



OPEN Maternal and paternal lineage analysis of Island Southeast Asian goats reveals continental propagation routes and introgression through the Indian ocean

Ryo Masuko¹, Ayin¹, Maho Masaoka¹, Fuki Kawaguchi¹, Shinji Sasazaki¹, Muhammad I. A. Dagong², Sri R. A. Bugiwati², Joseph S. Masangkay³, Takahiro Yonezawa⁴ & Hideyuki Mannen¹✉

The Philippines and Indonesia, located at the crossroads of historical human migrations in the Asia-Pacific region, lie in the southeasternmost part of Mainland Southeast Asia. Both countries are recognized as mega-diverse nations, hosting a wide variety of species. However, the history of goat propagation in Island Southeast Asia remains unclear due to limited molecular studies and the absence of archaeological evidence in this region. This study conducted phylogeographic analyses of mtDNA hypervariable region 1 and SRY 3'UTR sequences of goats from the Philippines and Indonesia, comparing them with Old World domestic goats. Philippine goat mtDNA haplotypes were classified into haplogroups B (152/206, 73.8%) and A (54/206, 26.2%), whereas all Indonesian goat mtDNA haplotypes belonged to haplogroup B (72/72, 100%). The distribution of mtDNA haplogroup B and regional differences suggest two propagation routes: one from China/Taiwan to the Philippines and another from Mainland Southeast Asia to Indonesia. The distribution of five SRY haplotypes demonstrated that haplotype Y2A, absent in Mainland Southeast Asia, was present in Island Southeast Asia, indicating gene introgression through the Indian Ocean from the African continent and/or southern Europe. This introgression may have been driven by human activities, such as European migrations between the 16th and 19th centuries.

Keywords Island Southeast Asia, Indian ocean, Propagation route, *Capra hircus*, MtDNA, SRY gene

Goats (*Capra hircus*), domesticated in the Fertile Crescent approximately 10,000 years ago, are bred in most parts of the world today^{1–3}. They are essential livestock for human life, providing milk, meat, hair, and other products. The genetic information of goats is crucial for understanding their genetic relationships, the origins of domestication, and the history of their propagation. Studies of goat genetics have primarily focused on mtDNA and the SRY gene on the Y chromosome^{4–8}.

Previous studies mainly based on sequences of hypervariable mtDNA control region (481 bp) have identified six maternal haplogroups (A, B, C, D, E, and G)^{1,2,9–11}. Haplogroup A is the most divergent globally, while the minor haplogroups C, D, E, and G are found sporadically at low frequencies across Asia, Europe, the Near East, and North Africa, respectively. The mtDNA haplogroups have been reported to show a weak phylogenetic structure due to extremely high mtDNA variation across regions and continent^{1,2,12,13}. In contrast, haplogroup B is found at a high frequency in Southeast Asia. Given the unique phylogeographical structure of haplogroup B, it

¹Laboratory of Animal Breeding and Genetics, Graduates School of Agricultural Science, Kobe University, Kobe, Japan. ²Faculty of Animal Science, Hasanuddin University, Makassar, South Sulawesi 90245, Indonesia. ³College of Veterinary Medicine, University of the Philippines, Los Baños, Philippines. ⁴Graduate School of Integrated Sciences for Life, Hiroshima University, Higashi-Hiroshima 739-8528, Japan. ✉email: mannen@kobe-u.ac.jp

is postulated that the Baluchistan region of Pakistan may be the origin of haplogroup B^{1,11}. However, the details of the origin and propagation routes of haplogroup B remain unclear.

As informative paternal genetic variations, five haplotypes (Y1AA, Y1AB, Y1B, Y2A, and Y2B) have been reported in previous studies using the Sex-determining region Y chromosome (SRY) 3'UTR sequence^{8,14–17}. Y1A is further classified into Y1AA and Y1AB; however, Y1A in certain parts of Europe and Africa has not been divided into these haplotypes. The available data suggest a geographic contrast, with Y1AB predominating in northern China and Y1AA along with Y2B in the south. Y1B, Y2A, and Y2B are primarily observed in Europe, Africa, and East Asia, respectively¹⁷. The distinct distribution of SRY haplotypes across regions indicates that SRY haplotypes exhibit a strong phylogeographic structure, in contrast to the weak phylogenetic structure seen in the major mtDNA haplogroups¹⁷.

The Philippines and Indonesia, located in Southeast Asia, are archipelagic nations consisting of numerous islands. The Philippines, situated at the crossroads of past human migrations in the Asia-Pacific region, is believed to have never been connected to the Asian continent, even during the major marine regressions of the Pleistocene. In contrast, Indonesia was part of Sundaland, which was connected to present-day Mainland Southeast Asia (MSEA) by land during the Pleistocene and is located in the southeasternmost part of MSEA. In Island Southeast Asia (ISEA), there are historical records of trade and Islamic missionary work by Muslim merchants beginning in the 9th century, as well as colonial rule by Spain in the Philippines and by the Netherlands in Indonesia from the 16th century onward^{18–20}.

It is generally thought that ISEA goats were propagated from the domestication center via MSEA and Maritime Silk Road. These small goats, called “Kambing Katjang”, are well known to be raised over a broad area of Southeast Asia^{17,21}. Given geographical and historical background of ISEA, Island Southeast Asian goats may have distinct genetic influences compared to those in Mainland Southeast Asia. Supporting this, several previous studies on Southeast Asian goats have reported that the SRY haplotype Y2A, primarily observed in Africa, is found in ISEA but not in MSEA^{17,22,23}. However, the genetic structure and introgression patterns of goats in ISEA remain unclear due to limited geographic coverage and sample numbers in previous studies.

Therefore, this study aimed to (1) provide a comprehensive understanding of the genetic structure of goats by sampling from a wide range of ISEA locations, (2) estimate the genetic differences among maternal and paternal lineages, and (3) infer the processes of propagation and gene introgression in ISEA goats, using sequences of mtDNA HV1 (hypervariable segment 1) and the SRY gene on the Y chromosome.

Results

We sequenced the mtDNA HV1 region (481 bp) of 176 Philippine goats and 72 Indonesian goats and analyzed these sequences in conjunction with previously published mtDNA sequences. Supplementary Figures S1 and S2 show the alignment of the D-loop sequence with the reference goat sequence (Accession number AF533441.1). The sequence analysis identified 59 and 45 variant sites, as well as 32 and 19 mitochondrial haplotypes, in Philippine goats and Indonesian goats, respectively (Supplementary Figures S1 and S2). Subsequently, neighbor-joining trees were constructed using 30 previously published Philippine goat sequences and 22 reference goat sequences^{2,22}. Consequently, 206 Philippine goats were classified into haplogroups B ($n = 152/206$) and A ($n = 54/206$), while all 72 Indonesian goats belonged to haplogroup B (Supplementary Figure S3).

Figure 1 shows the median-joining networks of mtDNA haplotypes for Philippine and Indonesian goats with previously published Philippine and Indonesian goat sequences^{22,23}. In Philippine goats, the median-joining network showed that the haplotypes were classified into haplogroups A and B, consistent with the neighbor-joining tree. In haplogroup A, two moderately frequent haplotypes (ISEA1 and ISEA2) were observed, differing by 19 base pairs. In contrast, haplogroup B showed a star-like appearance with haplotype ISEA12 being the most frequent ($n = 95/152$) as a center (Fig. 1a).

In Indonesian goats, the median-joining network showed that haplotype ISEA14 ($n = 31/72$) was the most frequent, followed by ISEA34 ($n = 9/72$) and ISEA33 ($n = 7/72$) (Fig. 1b). Compared with Philippine goat haplotypes, ISEA14 and ISEA13 were observed also in Philippine goats. However, the predominant haplotype ISEA12 found in Philippine goats was absent in Indonesian goats.

Nucleotide diversity (π) and haplotype diversity (H_d) were calculated for Southeast Asian goat populations. Table 1 shows the nucleotide and haplotype diversities for haplogroups A and B in Southeast Asia. The Philippine goat population showed higher genetic diversity in haplogroup A ($\pi = 0.01390$, $H_d = 0.813$) compared to haplogroup B ($\pi = 0.00224$, $H_d = 0.582$). In the Indonesian goat population, the nucleotide and haplotype diversities for haplogroup B were 0.00286 (π) and 0.789 (H_d), respectively. In ISEA, Indonesian goats show higher genetic diversity in haplogroup B than Philippine goats. The nucleotide diversity for haplogroup A in Philippine goats was relatively low compared to other Southeast Asian goat populations, while haplogroup B in the Philippines showed the lowest nucleotide diversity across Southeast Asia. The nucleotide diversity in Indonesia was also relatively low compared to other Southeast Asian goat populations.

Figure 2 shows the distribution of mtDNA haplogroups in the Old World and seven regions of ISEA. This map highlights the increase in the frequency of haplogroup B toward the southeast, as previously described²³. In ISEA, haplogroup B was particularly prevalent in Mindanao Island ($n = 29/30$) and South Sulawesi ($n = 110/112$).

Table 2 presents the SRY haplotype frequencies for each region in ISEA, while Fig. 3 shows the distribution of SRY haplotypes in the Old World and seven regions of ISEA. This distribution reveals that goats in ISEA possess haplotypes Y1AA, Y1B, Y2A, and Y2B. Haplotype Y1B was observed exclusively on Mindoro Island, Philippines, likely due to the introduction of modern European breeds by the Philippine Department of Agriculture. Haplotype Y2A, which is absent in MSEA, was observed in all ISEA regions, with the highest frequency in Mindanao Island, Philippines ($n = 6/16$).

Approximate Bayesian Computation (ABC) analysis results indicated posterior probability values for Scenario 1, Scenario 2, and Scenario 3 of 0.1762 (95% CI 0.1703–0.1820), 0.3586 (95% CI 0.3523–0.3650),

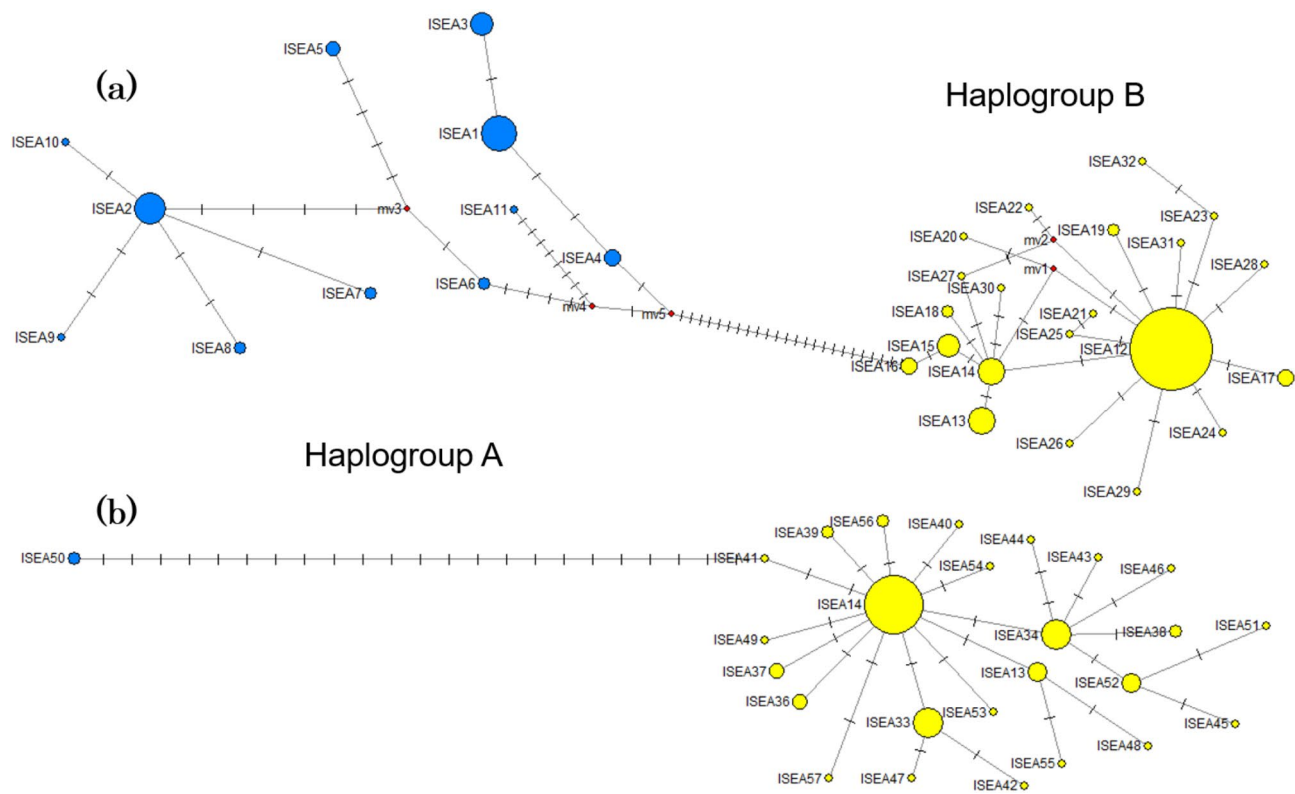


Fig. 1. Median joining network of 481 bp in the mtDNA HV1 sequence. Separator lines on the Network indicate mutation positions. Blue and yellow colors are shown haplogroup A and B, respectively. The size of nodes is proportional to the number of goats present in the node. Each short bar on the edges represents a nucleotide mutation. **(a)** Philippine goats; **(b)** Indonesian goats.

	Country	Individuals	Nucleotide diversity (π)	Haplotype diversity (h)
Haplogroup A	Bhutan	54	0.01758	0.856
	Myanmar	17	0.01835	0.993
	Laos	12	0.01711	0.909
	Cambodia	15	0.03485	0.837
	Vietnam	13	0.00981	0.859
	Philippines	55	0.01390	0.813
Haplogroup B	Bhutan	7	0.00337	0.905
	Myanmar	6	0.00804	1.000
	Laos	6	0.00430	0.933
	Cambodia	15	0.00230	0.552
	Vietnam	9	0.00289	0.806
	Malaysia	11	0.00318	0.745
	Indonesia	110	0.00286	0.789
	Philippines	152	0.00224	0.582

Table 1. Genetic diversity of haplogroup A and B in six Southeast Asian countries and eight Southeast Asian countries, respectively. Sequenced data used in this analysis are shown in Supplementary Table S5.

and 0.4652 (95% CI 0.4586–0.4717), respectively. Scenario 3, with the highest value (0.4652), indicates two propagation routes: one from China/Taiwan and another from MSEA via Indonesia to the Philippines, followed by admixture. However, the value of Scenario 3 is close to that of Scenario 2 (0.3586), so that the possibility of non-admixture history cannot be denied. In both cases, DIYABC analysis supports two propagation routes: from China/Taiwan to the Philippines and from MSEA to Indonesia.

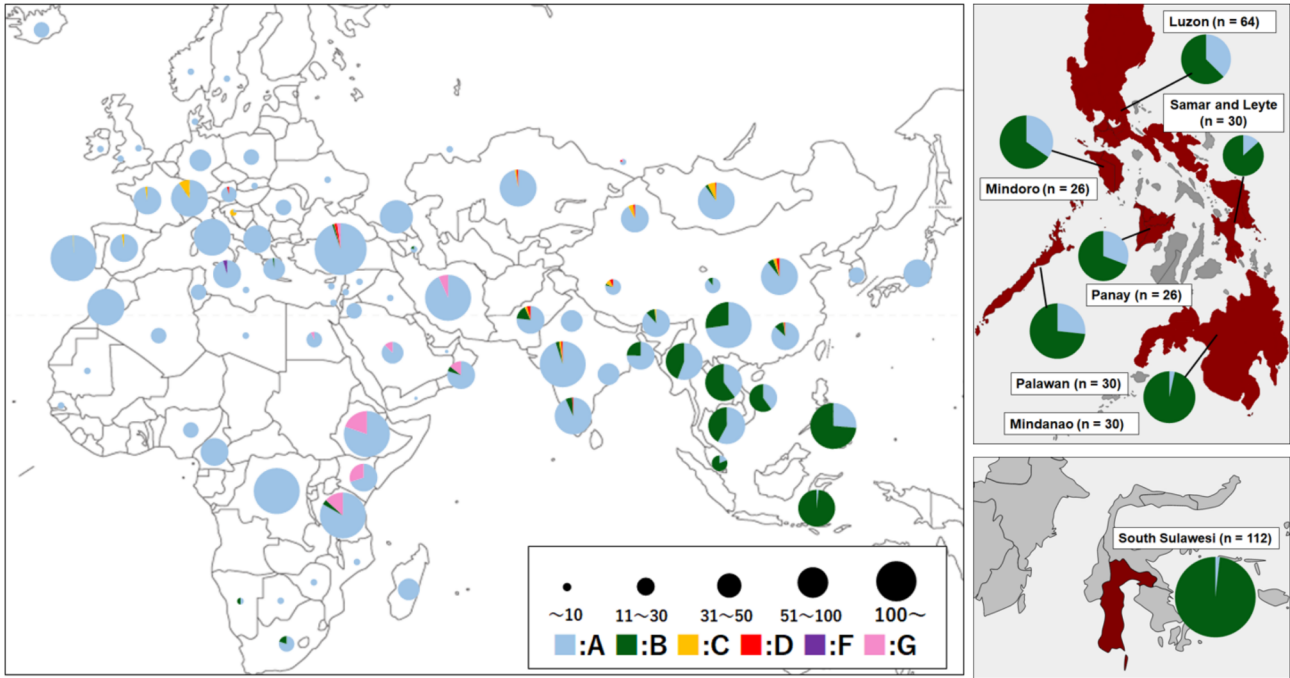


Fig. 2. Geographic distribution of caprine mtDNA haplogroups in the Old World and Island Southeast Asia. The size of nodes is proportional to the number of goats present in the node. The used data were shown in Table S5. The map was generated using Microsoft PowerPoint 2019 based on a free map website by 3kaku-K (<https://www.freemap.jp/item/world/world1.html>).

	Y1AA	Y1B	Y2A	Y2B
Philippines				
Luzon	16		1	7
Mindoro	11	7	1	4
Samar and Leyte	9		6	6
Panay	9		4	8
Palawan	17		4	6
Mindanao	8		6	2
Indonesia				
South Sulawesi	32		10	14

Table 2. SRY haplotype frequencies in Island Southeast Asia.

Discussion

The mtDNA haplogroup frequencies analyzed in this study revealed that haplogroup B in ISEA goats was more frequent than in MSEA (Fig. 2), although its genetic diversity was relatively lower compared to that of MSEA (Table 1). This lower genetic diversity may be attributed to a bottleneck effect in the propagation process due to the geographical location of ISEA. Haplogroup B was observed at higher frequencies particularly in the southeastern regions of ISEA, such as Mindanao and South Sulawesi. Additionally, the genetic diversity in the Philippines was lower than that in Indonesia, suggesting a migration event from South Sulawesi to Mindanao. Haplogroup B, predominantly observed in Southeast Asia, has also been reported at low frequencies in South African region: South Africa ($n=3/15$), Namibia ($n=2/4$), and Tanzania ($n=22/627$)^{2,24}. The absence of haplogroup B in African regions north than Tanzania suggests a potential propagation route by maritime trade in the Indian Ocean. The Indian Ocean has a long history of human maritime activity, with Islamic merchants trading and spreading Islam throughout the region since the 9th century, and European countries using the African coast as a transit point for maritime trade to Southeast Asia from the 16th century. Moreover, the migration of Austronesians from Indonesia to East Africa and the Arabian Peninsula due to this maritime trade has been documented^{25,26}. Therefore, the presence of haplogroup B in the South African region is likely associated with human migration and trade in the Indian Ocean since the Middle Ages. A previous study analyzing SRY haplotypes worldwide found that Y1AA, Y1AB, and Y2B spread from the Fertile Crescent to East and Southeast Asia, while Y1B and Y2A emerged during the expansion process in Europe

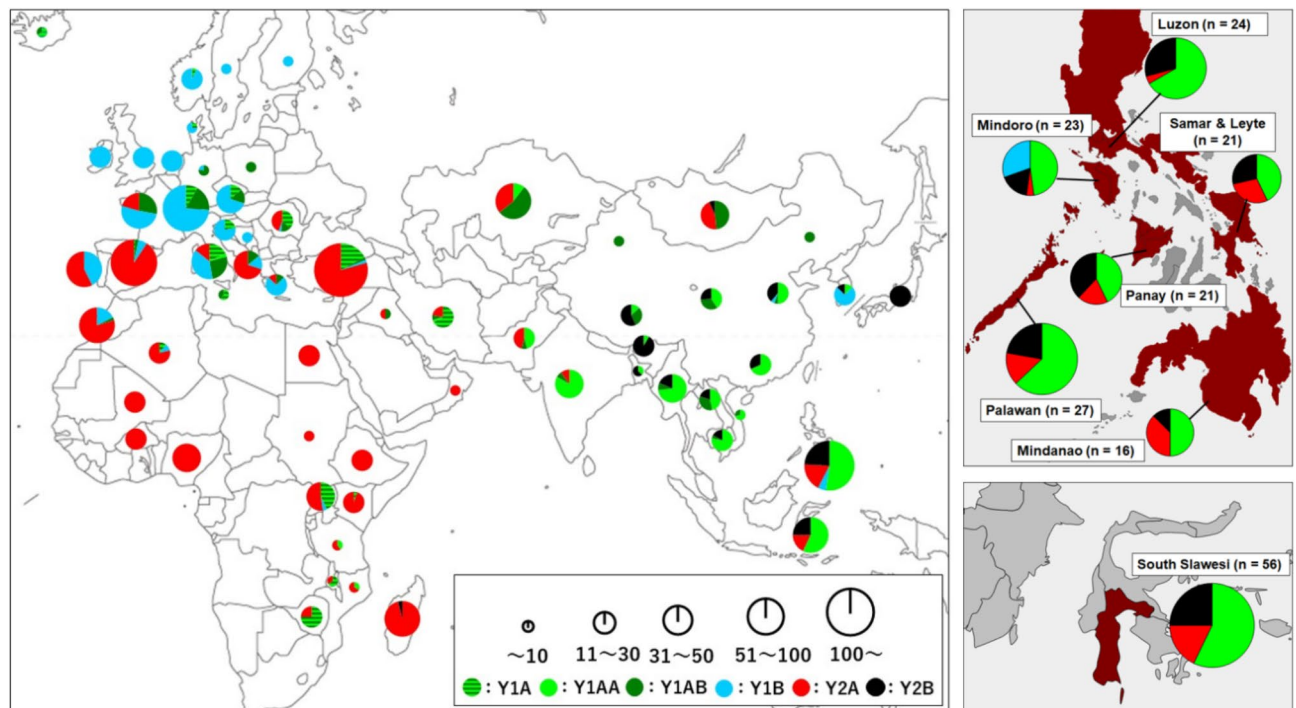


Fig. 3. Geographic distribution of caprine SRY haplotypes in the Old World and Island Southeast Asia. The size of nodes is proportional to the number of goats present in the node. The used data were shown in Table S6. The map was generated using Microsoft PowerPoint 2019 based on a free map website by 3kaku-K (<https://www.freemap.jp/item/world/world1.html>).

and Africa, respectively¹⁷. This study revealed that haplotype Y2A is prevalent across the entire ISEA (Fig. 3). Haplotype Y2A, a major haplotype in Africa and southern Europe, has not been observed in MSEA. Therefore, haplotype Y2A in ISEA has been genetically influenced by goats introduced from southern Europe and Africa, regions that served as base camps and relay ports during the Age of Discovery. Moreover, our results revealed regional differences in the frequency of haplotype Y2A in ISEA, which may reflect the gene flow process of Y2A. A worldwide SRY analysis also identified the East Asian-specific haplotype Y2B in northern Madagascar¹⁷.

Studies on the migration routes of canines and swine provide useful insights into the propagation of goats to Island Southeast Asia^{27,28}. Zhang et al.²⁷ suggested two possible routes for canine propagation: one through Taiwan to the Philippines and ultimately to Australia, and another through MSEA to Indonesia and then to Australia. These propagation are estimated to have occurred between 5000 and 2000 years ago. Similarly, Layos et al.²⁸ proposed two possible routes for swine propagation: one through Taiwan to the Philippines (during the Neolithic) and another through MSEA to the Philippines (during the prehistoric times). The propagation of domestic animals is thought to have accompanied the migration of the Austronesian language family, estimated to have occurred after 5000 years ago in the Philippines and between 4000 and 3000 years ago in Indonesia²⁹. The Philippines, located at the crossroads of past human migrations in the Asia-Pacific region, has shown genetic effects from several times of human migration³⁰. Indonesians are suggested to have experienced parallel introductions of MSEA- and Austronesian-related ancestry sources between 1000 and 2500 years ago^{31,32}.

Considering the migration and expansion of humans, dogs, and pigs, goats may have migrated through similar routes. The distribution of mtDNA haplogroups in ISEA showed that the frequency of haplogroup B tended to increase toward the southeast (Fig. 2). Within ISEA, haplogroup B is particularly prevalent in South Sulawesi and Mindanao, suggesting possible propagation between these regions due to their geographical proximity. Taking these results into consideration, a plausible hypothesis for goat propagation in ISEA is that haplogroup A arrived in the Philippines via China/Taiwan, while goats with haplogroup B flowed from the Malay Peninsula to the Philippines via Indonesian islands. Simulation results from DIYABC indicated a low probability for a single propagation route to ISEA via Indochina (Fig. 4). Although Scenario 3, with the highest posterior probability value, suggests genetic admixture in the Philippines, the actual occurrence of this admixture event remains unclear, given the close posterior probability values between Scenario 3 (0.4652) and Scenario 2 (0.3586).

Paternal analysis revealed that haplotype Y2A, which is not found in MSEA, was observed in ISEA. The ISEA has historically been an important trading hub, especially in the spice trade, linking Europe, China, India, and the Arab world. This was due to the activities of Muslim merchants from the ninth century, followed by the Spanish and Dutch colonization of the Philippines and Indonesia, respectively, from the 16th century^{19,33,34}. Therefore, it is possible that some exotic goats were introduced to the ISEA via maritime trade from the Middle Ages onwards in the Indian Ocean, from Europe, Africa, and India. Furthermore, the Maluku Islands in eastern Indonesia, the original source of clove and nutmeg, and South Sulawesi and Mindanao, which are located nearby,

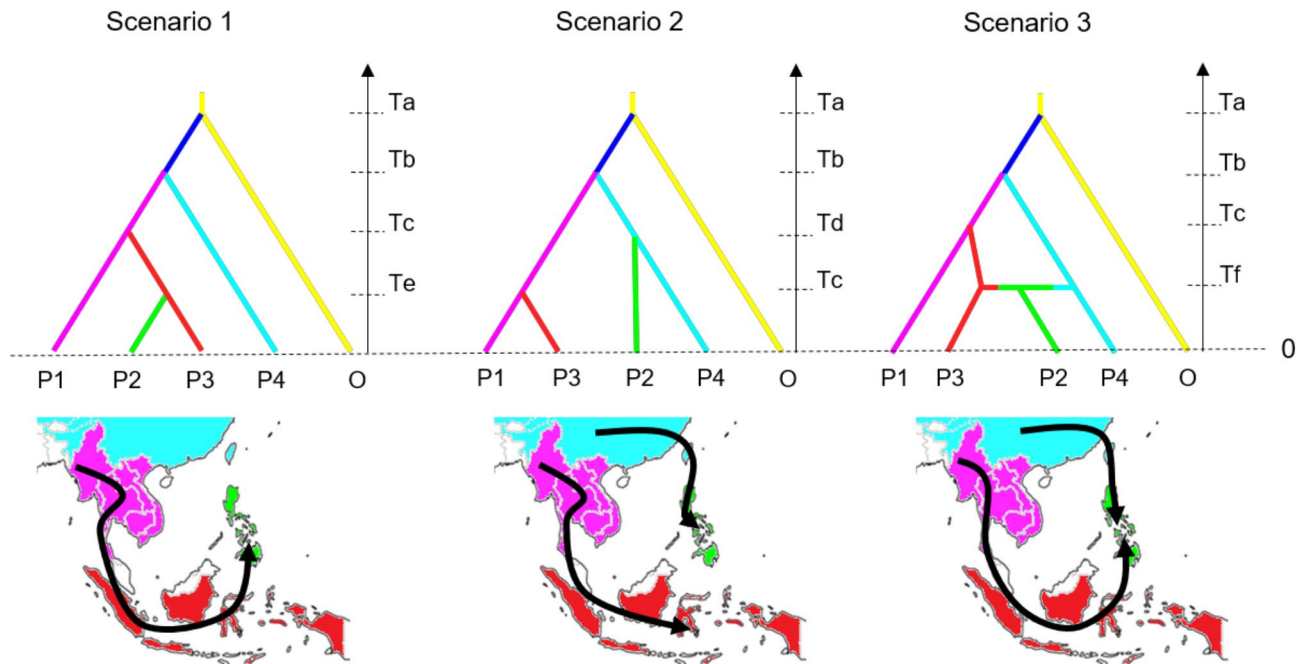


Fig. 4. Evolutionary scenarios for the propagation route of ISEA goats. Scenario 1: ISEA goats were introduced from MSEA. Scenario 2: Philippine and Indonesian goats were independently introduced from China/Taiwan and MSEA, respectively. Scenario 3: Philippine goats were introduced by two routes, from China/Taiwan and from MSEA via Indonesia, with admixture occurring in the Philippines. Upper figures show scenarios indicated by the trees; lower figures show geographic regions on the maps. Colors of the tree correspond to the geographic regions on the map (Green: Philippines; Red: Indonesia; Light blue: China; Pink: MSEA; Yellow: Outgroup). The map was generated using Microsoft PowerPoint 2019 based on a free map website by 3kaku-K (<https://www.freemap.jp/item/world/world1.html>).

were served as trading centers by European countries. This may explain the relatively high frequency of Y2A in these regions (Fig. 3). The presence of mtDNA haplogroup B and the SRY Y2B haplotype in the South African region and northern Madagascar further supports gene introgression between the South African region and ISEA through Indian Ocean maritime trade.

This hypothesis is supported by previous research on chickens using mtDNA, which demonstrated that human migrations across the Indian Ocean led to gene flow of chickens from Southeast Asia to East Africa³⁵. There are also historical records of chickens being transported aboard ships as a source of protein during voyages in the Medieval Period^{36,37}. Similarly, goats were used as a protein source on maritime expeditions during the Medieval Period³⁶, and comparable gene flow may have occurred in goats. Therefore, the genetic structure of domestic goats in the ISEA may reflect the influence of human maritime trade during the Medieval Period, suggesting that their genetic structures reflect the extent of maritime trade activities.

In this study, we estimated the genetic structure and distribution patterns of goats in the Old World using maternal and paternal analyses. Our results suggested (1) two main migration routes for ISEA goats, and (2) reciprocal gene flow between the ISEA and the South African region through transoceanic trade across the Indian Ocean. The genetic information obtained from this study provides important insights into the conservation of genetic resources and diversity of goats in the Old World, particularly in the ISEA.

Materials and methods

Ethical statement

This study is reported in accordance with ARRIVE guidelines (<https://arriveguidelines.org>). This study was approved by the University of the Philippines, Los Banos, under the attestations the H23Oct285/2–9 and the Hassanuddin University, Indonesia, under the attestations the 15/25Jun2018. All experiments were conducted in accordance with the Kobe University Animal Experimentation Regulations, and all protocols were approved by the Institutional Animal Care and Use Committee of Kobe University and by Association for the Promotion of Research Integrity (Approval Number: AP0000436777). All blood sample collections were approved by the animal owners with informed consent.

Animals

A total of 248 blood samples were collected from indigenous goats in the Philippines and Indonesia (Fig. 2 and Supplementary Fig. S4). In the Philippines, 176 blood samples were collected from seven islands and six regions: 34 from Luzon, 26 from Mindoro, 30 from Samar and Leyte, 26 from Panay, 30 from Palawan, and 30 from Mindanao (Table S1). In Indonesia, 72 blood samples were collected from South Sulawesi (Table S2). Efforts

were made to avoid sampling from related individuals. DNA was extracted from the blood samples using the standard phenol/chloroform method³⁸.

Sequencing

The amplification and sequencing of the mtDNA HV1 region (481 bp) and the SRY 3'UTR region (478 bp) were performed following the procedures described previously^{8,11,17,39,40} (Tables S3 and S4). All sequences have been deposited in the DDBJ database (accession numbers: LC849824–LC850069, LC856305, LC856306).

Statistical analysis

Sequence alignment of the HV1 region was performed according to a previous study³⁹. The sequence of the mtDNA HV1 region was aligned using the ClustalW program⁴¹ in MEGA 7.0⁴², based on the mtDNA reference sequence (DDBJ accession no. AF533441.1)⁴³. An unrooted neighbor-joining phylogenetic tree was constructed using the Tamura-Nei distance method⁴². The distance computation and phylogenetic tree construction were performed in MEGA 7.0. The phylogenetic trees were assessed by bootstrap percentages computed with 1000 replications⁴⁴. For phylogenetic tree construction, 22 reference sequences from six goat haplogroups were used as references² (DDBJ accession nos. AY155721, EF618134, EF617779, EF618200, EF617945, EF617965, AB044303, EF617706, AJ317833, DQ121578, AY155708, AJ317838, EF618413, DQ188892, AY155952, EF617701, DQ188893, DQ241349, DQ241351, EF618084, EF618535, EF617727). Median-joining networks⁴⁵ were constructed using Network 5.0.0.1 (https://www.fluxus-engineering.com/sharenet_rn.htm). Nucleotide diversity (π) and haplotype diversity (H_d) were calculated for HV1 sequences, in conjunction with sequences from Southeast Asian native goats deposited in public databases (Table S5) using DnaSP 6.12.03⁴⁶.

To infer relationships between mtDNA haplogroups in the ISEA and other regions, we analyzed the distribution of maternal haplogroups in the Eurasian and African continents using previously published mtDNA data from 5939 goats. The used data were shown in Table S5.

The distribution of paternal haplotypes in Eurasia and Africa was analyzed using SRY data from 2001 goats deposited in public databases (Table S6). The SRY haplotype data for native goats from the Philippines and Indonesia were previously published in Vargos Consortium et al.¹⁷. In this study, we calculated the frequency of paternal haplotypes in seven regions within the ISEA and inferred relationships between the ISEA and other regions.

Evolutionary scenarios on the propagation route of ISEA goats were evaluated using Approximate Bayesian computation (ABC) with the DIYABC program ver. 2.1.0⁴⁷, based on the mtDNA HV1 region. Three alternative scenarios were devised to assess the evolutionary relationships among four populations: the Philippines, Indonesia, China, and MSEA. Scenario 1: ISEA goats were propagated from MSEA. Scenario 2: Goats in the Philippines and Indonesia propagated from China/Taiwan and MSEA, respectively. Scenario 3: Philippine goats propagated via two routes—China/Taiwan and MSEA via Indonesia—and admixture occurred in the Philippines.

The following parameters were set: T_a , the time before goat domestication (10,000–470,000 years ago); T_b , the time when goat domestication occurred (10,000–15,000 years ago); T_c and T_d , the times when goats propagated from MSEA to Indonesia and from China/Taiwan to the Philippines, respectively. We assumed T_c to be from 4000 years ago and T_d to be between 5000 and 10,000 years ago. Other parameters were left as default. The mitochondrial genome sequence of haplogroup F of the Bezoar (*Capra aegagrus*) was used as the outgroup⁷. The mutation rate was estimated using the dataset from Colli et al.⁷ and was set between 3.75×10^{-8} /site/generations and 3.75×10^{-6} /site/generations. Simulations were conducted 3 million times with the HKY + I + Γ model under the three scenarios. To identify the most likely scenario, relative posterior probability with 95% confidence intervals (95% CI) was estimated for each scenario. Model validity was checked using the perform model checking function in DIYABC. Principal component analysis in DIYABC showed that the observed dataset belongs to the point cluster of posterior distribution (Supplementary Fig. S5), indicating that the model-posterior combination fits the observed data.

Data availability

The 278 mtDNA hypervariable 1 sequences are deposited and available in DDBJ database (accession numbers: LC849824–LC850069, LC856305, LC856306).

Received: 26 November 2024; Accepted: 7 March 2025

Published online: 19 March 2025

References

1. Luikart, G. et al. Multiple maternal origins and weak phylogeographic structure in domestic goats. *Proc. Natl. Acad. Sci.* **98**, 5927–5932 (2001).
2. Naderi, S. et al. Large-scale mitochondrial DNA analysis of the domestic goat reveals six haplogroups with high diversity. *PLoS One.* **2**, e1012 (2007).
3. Daly, K. G. et al. Ancient goat genomes reveal mosaic domestication in the fertile crescent. *Science* **361**, 85–88 (2018).
4. Mannen, H., Nagata, Y. & Tsuji, S. Mitochondrial DNA reveal that domestic goat (*Capra hircus*) are genetically affected by two subspecies of bezoar (*Capra aegagrus*). *Biochem. Genet.* (2001).
5. Lenstra, J. Evolutionary and demographic history of sheep and goats suggested by nuclear, mtDNA and Y-chromosome markers. (2005).
6. Cinar Kul, B. et al. Y-chromosomal variation of local goat breeds of Turkey close to the domestication centre. *J. Anim. Breed. Genet. Z. Tierzucht Zuchtungsbiologie.* **132**, 449–453 (2015).
7. Colli, L. et al. Whole mitochondrial genomes unveil the impact of domestication on goat matrilineal variability. *BMC Genom.* **16**, 1115 (2015).
8. Waki, A., Sasazaki, S., Kobayashi, E. & Mannen, H. Paternal phylogeography and genetic diversity of East Asian goats. *Anim. Genet.* **46**, 337–339 (2015).

9. Joshi, M. B. et al. Phylogeography and origin of Indian domestic goats. *Mol. Biol. Evol.* **21**, 454–462 (2004).
10. Chen, S. Y., Su, Y. H., Wu, S. F., Sha, T. & Zhang, Y. P. Mitochondrial diversity and phylogeographic structure of Chinese domestic goats. *Mol. Phylogenet. Evol.* **37**, 804–814 (2005).
11. Lin, B. Z. et al. Molecular phylogeography and genetic diversity of East Asian goats. *Anim. Genet.* **44**, 79–85 (2013).
12. Naderi, S. et al. The goat domestication process inferred from large-scale mitochondrial DNA analysis of wild and domestic individuals. *Proc. Natl. Acad. Sci.* **105**, 17659–17664 (2008).
13. Zhao, Y. et al. Genetic diversity and molecular phylogeography of Chinese domestic goats by large-scale mitochondrial DNA analysis. *Mol. Biol. Rep.* **41**, 3695–3704 (2014).
14. Cañón, J. et al. Geographical partitioning of goat diversity in Europe and the middle East. *Anim. Genet.* **37**, 327–334 (2006).
15. Pereira, F. et al. A multiplex primer extension assay for the rapid identification of paternal lineages in domestic goat (*Capra hircus*): Laying the foundations for a detailed caprine Y chromosome phylogeny. *Mol. Phylogenet. Evol.* **49**, 663–668 (2008).
16. Pereira, F. et al. Tracing the history of goat pastoralism: new clues from mitochondrial and Y chromosome DNA in North Africa. *Mol. Biol. Evol.* **26**, 2765–2773 (2009).
17. VarGoats et al. Geographical contrasts of Y-chromosomal haplogroups from wild and domestic goats reveal ancient migrations and recent introgressions. *Mol. Ecol.* **31**, 4364–4380 (2022).
18. Skowronek, R. K. The Spanish Philippines: archaeological perspectives on colonial economics and society. *Int. J. Hist. Archaeol.* **2**, 45–71 (1998).
19. Ricklefs, M. C., Lockhart, B. & Lau, A. *A New History of Southeast Asia* (Bloomsbury Academic, 2010).
20. Mbeki, L. & van Rossum, M. Private slave trade in the Dutch Indian ocean world: a study into the networks and backgrounds of the slavers and the enslaved in South Asia and South Africa. *Slavery Abolit.* **38**, 95–116 (2017).
21. Devendra, C. & Nozawa, K. Goats in South East Asia—their status and production. *Z. Für Tierz. Zücht.* **93**, 101–120 (1976).
22. Kato, T. et al. Mitochondrial genetic diversity of goat in South Eastern Asia. **84**, 149–155 (2013).
23. Mannen, H. et al. Indonesian native goats (*Capra hircus*) reveal highest genetic frequency of mitochondrial DNA haplogroup B in the world. *Anim. Sci. J.* **91**, (2020).
24. Nguluma, A. et al. Mitochondrial DNA D-loop sequence analysis reveals high variation and multiple maternal origins of Indigenous Tanzanian goat populations. *Ecol. Evol.* **11**, 15961–15971 (2021).
25. Brucato, N. et al. Evidence of Austronesian genetic lineages in East Africa and South Arabia: complex dispersal from Madagascar and Southeast Asia. *Genome Biol. Evol.* **11**, 748–758 (2019).
26. Choudhury, A., Aron, S., Sengupta, D., Hazelhurst, S. & Ramsay, M. African genetic diversity provides novel insights into evolutionary history and local adaptations. *Hum. Mol. Genet.* **27**, R209–R218 (2018).
27. Zhang, W. et al. Identification of signatures of selection by Whole-Genome resequencing of a Chinese native pig. *Front. Genet.* **11**, 566255 (2020).
28. Layos, J. K. N. et al. Origin and demographic history of Philippine pigs inferred from mitochondrial DNA. *Front. Genet.* **12**, 823364 (2022).
29. Bellwood, P. S. *Man's Conquest of the Pacific: the Prehistory of Southeast Asia and Oceania* (Oxford University Press, 1979).
30. Larena, M. et al. Multiple migrations to the Philippines during the last 50,000 years. *Proc. Natl. Acad. Sci.* **118**, e2026132118 (2021).
31. Hudjashov, G. et al. Complex patterns of admixture across the Indonesian Archipelago. *Mol. Biol. Evol.* **34**, 2439–2452 (2017).
32. Taufik, L. et al. Human genetic research in Wallacea and Sahul: recent findings and future prospects. *Genes* **13**, 2373 (2022).
33. Chias, P. & Abad, T. Colonial urban planning and land structures in the Philippines, 1521–1898. *J. Asian Archit. Build. Eng.* **11**, 9–16 (2012).
34. Zuhdi, S. Shipping routes and spice trade in Southeast Sulawesi in the 17th and 18th century. *J. Marit Stud. Natl. Integr.* **2**, 31–44 (2018).
35. Mwacharo, J. M. et al. Mitochondrial DNA reveals multiple introductions of domestic chicken in East Africa. *Mol. Phylogenet. Evol.* **58**, 374–382 (2011).
36. Lobo, J., Costa, M. G., da, Beckingham, C. F., Charles, F. & Lockhart, D. M. *Itinerário, e Outros Escritos Inéditos*. (Livreria Civilização, (1971).
37. Casimiro, T. M. & Borges, M. O. Life on board Portuguese ships in the 16th–18th centuries: Theorizing households through history and archaeology. *Heritage* **6**, 2020–2037 (2023).
38. Sambrook, J. & Russell, D. *Molecular Cloning: a Laboratory Manual* (Cold Spring Harbor Laboratory Press, 2001).
39. Tabata, R. et al. Phylogeographic analysis of Madagascan goats using MtDNA control region and SRY gene sequences. *Zoolog Sci.* **36**, 294 (2019).
40. Tabata, R. et al. The Eurasian steppe is an important goat propagation route: A phylogeographic analysis using mitochondrial DNA and Y-chromosome sequences of Kazakhstani goats. *Anim. Sci. J.* **90**, 317–322 (2019).
41. Thompson, J. D., Higgins, D. G. & Gibson, T. J. CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice.
42. Kumar, S., Stecher, G. & Tamura, K. MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. *Mol. Biol. Evol.* **33**, 1870–1874 (2016).
43. Pietro, P., Maria, F., GianFranco, G. & Giuseppe, E. The complete nucleotide sequence of goat (*Capra hircus*) mitochondrial genome: goat mitochondrial genome. *DNA Seq.* **14**, 199–203 (2003).
44. Felsenstein, J. Phylogenies and the comparative method. *Am. Nat.* **125**, 1–15 (1985).
45. Bandelt, H. J., Forster, P. & Rohl, A. Median-joining networks for inferring intraspecific phylogenies. *Mol. Biol. Evol.* **16**, 37–48 (1999).
46. Rozas, J. et al. DnaSP 6: DNA sequence polymorphism analysis of large data sets. *Mol. Biol. Evol.* **34**, 3299–3302 (2017).
47. Cornuet, J. M. et al. DIYABC v2.0: a software to make approximate bayesian computation inferences about population history using single nucleotide polymorphism, DNA sequence and microsatellite data. *Bioinform. Oxf. Engl.* **30**, 1187–1189 (2014).

Acknowledgements

The manuscript has been carefully reviewed by an experienced editor (Enago, Crimson Interactive Inc.) whose first language is English and who specializes in editing papers written by scientists whose native language is not English.

Author contributions

H.M. conceived and designed the experiments and supervised the project. R.M., A., M.M., F.K., S.S. performed the genetic analyses. R.M., T.Y., A., H.M. performed the phylogenetic analyses. M.I.A.D., S.R.A.B., J.S.M. conducted the field investigation and collected the samples. R.M., T.Y., H.M. wrote the manuscript. All of the authors discussed the results and contributed to the final manuscript.

Ethics declarations

Competing interests

The authors declare no competing interests.

Additional information

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1038/s41598-025-93651-9>.

Correspondence and requests for materials should be addressed to H.M.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Open Access This article is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License, which permits any non-commercial use, sharing, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if you modified the licensed material. You do not have permission under this licence to share adapted material derived from this article or parts of it. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by-nc-nd/4.0/>.

© The Author(s) 2025