### **REVIEW ARTICLE**

# Uncommon opportunistic fungal infections of oral cavity: A review

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#### ABSTRACT

The majority of opportunistic oral mucosal fungal infections are due to *Candida albicans* and *Aspergillus fumigatus* species. *Mucor* and *Cryptococcus* also have a major role in causing oral infections, whereas *Geotrichum, Fusarium, Rhodotorula, Saccharomyces* and *Penicillium marneffei* are uncommon pathogens in the oral cavity. The broad spectrum of clinical presentation includes pseudo-membranes, abscesses, ulcers, pustules and extensive tissue necrosis involving bone. This review discusses various uncommon opportunistic fungal infections affecting the oral cavity including their morphology, clinical features and diagnostic methods.

Key words: Immunocompromised patients, opportunistic fungi, oral cavity

# INTRODUCTION

Humans are exposed to hundreds of fungal spores daily, usually not producing any harmful effect on their health. This protection is by various pulmonary defense mechanisms that effectively eliminate the fungal spores.<sup>[1]</sup>

*Coccidioides immitis, Histoplasma capsulatum, Blastomyces dermatitidis, Paracoccidioides brasiliensis* and dermatophyte fungi can infect healthy, immunologically competent individuals.<sup>[1]</sup> By contrast, species such as *Candida, Aspergillus, Rhizopus* and *Fusarium* are normally avirulent in healthy people, but can cause disseminated fatal infections in patients with suppressed immunity. These are called opportunistic pathogenic fungi. The fungus *Cryptococcus neoformans* can be considered both as a true and opportunistic pathogen since it can cause infections in immunologically competent as well as immunocompromised hosts.<sup>[1]</sup>

The occurrence of superficial as well as invasive opportunistic fungal infections has increased significantly over the past two decades. This increase can be attributed to the growing number of immunocompromised patients- including those with AIDS,

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neoplastic disease, advanced age, long-standing diabetes mellitus, undergoing blood and marrow transplantation, solid-organ transplantation, major surgery, receiving immunosuppressive therapy and premature infants.<sup>[2]</sup> Genetic predisposition to invasive fungal infection has been reported recently owing to defective NADPH oxidase activity, abnormal production of tumor necrosis factor-  $\alpha$ , interleukin 10 and other cytokines.<sup>[3,4]</sup>

The spectrum of opportunistic fungal infections is changing. The majority of invasive fungal infections are still due to *Aspergillus* and *Candida* species; but infections due to mycelial fungi other than *Aspergillus* and non-*albicans* species of *Candida* are becoming increasingly common.<sup>[5]</sup> Any fungus present in the environment can be potentially pathogenic in immunocompromised patients.<sup>[6]</sup>

This review considers the main general and oral aspects of these emerging uncommon opportunistic fungal infections such as Candidiasis due to *Candida* species other than *C. albicans*, Aspergillosis due to non-fumigates species of *Aspergillus*, *Mucormycosis*, *Cryptococcosis*, *Geotrichosis*, *Rhodotorula* infection, *Saccharomyces* infection, Fusariosis and Penicilliosis.

# CANDIDIASIS DUE TO CANDIDA SPECIES OTHER THAN C. ALBICANS

Oral candidiasis is the most common human fungal infection.<sup>[7]</sup> Even though *C. albicans* is the most common pathogen responsible for candidiasis, other *Candida* species causing oral infections have also been identified including

*C. glabrata*, *C. krusei*, *C. parapsilosis*, *C. dublieniesis*, *C. tropicalis*, *C. kefyr* and *C. guilliermondii*.<sup>[8-11]</sup> Less commonly isolated species are *C. inconspicua*, *C. lusitaniae*, *C. norvegensis* and *C. rugosa*.<sup>[12]</sup> These species are resistant to the commonly used azole antifungal drugs.<sup>[13]</sup> These non-*Candida albicans Candida* species lack many of the virulence factors present in the virulent *C. albicans* i.e. the ability to form hyphae and phenotypic switching. They have low adherence capacity to buccal epithelial and vascular endothelial surfaces. They secrete less proteinases. Thus they are thought to cause candidiasis of less severity.<sup>[14]</sup>

*C. dublieniesis* is a species associated with oral lesions in HIV-infected individuals. It is morphologically and genotypically closely related to *C. albicans*.<sup>[15,16]</sup> *C. dublieniesis* is the only *Candida* species other than *Candida albicans* that forms true hyphae. They show decreased susceptibility to fluconazole.<sup>[17]</sup>

*C. glabrata* is emerging as an important pathogen in both mucosal and blood stream infections and is commonly isolated from the oral cavity of HIV-infected individuals.<sup>[11,14]</sup> *C. glabrata*-associated oropharyngeal candidiasis in HIV infection and cancer patients is more severe and more difficult to treat due to their quick development of resistance to fluconazole.<sup>[18]</sup>

*C. guilliermondii* cause infection in patients undergoing surgical procedures, endocarditis, in intravenous drug users and fungemia in immunocompromised patients.<sup>[19]</sup> They show resistance to amphotericin B.<sup>[20]</sup>

*C. krusei* infection occurs in critically ill patients mainly in hematology patients with severe neutropenia. It is an uncommon pathogen causing candidemia. The increase in *C. krusei* infection in HIV-infected patients is thought to be due to the widespread use of fluconazole prophylaxis.<sup>[21]</sup>

*C. lusitaniae* mainly causes infection in immunocompromised hosts with prolonged antibiotic administration, hospitalization, cytotoxic or corticosteroid treatment, granulocytopenia and low-birthweight babies.<sup>[22,23]</sup>

*C. parapsilosis* mainly affects critically ill neonates and patients in intensive care units i.e. prematurity and low birth weight are recognized as the risk factors.<sup>[24]</sup>

*C. tropicalis* is the most virulent of the non-*albicans Candida* species. This may be due to its ability to adhere to epithelial cells *in vitro* and to secrete moderate levels of proteinases.<sup>[14]</sup> It is usually isolated from oral cavity and skin and can cause infections in esophagus in patients with systemic diseases.<sup>[25]</sup>

# Diagnosis

The diagnosis of candidiasis is often made on the basis of clinical suspicion of the typical mucosal changes and angular

cheilitis. These are almost always associated with some degree of discomfort.<sup>[26]</sup> Microscopic examination of the smears stained with periodic acid Schiff's method (PAS), or KOH preparation can reveal candidal hyphae and blastospores.<sup>[27]</sup>

Qualitation is accomplished by culture on Sabouraud's dextrose agar (SDA) at 25-30°C for 48-72 hours.[28] Germ tube growth can be used for identification of C. albicans whereas carbohydrate assimilations can be employed for other species.<sup>[26]</sup> Culture on CHROMagar can be used to identify C. albicans, C. krusei and C. tropicalis. C. albicans forms green colonies, C. krusei form fuzzy, rose-colored colonies and C. tropicalis show steel-blue colonies.[13,29,30] CHROMagar-PAL is a new method which combines CHROMagar Candida and Pal agar that can easily differentiate between C. albicans and C. dublieniesis even in mixed cultures. ELISA and polymerase chain reaction (PCR) may also be used for the same. Multiplex PCR, which identifies yeast directly in clinical samples is a more recent advancement.<sup>[31]</sup> Further speciation can be done by conventional biotyping techniques or with the aid of commercially available systems like Micronaut-Candida, API ID32C, RapID yeast plus system, Auxocolor, Vitek, Api Candida, etc.<sup>[32,33]</sup> Quantitation is done by collecting whole saliva and plating on SDA, incubating and counting the colony-forming units.<sup>[28]</sup>

Biopsy is mainly done to rule out hyperplastic candidiasis. In hyperplastic candidiasis, histopathological examination will reveal epithelial parakeratosis with polymorphonuclear leukocytes in the superficial layers. PAS-staining will show the presence of Candidal hyphae in these areas.<sup>[34]</sup>

# **ASPERGILLOSIS**

Aspergillosis has been reported as the second most prevalent opportunistic fungal infection.<sup>[35]</sup> *Aspergillus* species are universally found in humid areas, damp soil, grain, cereal, mouldy flour and organic decaying or decomposing matter.<sup>[36]</sup> In the hospital setting, construction activities, rotten leaves or insufficient cleaning of dust can increase the risk of developing aspergillosis.<sup>[37]</sup> *Aspergillus fumigatus* is the most familiar pathogen of the species. Human infections are also caused by less common *Aspergillus* species, such as *Aspergillus flavus, Aspergillus glaucis, Aspergillus terrus, Aspergillus parasiticus, Aspergillus repens, Aspergillus nidulans, Aspergillus niger* and *Aspergillus tubingensis*.<sup>[38,39]</sup> In India, the most common species encountered is *A. flavus* followed by *Aspergillus fumigatus* and *A. niger*.<sup>[40]</sup>

Rowe Jones in 1994 classified aspergillosis into three chief variants: Invasive, non-invasive and non-invasive destructive type. Invasive type represents true fungal tissue invasion that can be either slow progressive and destructive (non-fulminant) or highly aggressive and lethal (fulminant). Non-invasive type is further classified into Aspergilloma, Fungal ball, Mycetoma (usually affecting one sinus) or allergic Aspergillus

sinusitis (involving more than one sinus). Destructive non-invasive variant is locally destructive but shows no tissue invasion.<sup>[41]</sup>

### **Clinical presentation**

Aspergillosis generally occurs after inhalation of spores, that can result in both upper and lower respiratory tract infection- bronchopulmonary aspergillosis.<sup>[42]</sup> From lungs, infections may spread to the brain, bone or endocardium.<sup>[43]</sup> Paranasal sinuses, larynx, eyes, ears and the oral cavity may be involved in primary aspergillosis.<sup>[43,44]</sup> *A. fumigatus* is the usual agent of sinus aspergillosis, whereas *A. flavus* is more common in invasive lesions in immunosuppressed individuals.<sup>[45]</sup> Orofacial aspergillosis is relatively common in patients undergoing treatment for malignancies of the blood and blood-forming organs.<sup>[44,46-48]</sup>

Aspergillus does not contain chlorophyll, thus light is not required for growth. Aspergillus exhibits a centrifugal linear growth unless inhibited by natural or artificial barriers. This is why Aspergillus in the paranasal sinus eventually develops into a ball-shaped mass.<sup>[49]</sup> The centre of the mass contains calcium phosphate and therefore mimics a foreign body on radiography.<sup>[50]</sup>

In most cases, oral aspergillosis lesions are yellow or black in color, with a necrotic ulcerated base, classically located on the palate or posterior tongue.<sup>[46,51]</sup> *Aspergillus hyphae* invade host tissues through the release of various toxins. These include various proteases, phospholipases, hemolysins, gliotoxin, aflatoxin, phthioic acid and other toxins.<sup>[52]</sup> The hyphal elements of the fungus may invade the oral mucosa and penetrate the walls of small to medium-sized blood vessels, producing thrombosis, infarction and necrosis, finally leading to systemic spread.<sup>[53]</sup>

# Diagnosis

Histopathologically, invasive lesions show chronic granulomatous reactions. In hematoxylin and eosin-stained sections hyphal forms can be seen faintly in the center of an area of necrosis. They may go unnoticed unless special stains like methanamine silver are used. The fungi appear as septate hyphae, showing branching at 45° angles and are about 2-4 mm in diameter.<sup>[54]</sup> Conidiospores and fruiting bodies are also seen. This fungus should be differentiated from mucor which shows broader non-septate hyphae with branching at 90°.<sup>[55,56]</sup>

# **MUCORMYCOSIS**

Mucormycosis (Zygomycosis or Phycomycosis) is an opportunistic mycotic infection caused by Mucorales.<sup>[57]</sup> It is considered as the third most common opportunistic fungal infection after candidiasis and aspergillosis.<sup>[58]</sup> The common

genera causing disease are Rhizopus, Rhizomucor and Absidia. Rhizopus is the chief pathogen accounting for 90% of the cases of rhinocerebral mucormycosis.<sup>[59]</sup>

This fungus is widespread in soil, manure, vegetables and as bread mould.<sup>[60]</sup> This pathogen may be cultured from the oral cavity, nasal passages, throat and stool of healthy patients without clinical signs of infection.<sup>[59]</sup> The infection usually results from inhalation of fungal spores, contamination of traumatized tissue, ingestion or direct inoculation.<sup>[61]</sup> An area of ulceration or an extraction socket in the mouth can be a port of entry for mucormycosis into the maxillofacial region, chiefly when the patient is immunocompromised.<sup>[62]</sup>

Up to 40-50% of patients suffering from mucormycosis have diabetes mellitus (DM).<sup>[63,64]</sup> Acidosis in DM reduce the phagocytic ability of granulocytes thereby affecting the immunological capability to resist mucormycosis.<sup>[65,66]</sup> During the state of diabetic ketoacidosis, the acidic environment and the increase in the levels of free ferric ions support the growth of mucorales.<sup>[67]</sup> In diabetic patients there is a high occurrence of mucormycosis caused by Rhizopus oryzae, because they produce the enzyme ketoreductase, which enables them to make use of the patient's ketone bodies.<sup>[68]</sup>

Mucorales have the ability to damage and penetrate endothelial lining of blood vessels. This accounts for the most characteristic feature of mucormycosis i.e. widespread angioinvasion resulting in thrombosis and tissue necrosis.<sup>[69]</sup>

# **Clinical presentation**

Eisenberg *et al.* described six clinical variants- rhinocerebral (rhinomaxillary), pulmonary, cutaneous, gastrointestinal, central nervous system and disseminated type.<sup>[68]</sup> Rhinocerebral form is the most common clinical variant which has been further divided into two subtypes: (i) A highly fatal rhino-orbito-cerebral form which is invasive involving the ophthalmic and internal carotid arteries, (ii) A less fatal rhino-maxillary form which involves the sphenopalatine and greater palatine arteries, resulting in thrombosis of the turbinate and necrosis of the palate.<sup>[59,70,71]</sup>

The clinical presentation of rhinocerebral mucormycosis includes malaise, headache, facial pain, swelling an irregular black eschar, exudation of pus from the eye and nose and low-grade fever.<sup>[64]</sup> The disease usually starts in the nasal mucosa or palate and spread through the surrounding vessels to the paranasal sinuses, frequently involving maxillary and ethmoid sinuses. In addition, mucormycosis can involve the retro-orbital region by direct extension.<sup>[61,63,72]</sup> Orbital involvement can impair the functions of cranial nerves III, IV and VI resulting in proptosis, ptosis, pupillary dilatation, orbital cellulitis and loss of vision. Direct penetration and growth of the fungi through the walls of blood vessels can result in thrombosis and extensive tissue necrosis.<sup>[57]</sup> Hematogenous

spread to the cavernous sinus can lead to fatal cavernous sinus thrombosis.<sup>[61,64,73]</sup> Rhinocerebral mucormycosis can also spread by perineural invasion.<sup>[74]</sup>

# Diagnosis

Suspicion of mucormycosis necessitates a CT scan of the maxilla, orbits and brain. In particular, evidence of intracranial brain abscesses and orbital extensions is critical. Sinus and orbital extensions are recognized by membrane or periosteal thickenings as well as bony disruption.<sup>[75]</sup>

Routine blood studies will show leukocytosis in the 12,000-20,000/ mm<sup>3</sup> range and usually a shift to the left. If the patient is diabetic, a full workup of serum glucose, electrolytes, blood chemistries and blood gases is required. Serological assays for Mucor antigens have been developed.<sup>[75]</sup>

Histopathologically broad, irregularly shaped, nonseptate hyphae with right angle branching are seen invading the tissue in H and E-stained sections; but are better visualized with PAS or methanamine silver stains. Ideally the biopsy specimen should be obtained from the junction of necrotic and non-necrotic tissue. The organisms are mainly seen within the walls of necrotic blood vessels.<sup>[75]</sup> Culturing can be done on Sabouraud's glucose agar and sporulation of fungal hyphae within 24-48 hours helps to identify mucor.<sup>[76]</sup>

# CRYPTOCOCCOSIS

*Cryptococcus neoformans* and *Cryptococcus gattii* are commonly considered as the causative agents of cryptococcosis.<sup>[34]</sup> *C. neoformans* generally affects immunocompromised hosts whereas *C. gattii* is isolated more from immunocompetent individuals.<sup>[34,77]</sup>

# **Clinical presentation**

*C. neoformans* infections usually occurs after inhalation of fungal spores from the soil and excreta of birds like pigeons, parrots and canaries.<sup>[78]</sup> In immunocompetent individuals the infection remains subclinical within the lungs. In the immunocompromised host, the fungus produces rapid disseminated infection involving central nervous system, skin, mucous membranes and many other tissues.<sup>[79]</sup>

The face, scalp and neck are the common sites of cutaneous lesions, presenting as papules, acne form pustules, abscesses, ulcers, superficial granulomas or sinus tracts.<sup>[80]</sup> Dissemination may occur from reactivation of dormant disease or a primary infection.<sup>[81]</sup> The most common clinical presentation is meningo encephalitis. The increased occurrence of cryptococcosis in HIV-infected individuals has been reduced by the implementation of Highly Active Anti Retro viral Therapy (HAART).<sup>[81]</sup>

Cryptococcosis in oral cavity may arise from hematogenous spread of the infection localized in the lungs of AIDS patients. However, oral cryptococcosis can be the initial presentation of a disseminated infection.<sup>[81]</sup> Violaceous nodules, swellings or ulcers have been reported on the gingiva, hard and soft palates, pharynx, oral mucosa, tonsillar pillar and in tooth socket after extraction.<sup>[82]</sup>

# Diagnosis

Histopathology varies according to the immunological status of the host. In immunocompetent hosts, typical granulomas are formed at the site of cryptococcal infection, with multinucleated giant cells containing intracytoplasmic cryptococci in budding forms. In immunosuppressed patients, proliferating cryptococci present as extra- and intracellular yeast cells with some budding forms with reactive macrophages, minor lymphocytic and neutrophilic infiltrate. The definitive diagnosis of cryptococcosis is established with periodic acid Schiff (PAS), methanamine silver and mucicarmine-stained preparations. The fungal cytoplasm appears bright magenta by PAS stain and mucicarmine stains the fungal capsule.<sup>[81,83]</sup> Culture and assay of serum or cerebrospinal fluid for capsular antigen is useful.<sup>[84]</sup>

# **GEOTRICHOSIS**

Geotrichosis is caused by *Geotrichum candidium* which is a component of the normal microflora of the skin and the mucosa of the respiratory and digestive tracts. It can also be isolated from vegetables, fruits, soil and plants.<sup>[85]</sup> Oral lesions are caused by *G. candidum* and *G. capitatum*.<sup>[86-89]</sup>

# **Clinical presentation**

Geotrichosis can present as pseudomembranes, mucosal ulcerations, edematous and erythematous gingivae. Easily scrapable creamy-white pseudomembranous plaques with an erythematous background can be seen mainly on the tongue, resulting in glossitis and on the cheeks. The most common symptoms are burning pain and impaired swallowing.<sup>[90]</sup> Angular cheilitis and palatal ulcers appearing similar to the ulcers caused by zygomycosis and aspergillosis have been reported. Palatal ulcers can result in a very aggressive palatine- cerebral condition with poor prognosis.<sup>[90,91]</sup>

# Diagnosis

Direct examination and staining will reveal multiple septate hyphae with rectangular arthroconidia. But some arthroconidia may have rounded appearance (clavata cells) and may be easily mistaken for Candida. Hence culture is recommended for the accurate diagnosis.<sup>[90]</sup>

In culture *G. candidum* and *G. capitatum* are seen as white, membranous, villous, wet colonies. In chromogenic culture

media their villous wet growth with slight pink pigmentation distinguishes them from the major *Candida* species.<sup>[90]</sup> Various biochemical tests can also aid the diagnosis; but molecular biology is the most accurate technique for species identification by internal transcriber spacer (rDNA).<sup>[86,92]</sup>

### **RHODOTORULA INFECTION**

The genus Rhodotorula is a pigmented yeast classified under the family Cryptococcaceae. Three main species are actually known: *Rhodotorula glutinis*, *Rhodotorula minuta* and *Rhodotorula mucilaginosa*.<sup>[93]</sup> *R. mucilaginosa* is the current name for the species formerly known as *Rhodotorula rubra*. Rhodotorula is found in air, soil, lakes, ocean water and dairy products. It may colonize plants, humans and other mammals.<sup>[94,95]</sup> Rhodotorula produce moist, smooth to mucoid, glistening, pigmented colonies. Salmon-pink to coral red color of the colony is due to the carotenoid pigment, torularhodin.<sup>[96]</sup> This pigment blocks certain wavelengths of light which can damage the yeast cell.<sup>[94,95]</sup>

#### **Clinical presentation**

Rhodotorula can cause meningitis, endocarditis, ventriculitis, peritonitis, fungemia, central venous catheter infection and keratitis.<sup>[97]</sup> Non-healing oral ulcers and white patches are the reported oral presentations of Rhodotorula infection.<sup>[98]</sup>

# Diagnosis

Gram stain will show Gram-positive round, budding yeast cells of 4-8  $\mu$ m in diameter with clear halo around them. India ink preparation of the smear will demonstrate encapsulated budding yeast cells.<sup>[99]</sup>

Rhodotorula are pigmented yeasts easily identified as coral pink, smooth, sometimes reticulate, rugose or corrugated and moist to mucoid yeast-like colony forms when grown on SDA. The germ tube test is negative. Rhodotorula produces urease enzyme and does not ferment carbohydrates.<sup>[100]</sup>

# SACCHAROMYCES INFECTION

*Saccharomyces cerevisiae* (also known as "baker's yeast" or "brewer's yeast") is widespread in nature and is a commensal inhabiting the gastrointestinal tract of humans, which has an important role in maintaining the normal homeostasis of the lower gastrointestinal tract.<sup>[101,102]</sup> *S. cerevisiae* is now included in some diet or health foods. Fungemia from *S. cerevisiae* can follow the use of live yeast capsules of *Saccharomyces boulardii* which are used as probiotics for the prevention and treatment of various diarrheal disorders.<sup>[103]</sup>

# **Clinical presentation**

Lesions resemble invasive candidiasis due to the presence of choreoretinitis and esophagitis in both conditions. Fever may be present in majority of patients. Deep site involvement with necrosis and granulomatous reaction has also been reported.<sup>[104]</sup> Intra-oral manifestations include ulcers with associated painful swallowing, dry mouth and burning sensation.<sup>[102]</sup>

### Diagnosis

Direct Gram stain from the swab will show majority of Gram-positive budding yeast cells. The culture will give creamy-white yeast-like growth. The Gram staining from the colony will show budding yeast cells without any capsule. The organism can be identified using corn meal agar and carbohydrate assimilation test.<sup>[102]</sup>

#### **FUSARIOSIS**

*Fusarium* species are important plant pathogens causing various diseases on cereal grains, occasionally causing infection in animals.<sup>[105,106]</sup> In humans, *Fusarium* species cause both superficial infections (such as keratitis and onychomycosis), allergic diseases and disseminated diseases.<sup>[107]</sup>

*Fusarium solani* was most frequent species followed by *Fusarium oxysporum* and *Fusarium verticillioidis* and *Fusarium moniliforme*.<sup>[105]</sup> *Fusarium* species are the second-most common mould causing invasive fungal infections in immunocompromised individuals.<sup>[108,109]</sup>

#### **Clinical presentation**

The clinical presentation of fusariosis depends principally on the immune status of the host and the portal of entry of the infection.<sup>[110]</sup> The main routes of entry for *Fusarium* species are the airways and the skin. Risk factors are persistent neutropenia, severe depletion of T- lymphocytes and previous fungal infections.<sup>[111]</sup> Disseminated infection and skin involvement leading to cellulitis are seen in immunocompromised patients. These lesions may frequently appear as multiple erythematous papules or nodules with central necrosis.<sup>[110]</sup>

Fusariosis presenting as a necrotic ulceration of the gingiva, extending to the alveolar bone has been reported in a granulocytopenic patient.<sup>[112]</sup>

### Diagnosis

Skin lesions (cellulitis and metastatic lesions) and positive blood cultures for mold can suggest disseminated fusariosis. Due to the dissemination of yeast-like structures, blood cultures are often positive in fusariosis.<sup>[113]</sup> In histopathology the hyphae are similar to those of Aspergillus species, with hyaline and septate filaments that typically dichotomize in acute and right angles. The finding of hyphae and yeast-like structures together is highly suggestive of fusariosis.<sup>[114]</sup> From cultures, *Fusarium* can be identified by the presence

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of hyaline, banana shaped multicellular macroconidia with a foot cell at the base.<sup>[115]</sup> But species identification is difficult and needs molecular techniques like PCR.<sup>[115]</sup>

#### PENICILLIOSIS

Penicilliosis is caused by *Penicillium marneffei* which can cause fatal infection in HIV-infected individuals.<sup>[116]</sup> It is a dimorphic fungus that can exist in mycelia form at 25°C and yeast form at 37°C.<sup>[117]</sup> It was first identified in 1973, as an opportunistic infection in a patient with Hodgkins lymphoma.<sup>[118]</sup> In South-east Asian countries like Thailand, Penicilliosis is the third-most common opportunistic infection in people with AIDS.<sup>[116]</sup>

#### **Clinical presentation**

Penicilliosis is mostly seen in late HIV infection with CD4+ count less than 100/ $\mu$ L.<sup>[119-121]</sup> Inhalation of air-borne conidia is the most common mode of transmission.<sup>[122]</sup> Most of the patients have fever, weight loss and malaise. Skin manifestation such as subcutaneous abscesses, molluscum contagiosum-like lesions and papule-like ulcers may be present.<sup>[123]</sup> Respiratory involvement is characterized by productive cough, dyspnea and hemoptysis.<sup>[124]</sup> Oral lesions include papules, erosions or shallow ulcers covered by yellow necrotic slough which are mainly seen on the palate, gingiva, labial mucosa, tongue and oropharynx.<sup>[125]</sup>

# Diagnosis

The diagnosis of penicilliosis may be made through examination of cytology or biopsy specimens.<sup>[126]</sup> In fungemia, yeast cells may be seen inside monocytes in peripheral blood smear and are best demonstrated by PAS/methanamine silver stain.<sup>[127]</sup> Detection of non-budding yeast cells with characteristic central transverse septum is the key to diagnosis which should be confirmed by microbiological culture.<sup>[126]</sup> isolation of *P. maneffei* is the gold standard for diagnosis. Culture on SDA at 25°C for 3 days will give granular colonies with greenish-yellow color and a characteristic red diffusible pigment.<sup>[126]</sup> In histopathologically occult cases, methanamine silver stain can aid in the diagnosis.<sup>[127]</sup> Various types of antigen-antibody testing and PCR assay specific to *P. marneffei* have been developed in research laboratories, but are not widely available.<sup>[126]</sup>

#### CONCLUSIONS

Fungi are opportunistic infectious agents and most of them are not usually pathogenic. But when they infect an immunocompromised host, can cause a wide range of diseases ranging from superficial to disseminated infections involving the vital internal organs. The outcome of invasive fungal infections depends on various factors such as the clinical condition of the patient, immunological status, pathogenicity and virulence factors of the invading fungal species and the location of the infected area.  $^{\left[ 128\right] }$ 

The range of patients at risk for invasive fungal infections continues to expand beyond the normal host to encompass patients with acquired immunodeficiency syndrome; diabetes mellitus, those undergoing therapy for cancer and organ transplantation and major surgical procedures. As the population at risk continues to expand, so also does the spectrum of opportunistic fungal pathogens infecting these patients.

Inhalation of spores of these microorganisms is the most common mode of infection in a susceptible host. Hence, prevention of the same would be an excellent prophylactic measure to contain opportunistic fungal infections especially in immunocompromised individuals.

# REFERENCES

- Odds FC, Gow NA, Brown AJ. Fungal virulence studies come of age. Genome Biol 2001;2:REVIEWS 1009.
- 2. Pfaller MA. Invasive fungal pathogens: Current epidemiological trends. Clin Infect Dis 2006;43:S3-14.
- 3. Segal BH, Romani LR. Invasive aspergillosis in chronic granulomatous disease. Med Mycol 2009:S282-90.
- Baskova L, Buchta V. Laboratory diagnostics of invasive fungal infections: An overview with emphasis on molecular approach. Folia Microbiol 2012;57:421-30.
- Singh N. Trends in the epidemiology of opportunistic fungal infections: Predisposing factors and the impact of antimicrobial use practices. Clin Infect Dis 2001;33:1692-6.
- Badiee P, Hashemizadeh Z. Opportunistic invasive fungal infections: Diagnosis and clinical management. Indian J Med Res 2014;139:195-204.
- Akpan A, Morgan R. Oral candidiasis. Postgrad Med J 2002;78:455.
- Cannon RD, Chaffin WL. Oral colonization by candida albicans. Crit Rev Oral Biol Med 1995;10:359.
- Belazi M, Velegraki A, Koussidou-Eremondi T, Andrealis D, Hini S, Arsenis G, *et al.* Oral Candida isolates in patients undergoing radiotherapy for head and neck cancer: Prevalence, azole susceptibility profiles and response to antifungal treatment. Oral Microbiol Immunol 2004;19:347.
- Odds FC. Candida and candidosis A review and bibliography. 2<sup>nd</sup> ed.vol 1. Bailliere Tindall –WB Saunders London; 1988.
- 11. Li L, Redding S, Dongari-Bagtzoglou A. Candida glabrata, an emerging opportunistic pathogen. J Dent Res 2007;86:204.
- Richardson MD, Warnock DW. Fungal infections: Diagnosis and management. 3<sup>rd</sup> ed.vol 1. Oxford (UK): Blackwell Publishing; 2003.
- Meurman JH, Siikala E, Richardson M, Rautemaa R. Non-candida albicans Candida yeasts of the oral cavity. Communicating Current Research and Educational Topics and Trends in Applied Microbiolgy. A Mendez-Vilas (Ed). 2007; 1: 719-731.
- Moran GP, Sullivan DJ, Coleman DC. Emergence of non-Candida albicans Candida species as pathogens. In: Candida and candidiasis. 4<sup>th</sup> ed.vol 1. Washington: ASM Press; 2002. p. 37-53.

- Sullivan DJ, Westernberg TJ, Haynes KA, Bennett DE, Coleman DC. Candida dublieniesis sp. Nov; Phenotypic and molecular characterization of a novel species associated with oral candidosis in HIV infected individuals. Microbiology 1995;141:1507.
- Coleman DC, Sullivan DJ, Bennett DE, Moran GP, Barry HJ, Shanley DB. Candidiasis; the emergence of a novel species, Candida dublieniesis. AIDS 1997;11:557.
- Pinjon E, Moran GP, Coleman DC, Sullivan DJ. Azole susceptibility and resistance in Candida dublieniesis. Biochem Soc Trans 2005;33:1210.
- Nucci M, Marr KA. Emerging Fungal Diseases. Clin Infect Dis 2005;41:521.
- Mardani M, Hanna HA, Girgawy E, Raad I. Nosocomial Candida guilliermondii fungemia in cancer patients. Infect Control Hosp Epidemiol 2000;21:336-7.
- Hazen KC. New and emerging yeast pathogens. Clin Microbiol Rev 1995;8:462.
- Samaranayke YH, Samaranayke LP. Candida krusei: Biology, epidemiology, pethogenecity and clinical manifestations as an emerging pathogen. J Med Microbiol 1994;41:295.
- 22. Viudes A, Peman J, Canton E, Salavert M, Ubeda P, Lopez-Ribot JL, *et al.* Two cases of fungemia due to Candida lusitaniae and a literature review. Eur J Clin Microbiol Infect Dis 2002;21:294.
- Smith PB, Steinbach WJ, Benjamin DK. Neonatal candidiasis. Infect Dis Clin North Am 2005;19:603.
- 24. Sarvikivi E, Lyytikainen O, Soll DR, Pujol C, Pfaller MA, Richrdson MA. Emergence of fluconazole resistance in Candida parapsilosis strain that caused infections in a neonatal intensive care unit. J Clin Microbiol 2005;43:2729.
- Maenza JR, Merz WG. Candida albicans and related species. In: Jonathan Cohen, William G.Powderly and Gill day. Infectious Diseases, 2<sup>nd</sup> ed, vol 1. Saunders; 1998. p. 2313-22.
- Epstein JB, Silverman S, Fleischmann J. Oral fungal infections. In: Silverman S, Eversole LR and Truelove EL. Essentials of Oral Medicine, 3<sup>rd</sup> ed vol 1. Hamilton (London):BC Decker Inc; 2002. p. 170-9.
- Neville BW, Damm DD, Allen CM and Bouquot JE. Oral and Maxillofacial Pathology, 3<sup>rd</sup> ed. .Philadelphia: Saunders Company Ltd; 2005.
- Samaranayake L. Essential Microbiology for Dentistry. 3<sup>rd</sup> ed., vol 1. Edinburgh: Churchill Livingstone; 2006. p. 255, 62-4.
- 29. CHROMagar microbiology, Paris, France. Available from: http://chromagar.com [Last accessed on 13 May 2013].
- Hospenthal DR, Beckius ML, Floyd KL, Horvath LL, Murray CK. Presumptive identification of Candida species other than C. albicans, C.krusei and C. tropicalis with the chromogenic medium CHROMagar candida. Ann Clin Microbiol Antimicrob 2006;5:1.
- Coronado-Castelotte L, Jimenez-Soriano Y. Clinical and microbiological diagnosis of oral candidiasis. J Clin Exp Dent 2013;5:e279-86.
- 32. Szabo Z, Toth B, Kovacs M, Kardoz G, Maraz A, Rozgonyi F, et al. Evaluation of the new Micronaut Candida system compared to the API ID 32C method for yeast identification. J Clin Microbiol 2008;46:1824-5.
- Verweji PE, Breuker IM, Rijs AJ, Meis JF. Comparative study of seven yeast identification systems. J Clin Pathol 1999;52:271-3.
- Krishnan PA. Fungal infections of the oral mucosa. Indian J Dent Res 2012;23:650-9.

- Hartwick RW, Batsakis JG. Sinus aspergillosis and allergic fungal sinusitis. Ann Otol Rhinol Laryngol 1991;100:427-30.
- Tamgadge AP, Mengi R, Tamgadge S, Bhalerao SS. Chronic invasive aspergillosis of paranasal sinuses: A case report with review of literature. J Oral Maxillofac Pathol 2012;16:460-4.
- 37. Marschmeyer G, Ruhnke M. Update on antifungal treatment of invasive Candida and Aspergillus infections. Mycoses 2004;47:263-6.
- 38. Scully C, de Almeida OP. Orofacial manifestations of then systemic mycoses. J Oral Pathol Med 1992;21:289-94.
- Bathoorn E, Salazar NE, Sepehrkhouy S, Meijer M, de Cock H, Haas P. Involvement of then opportunistic pathogen Aspergillus tubingensis in osteomyelitis of the maxillary bone: A case report. BMC Infect Dis 2013;13:59.
- 40. Bajwa SJ, Kulshrestha A. Fungal infections in intensive care unit: Challenges in diagnosis and management. Ann Med Health Sci Res 2013;3:238-44.
- Rowe-Jones JM, Meore-Gillon V. Destructive noninvasive paranasal sinus *aspergillosis*: Component of a spectrum of disease. J Otolaryngol 1994;23:92-6.
- 42. Dreizen S, Keating MJ, Beran M. Orofacial fungal infections. Nine pathogens that may invade during chemotherapy. Postgrad Med 1992;91:349-50, 353-44, 357-60.
- 43. Benson-Mitchell R, Tolley N, Croft CB, Gallimore A. Aspergillosis of the larynx. J Laryngol Otol 1994;108:883-5.
- Myoken Y, Sugata T, Kyo TI, Fujihara M. Pathological features of invasive oral aspergillosis in patients with hematologic malignancies. J Oral Maxillofac Surg 1996;54:263-70.
- 45. Emmanuelli JL. Infectious granulomatous diseases of the head and neck. Am J Otolaryngol 1993;14:155-67.
- 46. Chambers MS, Lyzak WA, Martin JW, Lyzak JS, Toth BB. Oral complications associated with aspergillosis in patients with a hematologic malignancy. Presentation and treatment. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 1995;79:559-63.
- Dreizen S, Bodey GP, McCredie KB, Keating MJ. Orofacial aspergillosis in acute leukemia. Oral Surg Oral Med Oral Pathol 1985;59:499-504.
- Sugata T, Myoken Y, Kyo T, Fujihara M. Invasive oral aspergillosis in immunocompromised patients with leukemia. J Oral Maxillofac Surg 1994;52:382-6.
- 49. Rossouw DP, Swart JG. Aspergillus fumigatus infection of the maxillary sinus. S Afr Med J 1988;73:47-78.
- Ogata Y, Okinaka Y, Takahashi M. Antrolith associated with Aspergillosis of the maxillary sinus: Report of a case. J Oral Maxillofac Surg 1997;55:1339-41.
- Napoli JA, Donegan JO. Aspergillosis and necrosis of the maxilla: A case report. J Oral Maxillofac Surg 1991;49:532-4.
- 52. Denning DW. Invasive Aspergillosis. Clin Infect Dis 1998;26:781-805.
- Dreizen S. Oral complications of cancer therapies: Description and incidence of oral complications. Monogr Natl Cancer Inst 1990;9:11-5.
- Dayananda BC, Vandana R, Rekha K, Kumar GS. Aspergillosis of the maxillary antrum. A case report. J Oral Maxillofac Pathol 2002;1:26-9.
- Sapp JP, Eversole LR, Wysocki GP. Contemporary Oral and Maxillofacial Pathology. 2<sup>nd</sup> ed., vol 1. Missouri: Mosby-An Affiliate of Elsevier; 2004. p. 207-51.
- Regezi JA, Sciubba JJ, Jordan RC, editors. Oral pathology-Clinical Pathologic Correlations. 4<sup>th</sup> ed., vol 1. Missouri: Saunders-An Imprint of Elsevier Science; 2003. p. 23-74.

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- 57. Bakathir AA. Mucormycosis of the jaw after dental extractions: Two case reports. Sultan Qaboos Univ Med J 2006;6:77-82.
- Perusquia-Ortiz AM, Vazquez-Gonzalez D, Bonifaz A. Opportunistic filamentous mycoses: Aspergillosis, mucormycosis, phaehyphomycosis and hyalohyphomycosis. J Dtsch Dermatol Grs 2012;10:611-21.
- Auluck A. Maxillary necrosis by mucormycosis. A case report and literature review. Med Oral Patol Oral Cir Bucal 2007;12:E360-4.
- Bouza E, Munoz P, Guinea J. Mucormycosis: An emerging disease? Clin Microbiol Infect 2006;12:7-23.
- 61. Sugar AM. Mucormycosis. Clin Infect Dis 1992;14:126-9.
- Huang JS, Kok SH, Lee JJ, Hsu WY, Chiang CP, Kuo YS. Extensive maxillary sequestration resulting from mucormycosis. Br J Oral Maxillofac Surg 2005;43:532-4.
- 63. McNulty JS. Rhinocerebral mucormycosis: Predisposing factors. Laryngoscope 1982;92:1140-4.
- Kim J, Fortson JK, Cook HE. A fatal outcome from rhinocerebral mucormycosis after dental extractions: A case report. J Oral Maxillofac Surg 2001;59:693-7.
- Gale GR, Welch AM. Studies of opportunistic fungi. Inhibition of Rhizopus oryzae by human serum. Am J Med Sci 1961;241:604-8.
- Bauer H, Flanagan JF, Sheldon WH. Experimental cerebral mucormycosis in rabbits with alloxan diabetes. Yale J Biol Med 1955;28:29-32.
- 67. Damante JH, Fleury RN. Oral and rhinocerebral mucormycosis: Case Report. J Oral Maxillofac Surg 1998;56:267-71.
- Hadzri MH, Azariman SM, Fauzi AR, Kahairi A. Invasive rhinocerebral mucormycosis with orbital extension in poorly controlled diabetes mellitus. Singapore Med J 2009;50:e107.
- 69. Tippu SR, Rahman F, Pilania D. mucormycosis of maxilla an overview and case report. THE CUSP 2013;10:23-6.
- Woo SB, Greenberg MS. Ulcerative, vesiculous and bullous lesions. In: Greenberg MS, editor. Glick Burket's Oral Medicine. 11<sup>th</sup> ed.vol 1. BC Decker Inc Hamilton; 2008. p. 74-5.
- 71. Cohen SG, Greenberg MS. Rhinomaxillary mucormycosis in kidney transplant patient. Oral Surg 1980;50:33-8.
- Jones AC, Bentsen TY, Freedman PD. Mucormycosis of the oral cavity. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 1993;75:455-60.
- 73. Peterson KL, Wang M, Canalis RF. Rhinocerebral mucormycosis: Evaluation of the disease and treatment options. Laryngoscope 1997;107:855-61.
- McLean FM, Ginsberg LE, Stanton CA. Perineural spread of rhinocerebral mucormycosis. Am J Neuroradiol 1996;17:114-6.
- Marx RE, Stern D. Oral and maxillofacial pathology; A rationale for diagnosis and treatment. 1<sup>st</sup> ed. Vol 1. United Kingdom. Quintessence Publishing Co. Inc; 2006. p. 104-6.
- Chayalkulkeeree M, Ghannoum MA, Perfect JR. Zygomycosis; The re-emerging fungal infection. Eur J Clin Microbiol Infect Dis 2006;25:215-9.
- Iatta R, Napoli C, Borghi E, Montagna MT. Rare mycoses of the oral cavity; a literature epidemiologic review. Oral Surg Oral Med Oral Pathol Oral Radiol End 2009;108:647-55.
- Cafarchia C, Rmoito D, Iatta R. Role of birds of prey as carriers and spreaders of C. neoformans and other zoonotic yeasts. Med Mycol 2006;44:485-92.
- Rippon JW. Medical Mycology. 3<sup>rd</sup> ed, vol 1. Philadelphia: WB Saunders; 1988.

- Moore M. Cryptococcosis with cutaneous manifestations; four cases with review of published reports. J Invest Dermatol 1957;28:159-82.
- Delgado WA, Leon ER. 14<sup>th</sup> International Congress IAOP/ AAOMP Clinical Pathology Conference Case 6. Head Neck Pathol 2008;2:298-301.
- Cawley EP, Grekin RH, Curtis AC. Torulosis: A review of the cutaneous and adjoining mucous membrane manifestations. J Invest Dermatol 1950;14:327-44.
- Shibuya K, Hirata A, Omuta J, Sugamata M, Katori S, Saito N, et al. Granuloma and Cryptococcosis. J Infect Chemother 2005;11:115-22.
- 84. Samaranayake LP, Leung WK, Jin L. Oral mucosal fungal infections. Periodontology 2000. 2009;49:39-59.
- Pottier I, Gente S, Vernoux JP. Safety assessment of diary microorganisms: Geotrichum candidum. Int J Food Microbiol 2008;126:327-32.
- Hattori H, Inoue C, Tomita Y, Kanbe T. A case of oral Geotrichosis caused by Geotrichum candidum in an old patient. Jpn J Infect Dis 2007;60:300-1.
- Heinic GS, Greenspan D. Oral Geotrichum candidum infection associated with HIV infection, a case report. Oral Surg Oral Med Oral Pathol 1992;73:726-8.
- Samaranayake LP. Oral mycoses in HIV infection. Oral Surg Oral Med Oral Pathol Oral Radiol End 1992;73:171-80.
- Listemann H, Schonrock-Nalbusi P, Kuse R. Geotrichosis of oral mucosa. Mycoses 1996;39:289-91.
- Bonifaz A, Gonzalez DZ, Macias B, Parades-Farrera F, Hernandez MA, Araiza J. Oral geotrichosis: Report of 12 cases. J Oral Sci 2010;52:477-83.
- 91. Bonifaz A, Macias B, Parades-Farrera F, Arias P, Ponce RM, Araiza J. Palatal zygomycosis; Experience of 21 cases. Oral Dis 2008;14:569-74.
- 92. Henrich TJ, Marty FM, Milner DA. Disseminated G. Candidum infection in patient with relapsed acute myelogenous leukemia following allogenic stem cell transplantation with review of literature. Transpl Infect Dis 2009;11:458-62.
- 93. Lanzafame M, De Chechhi G, Parinello A. Rhodotorula glutinis related meningitis. J Clin Microbiol 2001;39:410.
- 94. Available from: http://doctorfungus.org/the fungi rhodotorula- mucilaginosa.htm [Last accessed on 12 Feb 2013].
- De Hoog, Guarro JG. Atlas of clinical fungi. 2<sup>nd</sup> ed., vol 1. Centralbureau voor Schimmelcultures: The Netherlands; 2000.
- Lo Re V III, Fishman NO, Nachamkin I. Recurrent catheter related Rhodotorula rubra infection. Clin Microbiol Infect 2003;9:897-900.
- 97. Pamidimukkala U, Challa S, Lakshmi V. Sepsis and meningoencephalitis due to Rhodotorula glutinis in a patient with systemic lupus erythematosus, diagnosed at autopsy. Neurol India 2007;55:304-7.
- Kaur R, Wadhwa A, Agarwal S. Rhodotorula mucilaginosa; an unusual cause of oral ulcers in AIDS patients. AIDS 2007;21:1068-9.
- Shinde RS, Mantur BG, Patil G, Parande MV, Parande AM. Meningitis due to Rhodotorula lutinis in an HIV infected patient. Indian J Med Microbiol 2008;26:375-97.
- 100. Anaissie EJ, McGinnis MR, Pfaller MA. Clinical mycology. 2<sup>nd</sup> ed., vol 1. Churchill Livingstone; Philadelphia, 2003.
- 101. Kwon- Chung J, Bennett JE. Medical Mycology. 1<sup>st</sup> ed., vol 1. Philadelphia: Lea and Febinger; 1992.
- 102. Wadhwa A, Kaur R, Bhalla P. Saccharomyces cerevisiae as a cause of oral thrush and diarrhoea in an HIV/AIDS patient. Trop Gastroenterol 2010;31:227-9.

- 103. Marteau PR, de Vrese M, Cellier CJ, Schrezenmeir J. Protection from gastrointestinal disease with the use of probiotics. Am J Clin Nutr 2001;73 Suppl 2:430-6S.
- 104. Jensen DP, Smith DL. Fever of unknown origin secondary to brewer's yeast ingestion. Arch Intern Med 1976;136:332-3.
- 105. Nelson PE, Dignani MC, Annaissie EJ. Taxonomy, biology and clinical aspects of Fusarium species. Clin Microbiol Rev 1994;7:479-504.
- 106. Evans J, Levesque D, de LA, Jensen HE. Intracranial fusariosis: A novel cause of fungal meningoencephalitis in a dog. Vet Pathol 2004;41:510-4.
- 107. Nucci M, Annaisie E. Cutaneous infection by Fusarium species in healthy and immunocompromised patients. Clin Infect Dis 2002;35:909-20.
- 108. Boutati EI, Annaissie EJ. Fusarium, a significant emerging pathogen in patients with hematologic malignancy: Ten years' experience at a cancer centre and implications of management. Blood 1997;90:999-1008.
- Walsch TJ, Groll AH. Emerging fungal infections. Transpl Infect Dis 1999;1:247-61.
- 110. Nucci M, Annaisie E. Fusarium Infections in Immunocompromised Patients. Clin Microbiol Rev 2007;20:695-704.
- 111. Huprikar S, Shoham S. Emerging fungal infections in solid organ transplantation. Am J Transplant 2013;13:262-71.
- 112. Myoken Y, Sugata T, Kyo T, Fujihara M. Oral Fusarium infection in a granulocytopenic patient with acute myelogenous leukemia: A case report. J Oral Pathol Med 1995;24:237-40.
- 113. Liu K, Howell DN, Perfect JR, Schell WA. Morphologic criteria for the preliminary identification of Fusarium, Paecilomyces and Acremonium species by histopathology. Am J Clin Pathol 1998;109:45-54.
- 114. Hayden RT, Isotalo PA, Parrett T, Wolk DM, Qian X, Roberts GD, et al. In situ hybridization for the differentiation of Aspergillus, Fusarium and Pseudallescheria species in tissue sections. Diagn Mol Pathol 2003;12:21-6.
- 115. Healy M, Reece K, Walton D, Huong J, Frye S, Raad II. Use of Diversi Lab system for species and strain differentiation of Fusarium species isolates. J Clin Microbiol 2005;43:5278-80.

- Dahiya P. Penicilliosis in an HIV positive individual. Indian J Sex Trans Dis AIDS 2012;33:38-40.
- 117. Segretain G. Penicillium marneffei agent of mycosis of reticuloendothelial system. Mycopathologia 1959;11:327-53.
- 118. Tlamcani Z, Er-rami M. Fungal opportunistic infection: Common and emerging fungi in immunocompromised patients. J Immunol Tech Infect Dis 2013;2:1-5.
- 119. Sapparatpinyo K, Khamwan C, Baosoung V, Nelson KE, Sirisanthana T. Disseminated Penicillosis in Southeast Asia. Lancet 1994;344:110-3.
- 120. Wu TC, Chan JW, Ng CK, Tsang DN, Lee MP, Li PC. Clinical presentations and outcomes of Penicillium marneffei infections: A series from 1994 to 2004. Hong Kong Med J 2008;14:103-9.
- 121. Antinori S, Gianelli E, Bonaccorso C, Ridolfo AL, Croce F, Sollima S, *et al.* Disseminated Penicillium marneffei infection in HIV positive Italian patient and review of cases reported outside epidemic regions. J Travel Med 2006;13:181-8.
- Chitasombat M, Sapparatpinyo K. Penicillium marneffei infection in immunocompromised host. Curr Fungal Infect Rep 2013;7:44-50.
- 123. Cooper CR Jr, McGinnis MR. Pathology of Penicillium marneffei; an emerging acquired immunodeficiency syndrome- related pathogen. Arch Pathol Lab Med 1997;89:798-804.
- 124. Deesomchok A, Tanprawate S. A 12 case series of Penicillium marneffei pneumonia. J Med Assoc Thailand 2006;89:441-7.
- 125. Nittayanantha W. Penicilliosis marneffei another AIDS defining illness in Southeast Asia. Oral Dis 1999;5:286-93.
- 126. Wong SY, Wong KF. Penicillium marneffei infection in AIDS. Pathol Res Int Volume 2011;2011:764293.
- 127. Wong KF. Marrow penicilliosis, a readily missed diagnosis. Am J Clin Pathol 2010;134:214-8.
- 128. Ramana KV, Kandi S, Bharatkumar PV, Sharada CH, Rao R, Mani R, *et al.* Invasive fungal infections: A comprehensive review. Am J Infect Dis Microbiol 2013;1:64-9.

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