



FULL PAPER

Parasitology

Fasciola hepatica infection in water buffalo *Bubalus bubalis* in three provinces of the Nile Delta, Egypt: a cross-sectional study

Abdelgawad Salah EL-TAHAWY¹⁾, Nigel KWAN²⁾ and Katsuaki SUGIURA^{2)*}

¹⁾Department of Animal Husbandry and Wealth Development, Faculty of Veterinary Medicine, Damanhur University, Damanhour, Beheira Governorate, Egypt

²⁾Department of Global Agricultural Sciences, Graduate School of Agricultural and Life Sciences, The University of Tokyo, 1-1-1 Yayoi, Bunkyo-ku, Tokyo 113-8657, Japan

ABSTRACT. A cross-sectional study was conducted to estimate the prevalence of *Fasciola hepatica* (*F. hepatica*) infection in water buffalo (*Bubalus bubalis*) in Alexandria, Beheira, and Kafr el-Sheikh governorates (provinces) of the Nile Delta in Egypt and to identify the underlying risk factors associated with the infection. A total of 29 farms (10 in Alexandria, 10 in Beheira, and 9 in Kafr el-Sheikh) were randomly selected and all the buffaloes that resided on these farms from 21 February 2015 to 20 February 2016 were included in the study. The sampling approach was target-based where all the buffaloes were examined and screened for clinical signs of *Fasciola* infection. All suspected buffaloes were then subjected to fecal examination, and those positive for *Fasciola* eggs underwent antibody testing using indirect hemagglutination test. Consequently, data on 3,356 buffaloes from 29 farms in these governorates was analyzed using a multiple logistic regression model. The final model showed that the age and body condition score of the buffalo, location and type of the farm, application of prophylactic treatment, and temperature and relative humidity of the farm's location significantly affected the rate of infection. The highest prevalence was observed in buffaloes from Alexandria governorate (19.6%), followed by Beheira and Kafr el-Sheikh governorates (15.5 and 9.1%, respectively).

KEY WORDS: buffalo, cross-sectional study, Egypt, Fasciola hepatica, logistic regression

J. Vet. Med. Sci. 80(1): 28–35, 2018 doi: 10.1292/jvms.17-0282

Received: 26 May 2017 Accepted: 1 November 2017 Published online in J-STAGE: 20 November 2017

Fasciolosis is a global zoonotic disease caused by infection with the food- and water-borne trematodes, *Fasciola hepatica* (*F. hepatica*) and *Fasciola gigantica* (*F. gigantica*). The common liver fluke, *F. hepatica*, is widely distributed worldwide. It is found chiefly in cattle, sheep, goats, and buffaloes, but it also affects humans. It is estimated that over 17 million people are infected worldwide, and a further 180 million people in endemic areas are at risk of infection [8]. Human infection typically occurs via the consumption of aquatic vegetables such as watercress, or water contaminated with encysted cercariae. In Egypt, fasciolosis is a serious public health and economic issue consistently affecting human and animal health, and livestock production particularly cattle, buffalo, and sheep [32]. The presence of both, *F. hepatica* and *F. gigantica*, has been confirmed in local cattle and buffaloes [25].

Estimating the prevalence of fasciolosis and identifying the associated risk factors of *F. hepatica* infection will provide useful information in establishing effective preventive and control measures against this majorly neglected tropical disease. Fasciolosis in production animals in different regions of Egypt have been widely studied and the estimated prevalence varied greatly from 0.2–33.7% [2, 10, 11, 17, 27]. Various risk factors such as the management system, pasture management, and climatic/environmental factors like temperature, rainfall, and soil type have been identified in terms of *F. hepatica* infection in farm animals [13, 19, 31, 36]. However, information regarding the risk factors associated with *F. hepatica* infection in the domestic Asian water buffalo (*Bubalus bubalis*) in Egypt is very limited. Therefore, the aim of this study is to estimate the prevalence of *F. hepatica* infection in water buffalo in three governorates of the Nile Delta in Egypt and to identify the underlying risk factors associated with the infection.

*Correspondence to: Sugiura, K.: aksugiur@mail.ecc.u-tokyo.ac.jp ©2018 The Japanese Society of Veterinary Science



This is an open-access article distributed under the terms of the Creative Commons Attribution Non-Commercial No Derivatives (by-nc-nd) License. (CC-BY-NC-ND 4.0: https://creativecommons.org/licenses/by-nc-nd/4.0/)

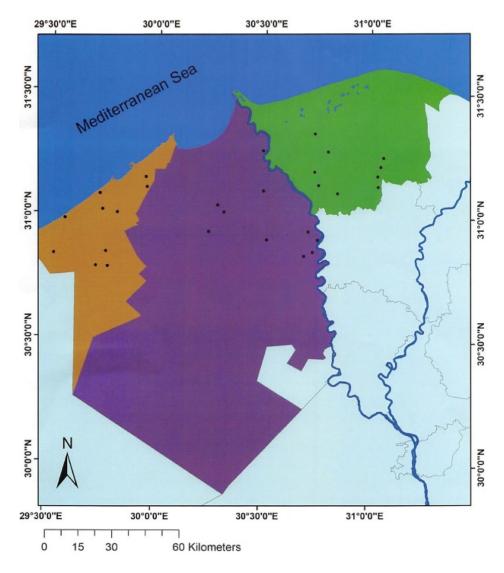


Fig. 1. Geographical distribution of the 29 farms (black dots) included in the current study in Alexandria (brown), Beheira (purple) and Kafr el-Sheikh (green) governorates in the Nile Delta, Egypt.

MATERIALS AND METHODS

Study population

The study population comprised animals from three governorates (provinces), Kafr el-Sheikh, Beheira, and Alexandria, in the Nile Delta of Egypt, where more than 75% of the Egyptian water buffaloes are farmed. A total of 29 farms, including 10 in Alexandria, 10 in Beheira, and 9 in Kafr el-Sheikh, were randomly selected for this study from an exhaustive list of buffalo farms (45 farms in total) provided by the Egyptian government (Fig. 1). Three farm owners declined to participate in the study and were replaced by three other farms from the list. In the selected governorates, female and male buffaloes were usually kept together for dairy and meat production, and *Fasciola* infection constituted a major economic problem that caused declines in productivity. To mitigate the potential economic loss, it was a common for farmers, particularly those on organic farms, to provide prophylactic treatment to the buffaloes using anthelmintic drugs such as triclabendazole, rafoxanid, and/or albendazole.

Case definition and sampling method

All the buffaloes (n=3,356) that resided on the 29 selected farms from 21 February 2015 to 20 February 2016 were included in the current study. Buffaloes that were born, moved out of the farm, or had died within the one-year study period were excluded. Local veterinarians, who assisted with the current study, visited each of the 29 farms from 26 February 2016 to 28 April 2016 and collected all the necessary samples and data. The sampling approach was target-based i.e., all the buffaloes were examined and screened for clinical signs of *Fasciola* infection including emaciation, rough coat, persistent poor appetite, weight loss and/ or diarrhea, icterus of the conjunctiva and/or vulva, decreased milk production, bitter taste of the milk, and/or anestrous features.

All the suspected buffaloes were then subjected to fecal examination and those positive for *Fasciola* eggs underwent antibody testing using the indirect hemagglutination test (IHAT) to diagnose *F. hepatica* infection. A total of 984 buffaloes, suspected of *Fasciola* infection, were subjected to coprological sampling with an average of 34 buffaloes (range, 23–50) being sampled from each farm. Three grams of fecal sample obtained from each buffalo was transferred to clean jugs protected with 10% formalin and fecal examination was performed utilizing the sedimentation strategy according to Hansen *et al.* [16]. The confirmation test for *F. hepatica* antibody in the serum samples was conducted using the Distomiasis Fumouze® IHAT kit (Fumouze Diagnostics, Strasbourg, France) based on the method detailed by Nossair *et al.* [29]: Approximately 5 ml of blood was collected aseptically in sterile tubes from each suspected buffalo. The blood samples were left to clot for 30 min at room temperature and centrifuged at 3,000 rpm for 15 min. The serum was then aspirated, labeled, and kept at -20° C until examination. This method was chosen owing to several advantages such as accuracy, short testing time, ready-to-use capability, and high sample stability. Sensitized red blood cells were composed of sheep red blood cells coated with an *F. hepatica* antigen. Serum antibodies against *F. hepatica* were revealed by an agglutination of the sensitized red blood cells i.e., a reddish-brown film was observed in the well. These red blood cells ensured the reaction specificity and eliminated interferences due to natural anti-sheep agglutinins (Forssman heteroantibodies, infectious mononucleosis antibodies).

Data collection

The following factors (independent variables) were considered for the logistic regression model (see section 2.4) to determine *F*. *hepatica* infection (dependent variable):

Age and sex of the buffalo: Information was collected by the attending veterinarians on the day of the visit.

Body condition (good, medium, or poor): Each buffalo was evaluated for its body condition score by the attending veterinarian according to Nicholson *et al.* [28]. The evaluation was based on a manual assessment of the thickness of fat cover and prominence of bone at the tail. Buffaloes with scores 3 and 4 (the short ribs could be felt by applying slight pressure; the overhanging shelf-like appearance of these bones had disappeared; the backbone was a rounded ridge; and the hook and pin bones were round and smoothed over) were considered to be in good body condition, buffaloes with a score of 2 (the ends of the short ribs could be felt, but they and the individual vertebrae were less visibly prominent; the short ribs did not form an obvious overhang or shelf effect; the hook and pin bones were prominent, but the depression of the thurl region between them was less severe) were considered to be in medium body condition, and buffaloes with scores of 1 (the ends of the short ribs were sharp to the touch and together give a prominent shelf-like appearance to the loin; the individual vertebrae of the backbone were prominent; the hook and pin bones were apparent over the ribs, thurl, and thighs) were considered to be in poor body condition.

Type of farm (organic or conventional): The farms that satisfied the criteria as per Sorge *et al.* [33] were considered as organic. Briefly, the criteria for organic production included a ban on the use of artificial fertilizer, grazing of animals, restriction on the use of concentrated feed, organic production of concentrated and roughage feed, and ban or restriction on the use of antibiotics and other hormones. The number of conventional and organic farms in each governorate was as follows: 8 conventional and 2 organic farms in Alexandria governorate, 6 conventional and 4 organic farms in Beheira governorate, and 3 conventional and 6 organic farms in Kafr el-Sheikh governorate.

Age and education level of the farmer (basic or higher): This information was collected by the attending veterinarian via face-toface interview. Farmers who attended high school and elementary school were considered to have basic education, whereas those that attended university were considered to have higher education.

Application of prophylactic treatment: Buffaloes that received at least one dose of the anthelmintic drugs, triclabendazole, rafoxanide (Rameda Pharmaceuticals, Cairo, Egypt), and/or albendazole (Arabcomed, Obour, Egypt), for prophylactic purposes during the 12-month study period were considered to have received prophylactic treatment. The prophylactic treatment was provided on all the conventional farms and two organic farms within the Alexandria governorate in anticipation of heavy infestation on these farms.

Temperature (°*C*) *and relative humidity* (%): The Nile Delta has a hot desert climate as the rest of Egypt, although its northernmost part has relatively moderate temperatures with an average not surpassing 31°C during the summer. Annual rainfall averages only 100–200 mm, most of which falls during the winter season. Average annual temperature and relative humidity were calculated for the 12-month study period based on monthly data of the respective regions where the farms were situated using Egyptian Meteorological Agency websites (http://www.wunderground.com and freemeteo.com).

Statistical analysis

Odds ratios for infection with *F. hepatica* were estimated using logistic regression analysis. We first conducted a univariate analysis to check for an association between the dependent variable (*F. hepatica* infection) and independent variables (potential risk factors), and only selected those that affected the dependent variable significantly (P<0.05). All independent variables (listed in the previous section) that passed this first screening were considered for the multiple logistic regression model. We used the Hosmer-Lemeshow goodness-of-fit test to decide the linear or categorical characteristic of the independent variable, and thus the farmer's age was excluded as a variable due to the poor fit with the final model (P=0.23). The significance level of the final model was set as 0.05. All statistical analyses were performed using SPSS ver 24.0 (IBM, Armonk, NY, U.S.A.).

Governorate	Study population (<i>n</i>)	Number of buffaloes tested (proportion of the study population, %)	Number of buffaloes positive for fecal examination and <i>F. hepatica</i> antibody ^{a)}	Within-herd prevalence (%)	95% confidence interval ^{b)} (%)
Alexandria	85	23 (27)	11	12.9	5.8-20.1
	96	28 (29)	10	10.4	4.3-16.5
	110	28 (25)	12	10.9	5.1-16.7
	80	28 (35)	13	16.3	8.2-24.3
	113	33 (29)	18	15.9	9.2-22.7
	125	34 (27)	19	15.2	8.9-21.5
	148	35 (24)	11	7.4	3.2-11.7
	111	42 (38)	13	11.7	5.7-17.7
	133	43 (32)	15	11.3	5.9-16.7
	90	45 (50)	18	20.0	11.7-28.3
Beheira	106	23 (22)	15	14.2	7.5-20.8
	144	26 (18)	15	10.4	5.4-15.4
	150	27 (18)	17	11.3	6.3-16.4
	133	33 (25)	17	12.8	7.1–18.5
	125	33 (26)	11	8.8	3.8-13.8
	120	33 (28)	14	11.7	5.9-17.4
	98	34 (35)	19	19.4	11.6-27.2
	114	40 (35)	13	11.4	5.6-17.2
	124	40 (32)	20	16.1	9.7-22.6
	120	43 (36)	17	14.2	7.9–20.4
Kafr el-Sheikh	100	22 (22)	13	13.0	6.4–19.6
	131	27 (21)	16	12.2	6.6-17.8
	107	29 (27)	12	11.2	5.2-17.2
	93	31 (33)	13	14.0	6.9-21.0
	122	33 (27)	16	13.1	7.1–19.1
	118	36 (31)	14	11.9	6.0-17.7
	83	41 (49)	15	18.1	9.8-26.4
	133	44 (33)	16	12.0	6.5-17.6
	144	50 (35)	19	13.2	7.7–18.7
Total	3,356	984 (29)	432	12.9	11.7-14.0

 Table 1. Number of buffaloes sampled on each farm and estimated within-herd prevalence of *F. hepatica* on the 29 farms included in the current study

a) All the 432 buffaloes positive for *Fasciola* eggs during the fecal examination were also positive for the *F. hepatica* antibody during the indirect hemagglutination test. b) The 95% confidence interval of prevalence (*Pre*) is given by: $Pre \pm 1.96 \times \sqrt{Pre \times (1 - Pre)/n}$

RESULTS

The within-herd prevalence of F. hepatica infection ranged from 7.4–20% in the 29 farms included in the current study, giving an overall prevalence of 12.9% (95% confidence interval (CI): 11.7-14%) in 3,356 individual buffaloes (Table 1). Further summary statistics of the study buffalo population is presented in Table 2 and the results of the multiple logistic regression model are shown in Table 3. The location of the farm had a significant effect on F. hepatica infection; buffaloes in Beheira and Kafr el-Sheikh governorates had lower risks of infection than those in Alexandria governorate with odds ratios of 0.36 (95% CI: 0.24–0.54) and 0.37 (95% CI: 0.26–0.54), respectively. In terms of individual factors, older buffaloes were less likely to be infected than the younger ones, with an odds ratio of 0.73 (95% CI: 0.65–0.82), and sex was not a significant risk factor. Buffaloes that received prophylactic treatment had a lower risk of infection than those that did not, with an odds ratio of 0.41 (95% CI: 0.28–0.59). In terms of management systems, buffaloes raised on organic farms had a lower risk of infection than those on conventional farms, with an odds ratio of 0.41 (95% CI: 0.28–0.59). In terms of climatic conditions, buffaloes raised in regions with annual average temperatures lower than 26°C and between 26 and 30°C had higher risks of infection than those raised in regions higher than 30°C, with odds ratios of 20.96 (95% CI: 13.42–32.74) and 6.49 (95% CI: 4.23–9.97), respectively. Moreover, buffaloes raised in regions with an annual average relative humidity (RH) lower than 50% and between 50% and 59% had higher risks of infection than those raised in regions with an RH higher than 59%, with odds ratios of 4.63 (95% CI: 3.12-6.85) and 3.21 (95% CI: 2.26-4.54), respectively. Finally, buffaloes with poor and medium body condition scores were more likely to be associated with infection than those with good body condition scores, with odds ratios of 3.71 (95% CI: 2.59–5.33) and 4.95 (95% CI: 3.30–7.43), respectively.

	Total population	Number of buffaloes tested in the population	Number of buffaloes that exhibited <i>F. hepatica</i> infection (prevalence in %)
Total	3,356	984	432 (12.8)
Sex			
Male	1,810	350	321 (17.7)
Female	1,546	634	111 (7.2)
Age (years)			
1	852	355	152 (17.8)
2	1,024	385	168 (16.4)
3	537	128	47 (8.8)
4	224	82	46 (20.5)
5	719	34	19 (2.6)
Governorates			
Alexandria	685	339	134 (19.6)
Beheira	860	332	133 (15.5)
Kafr El-Sheikh	1,811	313	165 (9.1)
Гуре of farm			
Conventional	1,159	534	384 (33.1)
Organic	2,197	450	48 (2.2)
Body condition score			
Good	2,281	290	195 (8.6)
Medium	309	344	108 (35.0)
Poor	766	350	129 (16.8)
Application of prophylactic tre	eatment		
Not applied	2,168	574	359 (16.6)
Applied	1,188	410	73 (6.1)
Age of farmer (years)			
<40	620	234	65 (10.5)
40–49	1,525	398	222 (14.6)
>49	1,211	352	145 (12.0)
Education of farmer			
Basic	2,572	660	407 (15.8)
Higher	784	324	25 (3.2)
Relative humidity (%)			
<50	596	288	123 (20.6)
50-59	509	374	159 (31.2)
>59	2,251	322	150 (6.7)
Temperature (°C)			
<26	494	291	216 (43.7)
26-30	872	321	164 (18.8)
>30	1,990	372	520 (2.6)

Table 2. Summary statistics of the buffalo population included in the current study

DISCUSSION

Fasciolosis is one of the most common parasitic diseases severely affecting public health and animal production, especially in developing countries such as Egypt. In terms of cattle production, economic losses associated with fasciolosis includes direct losses from condemnation of the liver and reduced carcass weight at slaughter [9]. Indirect losses in the form of poor feed conversion, weight loss, slow fattening, reduced milk yield, reproductive failure, and sometimes death also affect cattle production [4, 7]. The current study significantly associated *F. hepatica* infection with buffaloes having medium and poor body condition scores when compared to those with good body condition scores, which is consistent with previous reports on fasciolosis in cattle [5, 23].

The present study showed that the prevalence of *F. hepatica* infection in buffaloes in three governorates in the Nile Delta of Egypt was 12.87%. National level surveys in the late 1990s reported an infection prevalence of 3.5% for the whole country. Meanwhile, in the southern part of Egypt, the infection prevalence in the Assiut governorate (located about 400 km to the south of Cairo) and the Qena governorate (located about 600 km to the south of Cairo) was estimated to be 24.3 and 33.7%, respectively [2, 17, 27]. Our results revealed that the infection prevalence in Alexandria governorate (19.56%) was higher than that in Beheira (15.47%) and Kafr el-Sheikh (9.11%) governorates, which could be explained by the differences in the climatic/environmental conditions such as rainfall and/or the management systems (i.e., the scale of the farm and grazing habits of the cattle) between these governorates [6]. The farms included in the current study in Alexandria governorate were either situated close to the

Variable	Coefficient	Odds ratio	95% CI	P-value
Constant	-2.84	-		< 0.0001
Age (years)	-0.32	0.73	0.65-0.82	< 0.0001
Sex				
Male (reference)	0.00	1.00		
Female	0.102	1.11	0.79-1.55	0.55
Governorate				
Alexandria (reference)	0.00	1.00		
Beheira	-1.03	0.36	0.24-0.54	< 0.0001
Kafr el-Sheikh	-0.99	0.37	0.26-0.54	< 0.0001
Body condition score				
Good (reference)	0.00	1.00		
Medium	1.60	4.95	3.30-7.43	< 0.0001
Poor	1.31	3.71	2.59-5.33	< 0.0001
Prophylactic treatment				
Not applied (reference)	0.00	1.00		
Applied	-0.89	0.41	0.28-0.59	< 0.0001
Type of farm				
Conventional	0.00	1.00		
(reference)				
Organic	-0.89	0.41	0.28-0.59	< 0.0001
Education of farmer				
Basic (reference)	0.00	1.00		
Higher	-0.42	0.66	0.37 - 1.77	0.15
Temperature				
>30°C (reference)	0.00	1.00		
26–30°C	1.87	6.49	4.23-9.97	< 0.0001
<26°C	3.04	20.96	13.42-32.74	< 0.0001
Relative humidity				
>59% (reference)	0.00	1.00		
50-59%	1.17	3.21	2.26-4.54	< 0.0001
<50%	1.53	4.63	3.12-6.85	< 0.0001

Table 3. Association of age, sex, body condition score and underlying risk factors with *F. hepatica* infection predicted by multiple logistic regression for the study buffalo population in three governorates in the Nile Delta of Egypt

Model statistics: χ^2 (13)=1,290.016, P≤0.0001; Nagelkerke R²=0.595; Hosmer-Lemeshow test: P=0.709.

Mediterranean Sea or in areas filled with water channels having poor water management programs, all of which possibly promoted the development and spread of the intermediate snail hosts of fasciolosis.

Our estimated value for infection prevalence in Alexandria governorate appeared to be higher than the previously reported 13% by El-Sherif *et al.* [12], possibly due to the increase in contaminated vegetated areas in the governorate over the last 50 years. Contrastingly, most of the water channels running through Beheira and Kafr el-Sheikh governorates were periodically disinfected when compared to the water channels running through several areas in the Alexandria governorate.

In terms of management factors, we demonstrated that buffaloes raised on conventional farms had a higher risk of infection than those on organic farms, possibly due to poorer grassland management on conventional farms where buffaloes had easier access to contaminated pastures. However, this finding differed from the findings of Kantzoora et *al.* [19] which reported no significant difference in the prevalence of ovine fasciolosis between organic and conventional farms in Greece.

Our results suggested that sex was not a significant risk factor, which was consistent with several previous studies [18, 20–22, 30]. However, Yildirim *et al.* [36] reported that bovine fasciolosis in Turkey was more prevalent in females (70.7%) than in males (47.8%). The higher risk among females could be attributed to the fact that most female cattle are kept for breeding and milk production purposes, thereby increasing stress and reducing their resistance to infections. Furthermore, we demonstrated age to be a relevant risk factor for *F. hepatica* infection in buffaloes. This result contrasted the findings of Hansen *et al.* [13], which showed no significant difference between the prevalence of bovine fasciolosis in Ethiopia and young animals (33.3%), old animals (26.11%), or adult animals (24.7%). Nicholson *et al.* [15] also concluded that age was not a risk factor for bovine fasciolosis in Northern Ethiopia.

In terms of climatic conditions, our analysis revealed that temperatures lower than 30° C combined with a relative humidity (RH) lower than 60% had positive effects on fasciolosis in buffaloes. Abdelhamid *et al.* [1] investigated the prevalence of ovine fasciolosis in the Beheira governorate at different temperatures and RH, but found no significant difference between the following three conditions: group 1, temperatures <25°C and RH of 72.8%; group 2, temperatures between 25 and 30°C and RH of 70.25%;

and group 3, temperatures >30°C and RH of 68.96%. Temperature alone has also been proven as a consistent risk predictor. Torgerson *et al.* [34] and Urquhart *et al.* [35] demonstrated 10°C to be the minimum critical temperature for the development of *F. hepatica* eggs and the infection of the intermediate snail host, while Augot *et al.* [3] and Lee *et al.* [24] mentioned that the optimal temperature for metacercariae production in infected snails was between 22 and 25°C. More recently, Moayad *et al.* [26] revealed that the optimal temperature for metacercarial production in *F. gigantica*-infected snails was 25 ± 1 °C.

Finally, our analysis highlighted that buffaloes that did not receive prophylactic treatment were at a higher risk of fasciolosis than those that did. Likewise, Kantzoora *et al.* [19] also found a negative association between the use of anthelmintic treatment and the occurrence of *F. hepatica* infection (in terms of coproantigen detection) on sheep and goat farms. Given the fact that fasciolosis in cattle is mostly subclinical, the use of prophylactic treatment appears to be the most cost-effective control strategy in a developing country setting like Egypt.

The evidence-based information provided in this study could assist local veterinarians in targeting farms having high-risk factors such as conventional farms situated in Alexandria with an average temperature lower than 26°C, and in advising farmers to adopt a prophylactic strategy when necessary. However, anthelmintic resistance is a growing global problem and its development in livestock animals should not be overlooked [14].

A cross-sectional study was conducted to estimate the prevalence of *F. hepatica* infection and to identify its underlying risk factors in three governorates in the Nile Delta of Egypt. We found that the prevalence of *F. hepatica* infection was significantly associated with the location and type of the farm, age and body condition of the buffalo, the application of anthelmintic treatment, temperature, and relative humidity. These results are fundamental in formulating appropriate control strategies against bovine fasciolosis in Egypt and other areas with similar farming practices and climatic conditions.

DECLARATION OF INTERESTS. The authors declare that they have no competing interests.

ACKNOWLEDGMENTS. The authors would like to thank the veterinarians in Alexandria, Beheira, and Kafr el-Sheikh governorates who assisted in data collection for this study.

REFERENCES

- 1. Abdelhamid, M. S. 2009. Zoonotic importance of fascioliasis in sheep and human, Master Thesis, Faculty of Veterinary Medicine, Department of Animal Hygiene and Zoonosis. Menofia University, Egypt.
- Abdel-Nasser, A., Hussein, R. and Khalifa, M. A. 2010. Fascioliasis prevalences among animals and human in Upper Egypt. J. King. Saud. Univ. Sci. 22: 15–19. [CrossRef]
- Augot, D., Rondelaud, D., Dreyfuss, G., Cabaret, J., Bayssade-Dufour, C. and Albaret, J. L. 1998. Characterization of Fasciola hepatica redial generations by morphometry and chaetotaxy under experimental conditions. J. Helminthol. 72: 193–198. [Medline] [CrossRef]
- 4. Ayana, D., Gebreab, F. and Sori, H. 2009. Prevalence of Ovine and Caprine Fasciolosis in and around Assela Town of Oromia Regional State, Ethiopia. *Bull. Anim. Health Prod. Afr.* **57**: 109–116.
- 5. Bekele, M., Haftom, T. and Yehenew, G. 2010. Bovine fascioliosis: Prevalence and its economic loss due to liver condemnation at Adwa municipal abattoir, North Ethiopia. *Ethiop. J. Agri. Sci. Tech.* **1**: 39–47.
- Bennema, S. C., Ducheyne, E., Vercruysse, J., Claerebout, E., Hendrickx, G. and Charlier, J. 2011. Relative importance of management, meteorological and environmental factors in the spatial distribution of Fasciola hepatica in dairy cattle in a temperate climate zone. *Int. J. Parasitol.* 41: 225–233. [Medline] [CrossRef]
- 7. Biu, A., Paul, B., Konto, M. and Ya'Uba, A. 2013. Cross sectional and phenotypic studies on fasciolosis in slaughter cattle in Maiduguri. J. Agri. Vet. Sci. 5: 155–162.
- 8. Cwiklinski, K., O'Neill, S. M., Donnelly, S. and Dalton, J. P. 2016. A prospective view of animal and human Fasciolosis. *Parasite Immunol.* **38**: 558–568. [Medline] [CrossRef]
- 9. Degheidy, N. and Al-Malki, J. 2012. Epidemiological Studies of Fasciolosis in Human and Animals at Taif, Saudi Arabia. *World Appl. Sci. J.* 19: 1099–1104.
- Elmonir, W., Mousa, W. and Sultan, K. 2015. The prevalence of some parasitic zoonoses in different slaughtered animal species at Abattoir in the mid-delta of Egypt; with special reference to its economic implications. *Alex. J. Vet. Sci.* 47: 97–103.
- 11. el-Shazly, A. M., el-Wafa, S. A., Haridy, F. M., Soliman, M., Rifaat, M. M. and Morsy, T. A. 2002. Fascioliasis among live and slaugthered animals in nine centers of Dakahlia Governorate. J. Egypt. Soc. Parasitol. **32**: 47–57. [Medline]
- 12. El-Sherif, A. F., Abdou, A. H. and El-Sawi, M. F. 1959. The incidence of parasitic infestation among the farm animals of Faculty of Agriculture, University of Egypt. *Vet. Med. Assoc.* 19: 19–21.
- 13. Ephrem, B., Wassie, M. and Abadi, A. 2012. Prevalence and economic losses of bovine fasciolosis in dessie Municipal Abattoir, South Wollo Zone, Ethiopia. *Eur. J. Biol. Sci.* **4**: 53–59.
- 14. Furtado, L. F. V., de Paiva Bello, A. C. and Rabelo, E. M. L. 2016. Benzimidazole resistance in helminths: from problem to diagnosis. *Acta Trop.* **162**: 95–102. [Medline] [CrossRef]
- 15. Girmay, T., Teshome, Z. and Hailemikael, A. 2015. Prevalence and economic losses of bovine fasciolosis at Hawzien Abattoir, Tigray Region. Northern Ethiopia. J. Vet. Adv. 5: 945–951.
- 16. Hansen, J. and Perry, B. 1994. The epidemiology, diagnosis and control of helminth parasites of ruminants. A handbook, Food and Agricultural Organization of the United Nations, Rome.
- 17. Haridy, F. M., Ibrahim, B. B., Morsy, T. A. and El-Sharkawy, I. M. 1999. Fascioliasis an increasing zoonotic disease in Egypt. J. Egypt. Soc. Parasitol. 29: 35–48. [Medline]
- Kabir, M. H., Eliyas, M., Hashem, M. A. and Miazi, O. F. 2010. Prevalence of zoonotic parasitic diseases of domestic animals in different abattoir of Comilla and Brahman Baria region in Bangladesh. J. Zool. (Lond.) 28: 21–25.

- Kantzoura, V., Kouam, M. K., Demiris, N., Feidas, H. and Theodoropoulos, G. 2011. Risk factors and geospatial modelling for the presence of Fasciola hepatica infection in sheep and goat farms in the Greek temperate Mediterranean environment. *Parasitology* 138: 926–938. [Medline] [CrossRef]
- Kanyari, P. W., Kagira, J. M. and Mhoma, R. J. 2010. Prevalence of endoparasites in cattle with zoonotic potential within urban and peri-urban areas of Lake Victoria Basin, Kenya. J. Anim. Biomed. Sci. 4: 26–33.
- 21. Keyyu, J. D., Monrad, J., Kyvsgaard, N. C. and Kassuku, A. A. 2005. Epidemiology of Fasciola gigantica and amphistomes in cattle on traditional, small-scale dairy and large-scale dairy farms in the southern highlands of Tanzania. *Trop. Anim. Health Prod.* **37**: 303–314. [Medline] [CrossRef]
- 22. Khan, M. K., Sajid, M. S., Khan, M. N., Iqbal, Z. and Iqbal, M. U. 2009. Bovine fasciolosis: prevalence, effects of treatment on productivity and cost benefit analysis in five districts of Punjab, Pakistan. *Res. Vet. Sci.* 87: 70–75. [Medline] [CrossRef]
- 23. Kheider, Z. A. 2014. Prevalence and risk factors of bovine fasciolosis in North Kordofan State, Sudan, M.Sc., College of Veterinary Medicine, University of Khartoum, Khartoum.
- Lee, C. G., Cho, S. H. and Lee, C. Y. 1995. Metacercarial production of Lymnaea viridis experimentally infected with Fasciola hepatica. *Vet. Parasitol.* 58: 313–318. [Medline] [CrossRef]
- Lotfy, W. M., El-Morshedy, H. N., Abou El-Hoda, M., El-Tawila, M. M., Omar, E. A. and Farag, H. F. 2002. Identification of the Egyptian species of Fasciola. *Vet. Parasitol.* 103: 323–332. [Medline] [CrossRef]
- Moayad, M., Al-jibouri, S., Al-Mayah, H. and Hadi, R. H. 2011. The factors affecting metacercarial production of Fasciola gigantica from Lymnaeaauricularia snails. J. Basr. Res. 37: 9–16.
- 27. Monib, M. E. M. 1977. Study on some helminth parasites of ruminants in Assiut Governorate, M.Sc. Thesis, Faculty of Veterinary Medicine, Assiut University, Egypt.
- 28. Nicholson, M. J. and Butterworth, M. H. 1986. A guide to condition scoring of zebu cattle. International Livestock Center for Africa, Addis Ababa, Ethiopia.
- 29. Nossair, M. A. and Abdella, D. E. 2014. Serological detection of Fasciola hepatica antibodies among cattle and human in Behera Province, West Delta, Egypt. *Alex. J. Vet. Sci.* **40**: 16–23.
- 30. Phiri, A. M., Phiri, I. K., Sikasunge, C. S. and Monrad, J. 2005. Prevalence of fasciolosis in Zambian cattle observed at selected abattoirs with emphasis on age, sex and origin. J. Vet. Med. B Infect. Dis. Vet. Public Health **52**: 414–416. [Medline] [CrossRef]
- 31. Selemetas, N., Phelan, P., O'Kiely, P. and Waal, T. 2014. Weather and soil type affect incidence of fasciolosis in dairy cow herds. *Vet. Rec.* **175**: 371–375. [Medline] [CrossRef]
- 32. Soliman, M. F. 2008. Epidemiological review of human and animal fascioliasis in Egypt. *J. Infect. Dev. Ctries.* **2**: 182–189. [Medline] [CrossRef] 33. Sorge, U. S. R., Moon, R., Wolff, L. J., Michels, L., Schroth, S., Kelton, D. F. and Heins, B. 2016. Management practices on organic and
- conventional dairy herds in Minnesota. J. Dairy Sci. 99: 3183-3192. [Medline] [CrossRef]
- 34. Torgerson, P. and Claxton, J. 1999. Epidemiology and control. In: Fascioliosisb (Dalton, J. P.Eds), CABI Publishing, Oxfordshire.
- 35. Urquhart, G. M., Armour, J., Duncan, J. L., Dunn, A. M. and Jennings, F. W. 1996. Veterinary Parasitology, 2nd ed. Longman Scientific and Technical Press, Oxford.
- 36. Yildirim, A., Duzlu, O. and Inci, A. 2007. Prevalence and risk factors associated with *Fasciola hepatica* in cattle from Kayseri province, Turkey. *Revue. Méd. Vét.* **158**: 613–617.