



Draft Genome Sequence of *Chromobacterium violaceum* RDN09, Isolated from a Patient with a Wound Infection in Bangladesh

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ABSTRACT *Chromobacterium violaceum* is an emerging environmental opportunistic pathogen that causes life-threatening infections in humans. Here, we describe the draft genome sequence of *Chromobacterium violaceum* RDN09, isolated from the infected wound of an adult male patient in Bangladesh. The genome assembly consists of 4,736,739 bp spread across 84 contigs.

Chromobacterium violaceum is a betaproteobacterium and a member of the family *Neisseriaceae*. It is a saprophyte predominantly present in the natural ecosystems of tropical and subtropical countries of the world (1). This bacterium is known for the production of violacein, a purple-colored pigment produced as a result of quorum sensing (2). Human infections with *C. violaceum* are rare; however, if contracted, it can cause severe systemic infections with high mortality rates (~60%) (3). Infections manifest in the form of cellulitis and skin abscesses that can rapidly progress to sepsis, septic shock, and multiple abscesses in the vital organs (4). Its pathogenicity in the mammalian infection model is attributable to the presence of several virulence factors, including two type III secretion systems (T3SSs) (5).

The strain RDN09 was cultured from a pus specimen obtained from an infected wound in the leg of an adult male patient who had suffered an agricultural injury (6). The strain produced a violet pigment and was a Gram-negative coccobacillus (6). Genomic DNA was extracted from a 24-h culture that originated from a single colony of *C. violaceum* RDN09 grown at 37°C in Luria-Bertani broth, using the QIAamp DNA minikit (Qiagen) and following the Gram-negative bacterial DNA isolation procedure (6). DNA quality was assessed using a NanoDrop spectrophotometer (Thermo Scientific, USA), and quantification was carried out using a Qubit 2.0 fluorimeter (Life Technologies). The sequencing library was prepared from 1 ng genomic DNA using an Illumina Nextera XT DNA library preparation kit as per the manufacturer's instructions and sequenced on the Illumina NextSeq 500 platform employing the Illumina NextSeq v2.5 reagent kit (2 × 150 bp). Quality checks on the paired-end sequencing reads (150 bp) were performed using FastQC v0.11.11. The genome coverage was found to be 132× by mapping the reads against the reference genome *Chromobacterium violaceum* ATCC 12472 (7). Trimmomatic v1.01 was used for adapter trimming based on quality scores of Q30 with the following parameters applied: SLIDINGWINDOW:5:15; LEADING:5;TRAILING:5;MINLEN:36;ILLUMINACLIP:path_to_adaptors_sequences/adapter.fasta:2:30:10 (8). *De novo* assembly was conducted using SPAdes v3.11.1 (9). QUAST v5.0.2 was used for quality assessment of the assembly (10). Prokka v1.12 was utilized to annotate the genome with *C. violaceum* ATCC 12472 as a reference genome (GenBank accession number [NC_005085.1](https://ncbi.nlm.nih.gov/nuccore/NC_005085.1)) (11). Default parameters were applied for all software unless otherwise mentioned. The genome sequence of *C. violaceum* strain RDN09 is 4,736,739 bp with a G+C content of 64.83%. It is made up of 84 contigs, having 39 contigs larger than 1,000 bp, and the contig N_{50} value is 203,983 bp. A total of 4,316 coding DNA sequences (CDSs) were identified, of which 56% were assigned a putative function, and

Citation Mazumder R, Abdullah A, Hussain A, Ahmed D, Mondal D. 2020. Draft genome sequence of *Chromobacterium violaceum* RDN09, isolated from a patient with a wound infection in Bangladesh. *Microbiol Resour Announc* 9:e00957-20. <https://doi.org/10.1128/MRA.00957-20>.

Editor Frank J. Stewart, Georgia Institute of Technology

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Received 15 August 2020

Accepted 28 September 2020

Published 15 October 2020

the remaining CDSs were annotated as hypothetical. We identified 4 rRNA and 83 tRNA sequences in the *Chromobacterium violaceum* strain RDN09 genome sequence. We were not able to identify any intact prophage regions using the PHASTER algorithm, a new version of PHAST (12). The genome sequence of *Chromobacterium violaceum* strain RDN09 exhibited 92.97% similarity with *C. violaceum* strain ATCC 12472 and was found to harbor genetic elements associated with the production of secondary metabolites. This genome sequence showed the presence of a virulence-related type III secretion gene cluster. Unlike the genome of ATCC 12472, which showed the presence of two T3SSs encoded by Cpi-1/-1a and Cpi-2 (5), strain RDN09 showed the presence of only one T3SS encoded by Cpi-1/-1a. The genome sequence described in this article and future comparative genomic studies will be useful in improving our understanding of the biology and pathogenicity of *C. violaceum*. Such studies will also help decipher the virulence mechanisms and evolution of this emerging pathogen.

Data availability. The bacterial whole-genome shotgun project for *Chromobacterium violaceum* RDN09 has been deposited under the BioProject number [PRJNA640761](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA640761), BioSample number [SAMN15332061](https://www.ncbi.nlm.nih.gov/biosample/SAMN15332061), and SRA accession number [PRJNA640761](https://www.ncbi.nlm.nih.gov/sra/PRJNA640761) and in DDBJ/EMBL/GenBank under the accession number [JABXOB000000000](https://www.ncbi.nlm.nih.gov/nuccore/JABXOB000000000). The version described in this article is the first version ([JABXOB000000000.1](https://www.ncbi.nlm.nih.gov/nuccore/JABXOB000000000)).

ACKNOWLEDGMENTS

This research study was funded by core donors, who provide unrestricted support to icddr,b for its operations and research. The current donors providing unrestricted support include the governments of Bangladesh, Canada, Sweden, and the United Kingdom. We gratefully acknowledge our core donors for their support and commitment to icddr,b's research efforts.

REFERENCES

1. Sneath PHA, Singh RB, Whelan JPF, Edwards D. 1953. Fatal infection by *Chromobacterium violaceum*. *Lancet* 262:276–277. [https://doi.org/10.1016/S0140-6736\(53\)91132-5](https://doi.org/10.1016/S0140-6736(53)91132-5).
2. Antônio RV, Creczynski-Pasa TB. 2004. Genetic analysis of violacein biosynthesis by *Chromobacterium violaceum*. *Genet Mol Res* 3:85–91.
3. de Siqueira IC, Dias JP, Ruf H, Ramos EAG, Maciel EAP, Rolim A, Jabur L, Vasconcelos L, Silvany C. 2005. *Chromobacterium violaceum* in siblings, Brazil. *Emerg Infect Dis* 11:1443–1445. <https://doi.org/10.3201/eid1109.050278>.
4. Baker S, Campbell JI, Stabler R, Nguyen HVM, To DS, Nguyen DV, Farrar J. 2008. Fatal wound infection caused by *Chromobacterium violaceum* in Ho Chi Minh City, Vietnam. *J Clin Microbiol* 46:3853–3855. <https://doi.org/10.1128/JCM.01068-08>.
5. Batista JH, da Silva Neto JF. 2017. *Chromobacterium violaceum* pathogenicity: updates and insights from genome sequencing of novel *Chromobacterium* species. *Front Microbiol* 8:2213. <https://doi.org/10.3389/fmicb.2017.02213>.
6. Mazumder R, Sadique T, Sen D, Mozumder P, Rahman T, Chowdhury A, Halim F, Akter N, Ahmed D. 2020. Agricultural injury-associated *Chromobacterium violaceum* infection in a Bangladeshi farmer. *Am J Trop Med Hyg* 103:1039–1042. <https://doi.org/10.4269/ajtmh.20-0312>.
7. Andrews S. 2017. FastQC: a quality control tool for high throughput sequence data. <https://www.bioinformatics.babraham.ac.uk/projects/fastqc/>.
8. Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
9. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
10. Mikheenko A, Prjibelski A, Saveliev V, Antipov D, Gurevich A. 2018. Versatile genome assembly evaluation with QUAST-LG. *Bioinformatics* 34:i142–i150. <https://doi.org/10.1093/bioinformatics/bty266>.
11. Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. *Bioinformatics* 30:2068–2069. <https://doi.org/10.1093/bioinformatics/btu153>.
12. Arndt D, Marcu A, Liang Y, Wishart DS. 2019. PHAST, PHASTER and PHASTEST: tools for finding prophage in bacterial genomes. *Brief Bioinform* 20:1560–1567. <https://doi.org/10.1093/bib/bbx121>.