



Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.

Miscellaneous Antiviral Agents (Interferons, Imiquimod, Pleconaril)

Raphael Dolin

SHORT VIEW SUMMARY

INTERFERONS

- Interferons are potent cytokines that stimulate antiviral, immunomodulating, and antiproliferative effects.
- They are classified as type I (α , β), II (γ), or III (λ) (see Table 47-1) and administered subcutaneously or intramuscularly, but pharmacokinetic properties are not well linked to physiologic effects.
- Attachment to polyethylene glycol (pegylation) prolongs half-life and possibly decreases immunogenicity and results in more convenient dosing, but toxicities persist, including anemia and other cytopenias, depression, thyroid dysfunction, and fatigue.
- Major clinical use is in treatment of hepatitis B and hepatitis C (see Chapter 46).
- They are also approved for treatment of anogenital warts.

IMIQUIMOD AND RESIQUIMOD

- These agents are topically applied Toll-like receptor (TLR) 7 (imiquimod) and TLR7/TLR8 (resiquimod) agonists.
- Imiquimod is approved for treatment of anogenital warts and results in clearance of these lesions in 37% to 52% of cases.
- Local skin toxicity is common, consisting of erythema, burning, and tenderness.
- Resiquimod is an investigational compound with somewhat greater potency than imiquimod.
- Resiquimod has been studied in genital herpes simplex virus infection, with early suggestion of decreases in rates of recurrences, but a recent phase III study failed to support an effect on recurrences.

PLECONARIL

- An antipicornavirus drug, pleconaril inhibits viral replication by binding to a hydrophobic pocket on the viral capsid.
- In vitro activity is against almost all commonly isolated enteroviruses and 90% of rhinovirus clinical isolates.
- Orally administered and generally well tolerated, pleconaril induces CYP3A isoenzymes and therefore has multiple drug interactions.
- Its use has been studied in enteroviral meningitis, but effects on headache and illness duration were inconsistent.
- Pleconaril has reduced the duration of rhinovirus infection by 1 day but is not approved by the U.S. Food and Drug Administration for this indication.
- This agent is no longer under clinical development.

Antiviral agents are discussed here that have activity against a variety of viral infections in addition to those addressed in Chapters 43 to 46. The major clinical use of interferons is in treatment of hepatitis B and C, which is discussed in Chapter 46.

INTERFERONS Classification

Since their discovery in 1957 as mediators of the phenomenon of viral interference (i.e., inhibition of growth of one virus by another), interferons (IFNs) have become recognized as potent cytokines that are associated with complex antiviral, immunomodulating, and antiproliferative actions.¹⁻³ IFNs are proteins that are synthesized by eukaryotic cells in response to various inducers and that cause biochemical changes leading to a nonselective antiviral state in exposed cells of the same species. Three subfamilies of IFNs are recognized. Type I IFNs are the largest subfamily and include the IFN- α s (13 subtypes in humans) and the IFN- β s. The type II subfamily has only one member, IFN- γ . Type III is the subfamily most recently identified and includes IFN- λ ,⁴ of which there are three subtypes (λ 1, λ 2, λ 3), also known as interleukin (IL)-28, IL-29, and IL-28R.^{5,6} A fourth interferon λ subtype (λ 4) has been recently identified.^{6a} The type I IFNs are clustered on the short arm of chromosome 9 in humans,⁷ Type II IFN is on chromosome 12, and type III IFNs are encoded on chromosome 19.⁸ Formerly designated on the basis of the cell types from which they were derived, the IFN- α s, the IFN- β , and IFN- γ are the IFNs currently in clinical use (Table 47-1), whereas IFN- λ is being studied for hepatitis C. Each type is immunologically distinct and has different producer cells, inducers, and biologic effects and unique physicochemical characteristics.^{2,4,9}

The IFN- α s and IFN- β s are produced by almost all cells in response to viral infection and various other stimuli, including double-stranded RNA (dsRNA); bacteria; protozoa; mycoplasmas; polyanions; several low-molecular-weight organic compounds; and certain cytokines and growth factors, such as IL-1, IL-2, and tumor necrosis factor (TNF). IFN- γ production is restricted to T lymphocytes and natural killer cells responding to antigenic stimuli, mitogens, and certain cytokines, such

as IL-2. The IFN- λ s also appear to be produced by multiple cell types.¹⁰ The principal antiviral IFNs, IFN- α s and IFN- β s, are approximately 30% homologous at the amino-acid level. The human IFN- α s share a high degree of amino-acid sequence homology (>70%) but have differing in vitro antiviral and biologic effects on human cells.¹¹ Compared with the IFN- α s and IFN- β s, IFN- γ has less antiviral activity but more potent immunoregulatory effects, particularly with respect to macrophage activation, expression of class II major histocompatibility complex (MHC) antigens, and mediation of local inflammatory responses. Most IFNs in clinical use are produced by recombinant DNA techniques (see Table 47-1).

Mechanisms of Action

A wide range of animal viruses are sensitive to the antiviral actions of IFNs, although many DNA viruses are relatively insensitive and considerable differences in potency exist among viruses and assay systems. IFN activity is usually measured in terms of antiviral effects in cell culture. Typically, one unit of IFN activity is the amount present in a sample dilution that causes a 50% reduction in virus replication or expression in certain cell lines; this is generally expressed as international units (IU) relative to National Institutes of Health or World Health Organization reference standards.

IFNs are not directly antiviral but cause elaboration of effector proteins in exposed cells, which contribute to a state of viral resistance.^{11,12,13} The initial step involves IFN binding to specific cell surface receptors. For the type I IFNs (IFN- α s and IFN- β) the cognate receptor is IFNAR1/2. Type II IFNs (IFN- γ) use the homodimeric receptors, IFNGR1/2. Type III IFNs (IFN- λ) signal through a receptor complex consisting of IL10R2 and IFNLR1.^{6,14} IFN receptors are linked to the Janus kinase/signal transducer and activator of transcription (JAK-STAT) signaling pathways, which, through a multistep process, activate transcription factors that bind selectively to and upregulate approximately 100 IFN-regulated genes.¹⁵⁻¹⁷ The distinct pattern of STAT proteins activated by different IFNs is one mechanism for eliciting different cellular responses. For IFN- α and IFN- β , a three-protein complex

KEYWORDS

anogenital warts; common cold; enteroviral meningitis; imiquimod; interferon; interferon α ; interferon β ; interferon γ ; interferon λ ; pleconaril; resiquimod; Toll-like receptors 7 and 8 (TLR7, TLR8)

TABLE 47-1 Nomenclature and Classification of Human Interferons (IFNs)

SUBFAMILY	TYPE I	TYPE I	TYPE II	TYPE III
Class*	α	β	γ	λ
No. of subtypes	13	1	1	4
Receptor	IFNAR1/2	IFNAR1/2	IFNGR1/2	IL10R2, IFNLR1
Human chromosome	9	9	12	19
Commercial formulations	rIFN- α 2b (Intron A) IFN- α 2a (Roferon-A) Le-IFN- α n3 (Alferon N) Ly-IFN- α n1 (Wellferon) rIFNalfacon-1 (Infergen) Peg-IFN- α 2a (Pegasys) Peg-IFN- α 2b (PEG-Intron)	rIFN- β 1b (Betaseron) rIFN- β 1a (Avonex, Rebif)	rIFN- γ 1b (Actimmune) rIFN- γ (Immuneron)	Peg-IFN λ 1

*Type I classes in humans also include IFN ϵ and IFN ω .

known as IFN-stimulated gene factor 3 (ISGF-3) localizes to the nucleus and binds to a *cis*-acting DNA element (designated IRSE) that activates transcription of the target genes.¹

A family of IFN regulatory factors exists, and other pathways may contribute to regulation of the IFN response. Microarray analysis shows that many genes are upregulated by IFN- β but not by IFN- α or IFN- γ in vitro.¹⁵ The onset of IFN-induced antiviral action is rapid, and IFN exposure leads to production of more than two dozen cellular proteins. For many viruses, the primary antiviral effect of IFN in vitro is mediated by inhibition of viral protein synthesis. Depending on the virus and cell type, the antiviral actions of IFNs may also include inhibition of viral penetration or uncoating, synthesis or methylation of messenger (m)RNA, or viral assembly and release.

Among the better-characterized IFN-induced proteins are unique 2'-5'-oligoadenylate (2-5[A]) synthetases and protein kinase R (PKR), either of which can inhibit protein synthesis in the presence of dsRNA.¹ The 2-5(A) synthetase produces adenylate oligomers that activate a latent cellular endoribonuclease (RNase L) to cleave cellular and viral single-stranded (ss)RNAs, leading to inhibition of protein synthesis. Activated PKR selectively phosphorylates and inactivates eukaryotic initiation factor (eIF)-2 to impede translation. Activated PKR also phosphorylates the transcription factor inhibitor I κ B and mediates dsRNA-induced activation of nuclear factor (NF)- κ B, which is required for IFN- β synthesis. IFNs may also block mRNA capping by inhibiting transmethylation reactions. IFNs also induce human guanylate binding protein-1, which mediates antiviral activity for several RNA viruses; the soluble form of the low-density lipoprotein receptor inhibitory for rhabdovirus assembly¹⁸; the MxA protein (a guanosine triphosphatase with activity against orthomyxoviruses and certain RNA viruses)¹⁹; and the RNA-specific adenosine deaminase ADR1, which modifies RNA transcripts after transcription.¹

IFNs also inhibit hepatitis C virus (HCV) internal ribosome entry site-dependent RNA translation in vitro.²⁰ Induction of nitric oxide synthase seems to mediate a substantial antiviral effect of IFN- γ .²¹ Increased levels of 2-5(A) synthetase activity and MxA protein or mRNA in peripheral leukocytes are also used as a marker for IFN exposure or endogenous release.¹⁹

Except possibly for the Mx proteins and influenza viruses and for 2-5(A) synthetase/RNase-L and picornaviruses, no consistent correlations exist between induction of a particular protein and resistance to a specific virus across a range of cell types.¹³ A particular virus may be inhibited at several steps, and the principal inhibitory effect differs among virus families. Many viruses are able to counter IFN effects by blocking signaling and production or activity of selected IFN-inducible proteins.^{1,15} The NS5A protein of HCV represses the function of the IFN-induced PKR,²² and another hepatitis C virus protein E2 competitively inhibits PKR kinase activity. The NS1 gene of influenza is an IFN antagonist that binds dsRNA to inhibit IFN production and dsRNA-activated pathways. IFN exposure may also reduce the expression of certain cellular genes, including selected oncogenes and genes involved in collagen synthesis.

The viral and immune IFN systems are functionally nonredundant,^{1,23} and complex interactions exist between IFNs and between IFNs and other parts of the immune system.^{9,24} IFNs upregulate major

histocompatibility complex (MHC) class I expression and promote cytotoxic T-cell responses, regulate the expression of cytokines (IL-12, IL-15, IFN- γ) and chemokines that affect T-cell responses, alter expression of Toll-like receptors (TLRs), enhance natural killer cell cytotoxicity, and promote the differentiation of dendritic cells and T helper type 1 (Th1) lymphocytes.^{12,24} IFN- α is produced by macrophages and can modify macrophage functions, increasing phagocytosis and cytolytic activity. Consequently, IFNs may ameliorate viral infections by exerting direct antiviral effects and by modifying the immune response to infection. IFN-induced expression of MHC antigens may contribute to the antiviral actions of IFN by enhancing antigen presentation and the lytic effects of cytotoxic T lymphocytes. The IFN- α s, the IFN- β s, and IFN- γ lead to increased expression of class I MHC molecules, but only IFN- γ efficiently induces class II MHC molecules.¹ Several viruses, including cytomegalovirus and varicella-zoster virus, antagonize IFN- γ -induced MHC expression. In addition, proapoptotic and antiapoptotic genes are induced by IFNs¹⁵ and IFN- α and IFN- β are important mediators of apoptosis, including induction of TP53.^{1,25}

IFN titers generally appear at the sites of viral replication just after peak titers of virus and before humoral antibody responses. IFNs may mediate some of the systemic symptoms associated with viral infections and contribute to immunologically mediated tissue damage in certain viral diseases. High IFN titers are usually followed by a reduction of virus titers, although persistently elevated IFN titers have been recognized in certain chronic and acute viral infections (e.g., hemorrhagic fevers).

Pharmacokinetics

The prolonged biologic effects of IFNs are not easily related to serum concentrations or other conventional pharmacokinetic parameters. After intramuscular or subcutaneous injection of IFN- α , absorption is greater than 80%.^{26,27} Plasma levels are dose related, peaking at 4 to 10 hours and returning to baseline by 18 to 36 hours. Levels of 2-5(A) synthetase in peripheral blood mononuclear cells, which have been used as an index of biologic responsiveness to IFN, show increases beginning at 6 hours and lasting through 4 days after a single dose. An antiviral state in these cells is detectable at 1 hour, peaks at 24 hours, and slowly decreases to baseline by 6 days after injection. Intramuscular or subcutaneous injections of IFN- β result in negligible plasma levels, although increases in 2-5(A) synthetase may occur. Oral administration does not result in detectable serum IFN levels or increases in 2-5(A) synthetase activity in peripheral blood mononuclear cells.²⁸

After systemic administration, low levels of IFNs are detected in respiratory secretions, cerebrospinal fluid, eye, and brain. After intravenous dosing, cerebrospinal fluid levels average less than 1% of serum concentrations.²⁹ The IFN- α s are stable in most body fluids, whereas the IFN- β s and IFN- γ seem to lose activity readily. It is unknown, however, whether measurable IFN levels at a particular site accurately reflect its antiviral or other biologic activities. The IFN- α s and the IFN- β s are cleared rapidly in a complex fashion. Leukocyte and recombinant IFN- α species have a plasma elimination half-life ($t_{1/2\text{elim}}$) of 3 to 8 hours. The clearance of IFN includes inactivation by various body fluids, cellular uptake, and metabolism by body organs, primarily the kidney, although negligible biologically active IFN is excreted in the

urine. Clearance of IFN- α 2 is reduced by 64% to 79% in hemodialysis patients.³⁰

The attachment of polyethylene glycol to IFN slows absorption, decreases clearance, increases $t_{1/2\text{elim}}$, and results in higher and more sustained serum concentrations, so that once-weekly dosing is effective. Two types of pegylated (peg) IFN- α are currently approved: peg-IFN- α 2a has a 40-kDa branched polyethylene glycol moiety attached by a stable amide bond to lysine residues within the IFN protein, and peg-IFN- α 2b has a 12-kDa linear moiety attached to histidine residues. Peg-IFN- α 2a is more stable and dispensed in solution, whereas peg-IFN- α 2b requires reconstitution before use. Peg-IFN- α 2a is cleared primarily by the liver, whereas about 30% of peg-IFN- α 2b is cleared renally.³⁰ For peg-IFN- α 2a (multiple 180- μ g doses), a peak serum concentration of 26 ng/mL occurs at about 45 hours after the dosing, and $t_{1/2\text{elim}}$ is 80 to 90 hours. Steady-state serum levels are attained 5 to 8 weeks after initiation of weekly dosing. Moderate renal impairment and presence of cirrhosis do not affect pharmacokinetics, although clearance is reduced by 25% to 45% in patients with renal failure on hemodialysis. For peg-IFN- α 2b, dose-related maximal plasma concentrations (1.4 ng/mL with multiple doses of 1.5 μ g/kg) occur at 15 to 44 hours after dosing and decline with a $t_{1/2\text{elim}}$ of 30 to 40 hours, or about 10-fold longer than for IFN- α 2b.²⁸ Some accumulation occurs with repetitive dosing. Dosage reductions in both peg-IFNs are indicated in end-stage renal disease.

Interactions

IFN and its inducers reduce the metabolism of various drugs by the hepatic cytochrome P-450–dependent mixed-function oxidase system and specifically decrease CYP1A2-mediated clearance of theophylline. IFNs may increase the neurotoxic, hematotoxic, or cardiotoxic effects of other drugs, including increased risk for anemia with ribavirin.

Toxicity

Purified natural and recombinant IFNs are associated with dose-related immediate- and late-onset toxicities.³¹ Adverse effects are generally mild and reversible at dosages of less than 5 million IU/day.²⁷ Intramuscular and subcutaneous injections of IFN doses of 1 to 2 million IU or more are usually associated with an acute influenza-like syndrome, including fever, chills, headache, malaise, myalgia, arthralgia, nausea, vomiting, and diarrhea, especially during the first week of therapy. Symptoms begin several hours after administration and are most prominent 8 to 24 hours after dosing. Despite more prolonged blood levels, the duration of influenza-like symptoms after peg-IFN is similar to that after conventional IFNs.³⁰ Tolerance develops in most patients within several weeks. Febrile responses can be moderated by pretreatment with various antipyretics. Half of patients receiving intralesional therapy for genital warts experience the influenza-like illness. Intralesional IFN also causes discomfort at the injection site and leukopenia. Local reactions consisting of tenderness and erythema also occur after subcutaneous injection, and intranasal IFN causes local irritation.

Major toxicities that limit dosage and duration of IFN therapy are bone marrow suppression with granulocytopenia and thrombocytopenia; neuropsychiatric disturbance manifested by depression, anxiety, somnolence, confusion, behavioral disturbance, electroencephalographic changes, and, rarely, seizures; reversible neurasthenia with profound fatigue, anorexia, weight loss, and myalgia; thyroid dysfunction and autoimmune thyroiditis; and cardiotoxicity with hypotension, arrhythmias, and reversible cardiomyopathy. Psychiatric disturbance and depression are more common in patients with preexisting disorders but can also occur in otherwise healthy individuals. Elevations in hepatic enzymes and triglycerides and retinopathy are common.³² IFN may lead to the development of or exacerbate various immunologically mediated disorders, including sarcoidosis, systemic lupus erythematosus, psoriasis, vitiligo, lichen planus, and eczematoid skin lesions. Rare pulmonary manifestations include interstitial pneumonia, bronchiolitis obliterans, organizing pneumonia, asthma, and pleural effusion.³³ Alopecia, proteinuria, renal insufficiency, interstitial nephritis, autoantibody formation, bacterial infections, and hepatotoxicity occur.³⁴ Acute allergic reactions are rare. Patients with autoimmune chronic hepatitis, who may have false-positive enzyme immunoassay tests for

anti-HCV antibodies, can experience worsening of their disease if treated with IFN.³⁵

The adverse effects of peg-IFNs are similar to the adverse effects with conventional IFNs, although dose-related neutropenia and thrombocytopenia and injection site reactions are more common. About 50% of peg-IFN-treated patients with chronic hepatitis C develop fatigue and systemic symptoms after injections; 20% to 30% experience depression or other psychiatric reactions; and approximately 10% to 16% discontinue treatment because of adverse events, most commonly psychiatric disorders.^{36,37} Peg-IFN- α 2a may be associated with a lesser frequency of depression.³⁷

The development of serum neutralizing antibodies to exogenous IFNs varies with the IFN type, dosage, and route of administration but may be more common with IFN- α 2a.³⁸ Neutralizing antibodies may be associated infrequently with loss of clinical responsiveness.²⁷ Pegylation may reduce the immunogenicity of IFNs, and anti-PEG antibody seems to be rare.

IFNs may impair fertility and alter hormone levels in women. IFN is an abortifacient in monkeys at high dosages and has been used in small numbers of pregnant women, so safety during pregnancy is not established.³⁹ It is classified as pregnancy category C.

Clinical Studies

IFNs have undergone clinical studies in a broad variety of infections, malignancies, and other diseases. Depending on the IFN type, recombinant and natural IFN- α (see Table 47-1) are approved in the United States for treatment of condyloma acuminatum, chronic hepatitis C, chronic hepatitis B, Kaposi sarcoma in human immunodeficiency virus (HIV)-infected patients, and other malignancies. IFN- β is approved for management of multiple sclerosis. Recombinant IFN- γ is approved for treatment of chronic granulomatous disease. The major uses for interferons are treatments for hepatitis B and C, and the clinical studies of IFNs for those infections are discussed in detail in Chapters 46 and 119.

Herpesviruses

Although IFN is associated with antiviral effects against herpes simplex viruses (HSVs), no consistent reductions in symptoms or lesion duration have been observed with topical or systemic IFN treatment of genital herpes.⁴⁰ Topical IFN seems to have some activity in combination with trifluridine in drug-resistant mucocutaneous HSV infections. In superficial HSV keratitis, combined administration of topical IFN- α with trifluridine or acyclovir seems to be more effective than single-agent therapy.

In localized herpes zoster in cancer patients, early treatment with high-dose IFN- α (about 36 million IU/day for 5 to 7 days) reduces the risk for cutaneous or visceral dissemination, but systemic reactions are frequent, and more effective antiviral agents are available. IFN is ineffective in preventing cytomegalovirus (CMV) infection in bone marrow recipients or in treating CMV pneumonia.

Human Immunodeficiency Virus

HIV-infected patients frequently have detectable IFN levels, and plasma inhibitors of IFN activity are often present during the acquired immunodeficiency syndrome. High doses of IFN- α induce 10% to 40% response rates in patients with Kaposi sarcoma without benefiting concurrent herpesvirus infections or immune functions.⁴¹ IFN treatment is associated not only with dose-related antiretroviral effects, particularly in early-stage infection, but also with adverse effects.⁴² IFN seems to benefit HIV-related thrombocytopenia⁴³ and eosinophilic folliculitis.

Papillomavirus

Intralesional and systemic forms of administration of IFN produce some regression of anogenital warts,⁴⁴ although more cost-effective and better tolerated modalities are available (see Chapter 146).⁴⁵ Intralesional injection of various natural and recombinant IFNs is associated with complete clearance of injected warts in 42% to 62% of patients within 12 to 20 weeks.⁴⁶ Responders have low relapse rates (20% to 30%). Responsiveness is poor in HIV-infected patients and patients with chronic lesions. Intralesional IFN does not reliably increase the

response to other local therapies.⁴⁶ Mild to moderate systemic side effects (8% to 10% dropout rate), pain and irritation at injection site, and leukopenia ($\leq 30\%$) are common with intranasal IFN. Topical IFN gel provides inconsistent effects and does not seem to reduce the recurrence rate substantially after ablative therapies.⁴⁵

Systemic IFN may provide adjunctive benefit in recurrent juvenile laryngeal papillomatosis. Most children have some initial decrease in lesions, but recurrence rates are high after cessation of therapy, and the long-term response to parenteral IFN- α is variable.⁴⁷ Laryngeal disease in older patients seems to be more responsive.

Respiratory Viruses

Except against adenovirus, IFNs have broad-spectrum antiviral activity against respiratory viruses *in vitro*, including severe acute respiratory syndrome (SARS) coronavirus.⁴⁸ In experimentally induced infections in humans, intranasal administration of leukocyte or recombinant IFN- α is protective against rhinovirus, coronavirus 229E, respiratory syncytial virus (RSV), and, to a lesser extent, influenza virus infections.⁴⁹ Under natural conditions, prophylactic intranasal IFN- α is protective only against rhinovirus colds, however, and long-term use is limited by the occurrence of nasal side effects. Intranasal IFN- α is ineffective in treating rhinovirus colds. The IFN- α s and the IFN- β s and IFN- γ inhibit SARS coronavirus replication *in vitro*,⁴⁸ and the systemic IFN- α s have been used to treat SARS coronavirus illness, but the beneficial effect, if any, is unclear.^{50,51}

IMIQUIMOD AND RESIQUIMOD

Imiquimod (Aldara) is an imidazoquinoline compound (Fig. 47-1A) that is a TLR7 agonist; it acts as a topical immune response modifier lacking direct antiviral effects. Imiquimod exposure causes activation of immune cells (monocytes, macrophages, natural killer cells) to produce antiviral cytokines, particularly IFN- α and TNF- α and interleukin (IL)-12, IL-10, IL-1, IL-6, and IL-8.^{52,53} Imiquimod indirectly enhances acquired immune responses through activation of antigen-presenting dendritic cells, including Langerhans cells and Th1 lymphocytes. IFN- γ production from T cells stimulates cytotoxic T lymphocytes, which is important in clearance of virally infected cells. Clinical responses in anogenital warts are associated with decreases in human papillomavirus (HPV) DNA copies and RNA transcripts in treated skin.⁵⁴ Resiquimod is a structurally related investigational compound (see Fig. 47-1B) that is a TLR7/TLR8 agonist and is associated with greater stimulation of cytokines and with activation of dendritic cells.⁵⁵

Topical imiquimod 5% cream is approved for patient-applied treatment of anogenital warts and has been used in other mucocutaneous infections and dermatologic conditions.⁵² In immunocompetent patients, imiquimod (three overnight applications [for approximately 8 hours] weekly for up to 16 weeks) leads to complete wart clearance in 37% to 52%.^{45,52,56} Clearance rates (39% to 52%) are not different with 4, 8, 12, or 16 weeks of treatment.⁵⁷ Daily application increases local adverse effects without increasing clearance rates.⁵⁸ Imiquimod and podophyllin are similar in efficacy with 50% and 55% clearance rates.⁵⁹ Clearance rates are higher in women than in men and substantially lower ($<15\%$) in HIV-infected individuals.⁶⁰⁻⁶² In one study in women, clinical outcome was related to HPV type: complete response rates were 76% for HPV-6, 67% for HPV-11, 35% for HPV-6 and HPV-11 coinfection, and 6% for other HPV types.⁶³ The time to

clearance averages 8 to 10 weeks. Recurrences are less common (14% to 19%) than after ablative therapies, and re-treatment is frequently successful.⁵² Imiquimod may be useful as an adjunct to laser or surgical ablative therapies.⁵²

Imiquimod has been used for warts at nongenital sites⁵³ and in other types of HPV-related conditions. Both grade 2 and grade 3 intraepithelial neoplasia of the vulva respond to imiquimod therapy.⁶⁴ HPV anal infection in HIV-infected men may⁶⁵ or may not⁶⁶ respond. Imiquimod seems beneficial in refractory cutaneous leishmaniasis in combination with other drugs,^{67,68} although other authors did not observe any benefit from combining standard intravenous antimony therapy with imiquimod.⁶⁹ Depending on the level of immunocompetence, imiquimod may be beneficial for treatment of molluscum contagiosum.^{52,53} Imiquimod has been used successfully to treat chronic genital herpes resistant to acyclovir⁷⁰ and to foscarnet⁷¹ in immunocompromised individuals with HIV and drug-induced immunosuppression.⁷² No beneficial effects on lesions or recurrences are seen in immunocompetent adults with genital herpes.⁷³ Imiquimod 5% cream made recurrent herpes labialis lesions worse compared with the vehicle cream as control but increased the time to the next recurrence 80% from a median of 50 days to 91 days in the control and imiquimod-treated groups.⁷⁴ Other viral infections reported to respond to imiquimod include oral hairy leukoplakia,⁷⁵ orf,⁷⁶ and Kaposi sarcoma.⁷⁷ Tinea pedis resolved during imiquimod treatment of a contiguous basal cell carcinoma.⁷⁸

Because of encouraging preliminary findings, topical resiquimod was investigated for treatment of recurrent genital herpes in an effort to reduce recurrences.⁷⁹ However, a controlled study did not show an effect of imiquimod on the rate of recurrences.⁷³ Resiquimod 0.01% gel applied to recurrent genital herpes lesions two times a week for 3 weeks neither reduced healing time nor shedding of herpes simplex virus DNA by polymerase chain reaction assay compared with a control gel.⁸⁰ The same treatment regimen was associated in the subsequent 60 days with a reduction in lesional recurrence rate from 16% to 10%, and with a trend toward reduced shedding from 17% to 10%. Seven months later, recurrence rates were reduced from 26% to 10% in control and resiquimod groups.⁸¹ However, a phase III study of resiquimod did not show reduced rates of recurrence, and further development of the drug to treat anogenital herpes has been halted.^{80,82}

Topical application of imiquimod 5% cream to affected skin can result in systemic absorption with serum levels of 0.1 to 3.5 ng/mL. Patients treated with imiquimod need to wash the affected area on awakening to remove residual drug. About two thirds of treated patients experience local erythema. Application site reactions with erythema, irritation, pruritus, burning, tenderness, and scabbing (and less often with erosion or ulceration at the wart site and other exposed areas) are generally mild to moderate in intensity. These usually resolve within 2 weeks of cessation of the drug. The frequency of local reactions relates to frequency of application, and use in genital herpes may delay healing.⁷³ Severe local reactions, including pain, erythema, or scarring, are rare.⁶⁰ Unusual local reactions include erosive cheilitis⁸³ and aphthous ulcers,⁸⁴ angioedema and urticaria,⁸⁵ and worsening of psoriasis.⁸⁶ Systemic reactions to imiquimod, such as fatigue and influenza-like symptoms, have been reported infrequently.⁸⁷ Generalized psoriasis,⁸⁶ eczema,⁸⁸ exacerbations of myasthenia gravis,⁸⁹ and worsening of HLA-B27 spondyloarthritis⁹⁰ have been reported during topical therapy with imiquimod.

Systemic reactions may be caused not by the drug itself but rather by diffusion of cytokines from the skin into the systemic circulation. During repeated applications, small concentrations (<10 ng/mL of imiquimod) can be detected in the blood.⁹¹ Safety during pregnancy has not been established (pregnancy category B), but there are case reports of its safe use in pregnant women.^{92,93} Preclinical studies indicate that imiquimod is not genotoxic or teratogenic.

PLECONARIL

Pleconaril (3-[3,5-dimethyl-4([3-(3-methyl-5-isoxazolyl)propyl]oxy)phenyl]-5-[trifluoromethyl]-1,2,4-oxadiazole) is an orally active anti-picornaviral agent (Fig. 47-2). Pleconaril inhibits picornavirus replication by binding to a specific hydrophobic pocket within the viral capsid and preventing viral attachment or uncoating of the genome. In cell

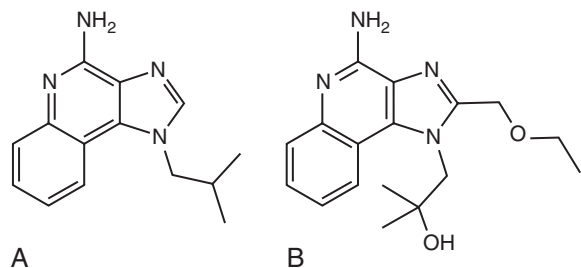


FIGURE 47-1 Chemical structure of (A) imiquimod and (B) resiquimod.

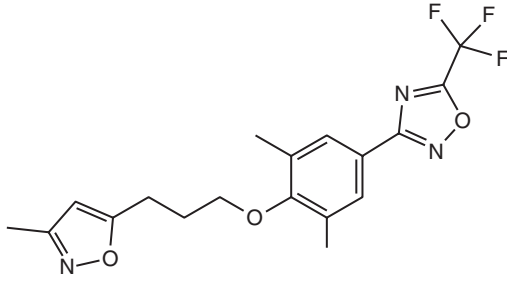


FIGURE 47-2 Chemical structure of pleconaril.

culture, pleconaril inhibits replication of almost all commonly isolated enterovirus serotypes⁹⁴ and approximately 90% of rhinovirus clinical isolates.⁹⁵ Pleconaril is active in murine and human models of coxsackievirus infection.^{94,96}

In adults, oral bioavailability is about 70% in the fed state, and peak plasma concentrations average 2.2 $\mu\text{g/mL}$ after doses of 400 mg.⁹⁷ Pleconaril undergoes hepatic metabolism, and less than 1% is excreted unchanged in the urine. The initial plasma $t_{1/2\text{elim}}$ averages 2 to 3 hours,

but there is a prolonged terminal $t_{1/2\text{elim}}$ of approximately 180 hours.⁹⁷ Single oral doses of 5 mg/kg in children provide maximal plasma concentrations of 1.3 $\mu\text{g/mL}$ and approximately 40% lower overall drug exposure because of a larger volume of distribution and more rapid clearance. Neonates seem to require higher dosages in part because of lower bioavailability.⁹⁸

Pleconaril has been generally well tolerated, and the most common adverse events have been headache, nausea, diarrhea, and abdominal discomfort. Pleconaril induces CYP3A isoenzymes, however, and consequently has the potential for multiple drug interactions, including with oral contraceptives.⁹⁹ In children or adults with enteroviral meningitis, pleconaril has inconsistent effects on headache and illness duration.⁹⁷ Pleconaril (400 mg three times a day for 5 days) reduces the duration of uncomplicated rhinovirus colds by about 1 day^{99,100} but was not approved by the U.S. Food and Drug Administration for this indication. Pleconaril antiviral effects and clinical outcomes are related to in vitro susceptibility.¹⁰⁰ Pleconaril treatment (15 mg/kg/day [children] and 600 to 1200 mg/day [adults] in divided doses for 7 to 10 days) seems to be beneficial in some patients with severe or life-threatening enteroviral syndromes, including chronic enteroviral meningoencephalitis in agammaglobulinemic patients and possibly neonatal enteroviral sepsis.^{97,101,102} The drug was formerly available for compassionate use, but it is no longer so.

Key References

The complete reference list is available online at Expert Consult.

- Ng D, Gommerman L. The regulation of immune responses by DC derived type I IFN. *Front Immunol*. 2013;4:94.
- Bonjardim CA, Ferreira PCP, Kroon EG. Interferons: signaling, antiviral and viral evasion. *Immunol Lett*. 2009;122:1-11.
- Kotenko SV, Gallagher G, Baurin V, et al. IFN- λ s mediate antiviral protection through a distinct class II cytokine receptor complex. *Nat Immunol*. 2003;4:1:69-77.
- Sheppard P, Kindsvogel W, Xu W, et al. IL-28, IL-29 and their class II cytokine receptor IL-28R. *Nat Immunol*. 2003;4:63-68.
- Hamming OJ, Terczynska-Dyla A, Hashaam S, et al. Characterization of the newly identified interferon lambda 4. *Cytokine*. 2013;63:269.
- Kotenko SV. IFN- λ s. *Curr Opin Immunol*. 2011;23:583-590.
- Donnelly RP, Kotenko SV. Interferon-lambda: a new addition to an old family. *J Interferon Cytokine Res*. 2010;30:8:555-564.
- O'Shea JJ, Plenge R. JAK and STAT signaling molecules in immunoregulation and immune-mediated disease. *Immunity*. 2012;36:542-550.
- Marsili G, Remoli AL, Sgarbanti M, et al. HIV-1, interferon and the interferon regulatory factor system: an interplay between induction, antiviral responses and viral evasion. *Cytokine Growth Factor Rev*. 2012;23:255-270.
- Ozaslan E, Yilmaz R, Simsek H, et al. Interferon therapy for acute hepatitis C during pregnancy. *Ann Pharmacother*. 2002;36:1715-1718.
- Beutner KR, Wiley DJ, Douglas JM, et al. Genital warts and their treatment. *Clin Infect Dis*. 1999;28(suppl 1):S37-S56.
- Higgins PG, Barrow GI, Tyrrell DA, et al. The efficacy of intranasal interferon alpha-2a in respiratory syncytial virus infection in volunteers. *Antiviral Res*. 1990;14:3-10.
- Loutfy MR, Blatt LM, Siminovitch KA, et al. Interferon alfacon-1 plus corticosteroids in severe acute respiratory syndrome: a preliminary study. *JAMA*. 2003;290:3222-3228.
- Zhao Z, Zhang F, Xu M, et al. Description and clinical treatment of an early outbreak of severe acute respiratory syndrome (SARS) in Guangzhou, PR China. *J Med Microbiol*. 2003;52:715-720.
- Garland SM. Imiquimod. *Curr Opin Infect Dis*. 2003;16:85-89.
- Meyer T, Surber C, French LE, et al. Resiquimod, a topical drug for viral skin lesions and skin cancer. *Expert Opin Investig Drugs*. 2013;22:149-159.
- Jaffary F, Musini V, Nilforoushadeh MA, et al. Systematic review of imiquimod for the treatment of external genital wart. *Int J Pharmacol*. 2007;3:1-10.
- Van Seters M, Van Beurden M, Ten Kate FJW, et al. Treatment of vulvar intraepithelial neoplasia with topical imiquimod. *N Engl J Med*. 2008;358:1465-1473.
- Arevalo I, Ward B, Miller R, et al. Successful treatment of drug-resistant cutaneous leishmaniasis in humans by use of imiquimod, an immunomodulator. *Clin Infect Dis*. 2001;33:1847-1851.
- Martinez S, Molina JM, Scieux C, et al. Topical imiquimod for recurrent acyclovir-resistant HSV infection. *Am J Med*. 2006;119:e9-e11.
- Lascaux AS, Caumes E, Deback C, et al. Successful treatment of acyclovir and foscarnet resistant herpes simplex virus lesions with topical imiquimod in patients infected with human immunodeficiency virus type 1. *J Med Virol*. 2012;84:194-197.
- Brummitt CF. Imiquimod 5% cream for the treatment of recurrent, acyclovir-resistant genital herpes. *Clin Infect Dis*. 2006;42:575.
- Schacker TW, Conant M, Thoming C, et al. Imiquimod 5-percent cream does not alter the natural history of recurrent herpes genitalis: a phase II, randomized, double-blind, placebo-controlled study. *Antimicrob Agents Chemother*. 2002;46:3243-3248.
- Bernstein DI, Spruance SL, Arora SS, et al. Evaluation of imiquimod 5% cream to modify the natural history of herpes labialis: a pilot study. *Clin Infect Dis*. 2005;41:808-814.
- Spruance SL, Tyring SK, Smith MH, et al. Application of a topical immune response modifier, resiquimod gel, to modify the recurrence rate of recurrent genital herpes: a pilot study. *J Infect Dis*. 2001;184:196-200.
- Fife KH, Meng TC, Ferris DG, et al. Effect of resiquimod 0.01% gel on lesion healing and viral shedding when applied to genital herpes lesions. *Antimicrob Agents Chemother*. 2008;52:477-482.
- Mark KE, Corey L, Meng TC, et al. Topical resiquimod 0.01% gel decreases herpes simplex virus type 2 genital shedding: a randomized, controlled trial. *J Infect Dis*. 2007;195:1324-1331.
- Mark KE, Spruance S, Kinghorn GR, et al. Three phase III randomized controlled trials of topical resiquimod gel 0.01% to prevent genital herpes recurrences [abstract P-256]. Presented at the 17th Annual Meeting of the International Society on Sexual Transmitted Diseases Research. Seattle, WA, 2007.
- Jacobs AA, Snavely N, Markus J, et al. Vasodilatory adverse events associated with topical imiquimod 5 percent cream. *Dermatol Online J*. 2008;14:4.
- Taylor CL, Maslen M, Kapembwa M. A case of severe eczema following use of imiquimod 5% cream. *Sex Transm Infect*. 2006;82:227-228.
- Pevear DC, Tull TM, Seipel ME, et al. Activity of pleconaril against enteroviruses. *Antimicrob Agents Chemother*. 1999;43:2109-2115.
- Hayden FG, Herrington DT, Coats TL, et al. Efficacy and safety of oral pleconaril for treatment of picornavirus colds in adults: results of two double-blind, randomized, placebo-controlled trials. *Clin Infect Dis*. 2003;36:1523-1532.
- Rotbart HA, Webster AD. Treatment of potentially life-threatening enterovirus infections with pleconaril. *Clin Infect Dis*. 2001;32:228-235.
- Webster AD. Pleconaril—an advance in the treatment of enteroviral infection in immunocompromised patients. *J Clin Virol*. 2005;32:1-6.

References

- Samuel CE. Antiviral actions of interferons. *Clin Microbiol Rev.* 2001;14:778-809.
- Baron S, Coppenhaver DH, Doanzani F. Introduction to the interferon system. In: Baron S, ed. *Interferon: Principles and Medical Applications*. Galveston, TX: University of Texas Medical Branch; 1992:1-15.
- Ng D, Gommerman L. The regulation of immune responses by DC derived type I IFN. *Front Immunol.* 2013;4:94.
- Bonjardim CA, Ferreira PCR, Kroon EG. Interferons: signaling, antiviral and viral evasion. *Immunol Lett.* 2009;122:1-11.
- Kotenko SV, Gallagher G, Baurin V, et al. IFN- λ s mediate antiviral protection through a distinct class II cytokine receptor complex. *Nat Immunol.* 2003;4:1:69-77.
- Sheppard P, Kindsvogel W, Xu W, et al. IL-28, IL-29 and their class II cytokine receptor IL-28R. *Nat Immunol.* 2003;4:63-68.
- Hamming OJ, Terczyńska-Dyla A, Hashaam S, et al. Characterization of the newly identified interferon lambda 4. *Cytokine.* 2013;63:269.
- Sadler AJ, Williams BR. Interferon-inducible antiviral effectors. *Nat Rev Immunol.* 2008;8:559-568.
- Kotenko SV. IFN- λ s. *Curr Opin Immunol.* 2011;23:583-590.
- Dianzani F, Antonelli G. Mechanisms of action of the interferons: biological basis. In: Stuart-Harris R, Penny R, eds. *Clinical Applications of the Interferons*. London: Chapman & Hall; 1997:20-31.
- Donnelly RP, Kotenko SV. Interferon-lambda: a new addition to an old family. *J Interferon Cytokine Res.* 2010;30:8:555-564.
- Finter NB. Why are there so many subtypes of alpha-interferons? *J Interferon Res.* 1991;(spec issue):185-194.
- Brierley MM, Fish EN. Review: IFN-alpha/beta receptor interactions to biologic outcomes—understanding the circuitry. *J Interferon Cytokine Res.* 2002;22:835-845.
- Sen GC, Ransohoff RM. Interferon-induced antiviral actions and their regulation. *Adv Virus Res.* 1993;42:57-102.
- Kotenko SV, Gallagher G, Baurin VV, et al. IFN-lambdas mediate antiviral protection through a distinct class II cytokine receptor complex. *Nat Immunol.* 2003;4:69-77.
- Der SD, Zhou A, Williams BR, et al. Identification of genes differentially regulated by interferon alpha, beta, or gamma using oligonucleotide arrays. *Proc Natl Acad Sci U S A.* 1998;95:15623-15628.
- Pfeffer LM, Mullersman JE, Pfeffer SR, et al. STAT3 as an adapter to couple phosphatidylinositol 3-kinase to the IFNAR1 chain of the type I interferon receptor. *Science.* 1997;276:1418-1420.
- O'Shea JJ, Plenge R. JAK and STAT signaling molecules in immunoregulation and immune-mediated disease. *Immunity.* 2012;36:542-550.
- Fischer DG, Tal N, Novick D, et al. An antiviral soluble form of the LDL receptor induced by interferon. *Science.* 1993;262:250-253.
- Roers A, Hochkeppel HK, Horisberger MA, et al. MxA gene expression after live virus vaccination: a sensitive marker for endogenous type I interferon. *J Infect Dis.* 1994;169:807-813.
- Kato J, Kato N, Moriyama M, et al. Interferons specifically suppress the translation from the internal ribosome entry site of hepatitis C virus through a double-stranded RNA-activated protein kinase-independent pathway. *J Infect Dis.* 2002;186:155-163.
- Karupiah G, Xie QW, Buller RM, et al. Inhibition of viral replication by interferon-gamma-induced nitric oxide synthase. *Science.* 1993;261:1445-1448.
- Gale MJ, Korth MJ, Tang NM, et al. Evidence that hepatitis C virus resistance to interferon is mediated through repression of the PKR protein kinase by the nonstructural 5A protein. *Virology.* 1997;230:217-227.
- Marsili G, Remoli AL, Sgarbanti M, et al. HIV-1, interferon and the interferon regulatory factor system: an interplay between induction, antiviral responses and viral evasion. *Cytokine Growth Factor Rev.* 2012;23:255-270.
- Biron CA. Interferons alpha and beta as immune regulators: a new look. *Immunity.* 2001;14:661-664.
- Takaoka A, Hayakawa S, Yanai H, et al. Integration of interferon-alpha/beta signalling to p53 responses in tumour suppression and antiviral defence. *Nature.* 2003;424:516-523.
- Wills RJ. Clinical pharmacokinetics of interferons. *Clin Pharmacokinet.* 1990;19:390-399.
- Haria M, Benfield P. Interferon-alpha-2a: a review of its pharmacological properties and therapeutic use in the management of viral hepatitis. *Drugs.* 1995;50:873-896.
- Witt PL, Goldstein D, Storer BE, et al. Absence of biological effects of orally administered interferon-beta ser. *J Interferon Res.* 1992;12:411-413.
- Smith RA, Norris F, Palmer D, et al. Distribution of alpha interferon in serum and cerebrospinal fluid after systemic administration. *Clin Pharmacol Ther.* 1985;37:85-88.
- Glue P, Fang JW, Rouzier-Panis R, et al. Pegylated interferon-alpha2b: pharmacokinetics, pharmacodynamics, safety, and preliminary efficacy data. Hepatitis C Intervention Therapy Group. *Clin Pharmacol Ther.* 2000;68:556-567.
- Quesada JR. Toxicity and side effects of interferons. In: Baron S, ed. *Interferon: Principles and Medical Applications*. Galveston, TX: University of Texas Medical Branch; 1992:426-432.
- Kawano T, Shigehira M, Uto H, et al. Retinal complications during interferon therapy for chronic hepatitis C. *Am J Gastroenterol.* 1996;91:309-313.
- Kumar KS, Russo MW, Borczyk AC, et al. Significant pulmonary toxicity associated with interferon and ribavirin therapy for hepatitis C. *Am J Gastroenterol.* 2002;97:2432-2440.
- Bayraktar Y, Bayraktar M, Gurakar A, et al. A comparison of the prevalence of autoantibodies in individuals with chronic hepatitis C and those with autoimmune hepatitis: the role of interferon in the development of autoimmune diseases. *Hepatology.* 1997;24:417-425.
- Papo T, Marcellin P, Bernuau J, et al. Autoimmune chronic hepatitis exacerbated by alpha-interferon. *Ann Intern Med.* 1992;116:51-53.
- Manns MP, McHutchison JG, Gordon SC, et al. Peginterferon alfa-2b plus ribavirin compared with interferon alfa-2b plus ribavirin for initial treatment of chronic hepatitis C: a randomized trial. *Lancet.* 2001;358:958-965.
- Fried MW, Shiffman ML, Reddy KR, et al. Peginterferon alfa-2a plus ribavirin for chronic hepatitis C virus infection. *N Engl J Med.* 2002;247:975-982.
- Antonelli G, Currenti M, Turriziani O, et al. Neutralizing antibodies to interferon-alpha: relative frequency in patients treated with different interferon preparations. *J Infect Dis.* 1991;163:882-885.
- Ozaslan E, Yilmaz R, Simsek H, et al. Interferon therapy for acute hepatitis C during pregnancy. *Ann Pharmacother.* 2002;36:1715-1718.
- Lebwohl M, Sacks S, Conant M, et al. Recombinant alpha-2 interferon gel treatment of recurrent herpes genitalis. *Antiviral Res.* 1992;17:235-243.
- Krown SE. The role of interferon in the therapy of epidemic Kaposi's sarcoma. *Semin Oncol.* 1987;14(suppl 3):27-33.
- Berglund O, Engman K, Ehrnst A, et al. Combined treatment of symptomatic human immunodeficiency virus type 1 infection with native interferon-alpha and zidovudine. *J Infect Dis.* 1991;163:710-715.
- Marroni M, Greslele P, Landonio G, et al. Interferon-alpha is effective in the treatment of HIV-1-related, severe, zidovudine-resistant thrombocytopenia: a prospective, placebo-controlled, double-blind trial. *Ann Intern Med.* 1994;121:423-429.
- Frazer IH, McMillan AJ. Papillomatosis and condylomata acuminata. In: Stuart-Harris R, Penny R, eds. *Clinical Applications of the Interferons*. London: Chapman & Hall; 1997:79-90.
- Wiley DJ, Douglas J, Beutner K, et al. External genital warts: diagnosis, treatment, and prevention. *Clin Infect Dis.* 2002;35(suppl 2):S210-S224.
- Beutner KR, Wiley DJ, Douglas JM, et al. Genital warts and their treatment. *Clin Infect Dis.* 1999;28(suppl 1):S37-S56.
- Leventhal BG, Kashima HK, Mounts P, et al. Long-term response of recurrent respiratory papillomatosis to treatment with lymphoblastoid interferon alpha-N1. Papilloma Study Group. *N Engl J Med.* 1991;325:613-617.
- Cinatl J, Morgenstern B, Bauer G, et al. Treatment of SARS with human interferons. *Lancet.* 2003;362:293-294.
- Higgins PG, Barrow GI, Tyrrell DA, et al. The efficacy of intranasal interferon alpha-2a in respiratory syncytial virus infection in volunteers. *Antiviral Res.* 1990;14:3-10.
- Loutfy MR, Blatt LM, Siminovich KA, et al. Interferon alfacon-1 plus corticosteroids in severe acute respiratory syndrome: a preliminary study. *JAMA.* 2003;290:3222-3228.
- Zhao Z, Zhang F, Xu M, et al. Description and clinical treatment of an early outbreak of severe acute respiratory syndrome (SARS) in Guangzhou, PR China. *J Med Microbiol.* 2003;52:715-720.
- Garland SM. Imiquimod. *Curr Opin Infect Dis.* 2003;16:85-89.
- Skinner RB. Imiquimod. *Dermatol Clin.* 2003;21:291-300.
- Tyring SK, Arany I, Stanley MA, et al. A randomized, controlled, molecular study of condylomata acuminata clearance during treatment with imiquimod. *J Infect Dis.* 1998;178:551-555.
- Meyer T, Surber C, French LE, et al. Resiquimod, a topical drug for viral skin lesions and skin cancer. *Expert Opin Investig Drugs.* 2013;22:149-159.
- Jaffary F, Musini V, Nilforoushadeh MA, et al. Systematic review of imiquimod for the treatment of external genital wart. *Int J Pharmacol.* 2007;3:1-10.
- Garland SM, Waddell R, Mindel A, et al. An open-label phase II pilot study investigating the optimal duration of imiquimod 5% cream for the treatment of external genital warts in women. *Int J STD AIDS.* 2006;17:448-452.
- Gotovtseva EP, Kapadia AS, Smolensky MH, et al. Optimal frequency of imiquimod (Aldara) 5% cream for the treatment of external genital warts in immunocompetent adults: a meta-analysis. *Sex Transm Dis.* 2008;35:346-351.
- Yan J, Chen SL, Wang HN, et al. Meta-analysis of 5% imiquimod and 0.5% podophyllotoxin in the treatment of condylomata acuminata. *Dermatology.* 2006;213:218-223.
- Gilson RJ, Shupack JL, Friedman-Kien AE, et al. A randomized, controlled, safety study using imiquimod for the topical treatment of anogenital warts in HIV-infected patients. Imiquimod Study Group. *AIDS.* 1999;13:2397-2404.
- Sauder DN, Skinner RB, Fox TL, et al. Topical imiquimod 5% cream as an effective treatment for external genital and perianal warts in different patient populations. *Sex Transm Dis.* 2003;30:124-128.
- Herrera S, Correa LA, Wolff JC, et al. Effect of imiquimod in anogenital warts from HIV-positive men. *J Clin Virol.* 2007;39:210-214.
- Dede M, Kubar A, Yenen MC, et al. Human papillomavirus-type predict the clinical outcome of imiquimod therapy for women with vulvar condylomata acuminata. *Acta Obstet Gynecol Scand.* 2007;86:968-972.
- Van Seters M, Van Beurden M, Ten Kate FJW, et al. Treatment of vulvar intraepithelial neoplasia with topical imiquimod. *N Engl J Med.* 2008;358:1465-1473.
- Wieland U, Brockmeyer NH, Weissenborn SJ, et al. Imiquimod treatment of anal intraepithelial neoplasia in HIV-positive men. *Arch Dermatol.* 2006;142:1438-1444.
- Pelletier F, Drobacheff-Thiebaut C, Aubin F, et al. Effects of imiquimod on latent human papillomavirus anal infection in HIV-infected patients. *Ann Dermatol Venerol.* 2004;131:947-951.
- Arevalo I, Ward B, Miller R, et al. Successful treatment of drug-resistant cutaneous leishmaniasis in humans by use of imiquimod, an immunomodulator. *Clin Infect Dis.* 2001;33:1847-1851.
- Miranda-Verastegui C, Llanos-Cuentas A, Arevalo I, et al. Randomized, double-blind clinical trial of topical imiquimod 5% with parenteral meglumine antimoniate in the treatment of cutaneous leishmaniasis in Peru. *Clin Infect Dis.* 2005;40:1395-1403.
- Firooz A, Khamisipour A, Ghoorchi MH, et al. Imiquimod in combination with meglumine antimoniate for cutaneous leishmaniasis: a randomized assessor-blind controlled trial. *Arch Dermatol.* 2006;142:1575-1579.
- Martinez S, Molina JM, Scieux C, et al. Topical imiquimod for recurrent acyclovir-resistant HSV infection. *Am J Med.* 2006;119:e9-e11.
- Lascaux AS, Caumes E, Debacq C, et al. Successful treatment of acyclovir and foscarnet resistant herpes simplex virus lesions with topical imiquimod in patients infected with human immunodeficiency virus type 1. *J Med Virol.* 2012;84:194-197.
- Brummitt CF. Imiquimod 5% cream for the treatment of recurrent, acyclovir-resistant genital herpes. *Clin Infect Dis.* 2006;42:575.
- Schacker TW, Conant M, Thoming C, et al. Imiquimod 5-percent cream does not alter the natural history of recurrent herpes genitalis: a phase II, randomized, double-blind, placebo-controlled study. *Antimicrob Agents Chemother.* 2002;46:3243-3248.
- Bernstein DI, Spruance SL, Arora SS, et al. Evaluation of imiquimod 5% cream to modify the natural history of herpes labialis: a pilot study. *Clin Infect Dis.* 2005;41:808-814.
- Allam JP, Erdsach T, Wenghoefer M, et al. Successful treatment of extensive human papillomavirus-associated oral leukoplakia with imiquimod. *Br J Dermatol.* 2008;158:644-646.
- Ara M, Zaballos P, Sanchez M, et al. Giant and recurrent orf virus infection in a renal transplant recipient treated with imiquimod. *J Am Acad Dermatol.* 2008;58(2 suppl 1):S39.
- Babel N, Eibl N, Ulrich C, et al. Development of Kaposi's sarcoma under sirolimus-based immunosuppression and successful treatment with imiquimod. *Transpl Infect Dis.* 2008;10:59-62.
- Stashower ME. Resolution of tinea pedis with imiquimod cream 5% in a patient with nodular basal cell carcinoma. *Cutis.* 2006;78:66-69.
- Spruance SL, Tyring SK, Smith MH, et al. Application of a topical immune response modifier, resiquimod gel, to modify the recurrence rate of recurrent genital herpes: a pilot study. *J Infect Dis.* 2001;184:196-200.
- Fife KH, Meng TC, Ferris DG, et al. Effect of resiquimod 0.01% gel on lesion healing and viral shedding when applied to genital herpes lesions. *Antimicrob Agents Chemother.* 2008;52:477-482.
- Mark KE, Corey L, Meng TC, et al. Topical resiquimod 0.01% gel decreases herpes simplex virus type 2 genital

- shedding: a randomized, controlled trial. *J Infect Dis.* 2007;195:1324-1331.
82. Mark KE, Spruance S, Kinghorn GR, et al. Three phase III randomized controlled trials of topical resiquimod gel 0.01% to prevent genital herpes recurrences [abstract P-256]. Presented at the 17th Annual Meeting of the International Society on Sexual Transmitted Diseases Research. Seattle, WA, 2007.
 83. Campanelli A, Lubbe J. Erosive cheilitis after facial application of imiquimod 5% cream. *J Eur Acad Dermatol Venerol.* 2007;21:1429-1430.
 84. Chakrabarty AK, Mraz S, Geisse JK, et al. Aphthous ulcers associated with imiquimod and the treatment of actinic cheilitis. *J Am Acad Dermatol.* 2005;52(2 suppl):35.
 85. Jacobs AA, Snavely N, Markus J, et al. Vasodilatory adverse events associated with topical imiquimod 5 percent cream. *Dermatol Online J.* 2008;14:4.
 86. Wu JK, Siller G, Strutton G. Psoriasis induced by topical imiquimod. *Australas J Dermatol.* 2004;45:47-50.
 87. Systemic reactions to imiquimod. *Med Lett.* 2004;40:92.
 88. Taylor CL, Maslen M, Kapembwa M. A case of severe eczema following use of imiquimod 5% cream. *Sex Transm Infect.* 2006;82:227-228.
 89. Wolfe CM, Tafuri N, Hatfield K. Exacerbation of myasthenia gravis during imiquimod treatment. *J Drug Dermatol.* 2007;6:745-746.
 90. Benson E. Imiquimod: Potential risk of an immunostimulant. *Australas J Dermatol.* 2004;45:123-124.
 91. Myhre PE, Levy ML, Eichenfield LF, et al. Pharmacokinetics and safety of imiquimod 5% cream in the treatment of molluscum contagiosum in children. *Pediatr Dermatol.* 2008;25:88-95.
 92. Audisio T, Roca FC, Piatti C. Topical imiquimod therapy for external anogenital warts in pregnant women. *Int J Gynecol Obstet.* 2008;100:275-276.
 93. Einarson A, Costei A, Kalra S, et al. The use of topical 5% imiquimod during pregnancy: a case series. *Reprod Toxicol.* 2006;21:1-2.
 94. Pevear DC, Tull TM, Seipel ME, et al. Activity of pleconaril against enteroviruses. *Antimicrob Agents Chemother.* 1999;43:2109-2115.
 95. Kaiser L, Crump CE, Hayden FG. In vitro activity of pleconaril and AG7088 against selected serotypes and clinical isolates of human rhinoviruses. *Antiviral Res.* 2000;47:215-220.
 96. Schiff GM, Sherwood JR. Clinical activity of pleconaril in an experimentally induced coxsackievirus A21 respiratory infection. *J Infect Dis.* 2000;181:20-26.
 97. Florea NR, Maglio D, Nicolau DP. Pleconaril, a novel anti-picornaviral agent. *Pharmacotherapy.* 2003;23:339-348.
 98. Kearns GL, Bradley JS, Jacobs RE, et al. Single dose pharmacokinetics of pleconaril in neonates. Pediatric Pharmacology Research Unit Network. *Pediatr Infect Dis J.* 2000;19:833-839.
 99. Hayden FG, Herrington DT, Coats TL, et al. Efficacy and safety of oral pleconaril for treatment of picornavirus colds in adults: results of two double-blind, randomized, placebo-controlled trials. *Clin Infect Dis.* 2003;36:1523-1532.
 100. Pevear DC, Hayden FG, Demenczuk TM, et al. Relationship of pleconaril susceptibility and clinical outcomes in treatment of common colds caused by rhinoviruses. *Antimicrob Agents Chemother.* 2005;49:4492-4499.
 101. Rotbart HA, Webster AD. Treatment of potentially life-threatening enterovirus infections with pleconaril. *Clin Infect Dis.* 2001;32:228-235.
 102. Webster AD. Pleconaril—an advance in the treatment of enteroviral infection in immunocompromised patients. *J Clin Virol.* 2005;32:1-6.