

# The Crosstalk Between Cancer Cells and Neutrophils Enhances Hepatocellular Carcinoma Metastasis via Neutrophil Extracellular Traps-Associated Cathepsin G Component: A Potential Therapeutic Target

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**Background:** Emerging evidences have highlighted the roles of neutrophils, as the major host microenvironment component, in the development of hepatocellular carcinoma (HCC). Neutrophils extracellular traps (NETs) produced in the infection can strengthen the behavior of cancer metastasis. Here, we investigated the roles of NETs in HCC metastasis and further explore the underlying mechanism of how NETs interact with cancer.

**Methods:** The neutrophils were isolated from whole blood of HCC patients and used to evaluate the formation of NETs. NET markers were detected in tissue samples, plasma and cell climbing slice. Mouse models were used to evaluate the roles of NETs in HCC metastasis in vivo, and the corresponding mechanisms were explored using in vivo and in vitro assays.

**Results:** An increase in the release of NETs in patients with HCC, particularly those with portal vein tumor thrombosis (PVTT). The presence of NETs in HCC tumor tissues closely correlated with a poor prognosis. Functionally, the invasion ability of HCC cells was enhanced by co-culture with HCC neutrophils, through NETs formation, while the neutrophils from a healthy donor (HD) exhibited the inhibition of the invasion ability. Furthermore, we observed an enhanced ability of forming NETs in neutrophils from HCC patients in vitro, especially patients with PVTT or extra-hepatic metastasis. An in-vivo animal study demonstrated that neutrophils of HCC facilitated the metastatic behavior towards the lung. The further mechanistic investigation unveiled that HCC cells-derived cytokine IL-8 triggered NETs formation in an NADPH oxidase-dependent manner, and NETs-associated cathepsin G (cG) promoted HCC metastasis in vitro as well as vivo. Clinically, the expression of the cG protein in tumor tissues displayed a close correlation with the disease prognosis of HCC patients.

**Conclusion:** Our findings implicated that the induction of NETs by HCC cells is a critical metastasis-supporting cancer–host interaction and that NETs may serve as an immune-based potential therapeutic target against HCC progression.

**Keywords:** hepatocellular carcinoma, neutrophils extracellular traps, cathepsin G, IL-8, E-cadherin, metastasis, NADPH

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## Introduction

Despite the significant progress has been made in its diagnosis and treatment in recent years, hepatocellular carcinoma (HCC) remains one of the leading causes of cancer-related death worldwide.<sup>4,6,7</sup> Systemic therapies for HCC, such as surgical

resection, transarterial chemoembolization, local ablation, and targeted molecular therapy, have markedly advanced; however, the survival rate remains low, mainly due to distant metastasis.<sup>12,35</sup> Thus, a better understanding of the molecular mechanism underlying HCC metastasis is urgently required.

The inflammatory tumor microenvironment is crucial in the development of HCC.<sup>11,12</sup> Single-cell sequencing analyses have revealed a landscape of infiltrating T cells and their functional properties in HCC tissues.<sup>45</sup> Myeloid derived suppressor cells (MDSCs) have been demonstrated to accumulate in several animal models of HCC.<sup>27,28</sup> Tumor-infiltrating leukocytes represent a significant proportion in the inflammation microenvironment, and influence nearly all steps in the progression of malignancies, including metastasis.<sup>13,14,17</sup> As a critical element among leukocytes, neutrophils serve as a paradigm for the interaction between inflammation and cancer.<sup>7,9,11,24</sup> The infiltration of tumor-associated neutrophils (TAN) and the elevated neutrophil–lymphocyte ratio are correlated with poor outcomes in HCC.<sup>12,13</sup> Neutrophils exhibit several functions, including phagocytosis, generation of reactive oxygen species, as well as the formation of neutrophils extracellular traps (NETs), which are DNA meshes with associated cytotoxic enzymes released into the extracellular space.<sup>8,18</sup> NETs represent a novel aspect of neutrophil biology and were initially discovered as a part of the process of innate immune defense to kill microorganisms.<sup>46,47</sup>

In the past decades, NETs formation has been closely associated with several cancer-related diseases.<sup>22,23</sup> NETs bridge the crosstalk between cancer cells and neutrophils.<sup>3,5,48</sup> Recent pieces of evidence suggest cancer cells-secreted factors trigger NETs formation, which in turn promotes cancer metastasis by inducing a pro-inflammatory response in tumor cells.<sup>12,40,47</sup> Emerging evidence shows that NETs-associated proteases, such as neutrophil elastase (NE) and matrix metalloproteinase 9, support metastasis colonization in the lung by remodeling the extracellular matrix.<sup>5</sup> Neutrophils also degranulate to release Ser proteases, such as cathepsin G (cG), to degrade thrombospondin-1, thereby enhancing metastatic outgrowth.<sup>20</sup> To date, the effect of NETs, especially the associated proteases, on HCC progression, remains poorly understood.

In the present study, we examined the expression level of NETs-CitH3 in HCC patient samples and assessed the relationship between CitH3 expression and the clinicopathological features with HCC. Furthermore, we explored the ability of cancer cells to induce NETs formation, in turn,

the effect of NETs on HCC metastasis behavior, and also how the NETs-associated cG contributed to this process. To summarize, our study revealed a previously unknown mechanism underlying the crosstalk between HCC and neutrophils, which would exhibit critical clinical implications.

## Patients and Methods

### Neutrophils Isolation

For human neutrophils isolation, EDTA-K<sub>2</sub> anticoagulation vein whole blood samples were obtained preoperatively from patients with HCC at Zhongshan Hospital. Approximately 10 mL of fasting peripheral blood was withdrawn from the selected patients in the morning. The blood sample was placed in the 0.25% ethylenediaminetetraacetic acid (EDTA) anticoagulant inside a tube for neutrophil separation. A double gradient was formed by layering an equal volume of Histopaque (Sigma)-1.077 over 1.119. The whole blood was carefully layered onto an equal volume of double Histopaque gradient (1.077 and 1.119). After centrifugation at room temperature, erythrocytes were aggregated and rapidly sediment. The neutrophils fraction was collected from the 1.077–1.119 interface. The neutrophils were isolated and extracted from the whole blood in less than 2 h.

### Tissue Collection

Eighty-five HCC tissues were obtained from patients with HCC who underwent hepatectomy between 2011 and 2019 at Zhongshan Hospital of Xiamen University. Tissue samples were immediately frozen in liquid N<sub>2</sub> and stored at –80°C for further experiments. Written informed consent for participation in this research was obtained from all patients. The consent and experimental procedures were approved by the ethics committee of Zhongshan Hospital of Xiamen University according to the World Medical Association Declaration of Helsinki.

### Serum NETs Detection

Human serum NETs marker myeloperoxidase-deoxyribonucleic acid (MPO-DNA) was measured using a capture ELISA assay with some modification. In brief, 5 µg/mL MPO monoclonal antibody (1:100, ab4704, Abcam) was coated into a 96-well plate, followed by overnight incubation at 4°C overnight. After blocking with 1% bovine serum albumin (BSA), serum was added into each well, followed by incubation at room temperature for 3 h. After washing four times with phosphate-

buffered saline (PBS), PicoGreen<sup>®</sup> dsDNA Quantitation Reagent was added into each sample according to the manufacturer's instructions, followed by incubation at room temperature for 5 min, protected from light. The values were then measured using instrument parameters with a filter setting of 480 nm/520 nm excitation/emission.

## Immunohistochemistry Staining

Briefly, tissues were fixed with 10% neutral formalin, embedded in paraffin, and then 4  $\mu\text{m}$ -thick sections were prepared. The sections were deparaffinized, hydrated, and soaked in 3%  $\text{H}_2\text{O}_2$  for 1 h, and then blocked with 5% BSA at room temperature overnight. The sections were incubated with anti-histone H3 (citrulline  $\text{R}_2+\text{R}_8+\text{R}_{17}$ , CitH3) (1:500, ab5103, Abcam), anti-Ly6G (1:100, ab25377, Abcam), anti-cathepsin G (1:200, ab192793, Abcam) or anti-IL-8 (1:150, #44133, Signalway Antibody) antibodies at 4°C overnight. The slides were incubated with biotinylated antibodies for 1 h and stained with diaminobenzidine (DAB; Maixin Biotechnology, Fuzhou, China), followed by counterstaining with hematoxylin (Maixin Biotechnology).

## Immunofluorescence

Immunofluorescence assay was performed to determine NETs formation in mice lung frozen sections, or neutrophils on a coverslip. NETs specific marker was stained with a primary antibody against anti-histone H3 (citrulline  $\text{R}_2+\text{R}_8+\text{R}_{17}$ , CitH3) (1:400, ab5103, Abcam). After washing with PBS, the sections were stained with fluorochrome-conjugated secondary antibodies (1:200, AS007, ABclonal) and finally stained with DAPI (Invitrogen, 0.05 mg/mL). All negative controls were designed without the primary antibody. The extension of NETs release was assessed by calculating the area occupied by CitH3 color spot under each confocal microscope visual field. For NETs area quantification, Image J software and the particle analyzer plugin were used. Particle analyzer plugin was used to form the images of CitH3 color spot, and Image J software could calculate the percent of area occupied by CitH3 color.

## Cell Culture

HCC cell lines (HuH7, MHCC-97L and Hepa1-6) were amplified in our laboratory and stored in liquid nitrogen to ensure that the cells used for experiments were passaged for fewer than six months. The authentication of these cell lines was performed through comparisons with the STR

database. The cells were maintained in high glucose Dulbecco's modified eagle medium (DMEM) supplemented with 10% fetal bovine serum and 1% penicillin/streptomycin at 37°C with 5%  $\text{CO}_2$  in the air.

## Transwell Assay

For the transwell assay, approximately  $5 \times 10^5$  species-matched HD or HCC neutrophils were mixed in 500  $\mu\text{L}$  serum-free DMEM medium, seeded onto the bottom chamber, supplemented with 10% fetal bovine serum (FBS), and then incubated for 30 min before the addition of DNaseI (Invitrogen, AM2222), NE inhibitor (MedChemExpress, 150374-95-1) or cG inhibitor (Sigma-Aldrich, 219372). The indicated  $4 \times 10^5$  HCC cells in 200  $\mu\text{L}$  serum-free DMEM medium were seeded onto an 8  $\mu\text{m}$  pore polycarbonate membrane with Matrigel in the upper chamber as per the manufacturer's instructions (Corning Incorporated; New York, NY, USA). To digest the structure of the NETs, 15  $\mu\text{L}$  DNaseI alone was added into the lower chamber culture medium. For PAD4 inhibition with GSK484, isolated neutrophils were pre-treated with the PAD4 inhibitor GSK484 (Cayman Chemical, 17488) at 37°C for 4h before seeding onto the bottom chamber. NETs-CM was added to the bottom chamber, supplemented with 10% FBS, and  $4 \times 10^5$  HCC cells were seeded in a serum-free DMEM medium to the upper chamber. DNaseI or cG inhibitor was added into the conditioned medium (CM) following the previous instruction.

After 24 h of incubation, the non-invasive cells from the upper surface of the membrane were wiped off, while the cells on the bottom surface of the membrane were fixed with 4% paraformaldehyde (PFA) and stained with hematoxylin. The cells in ten randomly selected views were counted.

## Immunoblot Analysis

Cells were lysed with the radioimmunoprecipitation assay (RIPA) lysis buffer (Beyotime; Beijing, China). Protein concentration in the lysates was measured with a BCA Protein Assay Kit (ThermoFisher). Protein samples were separated by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) and then transferred onto polyvinylidene difluoride (PVDF) membranes (Millipore). Membranes were blocked with 5% non-fat milk (Bio-Rad) at room temperature for 1 h, and then incubated with anti-E-cadherin (1:400, #3195, Cell Signaling) or anti-IL-8 (1:100, ab18672, Abcam) at 4°C overnight, followed by incubation with anti-rabbit or anti-

mouse secondary antibodies conjugated to horseradish peroxidase (HRP, Jackson). Immunoreactive proteins were subsequently visualized using the enhanced chemiluminescence detection system (Millipore).

## Preparation of NETs

Neutrophils were isolated and seeded onto 24-well plates ( $2.5 \times 10^5$  cells/well). Phorbol 12-myristate 13-acetate (PMA, 500 nM, Sigma-Aldrich) was added to the isolated neutrophils, followed by incubated at 37°C and 5% CO<sub>2</sub> for 4 hours to induce NETs generation. Then, the supernatants were obtained carefully and washed twice to eliminate residual PMA or NETs-associated substances without disturbing the NETs. The supernatants containing NETs were collected and centrifuged to eliminate cell debris. Isolated NETs were stored at -80°C for subsequent experiments.

## NETs Formation Study

In HCC cells-CM and patients serum-induced NETs formation assays, neutrophils were incubated with the corresponding CM (1:2) for 24h, or incubated with serum from HCC patients or HD for 24h. The NOX2 inhibitor apocynin was added into the culture medium at a final concentration of 10 μM. IL-8 neutralizing antibody was added into culture medium at the concentration of 5 μg/mL. For quantification, NETs on coverslips in 24-well plates were detected by immunofluorescence following the previously described instructions. The extension of NETs release was assessed by calculating the area percent of CitH3 color with Image J software.

## Animal Study

Male BALB/c nude mice (6–8 weeks old, weight 20–25 g) and male C57BL/6J mice (5–6 weeks old, weight 19–23 g) were used in animal studies and fed in the experimental animal center for specific pathogen-free animals at Xiamen University. For establishing the animal model,  $5 \times 10^6$  HuH7 or 97 L cells were injected intravenously into nude mice. A co-injection experiment was performed by injecting  $4 \times 10^6$  HuH7 cells with  $4 \times 10^5$  human neutrophils into nude mice via the tail vein. For male C57BL/6J mice, treatment of DNaseI (50 μL/mouse) or cG inhibitor was given daily via intraperitoneal injection. The dose of the inhibitor was 12 mg/kg (Sigma-Aldrich, 219372). Hepa1–6 cells ( $2 \times 10^6$ ) were intravascularly injected to establish the metastatic model at two days after beginning treatment administration. Metastatic burden was assessed

by calculating the area percentage of lung tissue covered by the tumor. In the H&E-stained histological sections, the tumor-occupied area was quantitatively assessed by Image J software. All animal experiments were approved by the Animal Care and Use Committee of Xiamen University.

## Statistical Analysis

All statistical analyses were performed using GraphPad Prism 6 software. Study data were analyzed using two-sided t-tests or unpaired *t*-test. Pearson's correlation test was used for correlation analysis and log-rank (Mantel-Cox) test was used for follow-up data and prognosis analysis. A P-value less than 0.05 indicated statistical significance.

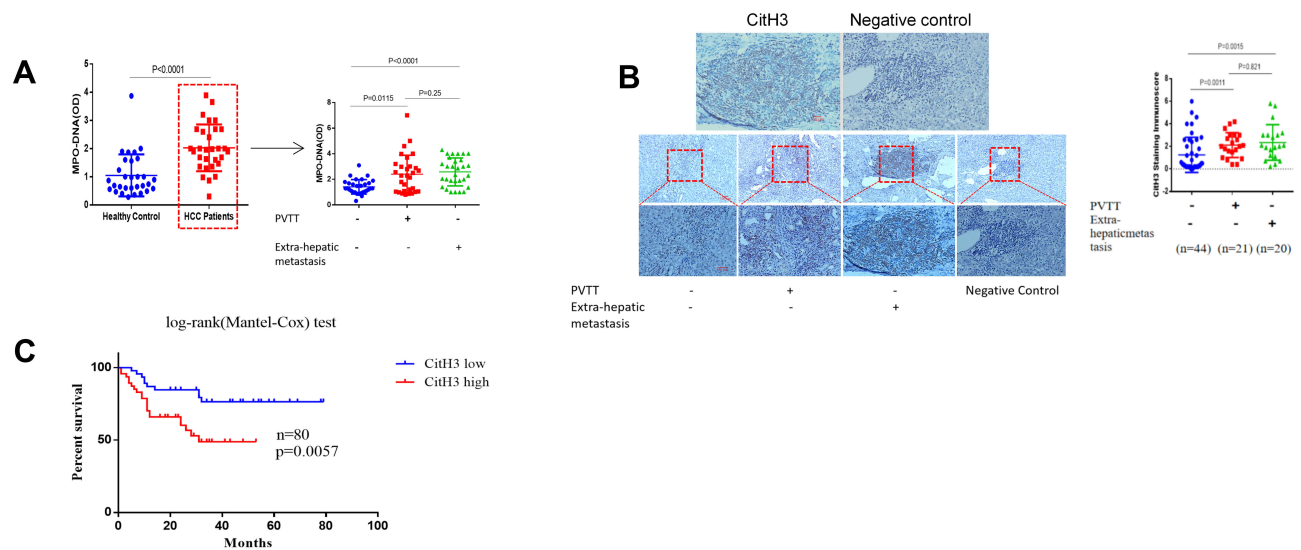
## Determination of the Dose of Treatments in Animal Studies

The dose of DNaseI in vivo was determined according to previously reported studies in the aspect of NETsosis (50 μL/mouse, via intraperitoneal injection). The dose of cG inhibitor in vivo (Sigma-Aldrich, 219372) was determined according to the inhibitor ((MCE, Sivelestat sodium tetrahydrate) of another NETs-carrying neutrophil elastase (NE) protease in vitro and vivo. The dose of NE inhibitor was reported to be 0.00456g/L in vitro, and 0.05mg/g in mouse. Accordingly, when cG inhibitor was reported at 0.00124g/L in vitro, a dose of cG inhibitor at about 0.012mg/g was used in mouse.

## Results

### An Increase in NETs Formation Indicates a Poor Prognosis of HCC Patients Associated with PVTT

When forming NETs, neutrophils release chromatin DNA associated with other neutrophil-derived proteins into extracellular space.<sup>48</sup> We examined the serum MPO-DNA level, a NETs product, of 30 preoperative patients with HCC and 30 healthy controls (HD) (Table S3). Our results suggested that serum MPO-DNA level increased in patients with HCC compared to HD. Among 90 HCC patients, further investigation indicated a remarkably higher level of MPO-DNA in HCC patients with portal vein tumor thrombosis (PVTT) or extra-hepatic metastasis compared to those without metastasis (Figure 1A). Indeed, during NETs formation, histone citrullination induced by peptidyl arginine deiminases (PADs) is an important post-translational modification that facilitates chromatin



**Figure 1** Increased NETs marker in HCC patients. **(A)** The Levels of MPO-DNA in HCC or healthy control serum samples were examined by the capture ELISA assay (n=90). The red dotted box points to HCC patients, indicating that MPO-DNA levels were particularly elevated in either PVTT or extra-hepatic metastasis among HCC patients. **(B)** The deposition of NETs-CitH3 in 85 HCC tissue samples were examined by the immunohistochemistry assay. NETs-CitH3 level elevated in HCC patients tissue with portal vein tumor thrombosis/extra-hepatic metastasis(n=41), compared to those without metastasis(n=44). Representative images of CitH3 in human tissues were shown. Scale bar: 100um. **(C)** Presence of NETs-CitH3 in HCC tissues correlated with disease prognosis. Log-rank (Mantel-Cox) disease-free survival curves of HCC patients was used basing on tissue CitH3 immunoscore.

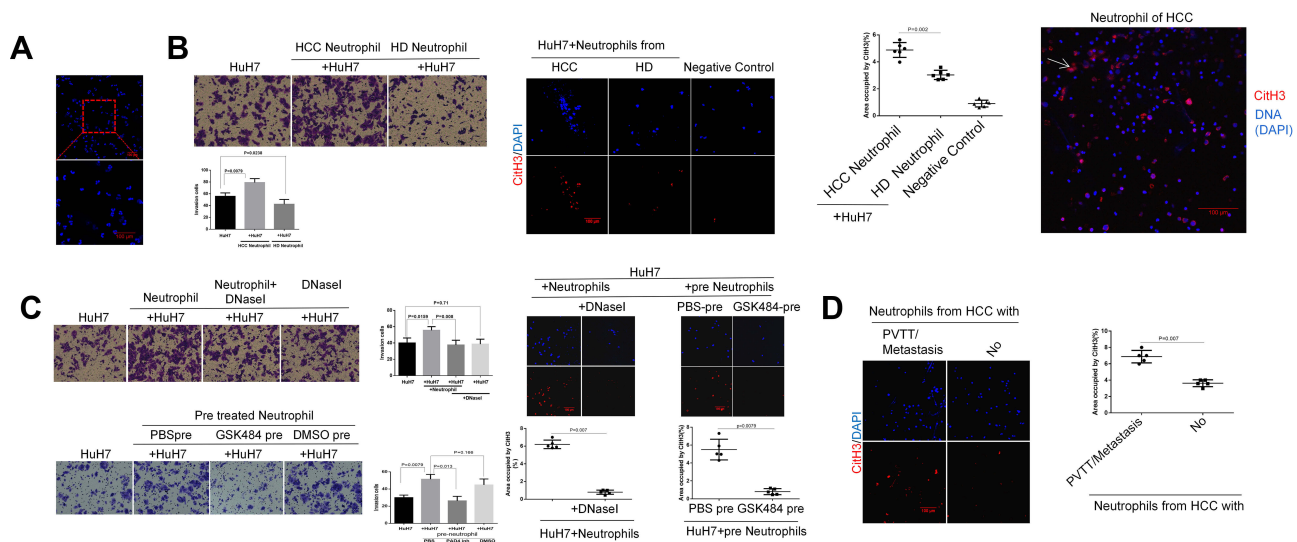
decondensation. Citrullinated histone H3 (CitH3) has been shown to be highly produced during NETsosis process mainly by peptidyl arginine deiminases 4 (PAD4), an enzyme which is crucial to NETs formation. CitH3 has been proposed as a target biomarker reflecting NETs formation, and several methods to quantify NETs in plasma samples and tissues through marker CitH3 have been developed.<sup>50,51</sup> By assessing 85 patients who underwent liver resection between 2011 and 2019, we gained histological evidence for CitH3 deposition in HCC tissues, suggesting that significantly stronger CitH3 expression was observed in the tissues specimens from HCC patients with PVTT or extra-hepatic metastasis than in those without metastasis (Figure 1B, Table S2). In order to further assess whether the presence of NETs in the tissue micro-environment was correlated with disease prognosis, the same sets of 80 tissue specimens were analyzed for correlation between the CitH3 level and the survival rate; the results indicated that patients with higher CitH3 tissue expression displayed a poorer survival percentage (Figure 1C). Collectively, these results showed that NETs formation was correlated with HCC malignant progression.

## Neutrophils of HCC Promote Cancer Cells Invasion via NETs Release in vitro

In order to investigate the invasion-inducing capacity of neutrophils, we performed a transwell chamber assay.

Isolation of peripheral blood neutrophils from patients or healthy donors routinely yielded a purity of greater than 90%, evaluated by white blood cell counting (Figure 2A, Table S1). Freshly isolated human neutrophils were plated into the lower chamber and cancer cells were added on the top of Matrigel-coated membranes in the upper wells. By co-culturing in this manner with HCC cells, neutrophils isolated from the HCC patients with metastasis effectively promoted HCC cell invasion, while those from an HD conversely inhibited the invasion phenotype (Figure 2B). Compared to HD neutrophils, HCC neutrophils exhibited an increased capacity of releasing the NETs structure into the extracellular space (Figure 2B). Here, we further observed that neutrophils from HCC patients with extra-hepatic metastasis showed an enhanced capacity of forming NETs compared to those without metastasis (Figure 2D). These findings demonstrated that neutrophils from patients with HCC but not HD efficiently promoted HCC cell invasion.

In order to determine whether NETs promote HCC cell invasion, we digested extracellular NETs structure by adding DNase I into the transwell system. The ability of neutrophils to stimulate the invasion of HuH7 was inhibited by the digestion of DNase I, whereas DNase I displayed no effect on fetal bovine serum (FBS)-stimulated HuH7 invasion. Next, the necessity of peptidyl arginine deiminase type 4 (PAD4), a crucial enzyme responsible for histone



**Figure 2** Neutrophil of HCC facilitates HCC cells invasion through NETs formation in vitro. **(A)** DAPI staining showed the purity of the neutrophils isolated from human peripheral whole blood. Scare bar: 100um. **(B)** The representative images of transwell assay for the effects of neutrophils on HCC cells invasion were shown. The cells were counted under a microscope in twenty randomly selected fields. NETs were identified by CitH3 component. The expression of NETs-CitH3 was detected by immunofluorescence staining on coverslip. The white arrow points to the expression and presence of NETs-CitH3. Scare bar: 100um. **(C)** The representative images of transwell assay for the effects of DNaseI or PAD4 inhibitor GSK484 on neutrophils-induced HCC cells invasion were shown. The expression of NETs-CitH3 was detected by immunofluorescence staining on coverslip. Neutrophils of HCC was pre-treated with GSK484 at 10uM for 4h before co-culture. 15µL DNaseI was added into per 500µL culture medium in lower chamber. Scare bar: 100um. **(D)** The expression of NETs-CitH3 was detected by immunofluorescence staining for neutrophils from HCC patients with metastasis or without. Scare bar: 100µm.

citrullination during NETosis, in NETs-stimulated cancer cell invasion was examined. As expected, pre-incubating neutrophils with the PAD4 inhibitor GSK484 before co-culture efficiently reduced the NETs-arousing capacity, which in turn blocked the neutrophils' ability to promote HCC cell invasion (Figure 2C). These results revealed that neutrophils of HCC, through NETs formation, contributed to HCC cell invasion.

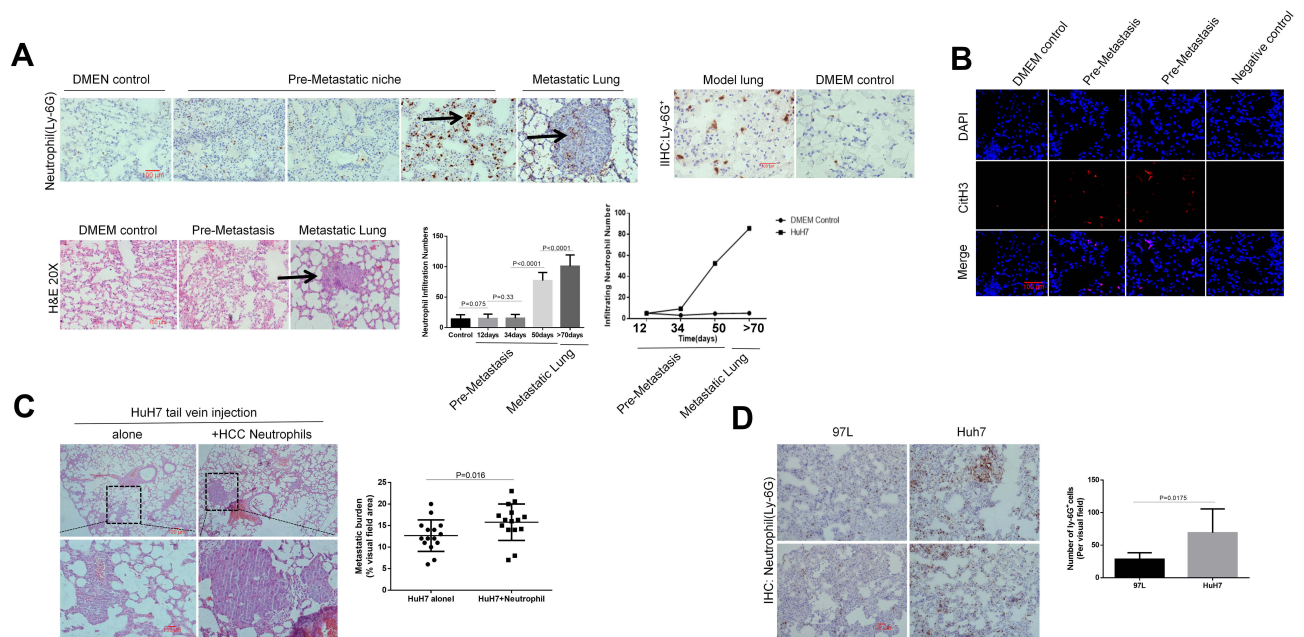
## NETs Form in HCC Pre-Metastatic Niche, While the Neutrophils of HCC Enhance Metastasis in vivo

HCC cells were intravenously injected into nude mice to establish a lung metastasis model. The success rate of this metastatic model reached about 83%. In detail, sixteen out of twenty mice developed lung detectable metastasis foci after the injection of HCC cells. Using this mice model, injection of HuH7 cells spontaneously resulted in gradual neutrophil infiltration into the site of the model lung at the pre-metastatic stage, in the absence of an infection trigger. We observed that neutrophil was the major immune component that accumulated and released the NETs structures in the lung, which preceded the microscopic metastasis foci. In detail, abundant neutrophil influx and NETs-CitH3 became detectable at about 40 days after injection, with the amounts

elevating for days, whereas multiple cancer cells reached the inside of the lung and accumulated there about eight weeks later (Figure 3A and B; Figure S2). Accordingly, we speculated that neutrophils contributed to the formation of the pre-metastatic niche during HCC metastasis through the NETosis process. Next, to investigate the effect of HCC neutrophils on metastasis in vivo, we injected HuH7 cells with or without human neutrophils of HCC intravenously into null mice. Enhanced metastatic burden was observed when HuH7 cells were co-injected with neutrophils, compared to the injection of HuH7 cells alone (Figure 3C). In addition, by tail vein injection of HCC cells with different metastatic capacity, we showed that the highly metastatic HCC cell line resulted in more ly-6G<sup>+</sup> cells penetrating into the lung at the pre-metastasis stage (Figure 3D).

## HCC Cells-Derived CXCL8/IL-8 Primes Normal Neutrophils Towards NETs Formation in an NADPH-Dependent Manner

We next focused on how the HCC cells elicited neutrophil sensitization for the formation of NETs. Indeed, cancer cell-induced NETs formation was dependent on the NADPH oxidase activity.<sup>10,42,49</sup> We performed Western blot and immunohistochemistry assays, confirming that



**Figure 3** Neutrophils of HCC enhances metastasis burden in vivo. **(A)** Huh7 cells were injected into nude mice via tail vein. At various time points (12, 34, 50 days) after HCC cells injection, the mice were killed and examined the density of Ly-6G<sup>+</sup> cells infiltration. Ly-6G<sup>+</sup> cells in nude mice lung were shown by immunohistochemistry staining with lung frozen section (n=7 each). Representative images of Ly-6G<sup>+</sup> neutrophils in the lung were shown. The black arrows point to Ly-6G<sup>+</sup> neutrophils (IHC staining) and metastasis (H&E staining). Scale bar: 100µm. **(B)** NETs-CitH3 were detected by immunofluorescence staining with lung frozen section. Representative images of CitH3 in the lung were shown (n=7 each). Scale bar: 100µm. **(C)** Huh7 cells were co-injected with HCC neutrophils (10:1) into nude mice (n=7 each). The metastatic burden in each group were calculated as the percentage of area occupied by metastatic tumor on each lung section, determined using Image J software. Representative images of metastatic foci in lung were shown. Scale bar: 100µm. **(D)** Ly-6G<sup>+</sup> cells were detected by immunohistochemistry with frozen lung section after Huh7 or 97L injection (n=5 each). Representative images were shown. Scale bar: 100µm.

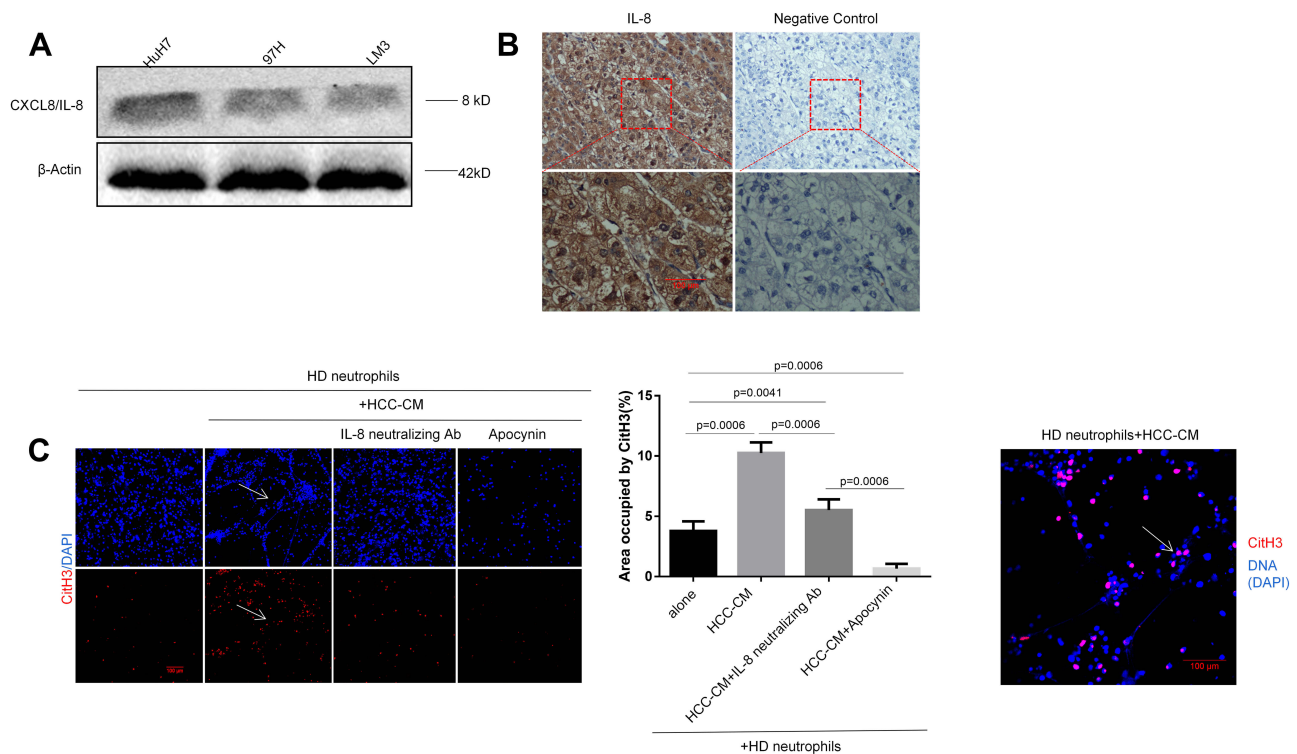
the pro-inflammatory cytokine IL-8 was produced remarkably in HCC cells and the patient's tissue, respectively (Figure 4A and B). By co-culturing HD neutrophils with the conditioned medium (CM) of HCC cells (HCC-CM), we observed that HCC-CM-induced HD neutrophils extruded extensive NETs, while the HD neutrophils alone exhibited a weak capacity of spontaneous NETs formation. Blockade of IL-8 with a neutralizing antibody (Ab) led to a significant reduction in NETs formation, although not a complete inhibition, suggesting additional factors in HCC-CM being involved in NETs induction (Figure 4C).

Moreover, the activation of the NADPH oxidase (NOX2) enzyme in neutrophils has been shown to be essential for NETosis.<sup>15,16</sup> A NOX2 inhibitor apocynin was previously found to strongly inhibit NETs formation and neutrophils-stimulated cancer cell invasion in vitro.<sup>46</sup> As expected, inhibiting NOX2 with apocynin completely prevented the HCC-CM from inducing HD neutrophils toward NETs formation (Figure 4C). Likewise, robust NETs formation was aroused when HD neutrophils were co-cultured with HCC patient serum, but not normal serum. Consistently, IL-8-neutralizing Ab reduced serum-induced NETs formation, while NOX2 inhibitor apocynin

exhibited a stronger inhibition capacity. But, treatment with apocynin did not completely prevent the NETs formation compared to the HD serum group (Fig. S1). Altogether, our data demonstrated that HCC cell-secreted CXCL8/IL-8 primed the HD neutrophils to release NETs structure through the NADPH pathway.

## NETs-Associated Cathepsin G Facilitates HCC Cells Invasion in vitro

NETs are characterized by the association of neutrophil protease with extracellular DNA-binding histone. It is speculated that the pro-invasive effects of NETs require the NETs-associated protease activity.<sup>44,46</sup> In order to elucidate what acts as a stimulus for the enhanced invasion phenotype of NETs-trapped HCC cells, we focused on the cathepsin G (cG) and neutrophil elastase (NE) components present on the NETs structure, providing a mechanistic insight by adding the indicated inhibitors into the co-culture medium. Indeed, inhibition of cG, but not NE, efficiently blocked the neutrophils' ability to promote HCC cell invasion, and also strikingly reduced the extension of NETs formation (Figure 5B). These findings suggested that cG is not just an invasion driver; it is also



**Figure 4** HCC cells-derived CXCL8/IL-8 triggers TAN towards NETs formation through NADPH pathway. **(A)** The protein expression of IL-8 was detected by Western blot assay in three HCC cell lines. Experiment was repeated three times. **(B)** The expression of IL-8 in HCC tissues was examined by the immunohistochemistry (n=30). A representative image was shown. Scare bar: 100µm. **(C)** Co-incubation of HD neutrophils ( $2.5 \times 10^6$ /mL) with the conditioned medium of HCC cells (HCC-CM) (1:2) was performed to explore the effect of IL-8 on HCC-CM-induced NETs formation. The expression of CitH3 was detected by immunofluorescence staining on coverslip for the identification of NETs formation. Representative images were shown. IL-8 neutralizing antibody was added into the culture medium at the concentration of 5µg/mL. NOX inhibitor apocynin was added into medium at the final concentration of 10µM. Experiment was repeated three times. Representative images were shown. The white arrows point to the presence of NETs-CitH3. Scare bar: 100µm.

essential in the NETs extrusion. Conversely, NE inhibitor sivelestat showed no significant effect on the neutrophils-stimulated invasion phenotype.

Furthermore, we generated NETs-containing conditioned medium (NETs-CM) from HCC neutrophils that formed NETs in response to PMA (Figure 5A). Similarly, NETs-CM markedly enhanced the invasion potential of HCC cells. Co-culturing NETs-CM with cG inhibitor equally blocked the invasion-arousing capacity (Figure 5C). As can be seen, cG protein was expressed during the PMA-stimulated NETosis process in vitro (Figure 6A); yet, digesting NETs-DNA with DNaseI prevented invasion, although cG was not deleted by DNaseI digestion (Figure 6A-C). These results suggested that co-localization of the cG component with NETs-CitH3 was essential for inducing HCC cell invasion.

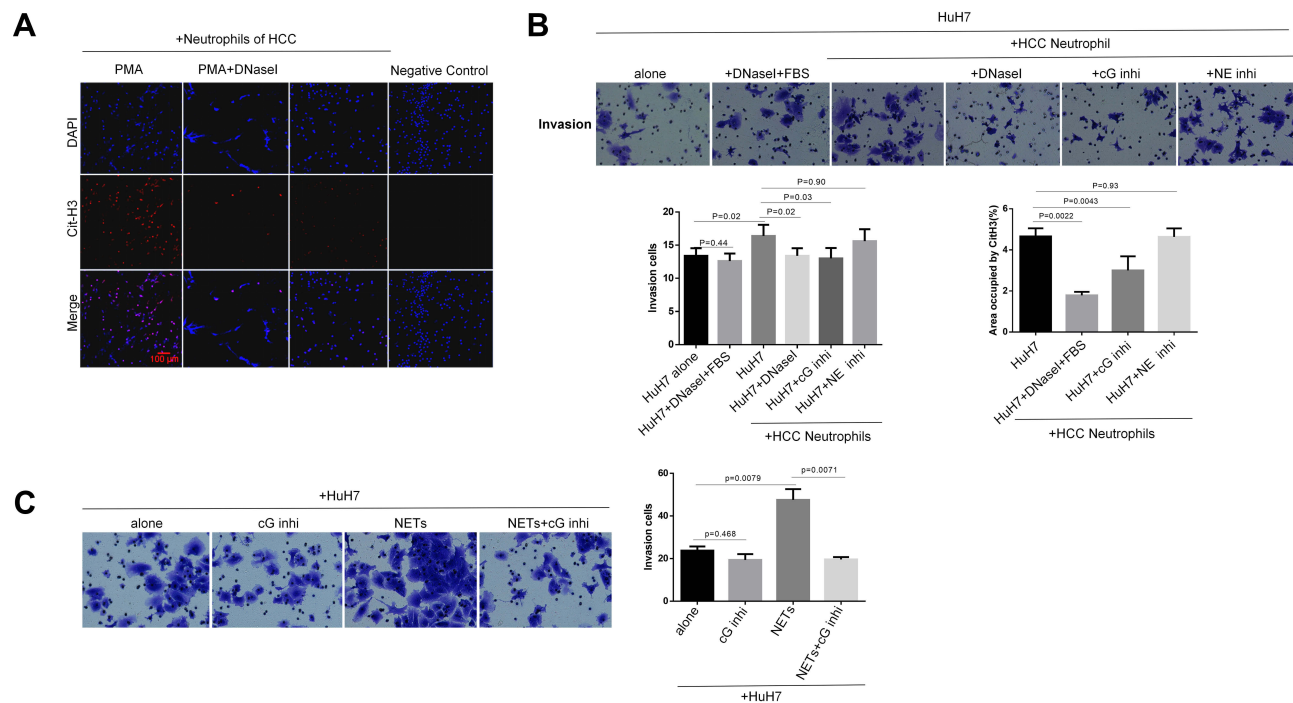
## Targeting the cG Component Effectively Reduces HCC Metastasis in vivo

In order to test the effect of cG inhibitor on tumor metastasis in vivo, we established an experimental metastasis

model by intravascular injection of Hep1-6 cells into C57BL/6J mice. In this model, NETs-CitH3 started to become detectable at the pre-metastatic stage in the model lung compared to the control arms, which could be abrogated by DNaseI treatment. However, as can be seen, the cG inhibitor exhibited little influence on the NETs formation in vivo, while significantly reducing the NETs-CitH3 release in vitro (Figure 7A). Thus, in line with the in vitro results, our findings indicated that the administration of either the cG inhibitor or the DNaseI treatment effectively led to significantly decreased lung metastasis burden compared to control arms (Figure 7B).

Clinically, with human HCC specimens, a higher density of cG protein was observed for metastatic patients than those without metastasis in the peritumoral tissues, but not in paracancerous tissues (Figure 8A and B). A Pearson correlation analysis confirmed that NETs-CitH3 expression positively correlated with cG in peritumoral tissues (Figure 8D). Moreover, by immunohistochemical staining, we found stronger co-localization between cG and CitH3 proteins in





**Figure 5** NETs-associated cathepsin G promotes HCC cells invasion. **(A)** Neutrophils generated NETs in response to PMA. PMA was co-incubated with neutrophils at a final concentration of 500nM for the preparation of NETs. DNaseI abolished the process of PMA-induced NETs formation. The expression of CitH3 was detected by immunofluorescence staining on coverslip for the identification of NETs formation. Representative images were shown. Scale bar: 100 $\mu$ m. **(B)** cG inhibitor reduced NETs extension and neutrophils-induced HCC cells invasion. NE inhibitor had no significant effect on NETs formation and neutrophils-induced invasion. Representative images were shown. cG inhibitor was added into culture medium at a final concentration of 2 $\mu$ M, and NE inhibitor was added into culture medium at a final concentration of 10 $\mu$ M. Experiment was repeated three times. **(C)** cG inhibitor abolished NETs-stimulated HCC cells invasion. cG inhibitor was added into culture medium at the final concentration of 2 $\mu$ M. Control group was set with DMEM culture medium. Representative images were shown. Experiment was repeated three times.

the peritumoral tissue from metastatic HCC compared to the metastasis-free ones (Figure 8C). It is noteworthy that the higher density of co-localization indicated poor disease prognosis in HCC (Figure 8E). Collectively, these findings suggested the significant contribution of cG to HCC metastasis and that the cG inhibitor effectively prevented HCC lung metastasis in vivo.

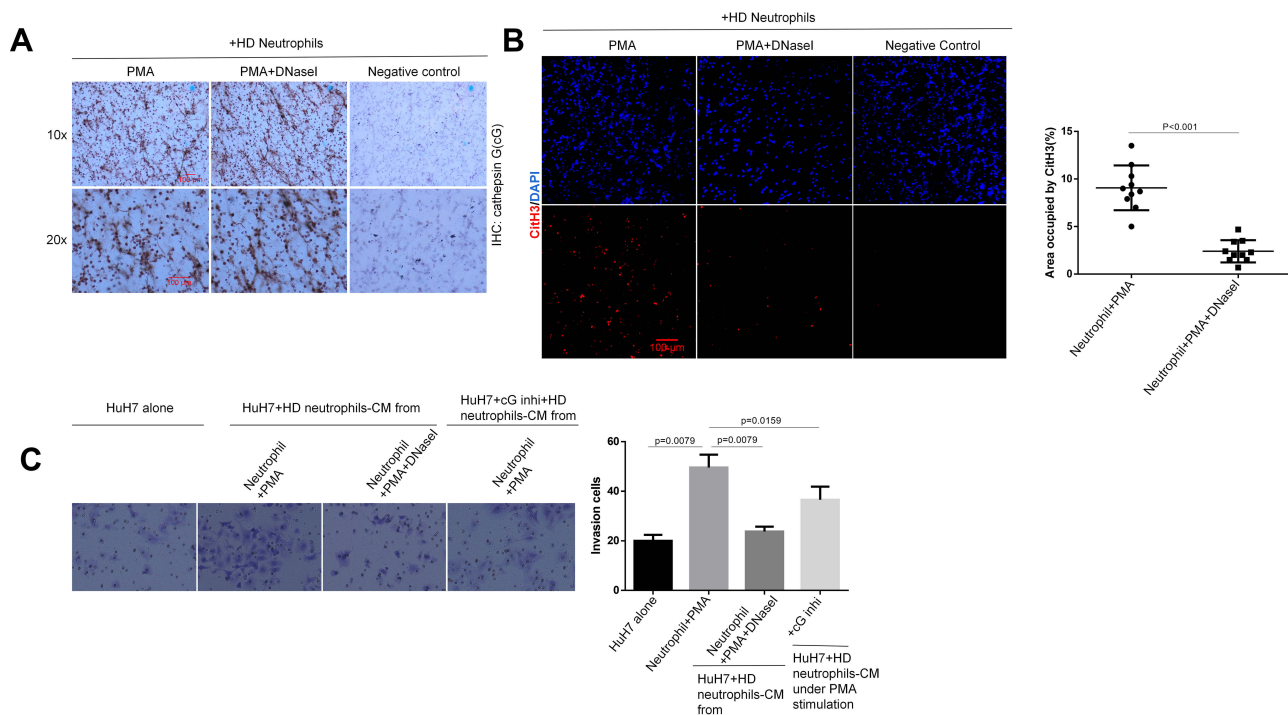
## NETs-Associated Cathepsin G Regulates E-Cadherin Expression in HCC Cells

It is reported that NETs-associated NE and MMP9 awaken dormant breast cancer cells by remodeling the extracellular matrix (ECM).<sup>5,19</sup> In this study, we next focused on the mechanism by which NETs-associated cG facilitated HCC cell invasion. We hypothesized that a likely target for proteases is the adhesion molecule expressed on the surface of HCC cells.<sup>31,41,43</sup> In order to test this hypothesis, HCC cell layers were incubated with NETs-CM for up to 20 h; then, the depleted area of HCC cells was quantified. We observed that incubation with NETs-CM resulted in significant dyshesion of the HCC cell layers (Figure 9C). Using Western blotting, we analyzed the level of CXCL8/

IL-8 and E-cadherin in HCC cells under NETs stimulation, which revealed that NETs-containing CM stimulation for an indicated interval led to elevated expression of CXCL8/IL-8 and decreased E-cadherin expression in HCC cells (Figure 9A and B). Immunofluorescence staining further confirmed the consistent alteration of E-cadherin protein on the surface of HCC cells (Figure 9D). Furthermore, the inhibitor of cG efficiently reversed the decline of E-cadherin but not of CXCL8/IL-8 (Figure 9A and B). Taken together, these results indicated that NETs-associated cG component promoted HCC cell invasion through decreasing E-cadherin expression in vitro.

## Discussion

Tumor metastasis affects a significant proportion of patients with HCC.<sup>11,34</sup> Here, our observations unveiled an inherent mechanism underlying the link between NETs and HCC progression. We began our investigations with the evaluation of human serum samples for the NETs marker MPO-DNA. We selected PVTT as an independent and critical indicator for HCC progression and found that patients with PVTT HCC developed higher NETs markers



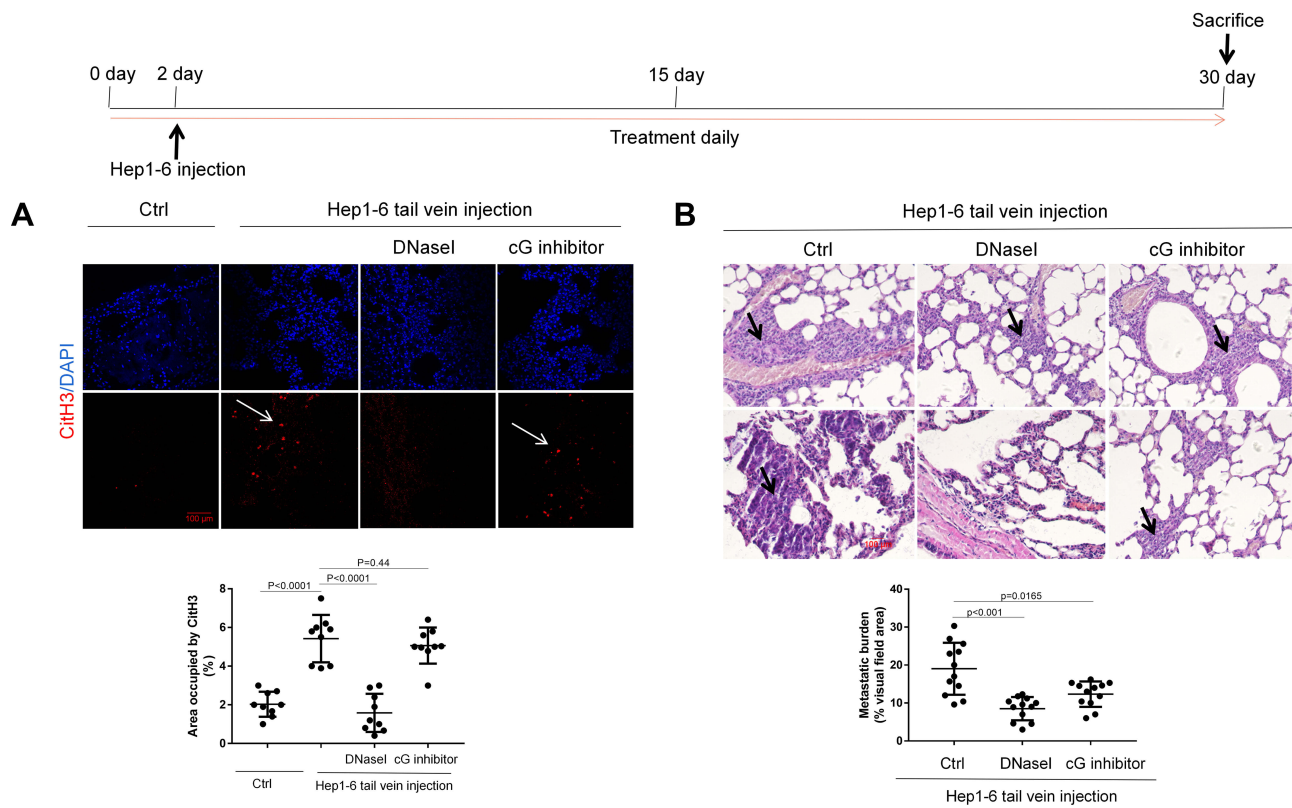
**Figure 6** Co-localization of cG and CitH3 is necessary to promote invasion in vitro. **(A)** The cathepsin G expression of HD neutrophils on coverslip were shown by immunohistochemistry staining. Representative images were shown. Scare bar: 100 $\mu$ m. **(B)** The expression of CitH3 was detected by immunofluorescence staining on coverslip for the identification of NETs formation. Representative images were shown. Scare bar: 100 $\mu$ m. **(C)** Transwell assay was performed to detect the invasion-inducing capacity of HD neutrophils-CM under different treatment. cG inhibitor was added into culture medium at the final concentration of 2 $\mu$ M. Control group was set with DMEM culture medium. Representative images were shown. Experiment was repeated three times.

compared to those without PVTT. We obtained consistent results through all tumor-tissue specimens. These analyses supported our hypothesis of increased NETs formation in patients with advanced HCC.

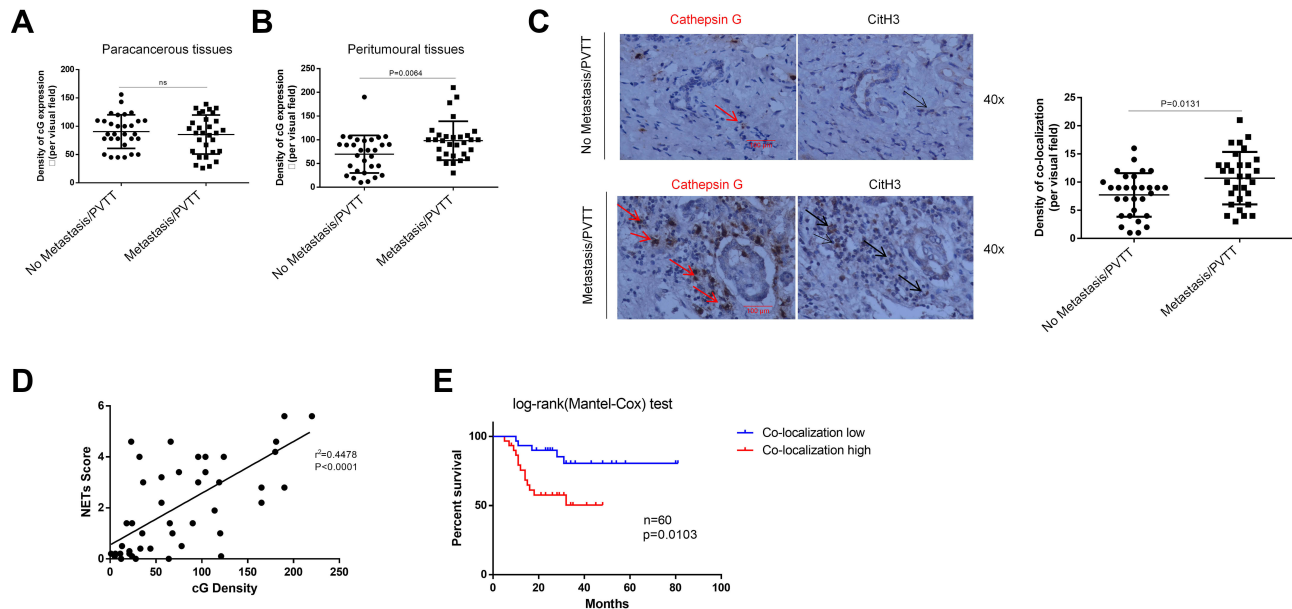
It was initially suggested that NETs aided in the defense against bacteria, fungi, viruses, and parasites.<sup>2,3</sup> However, if dysregulated, excessive NETs could contribute to several diseases, such as immune-related diseases and cancers.<sup>29,30,32</sup> The relationship between NETs-associated proteins and cancer metastasis is interesting. Understanding more mechanisms can lead to the discovery of effective therapeutic targets in HCC progression.<sup>1,3,4</sup> In this study, by blocking PAD4 and inhibiting cG in the transwell assay, we observed that neutrophils failed to enhance cancer cell invasion with reduced NETs formation, suggesting that PAD4 and cG mediated the key signaling pathways related to NETs-triggered invasion potential and HCC cells-inducing NETs release. We further demonstrated that NETs-derived cG promoted HCC cell invasion by reducing the levels of adhesion molecules and increasing the levels of inflammatory mediators in vitro. In addition to cG, other peptides were released during NETosis. MMP9, one of the NETs-

associated proteins, was initially demonstrated to promote the outgrowth of colorectal carcinoma micrometastasis after liver ischemia/reperfusion (I/R) injury.<sup>47</sup> However, the mechanism by which these proteins/peptides participate in HCC metastasis in vivo remains to be investigated.<sup>33,36</sup>

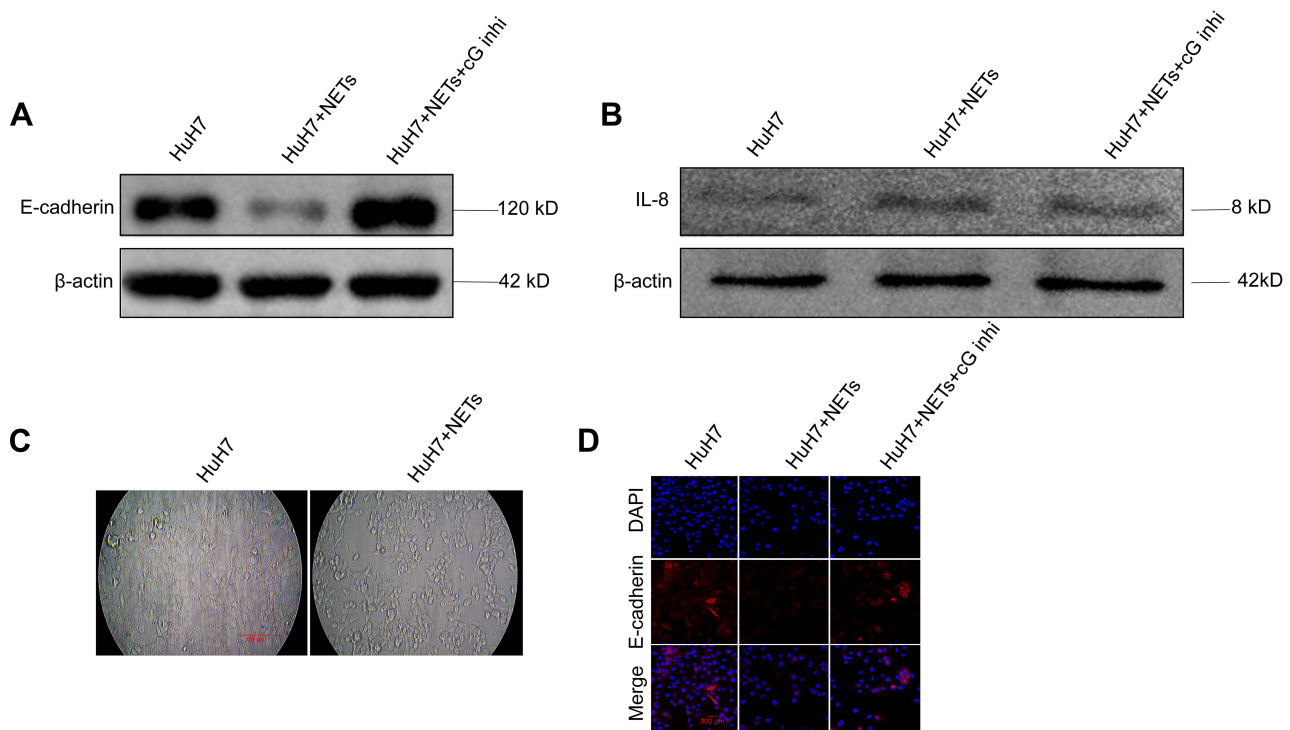
Inflammation has previously been reported to enhance cancer metastasis and is largely initiated by neutrophils.<sup>25,26</sup> NETs formed in response to systemic bacterial infection or stress can enhance cancer cell dissemination in the metastatic microenvironment.<sup>20,21</sup> Recently, a study has reported that increased NETs structure induced under inflammation conditions can promote HCC metastasis via pro-inflammatory TLR9-COX2 signaling pathways. In this study, lipopolysaccharide (LPS) injection into immune-component C57BL/6 mice was used to induce efficient NETs formation, which subsequently promoted lung metastasis. This study stressed the roles of inflammation factors in neutrophil-stimulated cancer invasion, and therefore, proposed that a combination of DNase I with the anti-inflammatory drug aspirin could serve as a novel therapeutic strategy against HCC metastasis.<sup>12</sup> Although additional studies on the mechanisms of NETs



**Figure 7** Inhibition of cG component alleviates lung metastasis in vivo. **(A)** The expression of NETs-CitH3 protein in model lung were shown by immunofluorescence (n=5 each). Representative images were shown. The white arrows point to the expression and presence of NETs-CitH3 in lung. Scare bar: 100µm. **(B)** Hep1-6 cells were injected into male C57BL/6 mice via tail vein (n=5 each), and the metastatic burden in each group were calculated as the percentage of area occupied by metastatic tumor on each lung tissue section, determined using Image J software. Representative images of metastatic foci were shown. The black arrows point to metastasis foci. Scare bar: 100µm.



**Figure 8** Co-localization between cG and NETs-CitH3 in HCC tissues indicates disease prognosis. **(A–C)** The expression of cathepsin G and CitH3 protein in patients tissues were shown by immunohistochemistry staining with serial sections (n=60). Representative images were shown. The red arrows point to protein cathepsin G and black arrows point to protein CitH3 in serial section, indicating the co-localization of protein cathepsin G and CitH3 in tissue. Scare bar: 100µm. **(D)** Pearson correlation analysis of NETs-CitH3 and cG expression in HCC samples (n=60). **(E)** 60 non-metastasis/PVTT HCC patients with different level of cG and NETs for metastasis/PVTT cumulative incidence analysis, and Log rank test was used.



**Figure 9** NETs-associated cathepsin G regulates the expression of E-cadherin in HCC cells in vitro. **(A, B)** Western blot analysis of E-cadherin and CXCL8/IL-8 level in HuH7 cells treated with NETs-CM. Cells were stimulated with NETs-CM for 20 hours. Experiment was repeated three times. **(C)** HuH7 ( $5 \times 10^5$  cells) were co-cultivated with NETs-CM or cultivated alone to confluence for 20 hours, and the area depleted of tumor cells was quantified as percentage of the total area. Representative images were shown. Scale bar: 100 $\mu$ m. **(D)** E-cadherin expression of HuH7 cells layer was shown by immunofluorescence staining. Representative images were shown. Scale bar: 100 $\mu$ m. Experiment was repeated three times.

in cancer progression are needed, some significant progress, particularly in the metastasis and invasion fields, had been made. There has been another strong evidence that NETs triggered HMGB1 release and activated TLR-9-dependent signaling pathway in cancer cells to enhance liver metastasis potential after surgical stress in colorectal cancer.<sup>47</sup> Therefore, whether the HMGB1-TLR9-COX2 signaling axis is affected in cancer metastasis deserves additional investigation.

In the present study, we demonstrated a previously unrecognized pathway on how NETs induced HCC metastasis as well as how HCC cells triggered NETs formation. We found that HCC neutrophils enhanced HCC cell invasion through NETs release in vitro. In line with this, the human HCC neutrophils elicited the HCC cell metastasis in vivo. Then, we sought to determine, which factors secreted from cancer cells accounted for increased NETs formation. Of note, the NETs-arousing ability of HCC-CM was observed to be reduced upon IL-8 blockade. We have shown that HCC-secreted IL-8 triggered neutrophils to initiate the NETosis process through the NADPH oxidase pathway, featuring a new mechanism between HCC cells

and their microenvironment. The dissemination of HCC cells has a strong predilection for the lung and has often been described as a passive process that is, in part, governed by the microenvironmental mechanisms.<sup>37–39</sup> Using the nude mice metastatic model, we observed that HCC cell injection could spontaneously result in neutrophil influx and NETs formation in the pre-metastatic lung in the absence of inflammation induction. Our findings suggested that NETs formation and neutrophil influx induced by cancer cells may be a critical driver for subsequent metastasis behavior, which could be prevented by DNase I. Altogether, with the release of all the pro-tumorigenic peptides, NETs act as fertilizers enriching the pre-metastatic niche to facilitate cancer metastasis in HCC.

NETs, with their large, extracellular, web-like structures, are composed of several granule proteins that are assembled on a scaffold of extruded DNA. These proteins are mainly histones, neutrophil elastase, cathepsin G, myeloperoxidase, defensins, and actin.<sup>3</sup> However, how the NETs composition impacts the NETs function in diseases remains to be identified. We, therefore, determined the cG protein on the PMA-stimulated NETs structure and then

explored the roles of cG in NETs-induced HCC cell invasion in vitro. Our results indicated that co-localization expression of cG and CitH3 was necessary for promoting HCC cell invasion. Using the HCC tissue samples, the co-localization expression between cG and NETs-CitH3 was also observed in the peritumoral tissues, and the density of co-localization was observed to be correlated with disease prognosis. These findings suggested cG as a key signaling target in NETs-triggered metastasis in vivo.

## Conclusions

In summary, our research identified a tumor-associated cellular, molecular, and clinical network, involving NETs, cG, NADPH signaling, and IL-8, that regulates the positive feedback loop between neutrophils and cancer progression. These findings supported the idea of the exploration of this network as a potential therapeutic target against HCC progression.

## Abbreviations

NETs, neutrophils extracellular traps; IL-8, interleukin-8; HCC, hepatocellular carcinoma; CM, conditioned medium; cG, Cathepsin G; PVTT, Portal vein tumor thrombosis; NE, neutrophil elastase; PAD4, peptidyl arginine deiminase type 4.

## Data Sharing Statement

All data generated or analyzed during this study are included in this published article.

## Ethics Approval

Samples were collected after approval by the medical ethics committee of Zhongshan Hospital affiliated to Xiamen University and written informed consents were obtained from each patient. All animal experiments were approved by the animal Ethics committee of Xiamen University, and all institutional and national guidelines for the care and use of laboratory animals were followed.

## Consent for Publication

All authors have seen and approved the manuscript and consent for publication.

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## Author Contributions

Xiangqian Guan performed the study, acquisition and analysis of data, and drafted the manuscript. Yuyan Lu participated in animal study and protein assays. Heping Zhu carried out the collection of patients blood specimens. Wenxiu Zhao participated in draft writing and data analysis. Xiaoqin Chi provided tissues specimens and clinicopathological characteristics of HCC patients. Shuqi Yu participated in the preparation of tissue sections, and helped to perform functional experiments and immunoassays. Chengrong Xie participated in data analysis and gave some technique assistance. Zhenyu Yin started the study and participated in experimental design. Chengrong Xie and Zhenyu Yin approved for the final version and submission. All authors made a significant contribution to the work reported, whether that is in the conception, study design, execution, acquisition of data, analysis and interpretation, or in all these areas; took part in drafting, revising or critically reviewing the article; gave final approval of the version to be published; have agreed on the journal to which the article has been submitted; and agree to be accountable for all aspects of the work.

## Disclosure

The authors reported no conflicts of interest for this work.

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