenic mouse model of Alzheimer's disease. *Pharm. Res.* **2013**, *30*, 2868–2879. [CrossRef] [PubMed]
- 147. Daneman, R.; Zhou, L.; Kebede, A.A.; Barres, B.A. Pericytes are required for blood-brain barrier integrity during embryogenesis. *Nature* **2010**, *468*, 562–566. [CrossRef]
- 148. Sagare, A.P.; Bell, R.D.; Zhao, Z.; Ma, Q.; Winkler, E.A.; Ramanathan, A.; Zlokovic, B.V. Pericyte loss influences Alzheimer-like neurodegeneration in mice. *Nat. Commun.* **2013**, *4*, 2932. [CrossRef]
- 149. Sengillo, J.D.; Winkler, E.A.; Walker, C.T.; Sullivan, J.S.; Johnson, M.; Zlokovic, B.V. Deficiency in mural vascular cells coincides with blood-brain barrier disruption in Alzheimer's disease. *Brain Pathol.* **2013**, 23, 303–310. [CrossRef]
- 150. Tai, L.M.; Thomas, R.; Marottoli, F.M.; Koster, K.P.; Kanekiyo, T.; Morris, A.W.; Bu, G. The role of APOE in cerebrovascular dysfunction. *Acta Neuropathol.* **2016**, *131*, 709–723. [CrossRef]
- 151. Brandon, J.A.; Farmer, B.C.; Williams, H.C.; Johnson, L.A. Apoe and Alzheimer's Disease: Neuroimaging of Metabolic and Cerebrovascular Dysfunction. *Front. Aging Neurosci.* **2018**, *10*, 180. [CrossRef] [PubMed]
- 152. Bell, R.D.; Winkler, E.A.; Singh, I.; Sagare, A.P.; Deane, R.; Wu, Z.; Holtzman, D.M.; Betsholtz, C.; Armulik, A.; Sallstrom, J.; et al. Apolipoprotein E controls cerebrovascular integrity via cyclophilin A. *Nature* 2012, 485, 512–516. [CrossRef] [PubMed]

- 153. Daneman, R.; Prat, A. The blood-brain barrier. *Cold Spring Harb. Perspect. Biol.* **2015**, 7, a020412. [CrossRef] [PubMed]
- 154. Zenaro, E.; Piacentino, G.; Constantin, G. The blood-brain barrier in Alzheimer's disease. *Neurobiol. Dis.* **2017**, *107*, 41–56. [CrossRef]
- 155. Sweeney, M.D.; Sagare, A.P.; Zlokovic, B.V. Blood-brain barrier breakdown in Alzheimer disease and other neurodegenerative disorders. *Nat. Rev. Neurol.* **2018**, *14*, 133–150. [CrossRef]
- 156. Greene, C.; Hanley, N.; Campbell, M. Claudin-5: Gatekeeper of neurological function. *Fluids Barriers CNS* **2019**, *16*, 3. [CrossRef]
- 157. Lee, H.; Pienaar, I.S. Disruption of the blood-brain barrier in Parkinson's disease: Curse or route to a cure? *Front. Biosci. (Landmark Ed.)* **2014**, *19*, 272–280. [CrossRef]
- 158. Pan, Y.; Nicolazzo, J.A. Impact of aging, Alzheimer's disease and Parkinson's disease on the blood-brain barrier transport of therapeutics. *Adv. Drug Deliv. Rev.* **2018**, *135*, 62–74. [CrossRef]
- 159. Farkas, E.; De Jong, G.I.; de Vos, R.A.; Jansen Steur, E.N.; Luiten, P.G. Pathological features of cerebral cortical capillaries are doubled in Alzheimer's disease and Parkinson's disease. *Acta Neuropathol.* **2000**, 100, 395–402. [CrossRef]
- 160. Guan, J.; Pavlovic, D.; Dalkie, N.; Waldvogel, H.J.; O'Carroll, S.J.; Green, C.R.; Nicholson, L.F. Vascular degeneration in Parkinson's disease. *Brain Pathol.* **2013**, *23*, 154–164. [CrossRef]
- 161. Hill, K.K.; Campbell, M.C.; McNeely, M.E.; Karimi, M.; Ushe, M.; Tabbal, S.D.; Hershey, T.; Flores, H.P.; Hartlein, J.M.; Lugar, H.M.; et al. Cerebral blood flow responses to dorsal and ventral STN DBS correlate with gait and balance responses in Parkinson's disease. *Exp. Neurol.* 2013, 241, 105–112. [CrossRef] [PubMed]
- 162. Horvath, S.; Langfelder, P.; Kwak, S.; Aaronson, J.; Rosinski, J.; Vogt, T.F.; Eszes, M.; Faull, R.L.; Curtis, M.A.; Waldvogel, H.J.; et al. Huntington's disease accelerates epigenetic aging of human brain and disrupts DNA methylation levels. *Aging (Albany Ny)* **2016**, *8*, 1485–1512. [CrossRef] [PubMed]
- 163. Okusaga, O.O. Accelerated aging in schizophrenia patients: The potential role of oxidative stress. *Aging Dis.* **2014**, *5*, 256–262. [CrossRef] [PubMed]



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