

# Healthspan Extension through Innovative Genetic Medicines

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**Summary:** Genetic medicine has emerged as an innovative class of therapeutics, allowing the development of new and powerful approaches to address a myriad of diseases that were previously untreatable. At the same time, our improved understanding of the mechanisms underlying aging has created novel opportunities to intervene therapeutically in the aging process itself through the targeting of key pathways driving this process. As individuals age, the onset of a multitude of age-related diseases can significantly impact lifespan. The ultimate goal of their treatment is the maximization of healthy, disease-free years, or healthspan. Here, we discuss a number of promising genetic medicine approaches to target both general and specific mechanisms of age-related disease, and their potential impact on healthspan extension. Essential to this topic is the challenge of nucleic acid delivery, and we discuss the technologies that have been developed to address this challenge in highly promising preclinical and clinical development efforts. In particular, we describe a next-generation delivery technology for healthspan applications called proteo-lipid vehicles. (*Plast. Reconstr. Surg.* 150: 49S, 2022.)

From a regulatory perspective, genetic medicine covers a broad area, comprising the manipulation of both genetic code and gene expression via the exogenous introduction of nucleic acids into a biological system. The first gene therapy clinical trial occurred in 1989, and it took 14 years for one to be approved. However, in the time since, several cell and gene therapies have been approved for clinical use, and at a much faster pace.<sup>1-3</sup> The first gene therapy approved in the United States entailed the ex vivo manipulation of patient cells, but recent improvements to delivery methods have enabled the use of genetic therapies that are administered in vivo, directly to patients. Notable examples are the latest genetic vaccines designed to immunize against COVID-19. These have set new benchmarks for speed and efficacy with respect to the design and deployment of commercial genetic medicines. These recent

successes have ushered in a new era of treatments that can sense and modify genetic pathways with a precision never before possible using small molecules and biologics, which rely on chemistry for their targeting and mechanism of action. The question has now become how to best utilize these new tools, especially for something as complex and multifactorial as healthspan.

Healthspan is commonly defined as the period of time in which an organism is relatively healthy and free from disease. Although this is not a strictly defined phenotype, maximizing healthspan remains a general goal to achieve for the longevity field.<sup>4</sup> From practical and regulatory standpoints, single age-related disease indications and quantifiable metrics are the most practical ways to assess the efficacy of interventions, with potential consequences for healthspan. An effective therapy targets the core aspects of aging that contribute to age-related disease to varying degrees.<sup>5</sup> For example, senolytics, a class of therapies that selectively eliminate senescent cells, has the potential to be beneficial in multiple age-related diseases.<sup>6-8</sup> In this mini-review, we will expand upon this class

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of therapies, provide an overview of the genetic medicine payloads that are being developed for healthspan extension, and discuss a major challenge of genetic medicines: payload delivery.

### SENOLYTICS

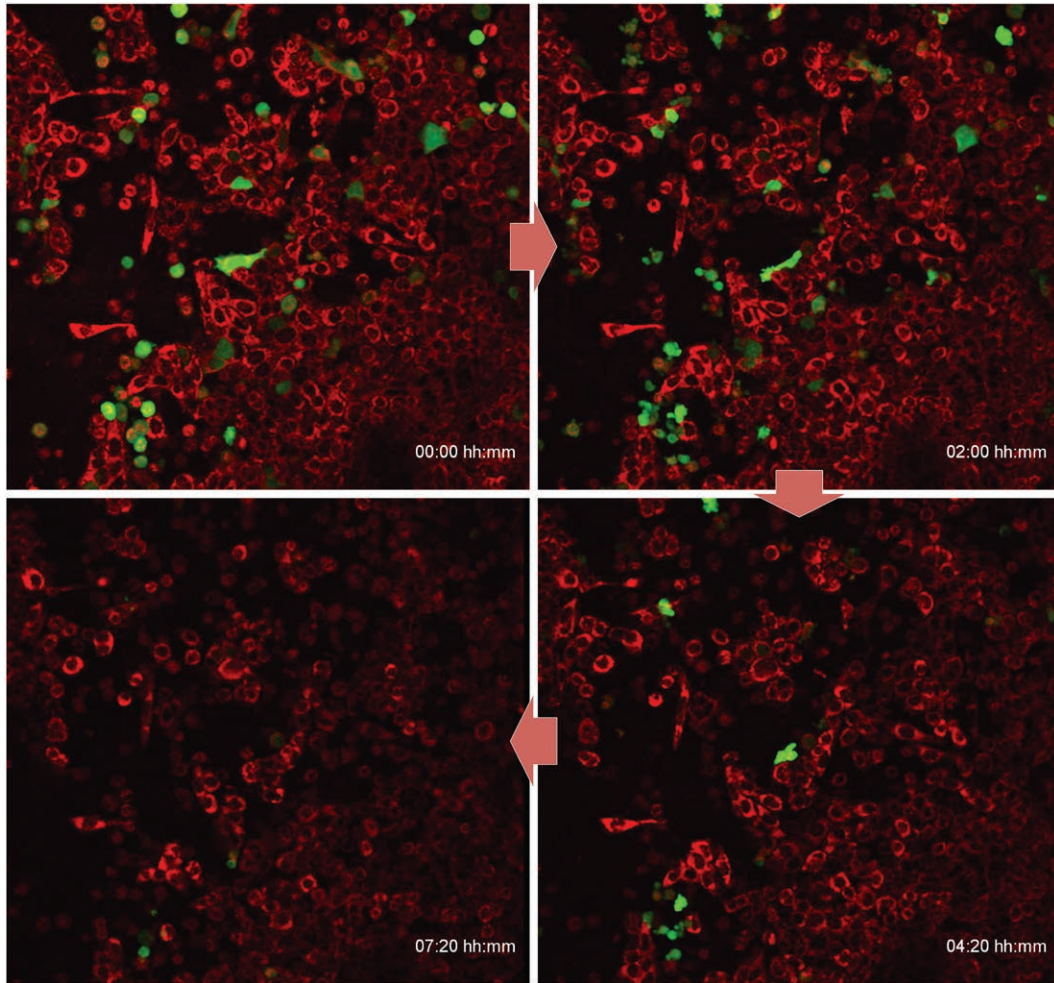
Senescence is a cell fate characterized by stable cell cycle arrest and secretion of proinflammatory cytokines and molecules known as the senescence-associated secretory phenotype (SASP). The SASP is comprised of cytokines, chemokines, proteases, and growth factors, which impact surrounding healthy cells and alter their function.<sup>9</sup> Senescent cells accumulate over time and contribute to a decline in organ function, implicating them in most age-related diseases. The seminal proof-of-concept that showed the removal of senescent cells *in vivo* can rejuvenate an organism was first shown in 2011. Since then, a rapid expansion of research in senolytic therapies have led to a number of clinical trials.<sup>7,10,11</sup> Many small molecule and biologic approaches have been used academically and clinically with varying degrees of success. Despite their broad effect, senescent cells only constitute a small percentage of cells in the body. Senescent cells themselves are also heterogeneous with features that closely resemble surrounding cells.<sup>12–14</sup> These challenges limit the utility of small molecule drugs due to their indiscriminate nature and resultant systemic toxicity, since many are based on drug repurposing. This is especially true in sensitive tissues like skin, where senescent cells contribute to health and resultant appearance.<sup>15–20</sup> Attempts have been made to target localized compartments to circumvent this, but so far this strategy has failed to achieve its purpose, illustrated by the failed trial for osteoarthritis using intra-knee injections of UBX0101.<sup>21,22</sup>

In contrast, genetic approaches provide a method to selectively target senescence-associated features, improving efficacy and limiting toxicity. Indeed, the first proof-of-concept for the beneficial effect of senolysis was demonstrated in transgenic mice where an inducible suicide gene was selectively activated in p16<sup>+</sup> senescent cells.<sup>10,23</sup> The selectivity and control of a genetic medicine approach enables the precise elimination of senescent cells and is a primary focus of the authors.<sup>24</sup> This novel senolytic approach utilizes a plasmid DNA construct encoding a late-stage apoptotic protein under the control of a senescence-associated promoter. Apoptosis induction is further controlled by activation via addition of a dimerizing agent (Fig. 1). Senescent cells contribute to the

onset and progression of age-related diseases, and a senolytic that can be safely used by healthy aged individuals for age-related disease prevention will have a broad impact. The ability of gene therapy to target-specific subtypes of cells makes it an ideal type of therapeutic to treat chronic and complex age-related diseases. For example, while senescent cells are important for wound healing, persistence of senescent cells leads to chronic wounds, which necessitates nonpersistent sporadic senescent clearance. Due to the complexity of wound healing, chronic wounds can be challenging for clinicians, especially in elderly patients, and senolytics offer another arrow in the quiver of therapeutic options.<sup>16,25–28</sup>

### DURABLE GENE REPLACEMENT THERAPIES FOR IMPROVING HEALTHSPAN

Age-related diseases are typically chronic, and it is reasonable to assume that their treatment will require prolonged or repeat dosing. Although the half-lives of small molecules and many biologics are relatively short, gene therapies are unique in that they can be engineered to persist indefinitely. The first approved gene therapy in the United States, Kymriah, introduced a gene for a chimeric antigen receptor on a patient's T cells (CAR-T) *ex vivo* before reinfusing them to treat their acute lymphoblastic leukemia. Since then, CAR-T research has expanded to multiple hematological malignancies, solid tumors, and more recently to target senescent cells based on a surface cell marker.<sup>29–31</sup> CAR-T cells are designed to persist indefinitely within the patient to exert their protective effects, so great caution must be taken with a persistent senescent cell clearance therapy.<sup>23</sup> Treatment of monogenic diseases via gene replacement therapies is an excellent example of the durable effect afforded by genetic medicines.<sup>32</sup> In addition to the approved gene replacement therapies, efforts are being made to understand the genes that decrease with either age or age-related diseases, and several have been identified that demonstrate protective antiaging effects and healthspan benefits. For example, a gene therapy is being developed to increase the expression of the DNA-repair enzyme, SIRT6.<sup>33</sup> There are multiple groups developing therapies to extend telomeres via the expression of telomerase.<sup>34–37</sup> A triple gene therapy approach to treat mitral valve disease is being developed for canines, with an aim to eventually progress to humans.<sup>38</sup> A therapy based on work from the University of South



**Fig. 1.** Specific elimination of target cells using senolytic gene therapy. Time-lapse microscopy of target cells expressing the senescence-associated gene p16 (green) in a field of normal cells stained with MitoTracker (red). Cells were treated with PLVs encoding an inducible caspase 9 tagged with GFP under the control of the p16 promoter. Addition of a dimerizing agent to activate caspase 9 resulted in all target cells being eliminated within 8 hours.

Alabama is introducing a gene in macrophages *ex vivo* that can metabolize oxidized cholesterol.<sup>39</sup> Efforts to treat epidermolysis bullosa have had dramatic results from two groups using either transgenic *ex vivo* keratinocytes or intradermal injection of collagen VII.<sup>40,41</sup> Using their genetic medicine platform, the authors are developing multiple gene replacement therapies to treat sarcopenia, mitochondrial diseases, collagen defects, and skin disease. Attempting to recapitulate the effects of gene replacement therapies via recombinant protein therapy is prohibitively expensive and often precluded by delivery challenges and complicated manufacturing.

The discovery of the CRISPR/Cas9 gene editing system to fix single genetic mutations rather than replace the gene outright has dramatically broadened the potential applications of genetic

medicines.<sup>42–44</sup> There are numerous age-related diseases driven by point mutations that could be treated via CRISPR/Cas9-based gene therapies, such as ALS, Alzheimer disease, Parkinson disease, and rheumatoid arthritis, and more as CRISPR-based research progresses.<sup>45</sup> However, although these tools hold great promise to correct genetic diseases, *in vivo* delivery of these tools to the cells of interest remains a significant challenge.

### GENETIC APPROACHES CAN BE TUNED FOR SPATIOTEMPORAL SPECIFICITY

Given the highly regulated spatial and temporal nature of gene expression, genetic medicines must be designed to express exogenous genes in the right cell population at the right time. Engineering highly selective DNA promoters and

enhancer/repressor combinations allows for a gene therapy to deliver throughout the body and still achieve precise cell and tissue.

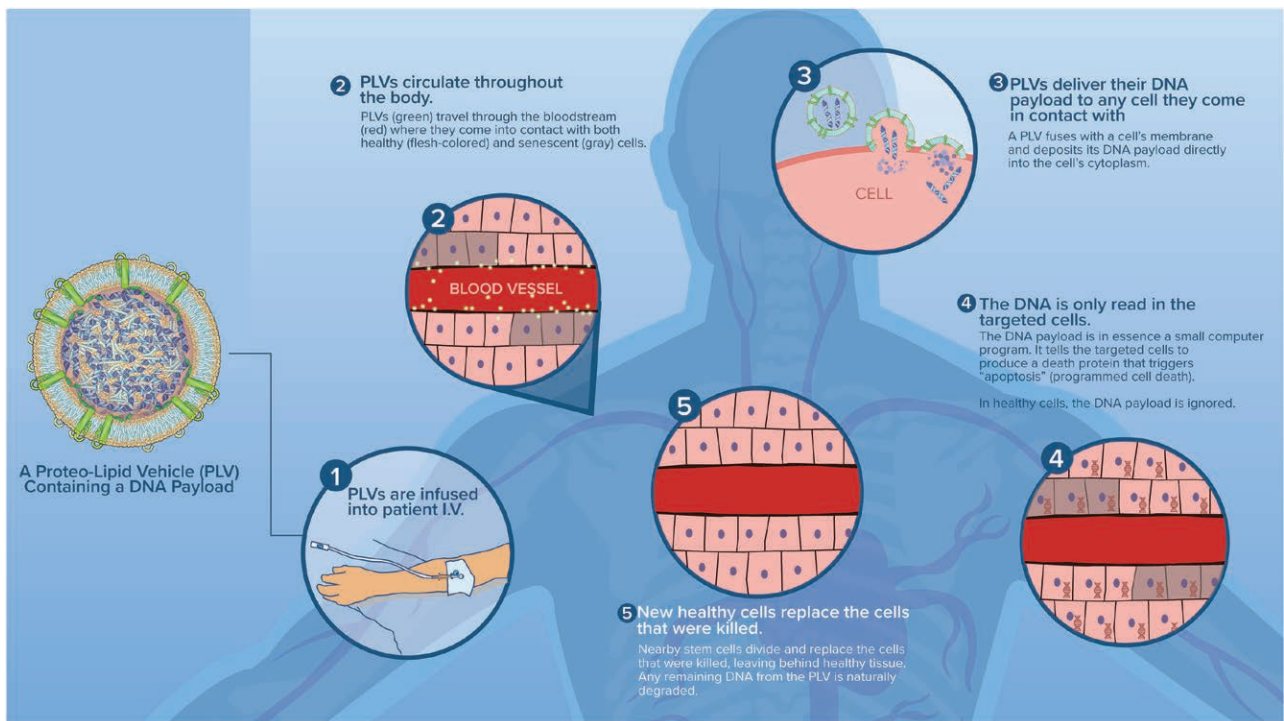
Temporal specificity is also important for gene therapies affecting healthspan. The type of payload can also affect temporal specificity, such as mRNA, which provides rapid and robust yet transient cargo expression. However, in some indications, this approach is desirable. For example, there are multiple efforts to utilize Yamanaka factors to partially reprogram cells into a more youthful state.<sup>46–50</sup> With their senolytic approach, the authors are eliminating senescent cells in a similar “hit-and-run” approach that can be delivered body-wide repeatedly at regular intervals, further reducing toxicity and effectively re-creating what has been shown by other studies involving transgenic animals (Fig. 2).<sup>10</sup>

### THE CHALLENGE OF NUCLEIC ACID DELIVERY

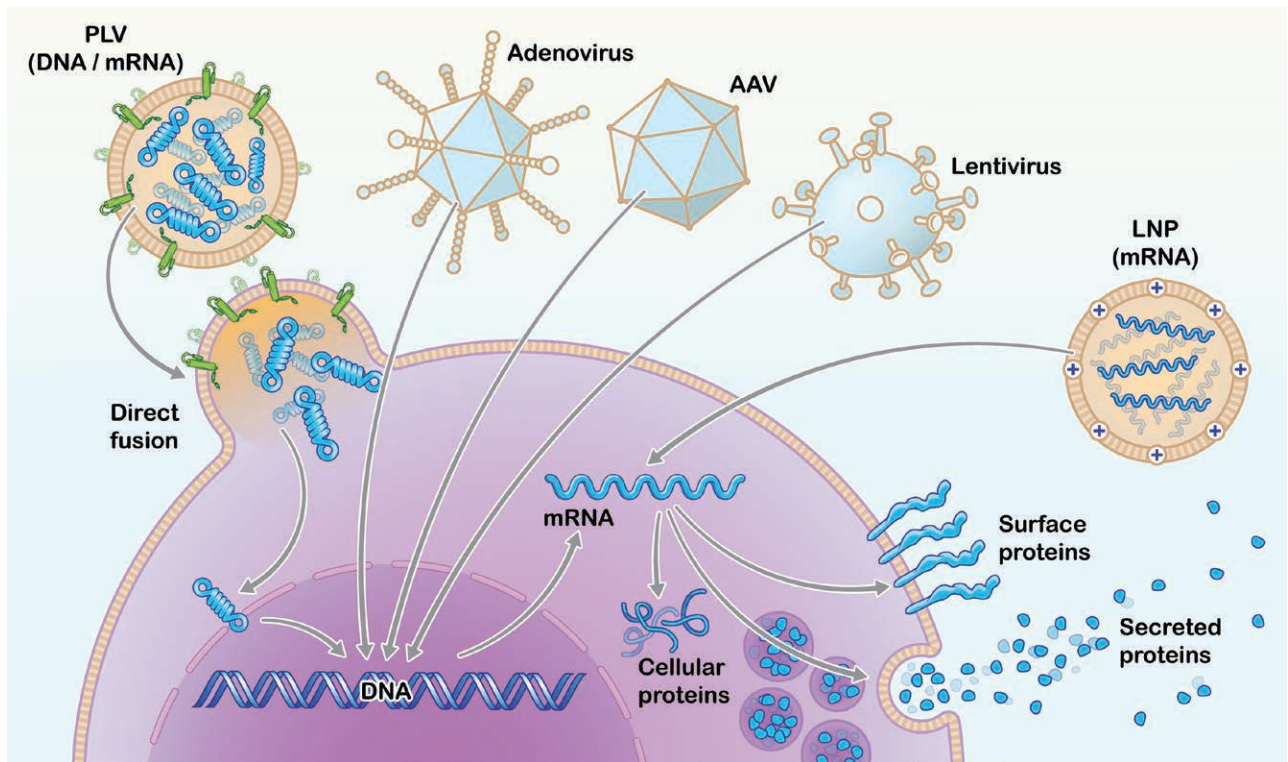
There are some significant hurdles that genetic medicines must overcome to gain more broad applicability. The plasma membrane is a highly effective physical barrier that actively repels or sequesters exogenous macromolecules such as negatively charged nucleic acids. This necessitates the use of a carrier vehicle to facilitate entry

into the cytosol to exert the intended therapeutic effect (Fig. 3). Viral vectors have dominated gene therapy efforts due to their high efficiency of gene expression, with adenoviral, retroviral/lentiviral, and adeno-associated viral (AAV) vectors leading the field.<sup>1,51</sup> The two subtypes of retroviruses,  $\gamma$ -retroviruses, and lentiviruses contain an RNA genome that undergoes reverse transcription into DNA upon entry into a transduced cell. The new DNA genome is then integrated into the host cell therefore enabling durable gene expression.<sup>52</sup> Retroviruses have demonstrated their clinical success via ex vivo gene transfer into hematopoietic stem cells (HSCs)<sup>53,54</sup> and T cells.<sup>55,56</sup> Genetically altered cells can be reintroduced into patients and reverse deficiencies in HSCs, or target cancer cells. The risk of insertional mutagenesis has hindered the systemic use of retroviruses<sup>52</sup> and further testing and validation will be required before this platform can be adopted to improve health span.

Adenoviral and AAV vectors have predominately been the chosen viral vectors for systemic gene therapy. These vectors have a DNA genome and possesses an ability to transduce quiescent cells, making them an ideal candidate for the treatment of monogenic diseases and potentially aging.<sup>57</sup> However, initial studies examining the use of adenoviruses as gene therapy vectors were



**Fig. 2.** Oisín Biotechnologies clinical approach. Schematic detailing the mechanism of action of a senolytic PLV infusion.



**Fig. 3.** Gene delivery technologies. Illustration of gene therapy delivery modalities. Viral-based delivery methods have efficient transduction but are immunogenic. Nonviral-based LNPs are relatively nonimmunogenic but have inefficient transduction due to endocytosis and utilize lipids that are toxic. PLVs utilize nontoxic lipids and a nonimmunogenic fusogen (*green*) that directly fuses the PLV with the cell membrane, enabling nonimmunogenic, nontoxic, and efficient transduction.

met with resistance after it was reported that serious and life-threatening side effects could occur,<sup>58</sup> causing a shift to the use of safer AAV vectors.<sup>59,60</sup> Sustained gene expression from a single dose is often required for therapeutic effect, as adaptive immune responses prevent AAV vectors from being administered repeatedly.<sup>60,61</sup> Additionally, preexisting neutralizing AAV antibodies can exist even without prior vector exposure further hindering gene expression.<sup>61–64</sup> Although immune stimulation be advantageous in the field of vaccines, where robust T-cell responses induced by the gene delivery vector can help facilitate long-lasting immunity to the antigen, this immunogenicity hinders repeat dosing.<sup>65</sup>

To overcome limitations surrounding viral vectors, the focus in recent years has largely shifted to the development and optimization of the safer nonviral delivery vectors.<sup>51</sup> Nonviral delivery vectors are easier and cheaper to manufacture than viral vectors, allowing for rational design and large-scale testing of novel materials and formulations with nucleic acid delivery potential.<sup>66,67</sup> Of the various nonviral delivery vectors, lipid nanoparticles (LNPs) have demonstrated

their success with the FDA approval of patisiran (Onpattro), an LNP-based RNA interference agent that treats amyloidosis caused by abnormal transthyretin production.<sup>68,69</sup> LNPs are formulated with cationic or ionizable lipids that by neutralizing the anionic charge of nucleic acids enable transport of nucleic acids across the charge-restrictive plasma membrane via endocytosis.<sup>70</sup> Despite the immunogenicity benefits LNP vectors have over viral vectors, LNPs containing cationic or ionizable lipids can stimulate toxic and inflammatory responses, limiting their usefulness clinically. At a cellular level, LNPs facilitate apoptotic cell death that translates to liver toxicity following systemic delivery.<sup>71,72</sup> Interactions between LNPs and the immune system can have systemic effects and lead to secretion of proinflammatory cytokines like tumor necrosis factor alpha (TNF- $\alpha$ ), interferon-gamma (IFN- $\gamma$ ), and interleukin-6 (IL-6).<sup>73,74</sup> Furthermore, LNPs can stimulate complement activation-related pseudoallergy (CARPA), a hypersensitivity reaction resulting in death in severe circumstances.<sup>75–77</sup>

The authors have taken an approach that combines the strengths and mitigates the weaknesses

of viral and nonviral delivery vectors by developing proteo-lipid vehicles (PLVs), a platform that incorporates a fusion-associated small transmembrane (FAST) protein into a lipid-based formulation.<sup>78–101</sup> FAST proteins are the smallest viral fusogens that have been described ( $\approx 100$ – $200$  residues), with a single transmembrane domain fixing the FAST protein in a <sup>N</sup>exoplasmic/<sup>C</sup>cytoplasmic type I membrane topology that exposes an even smaller ectodomain ( $\approx 20$ – $40$  residues).<sup>86</sup> FAST proteins function as fusion machines, where incorporation into a lipid-based nanoparticle is sufficient to facilitate nanoparticle-cell fusion and delivery of encapsulated cargo directly into the cytosol of target cells.<sup>87</sup>

This enables FAST protein-containing PLVs to be formulated with reduced ionizable and cationic lipids, substantially improving safety while retaining efficacy, which allows for genetic medicines that require high systemic doses. Furthermore, FAST proteins are nonimmunogenic enabling repeat dosing. Notably unlike other LNP platforms, FAST-PLVs have demonstrated the ability to successfully deliver plasmid DNA systemically after intravenous administration, similar to viral vectors.<sup>85</sup>

## DISCUSSION

Genetic medicines are rapidly becoming a distinct and revolutionary class of therapeutics. The durability, specificity, and ease of manufacturing allows for a plethora of creative and effective strategies. The COVID-19 pandemic has shown the world that a gene therapy can be rapidly developed and deployed globally. This has already begun to catalyze the interest and capital that will be leveraged toward gene therapies going forward. Much like LNPs, PLVs are relatively inexpensive to manufacture at scale compared to viral-based delivery methods, which enables the development of cost-effective therapeutics that can be more broadly applied.

Although there are still considerable efforts involved in designing genetic medicines, the ability to progress from target to preclinical testing in a matter of months is typically shorter than the development that many small molecules go through before preclinical in vivo testing. As a result, researchers developing genetic medicines can take a more iterative approach and de-risk their clinical efforts.

Next-generation gene delivery platforms such as PLVs enables the development of more sophisticated genetic medicine approaches

utilizing DNA or self-amplifying RNA. This allows for innovative gene therapies that can be exquisitely regulated. For example, a genetic program that expresses a gene of interest X exclusively in keratinocytes but is repressed when that keratinocyte expresses another gene Y is a kind of logic gate that is possible utilizing DNA. There is enormous potential in the gene therapy space, which can synergize with and supplement surgical efforts. Platform technologies like PLV are also likely essential for the treatment of chronic diseases of aging as individuals will require systemic treatments over the course of many years. We are focusing our initial efforts on the development of a first-in-class senolytic gene therapy while engineering innovative solutions to ameliorate age-related diseases and promote healthspan extension using multiple complementary approaches. The future of gene therapy in the context of healthspan could be seen as an accumulation of genetic programs that prevent or correct the normal trajectory of age-related disease and could be taken acutely and/or repeatedly as the demand arises.

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## REFERENCES

1. Ginn SL, Amaya AK, Alexander IE, et al. Gene therapy clinical trials worldwide to 2017: An update. *J Gene Med* 2018;20:e3015.
2. Food and Drug Administration. Approved cellular and gene therapy products. Available at <https://www.fda.gov/vaccines-blood-biologics/cellular-gene-therapy-products/approved-cellular-and-gene-therapy-products>. 2021.
3. Daley, J. Gene therapy arrives. *Nature*. 2019;576, S12–S13.
4. Kaerberlein M. How healthy is the healthspan concept? *Geroscience*. 2018;40:361–364.
5. López-Otín C, Blasco MA, Partridge L, et al. The hallmarks of aging. *Cell*. 2013;153:1194–1217.
6. Amaya-Montoya M, Pérez-Londoño A, Guatibonza-García V, et al. Cellular senescence as a therapeutic target for age-related diseases: A review. *Adv Ther*. 2020;37:1407–1424.
7. Borghesan M, Hoogaars WMH, Varela-Eirin M, et al. A senescence-centric view of aging: Implications for longevity and disease. *Trends Cell Biol*. 2020;30:777–791.

8. Tchkonja T, Zhu Y, van Deursen J, et al. Cellular senescence and the senescent secretory phenotype: Therapeutic opportunities. *J Clin Invest*. 2013;123:966–972.
9. Campisi J. Cellular senescence: Putting the paradoxes in perspective. *Curr Opin Genet Dev*. 2011;21:107–112.
10. Baker DJ, Wijshake T, Tchkonja T, et al. Clearance of p16Ink4a-positive senescent cells delays ageing-associated disorders. *Nature*. 2011;479:232–236.
11. McHugh D, Gil J. Senescence and aging: Causes, consequences, and therapeutic avenues. *J Cell Biol*. 2018;217:65–77.
12. Wiley CD, Flynn JM, Morrissey C, et al. Analysis of individual cells identifies cell-to-cell variability following induction of cellular senescence. *Aging Cell*. 2017;16:1043–1050.
13. Hernandez-Segura A, de Jong TV, Melov S, et al. Unmasking transcriptional heterogeneity in senescent cells. *Curr Biol*. 2017;27:2652–2660.e4.
14. Hernandez-Segura A, Nehme J, Demaria M. Hallmarks of cellular senescence. *Trends Cell Biol*. 2018;28:436–453.
15. Waaijer ME, Gunn DA, Adams PD, et al. P16INK4a positive cells in human skin are indicative of local elastic fiber morphology, facial wrinkling, and perceived age. *J Gerontol A Biol Sci Med Sci*. 2016;71:1022–1028.
16. Velarde MC, Demaria M. Targeting senescent cells: possible implications for delaying skin aging: A mini-review. *Gerontology*. 2016;62:513–518.
17. Demaria M, Desprez PY, Campisi J, et al. Cell autonomous and non-autonomous effects of senescent cells in the skin. *J Invest Dermatol*. 2015;135:1722–1726.
18. Lowry WE. Its written all over your face: The molecular and physiological consequences of aging skin. *Mech Ageing Dev*. 2020;190:111315.
19. Toutfaire M, Bauwens E, Debacq-Chainiaux F. The impact of cellular senescence in skin ageing: A notion of mosaic and therapeutic strategies. *Biochem Pharmacol*. 2017;142:1–12.
20. Bonté F, Girard D, Archambault JC, et al. Skin changes during ageing. *Subcell Biochem*. 2019;91:249–280.
21. Globenewswire.com. Unitybiotechnologyannounces 12-week data from UBX0101 phase 2 clinical study in patients with painful osteoarthritis of the knee. Available at <https://www.globenewswire.com/news-release/2020/08/17/2079116/0/en/UNITY-Biotechnology-Announces-12-week-data-from-UBX0101-Phase-2-Clinical-Study-in-Patients-with-Painful-Osteoarthritis-of-the-Knee.html>. 2020.
22. Unity Biotechnology. A study to assess the safety and efficacy of a single dose of UBX0101 in patients with osteoarthritis of the knee. Available at <https://clinicaltrials.gov/ct2/show/NCT04129944>. 2019.
23. Demaria M, Ohtani N, Youssef SA, et al. An essential role for senescent cells in optimal wound healing through secretion of PDGF-AA. *Dev Cell*. 2014;31:722–733.
24. Oisin Biotechnologies. Available at: <https://www.oisinfo.com/>. Accessed August 30, 2022.
25. Blair MJ, Jones JD, Woessner AE, et al. Skin structure-function relationships and the wound healing response to intrinsic aging. *Adv Wound Care (New Rochelle)*. 2020;9:127–143.
26. Wang Z, Shi C. Cellular senescence is a promising target for chronic wounds: A comprehensive review. *Burns Trauma*. 2020;8:tkaa021.
27. Eming SA, Martin P, Tomic-Canic M. Wound repair and regeneration: Mechanisms, signaling, and translation. *Sci Transl Med*. 2014;6:265sr6.
28. Wei X, Li M, Zheng Z, et al. Senescence in chronic wounds and potential targeted therapies. *Burns Trauma* 2022;10:tkab045.
29. Hong M, Clubb JD, Chen YY. Engineering CAR-T cells for next-generation cancer therapy. *Cancer Cell*. 2020;38:473–488.
30. Holstein SA, Lunning MA. CAR T-cell therapy in hematologic malignancies: A voyage in progress. *Clin Pharmacol Ther*. 2020;107:112–122.
31. Amor C, Feucht J, Leibold J, et al. Senolytic CAR T cells reverse senescence-associated pathologies. *Nature*. 2020;583:127–132.
32. Day JW, Finkel RS, Chiriboga CA, et al. Onasemnogene abeparvovec gene therapy for symptomatic infantile-onset spinal muscular atrophy in patients with two copies of SMN2 (STRIVE): An open-label, single-arm, multicentre, phase 3 trial. *Lancet Neurol*. 2021;20:284–293.
33. Roichman A, Kanfi Y, Glazz R, et al. SIRT6 overexpression improves various aspects of mouse healthspan. *J Gerontol A Biol Sci Med Sci*. 2017;72:603–615.
34. Bernardes de Jesus B, Vera E, Schneeberger K, Tejera AM, et al. Telomerase gene therapy in adult and old mice delays aging and increases longevity without increasing cancer. *EMBO Mol Med*. 2012;4:691–704.
35. Ramunas J, Yakubov E, Brady JJ, et al. Transient delivery of modified mRNA encoding TERT rapidly extends telomeres in human cells. *FASEB J*. 2015;29:1930–1939.
36. Muñoz-Lorente MA, Martínez P, Tejera Á, et al. AAV9-mediated telomerase activation does not accelerate tumorigenesis in the context of oncogenic K-Ras-induced lung cancer. *PLoS Genetics*. 2018;14:e1007562.
37. Fossel M. Cell senescence, telomerase, and senolytic therapy. *OBM Geriatrics*. 2018;3:034.
38. Davidsohn N, Pezone M, Vernet A, et al. A single combination gene therapy treats multiple age-related diseases. *Proc Natl Acad Sci U S A*. 2019;116:23505–23511.
39. D'Arcy BM, Swingle MR, Schambeau L, Pannell L, Prakash A, Honkanen RE. Development of a synthetic 3-ketosteroid  $\delta$  1 -dehydrogenase for the generation of a novel catabolic pathway enabling cholesterol degradation in human cells. *Sci Rep*. 2019;9:5969.
40. Hirsch T, Rothoef T, Teig N, et al. Regeneration of the entire human epidermis using transgenic stem cells. *Nature*. 2017;551:327–332.
41. Marinkovich MP, Tang JY. Gene therapy for epidermolysis bullosa. *J Invest Dermatol*. 2019;139:1221–1226.
42. Jinek M, Chylinski K, Fonfara I, et al. A programmable dual-RNA-guided DNA endonuclease in adaptive bacterial immunity. *Science*. 2012;337:816–821.
43. Lino CA, Harper JC, Carney JP, et al. Delivering CRISPR: A review of the challenges and approaches. *Drug Deliv*. 2018;25:1234–1257.
44. Li Q, Qin Z, Wang Q, et al. Applications of genome editing technology in animal disease modeling and gene therapy. *Comput Struct Biotechnol J*. 2019;17:689–698.
45. Caobi A, Dutta RK, Garbinski LD, et al. The impact of CRISPR-Cas9 on age-related disorders: From pathology to therapy. *Aging Dis*. 2020;11:895–915.
46. Sarkar TJ, Quarta M, Mukherjee S, et al. Transient non-integrative expression of nuclear reprogramming factors promotes multifaceted amelioration of aging in human cells. *Nat Commun*. 2020;11:1545.
47. Ocampo A, Reddy P, Belmonte JCI. Anti-aging strategies based on cellular reprogramming. *Trends Mol Med*. 2016;22:725–738.
48. Ocampo A, Reddy P, Martinez-Redondo P, et al. In Vivo amelioration of age-associated hallmarks by partial reprogramming. *Cell*. 2016;167:1719–1733.e12.
49. Wang C, Rabadan Ros, R Martinez-Redondo P, et al. In vivo partial reprogramming of myofibers promotes muscle regeneration by remodeling the stem cell niche. *Nature Commun*. 2021;12:3094.

50. Takahashi K, Yamanaka S. Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell*. 2006;126:663–676.
51. Yin H, Kanasty RL, Eltoukhy AA, et al. Non-viral vectors for gene-based therapy. *Nat Rev Genet*. 2014;15:541–555.
52. Milone MC, O’Doherty U. Clinical use of lentiviral vectors. *Leukemia*. 2018;32:1529–1541.
53. Ferrua F, Cicalese MP, Galimberti S, et al. Lentiviral haemopoietic stem/progenitor cell gene therapy for treatment of Wiskott-Aldrich syndrome: Interim results of a non-randomised, open-label, phase 1/2 clinical study. *Lancet Haematol*. 2019;6:e239–e253.
54. De Ravin SS, Wu X, Moir S, et al. Lentiviral hematopoietic stem cell gene therapy for X-linked severe combined immunodeficiency. *Sci Transl Med*. 2016;8:335ra57.
55. Maude SL, Frey N, Shaw PA, et al. Chimeric antigen receptor T cells for sustained remissions in leukemia. *N Engl J Med*. 2014;371:1507–1517.
56. Roex G, Timmers M, Wouters K, et al. Safety and clinical efficacy of BCMA CAR-T-cell therapy in multiple myeloma. *J Hematol Oncol*. 2020;13:164.
57. Bulcha JT, Wang Y, Ma H, et al. Viral vector platforms within the gene therapy landscape. *Signal Transduct Target Ther*. 2021;6:53.
58. Raper SE, Chirmule N, Lee FS, et al. Fatal systemic inflammatory response syndrome in a ornithine transcarbamylase deficient patient following adenoviral gene transfer. *Mol Genet Metab*. 2003;80:148–158.
59. Thomas CE, Ehrhardt A, Kay MA. Progress and problems with the use of viral vectors for gene therapy. *Nat Rev Genet*. 2003;4:346–358.
60. Wang D, Tai PWL, Gao G. Adeno-associated virus vector as a platform for gene therapy delivery. *Nat Rev Drug Discov*. 2019;18:358–378.
61. Masat E, Pavani G, Mingozzi F. Humoral immunity to AAV vectors in gene therapy: Challenges and potential solutions. *Discov Med*. 2013;15:379–389.
62. Boutin S, Monteilhet V, Veron P, et al. Prevalence of serum IgG and neutralizing factors against adeno-associated virus (AAV) types 1, 2, 5, 6, 8, and 9 in the healthy population: Implications for gene therapy using AAV vectors. *Hum Gene Ther*. 2010;21:704–712.
63. Nayak S, Herzog RW. Progress and prospects: immune responses to viral vectors. *Gene Ther*. 2010;17:295–304.
64. Ferreira V, Twisk J, Kwinkkers K, et al. Immune responses to intramuscular administration of alipogene tiparvovec (AAV1-LPL(S447X)) in a phase II clinical trial of lipoprotein lipase deficiency gene therapy. *Hum Gene Ther*. 2014;25:180–188.
65. Taxis N, Ertl HC. Adenoviruses as vaccine vectors. *Mol Ther*. 2004;10:616–629.
66. Miao L, Li L, Huang Y, et al. Delivery of mRNA vaccines with heterocyclic lipids increases anti-tumor efficacy by STING-mediated immune cell activation. *Nat Biotechnol*. 2019;37:1174–1185.
67. Kauffman KJ, Dorkin JR, Yang JH, et al. Optimization of lipid nanoparticle formulations for mRNA delivery in vivo with fractional factorial and definitive screening designs. *Nano Lett*. 2015;15:7300–7306.
68. Cullis PR, Hope MJ. Lipid nanoparticle systems for enabling gene therapies. *Mol Ther*. 2017;25:1467–1475.
69. Akinc A, Maier MA, Manoharan M, et al. The Onpatro story and the clinical translation of nanomedicines containing nucleic acid-based drugs. *Nat Nanotechnol*. 2019;14:1084–1087.
70. Wan C, Allen TM, Cullis PR. Lipid nanoparticle delivery systems for siRNA-based therapeutics. *Drug Deliv Transl Res*. 2014;4:74–83.
71. Xue HY, Liu S, Wong HL. Nanotoxicity: a key obstacle to clinical translation of siRNA-based nanomedicine. *Nanomedicine (Lond)*. 2014;9:295–312.
72. Sedic M, Senn JJ, Lynn A, et al. Safety evaluation of lipid nanoparticle-formulated modified mRNA in the Sprague-Dawley rat and cynomolgus monkey. *Vet Pathol*. 2018;55:341–354.
73. Zolnik BS, González-Fernández A, Sadrieh N, et al. Nanoparticles and the immune system. *Endocrinology*. 2010;151:458–465.
74. Zhang JS, Liu F, Huang L. Implications of pharmacokinetic behavior of lipoplex for its inflammatory toxicity. *Adv Drug Deliv Rev*. 2005;57:689–698.
75. Gonçalves GAR, Paiva RMA. Gene therapy: advances, challenges and perspectives. *Einstein (Sao Paulo)*. 2017;15:369–375.
76. Szebeni J, Bedocs P, Rozsnyay Z, et al. Liposome-induced complement activation and related cardiopulmonary distress in pigs: Factors promoting reactogenicity of Doxil and AmBisome. *Nanomedicine*. 2012;8:176–184.
77. Szebeni J. Complement activation-related pseudoallergy: A stress reaction in blood triggered by nanomedicines and biologicals. *Mol Immunol*. 2014;61:163–173.
78. Clancy EK, Duncan R. Helix-destabilizing, beta-branched, and polar residues in the baboon reovirus p15 transmembrane domain influence the modularity of FAST proteins. *J Virol*. 2011;85:4707–4719.
79. Clancy EK, Duncan R. Reovirus FAST protein transmembrane domains function in a modular, primary sequence-independent manner to mediate cell-cell membrane fusion. *J Virol*. 2009;83:2941–2950.
80. Corcoran JA, Clancy EK, Duncan R. Homomultimerization of the reovirus p14 fusion-associated small transmembrane protein during transit through the ER-Golgi complex secretory pathway. *J Gen Virol*. 2011;92(Pt 1):162–166.
81. Clancy EK, Barry C, Ciechonska M, et al. Different activities of the reovirus FAST proteins and influenza hemagglutinin in cell-cell fusion assays and in response to membrane curvature agents. *Virology*. 2010;397:119–129.
82. Dawe S, Corcoran JA, Clancy EK, et al. Unusual topological arrangement of structural motifs in the baboon reovirus fusion-associated small transmembrane protein. *J Virol*. 2005;79:6216–6226.
83. Corcoran JA, Salsman J, de Antueno R, et al. The p14 fusion-associated small transmembrane (FAST) protein effects membrane fusion from a subset of membrane microdomains. *J Biol Chem*. 2006;281:31778–31789.
84. Read J, Clancy EK, Sarker M, et al. Reovirus FAST proteins drive pore formation and syncytiogenesis using a novel helix-loop-helix fusion-inducing lipid packing sensor. *PLoS Pathog*. 2015;11:e1004962.
85. Brown DW, Raturi A, Bhandari P, et al. Selective ablation of solid tumors using a p53-targeted FAST-LNP gene therapy. *Cancer Res*. 2020;80(16S):4069.
86. Duncan R. Fusogenic reoviruses and their fusion-associated small transmembrane (FAST) proteins. *Annu Rev Virol*. 2019;6:341–363.
87. Top D, de Antueno R, Salsman J, et al. Liposome reconstitution of a minimal protein-mediated membrane fusion machine. *EMBO J*. 2005;24:2980–2988.
88. Corcoran JA, Syvitski R, Top D, et al. Myristoylation, a protruding loop, and structural plasticity are essential features of a nonenveloped virus fusion peptide motif. *J Biol Chem*. 2004;279:51386–51394.



89. Top D, Read JA, Dawe SJ, et al. Cell-cell membrane fusion induced by p15 fusion-associated small transmembrane (FAST) protein requires a novel fusion peptide motif containing a myristoylated polyproline type II helix. *J Biol Chem.* 2012;287:3403–3414.
90. Top D, Barry C, Racine T, et al. Enhanced fusion pore expansion mediated by the trans-acting endodomain of the reovirus FAST proteins. *PLoS Pathog.* 2009;5:e1000331.
91. Duncan R, Corcoran J, Shou J, et al. Reptilian reovirus: a new fusogenic orthoreovirus species. *Virology.* 2004;319:131–140.
92. Salsman J, Zimmerman N, Chen T, et al. Genome-wide screen of three herpesviruses for protein subcellular localization and alteration of PML nuclear bodies. *PLoS Pathog.* 2008;4:e1000100.
93. Salsman J, Top D, Boutilier J, Duncan R. Extensive syncytium formation mediated by the reovirus FAST proteins triggers apoptosis-induced membrane instability. *J Virol.* 2005;79:8090–8100.
94. Duncan R, Murphy FA, Mirkovic RR. Characterization of a novel syncytium-inducing baboon reovirus. *Virology.* 1995;212:752–756.
95. Parmar HB, Barry C, Duncan R. Polybasic trafficking signal mediates golgi export, ER retention or ER export and retrieval based on membrane-proximity. *PLoS One.* 2014;9:e94194.
96. Corcoran JA, Duncan R. Reptilian reovirus utilizes a small type III protein with an external myristylated amino terminus to mediate cell-cell fusion. *J Virol.* 2004;78:4342–4351.
97. Ciechonska M, Duncan R. Lysophosphatidylcholine reversibly arrests pore expansion during syncytium formation mediated by diverse viral fusogens. *J Virol.* 2014;88:6528–6531.
98. Shmulevitz M, Duncan R. A new class of fusion-associated small transmembrane (FAST) proteins encoded by the non-enveloped fusogenic reoviruses. *EMBO J.* 2000;19:902–912.
99. Barry C, Duncan R. Multifaceted sequence-dependent and -independent roles for reovirus FAST protein cytoplasmic tails in fusion pore formation and syncytiogenesis. *J Virol.* 2009;83:12185–12195.
100. Ciechonska M, Duncan R. Reovirus FAST proteins: virus-encoded cellular fusogens. *Trends Microbiol.* 2014;22:715–724.
101. Boutilier J, Duncan R. The reovirus fusion-associated small transmembrane (FAST) proteins: virus-encoded cellular fusogens. *Curr Top Membr.* 2011;68:107–140.