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Effect of urbanization on zoonotic gastrointestinal parasite prevalence in endemic toque macaque (*Macaca sinica*) from different climatic zones in Sri Lanka

Shehani U. Fernando^a, PreethiV. Udagama^a, Saminda P. Fernando^{b,*}

^a Department of Zoology and Environment Sciences, Faculty of Science, University of Colombo, Colombo 03, Sri Lanka
^b Department of Zoology, Faculty of Natural Sciences, The Open University of Sri Lanka, Nawala, Nugegoda, Sri Lanka

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ABSTRACT

Understanding variations in host-parasite relationships with urbanization is vital for both, public health management and conservation of endemic animals with high anthropogenic interactions. Toque macaques (Macaca sinica) are such endemic old-world monkeys in Sri Lanka. Three macaque sub species inhabit the main climatic zones of the island; M. s. sinica, M. s. aurifrons and M. s. opisthomelas inhabit the dry zone, wet zone, and montane regions of the island, respectively. This study aimed to examine parasite prevalence in this host in association with urbanization. A total of 180 fecal samples were collected from the three sub species of toque macaques inhabiting the main climatic zones (dry, wet, and montane) in Sri Lanka; 20 samples each were collected from urban, suburban, and wild populations representing each climatic zone. Twenty gastrointestinal (GI) parasite genera types i.e. five types of protozoan cysts, two types of trematode ova, four types of cestode ova, eight types of nematode ova, and a single type of acanthocephalan ova were identified. The overall prevalence of parasites was 62% (112/180) with the highest prevalence of Strongyloides infection. In all three sub species, toque macaque populations with proximity to human settlements, including urban and suburban populations, manifested a greater GI parasitic prevalence, mean ova/cyst counts and species richness, compared to their wild counterparts. Importantly, records of five parasite types in toques in Sri Lanka are reported for the first time, while Moniliformis type was reported as a first record in free ranging macaques, globally. This study clearly demonstrated that human contact and habitat modification may influence patterns of parasitic infections in macaques. As most of the parasite types identified manifest zoonotic potential, with public health implications, close associations of macaques may cause a threat to human well-being.

1. Introduction

During the past few decades, rapid intensification, and fragmentation of ecosystems through urbanization, were responsible for the rapid and comprehensive change in parasites of primates (Moll et al., 2019). Such modifications and alterations in natural habitats impact upon primate survivability, and susceptibility to parasitic infection, as parasites can influence the health (Johnson and Hoverman, 2012), population size (Tompkins et al., 2002), reproductive condition (Auld et al., 2016), behaviour and social interactions (Herbison et al., 2018) of primates. Due to the loss and fragmentation of their natural habitats, primates are forced to live in anthropogenically disturbed urban and suburban areas with proximity to humans. Thus, continuous human disturbances and encroachment, the ability of some parasites to infect multiple host species, and high level of genetic homology between humans and primates, increase the emergence and transmission of parasitic diseases from primates to humans and *vice versa* (Jones-Engel et al., 2004; Lane et al., 2011; Amarasinghe and Premathilake, 2014). Such interactions provide cause for serious concern on primate conservation as well as on public health. Thus, it is important to emphasize on the effects of human induced landscape changes on GI parasite infection in primates.

Several published studies had focused on parasites on free-ranging, semi-captive and captive orangutans, *M. fascicularis*, and *M. nemestrina* (Mul et al., 2007; Adrus et al., 2018) as well as on several semi-captive and wild primate species (Warren, 2001; Milozzi et al., 2012). Although

* Corresponding author. *E-mail addresses:* shehanisu1994@gmail.com (S.U. Fernando), preethi@zoology.cmb.ac.lk (PreethiV. Udagama), saminda@ou.ac.lk (S.P. Fernando).

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anthropogenic activities are likely to play a role in primate-parasite associations, none of the previous studies had examined the variations in parasitic infections in the same primate species that differed in their contacts and interactions with humans. Hence, to widen the scope of such investigations, this study specifically focused on the effects of human induced landscape changes on GI parasite infection in toque macaques.

Toque macaque (*Macaca sinica*) in Sri Lanka is a perfect model for assessing the influence of the anthropogenic landscape on GI parasite infections. They are one of the three primate species endemic to Sri Lanka, the others being purple-faced langur (*Trachypithecus vetulus*) and red slender loris (*Loris tardigradus*) (Huffman et al., 2013). Toque macaques are found as three geographically segregated subspecies across the six climatic zones in the country: the dry zone macaque or common macaque (*M. s. sinica,* Linnaeus, 1771) in the dry zone lowland, arid lowlands and intermediate zones; dusky or pale-fronted macaque (*M. s. aurifrons,* Pocock, 1931) in the rainforests of the lowland and midland of the wet zone, and the hill-zone macaque (*M. s. opisthomelas,* Hill, 1942) in the montane regions (Dittus, 1977; Nekaris and de Silva Wijeyeratne, 2009; Dittus, 2013) (Fig. 1).

Their populations declined gradually over the past 40 years, where the total population size reduced in half, largely due to habitat loss and persecution of M. sinica by humans (Nahallage et al., 2008; Nahallage and Huffman, 2013). Therefore, according to the IUCN Red List of Threatened Species, M. s. sinica and M. s. aurifrons are categorized as "Endangered" (Dittus, 2020; Dittus and Nekaris, 2020) whereas M. s. opisthomelas is listed as "Critically Endangered" (Dittus and Gamage, 2020). In addition, macaques are also listed in Appendix II of CITES (Convention on International Trade in Endangered Species) that includes species not necessarily threatened with extinction (CITES, 2005). Even though there are reports on the health, behaviour, and ecology of toque macaques (Dittus, 1984, 1986; Dittus and Ratnayeke, 1989; Keane et al., 1997; Weerasekara et al., 2021), parasite diversity of these animals is not well studied. A single study was reported on the macaque populations across the country, representing the three subspecies (Huffman et al., 2013) while others were mainly focused on macaques inhabiting specific regions, i.e Polonnaruwa (M. s. sinica), Peradeniya (M. s. aurifrons) (Dewit et al., 1991; Ekanayake et al., 2004, 2006; Thilakarathne et al., 2021), and in captivity (Gunasekera et al., 2012; Aviruppola et al., 2016). Hence, this study was undertaken to fulfill two objectives; Firstly, to determine how human influenced landscape modifications facilitated the transmission of GI parasites in macaques (in correlation to parasite prevalence, species-richness, and fecal ova/oocyst/cyst counts) in urban, suburban, and wild populations. Secondly, to establish baseline data for parasites of zoonotic potential of macaques, with a view to project significant impacts on the conservation of endangered macaques, and on the well-being of associated human populations.

2. Materials and methods

2.1. Ethics statement

This study was conducted with permission from the Department of Wildlife Conservation, Sri Lanka (WL/3/2/32/19) to enter National Parks in Sri Lanka, and to collect macaque fecal samples from these locations. The study was based on noninvasive sample collection following defecation, without causing disturbance or distress to the animals; hence, obtaining ethical clearance was not applicable.

2.2. Study sites

This study included the three endemic sub species of toque macaques, *Macaca sinica sinica*, *M. s. aurifrons*, and *M. s. opisthomelas*, inhabiting three different climatic zones, the dry zone, wet zone, and the montane region, of the island. Fourteen study sites were selected representing the dry and wet zones and montane regions in the island. The selection was based on earlier records on the presence of subspecies of toque macaques, and the ease of travel to these locations.

To assess the impact of urbanization on parasitism, each subspecies was again divided into three populations based on their level of human contact; urban/free ranging areas with the continuous industrialized expansions, where human contact was higher due to higher human density; suburban, areas adjacent to forests and with less frequent human contact, populated with a low level of human density (lesser than in urban areas); wild macaques that lived in their undisturbed natural habitats, sans involvement with humans. All these locations are geographically segregated and differed significantly in macaque habitats, the degree of anthropogenic disturbances, and climatic conditions.

Dry Zone (*M. s. sinica*): The dry zone area occupies approximately two-thirds of Sri Lanka's land area (southeast, east, and northern parts), which consists of variable forest structures and different types of habitats. Climatically, the annual rainfall varies between 1250 and 1800 mm and the temperature ranges between 29 and 32 °C. Five study sites in the dry zone were selected based on their level of human contact: Anuradhapura (8°18′46.12″N 80°24′11.38″E), Kurunegala (7°29′0.66″N 80°22′5.69″E) (urban/free ranging); Anuradhapura archeological sites (8°22′10.67″N 80°23′39.98″E), Dambulla (7°51′20.37″N 80°38′59.91″E) (suburban); and Udawalawa National park(6°28′23.91″N 80°53′55.36″E) (wild).

Wet zone (*M. s. aurifrons*): The southwestern part and the mountains of the island, well known as the "wet zone", receives an ample annual average rainfall of 2000–2500 mm with temperature that range from 24 to 27 °C. Six geographic locations with *M. s. aurifrons* macaque populations were selected encompassing urban, suburban, and wild habitats (Fig. 2). Samples of the urban/free ranging population were obtained from two locations, Awissawella (6°57′14.41"N 80°12′16.15"E) and Kandy (7°17′35.74"N 80°38′29.36"E), while Korathota (6°54′54.14"N 80° '7.69"E) and Kegalle (7° 7′36.80"N 80°24′13.01"E) harboured suburban macaque populations. Macaque populations inhabiting Udawattakele sanctuary (7°17′55.90"N 80°38′34.02"E) and Nachchimale

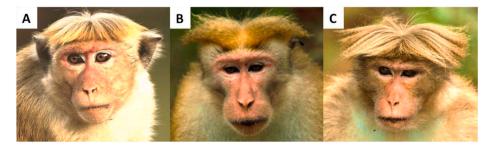


Fig. 1. The three subspecies of macaque's endemic to Sri Lanka. (A) Common macaque (*Macaca sinica sinica*), (B) dusky or pale-fronted macaque (*M. s. aurifrons*), and (C) hill-zone macaque (*M. s. opisthomelas*) (image courtesy: Madura De Silva).

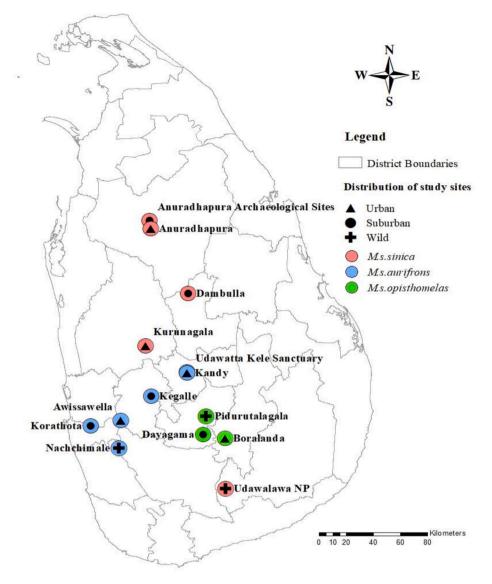


Fig. 2. Map of Sri Lanka with sampling localities in the dry and the wet zones and the montane region.

forest (6°45′31.70"N 80°11′22.52"E) were sampled as wild populations (Fig. 2).

Montane region (*M. s. opisthomelas*): The montane region located above 1000 m in the central highlands of Sri Lanka, receives 2500–5000 mm annual rainfall with the mean daily temperature being approximately 16 °C. Three sampling sites were selected according to the level of anthropogenic disturbances, including Boralanda (6°49'39.84"N 80°53'38.56"E) (Urban/free ranging), Dayagama (6°51'17.07"N 80°44'53.74"E) (suburban), and Pidurutalagala (6°59'10.25"N 80°46'13.32"E) (wild) (Fig. 2).

2.3. Sampling

A total of 180 macaque fecal samples were collected form all three subspecies of macaques, between January and November 2019: 20 samples each from urban, suburban, and wild locations amounting to 60 samples per subspecies. The activity patterns of free-ranging macaques were monitored daily from 8.00 a.m. to 4.00 p.m. Fecal samples of macaque troops were opportunistically collected by following the macaque troop till defecation took place. Upon observing that a macaque had defecated and before collection, the sample was examined for the presence of mucus, blood, and tapeworm proglottids. Then, with gloved hands and a disposable spoon, a sample from within the fecal mass was collected into a plastic tube. The Parafilm sealed tube was labeled with the date, time, location and habitat type, and samples were transported at +8 °C (in a cooler box with ice packs) to the laboratory and stored at 4 °C for a maximum of 96 h, prior to microscopic examination.

2.4. Coprological examination

Samples were analyzed at the Zoology Laboratory of the Department of Zoology and Environment Sciences of the Faculty of Science, University of Colombo. Fecal analysis was carried out essentially according to Jenkins et al. (2017), using modified Sheather's sucrose flotation method, suitable for detection and identification of various parasite ova and cysts. Briefly, 3 g of feces was mixed with distilled water and centrifuged thrice (at $2045 \times g$). The resultant pellet was emulsified and thoroughly suspended in a saturated sucrose solution. Approximately, 5 ml of the top suspensions were collected, topped up with distilled water and repeatedly centrifuged at $1370 \times g$ for 10 min. Finally, 1 ml of each suspension with the pellet was transferred to the Eppendorf® tubes. Distilled water was added to a final volume of 1.5 ml then centrifuged for 10 min at $1150 \times g$. The supernatants were discarded. The remaining 0.5 ml pellet thus obtained, was observed under standard light microscopy (Olympus, Japan) for the presence of parasite ova and cysts. Parasite stages were identified using available keys and previous references by morphological characteristics (shape, colour and content) and morphometric characteristics (length and width) (Greiner, 1989; WHO, 1994; Kouassi et al., 2015; Klaus et al., 2017; Li et al., 2017).

2.5. Parasite indices and statistical analysis

Parasite prevalence was calculated as a proportion of the number of infected individuals to the total number of examined hosts (Hodder and Chapman, 2012). Parasite richness was described as the total of parasite taxonomic groups found through the morphological identification, in each habitat type, of a given macaque subspecies (Gillespie et al., 2005). The total number of ova per gram (OPG) and cysts per gram (CPG) feces for each parasite type was considered as the parasite load (Hodder and Chapman, 2012).

Normality of distributions was tested by the Shapiro-Wilk normality test. Chi-square test determined differences in the overall parasite prevalence of each sub species, habitat type, the relative prevalence of each parasite type and prevalence of multiple infection between urbanized (urban and suburban) and wild habitats. Kruskal-Wallis test assessed differences in parasite richness and parasite ova per gram (OPG) or cysts per gram (CPG) feces, among the three habitat types within a climatic zone. Mann-Whitney U test to compare the differences in parasite richness and intensity, in urbanized habitats (urban and sub urban) and wild habitat. All statistical tests were performed at a significance level of p < 0.05, using SPSS Statistics V23.0 (IBM, USA). It was assumed that the habitat types the macaques were found during fecal sample collection to be consistent with parasitic infections at that time (Hodder and Chapman, 2012). However, due to the ranging behaviour of macaques, their distribution may have extended beyond a single habitat type.

3. Results

3.1. GI parasites of macaques

The three main parasite groups identified in this study were protozoans, helminths, and acanthocephalans. Helminth groups identified were trematodes, cestodes and nematodes, where the nematodes were the most common GI parasites among them. A total of 20 genera types of GI parasites were explicitly identified; (i) five types of protozoan cysts; *Balantidium, Endolimax, Isospora, Entamoeba*,and an unidentified protozoan; (ii) two types of unidentified trematode ova; (iii) four types of cestode ova; *Hymenolepis, Bertiella, Diphyllobothrium*, and an unidentified cestode; (iv) eight types of nematode ova; *Strongyloides, Trichuris, Trichostrongylus, Oesophagostomum, Ascaris, Enterobius, Strongyle*/hookworm types and an unidentified nematode type; and (v) a single type of acanthocephalan ova; *Moniliformis* (Fig. 3).

3.2. Overall parasite prevalence

Out of 180 samples of macaque fecal samples examined, 62.2% (112/180) were infected with at least a single GI parasite type. Among the samples, 62.2% (112/180) were infected with helminths while protozoan infections were detected in only 8.3% (15/180). There was no significant difference in the overall prevalence of GI parasites among *M. s. aurifrons* (56.7% [34/60]), *M. s. sinica* (68.3% [41/60]) and *M. s. opisthomelas* (61.7% [37/60]) ($\chi^2 = 1.749$, p = 0.471).

However, the overall GI parasite prevalence significantly differed in the three population types representing each macaque subspecies. The highest significant prevalence was found in macaques that inhabit urban habitats (76.7% [46/60]) followed by macaques in suburban habitats (68.3% [41/60]), with the lowest prevalence reported from wild habitats ($\chi^2 = 17.064$, p < 0.05). Overall, macaques in urban ($\chi^2 = 15.211$, p < 0.05) and sub urban ($\chi^2 = 8.620$, p < 0.05) habitats had the highest significant prevalence in contrast to their wild counterparts (Table 1).

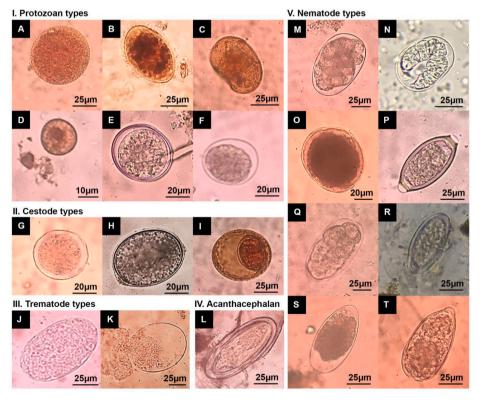


Fig. 3. GI parasite genera types identified from fecal samples of toque macaques. I. Protozoan types: (A) Balantidium cyst, (B) Balantidium trophozoite, (C) Endolimax cyst, (D) Entamoeba cyst, (E) Isospora cyst. (F) Unidentified protozoan cyst; II. Cestode types: (G) Bertiella ova, (H) Diphyllobothrium ova, (I) Hymenolepis ova; III. Trematode types: (J–K) Unidentified trematode ova; IV. Acanthocephalan type: (L) Moniliformis ova; V. Nematode types: (M) Oesophagostomum ova, (N) Strongyloides ova, (O) Ascaris ova, (P) Trichuris ova, (Q) Strongyle/ Hookworm ova, (R) Enterobius ova, (S)Trichostrongylus ova, (T) Unidentified nematode ova.

Table 1

Overall parasite prevalence and prevalence of multiple infections of GI parasites in urban, suburban, and wild macaque (Macaca sinica) populations.

Population type (n)	Overall	Prevalence of multiple infections % (n)						
	parasite prevalence	Parasite free	Single infection	Two infections	Three infections	Four infections		
Urban								
Overall urban (60)	76.7 (46) ^a	23.3 (14)	38.3 (23)	26.7 (16) ^e	5.0 (3)	6.7 (4)		
M.s. aurifrons (20)	70.0 (14)	30.0 (6)	30.0 (6)	35.0 (7)	5.0 (1)	-		
M.s. opisthomelas (20)	70.0 (14)	30.0 (6)	40.0 (8)	20.0 (4)	5.0 (1)	5.0 (1)		
M.s. sinica (20)	90.0 (18) ^d	10.0 (2)	45.0 (9) ^f	25.0 (5)	5.0 (1)	15.0 (3)		
Suburban								
Overall suburban (60)	$68.3 (41)^{b}$	31.7 (19)	35.0 (21)	21.7 (13)	8.3 (5)	3.3 (2)		
M.s. aurifrons (20)	60.0 (12)	40.0 (8)	35.0 (7)	5.0 (3)	5.0 (1)	5.0 (1)		
M.s. opisthomelas (20)	75.0 (15) ^c	25.0 (5)	35.0 (7)	20.0 (4)	5.0 (3)	5.0 (1)		
M.s. sinica (20)	70.0 (14)	30.0 (6)	35.0 (7)	30.0 (6)	5.0 (1)	_		
Wild								
Overall wild (60)	41.7 (25)	54.3 (35)	26.7 (16)	10.0 (6)	5.0 (3)	_		
M.s. aurifrons (20)	40.0 (8)	60.0 (12)	35.0 (7)	5.0 (1)	_	_		
M.s. opisthomelas (20)	40.0 (8)	60.0 (12)	30.0 (6)	5.0 (1)	5.0 (1)	-		
M.s. sinica (20)	45.0 (9)	55.0 (11)	5.0 (3)	20.0 (4)	3.3 (2)	-		

Significant difference between overall parasite prevalence in.

single parasite types in urban and wild populations of *M. s. sinica* ($\chi 2 = 4.286$, p = 0.038).

Two types of parasites in urban and wild populations of *M*. *s. aurifrons* ($\chi 2 = 5.625$, p = 0.017).

^a Urban and wild populations ($\chi 2 = 15.211$, p < 0.05).

 $^{\rm b}$ Suburban and wild populations (χ 2 = 8.620, p < 0.05).

^c Sub urban and wild populations of *M. s. opisthomelas* ($\chi 2 = 5.013$, p = 0.025).

^d Urban and wild populations of *M. s. sinica* ($\chi 2 = 9.231$, p = 0.002) Significant difference between macaques having infections with.

^e Two types of parasites in overall urban and overall wild populations ($\chi 2 = 5.566$, p = 0.018).

3.3. Mixed infections

Infected macaques harbored multiple parasitic infections, with numbers that ranged between one and four per individual. Of the overall number of macaques in the three population types, i.e., urban, suburban, and wild, most animals were infected with a single type of parasite, *i.e.*, urban (38.3% [23/60]), suburban (35% [21/60]) and wild, (26.7% [16/ 60]). Collectively, macaques having infection with two types of parasites in urban (26.7% [16/60]) were significantly higher when compared to the wild (10% [6/60]) ($\chi^2 = 5.566$, p = 0.018) but there was no significant difference between sub urban (21.7% [13/60]) and wild (γ^2 = 3.064, p = 0.08) populations. Conversely, no significant difference was recorded among the percentage of toque macaques harbouring three parasite types in urban (5% [3/60]), sub urban (8.3% [5/60]) and wild (5% [3/60]) habitats. The maximum score for multiple infections per single macaque was four types of parasites per individual recorded in urban (6.7% [4/60]) and suburban (3.3% [2/60]) populations. Conversely, the highest number of parasite types per infected macaque in the wild population, numbered three (Table 1).

With respect to single parasitic infections in each macaque sub species, only *M. s. sinica* in the urban habitat showed a significant difference compared to their wild counterparts ($\chi 2 = 4.286$, p = 0.038). Conversely, only urban population of *M. s. aurifrons* indicated a significant difference compared to their wild population, harbouring two types of parasitic infections in each macaque ($\chi 2 = 5.625$, p = 0.017) (Table 1).

3.4. Parasite prevalence and species richness of the three-macaque subspecies inhabiting diverse habitat types

Overall prevalence of infection did not differ among *M. s*, *s. aurifrons* in urban (70%), suburban (60%) and wild (40%) populations (χ^2 = 3.801, p = 0.15), or for any of the GI parasite genera types documented (Table 1). Of *M. s. aurifrons* populations in the three habitat types s, *Strongyloides* type was the most frequent nematode infection, followed by *Strongyle*/hookworm type and *Trichuris* type while *Balantidium* type was the commonest amongst protozoans. *M. s. aurifrons* from the urban population were exclusively infected with *Isospora* type and *Trichostrongylus* type. Infection by *Diphyllobothrium* type and *Ascaris* type were confined to suburban animals, while Enterobius

type was recorded in macaques inhabiting their natural habitat (Table 2). A significant difference existed between the total number of parasite genera types infecting *M. s. aurifrons* urban (n = 7), suburban(n = 6) and wild (n = 3) (p = 0.045) habitats. Urban living *M. s. aurifrons* population harbored more parasite species than those living in the wild (Z = -2.484, p = 0.013) (Fig. 4).

There was a significant difference in the overall prevalence of parasites among the urban (70%), suburban (75%) and wild (40%) populations of *M. s. opisthomelas* ($\chi^2 = 6.063$, p = 0.048). However, when comparisons were drawn among populations, macaques in the suburban population (75%) had the highest significant prevalence of parasites than in the wild (40%) ($\chi^2 = 5.013$, p = 0.025), while there was no significant difference between wild and urban living macaques ($\chi^2 =$ 3.636, p = 0.057) (Table 1).

M. s. opisthomelas was exceptional among the three-macaque subspecies, as prevalence of specific parasite types among the urban, suburban, and wild populations did not differ. Infection by *Isospora* type and *Entamoeba* type were confined to macaques in the suburban habitat (Table 2). There was a significant difference between the total number of parasite genera types infecting *M. s. opisthomelas* in urban (n = 7), suburban (n = 9) and wild (n = 3) (p = 0.028) habitats. Comparison of parasite richness in the different habitat types, was high in both suburban (n = 9) (Z = -2.022, p = 0.043) and urban populations (n = 7) (Z = -2.514, p = 0.012) compared to the wild (n = 3) (Fig. 4).

The overall prevalence of infection significantly differed among *M*. *s. sinica* urban (90%), suburban (70%), and wild (45%) populations ($\chi^2 =$ 9.397, p = 0.009). Parasite prevalence in the urban macaque population was significantly higher than of the wild ($\chi^2 =$ 9.231, p = 0.002) (Table 1). There was no significant difference in the relative prevalence of individual parasite types except for *Strongyloides* type in urban and wild populations ($\chi^2 =$ 10.417, p = 0.001). Comparable to the other two subspecies, *Strongyloides* type was the frequently recorded nematode infection followed by *Trichuris* type, in *M*. *s. sinica* in all three habitat types. It is noteworthy that *Hymenolepis* type, *Bertiella* type, and *Moniliformis* type were confined to *M*. *s. sinica* among the three subspecies of endemic macaques (Table 2). Overall parasite species high in the urban macaques (n = 8) compared to their suburban and wild counterparts (n = 6) (Z = -1.017, p = 0.02) (Fig. 4).

Table 2

Prevalence (%) of GI parasite infections in Macaca sinica aurifrons, M. s. sinica and M. s. opisthomelas in urban, suburban, and wild habitats.

Host	Prevalence of GI parasite type% (n) (N = 180)									
Habitat type Parasite genera types	<i>M.s. aurifrons</i> (n = 60) (n = 20 per habitat)			<i>M.s. sinica</i> $(n = 60)$ $(n = 20$ per habitat)			<i>M.s. opisthomelas</i> $(n = 60)$ $(n = 20$ per habitat)			
	U	SU	W	U	SU	W	U	SU	W	
Protozoan types										
Balantidium	20(4)	15(3)	-	-	-	-	5(1)	10(2)	5(1)	
Isospora	5(1)	-	-	-	_	-	_	5(1)	-	
Entamoeba	-	-	-	-	-	-	_	5(1)	-	
Endolimax	-	-	-	5(1)	-	5(1)	_	-	-	
Unidentified protozoa	-	-	-	-	-	-	-	5(1)	-	
Total protozoan infection	25(5)	15(3)	-	5(1)	-	5(1)	5(1)	20(4)	5(1)	
Trematode types										
Unidentified trematodes	-	-	-	-	5(1)	-	5(1)	-	-	
Cestode types										
Diphyllobothrium	-	5(1)	-	-	-	-	_	-	-	
Bertiella	-	-	-	-	5(1)	-	_	-	-	
Hymenolepis	-	-	-	-	-	25(5)	_	-	-	
Unidentified cestode	-	5(1)	-	5(1)	-	-	_	-	-	
Nematode types										
Strongyloides	45(9)	60(12)	35(7)	85(17)	70(14)	35(7)	60(12)	45(9)	30(6)	
Trichuris	15(3)	-	10(2)	25(5)	10(2)	15(3)	5(1)	20(4)	-	
Ascaris	-	5(1)	-	15(3)	15(3)	-	10(2)	10(2)	-	
Strongyle/Hook worm	20(4)	10(2)	-	15(3)	5(1)	-	25(5)	35(7)	20(4)	
Trichostrongylus	20(4)	-	-	10(2)	_	-	_	-	-	
Oesophagostomum	_	-	-	5(1)	-	-	5(1)	5(1)	-	
Enterobius	_	-	5(1)	-	-	5(1)	_	-	-	
Unidentified nematode	5(1)	_	_	-	-	-	_	_	_	
Total helminths infection	70(14)	60(12)	40(8)	90(18)	70(14)	45(9)	70(14)	75(15)	40(8)	
Acanthocephalan type										
Moniliformis	_	_	_	5(1)	_	_	_	_	_	

U, urban; S.U, suburban; W, wild.

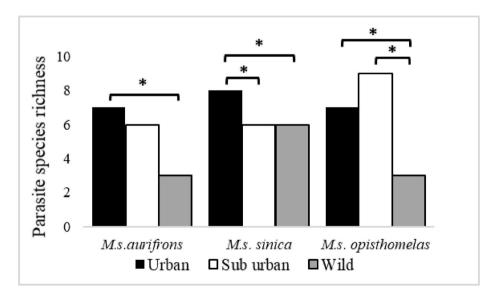


Fig. 4. Number of parasite genera types (species richness) infecting M. s. aurifrons, M. s. sinica and M. s. opisthomelas in urban, suburban, and wild habitats.

3.5. Parasite ova/cyst per gram (OPG/CPG) feces

The index OPG/CPG indicates the parasite load in a host. In *M. s.* aurifrons the highest OPG was of *Strongyloides* type (OPG = urban; 3.33 \pm 1.42, suburban; 1.72 \pm 1.14, wild; 1.00 \pm 0.51), though the maximum CPG was recorded for *Balantidium* type in both urban and suburban populations. All other analyzed OPG and CPG were relatively low. No significant variation among individual OPG was recorded except for the EPG of *Strongyloides* type (Table 3). The urban population of *M. s.* aurifrons manifested a significantly higher OPG of *Strongyloides* type than those from the wild (Z = -2.036, p = 0.042) population.

The highest OPG for M. s. opisthomelas was recorded for Strongyloides

type from urban and suburban habitats (OPG = urban; 4.83 \pm 7.67, suburban; 1.41 \pm 0.96). The highest protozoan CPG was of *Balantidium* type recorded in the suburban population (0.83 \pm 0.71). OPG of *Strongyloides* type differed among urban and wild populations (Z = -2.167, p = 0.03). No other significant comparisons were recorded among the individual parasite genus types in *M. s. opisthomelas* inhabiting the three examined habitats (Table 3).

Compared to the wild population, OPG of individual parasite types differed for *M. s. sinica* among both urban (Z = -3.625, p < 0.05) and sub urban (Z = -2.636, p = 0.008) populations. Nevertheless, the highest OPG of *Strongyloides* type was recorded from suburban *M. s. sinica* samples (5.14 \pm 5.81) than those from the other two macaque

Table 3

Mean parasite ova/cyst per gram (OPG/CPG) feces of different parasite types infecting Macaca sinica aurifrons, M.s. sinica and M.s. opisthomelas in urban, suburban and wild habitats.

Host	Mean parasite OPG/CPG \pm (SD)								
Habitat type Parasite genera types	M.s. aurifrons			M.s. sinica			M.s. opisthomelas		
	U	SU	W	U	SU	W	U	SU	W
Protozoan types									
Balantidium	$0.67\pm$	$0.89\pm$	-	-	-	-	0.67 ^a	$0.83\pm$	0.67 ^a
	(0.27)	(0.38)	-	-	-	-	-	(0.71)	-
Isospora	1.0^{a}	-	-	-	-	-	-	0.33 ^a	-
Entamoeba	-	-	-	1.00^{a}	-	1.00^{a} -	-	0.67 ^a	-
Endolimax	-	-	-	-	-	-	-	-	_
Unidentified protozoa	-	-	-	-	-	-	-	0.33 ^a	-
Trematode types									
Unidentified Trematodes	_	-	_	-	0.33 ^a	-	0.33 ^a	-	_
Cestode types									
Diphyllobothrium	_	1.33 ^a	_	-	-	-	-	-	_
Bertiella	-	_	_	_	0.33 ^a	_	-	-	-
Hymenolepis	-	-	-		-	$2.53 \pm (1.42)$	-	-	-
Unidentified cestode		2.00 ^a		1.00 ^a					
Nematode types									
Strongyloides	$3.33\pm$	$1.72\pm$	$1.00\pm$	$4.35\pm$	$5.14\pm$	$1.14\pm$	$4.83\pm$	$1.41\pm$	$0.67\pm$
	(1.42)	(1.14)	(0.51)	(5.33)	(5.81)	(0.77)	(7.67)	(0.96)	(0.37)
Trichuris	$1.00\pm$	_	$0.33\pm$	$1.00\pm$	$1.17\pm$	$1.33\pm$	1.00 ^a	$1.25\pm$	-
	(0.33)		(0)	(0.71)	(1.18)	(0.88)		(0.69)	-
Ascaris		1.67 ^a	_	0.33± (0)	$1.33\pm$	-	0.67±(0.47)	0.50±(0.24)	$0.83\pm$
	-		_		(1.21)	_			(0.43)
Strongyle/Hook worm	$1.50\pm$	$2.00\pm(0.47)$	_	4.11± (2.41)	4.33 ^a	_	$2.07 \pm (1.38)$	$1.14 \pm (0.43)$	-
	(0.88)	_	0.67 ^a	$0.33\pm$	_	1.00 ^a	-		-
Oesophagostomum	$1.50\pm$	_	_	(0)	_	_	1.67 ^a		-
	(0.69)	-	_	1.33 ^a	_	_	-	-	_
Enterobius	_	_		_	_		_	1.33 ^a	_
Unidentified nematode	-	-		_	-		_	-	
Acanthocephalan type									
Moniliformis	0.33 ^a			1.33 ^a	-			-	
-	_							_	

U-urban; S.U.- suburban; W- wild.

^a Infections were confined to a single individual macaque.

subspecies. The maximum CPG was of *Endolimax* type (1.00) which was recorded in a single individual macaque each from urban and wild populations.

4. Discussion

It is well established that demographic and anthropogenic changes exert influence on the emergence of zoonotic diseases (Jones et al., 2008; Lane et al., 2011). Hitherto, GI parasite infections in toque macaques, especially with the influence of urban landscape on the wildlife pathogen relationship received less emphasis. This is the first study of its kind, based on the impact of urbanization on GI parasites in the three endemic subspecies of toque macaques in Sri Lanka. As hypothesized, M. s. opisthomelas, M. s. aurifrons and M. s. sinica in urbanized habitats harboured a higher prevalence and parasite richness of GI parasites in contrast to their wild counterparts. It was observed that there is a marked difference in daily activity, ranging patterns, food, and water sources of macaque troops in urban settings, compared to those in the wild. These observations reiterate several studies on different macaque species and other primates, globally; these reports are indicative of alterations in population structure of parasite infections in primates with the level of anthropogenic interactions and environment modifications (Sleeman et al., 2000; Gillespie et al., 2005; Ekanayake et al., 2006; Wenz et al., 2010; Lane et al., 2011; Hodder and Chapman, 2012; Hussain et al., 2013; Wenz-mücke et al., 2013). Furthermore, climatic factors and altitude also can be considered as factors that influence the survival and infection of parasites in free-ranging macaques. Among the three sub species inhabiting different climatic zones of the island, M. s. sinica found in the dry zone harbour a higher prevalence of GI parasites compared to the other two subspecies. This may be due to increase infection success and faster development in parasites in the high

temperature than in relatively low temperature ranges (Labaude et al., 2020). Maximum temperature is an important variable in determining soil-transmitted helminth distribution owing to effect of heat and low humidity of the soil on the embryonation, development and survival of free-living infective stages (Brooker et al., 2003).

Overall, 20 parasite genera types were identified in the GI parasite survey that contributed to the parasite diversity of toque macaques; the Strongyloides type was responsible for the highest prevalence and parasite load (OPG) in all three subspecies of hosts followed by Strongyle/ hookworm and Trichuris (whipworm) types, respectively. Infection by Strongyloides occurs by accidental oral mucosa or skin penetration of third-stage filariform larvae in the soil (Grove, 1996). Even though macaques are primarily arboreal (Nekaris and de Silva Wijeyeratne, 2009), during the observation period from 8.00 to 16.00 h, both urban and suburban troops dwelled on the ground for about 80% of the time, while this was limited to an average of 15% by the toque groups in the wild (authors' personal observations). Therefore, predominant ground living behaviour and close association of individuals within the macaque group may be the major reason for the high prevalence of Strongyloides infection (Altmann and Muruthi, 1988; Ekanayake et al., 2006; Griffin and Nunn, 2012). Strongyloidiasis in humans is described as a potential zoonotic disease (Nutman, 2016). Pathological effects of this infection in primates vary from vomiting, diarrhea, and severe dehydration (Strait et al., 2012) while fibrosis of the intestine results in humans (Mukerjee et al., 2003). In severe infection, macaques may cope with similar clinical problems as humans, and several fatal cases have been reported in the gibbon, orangutan, and woolly monkeys (Pillers and Southwell, 1929; DePaoli and Johnsen, 1978; Uemura et al., 1979) elsewhere in the world.

Strongyle/hookworm type infection, the second-highest GI parasite prevalence was recorded in all three-macaque subspecies inhabiting

urbanized habitats. However, in the wild, this infection was restricted to *M. s. opisthomelas*. Individual primates with more grooming partners can have a significantly higher infection of hookworms (Wren et al., 2016) and grooming related with the physiological stress levels in macaques (Shutt et al., 2007). Therefore, when individuals in the population are more stressed and defense mechanisms fail (Mul et al., 2007) it could lead to greater susceptibility to infections in such individuals than in others. It eventually poses a threat to macaque populations as this infection causes tissue damage, retarded growth, anemia, and inflammation (Hussain et al., 2013; Seguel and Gottdenker, 2017). More importantly, macaques in urbanized habitats create human-macaque interfaces that disturb the dynamics of hookworm infection and infected macaques can be a threat to closely interacting humans (Seguel and Gottdenker, 2017).

Trichuris (whipworm) infection was reported in all three-macaque subspecies in habitats except for M. *s. opisthomelas* wild populations (in montane regions), and suburban populations of *M. s. aurifrons* in the wet zone. This reiterates alteration of parasite patterns due to geographic and climatic conditions (Sharma et al., 2013). In previous studies, *Trichuris* infection in Asian primates was found to be less common, and macaques may acquire it because of the close association with grey langurs (Dewit et al., 1991). This may explain the relatively high prevalence of *Trichuris* type in *M. s. sinica* in this study, as their troops were mostly observed foraging together with grey langurs. *Trichuris* infection occurs orally, and ova can survive for a longer period outside the host, in severe environmental conditions. Severe whipworm infections can produce clinical signs such as anorexia, diarrhea resulting sometimes in death (Le et al., 2020).

Most of the suburban and a single urban population of macaques were recorded in places of religious worship, where local pilgrims and tourists visited throughout the year. Consequently, the presence of infected macaques with their randomly dispersed feces found in those areas may pose a threat to public health, as in those religious and archeological sites, people usually walk barefooted. This may increase the possibility of humans to acquire soil-transmitted helminth infections.

Nematode infections pose high potential for zoonotic transmission with their direct and simple life cycles while the infections by cestodes and acanthocephalan are normally rare and only causes diarrhea and abdominal pain in primates (Torgerson and Macpherson, 2011; Strait et al., 2012). Even though *Bertiella and Moniliformis* infections were restricted to individual M. *s. sinica* macaques, it may indicate the possible transmission of these parasites from other animals such as rodents and mites to macaques (Elliott, 2007; Kowalewski et al., 2017).

Confirming previous studies, the prevalence of protozoans was lower compared to the helminth infections in macaques in the current study (Kouassi et al., 2015; Adhikari and Dhakal, 2018; Adrus et al., 2018; Kumar et al., 2018; Zhang et al., 2019). The contrary was reported by Li et al. (2017) with 40.1% of macaques screening positive for protozoan and 29.6% for helminth infections. Kurniawati et al. (2020) reiterated protozoan was the most prevalent infection (89%) compared to helminth infections (66%) in long-tailed macaques. All five types of protozoans detected in the current study, were of relatively high prevalence; Balantidium type > Isospora type in both M. s. aurifrons and M. s. opisthomelas. All recorded protozoan's manifest zoonotic transmission and are human pathogens except for Isospora type which is a common parasite of cats and dogs (Youn, 2009; Ferreira et al., 2011). In general, protozoan infections are highly prevalent due to the capability of cysts to survive even in harsh environments, and their one host-life cycle. These recorded protozoans can transmit directly through the fecal-oral route via contaminated food or water sources (Osman et al., 2016). Balantidium sp. is known as the only ciliated protozoan that commonly infects humans and several animals. It can be pathogenic to both macaques and humans by causing clinical symptoms of Balantidiasis which include diarrhea and dysentery (Hussain et al., 2013). During the current study macaque troops were observed using the same

water source for drinking as well as for bathing which can be the major reason that facilitates transmission of protozoan infections. Ekanayake et al. (2006) observed that wild primates using land areas heavily soiled by human and livestock feces, harbored more parasitic protozoans common in humans, than their wild counterparts. More importantly, *Balantidium* infection in lactating female macaques can reduce the fat of milk and inhibit the development of lactating mothers by reducing activity levels and impairing their foraging behaviour (Hinde, 2007). Importantly, it can compromise the growth of infants which can cause several health implications to macaque populations.

Several studies conducted in both children and adults in Sri Lanka from 1924 to 2019, found that school children across the country were positive for hookworm, whipworm, roundworm and Bertiella infection, especially those living in urban areas (Amarasinghe et al., 2020; De Silva et al., 1994; Sorensen et al., 1996; Gunawardena et al., 2011; Galgamuwa et al., 2016; Ediriweera et al., 2019)., The reason attributed to Bertiella infections was that the main reservoir hosts, the toque macaques, were regular visitors to human settlements (Amarasinghe et al., 2020), which may pose a threat, especially to small children as well as to domestic animals. Furthermore, alternations to the availability of foraging and roosting sites and the quality and quantity of food macaques consumed due to anthropogenic activities and urbanization, may lead in several ways to increase the parasite burden in macaque populations (Weyher et al., 2006). Importantly, crowding in a fragmented area with small space, frequent locomotion on the ground, grooming behaviour and repeated use of the same area may not only increase the possibility of parasite transmission but also, increase chances to re-infect within the troop members. Although there are no fatality reports on diseases caused by GI parasites in macaques and humans in Sri Lanka, it is prudent to take precautionary measures. Therefore, it is important to encourage creating more natural habitats suitable for macaques within urban areas, the introduction of proper waste disposal methods, avoid feeding macaques, continuous deworming practices and health education, to reduce health impacts on both humans and co-existing macaque populations.

Several novel host-parasite associations were established through the current study; this is the first report of *Moniliformis* type infection in macaques globally and the first report of *Isospora* type, *Trichostrongylus* type, *Enterobius* type and *Diphyllobothrium* type infection in *M.s.aurifrons* (Huffman et al., 2013; Thilakarathne et al., 2021), *Endolimax* type, *Moniliformis* type and *Bertiella* type infection in *M.s.sinica*(Dewit et al., 1991; Ekanayake et al., 2006; Huffman et al., 2013; Thilakarathne et al., 2021) and *Isospora* type, *Balantidium* type, *Strongyloides* type, *Ascaris* type and *Oesophagostomum* type infections in *M. s. opisthomelas* (Huffman et al., 2013).

In summation, results from this study lead to two conclusions; Firstly, macaques that live in urban and suburban areas, closer to human settlements harboured increased prevalence and parasite richness compared to wild macaques in their natural habitat. Secondly, infections by soil-transmitted parasites in macaques were relatively high. As two of the subspecies of macaques (M. s. aurifrons and M. s. sinica) are endangered and the other (M. s. opisthomelas) is critically endangered, further studies are required to gauge whether the higher prevalence of parasites encountered may pose a threat to their population abundance and daily activity patterns. Further, identification of recorded parasites up to the species level is also important in assessing major health risks of these parasitic infections. Almost all the parasites identified in the current study are of zoonotic potential. Given the genetic relatedness of humans and macaques, disease transmission between them may be potent. Therefore, to verify the existent risk, it is necessary to prove whether humans who share the same environment are indeed infected with any diseases caused by zoonotic GI parasites found in co-existing macaque populations. Such future efforts will help prevent transmission of zoonotic parasites, which is an important implication for primate conservation efforts while ensuring public health and safety.

Declaration of interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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References

- Adhikari, P.P., Dhakal, P., 2018. Prevalence of gastro-intestinal parasites of rhesus macaque (*macaca mulatta zimmermann*, 1780) and hanuman langur (*semnopithecus entellus dufresne*, 1797)indevghat, Chitwan, Nepal. JIST 22 (2), 12–18. https://doi. org/10.3126/jist.v22i2.19590.
- Adrus, M., Zainudin, R., Ahamad, M., Jayasilan, M., Abdullah, M.T., 2018. Gastrointestinal parasites of zoonotic importance observed in the wild, urban, and captive populations of non-human primates in Malaysia. J. Med. Primatol. 48 (1), 22–31. https://doi.org/10.1111/jmp.12389.
- Altmann, J., Muruthi, P., 1988. Differences in daily life between semiprovisioned and wild-feeding baboons. Am. J. Primatol. 15 (3), 213–221. http://doi:10.1002/ajp.1 350150304.
- Amarasinghe, L.D., Premathilake, E.M.D.L., 2014. Parasites of domestic animals and their possible zoonoses-a study from selected sites of western province, Sri Lanka. J. Exp. Biol. Agric. Sci. 2 (2), 182–187.
- Amarasinghe, A., Le, T.H., Wickramasinghe, S., 2020. Bertiella studeri infection in children SriLanka. Emerg. Infect. Dis. 26 (8), 1889. https://dx.doi.org/10.3201/eid 2608.200324.
- Auld, S.K.J.R., Tinkler, S.K., Tinsley, M.C., 2016. Sex as a strategy against rapidly evolving parasites. Proc. Royal Soc. B-Bio. Sci. 283 (1845), 20162226. https://doi. org/10.1098/rspb.2016.2226.
- Aviruppola, A.J.M.K., Rajapakse, R.P.V.J., Rupika, S., 2016. Coprological survey of gastrointestinal parasites of mammals in Dehiwala National Zoological Gardens, Sri Lanka. Ceylon J. Sci. 45, 83–96. https://doi.org/10.4038/cjs.v45i1.7367.
- Brooker, S., Singhasivanon, P., Waikagul, J., Supavej, S., Kojima, S., Takeuchi, T., Luong, T.V., Looareesuwan, S., 2003. Mapping soil-transmitted helminths in Southeast Asia and implications for parasite control. Southeast Asian J. Trop. Med. Publ. Health 34 (1), 24–36. PMID: 12971511.
- Cites, 2005. Convention on International trade in endangered species of wild Flora and Fauna. https://checklist.cites.org/#/en.
- DePaoli, A., Johnsen, D.O., 1978. Fatal strongyloidiasis in gibbons (Hylobates lar). Veterinary Pathology 15 (1), 31–39. https://doi.org/10.1177/030098587 801500104.
- De Silva, N.R., De Silva, H.J., Jayapani, V.P.P., 1994. Intestinal parasitoses in the Kandy area, Sri Lanka. Southeast Asian J. Trop. Med. Pub. Health 25, 469-469.
- Dewit, I., Dittus, W.P.J., Vercruysse, J., Harris, E.A., Gibson, D.I., 1991. Gastro-intestinal helminths in a natural population of *Macaca sinica* and *Presbytis* spp. at Polonnaruwa, Sri Lanka. Primates 32, 391–395. https://doi.org/10.1007/ BF02382681.
- Dittus, W.P.J., 1977. The socioecological basis for the conservation of the toque monkey (Macaca sinica) of Sri Lanka (Ceylon). Primate Con 237–265. https://doi.org/ 10.1016/b978-0-12-576150-5.50012-1.
- Dittus, W.P., 1984. Toque macaque food calls: semantic communication concerning food distribution in the environment. Anim. Behav. 32 (2), 470–477. https://doi.org/ 10.1016/S0003-3472(84)80283-3.
- Dittus, W.P.J., 1986. Sex differences in fitness following a group take-over among toque macaques: testing models of social evolution. Behav. Ecol. Sociobiol. 19, 257–266. https://doi.org/10.1007/BF00300640.
- Dittus, W.P., Ratnayeke, S.M., 1989. Individual and social behavioral responses to injury in wild toque macaques (*Macaca sinica*). Int. J. Primatol. 10 (3), 215–234. https://doi.org/10.1007/BF02735201.
- Dittus, W.P.J., 2013. Subspecies of Sri Lankan mammals as units of biodiversity conservation , with special reference to the primates. Ceylon J. Sci. 42 (2), 1–27. https://doi.org/10.4038/cjsbs.v42i2.6606.
- Dittus, W., 2020. Macaca sinica ssp. sinica. The IUCN red list of threatened species 2020. e.T39798A17985798. https://dx.doi.org/10.2305/IUCN.UK.2020-2.RLTS.T39798 A17985798.
- Dittus, W., Gamage, S.N., 2020. Macaca sinica ssp. opisthomelas. The IUCN Red List of Threatened Species 2020: e.T39800A17985750. https://dx.doi.org/10.2305/IUCN. UK.2020-2.RLTS.T39800A17985750.
- Dittus, W., Nekaris, K.A.I., 2020. Macaca sinica ssp. aurifrons. The IUCN red list of threatened Species2020. e.T39799A17985774. https://dx.doi.org/10.2305/IUCN. UK.2020-2.RLTS.T39799A17985774.en.
- Ediriweera, D.S., Gunawardena, S., Gunawardena, N.K., Iddawela, D., Kannathasan, S., 2019. Articles Reassessment of the prevalence of soil-transmitted helminth infections

in Sri Lanka to enable a more focused control programme : a cross-sectional national school survey with spatial modelling. Lancet Global Health 1–10. https://doi.org/10.1016/S2214-109X(19)30253-0.

- Ekanayake, D.K., Rajapakse, R.P.V.J., Dubey, J.P., Dittus, W.P.J., 2004. Seroprevalence of *Toxoplasma gondii* in wild toque macaques (*Macaca sinica*) at Polonnaruwa, Sri Lanka. J. Parasitol. 90, 870–871. https://doi.org/10.1645/ge-291r.
- Ekanayake, D.K., Arulkanthan, A., Horadagoda, N.U., Sanjeevani, G.K.M., Kieft, R., Gunatilake, S., Dittus, W.P.J., 2006. Prevalence of *Cryptosporidium* and other enteric parasites among wild non-human primates in Polonnaruwa, Sri Lanka. Am. J. Trop. Med. Hyg. 74, 322–329. https://doi.org/10.4269/ajtmh.2006.74.322.
- Elliott, S.P., 2007. Rat bite fever and Streptobacillus moniliformis. Clin. Microbiol. Rev. 20 (1), 13–22. https://dx.doi.org/10.1128/CMR.00016-06.
- Ferreira, F.S., Pereira-baltasar, P., Parreira, R., Padre, L., Vilhena, M., Tavira, L.T., Atouguia, J., Centeno-lima, S., 2011. Veterinary Parasitology Intestinal parasites in dogs and cats from the district of Évora, Portugal. Vet. Parasitol. 179, 242–245. https://doi.org/10.1016/j.vetpar.2011.02.003.
- Galgamuwa, L., Iddawela, D., Dharmaratne, S.D., 2016. Factors associated with the prevalence of Ascaris lumbricoides infection among preschool children in a plantation community, Kandy District, Sri Lanka. Southeast Asian J. Trop. Med. Publ. Health 47 (6), 1143–1152.
- Gillespie, T.R., Chapman, C.A., Greiner, E.C., 2005. Effects of logging on gastrointestinal parasite infections and infection risk in African primates. J. Appl. Ecol. 42, 699–707. https://doi.org/10.1111/j.1365-2664.2005.01049.x.
- Greiner, E.C., 1989. Parasite diagnosis by fecal examination. J. Assoc. Avian Vet. 3 (2), 69–72. https://doi.org/10.2307/30134907.
- Griffin, R.H., Nunn, C.L., 2012. Community structure and the spread of infectious disease in primate social networks. Evol. Ecol. 26, 779–800. https://doi.org/10.1007/ s10682-011-9526-2.
- Grove, D.I., 1996. Human strongyloidiasis. Adv. Parasitol. 38, 251–309. https://doi.org/ 10.1016/S0065-308X(08)60036-6.
- Gunasekera, U.C., Wickramasinghe, S., Wijesinghe, G., Rajapakse, R.P.V.J., 2012. Gastrointestinal parasites of captive primates in the national zoological gardens of Sri Lanka. Taprobanica J. Asian Biodivers. 4, 37–41. https://doi.org/10.4038/tapro. v4i1.4380.
- Gunawardena, K., Kumarendran, B., Ebenezer, R., Sanjeewa, M., Pathmeswaran, A., De Silva, N., 2011. Soil-transmitted helminth infections among plantation sector schoolchildren in Sri Lanka : prevalence after ten years of preventive chemotherapy. PLoS Nevlected Tron. Dis. 5: e1341 https://doi.org/10.1371/journal.pntd.0001341.
- Herbison, R., Largue, C., Poulin, R., 2018. The missing link in parasite manipulation of host behaviour. Parasites and vect 11 (1), 1–6. https://doi.org/10.1186/s13071-018-2805-9.
- Hinde, K., 2007. Milk composition varies in relation to the presence and abundance of Balantidium coli in the mother in captive rhesus macaques (Macaca mulatta). Am. J. Primatol. 69 (6), 625–634. https://doi.org/10.1002/ajp.
- Hodder, S.A.M., Chapman, C.A., 2012. Do nematode infections of red Colobus (Procolobus rufomitratus) and black-and-white Colobus (*Colobus guereza*) on Humanized forest edges differ from those on nonhumanized forest edges. htt ps://doi.org/10.1007/s10764-012-9619-y, 845-859.
- Huffman, M.A., Nahallage, C.A.D., Hasegawa, H., Ekanayake, S., Silva, L.D.G.G. De, 2013. Preliminary survey of the distribution of four potentially zoonotic parasite species among primates in Sri Lanka. J Natl Sci Found 41, 319–326. https://doi.org/ 10.4038/jnsfsr.v41i4.6246.
- Hussain, S., Ram, M.S., Kumar, A., Shivaji, S., Umapathy, G., 2013. Human presence increases parasitic load in endangered lion-tailed macaques (*Macaca silenus*) in its fragmented rainforest habitats in southern India. PLoS One 8 (5), e63685. https:// doi.org/10.1371/journal.pone.0063685.

Jenkins, T.P., Rathnayaka, Y., Perera, P.K., Peachey, L.E., Nolan, M.J., Krause, L., Rajakaruna, R.S., Cantacessi, C., 2017. Infections by human gastrointestinal helminths are associated with changes in faecal microbiota diversity and composition. PLoS One 12, 1–18. https://doi.org/10.1371/journal.pone.0184719.

- Johnson, P.T.J., Hoverman, J.T., 2012. Parasite diversity and coinfection determine pathogen infection success and host fitness. Proc. Natl. Acad. Sci. Unit. States Am. 109 (23), 9006–9011. https://doi.org/10.1073/pnas.1201790109.
- Jones-Engel, L., Engel, G.A., Schillaci, M.A., Kyes, K., Froehlich, J., Paputungan, U., Kyes, R.C., 2004. Prevalence of enteric parasites in Pet macaques in Sulawesi, Indonesia. Am. J. Primatol. 62 (2), 71–82. https://doi.org/10.1002/ajp.20008.
- Jones, K.E., Patel, N.G., Levy, M.A., Storeygard, A., Balk, D., Gittleman, J.L., Daszak, P., 2008. Global Trends in Emerging Infectious Diseases, 451, pp. 990–994. https://doi. org/10.1038/nature06536.
- Keane, B., Dittus, W.P.J., Melnick, D.J., 1997. Paternity assessment in wild groups of toque macaques *Macaca sinica* at Polonnaruwa, Sri Lanka using molecular markers. Mol. Ecol. 6 (3), 267–282. https://doi.org/10.1046/j.1365-294X.1997.00178.x.
- Klaus, A., Zimmermann, E., Roper, M.K., Radespiel, U., Nathan, S., Goossens, B., Strube, C., 2017. Co-infection patterns of intestinal parasites in arboreal primates (proboscis monkeys, *Nasalis larvatus*) in Borneo. Int. J. Parasitol. Parasit. Wildl. 6, 320–329. https://doi.org/10.1016/j.ijppaw.2017.09.005.
- Kouassi, R.Y.W., McGraw, S.W., Yao, P.K., Abou-Bacar, A., Brunet, J., Pesson, B., Bonfoh, B., N'Goran, E.K., Candolfi, E., 2015. Diversity and prevalence of gastrointestinal parasites in seven non-human primates of the Taï National Park, C^ote d'Ivoire. Parasite 22, 1–12. https://doi.org/10.1051/parasite/2015001.
- Kowalewski, M., Torres, J., Milozzi, C., Gillespie, T.R., 2017. *Bertiella* sp. infection patterns in black and gold howler monkeys across their distribution. Primatol. Argentina 219–233.
- Kumar, S., Sundararaj, P., Kumara, H.N., Pal, A., Santhosh, K., Vinoth, S., 2018. Prevalence of gastrointestinal parasites in bonnet macaque and possible

S.U. Fernando et al.

consequences of their unmanaged relocations. PLoS One 13 (11), e0207495. https://doi.org/10.1371/journal.pone.0207495.

- Kurniawati, D.A., Suwanti, L.T., Lastuti, N.D.R., Koesdarto, S., Suprihati, E., Mufasirin, M., Pratiwi, A., 2020. Zoonotic potential of gastrointestinal parasite in long-tailed macaque *Macaca fascicularis* at Baluran national park, Situbondo, east Java, Indonesia. Aceh J. Anim. Sci. 5 (1), 47–56.
- Labaude, S., Cézilly, F., De Marco, L., 2020. Increased temperature has no consequence for behavioral manipulation despite effects on both partners in the interaction between a crustacean host and a manipulative parasite. Sci. Rep. 10, 11670. https:// doi.org/10.1038/s41598-020-68577-z.
- Lane, K.E., Holley, C., Hollocher, H., 2011. The anthropogenic environment lessens the intensity and prevalence of gastrointestinal parasites in Balinese long-tailed macaques (*Macaca fascicularis*). Primates 52 (2), 117–128. https://doi.org/ 10.1007/s10329-010-0230-6.
- Le, L., Khatoon, S., Jiménez, P., 2020. Chronic whipworm infection exacerbates Schistosoma mansoni egg-induced hepatopathology in non-human primates. Parasites Vectors 13, 109. https://doi.org/10.1186/s13071-020-3980-z.
- Li, J., Dong, H., Wang, R., Yu, F., Wu, Y., 2017. An investigation of parasitic infections and review of molecular characterization of the intestinal protozoa in nonhuman primates in China from 2009 to 2015. Int. J. Parasitol. Parasites Wildl. 6, 8–15. https://doi.org/10.1016/j.ijppaw.2016.12.003.
- Milozzi, C., Bruno, G., Cundom, E., Mudry, M.D., Navone, G.T., 2012. Intestinal parasites of *Alouatta caraya* (Primates, Ceboidea): preliminary study in semi-captivity and in the wild in Argentina. Mastozool. Neotrop. 19 (2), 271–278. https://www.redalyc. org/articulo.oa/id=45725085018.
- Moll, R.J., Cepek, J.D., Lorch, P.D., Dennis, P.M., Tans, E., Robison, T., Millspaugh, J.J., Montgomery, R.A., 2019. What does urbanization actually mean ? A framework for urban metrics in wildlife research. J. Appl. Ecol. 56 (5), 1289–1300. https://doi.org/ 10.1111/1365-2664.13358.
- Mukerjee, C.M., Carrick, J., Walker, J.C., Woods, R.L., 2003. CASE REPORT Pulmonary strongyloidiasis presenting as chronic bronchitis leading to interlobular septal fibrosis and cured by treatment. Respirology 8 (4), 536–540. https://doi.org/ 10.1046/j.1440-1843.2003.00505.x.
- Mul, I.F., Paembonan, W., Singleton, I., Wich, S.A., Van Bolhuis, H.G., 2007. Intestinal parasites of free-ranging , Semicaptive , and captive Pongo abelii in Sumatra , Indonesia. J. Primatol. 28 (2), 407–420. https://doi.org/10.1007/s10764-007-9119-7.
- Nahallage, C.A., Huffman, M.A., Kuruppu, N., Weerasingha, T., 2008. Diurnal primates in Sri Lanka and people's perception of them. Primate Conserv. 23 (1), 81–87. https://doi.org/10.1896/052.023.0109.
- Nahallage, C.A.D., Huffman, M.A., 2013. Macaque–human interactions in past and present-day Sri Lanka. In: Radhakrishna, S., Huffman, M., Sinha, A. (Eds.), The Macaque Connection. Dev. In Primatol.: Progress and Prospects, 43, pp. 135–148. https://doi.org/10.1007/978-1-4614-3967-7_9.
- Nekaris, K.A.I., de Silva Wijeyeratne, G., 2009. Toque macaque. In: The Primates of Sri Lanka. Sri Lanka Tourism Promotion Bureau, Colombo, pp. 100–119.
- Nutman, T.B., 2016. Human infection with Strongyloides stercoralis and other related Strongyloides species. Parasitology 263–273. https://doi.org/10.1017/ S0031182016000834.
- Osman, M., Safadi, D.E., Cian, A., Benamrouz, S., Nourrisson, C., Poirier, P., Pereira, B., Razakandrainibe, R., Delbac, F., Hamze, M., Viscogliosi, E., 2016. Prevalence and risk factors for intestinal Protozoan infections with cryptosporidium, Giardia, Blastocystis and Dientamoeba among schoolchildren in Tripoli, Lebanon. PLoS Neglected Trop. Dis. 10 (3), e0004496 https://doi.org/10.1371/journal. pntd.0004496, 1–17.
- Pillers, A.W.N., Southwell, T., 1929. Strongyloidosis of the woolly monkey (Lagothrix Humboldti). Ann. Trop. Med. Parasitol. 23 (1), 129. https://doi.org/10.1080/ 00034983.1929.11684598.
- Seguel, M., Gottdenker, N., 2017. The diversity and impact of hookworm infections in wildlife. Int. J. Parasitol. Parasites Wildl. 6 (3), 177–194. https://doi.org/10.1016/j. ijppaw.2017.03.007.

- Sharma, P., Kumar, V., Pal, B., Mandial, R.K., Jithendran, K.P., Rastogi, S., 2013. Prevalence of gastrointestinal parasitism in free ranging rhesus macaque (Macaca mulatta) of Himachal Pradesh. Rev. Inst. Med. trop. S. Paulo. 45, 193–195. http ://doi=10.1.1.733.6666.
- Shutt, K., Maclarnon, A., Heistermann, M., Semple, S., Shutt, K., Maclarnon, A., Heistermann, M., Semple, S., 2007. Grooming in Barbary macaques : better to give than to receive ? Grooming in Barbary macaques : better to give than to receive. Biol. Lett. 3 (3), 231–233. https://doi.org/10.1098/rsbl.2007.0052.
- Sleeman, J.M., Meader, L.L., Mudakikwa, A.B., Foster, J.W., Patton, S., 2000. Gastrointestinal parasites of mountain gorillas (Gorilla gorilla beringei) in the Parc National des Volcans, Rwanda. J. Zoo Wildl. Med. 31 (3), 322–328. https://doi.org/ 10.1638/1042-7260(2000)031[0322:GPOMGG]2.0.CO;2.
- Sorensen, E., Ismail, M., Amarasinghe, D.K., Hettiarachchi, I., Dassenaieke, T.S., 1996. The prevalence and control of soil-transmitted nematode infections among children and women in the plantations in Sri Lanka. Ceylon Med. J 41 (2), 37–41. PMID: 8771940.
- Strait, K., Else, J.G., Eberhard, M.L., 2012. Parasitic diseases of nonhuman primates. In: Nonhuman Primates in Biomed. Research, pp. 197–297. https://doi.org/10.1016/ B978-0-12-381366-4.00004-3.
- Thilakarathne, S.S., Rajakaruna, R.S., Fernando, D.D., Rajapakse, J., Perera, P.K., 2021. Veterinary Parasitology : regional Studies and Reports Gastro-intestinal parasites in two subspecies of toque macaque (*Macaca sinica*) in Sri Lanka and their zoonotic potential. Vet. Parasitol. Reg. Stud. Rep. 24, 100558. https://doi.org/10.1016/j. vprsr.2021.100558.
- Tompkins, D.M., Dobson, A.P., Arneberg, P., Begon, M.E., Cattadori, I.M., Greenman, J. V., Hudson, P.J., Newborn, D., Pugliese, A., Rizzoli, A.P., Rosa, R., Rosso, F., Wilson, k., 2002. Parasites and host population dynamics. Eco. Wildlife Dis. 45–62.
- Torgerson, P.R., Macpherson, C.N.L., 2011. Veterinary Parasitology the socioeconomic burden ofparasitic zoonoses. Global trends 182, 79–95. https://doi.org/10.1016/j. vetpar.2011.07.017.
- Uemura, E., Houser, W.D., Cupp, C.J., 1979. Strongyloidiasis in an infant orangutan (Pongo pygmaeus). J. Med. Primatol. 8, 282–288. https://doi.org/10.1159/ 000460212.
- Warren, K.S., 2001. Orang-utan Conservation: Epidemiological Aspects of Health

Management and Population Genetics. Doctoral dissertation, Murdoch University. Weerasekara, L.S., Wijesooriya, K.B., Ranawana, K.B., Vandercone, R.P., 2021. Survey of

- Weerasekara, L.S., Wijesooriya, K.B., Ranawana, K.B., Vandercone, R.P., 2021. Survey or relative abundance of diurnal primates in the giritale nature reserve, Sri Lanka. Asian Primates J. 9 (1), 2–9.
- Wenz, A., Heymann, E.W., Petney, T.N., Taraschewski, H.F., 2010. The influence of human settlements on the parasite community in two species of Peruvian tamarin. Parasitology 137 (4), 675–684. https://doi.org/10.1017/S0031182009991570.
- Wenz-mücke, A., Sithithaworn, P., Petney, T.N., Taraschewski, H., 2013. Human contact in fl uences the foraging behaviour and parasite community in long-tailed macaques. Parasitology 140 (6), 709–718. https://doi.org/10.1017/S003118201200203X.
- Weyher, A., Chapman, C.A., Ryan, S.J., Muriuki, S., 2006. Gastrointestinal parasites in crop raiding and wild foraging Papio anubis in Nigeria. Int. J. Primatol. 27 (6), 1519–1534. https://doi.org/10.1007/s10764-006-9089-1.

World Health Organization, 1994. Part 2-Basic Laboratory Methods in Medical Parasitology. World Health Organization, Geneva.

Wren, B.T., Remis, J., Camp, J.W., Gillespie, T.R., 2016. Number of grooming partners is associated with hookworm infection in wild Vervet monkeys (*Chlorocebus aethiops*). https://doi.org/10.1159/000448709, 47907, 168-179.

Youn, H., 2009. Review of zoonotic parasites in medical and veterinary fields in the Republic of Korea. https://doi.org/10.3347/kjp.2009.47.S.S133, 47, 133-142.

Zhang, Q., Han, S., Liu, K., Luo, J., Luo, J., He, H., 2019. Occurrence of selected zoonotic fecal pathogens and first molecular identification of Hafnia paralvei in wild Taihangshan macaques (*Macaca mulatta tcheliensis*) in China 2019. BioMed Res. Int. https://doi.org/10.1155/2019/2494913.