



Recent advances in the understanding of the molecular pathogenesis and targeted therapy options in Langerhans cell histiocytosis

Jin Kyung Suh, Sunghan Kang, Hyery Kim, Ho Joon Im, Kyung-Nam Koh

Division of Pediatric Hematology/Oncology, Department of Pediatrics, Asan Medical Center Children's Hospital, University of Ulsan College of Medicine, Seoul, Korea

p-ISSN 2287-979X / e-ISSN 2288-0011
<https://doi.org/10.5045/br.2021.2021013>
Blood Res 2021;56:S65-S69.

Received on January 21, 2021

Revised on April 2, 2021

Accepted on April 2, 2021

*This study was supported by a grant from the Korea Disease Control and Prevention Agency (2019ER690301).

Correspondence to

Kyung-Nam Koh, M.D., Ph.D.

Ho Joon Im, M.D., Ph.D.

Division of Pediatric Hematology/Oncology, Department of Pediatrics, Asan Medical Center Children's Hospital, University of Ulsan College of Medicine, 88, Olympic-ro, 43-gil, Songpa-gu, Seoul 05505, Korea (K.N.K. and H.J.I.)

E-mail: K.N.K., pedkkn@amc.seoul.kr
H.J.I., hojim@amc.seoul.kr

© 2021 Korean Society of Hematology

Abstract

Langerhans cell histiocytosis (LCH) is the most common histiocytic disorder caused by the clonal expansion of myeloid precursors that differentiate into CD1a+/CD207+ cells in the lesion. Advances in genomic sequencing techniques have improved our understanding of the pathophysiology of LCH. Activation of the mitogen-activated protein kinase (MAPK) pathway is a key molecular mechanism involved in the development of LCH. Recurrent BRAF mutations and MAP2K1 mutations are the major molecular alterations involved in the activation of the MAPK pathway. Recent studies have supported the “misguided myeloid differentiation model” of LCH, where the extent of disease is defined by the differentiation stage of the cell in which the activating somatic MAPK mutation occurs, suggesting LCH. Several studies have advocated the efficacy of targeted therapy using BRAF inhibitors with a high response rate, especially in patients with high-risk or refractory LCH. However, the optimal treatment scheme for children remains unclear. This review outlines recent advances in LCH, focusing on understanding the molecular pathophysiology, emerging targeted therapy options, and their clinical implications.

Key Words Langerhans cell histiocytosis, Therapeutics, Pathology

INTRODUCTION

Langerhans cell histiocytosis (LCH) is a myeloproliferative disorder characterized by inflammatory lesions with the accumulation of CD1a+/CD207+ histiocytes, leading to the destruction of affected tissues [1, 2]. While LCH invades bones and skin commonly, it can involve any organ, including the liver, spleen, lungs, lymph nodes, central nervous system, and hematopoietic system. The clinical course varies greatly from a single bone lesion to a fatal multi-organ disease [3]. Although the precise origin of the pathological histiocytes in LCH is unclear, the current hypothesis is that these pathological cells are more likely to arise from dysregulated differentiation or the recruitment of bone marrow-derived precursor cells or yolk sac progenitors than from epidermal Langerhans cells [4, 5]. Recent advances in genomic sequenc-

ing have highlighted the activation of the mitogen-activated protein kinase (MAPK) pathway at critical stages in myeloid differentiation as an essential driver of LCH pathology [4]. In this review, we overview the recently growing understanding of LCH pathogenesis and a novel therapeutic option to optimize and personalize therapy through improved risk stratification, targeted therapy, and the assessment of therapy responses based on specific molecular features.

Pathological considerations of LCH

Diagnosis and clinical overview of LCH: Histiocytic disorders are diseases associated with cells of the mononuclear phagocyte system, which include hematopoietic cells with mononuclear morphology and phagocytic activity. LCH is the most common histiocytic disorder, and its clinical presentation varies from limited, indolent, and self-regressive to life-threatening disseminated disease. The diagnosis of

LCH is based on clinical and radiological findings in combination with histopathological analysis, identifying the infiltration of tissue by histiocytes using the ultrastructural or immunophenotypic characteristics of Langerhans cells, which include the surface expression of CD207 (Langerin) and CD1a, as well as the detection of Birbeck granules using electron microscopy [6]. In routine clinical practice, the identification of Birbeck granules has recently been replaced with positive immunohistochemical staining for CD1a and CD207.

LCH as an immunoreactive disorder versus as a neoplastic disorder: In LCH lesions, pathological CD1a+CD207+ histiocytes are present in a small proportion, from less than 1% to more than 70% (median 8%), and are mixed with a variety of inflammatory cells, including eosinophils, macrophages, multinucleated giant cells, and lymphocytes (enriched with regulatory T cells) [7, 8]. In addition, these pathological histiocytes have a benign morphology and low mitotic activity. In this regard, LCH is considered an immune disorder in which normal cells proliferate in response to environmental stimuli.

The findings of Badalian-Vey *et al.* [9], in 2010, showed that 57% of patients with LCH had recurrent *BRAF*V600E mutations, and subsequent studies have identified recurrent somatic mutations in pathological LCH cells, revealing the neoplastic nature of LCH and leading to the definition of LCH as an inflammatory myeloid neoplasm [1].

Misguided myeloid differentiation model of LCH pathogenesis: The identification of the *BRAF*V600E mutation provided a molecular tag for lineage tracking to test the hypothesis that LCH originated from early myeloid progenitors rather than epidermal Langerhans cells. An analysis of peripheral

blood mononuclear cells (PBMCs) identified the presence of a small (<1%) but consistent proportion of *BRAF*V600E mutations in myeloid cells (CD11c+ myeloid dendritic cell precursors and CD14+ monocytes) in patients with high-risk LCH. Similarly, in bone marrow samples, *BRAF*V600E was identified in CD34+ hematopoietic stem cells from many patients with high-risk LCH, of which up to 50% were reported to have normal histology. In contrast, *BRAF*V600E was generally absent in PBMCs from patients with single lesion LCH and rare in the PBMCs of patients with multifocal low-risk LCH [4, 7, 10]. These observations support the “misguided myeloid differentiation model” of LCH, where the extent of disease is defined by the differentiation stage of the cell in which the activating somatic MAPK mutation occurs (Fig. 1) [4]. According to this model, the high-risk multisystem LCH is caused by somatic activating mutations in the MAPK gene of hematopoietic stem/progenitor cells from bone marrow; the low-risk multisystem LCH is caused by somatic MAPK mutations of committed dendritic cell (DC) precursor cells in the blood, and the low-risk single-system LCH is caused by somatic MAPK mutations of more differentiated DC precursors from blood residing in a single lesion [4, 7, 10].

Recent progress in molecular pathogenesis of LCH

Activation of MAPK pathway in LCH: The improvement of sequencing techniques that amplify the sequencing depth has enabled simultaneous studies of multiple target genomic regions per sample, which has led to the identification of the recurrent oncogenic somatic mutations in LCH that activate the MAPK pathway. The MAPK pathway comprising RAS/RAF/MEK/ERK, which regulates cell differentiation,

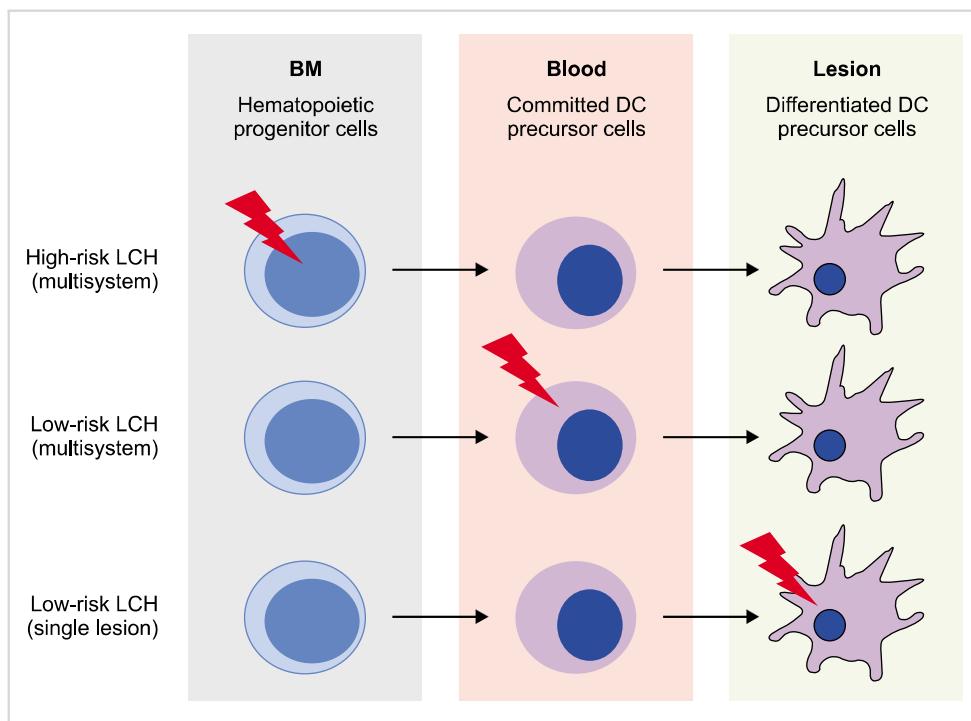


Fig. 1. Misguided myeloid differentiation model for LCH ontogeny according to different risk groups. According to this model, the extent of disease is defined by the differentiation stage of the cell in which the activating somatic MAPK mutation occurs.

Abbreviations: DC, dendritic cell; LCH, Langerhans cell histiocytosis.

proliferation, and apoptosis, plays a vital role in myeloid cell differentiation and maturation (Fig. 2). Previous reports have shown evidence of activation of the MAPK pathway in all the LCH samples [9, 11]. Mutations in the MAPK pathway proteins are associated with the differentiation of DC progenitors and inhibit the maturation of DCs. This role of activation of the MAPK pathway in myeloid cell differentiation and maturation may have implications regarding the origin and functional alterations of histiocytes in LCH [4].

Somatic mutations associated with LCH: Badalian-Vey *et al.* [9] identified the *BRAF*-V600E mutation in pathological LCH cells and subsequent studies explored the genomic landscape of childhood LCH using targeted and whole-exome next-generation sequencing (NGS) approaches. The heterozygous *BRAF*-V600E mutation is a predominant genomic alteration in LCH, estimated in 38–64% of patients with LCH [7, 9, 12, 13]. Recurrent mutations in exons 2 and 3 of the *MAP2K1* gene, which encodes MEK1, a downstream signaling protein of *BRAF* in the MAPK pathway, were reported in 15–20% of LCH cases [14, 15]. In addition, small in-frame *BRAF* deletions or insertions localized to exon 12, which encodes the small N-terminal lobe of the *BRAF* kinase domain, have been reported in 5–10% of LCH cases [14, 16]. In addition, rare cases of mutations of other genes encoding MAPK pathway proteins have been reported, such as FAM73A-*BRAF* fusion [14], *MAP3K1* mutation [15], and *ARAF* mutation [17]. A somatic oncogenic PIK3CA mutation associated with the PI3K-AKT pathway has been reported

in a pediatric LCH case and in an adult non-LCH disease (Erdheim-Chester disease) [18, 19]. No genetic alterations were found in approximately 20% of the patients with LCH. In these cases, copy number changes or epigenetic alterations may be additional mechanisms for MAPK pathway activation [1].

Targeted therapy in LCH

***BRAF* kinase inhibitors: vemurafenib and dabrafenib:** Identification of *BRAF* and *MAP2K1* mutations in LCH led to targeted therapies acting on the MAPK pathway. First-generation *BRAF* inhibitors, vemurafenib and dabrafenib, for the treatment of high-risk or refractory LCH, have been studied. The efficacy of *BRAF* inhibitors in the treatment of LCH was first reported in the off-label clinical use of vemurafenib in three patients with multisystem and refractory ECD carrying the *BRAF*-V600E mutation, two of whom also had combined LCH, and the other being a young infant with refractory, multisystem LCH. In these reports, patients receiving vemurafenib showed a dramatic response and good short-term tolerability [20, 21]. A recent international study of vemurafenib in 54 pediatric patients with refractory multisystem LCH showed a 100% overall response rate (complete response in 70% and partial response in 30%) after at least eight weeks of vemurafenib (20 mg/kg/day) monotherapy. However, 80% of the patients experienced LCH reactivation after the discontinuation of vemurafenib. The re-introduction of vemurafenib was effective in 75% of the reactivated patients [22]. Previous studies on vemurafenib have

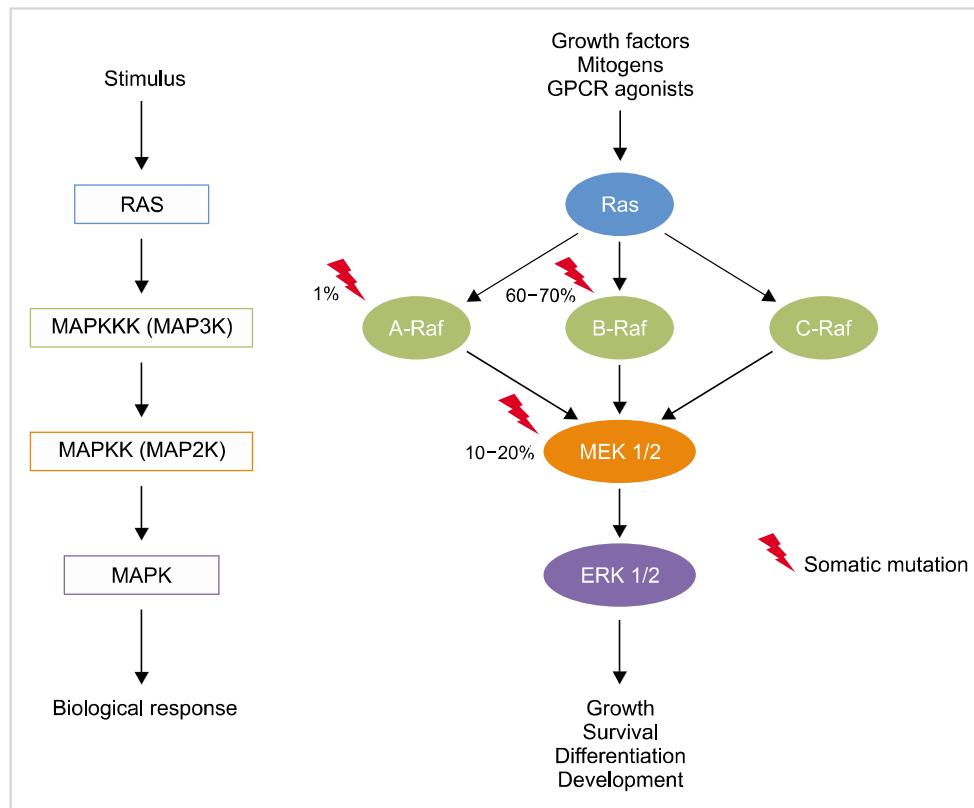


Fig. 2. MAPK pathway mutations in LCH. Activating MAPK pathway mutations in LCH drive pathologic ERK activation.
Abbreviations: A-Raf, rapidly accelerated fibrosarcoma isoform A; B-Raf, rapidly accelerated fibrosarcoma isoform B; C-Raf, rapidly accelerated fibrosarcoma isoform C; ERK, extracellular signal-regulated kinase; GPCR, G-protein-coupled receptor; LCH, Langerhans cell histiocytosis; MAPK, mitogen-activated protein kinase; MEK, mitogen-activated protein kinase kinase; Ras, rat sarcoma.

found that while patients achieved excellent response rates, frequent recurrences were observed after the discontinuation of vemurafenib [23, 24]. Dabrafenib is another treatment option for high-risk and refractory LCH. Dabrafenib is a selective *BRAF* kinase inhibitor that has been reported to have fewer adverse effects [25]. In a recently reported Chinese pediatric study, with 20 enrolled patients with refractory LCH that were treated with dabrafenib monotherapy, the overall objective response rate was 65%, and the overall disease control rate was 75% [26].

Drawbacks of *BRAF* kinase inhibitors and future perspectives of target therapy: Although *BRAF*-V600E targeted therapy is an important advance, especially for refractory or high-risk LCH, the use of *BRAF* kinase inhibitors has several drawbacks that need to be overcome. *BRAF* inhibitors were initially used in the treatment of metastatic melanoma and other adult *BRAF*-mutated cancers. According to the adult experience of *BRAF* inhibitors, *BRAF* inhibitors were ineffective for many other *BRAF* mutations, such as *BRAF* fusion with a partner gene, insertion/deletion of exon 12, and ineffective in activating mutations of genes that encode proteins acting upstream of RAF (such as RAS) [27]. This ineffectiveness can trigger a paradoxical transactivation mechanism, which is responsible for serious cutaneous adverse effects, including keratoacanthomas and squamous-cell carcinomas [28]. In pediatric cases, rapid LCH reactivation has been observed after the discontinuation of *BRAF* inhibitors, and it has been found to be ineffective in sclerosing cholangitis and advanced neurodegenerative disease [1, 22, 29]. In this regard, prospective trials should be performed to determine an optimal duration of use and potential combinations with other targeted or cytotoxic therapies. An approach combining a *BRAF* inhibitor with a MEK inhibitor that is used in adult malignancies can be effective in LCH. There is currently an ongoing international clinical trial to evaluate the efficacy of a combination of dabrafenib and the MEK inhibitor, trametinib, for adults and children with refractory or relapsed LCH (MEK inhibitor±*BRAF* inhibitor, NCT02124772).

Clinical considerations according to molecular pathogenesis

***BRAF*-V600E mutation as a risk factor of LCH:** Several previous studies have shown that the *BRAF*-V600E mutation is associated with the severity of LCH and with treatment failure. Berres *et al* [7] reported that patients with a *BRAF*-V600E mutation had an approximately two-fold risk of reactivation. Héritier *et al* [30] found that patients with *BRAF*-V600E had more severe disease, tended to resist standard therapy, had a higher reactivation rate, and more permanent, long-term sequelae after treatment. In this regard, the French group recommended that a molecular analysis be performed at least once for the following presentations at the initial biopsy: 1) LCH with risk organ involvement; 2) LCH diagnosis before two years of age; 3) LCH with pituitary involvement or skull base or orbit bone lesions; 4) resistance to first-line therapy; and 5) severe complications and sequelae [1].

Circulating cell-free *BRAF*-V600E as a biomarker in LCH: Héritier *et al* [30] quantified the *BRAF*-V600E allele in the circulating cell-free DNA from plasma samples from pediatric patients with LCH. This study reported a higher *BRAF* ccf load in patients with risk organ involvement and a sequential decrease in the *BRAF* ccf load after treatment [30]. Recent studies of *BRAF* inhibitors in patients with LCH included follow-up tests of *BRAF* ccf load and showed that *BRAF* ccf load reduced rapidly after *BRAF* inhibitor treatment [22, 26]. These results indicated that the *BRAF* ccf load was associated with high-risk features and was responsive to chemotherapy. In this regard, the ccf *BRAF* load should be considered as a blood biomarker of LCH in future clinical trials.

CONCLUSION

LCH has long been an enigma with respect to its origin and pathogenesis. Recent advances in molecular genomic analysis techniques have revealed the pathogenic molecular mechanism of LCH. The understanding of the origin and biology of LCH has increased rapidly in the last decade, leading to the introduction of promising targeted therapies for high-risk patients. However, high-risk patients still have suboptimal outcomes, and targeted therapy for such patients cannot eradicate the malignant clone and provide long-lasting remission after discontinuation. Future research should focus on improving outcomes and reducing permanent sequelae in high-risk LCH patients based on the recent understanding of LCH.

ACKNOWLEDGMENTS

We thank Lee Changik, Park Yerang, and Lee Sangmin for their generous support for this research.

Authors' Disclosures of Potential Conflicts of Interest

No potential conflicts of interest relevant to this article were reported.

REFERENCES

1. Héritier S, Emile JF, Hélias-Rodzewicz Z, Donadieu J. Progress towards molecular-based management of childhood Langerhans cell histiocytosis. *Arch Pediatr* 2019;26:301-7.
2. Koh KN, Im HJ, Seo JJ. Recent updates in Langerhans cell histiocytosis. *Clin Pediatr Hematol Oncol* 2015;22:15-21.
3. Kim BE, Koh KN, Suh JK, et al. Clinical features and treatment outcomes of Langerhans cell histiocytosis: a nationwide survey from Korea histiocytosis working party. *J Pediatr Hematol Oncol* 2014;36:125-33.
4. Berres ML, Merad M, Allen CE. Progress in understanding the

- pathogenesis of Langerhans cell histiocytosis: back to Histiocytosis X? *Br J Haematol* 2015;169:3-13.
5. Collin M, Bigley V, McClain KL, Allen CE. Cell(s) of origin of Langerhans cell histiocytosis. *Hematol Oncol Clin North Am* 2015;29:825-38.
 6. Emile JF, Abla O, Fraitag S, et al. Revised classification of histiocytoses and neoplasms of the macrophage-dendritic cell lineages. *Blood* 2016;127:2672-81.
 7. Berres ML, Lim KP, Peters T, et al. BRAF-V600E expression in precursor versus differentiated dendritic cells defines clinically distinct LCH risk groups. *J Exp Med* 2014;211:669-83.
 8. Senechal B, Elain G, Jeziorski E, et al. Expansion of regulatory T cells in patients with Langerhans cell histiocytosis. *PLoS Med* 2007;4:e253.
 9. Badalian-Very G, Vergilio JA, Degar BA, et al. Recurrent BRAF mutations in Langerhans cell histiocytosis. *Blood* 2010;116: 1919-23.
 10. Rodriguez-Galindo C, Allen CE. Langerhans cell histiocytosis. *Blood* 2020;135:1319-31.
 11. Chakraborty R, Hampton OA, Shen X, et al. Mutually exclusive recurrent somatic mutations in MAP2K1 and BRAF support a central role for ERK activation in LCH pathogenesis. *Blood* 2014;124:3007-15.
 12. Sahm F, Capper D, Preusser M, et al. BRAFV600E mutant protein is expressed in cells of variable maturation in Langerhans cell histiocytosis. *Blood* 2012;120:e28-34.
 13. Satoh T, Smith A, Sarde A, et al. B-RAF mutant alleles associated with Langerhans cell histiocytosis, a granulomatous pediatric disease. *PLoS One* 2012;7:e33891.
 14. Chakraborty R, Burke TM, Hampton OA, et al. Alternative genetic mechanisms of BRAF activation in Langerhans cell histiocytosis. *Blood* 2016;128:2533-7.
 15. Nelson DS, van Halteren A, Quispel WT, et al. MAP2K1 and MAP3K1 mutations in Langerhans cell histiocytosis. *Genes Chromosomes Cancer* 2015;54:361-8.
 16. Héritier S, Hélias-Rodzewicz Z, Chakraborty R, et al. New somatic BRAF splicing mutation in Langerhans cell histiocytosis. *Mol Cancer* 2017;16:115.
 17. Nelson DS, Quispel W, Badalian-Very G, et al. Somatic activating ARAF mutations in Langerhans cell histiocytosis. *Blood* 2014;123:3152-5.
 18. Héritier S, Saffroy R, Radosevic-Robin N, et al. Common cancer-associated PIK3CA activating mutations rarely occur in Langerhans cell histiocytosis. *Blood* 2015;125:2448-9.
 19. Emile JF, Diamond EL, Hélias-Rodzewicz Z, et al. Recurrent RAS and PIK3CA mutations in Erdheim-Chester disease. *Blood* 2014; 124:3016-9.
 20. Haroche J, Cohen-Aubart F, Emile JF, et al. Dramatic efficacy of vemurafenib in both multisystemic and refractory Erdheim-Chester disease and Langerhans cell histiocytosis harboring the BRAF V600E mutation. *Blood* 2013;121:1495-500.
 21. Héritier S, Jehanne M, Leverger G, et al. Vemurafenib use in an infant for high-risk Langerhans cell histiocytosis. *JAMA Oncol* 2015;1:836-8.
 22. Donadieu J, Larabi IA, Tardieu M, et al. Vemurafenib for refractory multisystem langerhans cell histiocytosis in children: an international observational study. *J Clin Oncol* 2019;37: 2857-65.
 23. Heisig A, Sörensen J, Zimmermann SY, et al. Vemurafenib in Langerhans cell histiocytosis: report of a pediatric patient and review of the literature. *Oncotarget* 2018;9:22236-40.
 24. Kolenová A, Schwentner R, Jug G, et al. Targeted inhibition of the MAPK pathway: emerging salvage option for progressive life-threatening multisystem LCH. *Blood Adv* 2017;1:352-6.
 25. Hauschild A, Grob JJ, Demidov LV, et al. Dabrafenib in BRAF-mutated metastatic melanoma: a multicentre, open-label, phase 3 randomised controlled trial. *Lancet* 2012;380:358-65.
 26. Yang Y, Wang D, Cui L, et al. Effectiveness and safety of dabrafenib in the treatment of 20 Chinese children with BRAFV600E-mutated Langerhans cell histiocytosis. *Cancer Res Treat* 2021; 53:261-9.
 27. Karoulia Z, Gavathiotis E, Poulikakos PI. New perspectives for targeting RAF kinase in human cancer. *Nat Rev Cancer* 2017; 17:676-91.
 28. Su F, Viros A, Milagre C, et al. RAS mutations in cutaneous squamous-cell carcinomas in patients treated with BRAF inhibitors. *N Engl J Med* 2012;366:207-15.
 29. McClain KL, Picarsic J, Chakraborty R, et al. CNS Langerhans cell histiocytosis: common hematopoietic origin for LCH-associated neurodegeneration and mass lesions. *Cancer* 2018;124:2607-20.
 30. Héritier S, Emile JF, Barkaoui MA, et al. BRAF mutation correlates with high-risk Langerhans cell histiocytosis and increased resistance to first-line therapy. *J Clin Oncol* 2016;34:3023-30.