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Genome Sequence of *Fusarium* graminearum ITEM 124 (ATCC 56091), a

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Mycotoxigenic Plant Pathogen

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ABSTRACT Fusarium graminearum is among the main causal agents of Fusarium head blight (FHB), or scab, of wheat and other cereals, caused by a complex of Fusarium species, worldwide. Besides causing economic losses in terms of crop yield and quality, *F. graminearum* poses a severe threat to animal and human health. Here, we present the first draft whole-genome sequence of the mycotoxigenic Fusarium graminearum strain ITEM 124, also providing useful information for comparative genomics studies.

Fusarium head blight (FHB) of wheat is a major disease worldwide, with *Fusarium* graminearum being the main causal agent of the species complex (1). Notably, *F. graminearum* was recently ranked fourth in a top 10 fungal plant pathogen list (2). FHB not only directly affects the grain production but also poses a severe threat to plant, animal, and human health due to the accumulation of mycotoxins on kernels and wheat products.

F. graminearum ITEM 124 (http://server.ispa.cnr.it/ITEM/Collection/), also known as ATCC 56091, was isolated in 1976 from rice harvested in Vercelli, Piemonte, Italy. *F. graminearum* ITEM 124 is able to produce deoxynivalenol (DON), the best-known mycotoxin belonging to trichothecenes (3). This class of mycotoxins acts as an inhibitor of cell protein synthesis (4). The ability of *F. graminearum* ITEM 124 to produce DON on wheat and rice kernels has been assessed previously (5, 6). Molecular chemotype detection, performed according to Somma et al. (7), identified the strain *F. graminearum* ITEM 124 as chemotype 15-acetyl DON (15ADON). Chemical analyses confirmed the production of DON by ultraperformance liquid chromatography (UPLC) with a photodiode array (PDA) assay (8).

A competition test on rice kernels inoculated with *F. graminearum* ITEM 124 and the beneficial fungus *Trichoderma gamsii* T6085 highlighted a reduction of the amount of DON to almost 92% as a direct consequence of a decreased biomass of the pathogen. Moreover, the growth of *F. graminearum* ITEM 124 has been reduced by the presence of *T. gamsii* T6085 in dual confrontation assays. *F. graminearum* ITEM 124 is overgrown by *T. gamsii* T6085 under *in vitro* conditions, and the latter produces short loops and coilings on pathogen hyphae, which are typical mycoparasitic traits (5, 6, 9).

The genome of *F. graminearum* ITEM 124 was sequenced using Illumina paired-end sequencing technology by BMR Genomics (Padua, Italy). Paired reads of 300 bp (3.16 Gbp; average coverage, $38\times$) were assembled using SPAdes v3.8.2 (10). The nuclear genome of *F. graminearum* ITEM 124 consists of 67 sequence scaffolds with a total assembly length of 36.88 Mbp (N_{50} , 1,518,396; L_{50} , 6), 48.18% GC content, and a maximum scaffold size of 7,305,194 bp. The completeness of the assembly was assessed using BUSCO v12 (11), which estimated the genome sequence to be 99.86%

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complete. The nuclear genome was annotated using the MAKER2 pipeline (12). Overall, 11,827 protein-coding gene models were predicted. Analysis with SignalP 4.1 (13) revealed that 1,288 predicted proteins (10.9% of the proteome) contain a secretion signal peptide.

The genome sequence of the mycotoxigenic *F. graminearum* ITEM 124 provides a novel source for comparative genomics studies and also represents a useful platform to study fungal-fungal interactions, such as the mycoparasitic relationship established with the beneficial isolate *T. gamsii* T6085, whose genome is also available (14).

Accession number(s). This whole-genome shotgun project has been deposited in GenBank under accession number NQOC00000000 (BioProject PRJNA397367; Bio-Sample SAMN07455307). The version described in this paper is NQOC01000000.

REFERENCES

- Parry DW, Jenkinson P, McLeod L. 1995. Fusarium ear blight (scab) in small grain cereals—a review. Plant Pathol 44:207–238. https://doi.org/ 10.1111/j.1365-3059.1995.tb02773.x.
- Dean R, Van Kan JA, Pretorius ZA, Hammond-Kosack KE, Di Pietro A, Spanu PD, Rudd JJ, Dickman M, Kahmann R, Ellis J, Foster GD. 2012. The top 10 fungal pathogens in molecular plant pathology. Mol Plant Pathol 13:414–430. https://doi.org/10.1111/j.1364-3703.2011.00783.x.
- Desjardins AE. 2006. Trichothecenes, p 13–64. *In Fusarium* mycotoxins chemistry, genetics, and biology. The American Phytopathological Society, St. Paul, MN.
- 4. Westerberg UB, Bolcsfoldi G, Eliasson E. 1976. Control of transfer RNA synthesis in the presence of inhibitors of protein synthesis. Biochim Biophys Acta 447:203–213. https://doi.org/10.1016/0005-2787(76)90343-9.
- Matarese F. 2010. Biocontrol of *Fusarium* head blight: molecular interactions between *Trichoderma* and mycotoxigenic *Fusarium*. Ph.D. thesis. University of Pisa, Pisa, Italy.
- Matarese F, Sarrocco S, Gruber S, Seidl-Seiboth V, Vannacci G. 2012. Biocontrol of *Fusarium* head blight: interactions between *Trichoderma* and mycotoxigenic *Fusarium*. Microbiology 158:98–106. https://doi.org/ 10.1099/mic.0.052639-0.
- Somma S, Petruzzella AL, Logrieco AF, Meca G, Cacciola OS, Moretti A. 2014. Phylogenetic analyses of *Fusarium graminearum* strains from cereals in Italy, and characterization of their molecular and chemical chemotypes. Crop Pasture Sci 65:52–60.
- 8. Pascale M, Panzarini G, Powers S, Visconti A. 2014. Determination of deoxynivalenol and nivalenol in wheat by ultra-performance liquid

chromatography/photodiode-array detector and immunoaffinity column cleanup. Food Anal Methods 7:555–562. https://doi.org/10.1007/ s12161-013-9653-1.

- Sarrocco S, Matarese F, Moncini L, Pachetti G, Ritieni A, Moretti A, Vannacci G. 2013. Biocontrol of *Fusarium* head blight by spike application of *Trichoderma gamsii*. J Plant Pathol S1:19–27.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.
- Simão FA, Waterhouse RM, Ioannidis P, Kriventseva EV, Zdobnov EM. 2015. BUSCO: assessing genome assembly and annotation completeness with single-copy orthologs. Bioinformatics 31:3210–3212. https:// doi.org/10.1093/bioinformatics/btv351.
- Holt C, Yandell M. 2011. MAKER2: an annotation pipeline and genomedatabase management tool for second-generation genome projects. BMC Bioinformatics 12:491. https://doi.org/10.1186/1471-2105-12-491.
- Petersen TN, Brunak S, von Heijne G, Nielsen H. 2011. SignalP 4.0: discriminating signal peptides from transmembrane regions. Nat Methods 8:785–786. https://doi.org/10.1038/nmeth.1701.
- Baroncelli R, Zapparata A, Piaggeschi G, Sarrocco S, Vannacci G. 2016. Draft whole-genome sequence of *Trichoderma gamsii* T6085, a promising biocontrol agent of *Fusarium* head blight on wheat. Genome Announc 4(1):e01747-15. https://doi.org/10.1128/genomeA.01747-15.