Contents lists available at ScienceDirect

Heliyon

journal homepage: www.cell.com/heliyon

Research article

CelPress

A novel lnc-LAMC2-1:1 SNP promotes colon adenocarcinoma progression by targeting miR-216a-3p/HMGB3



Fulong Ji ^{a,1}, Zhiwei Yao ^{a,1}, Chunxiang Liu ^{a,1}, Siqi Fu ^a, Bingbing Ren ^a, Yong Liu ^a, Lushun Ma ^a, Jianming Wei ^{b,**}, Daqing Sun ^{a,*}

^a Department of Pediatric Surgery, Tianjin Medical University General Hospital, Tianjin, 300052, China
^b Department of General Surgery, Tianjin Medical University General Hospital, Tianjin, 300052, China

ARTICLE INFO

Keywords: Colon adenocarcinoma Inc-LAMC2-1:1 SNP miR-216a-3p HMGB3 Progression

ABSTRACT

Single nucleotide polymorphisms (SNPs) was associated with altering the secondary structure of long non-coding RNA (IncRNA). Increasing reports showed that *Inc-LAMC2-1:1 SNP* played an important role in cancer development and invasion. This study is to elucidate the molecular function of *Inc-LAMC2-1:1 SNP rs2147578* promoting tumor progression in colon adenocarcinoma (COAD). In this study, we found that the *Inc-LAMC2-1:1 SNP rs2147578* promoted colon cancer migration, invasion, and proliferation. Interestingly, *Inc-LAMC2-1:1 SNP rs2147578* promoted colon cancer migration, invasion, and proliferation. Interestingly, *Inc-LAMC2-1:1 SNP rs2147578* positively regulated *HMGB3* expression via *miR-216a-3p* in colon cancer cells. Functional enrichment analysis showed that targeting genes of *miR-216a-3p* were enriched in regulating the pluripotency of stem cells, MAPK signaling pathway, TNF signaling pathway, neurotrophin signaling pathway, relaxin signaling pathway, and FoxO signaling pathway. Tumor Immune Estimation Resource (TIMER) database revealed that there was a significantly positive correlation between *HMGB3* expression and the infiltration of CD8⁺ T cells, B cells, neutrophils, macrophages, and CD4⁺ T cells. Finally, *HMGB3* overexpression was validated in external data. In conclusions, *Inc-LAMC2-1:1 SNP rs2147578* was involved in promoting COAD progression by targeting *miR-216a-3p/HMGB3*, and this study will provide a novel molecular target for COAD.

1. Introduction

Colorectal cancer (CRC) is a frequent malignancy that ranks third in terms of incidence, but second in terms of mortality in 2021 [1]. Colon adenocarcinoma (COAD) is a common malignant tumor with the highest incidence and the leading causes of cancer death in CRC. Although surgery, radiotherapy, chemotherapy, and immunotherapy have developed in recent years, the overall survival rate of COAD patients is still unsatisfactory [2]. This needs to explore effective therapies and identify novel prognostic biomarkers for COAD patients. Over 100 CRC risk loci have been identified by genome-wide association studies (GWASs) [3]. Single nucleotide polymorphisms (SNPs) usually consist of intronic and exonic sites. However, most of the susceptibility loci are located in noncoding regions [4]. Long non-coding RNA (lncRNA) is an RNA transcript with a

length of more than 200 nucleotides and no protein coding ability [5]. LncRNA SNP plays an important role in tumor growth [6]. Increasing recent evidence has revealed that SNPs can influence susceptibility to disease by changing the expression of lncRNAs [7]. Previous studies have proved that *lnc-LAMC2-1:1 SNP rs2147578* affected tumor invasion and progression [8, 9, 10]. However, the role and molecular mechanism of *lnc-LAMC2-1:1 SNP rs2147578* in COAD have not been elucidated.

In this study, we investigated the role of *lnc-LAMC2-1:1 SNP rs2147578* promoting the proliferation, migration, and invasion in COAD. Our results showed that *lnc-LAMC2-1:1 SNP rs2147578* could positively regulated high-mobility group-box 3 (HMGB3) expression via binding *miR-216a-3p* in COAD. Moreover, *miR-216a-3p*-mediated regulatory network was constructed. *HMGB3* overexpression was validated in external data. Besides, we explored the correlation of the expression of

** Corresponding author.

https://doi.org/10.1016/j.heliyon.2022.e12342

Received 7 March 2022; Received in revised form 1 September 2022; Accepted 6 December 2022



^{*} Corresponding author.

E-mail addresses: weijianming@tmu.edu.cn (J. Wei), sdqchris2019@tmu.edu.cn (D. Sun).

¹ Authors contributed equally.

^{2405-8440/© 2022} The Author(s). Published by Elsevier Ltd. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

HMGB3 with tumor immune infiltrating cells. This study revealed a novel mechanism of *lnc-LAMC2-1:1 SNP* promoting tumor behavior and suggested a potential new molecular target in COAD.

2. Materials and methods

2.1. Network construction of miR-216a-3p-mRNAs

To ananlyzed the mechanism of *miR-216a-3p*, firstly, the target genes of miR-216a-3p were predicted from the miRDB, miRTarBase and TargetScan databases using the Perl (5.26.3.0000) software program. Then, the regulatory network of miRNA-mRNA and hub genes was constructed using Cytoscape software 3.6.0 [11].

2.2. Functional enrichment analysis of target genes of miR-216a-3p

Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway analysis were performed using R (v3.6.3) and Perl (5.26.3.0000) software. The packages "colorspace" "dose" biocLite ("DOSE") "clusterProfiler" and "pathview" were installed. The selection criterion for significant GO and KEGG pathway terms was P < 0.05.

2.3. Cell culture and transfection

The SW480 cell line was obtained from COSMOBIO Company in China and was grown in DMEM supplemented with 10% FBS (GIBCO) and 1% antibiotics (50 U/mL penicillin and 50 μ g/mL streptomycin) in a humidified atmosphere of 5% CO2 at 37 °C. For transfection assays, cells were seeded in 96-well plates and simultaneously transfected with the psiCHECK-2 vector and miRNA mimics using Lipofectamine 3000 (Invitrogen). All experiments were independently performed in triplicates.

2.4. Real-time qRT-PCR analysis

Total RNA was extracted using TRIzol reagent (Invitrogen, USA) and transcribed into cDNA using M-MLV reverse transcriptase (TaKaRa Bio, Japan) following the manufacturer's instructions. The primer sequences were as follows:

IncRNA LAMC2-wt: 5'-CATAGTCCCTCAGTGTGGGTCATTTTCATTAG-3' IncRNA LAMC2 SNP: 5'-CATAGTCCCTCACTGTGGGTCATTTTCATTAG-3' β-actin-S: 5'-CGTGACATTAAGGAGAAGCTG-3' β-actin-AS: 5'-CTAGAAGCATTTGCGGTGGAC-3' HMGB3-S: 5'-ATTCGGAATTCCGTATCTGGCCTTTTGAC-3' HMGB3-AS: 5'-CGGTTACTCGGCTTACGCTTGGACTG-3'

2.5. Luciferase activity assay

StarBase online predicted the potential targets of *miR-216a-3p*. The wild-type (wt) or mutant (MUT) *HMGB3*-binding *miR-216a-3p* was subcloned into the pGL3 Basic vector (Promega). A total of SW480 cells were seeded in 24-well plates for 48 h. Mimics or inhibitors of *miR-216a-3p* (RiboBio, Guangzhou, China) were cotransfected with 10 μ g pLUC-wt-HMGB3 or p LUC-MUT- HMGB3. The same procedure was used to assess the combined effect of *HMGB3* and *miR-216a-3p*. Luciferase reporter system (Promega) was used to assess the luciferase activity in SW480 cells.

2.6. TIMER database

TIMER database could provide the association between immune cells infiltrating and clinical factors, including gene expression, clinical outcomes, somatic mutations, and somatic copy number alterations [12]. We

comprehensively investigated molecular characterization of tumor-immune interactions using TIMER database.

2.7. Validation of HMGB3 expression in The Human Protein Atlas database

The Human Protein Atlas (HPA; http://www.proteinatlas.org/), GSE8671, GSE9348, online databases TCGA, GEPIA and The Human Protein Atlas Database were used to validate the expression of *HMGB3* in COAD samples [13].

2.8. Statistical analysis

All data are presented as the mean \pm standard deviation (SD). Student's t-test and one-way analysis of variance were carried out to evaluate significant differences. P values $<\!0.05$ were considered to indicate statistical significance.

3. Results

3.1. Lnc-LAMC2-1:1 SNP rs2147578 could promote the progression and invasion in COAD

Accumulating evidences showed that *lnc-LAMC2-1:1 SNP rs2147578* increased the risk of ovarian cancer [9]. To evaluate the role of *lnc-LAMC2-1:1 SNP rs2147578*, transwell and 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2-H-tetrazolium bromide (MTT) assays were performed. Here, we found that *lnc-LAMC2-1:1 SNP rs2147578* significantly promoted proliferation in SW480 cells (Figure 1A). Moreover, the result showed that *lnc-LAMC2-1:1 SNP rs2147578* markedly increased the migration and invasion in SW480 cells (Figure 1B and 1C). These results showed that *lnc-LAMC2-1:1 SNP rs2147578* could promote the tumor development in COAD.

3.2. The lnc-LAMC2-1:1 SNP is a sponge of miR-216a-3p

A recent study revealed that IncRNA SNPSNP rs140618127 contained a binding site for miR-539-5p promoting cell proliferation and tumor progression [14]. Previous study showed that Inc-LAMC2-1:1 SNP rs2147578 can interact with hsa-miR-128-3p, hsa-miR-216a-3p, and hsa-miR-368-3p [10]. In this study, we revealed that the expression of miR-216a-3p was the most significantly downregulated. Moreover, miR-216a-3p was significantly in the lnc-LAMC2-1:1 SNP rs2147578 group compared to the *lnc-LAMC2-1:1-wt* group (Figure 2A). To explore whether Inc-LAMC2-1:1 SNP rs2147578 bind with miR-216a-3p, StarBase database online was used to predict the binding site between Inc-LAMC2-1:1 SNP rs2147578 and miR-216a-3p. To validate this relationship, Inc-LAMC2-1:1-wt and Inc-LAMC2-1:1-SNP rs2147578 were constructed and transfected into SW480 cells. As shown in Figure 2B, overexpression of miR-216a-3p led to obvious loss of luciferase activity in the Inc-LAMC2-1:1-SNP rs2147578 group in the two cell lines, while it did not affect those in the *lnc-LAMC2-1:1-wt* group.

3.3. Regulatory network of miR-216a-3p

miRNAs post-transcriptionally suppress the target mRNA expression, mostly through interaction with the 3' UTR [15]. To find the target gene of *miR-216a-3p*, twenty-six target genes of *miR-216a-3p* were predicted using miRDB, miRTarBase, and TargetScan by Perl software. Investigating the interaction among genes, we used Cytoscape [16] (version 3.6.1) to construct the regulatory network, as shown in Figure 3A. The PPI network was constructed using the STRING database [17] (http://stri ng-db.org) with a total of 26 target genes of *miR-216a-3p*. After removing the isolated and partially connected nodes, a complex network of target genes was identified as the top 10 hub genes using the cytoHubba [18] (Figure 3B). a



Figure 1. The *lnc-LAMC2-1:1 SNP* is increased and promotes colon cancer progression. Knockdown of *lnc-LAMC2-1:1 SNP* inhibits (a) proliferation, (b) migration, and (C) invasion in COAD. *P < 0.05, **P < 0.01 compared with the indicated control group.

F. Ji et al.



Figure 2. The *lnc-LAMC2-1:1 SNP* is a sponge of *miR-216a-3p*. Bar graphs of (a) and (b) show the relative luciferase activity of vectors containing SW480. A dualluciferase reporter assay was performed, and the co-transfection of *lnc-LAMC2-1:1 SNP* and *miR-216a-3p* reduced the luciferase activity.

3.4. Functional enrichment analysis

In the present study, GO functional enrichment and KEGG pathway analyses were performed to explore the molecular function of the target genes of *miR-216a-3p*. Functional enrichment analysis with a P-value of 0.05 was obtained. The results were shown in Figure 4A. We found that the phospholipid transporter activity of the target gene was the most abundant. KEGG pathways analysis were mainly enriched in pathways regulating the pluripotency of stem cells, MAPK signaling pathway, TNF signaling pathway, neurotrophin signaling pathway, relaxin signaling pathway, and FoxO signaling pathway. These results were shown in Figure 4B.

3.5. The Inc-LAMC2-1:1 SNP positively regulates HMGB3 by sponging miR-216a-3p

High-mobility group box 3 (HMGB3), a member of the high-mobility group box (HMGB) family, was reported to be over-expressed in cancers [19, 20]. StarBase online predicted the targets of *miR-216a-3p*, and the results showed *HMGB3* had potential complementary sequences of *miR-216a-3p* (Figure 5A). qRT-PCR assay displayed that *HMGB3* was negatively regulated by *miR-216a-3p* (Figure 5B). In addition, this result showed that the expression of *HMGB3* level in SW480 cells was positively regulated by *lnc-LAMC2-1:1 SNP rs2147578* (Figure 5C). To validate the

relationship of *miR-216a-3p* with *HMGB3*, the luciferase reporter assay showed that *miR-216a-3p* upregulation significantly led to reduction in luciferase activity in the *HMGB3-WT* group, while its efficacy was lost when the binding sites were mutated (Figure 5D).

3.6. Correlation of the expression of HMGB3 with immune cell infiltration

Tumor immune cell was closely associated with COAD progression. In this study, we explored the correlation between the expression of *HMGB3* and immune cell infiltration using the TIMER database. There was a significantly positive correlation between *HMGB3* expression and the infiltration of CD8⁺ T cells (Cor = 0.12, p = 1.57e - 2; Figure 6A), CD4⁺ T cells (Cor = 0.162, p = 1.1e - 3; Figure 6B), macrophages (Cor = 0.185, p = 1.79e - 4; Figure 6C), neutrophils (Cor = 0.122, p = 1.47e - 2; Figure 6D), and B cells (Cor = 0.122, p = 1.42e - 2; Figure 6F). However, dendritic cells did not show significant expression of *HMGB3* (Cor = 0.088, p = 7.71e - 2; Figure 6E).

3.7. Validation of HMGB3 expression in external data

To test *HMGB3* expression in the present study, a total of 64 samples in GSE8671 including 32 COAD samples and 32 normal samples were screened out. The results revealed that COAD patients had markedly high expression of *HMGB3* (Figure 7A). Moreover, we found



Figure 3. The regulatory network of *miR-126-3p* and PPI (a) The network of *miR-126-3p* target genes. Red indicates *miR-126-3p*, and green indicates genes. (b) protein-protein interaction of target genes.



Figure 4. GO and KEGG functional enrichment analysis (a) GO enrichment significance items (b) Gene ratio and KEGG pathway items. Red indicates upregulated genes, and green indicates downregulated genes.



Figure 5. The *lnc-LAMC2-1:1 SNP* positively regulates *HMGB3* by sponging *miR-126-3p* (a) mRNA expression of *HMGB3* in SW480 cells. Cells were transfected with mimics of NC, *miR-126-3p* mimics, ASO-NC, and ASO-miR-126-3p. (b) mRNA expression of *HMGB3* in SW480 cells. Cells were transfected with pcDNA3.1, *lnc-LAMC2-1:1 SNP*, and *lnc-LAMC2-1:1-wt*. (c) Complementary sequences of *HMGB3* and *miR-126-3p* in the StarBase database. (d) A dual-luciferase reporter assay was performed, and the co-transfection of *miR-126-3p* and *HMGB3* reduced the luciferase activity.

that *HMGB3* expression was upregulated in tumor samples compared to that in normal samples of GSE9348 (Figure 7B). This result was also found that *HMGB3* expression was validated in 286 tumor samples compared to 41 normal samples in online TCGA database (Figure 7D). We used the GEPIA online database to validate the expression of *HMGB3*, and we found that the results were consistent with those mentioned above (Figure 7C). Immunohistochemistry showed that *HMGB3* was performed based on the HPA database, and the results

revealed that *HMGB3* was overexpressed in tumor samples than in normal samples (Figure 7E-F).

4. Discussion

LncRNAs are regulators of transcription and are increasingly recognized to play a role in cancer biology [21]. For example, Zhong et al found that LINC00636 also promotes lymph node metastasis and carcinogenesis



Figure 6. Correlation of *HMGB3* expression with immune cell infiltration. Association of the expression of (a) $CD8^+$ T cells, (b) $CD4^+$ T cells, (c) macrophages, (d) neutrophils, (e) dendritic cells and (f) B cells with *HMGB3* expression were shown in this study.



Figure 7. External data validation of HMGB3 expression. HMGB3 expression was validated in (a) GSE8671, (b) GSE9348, and (c) GEPIA database and (d) TCGA database. (e) and (f) show the expression of HMGB3 in COAD tissues on immunohistochemistry staining.

by regulating NM23 [21]. Meanwhile, the *lnc-LAMC2-1:1 SNP rs2147578* plays essential roles in many diseases [9, 22, 23]. To the best of our knowledge, no study has investigated the mechanism of *lnc-LAMC2-1:1 SNP rs2147578* in colorectal cancer. In this study, we found that *lnc-LAMC2-1:1 SNP rs2147578* could promote colon cancer proliferation, migration, and invasion. This study aimed to analyze the mechanism of *lnc-LAMC2-1:1 SNP rs2147578* in COAD and explore the novel ceRNA network of *lnc-LAMC2-1:1 SNP rs2147578/miR-216a-3p/HMGB3*.

There are abundant miRNA binding sites in LncRNA molecules, which act as sponges, counteracting the effect of miRNA on its target genes and increasing the expression level of target genes [24]. We studied the association of miRNAs in COAD affected by *lnc-LAMC2-1:1 SNP rs2147578*. A previous study reported that *lnc-LAMC2-1:1 SNP rs2147578* may affect *miR-128-3p*, *hsa-miR-216a-3p*, and *hsa-miR-368-3p* binding and confer a high risk of colorectal cancer [10]. Hence, we selected these three miRNAs for our study. We used a luciferase reporter assay to validate the target association between *lnc-LAMC2-1:1 SNP rs2147578* and *miR-216a-3p*. The results showed that *lnc-LAMC2-1:1 SNP rs2147578* significantly repressed the expression of *miR-216a-3p* compared to that in the *lnc-LAMC2-1:1-wt* group. *MiR-216a-3p* has been reported as a tumor suppressor in colon cancer [24], gastric cancer [25, 26], breast cancer [27] and pancreatic ductal adenocarcinoma [28]. In order to verify the

biological relationship between *lnc-LAMC2-1:1 SNP rs2147578* and *miR-216a-3p* in COAD, we carried out functional studies. The activity, invasion and proliferation of SW480 cells transfected with *lnc-LAMC2-1:1SNP rs2147578* were significantly enhanced. Therefore, *lnc-LAMC2-1:1 SNP rs2147578* acts as a sponge to regulate COAD by combining with *miR-216a-3p*.

Previous studies have suggested that HMGB3 is an oncogene in multiple cancers, including breast cancer [20, 29, 30], gastric cancer [31, 32], colon carcinoma [19], esophageal squamous cell carcinoma [33], ovarian cancer [34], non-small cell lung cancer [35, 36], liver cancer [37, 38, 39], thyroid carcinoma [40] and prostate cancer [41]. We predicted that HMGB3 had miR-216a-3p-binding sites in the StarBase database. In this study, we found that HMGB3 was downregulated in the miR-216a-3p mimics group, and the luciferase activity showed that HMGB3 could bind to miR-216a-3p. In our experiment, the RT-qPCR assay showed that Inc-LAMC2-1:1 SNP rs2147578 could promote HMGB3 expression, suggesting the potential ceRNA of Inc-LAMC2-1:1 SNP rs2147578/miR-216a-3p/HMGB3. We also investigated that HMGB3 was highly expressed in COAD, which is also in agreement with the data of GSE8671, GSE9348, TCGA database, the GEPIA database and the HPA database online, indicating that high expression of HMGB3 might contribute to COAD development. To sum up, the role of Inc-LAMC2-1:1 SNP rs2147578 in COAD may be related to the expression of miR-216a-3p/HMGB3.

Moreover, previous studies have suggested that *HMGB3* is associated with immune regulation [42, 43, 44, 45]. In the current study, we also found that *HMGB3* affected the expression of $CD8^+$ T cells, B cells, neutrophils, macrophages and $CD4^+$ T cells, which is expected to be explored in future studies.

5. Conclusion

In conclusion, our study highlighted that *lnc-LAMC2-1:1SNP rs2147578* promoted the progression of COAD by regulating *miR-216a-3p/HMGB3* axis as a ceRNA. This study elucidated a new mechanism for development of COAD and indicated a novel target for treatment of COAD. Furthermore, future studies are required to investigate the molecular mechanism, biomarker value and therapeutic significance of lnc-LAMC2-1:1SNP rs2147578 in COAD.

Declarations

Author contribution statement

Daqing Sun and Jianming Wei: Contributed reagents, materials, analysis tools and data.

Fulong Ji and Zhiwei Yao: Conceived and designed the experiments; Wrote the paper.

Chunxiang Liu and Lushun Ma: Performed the experiments.

Siqi Fu, Bingbing Ren and Yong Liu: Analyzed and interpreted the data; Wrote the paper.

Funding statement

This research was funded by the General Program of the National Natural Science Foundation of China (81770537 and 82070554).

Data availability statement

Data will be made available on request.

Declaration of interest's statement

The authors declare no conflict of interest.

Additional information

No additional information is available for this paper.

Acknowledgements

Thanks to the platforms provided by TCGA, GEPIA, and The Human Protein Atlas databases, and contributors for uploading their meaningful datasets.

References

- H. Sung, J. Ferlay, R.L. Siegel, et al., Global Cancer Statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries [J], CA (2021).
- [2] Z. Zhang, Trefoil factor 3 knock-down prevents autophagy-related gene 12 elevation in colon adenocarcinoma [J], J. Histotechnol. (2019) 1–8.
- [3] J. Tian, J. Lou, Y. Cai, et al., Risk SNP-mediated enhancer-promoter interaction drives colorectal cancer through both FADS2 and AP002754.2 [J], Cancer Research (2020).
- [4] C. Shen, T. Yan, Z. Wang, et al., Variant of SNP rs1317082 at CCSlnc362 (RP11-362K14.5) creates a binding site for miR-4658 and diminishes the susceptibility to CRC [J], Cell Death Dis. 9 (12) (2018) 1177.
- [5] A. Bhan, M. Soleimani, S.S. Mandal, Long noncoding RNA and cancer: a new paradigm [J], Cancer Res. 77 (15) (2017) 3965–3981.
- [6] Y. Fu, Y. Zhang, J. Cui, et al., SNP rs12982687 affects binding capacity of lncRNA UCA1 with miR-873-5p: involvement in smoking-triggered colorectal cancer progression [J], Cell Commun. Signal. : CCS 18 (1) (2020) 37.
- [7] X.F. Chen, D.L. Zhu, M. Yang, et al., An osteoporosis risk SNP at 1p36.12 acts as an allele-specific enhancer to modulate LINC00339 expression via long-range loop formation [J], Am. J. Hum. Genet. 102 (5) (2018) 776–793.
- [8] P. Cui, Y. Zhao, X. Chu, et al., SNP rs2071095 in LincRNA H19 is associated with breast cancer risk [J], Breast Cancer Res. Treat. 171 (1) (2018) 161–171.
- [9] Q. Wang, X.P. Li, X. Zhou, et al., A single-nucleotide polymorphism in lnc-LAMC2-1: 1 interferes with its interaction with miR-128 to alter the expression of deleted in colorectal cancer and its effect on the survival rate of subjects with ovarian cancer [J], J. Cell. Biochem. (2020).
- [10] J. Gong, J. Tian, J. Lou, et al., A functional polymorphism in lnc-LAMC2-1:1 confers risk of colorectal cancer by affecting miRNA binding [J], Carcinogenesis 37 (5) (2016) 443–451.
- [11] J. Wang, H. Liu, G. Xie, et al., Identification of hub genes and key pathways of dietary advanced glycation end products induced nonalcoholic fatty liver disease by bioinformatics analysis and animal experiments [J], Mol. Med. Rep. 21 (2) (2020) 685–694.
- [12] T. Li, J. Fan, B. Wang, et al., TIMER: a web server for comprehensive analysis of tumor-infiltrating immune cells [J], Cancer Res. 77 (21) (2017) e108–e110.
- [13] M. Uhlen, L. Fagerberg, B.M. Hallstrom, et al., Proteomics. Tissue-based map of the human proteome [J], Science (New York, NY) 347 (6220) (2015), 1260419.
- [14] T. Feng, N. Feng, T. Zhu, et al., A SNP-mediated lncRNA (LOC146880) and microRNA (miR-539-5p) interaction and its potential impact on the NSCLC risk [J], J. Exp. Clin. Cancer Res. : CR (Clim. Res.) 39 (1) (2020) 157.
- [15] Y.S. Lee, A. Dutta, MicroRNAs in cancer [J], Annual review of pathology 4 (2009) 199–227.
- [16] H. Zhang, J. Zhong, Y. Tu, et al., Integrated bioinformatics analysis identifies hub genes associated with the pathogenesis and prognosis of esophageal squamous cell carcinoma [J], BioMed Res. Int. 2019 (2019), 2615921.
- [17] D. Szklarczyk, J.H. Morris, H. Cook, et al., The STRING database in 2017: qualitycontrolled protein-protein association networks, made broadly accessible [J], Nucleic Acids Res. 45 (D1) (2017) D362–d368.
- [18] Z. Zhou, Y. Li, H. Hao, et al., Screening hub genes as prognostic biomarkers of hepatocellular carcinoma by bioinformatics analysis [J], Cell Transplantation (2019), 963689719893950.
- [19] Z. Zhang, Y. Chang, J. Zhang, et al., HMGB3 promotes growth and migration in colorectal cancer by regulating WNT/beta-catenin pathway [J], PLoS One 12 (7) (2017), e0179741.
- [20] J. Gu, T. Xu, Q.H. Huang, et al., HMGB3 silence inhibits breast cancer cell proliferation and tumor growth by interacting with hypoxia-inducible factor 1alpha [J], Cancer Manag. Res. 11 (2019) 5075–5089.
- [21] N.A. Al-Tassan, N. Whiffin, F.J. Hosking, et al., A new GWAS and meta-analysis with 1000Genomes imputation identifies novel risk variants for colorectal cancer [J], Sci. Rep. 5 (2015), 10442.
- [22] T. Yang, Z. Zhang, J. Zhang, et al., The rs2147578 C > G polymorphism in the Inc-LAMC2-1:1 gene is associated with increased neuroblastoma risk in the Henan children [J], BMC Cancer 18 (1) (2018) 948.
- [23] M. Hashemi, G. Bahari, M. Naderi, et al., Association of lnc-LAMC2-1:1 rs2147578 and CASC8 rs10505477 polymorphisms with risk of childhood acute lymphoblastic leukemia [J], Asian Pac. J. Cancer Prev. 17 (11) (2016) 4985–4989.
- [24] D. Wang, Y. Li, C. Zhang, et al., MiR-216a-3p inhibits colorectal cancer cell proliferation through direct targeting COX-2 and ALOX5 [J], J. Cell. Biochem. 119 (2) (2018) 1755–1766.

Heliyon 8 (2022) e12342

- [25] H. Song, H.N. Lu, X. Chen, et al., MiR-216a-3p promotes differentiation of BMMSCs into ACE II cells via Wnt/beta-catenin pathway [J], Eur. Rev. Med. Pharmacol. Sci. 22 (22) (2018) 7849–7857.
- [26] Y. Wu, J. Zhang, Y. Zheng, et al., miR-216a-3p inhibits the proliferation, migration, and invasion of human gastric cancer cells via targeting RUNX1 and activating the NF-kappaB signaling pathway [J], Oncology Research 26 (1) (2018) 157–171.
- [27] Z. Su, C. Wang, D. Chang, et al., Limonin attenuates the stemness of breast cancer cells via suppressing MIR216A methylation [J], Biomed. Pharmacother. 112 (2019), 108699.
- [28] T.F. Felix, R.M. Lopez Lapa, M. De Carvalho, et al., MicroRNA modulated networks of adaptive and innate immune response in pancreatic ductal adenocarcinoma [J], PLoS One 14 (5) (2019), e0217421.
- [29] O.A. Elgamal, J.K. Park, Y. Gusev, et al., Tumor suppressive function of mir-205 in breast cancer is linked to HMGB3 regulation [J], PLoS One 8 (10) (2013), e76402.
- [30] X. Li, Y. Wu, A. Liu, et al., MiR-27b is epigenetically downregulated in tamoxifen resistant breast cancer cells due to promoter methylation and regulates tamoxifen sensitivity by targeting HMGB3 [J], Biochem. Biophys. Res. Commun. 477 (4) (2016) 768–773.
- [31] J. Fang, X. Ge, W. Xu, et al., Bioinformatics analysis of the prognosis and biological significance of HMGB1, HMGB2, and HMGB3 in gastric cancer [J], J. Cell. Physiol. 235 (4) (2020) 3438–3446.
- [32] S. Guo, Y. Wang, Y. Gao, et al., Knockdown of high mobility group-box 3 (HMGB3) expression inhibits proliferation, reduces migration, and affects chemosensitivity in gastric cancer cells [J], Med. Sci. Mon. Int. Med. J. Exp. Clin. Res. 22 (2016) 3951–3960.
- [33] J. Gao, Z. Zou, J. Gao, et al., Increased expression of HMGB3: a novel independent prognostic marker of worse outcome in patients with esophageal squamous cell carcinoma [J], Int. J. Clin. Exp. Pathol. 8 (1) (2015) 345–352.
- [34] A. Mukherjee, V. Huynh, K. Gaines, et al., Targeting the high-mobility group box 3 protein sensitizes chemoresistant ovarian cancer cells to cisplatin [J], Cancer Res. 79 (13) (2019) 3185–3191.

- [35] G.H. Zhou, Y.Y. Lu, J.L. Xie, et al., Overexpression of miR-758 inhibited proliferation, migration, invasion, and promoted apoptosis of non-small cell lung cancer cells by negatively regulating HMGB [J], Biosci. Rep. 39 (1) (2019).
- [36] J. Wang, Z. Sheng, Y. Cai, Effects of microRNA-513b on cell proliferation, apoptosis, invasion, and migration by targeting HMGB3 through regulation of mTOR signaling pathway in non-small-cell lung cancer [J], J. Cell. Physiol. 234 (7) (2019) 10934–10941.
- [37] W.J. Zheng, M. Yao, M. Fang, et al., Abnormal expression of HMGB-3 is significantly associated with malignant transformation of hepatocytes [J], World J. Gastroenterol. 24 (32) (2018) 3650–3662.
- [38] W. Zheng, J. Yang, Z. Dong, et al., High mobility group box 3 as an emerging biomarker in diagnosis and prognosis of hepatocellular carcinoma [J], Cancer Manag. Res. 10 (2018) 5979–5989.
- [39] L.K. Wang, X.N. Xie, X.H. Song, et al., Upregulation of miR-200b inhibits hepatocellular carcinoma cell proliferation and migration by targeting HMGB3 protein [J], Technol. Cancer Res. Treat. 17 (2018), 1533033818806475.
- [40] J.W. Yu, W. Mai, Y.L. Cui, et al., Genes and pathways identified in thyroid carcinoma based on bioinformatics analysis [J], Neoplasma 63 (4) (2016) 559–568.
- [41] Y. Yamada, R. Nishikawa, M. Kato, et al., Regulation of HMGB3 by antitumor miR-205-5p inhibits cancer cell aggressiveness and is involved in prostate cancer pathogenesis [J], J. Hum. Genet. 63 (2) (2018) 195–205.
- [42] D.C. Avgousti, C. Herrmann, K. Kulej, et al., A core viral protein binds host nucleosomes to sequester immune danger signals [J], Nature 535 (7610) (2016) 173–177.
- [43] H.W. Choi, D.F. Klessig, DAMPs, MAMPs, and NAMPs in plant innate immunity [J], BMC Plant Biol. 16 (1) (2016) 232.
- [44] L. Li, J. Huang, Z. Ju, et al., Multiple promoters and targeted microRNAs direct the expressions of HMGB3 gene transcripts in dairy cattle [J], Anim. Genet. 44 (3) (2013) 241–250.
- [45] C. Yang, L. Chen, J. Su, et al., Two novel homologs of high mobility group box 3 gene in grass carp (Ctenopharyngodon idella): potential roles in innate immune responses [J], Fish Shellfish Immunol. 35 (5) (2013) 1501–1510.