



Article

# Stereoselective Synthesis of 24-Fluoro-25-Hydroxyvitamin D<sub>3</sub> Analogues and Their Stability to hCYP24A1-Dependent Catabolism

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**Abstract:** Two 24-fluoro-25-hydroxyvitamin D<sub>3</sub> analogues (**3,4**) were synthesized in a convergent manner. The introduction of a stereocenter to the vitamin D<sub>3</sub> side-chain C24 position was achieved via Sharpless dihydroxylation, and a deoxyfluorination reaction was utilized for the fluorination step. Comparison between (24*R*)- and (24*S*)-24-fluoro-25-hydroxyvitamin D<sub>3</sub> revealed that the C24-*R*-configuration isomer **4** was more resistant to CYP24A1-dependent metabolism than its 24*S*-isomer **3**. The new synthetic route of the CYP24A1 main metabolite (24*R*)-24,25-dihydroxyvitamin D<sub>3</sub> (**6**) and its 24*S*-isomer (**5**) was also studied using synthetic intermediates (**30,31**) in parallel.



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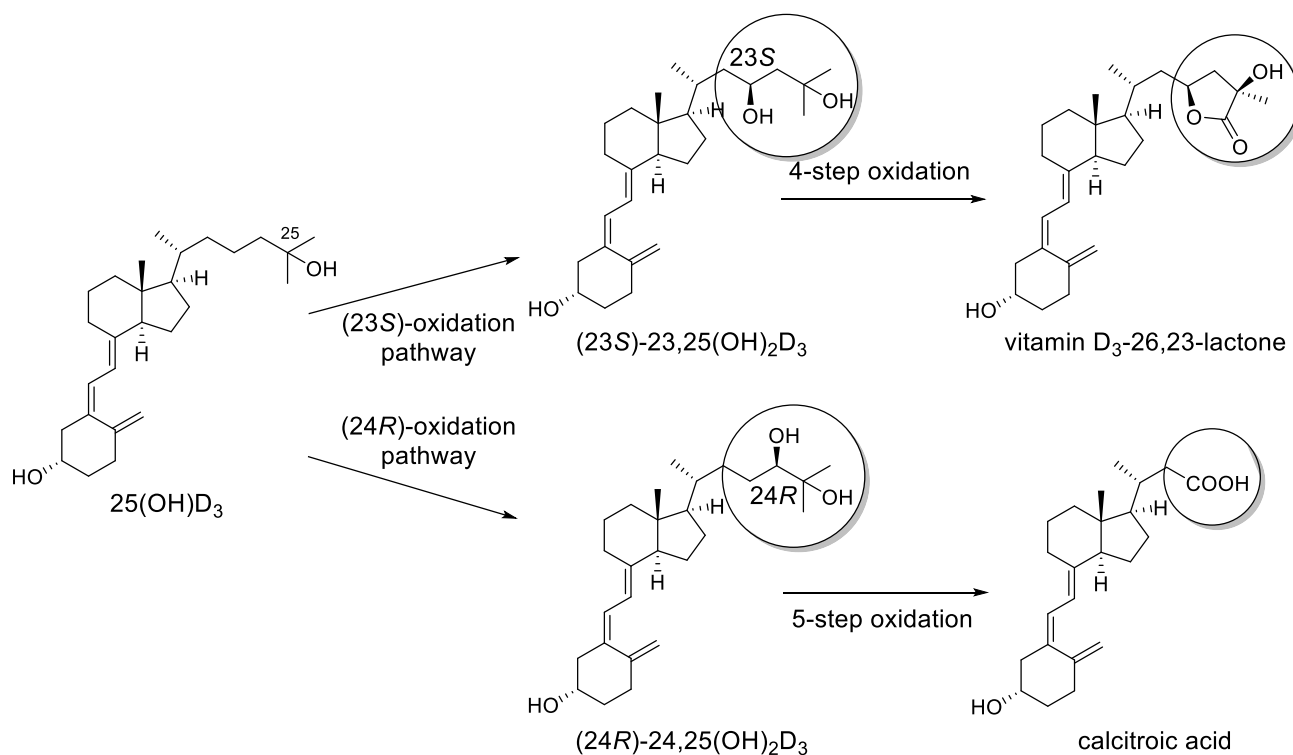
**Keywords:** human CYP24A1; synthesis; vitamin D<sub>3</sub> metabolite; 24-fluoro-25-hydroxyvitamin D<sub>3</sub> analogues; Sharpless dihydroxylation

## 1. Introduction

Vitamin D<sub>3</sub> is a lipophilic vitamin, and hydroxylation steps promoted by the cytochrome P450 family are essential for both activation and deactivation pathways. In the deactivation step, human cytochrome P450 24A1 (hCYP24A1) is one of the main enzymes catalyzing hydroxylation at the C23 or C24 positions of the 25-hydroxyvitamin D<sub>3</sub> [25(OH)D<sub>3</sub>] side-chain, and several subsequent hydroxylation steps lead to vitamin D<sub>3</sub>-23,26-lactone or calcitric acid (Scheme 1) [1–8].

Recently, we developed a new methodology to synthesize 23-fluorinated vitamin D<sub>3</sub> analogues (**1,2**), and identified their unique biological activities (Figure 1). The 23*S*-fluorinated isomer (**1**) showed higher metabolic resistance against hCYP24A1 than its 23*R*-isomer (**2**) [9,10]. On the other hand, the 23*R*-isomer (**2**) showed a greater binding affinity for human vitamin D receptor (hVDR) than its 23*S* isomer and natural 25(OH)D<sub>3</sub> (unpublished data). Encouraged by these results, we have been interested in 24-substituted vitamin D<sub>3</sub> analogues, 24-fluoro-25-hydroxyvitamin D<sub>3</sub> (**3,4**), to study elongation of the half-life time of 25(OH)D<sub>3</sub> against CYP24A1-dependent metabolism [11].

There have been several reports on the synthesis of 24-fluorinated vitamin D<sub>3</sub> analogues. For example, in 1979, a 24-fluorinated vitamin D<sub>3</sub> analogue was first reported by Ikekawa et al. [12,13]; they described 24-fluoro-25-hydroxyvitamin D<sub>3</sub> (**7**) as a C24 diastereomeric mixture (Figure 2). Later, Uskoković et al. synthesized (24*R*)-24-fluoro-1 $\alpha$ ,25-dihydroxyvitamin D<sub>3</sub> (**8**) from a steroid skeleton in 1985 and from a CD-ring fragment in 1988 [14,15]. However, selective synthesis of the 24*S*-fluorinated vitamin D<sub>3</sub> analogue has not been reported, and the route to synthetic modification at C24 is still limited. Considering the importance of the C24 position of vitamin D<sub>3</sub>—including its stereochemistry—the practical synthetic methodology for 24-fluorinated vitamin D<sub>3</sub> analogues is an essential topic.



Scheme 1. 25(OH)D<sub>3</sub> deactivation pathways catalyzed by human CYP24A1.

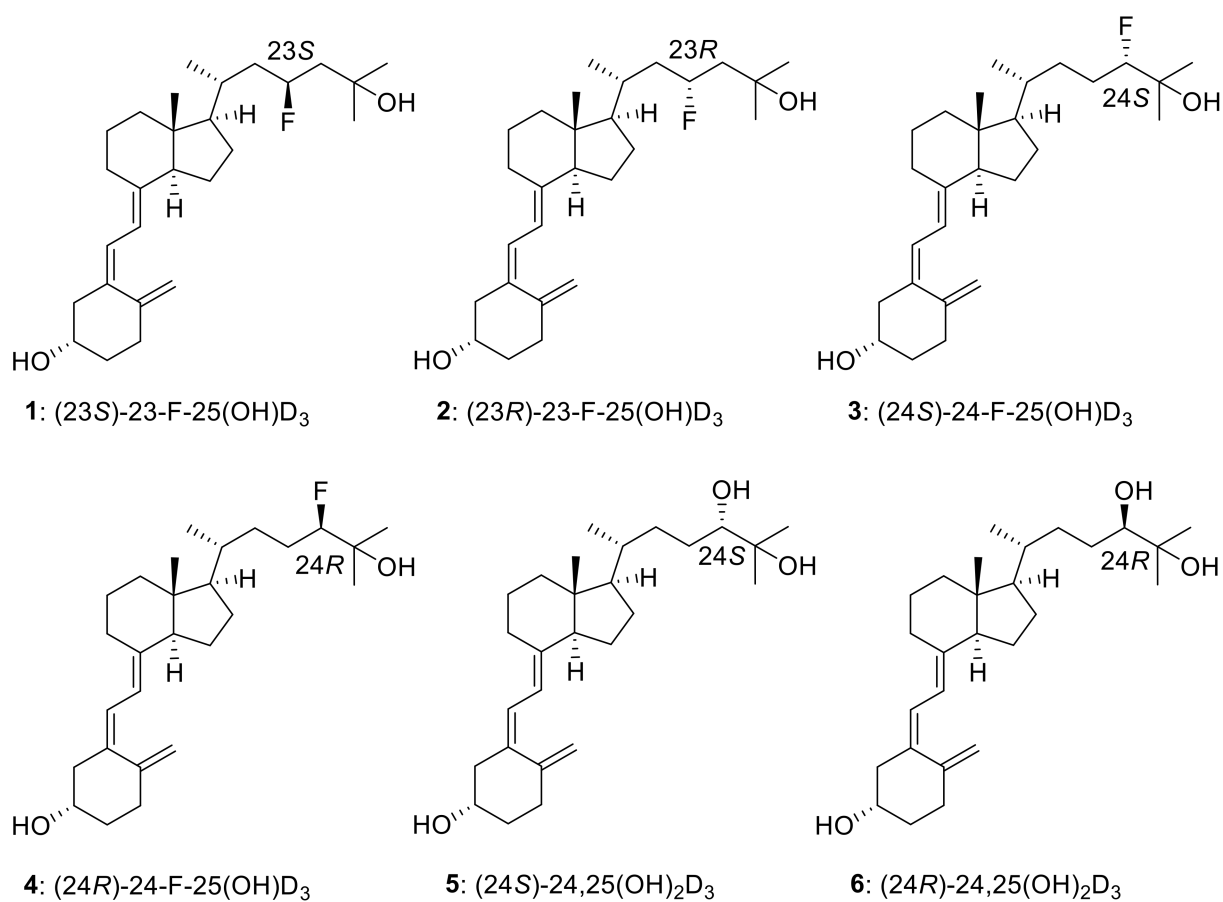
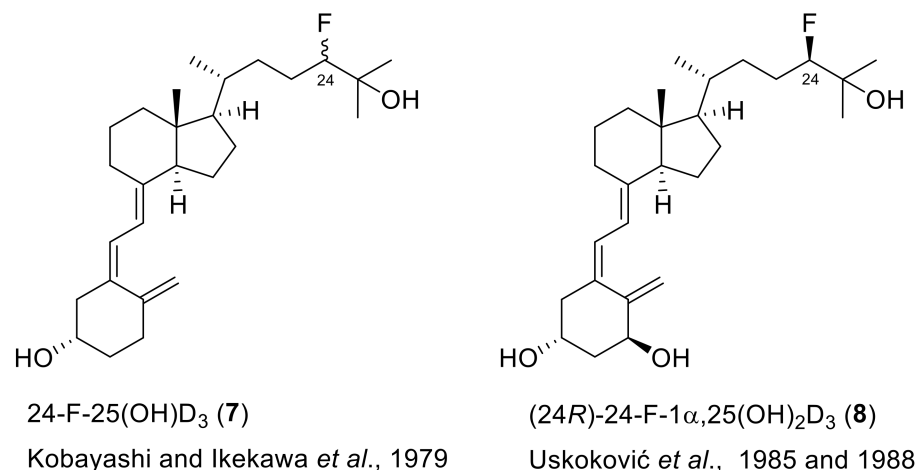
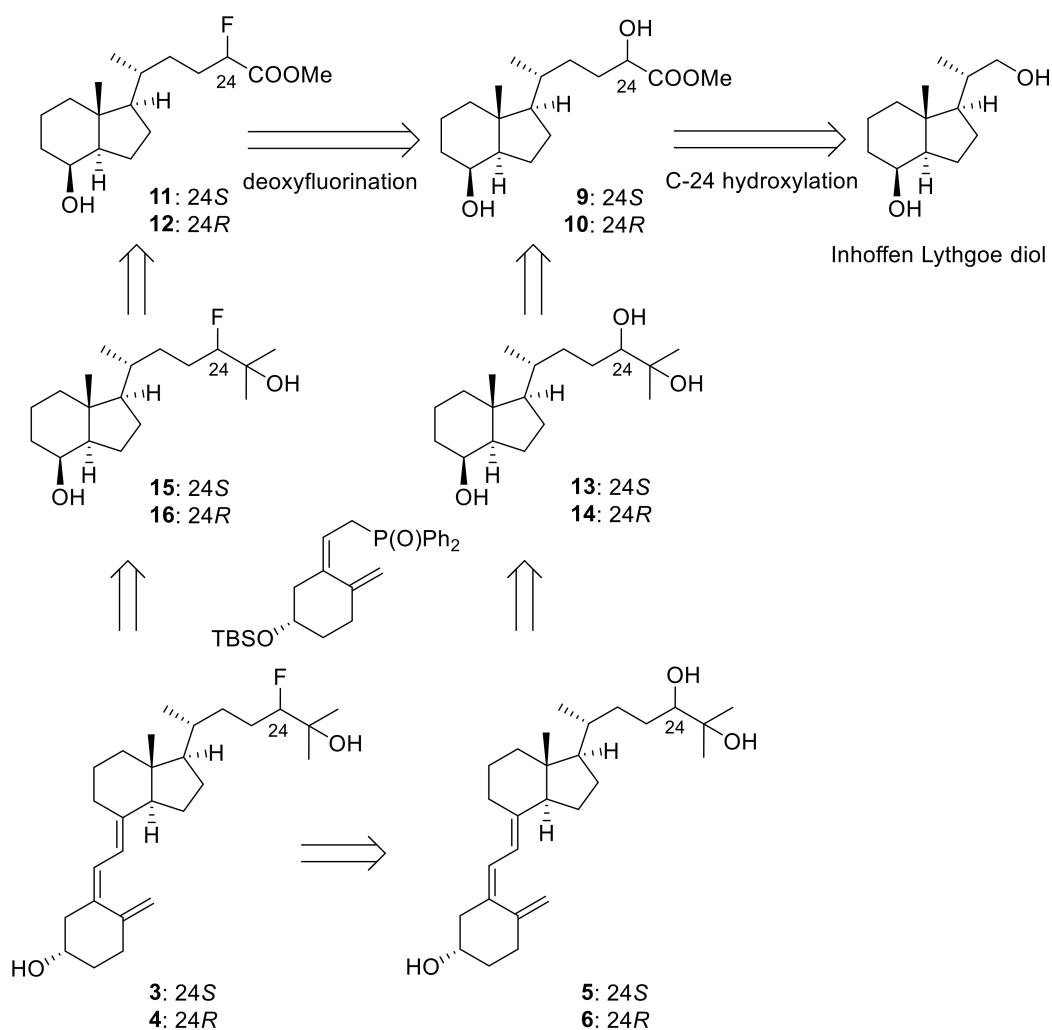


Figure 1. Structures of C23- and C24-substituted vitamin D<sub>3</sub> analogues.



**Figure 2.** Structures of C24-fluorinated vitamin D<sub>3</sub> analogues.

To solve the problems above, we herein report a new stereoselective synthetic methodology for 24-fluoro-25-hydroxyvitamin D<sub>3</sub> (**3,4**) through the chiral CD-ring part of 24,25-dihydroxyvitamin D<sub>3</sub> (**5,6**), and reveal their preliminary biological activities. We considered that 24-substituted CD-ring fragments (**13–16**) may be useful units to synthesize numerous 24-substituted vitamin D<sub>3</sub> analogues if coupled with various A-ring fragments [16] (Scheme 2).

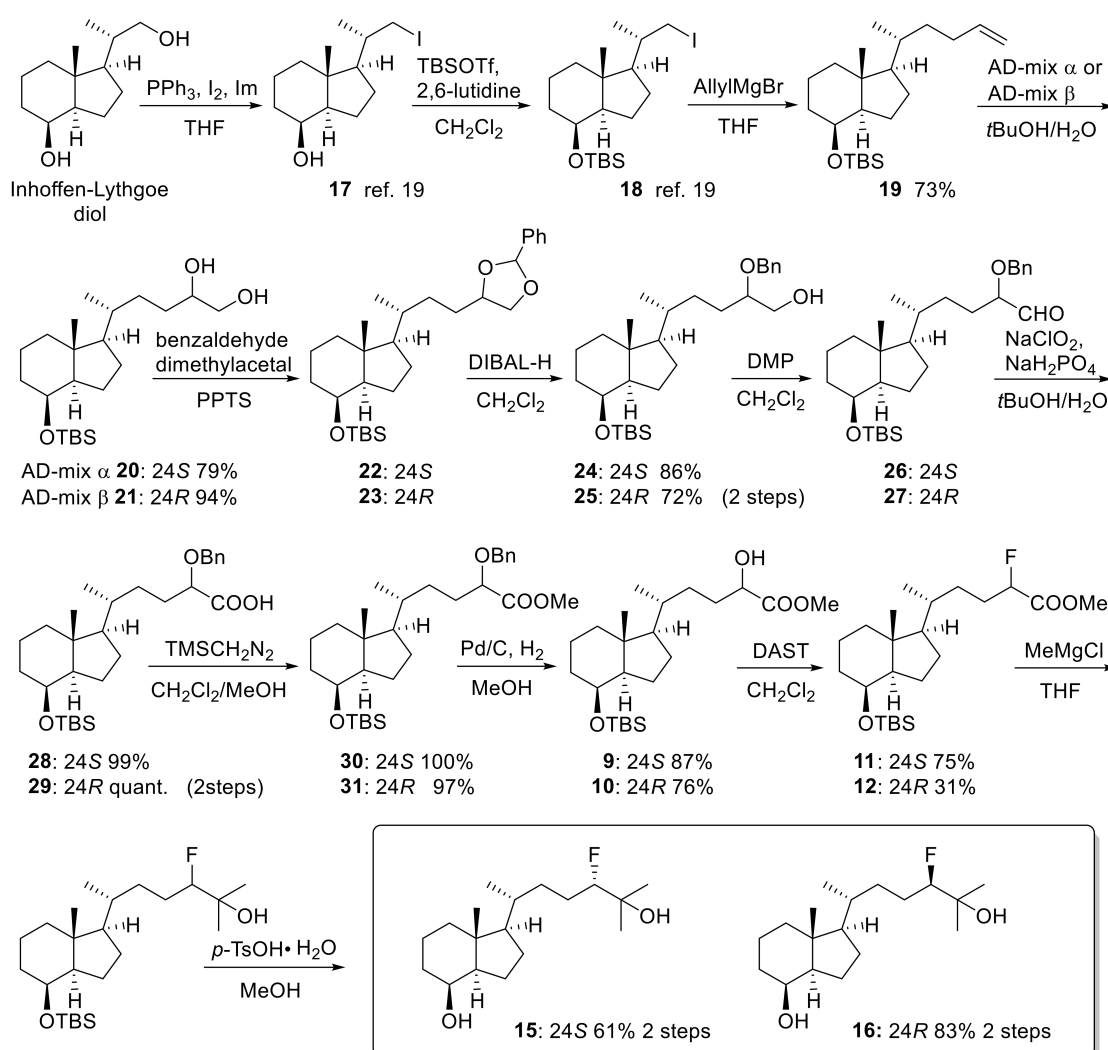


**Scheme 2.** Retrosynthetic analysis of C24-substituted vitamin D<sub>3</sub> analogues (**3–6**).

Synthesis of CD-ring fragments was achieved by side-chain elongation of Inhoffen–Lythgoe diol. Stereoselective introduction of the 24-hydroxy group was performed by Sharpless dihydroxylation reaction [17,18], and the fluorination step was achieved by deoxyfluorination reaction using *N,N*-diethylaminosulfur trifluoride (DAST).

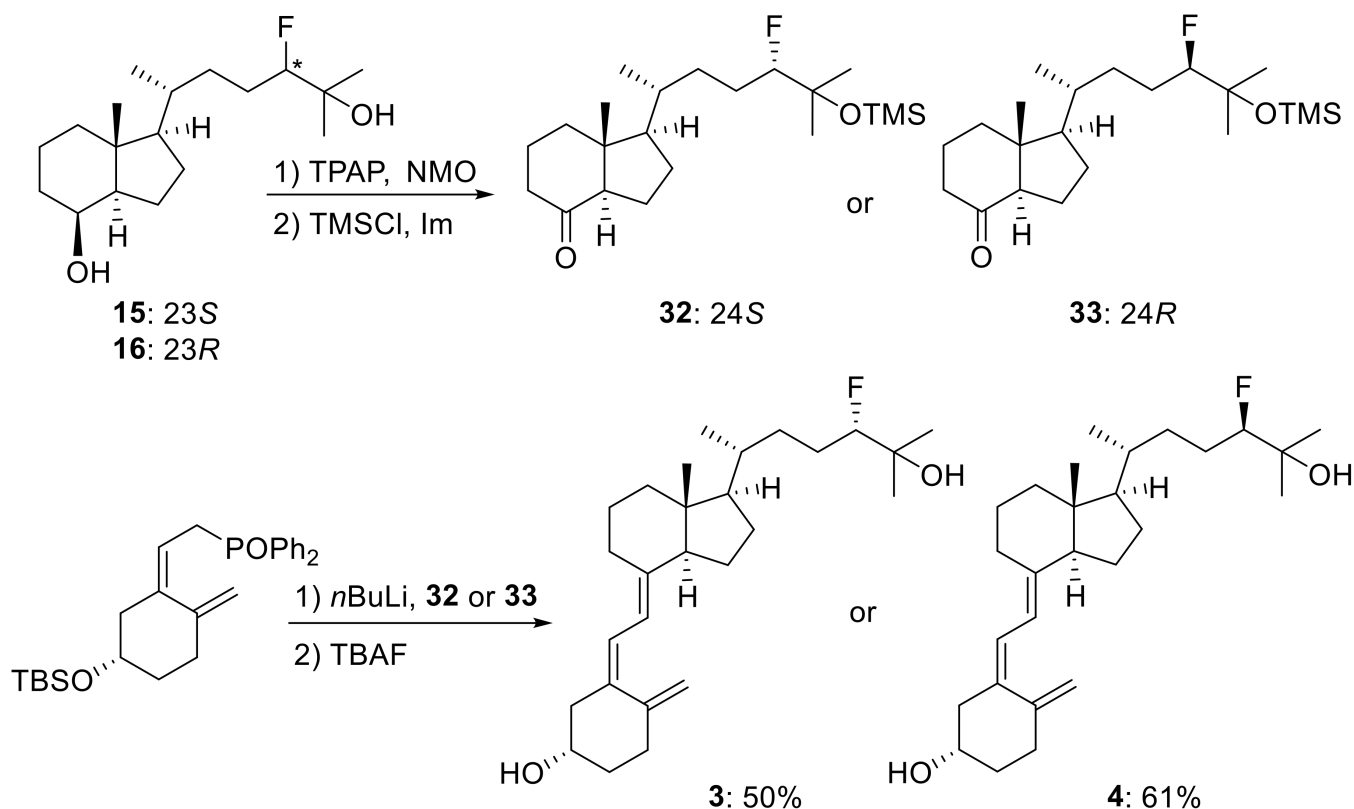
## 2. Results and Discussion

For the synthesis of C24-substituted CD-ring fragments (13–16), commercially available Inhoffen–Lythgoe diol was chosen as a starting material (Scheme 3). Iodination at C22-OH and hydroxy protection at the C8 position yielded iodide 18 [19]. After replacement of iodine with an allyl group utilizing allyl magnesium bromide, stereoselective dihydroxylation was achieved via Sharpless asymmetric dihydroxylation using AD-mix  $\alpha$  and  $\beta$  to yield diols with 24*S*-OH (20) and 24*R*-OH (21), respectively. Protection of the C24 position with benzyl ether and two-step oxidation afforded carboxylic acids (28,29). These were treated with trimethylsilyl diazomethane in methanol to produce methyl esters (30,31), which were subsequently hydrogenated to afford 24-hydroxylated methyl esters (9,10). Next, introduction of a fluorine atom was achieved via deoxyfluorination reaction using DAST. The addition of an excess of methyl magnesium chloride to the resulting fluoro methyl esters (11,12) in THF, followed by desilylation at the C8 position in the presence of *p*-toluenesulfonic acid, yielded 24-fluorinated CD-ring fragments (15,16).



**Scheme 3.** Stereoselective introduction of C24-hydroxy and -fluoro groups to the CD-ring side-chain using Sharpless asymmetric dihydroxylation and deoxyfluorination.

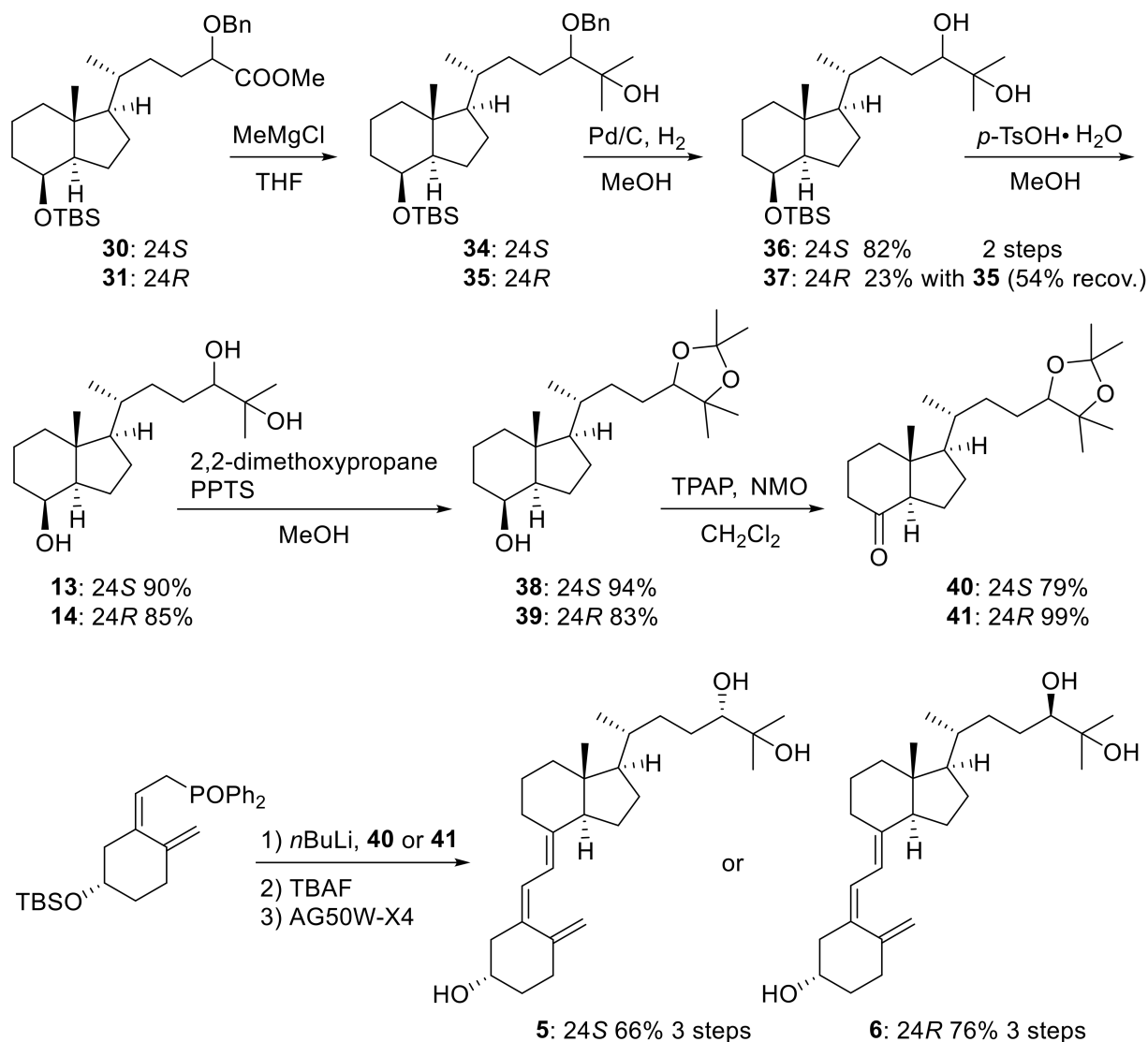
Oxidation of 24-fluorinated CD-ring fragments (**15,16**) with tetrapropylammonium perruthenate (TPAP) in the presence of 4-methylmorpholine *N*-oxide in methylene chloride, followed by protection of the C25-hydroxy group utilizing trimethylsilyl chloride (TMSCl), yielded 8-ketones (**32,33**) (Scheme 4). The Wittig–Horner coupling reaction with the lithium salt of the A-ring phosphine oxide [16] produced the coupling products. The final deprotection with tetrabutylammonium fluoride (TBAF) afforded the desired 24-fluoro-25-hydroxyvitamin D<sub>3</sub> (**3** and **4**) in 50 and 61% overall yields from **15** and **16**, respectively.



**Scheme 4.** Coupling reaction and desilylation steps for **3** and **4**.

There are several methods to synthesize 24-hydroxyvitamin D<sub>3</sub> analogues [20–27]. In this study, we also explored the possibility of using 24-*O*-benzyl methyl esters (**30,31**) to synthesize their important precursors (**13,14**). As shown in Scheme 5, the 24-*O*-benzyl methyl esters were subsequently reacted with methyl magnesium chloride to produce **34** and **35**. Deprotection of the benzyl group afforded **36** and **37**, respectively, and desilylation at the C8-OH with *p*-toluenesulfonic acid yielded 24-hydroxy CD-rings (**13,14**).

To construct triene structures, we took advantage of a method that Sarandeses et al. developed in 2002 [25]. 24,25-Diol protection of the 24-hydroxylated CD-ring fragments (**13,14**) as a ketal was performed with 2,2-dimethoxypropane in the presence of pyridinium *p*-toluenesulfonate (PPTS) as an acid catalyst, and subsequent oxidation with TPAP and NMO of C8-hydroxy groups afforded the desired 8-ketones (**40,41**). The coupling reaction between the CD-rings (**40,41**) and A-ring phosphine oxide [16] was performed via the Wittig–Horner reaction to yield the protected vitamin D<sub>3</sub>. Deprotection with TBAF followed by cationic exchange resin (AG 50W-X4, H<sup>+</sup> form) treatment afforded 24,25-dihydroxyvitamin D<sub>3</sub> (**5,6**).



**Scheme 5.** Alternative synthesis of 24,25(OH)<sub>2</sub>D<sub>3</sub> (**5,6**) via protected 24,25-dihydroxy CD-ring fragments (**40,41**).

### Biological Evaluation

The binding affinities of the three 24-fluorinated vitamin D<sub>3</sub> analogues—(24S)-24-F-25(OH)D<sub>3</sub> (**3**), (24R)-24-F-25(OH)D<sub>3</sub> (**4**), and 24,24-difluoro-25(OH)D<sub>3</sub> [28]—for hVDR are summarized in Table 1. For hVDR, **3** and **4** showed similar binding affinities, but slightly lower than that of natural 25(OH)D<sub>3</sub>. These results demonstrate that a fluorine atom at the C24 position could mildly impair the binding with hVDR. However, unexpectedly, 24,24-difluoro-25(OH)D<sub>3</sub> showed higher binding affinity for hVDR than those of the 24-fluorinated vitamin D<sub>3</sub> analogues **3** and **4**.

**Table 1.** Relative hVDR binding affinity of 24-fluorinated 25(OH)D<sub>3</sub>.

Compound	Relative hVDR Binding Affinity (%)
25(OH)D <sub>3</sub>	100
(24S)-24-F-25(OH)D <sub>3</sub> ( <b>3</b> )	64
(24R)-24-F-25(OH)D <sub>3</sub> ( <b>4</b> )	73
24,24-F <sub>2</sub> -25(OH)D <sub>3</sub> [28]	180

We next analyzed the metabolism of three analogues and 25(OH)D<sub>3</sub> by hCYP24A1. Hydroxylation activities of hCYP24A1 toward these analogues are shown in Table 2. The hCYP24A1 showed nearly the same activity toward (24S)-24-F-25(OH)D<sub>3</sub> as that toward 25(OH)D<sub>3</sub>, whereas 24,24-F<sub>2</sub>-25(OH)D<sub>3</sub> showed marked resistance to hCYP24A1-dependent metabolism. These results demonstrate that the 24R fluorine substitution allows 25(OH)D<sub>3</sub> to achieve stronger catabolic resistance than its 24S counterpart. In contrast, we demonstrated that (23S)-23-F-25(OH)D<sub>3</sub> (**1**) showed stronger resistance to CYP24A1 metabolism than (23R)-23-F-25(OH)D<sub>3</sub> (**2**), as described in our previous study [9]. These results can be explained by the direction of hydroxylation at the C23 and C24 positions by CYP24A1 [1–8].

**Table 2.** Hydroxylation activities of human CYP24A1 toward 25(OH)D<sub>3</sub> and its C24-fluorinated analogues.

Substrate	(nmol/min/nmol-P450)
25(OH)D <sub>3</sub>	5.0 ± 1.8
(24S)-24-F-25(OH)D <sub>3</sub> ( <b>3</b> )	4.8 ± 1.5
(24R)-24-F-25(OH)D <sub>3</sub> ( <b>4</b> )	1.6 ± 0.5
24,24-F <sub>2</sub> -25(OH)D <sub>3</sub> [28]	0.53 ± 0.12

Data were obtained at a substrate concentration of 5 μM. Each value is the mean ± SD of three separate experiments.

### 3. Experimental Section

<sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded on JEOL AL-400 NMR (400 MHz) and ECP-600 NMR (600 MHz) spectrometers (Tokyo, Japan). <sup>1</sup>H NMR spectra were referenced with (CH<sub>3</sub>)<sub>4</sub>Si (δ 0.00 ppm) or CHCl<sub>3</sub> (δ 7.26 ppm) as internal standards. <sup>13</sup>C NMR spectra were referenced with deuterated solvent (δ 77.0 ppm for CDCl<sub>3</sub>). IR spectra were recorded on a JASCO FT-IR-800 Fourier-transform infrared spectrophotometer (Tokyo, Japan). High-resolution mass spectra were obtained on a SHIMADZU LCMS-IT-TOF mass spectrometer (Kyoto, Japan) with an electrospray ionization (ESI) method or atmospheric-pressure chemical ionization (APCI). Optical rotations were measured on a JASCO DIP-370 digital polarimeter (Tokyo, Japan). Column chromatography was performed on silica gel 60N (40–50 μm, Kanto Chemical Co., Inc., Tokyo, Japan) or silica gel 60 (0.040–0.063 mm, Merck, Tokyo, Japan). All experiments were performed under anhydrous conditions in an atmosphere of argon, unless otherwise stated. The supporting information of <sup>1</sup>H and <sup>13</sup>C NMR spectra of all new compounds: **19–21**, **24**, **25**, **28–31**, **9–12**, **15**, **16**, **3**, **4**, **36**, and **37** is available at the link in Supplementary Materials.

#### 3.1. *tert*-Butyl(1*R*,3*aR*,4*S*,7*aR*)-1-[(*R*)-hex-5-en-2-yl]-7*a*-methyloctahydro-1*H*-inden-4-yl)oxy) Dimethylsilane (**19**)

To a solution of compound **18** [19] (180.0 mg, 0.412 mmol) in THF (4 mL), allyl magnesium bromide (3.3 mL, 1 M in Et<sub>2</sub>O, 3.3 mmol) was added at 0 °C, and it was stirred at room temperature for 23 h. After the reaction was quenched with water and aqueous saturated NH<sub>4</sub>Cl, the mixture was extracted with EtOAc three times, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane only) to obtain **19** (105.1 mg, 73%) as a colorless oil.

**19**: [α]<sub>D</sub><sup>27</sup> +52.7 (c 1.82, CHCl<sub>3</sub>); IR (neat) 1471, 1371, 1252, 1162, 1085, 1027, 837, 771 cm<sup>-1</sup>; <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ −0.01 (s, 3H), 0.01 (s, 3H), 0.89 (s, 9H), 0.90 (d, *J* = 6.6 Hz, 3H), 0.91 (s, 3H), 0.99–1.13 (m, 3H), 1.21–1.28 (m, 2H), 1.30–1.43 (m, 4H), 1.46–1.58 (m, 2H), 1.63–1.70 (m, 1H), 1.74–1.84 (m, 2H), 1.90–1.97 (m, 2H), 2.08–2.14 (m, 1H), 3.99–4.00 (m, 1H), 4.90–4.92 (m, 1H), 4.97–5.04 (m, 1H), 5.80 (tdd, *J* = 6.0, 10.2, 16.2 Hz, 1H); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>) δ −5.2, −4.8, 13.7, 17.7, 18.0, 18.5, 23.1, 25.8, 27.3, 30.5, 34.5, 34.9, 35.1, 40.7, 42.2, 53.1, 56.8, 69.5, 113.8, 139.7; HRMS (ESI<sup>+</sup>) calcd for C<sub>22</sub>H<sub>42</sub>OSi [M]<sup>+</sup> 350.2999, found 350.2992.

3.2. (2*S*,5*R*)-5-((1*R*,3*aR*,4*S*,7*aR*)-4-[(*tert*-Butyldimethylsilyl)oxy]-7*a*-methyloctahydro-1*H*-inden-1-yl)hexane-1,2-diol (**20**)

A mixture of AD-mix  $\alpha$  (4.01 g) in *t*BuOH (10 mL) and H<sub>2</sub>O (10 mL) was stirred at 0 °C for 25 min; **19** (303.5 mg, 0.255 mmol) was added to the mixture at 0 °C, and it was stirred at the same temperature for 5 h, and then at room temperature for 15 h under air. After the reaction was quenched with water, the mixture was extracted with EtOAc three times, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 1:1) to obtain **20** (253.9 mg, 79%) as a colorless oil.

**20**: [ $\alpha$ ]<sub>D</sub><sup>27</sup> +44.4 (c 1.55, CHCl<sub>3</sub>); IR (neat) 3402, 1645, 1469, 1374, 1265, 1160, 1066, 1032, 840, 776, 743 cm<sup>-1</sup>; <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  -0.01 (s, 3H), 0.00 (s, 3H), 0.88–0.90 (m, 15H), 0.98–1.12 (m, 3H), 1.20–1.43 (m, 7H), 1.47–1.58 (m, 3H), 1.64–1.67 (m, 1H), 1.75–1.83 (m, 2H), 1.92–1.95 (m, 1H), 2.27 (s, 3H), 3.41–3.44 (m, 1H), 3.62–3.67 (m, 2H), 3.98–3.99 (m, 1H); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>)  $\delta$  -5.2, -4.8, 13.7, 17.6, 18.0, 18.6, 23.0, 25.8, 27.3, 29.7, 31.5, 34.4, 35.3, 40.7, 42.1, 53.0, 56.5, 66.7, 69.4, 73.0; HRMS (APCI<sup>-</sup>) calcd for C<sub>22</sub>H<sub>44</sub>O<sub>3</sub>SiCl [M+Cl]<sup>-</sup> 419.2754, found 419.2764.

3.3. (2*R*,5*R*)-5-((1*R*,3*aR*,4*S*,7*aR*)-4-[(*tert*-Butyldimethylsilyl)oxy]-7*a*-methyloctahydro-1*H*-inden-1-yl)hexane-1,2-diol (**21**)

A mixture of AD-mix  $\beta$  (4.62 g) in *t*BuOH (15 mL) and H<sub>2</sub>O (15 mL) was stirred at 0 °C for 25 min; **19** (418.4 mg, 0.255 mmol) was added to the mixture at 0 °C, and it was stirred at the same temperature for 1 h 35 min under air. After the reaction was quenched with water, the mixture was extracted with EtOAc three times, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 1:1) to obtain **21** (433.1 mg, 94%) as a colorless oil.

**21**: [ $\alpha$ ]<sub>D</sub><sup>27</sup> +41.9 (c 2.05, CHCl<sub>3</sub>); IR (neat) 3294, 1223, 1076, 1026, 837, 764 cm<sup>-1</sup>; <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  -0.01 (s, 3H), 0.00 (s, 3H), 0.88–0.90 (m, 15H), 1.01–1.58 (m, 11H), 1.65–1.67 (m, 1H), 1.75–1.84 (m, 2H), 1.91–1.95 (m, 4H), 3.42–3.45 (m, 1H), 3.64–3.69 (m, 2H), 3.99 (dd, *J* = 2.4, 5.4 Hz, 1H); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>)  $\delta$  -5.2, -4.8, 13.7, 17.7, 18.0, 18.5, 23.0, 25.8, 27.3, 29.6, 31.4, 34.4, 35.1, 40.7, 42.1, 53.0, 56.5, 67.0, 69.4, 72.7; HRMS (ESI<sup>-</sup>) calcd for C<sub>22</sub>H<sub>44</sub>O<sub>3</sub>SiCl [M+Cl]<sup>-</sup> 419.2754, found 419.2773.

3.4. (2*S*,5*R*)-2-(Benzylloxy)-5-((1*R*,3*aR*,4*S*,7*aR*)-4-[(*tert*-butyldimethylsilyl)oxy]-7*a*-methyloctahydro-1*H*-inden-1-yl)hexan-1-ol (**24**)

Benzaldehyde dimethyl acetal (374.4 mg, 369  $\mu$ L, 2.46 mmol) and pyridinium *p*-toluenesulfonate (PPTS) (158.6 mg, 0.63 mmol) were added to a solution of **20** (472.0 mg, 1.23 mmol) in toluene (15 mL) at room temperature, and the mixture was stirred at the same temperature for 2 h. After the reaction was quenched with water and saturated aqueous NaHCO<sub>3</sub>, the mixture was extracted with EtOAc three times, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 4:1) to obtain the crude acetal **22**, which was used for the next reaction without further purification. To a solution of the above crude acetal **22** in CH<sub>2</sub>Cl<sub>2</sub> (15 mL), we added DIBAL-H (4.8 mL, 1.03 M in hexane solution, 4.92 mmol) at 0 °C, and the mixture was stirred at the same temperature for 20 min. After the reaction was quenched with MeOH at 0 °C, H<sub>2</sub>O and saturated aqueous potassium sodium tartrate were added at room temperature. The mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub> four times, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 4:1) to obtain **24** (501.1 mg, 86%) as a colorless oil.

**24**: [ $\alpha$ ]<sub>D</sub><sup>27</sup> +49.0 (c 3.62, CHCl<sub>3</sub>); IR (neat) 3420, 1465, 1453, 1374, 1254, 1085, 1028, 840, 776, 739 cm<sup>-1</sup>; <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  0.00 (s, 3H), 0.01 (s, 3H), 0.90–0.91 (m, 15H), 0.99–1.13 (m, 3H), 1.21–1.60 (m, 10H), 1.66–1.68 (m, 1H), 1.75–1.84 (m, 2H), 1.94–1.96 (m, 2H), 3.45–3.48 (m, 1H), 3.51–3.54 (m, 1H), 3.69 (dd, *J* = 3.0, 11.4 Hz, 1H), 4.00–4.00 (m, 1H), 4.53 (d, *J* = 11.7 Hz, 1H), 4.63 (d, *J* = 11.7 Hz, 1H), 7.28–7.32 (m, 1H), 7.35–7.36 (m, 4H); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>)  $\delta$  -5.2, -4.8, 13.7, 17.6, 18.0, 18.5, 23.0, 25.8,



27.1, 27.3, 31.2, 34.4, 35.3, 40.7, 42.1, 53.0, 56.5, 64.2, 69.4, 71.4, 80.4, 127.7, 127.8, 128.4, 138.5; HRMS (ESI<sup>+</sup>) calcd for C<sub>29</sub>H<sub>50</sub>NaO<sub>3</sub>Si [M + Na]<sup>+</sup> 497.3421, found 497.3433.

3.5. (2*R*,5*R*)-2-(Benzyloxy)-5-((1*R*,3*aR*,4*S*,7*aR*)-4-[(*tert*-butyldimethylsilyl)oxy]-7*a*-methyloctahydro-1*H*-inden-1-yl)hexan-1-ol (**25**)

Benzaldehyde dimethyl acetal (39.3 mg, 39  $\mu$ L, 0.258 mmol) and pyridinium *p*-toluenesulfonate (PPTS) (4.5 mg, 0.018 mmol) were added to a solution of **21** (49.6 mg, 0.129 mmol) in toluene (0.7 mL) at room temperature, and the mixture was stirred at the same temperature for 2 h. After the reaction was quenched with water and saturated aqueous NaHCO<sub>3</sub>, the mixture was extracted with EtOAc three times, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 5:1) to obtain the crude acetal **23**, which was used for the next reaction without further purification. To a mixture of the above crude acetal **23** in CH<sub>2</sub>Cl<sub>2</sub> (2 mL), we added DIBAL-H (313  $\mu$ L, 1.03 M in hexane solution, 0.322 mmol) at  $-40$  °C, and the mixture was stirred at the same temperature for 1 h, and then at room temperature for 1 h. After the reaction was quenched with MeOH, H<sub>2</sub>O and saturated aqueous potassium sodium tartrate were added at room temperature. The mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub> three times, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 4:1) to obtain **25** (44.4 mg, 72%) as a colorless oil.

**25**: [ $\alpha$ ]<sub>D</sub><sup>27</sup> +31.1 (c 0.91, CHCl<sub>3</sub>); IR (neat) 3332, 1462, 1369, 1257, 1076, 1030, 837, 771 cm<sup>-1</sup>; <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  0.00 (s, 3H), 0.01 (s, 3H), 0.89–0.90 (m, 15H), 1.00–1.14 (m, 3H), 1.20–1.26 (m, 2H), 1.30–1.45 (m, 6H), 1.51–1.58 (m, 2H), 1.65–1.71 (m, 2H), 1.75–1.84 (m, 2H), 1.93–1.95 (m, 1H), 3.46–3.50 (m, 1H), 3.52–3.55 (m, 1H), 3.68 (dd, *J* = 3.0, 12.0 Hz, 1H), 3.99–4.00 (m, 1H), 4.54 (d, *J* = 10.8 Hz, 1H), 4.62 (d, *J* = 10.8 Hz, 1H), 7.28–7.32 (m, 1H), 7.35–7.36 (m, 4H); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>)  $\delta$   $-5.2$ ,  $-4.8$ , 13.7, 17.7, 18.0, 18.6, 23.0, 25.8, 27.2, 27.3, 31.1, 34.4, 35.3, 40.7, 42.1, 53.0, 56.4, 64.4, 69.4, 71.6, 80.3, 127.7, 127.8, 128.5, 138.5; HRMS (APCI<sup>+</sup>) calcd for C<sub>29</sub>H<sub>50</sub>NaO<sub>3</sub>Si [M + Na]<sup>+</sup> 497.3421, found 497.3450.

3.6. (2*S*,5*R*)-2-(Benzyloxy)-5-((1*R*,3*aR*,4*S*,7*aR*)-4-[(*tert*-butyldimethylsilyl)oxy]-7*a*-methyloctahydro-1*H*-inden-1-yl)hexanoic acid (**28**)

Dess–Martin periodinane (1.42 g, 3.35 mmol) was added to a mixture of **24** (490.2 mg, 1.03 mmol) and 4Å molecular sieves (321.9 mg) in CH<sub>2</sub>Cl<sub>2</sub> (10 mL) at 0 °C, and the mixture was stirred at the same temperature for 2 h. After the reaction was quenched with water and saturated aqueous NaHCO<sub>3</sub>, the mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub> three times, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 10:1) to obtain the crude aldehyde **26**, which was used for the next reaction without further purification. To a mixture of the above crude aldehyde **26** and NaH<sub>2</sub>PO<sub>4</sub> (1.216 g, 8.11 mmol) in H<sub>2</sub>O (9 mL) and *t*-BuOH (18 mL), NaClO<sub>2</sub> (575.9 mg, 6.37 mmol) was added at 0 °C under air and stirred at the same temperature for 30 min. After the reaction was quenched with aqueous saturated NH<sub>4</sub>Cl and aqueous saturated sodium thiosulfate, the mixture was extracted with EtOAc three times, washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 1:1) to obtain **28** (960.6 mg, 99%) as a colorless oil.

**28**: [ $\alpha$ ]<sub>D</sub><sup>27</sup> +21.7 (c 1.32, CHCl<sub>3</sub>); IR (neat) 1720, 1469, 1254, 1089, 1032, 840, 780 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$   $-0.01$  (s, 3H), 0.01 (s, 3H), 0.89–0.90 (m, 15H), 0.99–1.94 (m, 17H), 3.94–3.99 (m, 2H), 4.50 (d, *J* = 11.6 Hz, 1H), 4.70 (d, *J* = 11.6 Hz, 1H), 7.29–7.39 (m, 5H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$   $-5.2$ ,  $-4.8$ , 13.7, 17.6, 18.0, 18.5, 23.0, 25.8, 27.2, 29.1, 31.1, 34.4, 35.0, 40.7, 42.1, 53.0, 56.4, 69.4, 72.5, 78.3, 128.1, 128.1, 128.5, 137.0, 176.7; HRMS (ESI<sup>-</sup>) calcd for C<sub>29</sub>H<sub>47</sub>O<sub>4</sub>Si [M-H]<sup>-</sup> 487.3249, found 487.3278.

3.7. (2*R*,5*R*)-2-(Benzyloxy)-5-((1*R*,3*aR*,4*S*,7*aR*)-4-[(*tert*-butyldimethylsilyl)oxy]-7*a*-methyloctahydro-1*H*-inden-1-yl)hexanoic acid (**29**)

Dess–Martin periodinane (2.76 g, 6.51 mmol) was added to a mixture of **25** (1.03 g, 2.17 mmol) and 4Å molecular sieves (600.0 mg) in CH<sub>2</sub>Cl<sub>2</sub> (10 mL) at 0 °C, and the mixture was stirred at the same temperature for 2 h. After the reaction was quenched with water and saturated aqueous NaHCO<sub>3</sub>, the mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub> three times, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The crude residue **27** was used for the next reaction without further purification. To a mixture of the above crude aldehyde **27** in H<sub>2</sub>O (3 mL) and *t*-BuOH (6 mL), NaH<sub>2</sub>PO<sub>4</sub> (134.8 mg, 0.898 mmol) and NaClO<sub>2</sub> (24.6 mg, 0.272 mmol) were added at 0 °C under air and stirred at the same temperature for 30 min. After the reaction was quenched with aqueous saturated NH<sub>4</sub>Cl and aqueous saturated sodium thiosulfate, the mixture was extracted with EtOAc three times, washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 1:1) to obtain **29** (125.3 mg, quantitative yield) as a colorless oil.

**29**: [ $\alpha$ ]<sub>D</sub><sup>27</sup> +47.1 (c 1.88, CHCl<sub>3</sub>); IR (neat) 1720, 1469, 1250, 1085, 1028, 840, 776 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  0.00 (s, 3H), 0.01 (s, 3H), 0.88–0.89 (m, 15H), 0.99–1.95 (m, 17H), 3.97–4.00 (m, 2H), 4.49 (d, *J* = 11.9 Hz, 1H), 4.71 (d, *J* = 11.9 Hz, 1H), 7.29–7.37 (m, 5H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  -5.2, -4.8, 13.7, 17.6, 18.0, 18.4, 23.0, 25.8, 27.2, 28.9, 30.6, 34.4, 34.7, 40.7, 42.1, 53.0, 56.3, 69.4, 72.6, 77.8, 128.1, 128.2, 128.5, 136.9, 176.2; HRMS (ESI<sup>-</sup>) calcd for C<sub>29</sub>H<sub>47</sub>O<sub>4</sub>Si [M-H]<sup>-</sup> 487.3249, found 487.3269.

3.8. Methyl (2*S*,5*R*)-2-(Benzyloxy)-5-((1*R*,3*aR*,4*S*,7*aR*)-4-[(*tert*-butyldimethylsilyl)oxy]-7*a*-methyloctahydro-1*H*-inden-1-yl)hexanoate (**30**)

Trimethylsilyl diazomethane (1.1 mL, 2.0 M in diethyl ether, 2.16 mmol) was added to a solution of **28** (490.2 mg, 1.03 mmol) in MeOH (2 mL) and CH<sub>2</sub>Cl<sub>2</sub> (6 mL) at 0 °C, and the mixture was stirred at the same temperature for 17 min. After the reaction was quenched with acetic acid and saturated aqueous NaHCO<sub>3</sub>, the mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub> three times, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 4:1) to obtain **30** (387 mg, 100%) as a colorless oil.

**30**: [ $\alpha$ ]<sub>D</sub><sup>27</sup> +17.0 (c 2.70, CHCl<sub>3</sub>); IR (neat) 1750, 1465, 1254, 1028, 840, 772 cm<sup>-1</sup>; <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  0.01 (s, 3H), 0.01 (s, 3H), 0.88–0.89 (m, 15H), 1.01–1.10 (m, 3H), 1.19–1.26 (m, 2H), 1.30–1.42 (m, 2H), 1.50–1.67 (m, 4H), 1.73–1.85 (m, 3H), 1.91–1.94 (m, 1H), 3.75 (s, 3H), 3.89 (dd, *J* = 5.4, 7.8 Hz, 1H), 3.99–3.99 (m, 1H), 4.41 (d, *J* = 11.4 Hz, 1H), 4.68 (d, *J* = 11.4 Hz, 1H), 7.27–7.31 (m, 1H), 7.33–7.36 (m, 4H); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>)  $\delta$  -5.2, -4.8, 13.7, 17.6, 18.0, 18.5, 23.0, 25.8, 27.1, 29.6, 31.2, 34.4, 35.0, 40.7, 42.1, 51.8, 53.0, 56.4, 69.4, 72.2, 78.9, 127.8, 127.9, 128.3, 137.6, 173.4; HRMS (ESI<sup>+</sup>) calcd for C<sub>30</sub>H<sub>49</sub>O<sub>4</sub>SiNa [M + Na]<sup>+</sup> 525.3371, found 525.3389.

3.9. Methyl (2*R*,5*R*)-2-(Benzyloxy)-5-((1*R*,3*aR*,4*S*,7*aR*)-4-[(*tert*-butyldimethylsilyl)oxy]-7*a*-methyloctahydro-1*H*-inden-1-yl)hexanoate (**31**)

Trimethylsilyl diazomethane (362  $\mu$ L, 2.0 M in diethyl ether, 0.73 mmol) was added to a solution of **29** (125.3 mg, 1.03 mmol) in MeOH (1.5 mL) and CH<sub>2</sub>Cl<sub>2</sub> (4.5 mL) at 0 °C, and the mixture was stirred at the same temperature for 20 min. After the reaction was quenched with acetic acid and saturated aqueous NaHCO<sub>3</sub>, the mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub> three times, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 4:1) to obtain **31** (125.1 mg, 97%) as a colorless oil.

**31**: [ $\alpha$ ]<sub>D</sub><sup>27</sup> +57.5 (c 1.71, CHCl<sub>3</sub>); IR (neat) 1750, 1471, 1253, 1029, 838, 774 cm<sup>-1</sup>; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  0.00 (s, 3H), 0.01 (s, 3H), 0.86–0.89 (m, 15H), 1.90–1.95 (m, 1H), 3.75 (s, 3H), 3.90 (dd, *J* = 4.6, 8.2 Hz, 1H), 3.98–3.99 (m, 1H), 4.40 (d, *J* = 12.0 Hz, 1H),

4.69 (d,  $J = 12.0$  Hz, 1H), 7.27–7.36 (m, 5H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  –5.2, –4.8, 13.7, 17.6, 18.0, 18.4, 23.0, 25.8, 27.1, 29.5, 31.0, 34.4, 34.7, 40.7, 42.1, 51.8, 53.0, 56.4, 69.4, 72.3, 78.3, 127.8, 128.0, 128.3, 137.6, 173.6; HRMS (ESI<sup>+</sup>) calcd for  $\text{C}_{30}\text{H}_{50}\text{O}_4\text{SiNa}$  [ $\text{M} + \text{Na}$ ]<sup>+</sup> 525.3371, found 525.3399.

**3.10. Methyl (2S,5R)-5-((1R,3aR,4S,7aR)-4-[(tert-butyl)dimethylsilyloxy]-7a-methyloctahydro-1H-inden-1-yl)-2-hydroxyhexanoate (9)**

To a solution of **30** (109.0 mg, 0.22 mmol) in MeOH (10 mL) and EtOAc (2 mL), we added 10% Pd/C catalyst (22.6 mg). The mixture was stirred for 45 h at room temperature, and then for 68 h at 50 °C, under a hydrogen atmosphere. The reaction mixture was diluted with AcOEt, filtered through a Celite pad, and concentrated under reduced pressure. Purification via flash column chromatography on silica gel (hexane:EtOAc = 3:1) yielded **9** (78.0 mg, 87%) as a colorless oil.

**9**:  $[\alpha]_{\text{D}}^{27} +44.8$  (c 1.67,  $\text{CHCl}_3$ ); IR (neat) 3488, 1742, 1461, 1370, 1257, 1081, 1020, 840, 776, 686  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (600 MHz,  $\text{CDCl}_3$ )  $\delta$  –0.01 (s, 3H), 0.00 (s, 3H), 0.88–0.90 (m, 15H), 0.98–1.57 (m, 13H), 1.64–1.70 (m, 1H), 1.75–1.88 (m, 3H), 1.92–1.95 (m, 1H), 2.14 (brs, 1H), 3.78 (s, 3H), 3.99–4.00 (m, 1H), 4.15 (dd,  $J = 3.9, 6.9$  Hz, 1H);  $^{13}\text{C}$  NMR (150 MHz,  $\text{CDCl}_3$ )  $\delta$  –5.2, –4.8, 13.7, 17.6, 18.0, 18.6, 23.0, 25.8, 27.2, 30.6, 31.1, 34.4, 35.0, 40.7, 42.1, 52.4, 53.0, 56.4, 69.4, 71.0, 175.9; HRMS (ESI<sup>+</sup>) calcd for  $\text{C}_{23}\text{H}_{44}\text{O}_4\text{SiNa}$  [ $\text{M} + \text{Na}$ ]<sup>+</sup> 435.2901, found 435.2897.

**3.11. Methyl (2R,5R)-5-((1R,3aR,4S,7aR)-4-[(tert-butyl)dimethylsilyloxy]-7a-methyloctahydro-1H-inden-1-yl)-2-hydroxyhexanoate (10)**

To a solution of **31** (219.1 mg, 0.44 mmol) in isopropanol (10 mL), we added 10% Pd/C catalyst (62.9 mg). The mixture was stirred for 45 h at room temperature, and then for 68 h at 50 °C, under a hydrogen atmosphere. The reaction mixture was diluted with EtOAc, filtered through a Celite pad, and concentrated under reduced pressure. Purification via flash column chromatography on silica gel (hexane:EtOAc = 3:1) yielded **10** (136.5 mg, 76%) as a colorless oil.

**10**:  $[\alpha]_{\text{D}}^{27} +33.2$  (c 0.61,  $\text{CHCl}_3$ ); IR (neat) 3506, 1739, 1468, 1253, 1085, 1025, 838, 778  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (600 MHz,  $\text{CDCl}_3$ )  $\delta$  –0.01 (s, 3H), 0.00 (s, 3H), 0.88–0.90 (m, 15H), 1.00–1.12 (m, 2H), 1.20–1.27 (m, 2H), 1.30–1.45 (m, 3H), 1.49–1.58 (m, 2H), 1.65–1.71 (m, 2H), 1.75–1.83 (m, 2H), 1.92–1.95 (m, 1H), 3.78 (s, 3H), 3.99–3.99 (m, 1H), 4.17–4.18 (dd,  $J = 5.6, 6.0$  Hz, 1H);  $^{13}\text{C}$  NMR (150 MHz,  $\text{CDCl}_3$ )  $\delta$  –5.2, –4.8, 13.7, 17.7, 18.0, 18.5, 23.0, 25.8, 27.2, 30.5, 30.9, 34.4, 34.8, 40.7, 42.1, 52.4, 53.0, 56.4, 69.4, 70.7, 175.9; HRMS (ESI<sup>+</sup>) calcd for  $\text{C}_{23}\text{H}_{44}\text{O}_4\text{Si}$  [ $\text{M} + \text{Na}$ ]<sup>+</sup> 435.2901, found 435.2887.

**3.12. Methyl (2S,5R)-5-((1R,3aR,4S,7aR)-4-[(tert-butyl)dimethylsilyloxy]-7a-methyloctahydro-1H-inden-1-yl)-2-fluorohexanoate (11)**

DAST (48.0 mg, 43  $\mu\text{L}$ , 0.30 mmol) was added to a solution of **10** (20.5 mg, 0.05 mmol) in  $\text{CH}_2\text{Cl}_2$  (5 mL) at 0 °C, and the mixture was stirred at the same temperature for 90 min. After the reaction was quenched with MeOH,  $\text{H}_2\text{O}$ , and saturated aqueous  $\text{NaHCO}_3$  at 0 °C, the mixture was extracted with  $\text{CH}_2\text{Cl}_2$  three times, dried over  $\text{Na}_2\text{SO}_4$ , filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 8:1) to obtain **11** (15.5 mg, 75%) as a colorless oil.

**11**:  $[\alpha]_{\text{D}}^{27} +34.4$  (c 1.03,  $\text{CHCl}_3$ ); IR (neat) 1766, 1746, 1469, 1442, 1378, 1254, 1212, 1089, 1024, 836, 776  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  –0.02 (s, 3H), 0.00 (s, 3H), 0.88–0.90 (m, 15H), 1.00–2.03 (m, 19H), 3.78 (s, 3H), 3.98–3.99 (m, 1H), 4.85 (ddd,  $J = 4.1, 7.3, 49.0$  Hz, 1H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  –5.2, –4.8, 13.7, 17.6, 18.0, 18.4, 23.0, 25.8, 27.1, 29.1 (d,  $J = 20.0$  Hz), 30.3 (d,  $J = 2.9$  Hz), 34.4, 34.9, 40.7, 42.1, 52.2, 53.0, 56.3, 69.4, 89.6 (d,  $J = 183.1$  Hz), 170.5 (d,  $J = 23.8$  Hz); HRMS (ESI<sup>+</sup>) calcd for  $\text{C}_{23}\text{H}_{43}\text{O}_3\text{FSiNa}$  [ $\text{M} + \text{Na}$ ]<sup>+</sup> 437.2858, found 437.2869.

3.13. Methyl (2R,5R)-5-[(1R,3aR,4S,7aR)-4-[(tert-butyl(dimethyl)silyloxy]-7a-methyloctahydro-1H-inden-1-yl]-2-fluorohexanoate (**12**)

DAST (195.0 mg, 173  $\mu$ L, 1.21 mmol) was added to a solution of **9** (99.7 mg, 0.24 mmol) in  $\text{CH}_2\text{Cl}_2$  (3 mL) at 0  $^\circ\text{C}$ , and the mixture was stirred at the same temperature for 2 h 15 min. After the reaction was quenched with MeOH,  $\text{H}_2\text{O}$ , and saturated aqueous  $\text{NaHCO}_3$  at 0  $^\circ\text{C}$ , the mixture was extracted with  $\text{CH}_2\text{Cl}_2$  three times, dried over  $\text{Na}_2\text{SO}_4$ , filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 8:1) to obtain **12** (31.0 mg, 31%) as a colorless oil.

**12**:  $[\alpha]_{\text{D}}^{27} +44.7$  (c 2.39,  $\text{CHCl}_3$ ); IR (neat) 1769, 1746, 1465, 1445, 1370, 1254, 1208, 1081, 1024, 836, 769  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (600 MHz,  $\text{CDCl}_3$ )  $\delta$  -0.01 (s, 3H), 0.00 (s, 3H), 0.88–0.90 (m, 15H), 0.99–1.12 (m, 2H), 1.15–1.26 (m, 3H), 1.29–1.37 (m, 3H), 1.39–1.48 (m, 1H), 1.51–1.59 (m, 2H), 1.64–1.68 (m, 1H), 1.73–1.84 (m, 3H), 1.73–1.84 (m, 3H), 1.86–1.95 (m, 2H), 3.79 (s, 3H), 3.99–3.99 (m, 1H), 4.89 (ddd,  $J = 4.2, 8.4, 49.8$  Hz, 1H);  $^{13}\text{C}$  NMR (150 MHz,  $\text{CDCl}_3$ )  $\delta$  -5.2, -4.8, 13.7, 17.6, 18.0, 18.4, 23.0, 25.8, 27.1, 29.1 (d,  $J = 20.1$  Hz), 30.1, 34.4, 34.6, 40.7, 42.1, 52.2, 53.0, 56.3, 69.4, 89.3 (d,  $J = 182.4$  Hz), 170.6 (d,  $J = 24.5$  Hz); HRMS ( $\text{ESI}^+$ ) calcd for  $\text{C}_{23}\text{H}_{43}\text{O}_3\text{FSiNa}$   $[\text{M} + \text{Na}]^+$  437.2858, found 437.2874.

3.14. (1R,3aR,4S,7aR)-1-[(2R,5S)-5-Fluoro-6-hydroxy-6-methylheptan-2-yl]-7a-methyloctahydro-1H-inden-4-ol (**15**)

To a solution of **11** (82.2 mg, 0.20 mmol) in THF (3 mL), we added MeMgCl (264  $\mu$ L, 3.0 M THF solution, 0.79 mmol) at 0  $^\circ\text{C}$ , and the mixture was stirred at 0  $^\circ\text{C}$  for 10 min. MeMgCl (264  $\mu$ L, 3.0 M THF solution, 0.79 mmol) was added to the mixture at 0  $^\circ\text{C}$  and stirred at the same temperature for 5 min. After the reaction was quenched with  $\text{H}_2\text{O}$ , the mixture was extracted with EtOAc three times, washed with saturated aqueous  $\text{NH}_4\text{Cl}$ , dried over  $\text{Na}_2\text{SO}_4$ , filtered, and concentrated. The crude residue was used for the next reaction without further purification. To the above crude residue in MeOH (10 mL) and  $\text{CH}_2\text{Cl}_2$  (5 mL), we added *p*-toluenesulfonic acid monohydrate (399.2 mg, 2.10 mmol), and the mixture was stirred at room temperature for 24 h under air. After the reaction was quenched with  $\text{H}_2\text{O}$  and saturated aqueous  $\text{NaHCO}_3$  at room temperature, the mixture was extracted with  $\text{CH}_2\text{Cl}_2$  three times, dried over  $\text{Na}_2\text{SO}_4$ , filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 1:1) to obtain **15** (36.3 mg, 61%, in 2 steps) as a white powder.

**15**:  $[\alpha]_{\text{D}}^{27} +17.5$  (c 1.30,  $\text{CHCl}_3$ ); IR (neat) 3412, 1465, 1378, 1250, 1168, 1066, 990, 731  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (600 MHz,  $\text{CDCl}_3$ )  $\delta$  0.92 (d,  $J = 6.0$  Hz, 3H), 0.94 (s, 3H), 1.03–1.17 (m, 3H), 1.20–1.21 (m, 6H), 1.29–1.36 (m, 2H), 1.42–1.90 (m, 13H), 1.98–2.01 (m, 1H), 4.07–4.08 (m, 1H), 4.14 (ddd,  $J = 1.8, 10.2, 48.0$  Hz, 1H);  $^{13}\text{C}$  NMR (150 MHz,  $\text{CDCl}_3$ )  $\delta$  13.5, 17.4, 18.5, 22.5, 24.3 (d,  $J = 4.4$  Hz), 25.3 (d,  $J = 4.4$  Hz), 26.4 (d,  $J = 21.6$  Hz), 27.1, 32.1, 33.6, 35.3, 40.4, 41.9, 52.6, 56.5, 69.4, 72.0 (d,  $J = 20.1$  Hz), 100.7 (d,  $J = 172.4$  Hz); HRMS ( $\text{ESI}^-$ ) calcd for  $\text{C}_{18}\text{H}_{32}\text{O}_2\text{FSi}$   $[\text{M}-\text{H}]^-$  299.2392, found 299.2388.

3.15. (1R,3aR,4S,7aR)-1-[(2R,5R)-5-Fluoro-6-hydroxy-6-methylheptan-2-yl]-7a-methyloctahydro-1H-inden-4-ol (**16**)

To a solution of **12** (31.0 mg, 0.075 mmol) in THF (1 mL), we added MeMgCl (150  $\mu$ L, 3.0 M THF solution, 0.45 mmol) at 0  $^\circ\text{C}$ , and the mixture was stirred at 0  $^\circ\text{C}$  for 10 min. MeMgCl (264  $\mu$ L, 3.0 M THF solution, 0.79 mmol) was added to the mixture at 0  $^\circ\text{C}$  and further stirred for 10 min. After the reaction was quenched with  $\text{H}_2\text{O}$ , the mixture was extracted with EtOAc three times, washed with saturated aqueous  $\text{NH}_4\text{Cl}$ , dried over  $\text{Na}_2\text{SO}_4$ , filtered, and concentrated. The crude residue was used for the next reaction without further purification. To the above crude residue in MeOH (10 mL) and  $\text{CH}_2\text{Cl}_2$  (5 mL), we added *p*-toluenesulfonic acid monohydrate (380.7 mg, 2.0 mmol), and the mixture was stirred at room temperature for 24 h under air. After the reaction was quenched with  $\text{H}_2\text{O}$  and saturated aqueous  $\text{NaHCO}_3$  at room temperature, the mixture was extracted

with  $\text{CH}_2\text{Cl}_2$  three times, dried over  $\text{Na}_2\text{SO}_4$ , filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 2:1) to obtain **16** (18.8 mg, 83%, in 2 steps) as a white powder.

**16**:  $[\alpha]_{\text{D}}^{27} +43.4$  (c 1.45,  $\text{CHCl}_3$ ); IR (neat) 3402, 1469, 1374, 1168, 1073, 994  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (600 MHz,  $\text{CDCl}_3$ )  $\delta$  0.91 (d,  $J = 6.0$  Hz, 3H), 0.94 (s, 3H), 1.01–1.74 (m, 21H), 1.78–1.91 (m, 3H), 1.98–2.00 (m, 1H), 4.07–4.07 (m, 1H), 4.18 (ddd,  $J = 2.1, 10.5, 48.6$  Hz, 1H);  $^{13}\text{C}$  NMR (150 MHz,  $\text{CDCl}_3$ )  $\delta$  13.5, 17.4, 18.3, 22.5, 24.2 (d,  $J = 4.4$  Hz), 25.4 (d,  $J = 4.4$  Hz), 26.0 (d,  $J = 21.6$  Hz), 27.1, 31.7, 33.6, 34.9, 40.4, 41.9, 52.6, 56.4, 69.4, 72.0 (d,  $J = 20.1$  Hz), 99.9 (d,  $J = 170.9$  Hz); HRMS (APCI<sup>-</sup>) calcd for  $\text{C}_{18}\text{H}_{33}\text{O}_2\text{FSi}$   $[\text{M}-\text{H}]^-$  299.2392, found 299.2420.

### 3.16. (24S)-24-Fluoro-25-hydroxyvitamin D<sub>3</sub> (**3**)

4-Methylmorpholine *N*-oxide (32.6 mg, 0.28 mmol) was added to a solution of **15** (22.2 mg, 0.074 mmol) in  $\text{CH}_2\text{Cl}_2$  (2 mL), and the mixture was cooled to 0 °C. Tetrapropylammonium perruthenate (TPAP, 15.2 mg, 0.043 mmol) was added to the mixture, and the mixture was stirred at room temperature for 1 h. The reaction was diluted with  $\text{Et}_2\text{O}$ , and the mixture was directly purified via flash column chromatography on silica gel ( $\text{Et}_2\text{O}$  only) to obtain the crude ketone, which was used for the next reaction without further purification.

TMSCl (80.4 mg, 94  $\mu\text{L}$ , 0.74 mmol) was added to the 0 °C cooled solution of crude ketone and imidazole (66.5 mg, 0.98 mmol) in  $\text{CH}_2\text{Cl}_2$  (2 mL), and the mixture was stirred at the same temperature for 15 min. After the reaction was quenched with  $\text{H}_2\text{O}$  and saturated aqueous  $\text{NH}_4\text{Cl}$  at 0 °C, the mixture was extracted with  $\text{CH}_2\text{Cl}_2$  three times, dried over  $\text{Na}_2\text{SO}_4$ , filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 10:1) to obtain crude **32**.

*n*BuLi (191  $\mu\text{L}$ , 1.55 M hexane solution, 0.30 mmol) was added to a solution of A-ring phosphine oxide [16] (132.6 mg, 0.29 mmol) in THF (1.5 mL) at −78 °C. After stirring for 15 min, the solution of crude **32** in THF (2 mL) was added, and the mixture was stirred at −78 °C for 15 min and 0 °C for 5 min. After the reaction was quenched with  $\text{H}_2\text{O}$  and saturated aqueous  $\text{NH}_4\text{Cl}$  at the same temperature, the mixture was extracted with EtOAc three times, washed with brine, dried over  $\text{Na}_2\text{SO}_4$ , filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 10:1) to obtain the crude coupling product (34.9 mg), which was used for the next reaction without further purification. Tetrabutylammonium fluoride (370  $\mu\text{L}$ , 1 M THF solution, 0.37 mmol) was added to a solution of the crude coupling product (34.9 mg) in THF (2 mL), and the mixture was stirred at room temperature for 16 h. After the reaction was quenched with  $\text{H}_2\text{O}$  at room temperature, the mixture was extracted with EtOAc three times, dried over  $\text{Na}_2\text{SO}_4$ , filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 1:1) to obtain **3** (15.6 mg, 50%, in 4 steps) as a white powder.

**3**:  $[\alpha]_{\text{D}}^{27} +85.1$  (c 1.20, EtOH); IR (neat) 3369, 1455, 1375, 1168, 1054, 893  $\text{cm}^{-1}$ ;  $^1\text{H}$  NMR (400 MHz,  $\text{CD}_3\text{OD}$ )  $\delta$  0.61 (s, 3H), 1.02 (d,  $J = 6.4$  Hz, 3H), 1.11–1.19 (m, 1H), 1.21 (s, 3H), 1.23 (d,  $J = 1.8$  Hz, 3H), 1.35–2.26 (m, 21H), 2.45 (dt,  $J = 4.8, 13.8$  Hz, 1H), 2.58 (dd,  $J = 3.7, 12.8$  Hz, 1H), 2.89–2.92 (m, 1H), 3.77–3.84 (m, 1H), 4.03–4.18 (m, 1H), 4.79 (d,  $J = 1.8$  Hz, 1H), 5.08 (brs, 1H), 6.08 (d,  $J = 11.4$  Hz, 1H), 6.26 (d,  $J = 11.4$  Hz, 1H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CD}_3\text{OD}$ )  $\delta$  12.7, 19.7, 23.6, 24.9, 25.0 (d,  $J = 2.9$  Hz), 26.0 (d,  $J = 2.8$  Hz), 27.5, 27.8, 29.0, 30.2, 33.9, 34.0, 36.9, 37.8, 42.2, 47.3 (d,  $J = 13.4$  Hz), 57.8, 58.1, 70.9, 72.7 (d,  $J = 21.0$  Hz), 101.6 (d,  $J = 173.5$  Hz), 113.0, 119.3, 122.9, 137.6, 142.8, 147.3; HRMS (ESI<sup>+</sup>) calcd for  $\text{C}_{27}\text{H}_{43}\text{O}_2\text{FNa}$   $[\text{M} + \text{Na}]^+$  441.3139, found 441.3106.

### 3.17. (24R)-24-Fluoro-25-hydroxyvitamin D<sub>3</sub> (**4**)

4-Methylmorpholine *N*-oxide (26.1 mg, 0.22 mmol) was added to a solution of **16** (18.8 mg, 0.063 mmol) in  $\text{CH}_2\text{Cl}_2$  (2 mL), and the mixture was cooled to 0 °C. TPAP (11.2 mg, 0.032 mmol) was added to the mixture, and the mixture was stirred at 0 °C for 10 min and room temperature for 20 min. The reaction was diluted with  $\text{Et}_2\text{O}$ , and the mixture was

directly purified via flash column chromatography on silica gel (Et<sub>2</sub>O only) to obtain the crude ketone, which was used for the next reaction without further purification.

TMSCl (68.4 mg, 80  $\mu$ L, 0.63 mmol) was added to the 0 °C cooled solution of crude ketone and imidazole (43.7 mg, 0.64 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (2 mL), and the mixture was stirred for 7 min at room temperature. After the reaction was quenched with H<sub>2</sub>O and saturated aqueous NH<sub>4</sub>Cl, the mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub> three times, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 10:1) to obtain crude **33**.

*n*BuLi (163  $\mu$ L, 1.55 M hexane solution, 0.25 mmol) was added to a solution of A-ring phosphine oxide [16] (117.4 mg, 0.26 mmol) in THF (1.5 mL) at –78 °C. After stirring for 15 min, a solution of crude **33** in THF (2 mL) was added, and the mixture was stirred at –78 °C for 15 min and 0 °C for 5 min. After the reaction was quenched with H<sub>2</sub>O at the same temperature, the mixture was extracted with EtOAc three times, washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 10:1) to obtain the crude coupling product (24.7 mg), and it was used for the next reaction without further purification. Tetrabutylammonium fluoride (315  $\mu$ L, 1 M THF solution, 0.32 mmol) was added to a solution of the crude coupling product (24.7 mg) in THF (2 mL), and the mixture was stirred at room temperature for 16 h. After the reaction was quenched with H<sub>2</sub>O at room temperature, the mixture was extracted with EtOAc three times, dried over MgSO<sub>4</sub>, filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 1:1) to obtain **4** (16.0 mg, 61%, in 4 steps) as a white powder.

**4**: [ $\alpha$ ]<sub>D</sub><sup>27</sup> +84.2 (c 1.24, EtOH); IR (neat) 3381, 1455, 1375, 1168, 1054, 881 cm<sup>–1</sup>; <sup>1</sup>H NMR (600 MHz, CD<sub>3</sub>OD)  $\delta$  0.62 (s, 3H), 1.02 (d, *J* = 6.6 Hz, 3H), 1.22 (d, *J* = 1.2 Hz, 3H), 1.23 (d, *J* = 1.2 Hz, 3H), 1.34–1.42 (m, 4H), 1.48–1.77 (m, 10H), 1.94–2.09 (m, 4H), 2.14–2.25 (m, 2H), 2.45 (dt, *J* = 5.1, 13.8 Hz, 1H), 2.58 (dd, *J* = 3.9, 12.6 Hz, 1H), 2.89–2.92 (m, 1H), 3.79–3.83 (m, 1H), 4.15 (ddd, *J* = 1.5, 10.8, 48.6 Hz, 1H), 4.79 (d, *J* = 1.2 Hz, 1H), 5.08 (brs, 1H), 6.09 (d, *J* = 11.1 Hz, 1H), 6.27 (d, *J* = 11.1 Hz, 1H); <sup>13</sup>C NMR (150 MHz, CD<sub>3</sub>OD)  $\delta$  12.7, 19.5, 23.5, 24.9, 25.2 (d, *J* = 2.9 Hz), 25.9 (d, *J* = 2.9 Hz), 27.2, 27.3, 29.0, 30.2, 33.5, 33.9, 36.9, 37.3, 42.2, 47.3 (d, *J* = 17.3 Hz), 57.8, 58.1, 70.9, 72.7 (d, *J* = 20.1 Hz), 100.7 (d, *J* = 173.7 Hz), 112.9, 119.3, 122.9, 137.7, 142.8, 147.3; HRMS (ESI+) calcd for C<sub>27</sub>H<sub>43</sub>O<sub>2</sub>FNa [M + Na]<sup>+</sup> 441.3139, found 441.3133.

**3.18.** (3*S*,6*R*)-6-((1*R*,3*aR*,4*S*,7*aR*)-4-[(*tert*-Butyldimethylsilyl)oxy]-7*a*-methyloctahydro-1*H*-inden-1-yl)-2-methylheptane-2,3-diol (**36**)

MeMgCl (0.53 mL, 3.0 M THF solution, 1.59 mmol) was added to a solution of **30** (133.4 mg, 0.265 mmol) in THF (4 mL) at 0 °C, and the mixture was stirred at 0 °C for 11 min. After the reaction was quenched with H<sub>2</sub>O, the mixture was extracted with EtOAc three times, washed with saturated aqueous NH<sub>4</sub>Cl, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 4:1) to obtain crude **34** (130.0 mg), and it was used for the next reaction without further purification.

To a solution of crude **34** (130.0 mg) in MeOH (4 mL), we added 10% Pd/C catalyst (20.0 mg). The mixture was stirred for 6 days at room temperature under a hydrogen atmosphere. The reaction mixture was diluted with EtOAc, filtered through a Celite pad, and concentrated under reduced pressure. Purification via flash column chromatography on silica gel (hexane:EtOAc = 2:1) yielded **36** (87.2 mg, 82%) as a colorless oil [25].

**36**: [ $\alpha$ ]<sub>D</sub><sup>27</sup> +31.9 (c 6.71, CHCl<sub>3</sub>); IR (neat) 3398, 1469, 1374, 1250, 1164, 1085, 1069, 1024, 832, 776, 739 cm<sup>–1</sup>; <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  –0.02 (s, 3H), –0.01 (s, 3H), 0.87–0.90 (m, 15H), 0.97–1.41 (m, 16H), 1.50–1.57 (m, 2H), 1.64–1.81 (m, 4H), 1.92–1.95 (m, 1H), 2.40 (s, 2H), 3.25 (dd, *J* = 2.1, 9.9 Hz, 1H), 3.98–3.98 (m, 1H); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>)  $\delta$  –5.2, –4.8, 13.7, 17.6, 18.0, 18.7, 23.0, 23.1, 25.8, 26.5, 27.3, 28.3, 33.1, 34.4, 35.4, 40.7, 42.1, 53.0, 56.6, 69.4, 73.3, 79.6; HRMS (ESI+) calcd for C<sub>24</sub>H<sub>48</sub>O<sub>3</sub>SiNa [M + Na]<sup>+</sup> 435.3265, found 435.3271.

3.19. (3R,6R)-6-[(1R,3aR,4S,7aR)-4-[(tert-Butyldimethylsilyl)oxy]-7a-methyloctahydro-1H-inden-1-yl]-2-methylheptane-2,3-diol (**37**)

MeMgCl (415  $\mu$ L, 3.0 M THF solution, 1.25 mmol) was added to a solution of **31** (125.1 mg, 0.249 mmol) in THF (3 mL) at 0 °C, and the mixture was stirred at 0 °C for 7 min. After the reaction was quenched with H<sub>2</sub>O, the mixture was extracted with EtOAc three times, washed with saturated aqueous NH<sub>4</sub>Cl, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 4:1) to obtain crude **35**, which was used for the next reaction without further purification.

To a solution of crude **35** in MeOH (4 mL), we added 10% Pd/C catalyst (20.0 mg). The mixture was stirred for 68 h at room temperature under a hydrogen atmosphere. The reaction mixture was diluted with EtOAc, filtered through a Celite pad, and concentrated under reduced pressure. Purification via flash column chromatography on silica gel (hexane:EtOAc = 2:1) yielded **37** (24.1 mg, 23%, **35** recovery 54%) as a colorless oil [25].

**37**: [ $\alpha$ ]<sub>D</sub><sup>27</sup> +62.5 (c 1.85, CHCl<sub>3</sub>); IR (neat) 3409, 1469, 1378, 1254, 1164, 1073, 1024, 840, 772, 739 cm<sup>-1</sup>; <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  -0.01 (s, 3H), 0.00 (s, 3H), 0.88–0.91 (m, 15H), 0.98–1.48 (m, 18H), 1.51–1.58 (m, 1H), 1.65–1.67 (m, 1H), 1.76–1.86 (m, 2H), 1.93–1.96 (m, 4H), 3.32–3.34 (m, 1H), 4.00–4.00 (m, 1H); <sup>13</sup>C NMR (150 MHz, CDCl<sub>3</sub>)  $\delta$  -5.2, -4.8, 13.7, 17.7, 18.0, 18.5, 23.0, 23.2, 25.8, 26.6, 27.4, 28.1, 32.7, 34.4, 35.1, 40.7, 42.1, 53.0, 56.7, 69.5, 73.2, 78.8; HRMS (ESI<sup>+</sup>) calcd for C<sub>24</sub>H<sub>48</sub>O<sub>3</sub>SiNa [M + Na]<sup>+</sup> 435.3265, found 435.3282.

3.20. (3S,6R)-6-[(1R,3aR,4S,7aR)-4-Hydroxy-7a-methyloctahydro-1H-inden-1-yl]-2-methylheptane-2,3-diol (**13**)

*p*-Toluenesulfonic acid monohydrate (199.1 mg, 1.01 mmol) was added to a solution of **36** (46.5 mg, 0.11 mmol) in MeOH (4 mL) and CH<sub>2</sub>Cl<sub>2</sub> (4 mL), and the mixture was stirred at room temperature for 45 h under air. After the reaction was quenched with H<sub>2</sub>O and saturated aqueous NaHCO<sub>3</sub> at room temperature, the mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub> three times, washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (EtOAc only) to obtain **13** (30.4 mg, 90%) as a colorless oil. The spectral data of the product matched those reported in the literature [25].

3.21. (3R,6R)-6-[(1R,3aR,4S,7aR)-4-Hydroxy-7a-methyloctahydro-1H-inden-1-yl]-2-methylheptane-2,3-diol (**14**)

*p*-Toluenesulfonic acid monohydrate (192.9 mg, 1.01 mmol) was added to a solution of **37** (49.2 mg, 0.12 mmol) in MeOH (5 mL) and CH<sub>2</sub>Cl<sub>2</sub> (5 mL), and the mixture was stirred at room temperature for 53 h under air. After the reaction was quenched with H<sub>2</sub>O and saturated aqueous NaHCO<sub>3</sub> at room temperature, the mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub> three times, washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (EtOAc only) to obtain **14** (30.4 mg, 85%) as a colorless oil. The spectral data of the product matched those reported in the literature [25].

3.22. (1R,3aR,4S,7aR)-7a-Methyl-1-[(R)-4-[(S)-2,2,5,5-tetramethyl-1,3-dioxolan-4-yl]butan-2-yl]octahydro-1H-inden-4-ol (**38**)

PPTS (15.8 mg, 0.06 mmol) was added to the solution of **13** (30.4 mg, 0.10 mmol) in acetone (1 mL) and 2,2-dimethoxypropane (1 mL), and the mixture was stirred at room temperature for 19 h under air. After the reaction was quenched with H<sub>2</sub>O and saturated aqueous NaHCO<sub>3</sub> at room temperature, the mixture was extracted with EtOAc three times, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 1:1) to obtain **38** (32.3 mg, 94%) as a colorless oil. The spectral data of the product matched those reported in the literature [25].

3.23. (1R,3aR,4S,7aR)-7a-Methyl-1-[(R)-4-[(R)-2,2,5,5-tetramethyl-1,3-dioxolan-4-yl]butan-2-yl]octahydro-1H-inden-4-ol (**39**)

PPTS (19.4 mg, 0.08 mmol) was added to a solution of **14** (30.4 mg, 0.10 mmol) in acetone (1 mL) and 2,2-dimethoxypropane (1 mL), and the mixture was stirred at room temperature for 4 h under air. After the reaction was quenched with H<sub>2</sub>O and saturated aqueous NaHCO<sub>3</sub> at room temperature, the mixture was extracted with EtOAc three times, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 1:1) to obtain **39** (28.6 mg, 83%) as a colorless oil. The spectral data of the product matched those reported in the literature [25].

3.24. (1R,3aR,7aR)-7a-Methyl-1-[(R)-4-[(S)-2,2,5,5-tetramethyl-1,3-dioxolan-4-yl]butan-2-yl]octahydro-4H-inden-4-one (**40**)

4-Methylmorpholine *N*-oxide (31.2 mg, 0.27 mmol) was added to a solution of **38** (32.3 mg, 0.095 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (2 mL), and the mixture was cooled to 0 °C. TPAP (18.3 mg, 0.052 mmol) was added to the mixture, and the mixture was stirred at 0 °C for 1 h. The reaction was diluted with an excess amount of Et<sub>2</sub>O. The mixture was directly purified via flash column chromatography on silica gel (Et<sub>2</sub>O only), followed by purification via flash column chromatography on silica gel (hexane:EtOAc = 4:1), to obtain **40** (25.2 mg, 79%) as a colorless oil. The spectral data of the product matched those reported in the literature [25].

3.25. (1R,3aR,7aR)-7a-Methyl-1-[(R)-4-[(R)-2,2,5,5-tetramethyl-1,3-dioxolan-4-yl]butan-2-yl]octahydro-4H-inden-4-one (**41**)

4-Methylmorpholine *N*-oxide (28.8 mg, 0.25 mmol) was added to a solution of **39** (28.6 mg, 0.085 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (1 mL), and the mixture was cooled to 0 °C. TPAP (13.9 mg, 0.04 mmol) was added to the mixture, and the mixture was stirred at 0 °C for 40 min. The reaction was diluted with Et<sub>2</sub>O, and the mixture was directly purified via flash column chromatography on silica gel (Et<sub>2</sub>O only), followed by purification via flash column chromatography on silica gel (hexane:EtOAc = 4:1), to obtain **41** (28.2 mg, 99%) as a colorless oil. The spectral data of the product matched those reported in the literature [25].

3.26. (24S)-24,25-Dihydroxyvitamin D<sub>3</sub> (**5**)

*n*BuLi (145 µL, 1.55 M hexane solution, 0.225 mmol) was added to a solution of A-ring phosphine oxide [16] (101.4 mg, 0.22 mmol) in THF (1 mL) at −78 °C. After stirring for 15 min, a solution of **40** (25.2 mg, 0.075 mmol) in THF (1.5 mL) was added, and the mixture was stirred at −78 °C for 2 h. After the reaction was quenched with H<sub>2</sub>O at the same temperature, the mixture was extracted with EtOAc three times, washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 10:1) to obtain the crude coupling product (39.4 mg), which was used for the next reaction without further purification. Tetrabutylammonium fluoride (414 µL, 1 M THF solution, 0.414 mmol) was added to the solution of the crude coupling product (39.4 mg) in THF (3 mL), and the mixture was stirred at room temperature for 15 h. After the reaction was quenched with H<sub>2</sub>O and aqueous saturated NH<sub>4</sub>Cl at room temperature, the mixture was extracted with EtOAc three times, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 1:2) to obtain the crude product, which was used for the next reaction without further purification.

The above crude residue was dissolved in MeOH (10 mL), and AG 50W-X4 resin (177.2 mg) was added. The mixture was then stirred for 26 h, and the solids were filtered off, washed with MeOH, and the solution was concentrated in vacuo. The residue was purified via flash column chromatography (hexane:EtOAc = 1:2) to obtain **5** (20.7 mg, 66%) as a white powder. The spectral data of the product matched those reported in the literature [25].



### 3.27. (24*R*)-24,25-Dihydroxyvitamin D<sub>3</sub> (6)

*n*BuLi (163  $\mu$ L, 1.55 M hexane solution, 0.252 mmol) was added to a solution of A-ring phosphine oxide [16] (110.7 mg, 0.24 mmol) in THF (1 mL) at  $-78$  °C. After stirring for 20 min, a solution of **41** (28.2 mg, 0.084 mmol) in THF (1 mL) was added, and the mixture was stirred at  $-78$  °C for 2 h 30 min. After the reaction was quenched with H<sub>2</sub>O at the same temperature, the mixture was extracted with EtOAc three times, washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 10:1) to obtain the crude coupling product (43.3 mg), which was used for the next reaction without further purification. Tetrabutylammonium fluoride (420  $\mu$ L, 1 M THF solution, 0.42 mmol) was added to the solution of the crude coupling product (43.3 mg) in THF (3 mL), and the mixture was stirred at room temperature for 17 h. After the reaction was quenched with H<sub>2</sub>O and aqueous saturated NH<sub>4</sub>Cl at room temperature, the mixture was extracted with EtOAc three times, dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The residue was purified via flash column chromatography on silica gel (hexane:EtOAc = 1:1) to obtain the crude product, which was used for the next reaction without further purification.

The above crude residue was dissolved in MeOH (5 mL), and AG 50W-X4 resin (167.5 mg) was added. The mixture was stirred for 24 h, and the solids were filtered off, washed with MeOH, and the solution was concentrated in vacuo. The residue was purified via flash column chromatography (hexane:EtOAc = 1:2) to obtain **6** (26.6 mg, 76%, in 3 steps) as a white powder. The spectral data of the product matched those reported in the literature [25].

### 3.28. Measurement of the hVDR Binding Affinity of **3**, **4**, and 24,24-Difluoro-25(OH)D<sub>3</sub>

The binding affinity of each analogue for hVDR was evaluated using an in vitro system based on the split-luciferase technique described in our previous study [29]. Briefly, 50  $\mu$ L of cell lysate prepared from recombinant *Escherichia coli* expressing split-luciferase vitamin D biosensor protein [29] was added to each well of a 96-well plate, and left for 10 min at room temperature. Then, 50  $\mu$ L of the luciferin solution containing 20 mM MgSO<sub>4</sub>, 2 mM D-luciferin, and 4 mM adenosine triphosphate in 25 mM Tris-HCl (pH 7.4) was injected into each well and incubated for 15 min at room temperature. The luminescence (photon counts) was measured using a luminometer. The relative hVDR binding affinity of each analogue was evaluated based on the concentration at which the luminescence showed 50% of the maximum value.

### 3.29. Metabolism of 25(OH)D<sub>3</sub> and Its Analogues by Recombinant hCYP24A1

The metabolism of 25(OH)D<sub>3</sub> and its analogues **3** and **4** by CYP24A1 was analyzed using the membrane fraction prepared from the recombinant *Escherichia coli* cells expressing human CYP24A1, as described in our previous study [30]. Briefly, the reaction mixture containing 0.02  $\mu$ M human CYP24A1, 2.0  $\mu$ M adrenodoxin (ADX), 0.2  $\mu$ M NADPH-adrenodoxin reductase (ADR), 1 mM EDTA, 1 mM NADPH, and 5.0  $\mu$ M of each substrate in 100 mM Tris-HCl (pH 7.4) was incubated at 37 °C for 5 or 15 min. The metabolites were extracted with 4 volumes of CHCl<sub>3</sub>-CH<sub>3</sub>OH (3:1) and analyzed via HPLC under the following conditions: column, CAPCELL PAK C18 UG120 (5  $\mu$ m) (4.6 mm  $\times$  250 mm) (SHISEIDO, Tokyo, Japan); UV detection, 265 nm; flow rate, 1.0 mL min<sup>-1</sup>; column temperature, 40 °C; mobile phase, CH<sub>3</sub>CN: a linear gradient of 20–100% CH<sub>3</sub>CN aqueous solution per 25 min and 100% CH<sub>3</sub>CN for 10 min.

## 4. Conclusions

In summary, in this paper we described novel stereoselective syntheses of 24-fluoro-25-hydroxyvitamin D<sub>3</sub> (**3** and **4**) and 24,25-dihydroxyvitamin D<sub>3</sub> (**5** and **6**). To our knowledge, this is the first reported study to synthesize both 24*R*- and 24*S*-24-fluorinated vitamin D<sub>3</sub> analogues. This approach also provides a practical synthetic route to one of the main natural metabolites of 25(OH)D<sub>3</sub> by hCYP24A1—(24*R*)-24,25-dihydroxyvitamin D<sub>3</sub> (**6**).

This synthetic method paves the way for efficient access to 24-substituted vitamin D<sub>3</sub> analogues. Synthesis of new 24-substituted vitamin D<sub>3</sub> analogues utilizing this method, along with evaluation of their biological activities, is in progress.

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## References

1. Sakaki, T.; Kagawa, N.; Yamamoto, K.; Inouye, K. Metabolism of vitamin D<sub>3</sub> by cytochromes P450. *Front. Biosci.* **2005**, *10*, 119–134. [[PubMed](#)]
2. Sakaki, T.; Sawada, N.; Komai, K.; Shiozawa, S.; Yamada, S.; Yamamoto, K.; Ohyama, Y.; Inouye, K. Dual metabolic pathway of 25-hydroxyvitamin D<sub>3</sub> catalyzed by human CYP24. *Eur. J. Biochem.* **2000**, *267*, 6158–6165. [[CrossRef](#)] [[PubMed](#)]
3. Inouye, K.; Sakaki, T. Enzymatic studies on the key enzymes of vitamin D metabolism; 1 $\alpha$ -hydroxylase (CYP27B1) and 24-hydroxylase (CYP24). *Biotechnol. Ann. Rev.* **2001**, *7*, 179–194.
4. Jones, G.; Strugnell, S.A.; DeLuca, H.F. Current understanding of the molecular actions of vitamin D. *Physiol. Rev.* **1998**, *78*, 1193–1231. [[CrossRef](#)]
5. Jones, G.; Prosser, D.E.; Kaufmann, M. 25-Hydroxyvitamin D-24-hydroxylase (CYP24A1): Its important role in the degradation of vitamin D. *Arch. Biochem. Biophys.* **2012**, *52*, 9–18. [[CrossRef](#)] [[PubMed](#)]
6. Tashiro, K.; Abe, T.; Oue, N.; Yasui, W.; Ryoji, M. Characterization of vitamin D-mediated induction of the CYP 24 transcription. *Mol. Cell. Endocrinol.* **2004**, *226*, 27–32. [[CrossRef](#)] [[PubMed](#)]
7. Ohyama, Y.; Noshiro, M.; Okuda, K. Cloning and expression of cDNA encoding 25-hydroxyvitamin 24-hydroxylase. *FEBS Lett.* **1991**, *278*, 195–198. [[CrossRef](#)]
8. Yasuda, K.; Nishikawa, M.; Okamoto, K.; Horibe, K.; Mano, H.; Yamaguchi, M.; Okon, R.; Nakagawa, K.; Tsugawa, N.; Okano, T.; et al. Elucidation of metabolic pathways of 25-hydroxyvitamin D<sub>3</sub> mediated by Cyp24A1 and Cyp3A using Cyp24a1 knockout rats generated by CRISPR/Cas9 system. *J. Biol. Chem.* **2021**, *296*, 100668. [[CrossRef](#)]
9. Kawagoe, F.; Yasuda, K.; Mototani, S.; Sugiyama, T.; Uesugi, M.; Sakaki, T.; Kittaka, A. Synthesis and CYP24A1-dependent metabolism of 23-fluorinated vitamin D<sub>3</sub> analogues. *ACS Omega* **2019**, *4*, 11332–11337. [[CrossRef](#)]
10. Kawagoe, F.; Sugiyama, T.; Yasuda, K.; Uesugi, M.; Sakaki, T.; Kittaka, A. Concise synthesis of 23-hydroxylated vitamin D<sub>3</sub> metabolites. *J. Steroid Biochem. Mol. Biol.* **2019**, *186*, 161–168. [[CrossRef](#)]
11. Kawagoe, F.; Mendoza, A.; Hayata, Y.; Asano, L.; Kotake, K.; Mototani, S.; Kawamura, S.; Kurosaki, S.; Akagi, Y.; Takemoto, Y.; et al. Discovery of a vitamin D receptor-silent vitamin D derivative that impairs sterol regulatory element-binding protein in vivo. *J. Med. Chem.* **2021**, *64*, 5689–5709. [[CrossRef](#)]
12. Kobayashi, Y.; Taguchi, T.; Terada, T.; Oshida, J.; Morisaki, M.; Ikekawa, N. Synthesis of 24,24-difluoro- and 24 $\epsilon$ -fluoro-25-hydroxyvitamin D<sub>3</sub>. *Tetrahedron Lett.* **1979**, *22*, 2023–2026. [[CrossRef](#)]
13. Kobayashi, Y.; Taguchi, T.; Terada, T.; Oshida, J.; Morisaki, M.; Ikekawa, N. Studies on organic fluorine compounds. Part 37. Studies on steroids. Part 78. Synthesis of 24,24-difluoro- and 24( $\epsilon$ )-fluoro-25-hydroxyvitamin D<sub>3</sub>. *J. Chem. Soc. Perkin Trans.* **1982**, *1*, 85–91. [[CrossRef](#)]
14. Shiuey, S.J.; Partridge, J.J.; Chadha, N.K.; Boris, A.; Uskoković, M.R. Stereospecific synthesis of 1 $\alpha$ ,25-dihydroxy-24R-fluorocholecalciferol (Ro 23-0233). *Proc. Workshop Vitamin D* **1985**, *6*, 765–766.
15. Shiuey, S.; Partridge, J.J.; Uskoković, M.R. Triply convergent synthesis of 1 $\alpha$ ,25-dihydroxy-24(R)-fluorocholecalciferol. *J. Org. Chem.* **1988**, *53*, 1040–1046. [[CrossRef](#)]
16. Toh, H.T.; Okamura, W.H. Studies on a convergent route to side-chain analogues of vitamin D: 25-hydroxy-23-oxavitamin D<sub>3</sub>. *J. Org. Chem.* **1983**, *48*, 1414–1417. [[CrossRef](#)]

17. Sharpless, K.B.; Amberg, W.; Bennani, Y.L.; Crispino, G.A.; Hartung, J.; Jeong, K.S.; Kwong, H.L.; Morikawa, K.; Wang, Z.M.; Xu, D.; et al. The osmium catalyzed asymmetric dihydroxylation: A new ligand class and a process improvement. *J. Org. Chem.* **1992**, *57*, 2768–2771. [[CrossRef](#)]
18. Heravi, M.M.; Zadsirjan, V.; Esfandyari, M.; Lashaki, T.B. Applications of Sharpless asymmetric dihydroxylation in the total synthesis of natural products. *Tetrahedron Asymm.* **2017**, *28*, 987–1043. [[CrossRef](#)]
19. Gandara, Z.; Suarez, P.L.; Gonzalez, M.; Gomez, G.; Fall, Y. Vitamin D heterocyclic analogues; part 2: Synthesis of the first vitamin D analogues with a tetrazole ring at the side chain. *Synthesis* **2011**, *23*, 3887–3893.
20. Lam, H.-Y.; Schnoes, H.K.; DeLuca, H.F.; Chen, T.C. 24,25-Dihydroxyvitamin D<sub>3</sub>. Synthesis and biological activity. *Biochemistry* **1973**, *12*, 4851–4855. [[CrossRef](#)] [[PubMed](#)]
21. Takayama, H.; Ohmori, M.; Yamada, S. Facile, stereoselective synthesis of (24R)-24,25-dihydroxyvitamin D<sub>3</sub> using D-glyceric acid as a chiral synthon. *Tetrahedron Lett.* **1980**, *21*, 5027–5028. [[CrossRef](#)]
22. Sterling, J.; Slovin, E.; Barasch, D. Use of malic acid as a chiral synthon: 24,25-dihydroxycholecalciferol. *Tetrahedron Lett.* **1987**, *28*, 1685–1688. [[CrossRef](#)]
23. Schrötter, E.; Schröneck, B.; Hauschild, U.; Droscher, P.; Schick, H. An improved synthesis of (24R)-24,25-dihydroxyprovitamin D<sub>3</sub>. *Synthesis* **1990**, 193–195. [[CrossRef](#)]
24. Stepanenko, W.; Wicha, J. Enantioselective synthesis of 24,25-dihydroxy vitamin D<sub>3</sub> northern portion from (S)-3-hydroxy-2,2-dimethylcyclohexane-1-one. Remote asymmetric induction in an acid-catalysed conjugate addition. *Tetrahedron Lett.* **1998**, *39*, 885–888. [[CrossRef](#)]
25. Sestelo, J.P.; Cornella, I.; de Uña, O.; Mouriño, A.; Sarandeses, L.A. Stereoselective convergent synthesis of 24,25-dihydroxyvitamin D<sub>3</sub> metabolites: A practical approach. *Chem. Eur. J.* **2002**, *8*, 2747–2752. [[CrossRef](#)]
26. Fernández, C.; Gándara, Z.; Gómez, G.; Covelo, B.; Fall, Y. D- and L-Serine, useful synthons for the synthesis of 24-hydroxyvitamin D<sub>3</sub> metabolites. A formal synthesis of 1 $\alpha$ ,24R,25-(OH)<sub>3</sub>-D<sub>3</sub>, 24R,25-(OH)<sub>2</sub>-D<sub>3</sub> and 24S,25-(OH)<sub>2</sub>-D<sub>3</sub>. *Tetrahedron Lett.* **2007**, *48*, 2939–2942.
27. Nicoletti, D.; Gregorio, C.; Mouriño, A.; Maestro, M. A short practical approach to 24R,25-dihydroxyvitamin D<sub>3</sub>. *J. Steroid Biochem. Mol. Biol.* **2010**, *121*, 43–45. [[CrossRef](#)]
28. Kawagoe, F.; Mototani, S.; Yasuda, K.; Nagasawa, K.; Uesugi, M.; Sakaki, T.; Kittaka, A. Introduction of fluorine atoms to vitamin D<sub>3</sub> side-chain and synthesis of 24,24-difluoro-25-hydroxyvitamin D<sub>3</sub>. *J. Steroid Biochem. Mol. Biol.* **2019**, *195*, 105477. [[CrossRef](#)]
29. Mano, H.; Ikushiro, S.; Saito, N.; Kittaka, A.; Sakaki, T. Development of a highly sensitive in vitro system to detect and discriminate between vitamin D receptor agonists and antagonists based on split-luciferase technique. *J. Steroid Biochem. Mol. Biol.* **2018**, *178*, 55–59. [[CrossRef](#)] [[PubMed](#)]
30. Kusudo, T.; Sakaki, T.; Abe, D.; Fujishima, T.; Kittaka, A.; Takayama, H.; Hatakeyama, S.; Ohta, M.; Inouye, K. Metabolism of A-ring diastereomers of 1 $\alpha$ ,25-dihydroxyvitamin D<sub>3</sub> by CYP24A1. *Biochem. Biophys. Res. Commun.* **2004**, *321*, 774–782. [[CrossRef](#)] [[PubMed](#)]