

Effects of grape seed proanthocyanidin on Alzheimer's disease *in vitro* and *in vivo*

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Abstract. Grape seed proanthocyanidin (GSPA) consists of catechin, epicatechin and epicatechin gallate, which are strong antioxidants that are beneficial to health and may attenuate or prevent Alzheimer's disease (AD). In the present study, the effects of GSPA on pheochromocytoma (PC12) cell viability were determined using cell counting kit-8 and lactate dehydrogenase (LDH) assays, whereas apoptosis and mitochondrial membrane potential (Ψ_m) were measured via flow cytometry analysis. The effect of GSPA administration on the behavior and memory of amyloid precursor protein (APP)/presenilin-1 (PS-1) double transgenic mice was assessed using a Morris water maze. APP A β peptides and tau hyperphosphorylation were examined by western blotting; whereas the expression levels of PS-1 were evaluated by reverse transcription-quantitative polymerase chain reaction and compared with pathological sections stained with hematoxylin-eosin and Congo red. Data from the *in vitro* experiments demonstrated that GSPA significantly alleviated A β 25-35 cytotoxicity and LDH leakage ratio, inhibited apoptosis and increased Ψ_m . The findings from the *in vivo* experiments showed a significant enhancement in cognition and spatial memory ability, an improvement in the pathology of APP and tau protein and a decrease in PS-1 mRNA expression levels. Therefore, the results of the present study indicated that GSPA may be a novel therapeutic strategy for the treatment of AD or may, at the very least, improve the quality of life of patients with AD.

Introduction

Alzheimer's disease (AD) is a progressive neurodegenerative disorder which is characterized by the loss of cognition and memory capacity, and decreased visual-spatial skills. Pathologically, AD is characterized by increased amyloid-beta (A β), hyperphosphorylation and the aggregation of tau protein (1,2). A β peptides are produced from amyloid precursor protein (APP) when it is cleaved by β - and γ -secretases into amino acid peptides within the cerebral cortex and hippocampus (3). Accumulation of insoluble A β induces the aggregation of peptide-forming amyloid fibrils, which have been demonstrated to be neurotoxic *in vitro* and *vivo* (4). The γ -secretase enzyme induces intramembrane cleavage of APP, and is part of a multi-subunit intramembranous protein complex that includes PS-1 (5,6). Moreover, excessive deposition of A β stimulates microtubule-associated protein tau aggregation into abnormally hyperphosphorylated tau, which assemble into paired helical filaments (PHFs) are significantly increased in patients with AD (7).

A β may trigger neurodegeneration via oxidative stress. Previous studies have demonstrated that the production of excessive reactive oxygen species (ROS) and signs of oxidative stress were detected in the AD brain (8-11). In addition, it has been demonstrated that oxidative stress has a critical role in A β -mediated neuronal cytotoxicity by triggering or facilitating neurodegeneration (12). Oxidative damage may initiate during the earlier stages of AD and induce amnesic mild cognitive impairment and a reduced antioxidant capacity (13-15). Furthermore, oxidative damage is associated with apoptosis, mitochondrial membrane damage and mitochondrial dysfunction (16); therefore, reducing oxidative stress may decrease A β -induced neurotoxicity (17). We hypothesized that strong antioxidants were capable of attenuating oxidative stress, and may represent a therapeutic strategy to treat A β -induced neurotoxicity and improve the symptoms of AD.

A previous study in transgenic animals have shown that moderate consumption of red wine may alleviate AD-like neuropathology and cognitive deterioration (18). Furthermore, Ono *et al* (19) demonstrated that a specific grape-derived polyphenolic extract (GSPE) reduced A β peptide oligomerization

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and fibril development *in vitro*. Previous studies have also indicated that families of polyphenols may interfere with tau fibril formation *in vitro* and in cultured cells (20,21). Notably, it has been demonstrated that GSPE attenuates the aggregation of tau-mediated neuropathology in the brain of the Thy-1 mutated human tau mouse model of tauopathy (22). GSPE is a complex mixture of proanthocyanidin monomers, oligomers and polymers, which are strong antioxidants; therefore, the antioxidant activities of these compounds may be beneficial to patients with AD (23,24). We hypothesize that using monomers of aqueous grape seed proanthocyanidin (GSPA) may induce a stronger antioxidant effect than a complex mixture of GSPE. This may provide a new strategy for the treatment of AD or, may at least improve the quality of life of patients with AD.

Materials and methods

Ethics statement. All experimental protocols were approved by the Ethics Committee of the Animal Center of Guangzhou University of Traditional Chinese Medicine (Guangzhou, China).

Grape seed extraction. Grape seed plant material was purchased (Biovin Naturprodukte, Ilbesheim, Germany), which was extracted for 2 h in an ethanol:water (13:7; v/v) mixture. Following filtration of the extracting solution, the filtrate was recovered using ethanol and ultrafiltered. The eluent was concentrated by rapid drying with hot gas, and the resulting eluate was lyophilized and reconstituted in phosphate-buffered saline (PBS) at various concentrations for biological assays and is referred to as aqueous GSPA. High-performance liquid chromatography (HPLC) analysis quantification of GSPA is shown in Fig. 1. Preliminary studies have demonstrated that GSPA is non-toxic to normal C57 mice at single doses (≤ 7000 mg/kg) within the first 14 days (data not shown).

Reagents. Beta-amyloid peptide (A β 25-35) and rhodamine 123 were purchased from Sigma-Aldrich (St. Louis, MO, USA). Dulbecco's Modified Eagle Medium (DMEM) and PBS were obtained from Hyclone (GE Healthcare Life Sciences, Logan, UT, USA). Fetal bovine serum (FBS), horse serum (HS), 0.25% trypsin and phenol red were purchased from Gibco (Thermo Fisher Scientific, Waltham, MA, USA). Annexin V-fluorescein isothiocyanate (FITC) and propidium iodide (PI) were purchased from eBioscience, Inc., (San Diego, CA, USA). Cell counting kit-8 (CCK-8) was purchased from Dojindo Molecular Technologies, Inc., (Kumamoto, Japan). Penicillin and streptomycin were purchased from Solarbio Science & Technology Co., Ltd., (Beijing, China). Lactate dehydrogenase (LDH) assay kit was purchased from Nanjing Jiancheng Bioengineering Institute (Nanjing, China). Immobilon-P polyvinylidene difluoride membrane was purchased from EMD Millipore (Billerica, MA, USA). Anti-APP antibody (cat. no. ab2072) and phosphorylated (phospho) tau (cat. no. s396) were purchased from Abcam (Cambridge, MA, USA). Anti- β -actin (cat. no. sc-47778) was purchased from Santa Cruz Biotechnology, Inc., (Dallas, Texas, USA). Horseradish peroxidase (HRP)-conjugated anti-mouse

IgG (cat. no. PA43002) was purchased from Kirkegaard & Perry Laboratories, Inc., (Gaithersburg, Maryland, USA). Donepezil hydrochloride was purchased from Eisai China Inc., (Shanghai, China). All other reagents and chemicals used in the present study were of analytical grade.

Animals and drug administration. A total of 30 APP/PS1 male heterozygous mice [Swedish mutant β -amyloid precursor protein, a 9 exon presenilin 1 gene mutation; license number: SCXK (Su) (2010-0001)], aged 5-6 months and weighing 22 ± 0.94 g, were purchased from Nanjing University (Nanjing, China), of which 24 were APP/PS1 double transgenic mice and remaining 6 were normal. Mice were housed according to a 12-h light-dark cycle with *ad libitum* access to food and water. Mice were randomly divided into the following five groups: Control group [6 normal mice administered saline by oral gavage (OG) for 2 months]; APP/PS1 model group (6 double transgenic mice administered saline by OG for 2 months); APP/PS1 donepezil control group (6 double transgenic mice administered donepezil hydrochloride by OG at 2 mg/kg/day for 2 months); low dose APP/PS1-treated group (6 double transgenic mice administered GSPA by OG at 50 mg/kg/day for 2 months); and high dose APP/PS1-treated group (6 double transgenic mice administered GSPA by OG at 100 mg/kg/day for 2 months).

Preparation of aggregated A β 25-35. A β 25-35 peptide was dissolved in deionized distilled water at 1 mM and incubated for 7 days at 37°C to induce aggregation, as previously described (10,25,26). Following aggregation, the solution was stored at -20°C until use.

Cell culture and treatment. Pheochromocytoma (PC12) cells were obtained from the College of Pharmacy of Sun Yat-Sen University (Guangzhou, China). Cells were seeded in 25 cm² flasks at a density of 1×10^5 cells and maintained in DMEM supplemented with 100 U/ml penicillin, 100 U/ml streptomycin, 5% HS and 5% FBS at 37°C in a humidified atmosphere of 95% air and 5% CO₂. At 80% confluence, cells were subcultured for 24 h in the same conditions prior to incubation with various concentrations of GSPA (12.5, 25, 50 and 100 μ g/ml) for 2 h. Following this, 20 μ M A β 25-35 was added to the culture medium for an additional 24 h prior to the initiation of the assays.

Cell viability assay. Cell viability was evaluated via quantitative colorimetric CCK-8 and LDH assays following treatment with GSPA, A β 25-35 or combination therapy with both. Briefly, cells were seeded onto 96-well culture plates at 1×10^4 cells/well in DMEM and, following drug treatment, the cell cultures were supplemented with 10 μ l/well CCK-8 solution and incubated at 37°C for 1 h. Optical density of each well was determined at 450 nm using a microplate reader (FSA-1510; Thermo Fisher Scientific, Inc.). Cell viability was expressed as a percentage of the untreated controls.

For the LDH assay, 150 μ l incubation medium was collected from each well and added to an LDH assay solution. LDH activity was measured using a microplate reader, according to the manufacturer's instructions (Nanjing Jiancheng Bioengineering Institute).

Assessment of apoptosis. Annexin V-FITC and PI staining was used to detect apoptosis and necrosis following drug treatment. During the early stages of apoptosis, membrane phosphatidylserine (PS) is translocated from the inner lipid layer of the plasma membrane to the outer layer in various cell types, including PC12 cells (27). Once on the cell surface, PS can be easily detected by staining with annexin, which is a protein with a strong affinity to PS. Annexin was conjugated to the highly photostable FITC. Cells in the late stage of apoptosis were measured by conventional PI staining. The assay was directly performed on live cells and the relative number of early and late apoptotic cells was measured using flow cytometry. Following drug treatment, cells (1×10^5 cells/plate) were washed with PBS, harvested and subsequently centrifuged at $2,000 \times g$ for 5 min at 37°C . Cells were then resuspended in $1000 \mu\text{l}$ buffer and an aliquot ($190 \mu\text{l}$) of the cell suspension was mixed with $5 \mu\text{l}$ Annexin V-FITC and $10 \mu\text{l}$ PI and incubated for 25 min at room temperature in the dark. Cell staining was measured with a fluorescence-activated cell sorting (FACS) flow cytometer (BD FACSCanto II; BD Biosciences, San Jose, CA, USA) at $\text{Ex}=488 \text{ nm}$ and $\text{Em}=530 \text{ nm}$. Cell apoptosis was expressed as the percentage of control cultures incubated with Annexin V-FITC + PI, but not treated with GSPA or A β 25-35.

Measurement of mitochondrial membrane potential (Ψ_m). Mitochondrial Ψ_m was measured using rhodamine 123. Rhodamine 123 can enter the mitochondrial matrix and induce photoluminescent quenching that is dependent on mitochondrial transmembrane potential (28). Following drug treatment, PC12 cells were incubated with 5 mg/l rhodamine 123 for 30 min at 37°C in the dark. Subsequently, cells were washed three times with PBS and the fluorescence emission intensity was measured with a flow cytometer at $\text{Ex}=488 \text{ nm}$ and $\text{Em}=535 \text{ nm}$. Intensity of fluorescence emission was expressed as the percentage of control cells incubated in rhodamine 123 but not treated with GSPA or A β 25-35.

Morris water maze (MWM) test. The MWM test was used to assess alterations in the behavior of the mice, as previously described (29). The maze was a circular pool (diameter, 160 cm; depth, 50 cm) filled with water at $24\text{-}26^\circ\text{C}$ to a depth of 35 cm. Soluble skim milk was used to ensure the water was opaque. A hidden platform (diameter, 8 cm) was submerged $\sim 2 \text{ cm}$ below the surface of the water in the center of the designated target quadrant. Visual cues were placed around the water maze. The two phases of the MWM tests included an oriented navigation trial and a spatial probe trial. Oriented navigation trials were conducted four times/day for five days. In each trial, mice were placed into the water in a different quadrant and given 120 sec to find the platform and remain on the platform for 10 sec. If the mouse failed to locate the platform within the given time, it was guided to the platform and remained on the platform for 10 sec. During the oriented navigation trials, behavior and the time required for the mouse to locate the hidden platform (escape latency) were recorded via a computerized video tracking system. Mice that did not locate the hidden platform within the allotted time scored a maximum of 120 sec. During the spatial probe trials, the platform was removed and the mice were allowed to swim freely for 120 sec. The mean search time each mouse spent

in the target quadrant was recorded to assess spatial memory ability.

Hematoxylin-eosin and Congo red staining. All mice were sacrificed with an overdose (150 mg/kg) of pentobarbital sodium (Sigma-Aldrich, St. Louis, MO, USA). Paraffin-embedded tissue sections were placed on slides, dewaxed and stained with hematoxylin for 3 min, followed by eosin staining for 3 sec. Sections were then dehydrated with alcohol, fixed with xylene and sealed. Hippocampal histopathological abnormalities were investigated under a light microscope. The number of cells in the hippocampal CA1 region of each section was examined by three independent pathologists in a blinded manner. The average number of cells was used as the final result.

Three slides from each mouse were rehydrated in a graded alcohol series, immersed in hematoxylin for 2 min, and subsequently submerged in hydrochloric acid for 10 sec. Following this, sections were rinsed in running tap water for 10 min until they turned blue, washed twice in distilled water and stained with Congo red for 40 min at room temperature prior to rinsing in running tap water. Subsequently, the slides were submerged in lithium carbonate for 5 sec and washed in running tap water. Alcohol (80%) was used to differentiate between true staining and nonspecific background staining. Sections were rinsed in running tap water for 10 min, cleared in xylene, and covered with neutral gum. Predetermined marks were used to position the light microscope field for image collection (Olympus BX41; Olympus Corporation, Tokyo, Japan). For blind analysis, this and all subsequent steps were performed by an investigator unaware of the treatment condition of each sample using an additional numbering code. Images were collected using the x20 objective (providing an overall magnification of x200) of the region of interest. For ease of processing, all images were of the same size. The intensity of each object was analyzed using Image J software version 1.37 software.

Western blot analysis. The right hippocampus was harvested from the mice and stored in liquid nitrogen. Following tissue homogenization, total proteins were extracted using the total protein extraction reagents kit (EMD Millipore). Protein concentration was measured using a bicinchoninic acid protein assay kit (Qian Chen Biotechnology Company, Shanghai, China). Protein samples ($20 \mu\text{l}$) were separated by SDS-PAGE for 70 min at 90 V and transferred to PVDF membranes using transblotting apparatus (Bio-Rad Laboratories, Inc., Hercules, CA, USA) for 90 min at 90 V. Membranes were blocked with 5% (w/v) skim milk at room temperature for 30 min and subsequently incubated at room temperature for 90 min with anti-APP antibody (1:1,000) and phospho tau (1:2,000) primary antibodies. The immunolabeled membranes were washed once with Tris-buffered saline with Tween 20 (TBS-T) for 30 min followed by three separate washes (10 min/wash). Following this, the membranes were probed with a HRP-conjugated secondary antibody (1:2,000) at room temperature for 1 h. To verify equal protein loading, the membranes were incubated with monoclonal β -actin antibody (1:1,500) at room temperature and then the same HRP-conjugated goat anti-mouse IgG (1:2,000) at 37°C for 2 h. The membrane was washed three times with TBS-T and the protein bands were visualized

with enhanced chemiluminescence western blotting detection reagents. The intensity of each target protein band was analyzed using Image J software version 1.37 (National Institutes of Health, Bethesda, MA, USA) and expressed relative to β -actin density.

Reverse transcription-quantitative polymerase chain reaction (RT-qPCR). Hippocampal tissue was homogenized using an automated homogenizer at 4°C. Total RNA was harvested from the hippocampus, isolated using TRIzol reagent and reverse transcribed into cDNA using RT-PCR kits according to the manufacturer's instructions. The contents of the reaction mixture were as follows: 2 μ l total RNA, 1 μ l Oligo dT, 9 μ l DEPC-treated water, 4 μ l 5X reaction buffer, 1 μ l Ribolcok TMRNase inhibitor, 2 μ l 10 mM dNTP and 1 μ l RevertAid TMV reverse transcriptase. cDNA was amplified by RT-qPCR on an ABI Prism 7500 system (Thermo Fisher Scientific, Inc.) using 12.5 μ l 2X Maxima SYBR Green/Rox Master Mix reagent, 0.75 μ l forward primer, 0.75 μ l reverse primer, 2 μ l template DNA and 9 μ l nuclease-free water. Expected RT-qPCR product sizes and the primers used in this study are presented in the Table I. Samples were inactivated for 2 min at 50°C prior to hot-start amplification. Amplification cycles were performed as follows: 95°C for 15 min, followed by 40 cycles of 55°C for 15 sec, 60°C for 30 sec and 72°C for 30 sec. RT-qPCR was repeated in triplicate. Data from the reactions were collected and analyzed using ABI Prism 7500 software. Relative gene expression levels were calculated according to the $2^{-\Delta\Delta C_q}$ method and were normalized to β -actin expression in each sample (30). **Statistical analysis.** Data are expressed as means \pm standard error of the mean. Multiple group comparisons were performed using one-way analysis of variance followed by Dunnett's test for pair-wise comparisons between groups. SPSS 15.0 (SPSS, Inc., Chicago, IL, USA) was used for all statistical analyses in this study. $P < 0.05$ was considered to indicate a statistically significant result.

Results

HPLC analysis of GSPA. Normal-phase-HPLC chromatograms of GSPA are presented in Fig. 1A. The most abundant peaks of the most active subfractions were purified using HPLC in isocratic elution mode to yield active fractions. Catechin retention time was 25.6 min, whereas epicatechin retention time was 32.8 min. Normal-phase-HPLC chromatograms of catechin and epicatechin are presented in Fig. 1B. The results suggest that catechin and epicatechin are abundant in GSPA.

Effect of GSPA on PC12 cell viability. Cell viability was assessed using a CCK-8 reduction assay (Fig. 2). Treatment of PC12 cells with $< 200 \mu\text{g/ml}$ GSPA alone for 24 h was demonstrated to be non-toxic. CCK-8 reduction is linearly correlated with cell number and only 200 $\mu\text{g/ml}$ GSPA, which was the highest concentration tested, reduced the viable cell number ($\sim 92\%$ of control). The results suggest that GSPA $< 200 \mu\text{g/ml}$ is non-toxic to PC12 cells.

Treatment with GSPA protects against A β 25-35-induced cytotoxicity in PC12 cells. Treatment of PC12 cells with 20 μM A β 25-35 for 24 h significantly reduced the estimated

Table I. Primers for reverse transcription-quantitative polymerase chain reaction analysis of β -actin and presenilin-1.

cDNA product	Sequence (5'-3')
β -actin	Forward, CTGGTGAAACTCTGCGTCTG Reverse, AGAACAAGCGCCATACGACT
Presenilin-1	Forward, GTGGTGAAACTCTGCGTCTG Reverse, AGAACAAGCGCCATACGACT

viable cell number to 65.9% of the control ($P < 0.01$; Fig. 3A). However, when PC12 cells were pretreated for 2 h with GSPA (12.5, 25, 50 and 100 $\mu\text{g/ml}$, respectively) A β 25-35-induced cytotoxicity was significantly reduced to 69.5% of control viability at 12.5 $\mu\text{g/ml}$ ($P < 0.05$), 70.7% at 25 $\mu\text{g/ml}$ ($P < 0.01$), 84.8% at 50 $\mu\text{g/ml}$ ($P < 0.01$), and 86% at 100 $\mu\text{g/ml}$ ($P < 0.01$).

Analysis of the levels of LDH released into the culture media from dead/dying cells confirmed that GSPA partially blocked A β 25-35-induced cytotoxicity (Fig. 3B). Compared with the control group, A β 25-35 treatment significantly increased LDH leakage ($P < 0.01$) and this release was significantly attenuated by GSPA administration ($P < 0.01$). The results suggest that GSPA is able to partially block A β 25-35-induced cytotoxicity.

Treatment with GSPA reduces A β 25-35-induced apoptosis. Annexin V-FITC and PI double staining can distinguish healthy cells (annexin-negative; PI-negative) from early apoptotic (annexin-positive; PI-negative), late apoptotic (annexin-positive; PI-positive), and necrotic (annexin-negative; PI-positive) cells (31). The apoptotic (early + late) rate of PC12 cells following A β 25-35 treatment was 45.1%; however, GSPA pretreatment significantly reduced the apoptotic rate to 39.9% at 50 $\mu\text{g/ml}$ ($P < 0.05$) and 33.9% at 100 $\mu\text{g/ml}$ ($P < 0.01$). The healthy cell rate following A β 25-35 treatment was 53.3%. However, GSPA pretreatment increased the healthy cell rate to 60% at 50 $\mu\text{g/ml}$ and 65.1% at 100 $\mu\text{g/ml}$ (both $P < 0.01$, as compared with A β 25-35 alone) (Fig. 4). The results suggest that GSPA reduces A β 25-35-induced apoptosis.

Treatment with GSPA reverses depolarized mitochondrial Ψ_m in PC12 cells. Cells exposed to 20 μM A β 25-35 for 24 h exhibited significantly depolarized mitochondrial Ψ_m (45% loss of Ψ_m ; $P < 0.01$, as compared with control group), which indicated impending apoptosis or necrosis (Fig. 5). Pretreatment with GSPA partially reversed the response to subsequent treatment with 20 μM A β 25-35 for 24 h. Rhodamine 123 fluorescence percentage rate was 56% at 50 $\mu\text{g/ml}$ and 88.3% at 100 $\mu\text{g/ml}$ (both $P < 0.001$, as compared with the A β 25-35 model group). The results suggest that GSPA is able to reverse depolarized mitochondrial Ψ_m in PC12 cells.

Treatment with GSPA improves learning ability in APP/PS1 double transgenic mice. The results of the oriented navigation trials are presented in Table II. No significant differences in swim speed were detected among the groups (data not shown). The results of the MWM test demonstrated that APP/PS1 mutant mice exhibited significantly impaired learning ability in the water maze, as compared with the control group from

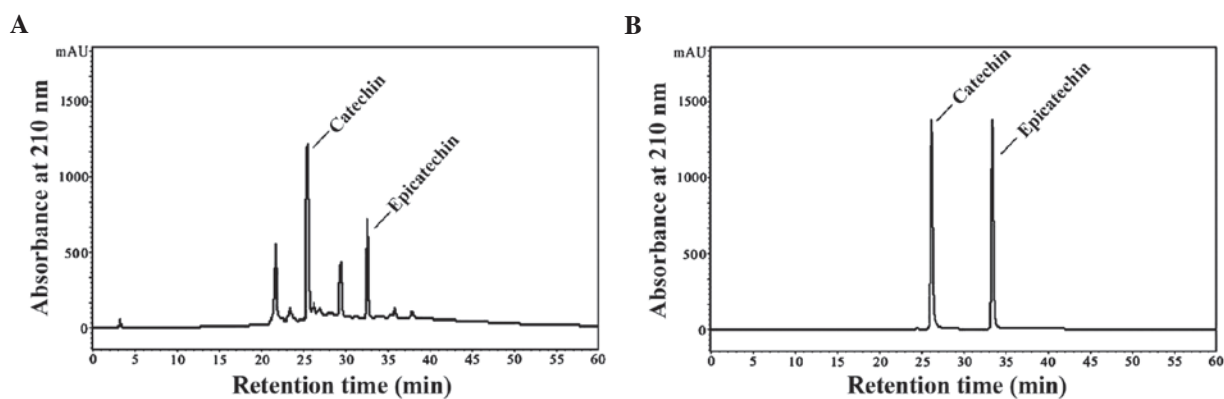


Figure 1. HPLC analysis of GSPA. (A) Normal-phase-HPLC chromatograms of GSPA. (B) Normal-phase-HPLC chromatograms of catechin and epicatechin. HPLC, high-performance liquid chromatography; GSPA, grape seed proanthocyanidin.

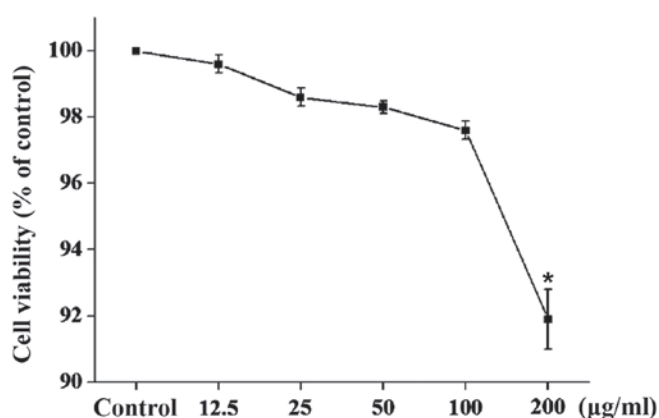


Figure 2. Grape seed proanthocyanidin treatment was non-toxic to PC12 cells at concentrations <200 µg/ml. Cell viability was assessed using a cell counting kit-8 reduction assay and expressed relative to untreated control cultures (100% viability). Data are expressed as the mean \pm standard error of the mean (n=6). *P<0.05, compared with the control group.

days 1-5 ($P<0.05$). The results of the spatial probe trials are shown in Fig. 6. As compared with the control group, the APP/PS1 model group exhibited significantly increased escape latency times ($P<0.01$) and a significantly decreased number of platform crossings ($P<0.01$). The APP/PS1 donepezil group exhibited significantly decreased latency ($P<0.01$) and a significantly increased number of platform crossings ($P<0.01$), as compared with the APP/PS1 model group. The APP/PS1-treated low and high dose GSPA groups also exhibited significantly reduced latency periods ($P<0.05$). Notably, the high dose GSPA group exhibited an increased number of platform crossings, as compared with the low dose group. Therefore, these results suggested that GSPA treatment may improve the learning ability of mice in a concentration-dependent manner. The results suggest that GSPA is able to improve learning ability in APP/PS1 double transgenic mice.

Treatment with GSPA alleviates amyloid plaques in the hippocampus of APP/PS1 double transgenic mice. HE staining revealed no remarkable neuronal abnormalities in the hippocampus of mice in the control group. The pyramidal cells in the CA1 region were arranged neatly and tightly, the

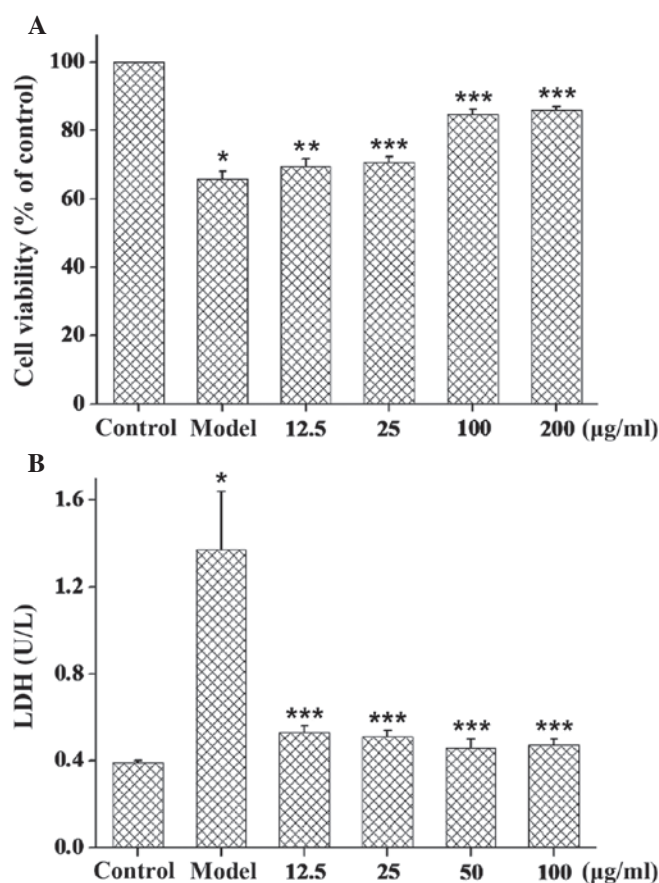


Figure 3. Grape seed proanthocyanidin partially reversed A β 25-35-induced cytotoxicity. (A) Cell viability was assessed using a cell counting kit-8 reduction assay. (B) Cell death was determined by LDH release into the culture media during amyloid β 25-35 treatment. Data are expressed as the mean \pm standard error of the mean (n=6). *P<0.01, as compared with the control group; **P<0.05 and ***P<0.01, as compared with the A β 25-35-treated group. LDH, lactate dehydrogenase.

cells were round and intact with clear dark blue stained nuclei. However, obvious hippocampal histopathological damage was demonstrated in the APP/PS1 model group. The pyramidal layered structure was disintegrated and neuronal loss was detected in the CA1 region. Neurons exhibited pyknotic nuclei with a shrunken or irregular shape. Following GSPA or donepezil hydrochloride oral administration for 2 months

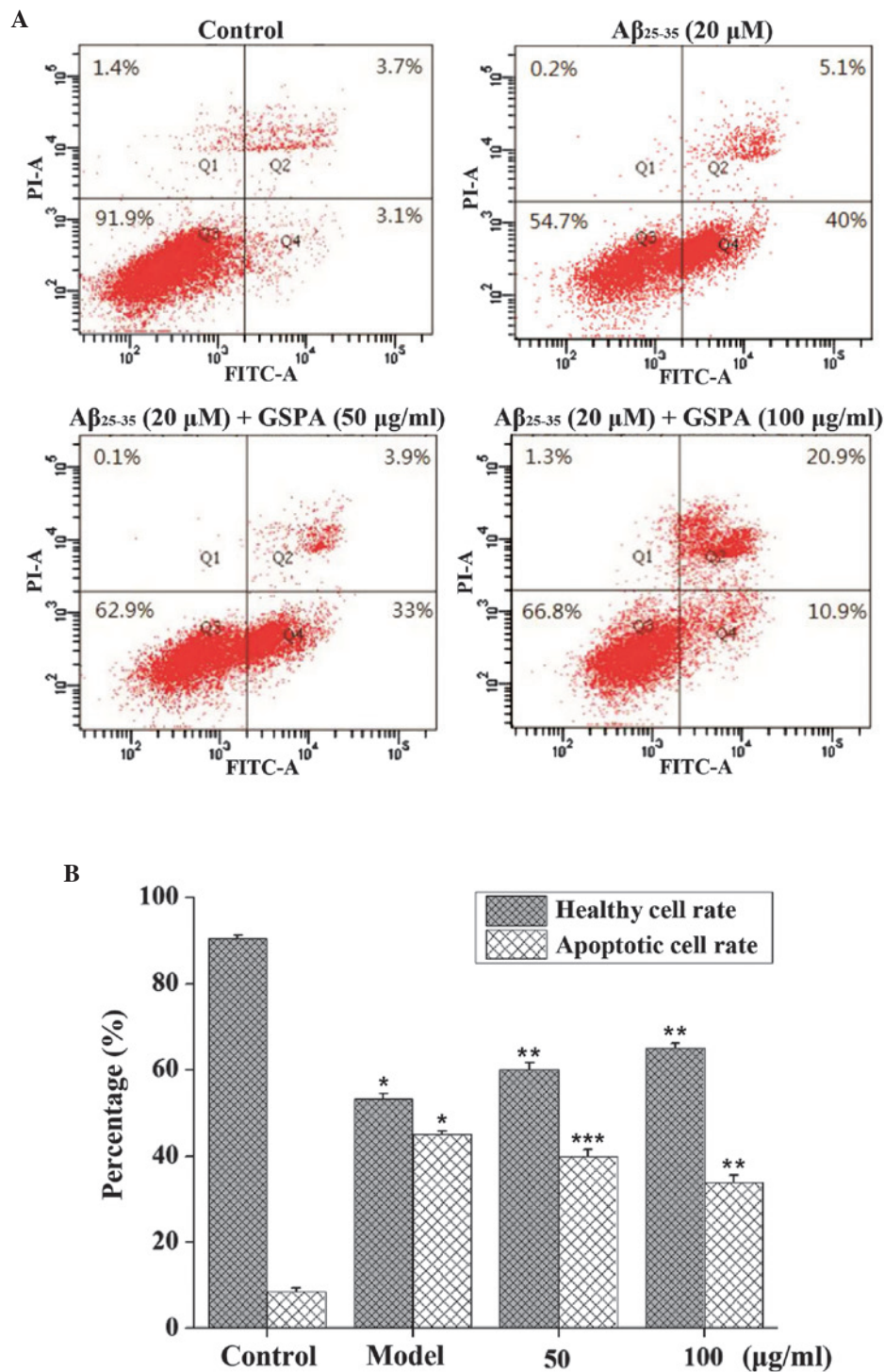


Figure 4. GSPA administration reduced A β ₂₅₋₃₅-induced apoptosis. (A) Individual values (%) are listed in representative dot plots and living PC12 cells were gated as shown in the fluorescence-activated cell sorting histograms for control PC12 cells, PC12 cells exposed to 20 μ M A β ₂₅₋₃₅, 20 μ M A β ₂₅₋₃₅ + 50 μ g/ml GSPA and 20 μ M A β ₂₅₋₃₅ + 100 μ g/ml GSPA. (B) A decrease in annexin-positive, PI-negative cells (lower right quadrant) and annexin-positive, PI-positive cells (higher right quadrant) was detected, which is indicative of reduced apoptosis. The increase in annexin-negative and PI-negative cells (lower left quadrant) is indicative of healthy non-apoptotic cells. Data are expressed as the mean \pm standard error of the mean (n=3). *P<0.01, as compared with the control group; **P<0.01 and ***P<0.05, as compared with the A β ₂₅₋₃₅-treated group. GSPA, grape seed proanthocyanidin; A β ₂₅₋₃₅, amyloid β ₂₅₋₃₅ peptide, PI, propidium iodide; FITC, fluorescein isothiocyanate.

these abnormalities were attenuated, as compared with the APP/PS1 model group. Cell morphology was regular, the cells were arranged more neatly and tightly, and cell nuclei stained clear. The APP/PS1-treated high dose GSPA group exhibited a stronger attenuation of these abnormalities, as compared with

the low group, suggesting a concentration-dependent effect (Fig. 7).

Congo red was used to stain the amyloid plaques in the hippocampus of APP/PS1 double transgenic mice. The results demonstrated that amyloid plaques were distributed in the

molecular layer of the hippocampus, and rare amyloid plaques were detected in the pyramidal cell layer. Amyloid plaques exhibited a light red dispersion without distinct boundaries and plaque staining was demonstrated to be denser in the hippocampi of the APP/PS1 model group, as compared with the hippocampi of the control group. The molecular layer exhibited an increased percentage of positive amyloid plaque areas, as compared with the control group. Plaque staining of APP/PS1 in the donepezil group and the low dose and high dose GSPA groups were lighter after GSPA or donepezil hydrochloride oral administration for 2 months. Positively stained areas of amyloid plaques were markedly reduced, as compared with the APP/PS1 model group (Fig. 8). The results suggest that GSPA is able to alleviate amyloid plaques in the hippocampus of APP/PS1 double transgenic mice.

Treatment with GSPA decreases APP and Tau protein expression levels in the hippocampi of APP/PS1 double transgenic mice. Western blot analyses of APP and tau protein expression levels are shown in Fig. 9. Low APP and tau protein expression levels were detected in the control group. Significantly increased APP and tau protein expression levels were detected in the hippocampi of mice in the APP/PS1 model group ($P < 0.05$ and $P < 0.01$, respectively). Treatment with oral donepezil hydrochloride for 2 months significantly decreased APP and tau protein expression levels in the hippocampi of the APP/PS1 donepezil group, as compared with the APP/PS1 model group ($P < 0.01$ and $P < 0.05$, respectively). Furthermore, 2-month administration of oral GSPA significantly decreased APP and tau protein expression levels in the hippocampi of mice in the APP/PS1-treated high dose group, as compared with the APP/PS1 model group ($P < 0.05$). The results suggest that GSPA is able to decrease APP and Tau protein expression levels in the hippocampi of APP/PS1 double transgenic mice.

Treatment with GSPA decreases PS-1 mRNA expression levels in the hippocampi of APP/PS1 double transgenic mice. Reverse transcription-quantitative polymerase chain reaction analysis of PS-1 mRNA expression levels was performed, as shown in Fig. 10. PS-1 mRNA expression levels were significantly increased in the APP/PS1 model group, as compared with the control group ($P < 0.01$). PS-1 expression levels in the APP/PS1 donepezil and high dose GSPA groups were significantly decreased, as compared with the APP/PS1 model group (both $P < 0.01$). The results suggest that GSPA is able to decrease PS-1 mRNA expression levels in the hippocampi of APP/PS1 double transgenic mice.

Discussion

AD is a growing public health concern with devastating impacts. The disease is characterized by the hallmarks of A β peptide accumulation with hyperphosphorylation and aggregation of tau protein. Previous studies have indicated that the accumulation of neurotoxic A β peptides has a causal role in AD dementia and memory deficits in the Tg2576 mouse AD model (32,33). It has also been demonstrated that certain GSPEs may interfere with the aggregations of synthetic A β peptides *in vitro* (34). Furthermore, according to the results

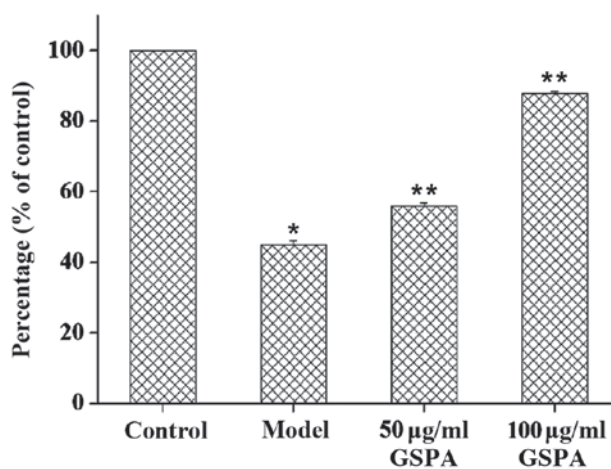


Figure 5. Grape seed proanthocyanidin suppressed mitochondrial membrane depolarization. Reduced rhodamine 123 fluorescence is indicative of mitochondrial membrane depolarization. Data are expressed as the mean \pm standard error of the mean (n=3). * $P < 0.01$ compared to the control group; ** $P < 0.01$ compared to the A β 25-35 model group.

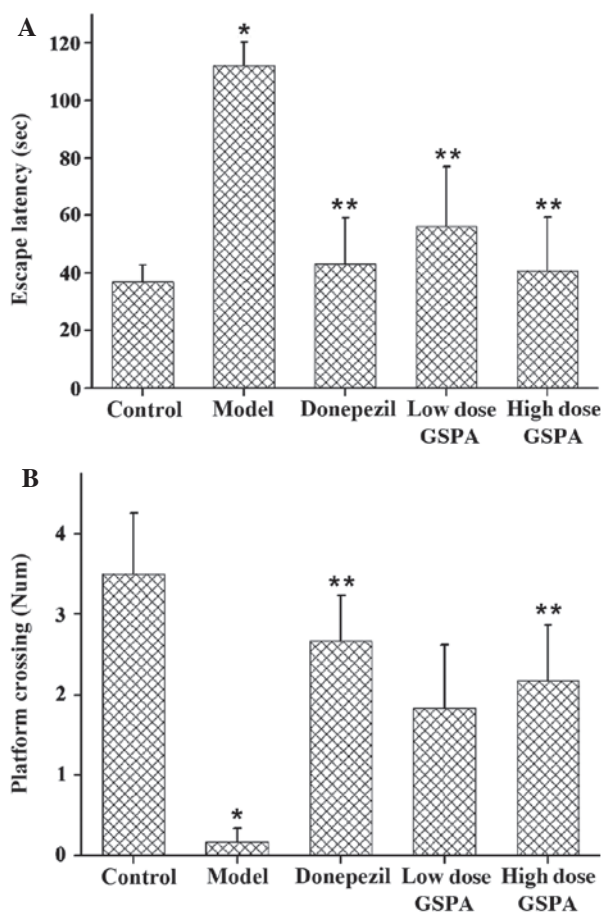


Figure 6. Comparisons of (A) average escape latency and (B) platform crossing in the spatial probe trials of amyloid precursor protein/ presenilin-1 heterozygous mouse model groups. Data are expressed as the mean \pm standard error of the mean (n=6). * $P < 0.01$, as compared with the control group; ** $P < 0.05$, as compared with the model group. GSPA, grape seed proanthocyanidin.

of a previous binding assay, GSPE is capable of inhibiting tau peptide polymerization and scattering pre-aggregated tau peptide (35).

Table II. Improved learning ability in amyloid precursor protein/presenilin-1 double transgenic mice treated with grape seed proanthocyanidin (GSPA), as determined by the Morris water maze test.

Group	Time taken to find the platform (sec)				
	Day 1	Day 2	Day 3	Day 4	Day 5
Control	117.50±2.50	75.83±20.04	67.17±23.70	58.17±20.02	52.67±11.11
Model	120.00±0.00	117.00±3.00	102.50±11.34	114.17±5.83	107.17±8.64 ^a
Donepezil	120.00±0.00	102.33±17.67	81.67±13.64	78.83±18.36	57.67±14.05 ^b
Low GSPA	120.00±0.00	97.67±7.31	88.33±13.18	81.67±10.84	70.17±8.66 ^b
High GSPA	120.00±0.00	80.00±14.09	82.50±13.20	74.50±8.72 ^b	67.83±3.96 ^b

Data are expressed as the mean ± standard error of the mean. ^aP<0.05, as compared with the control group; ^bP<0.05, as compared with the model group.

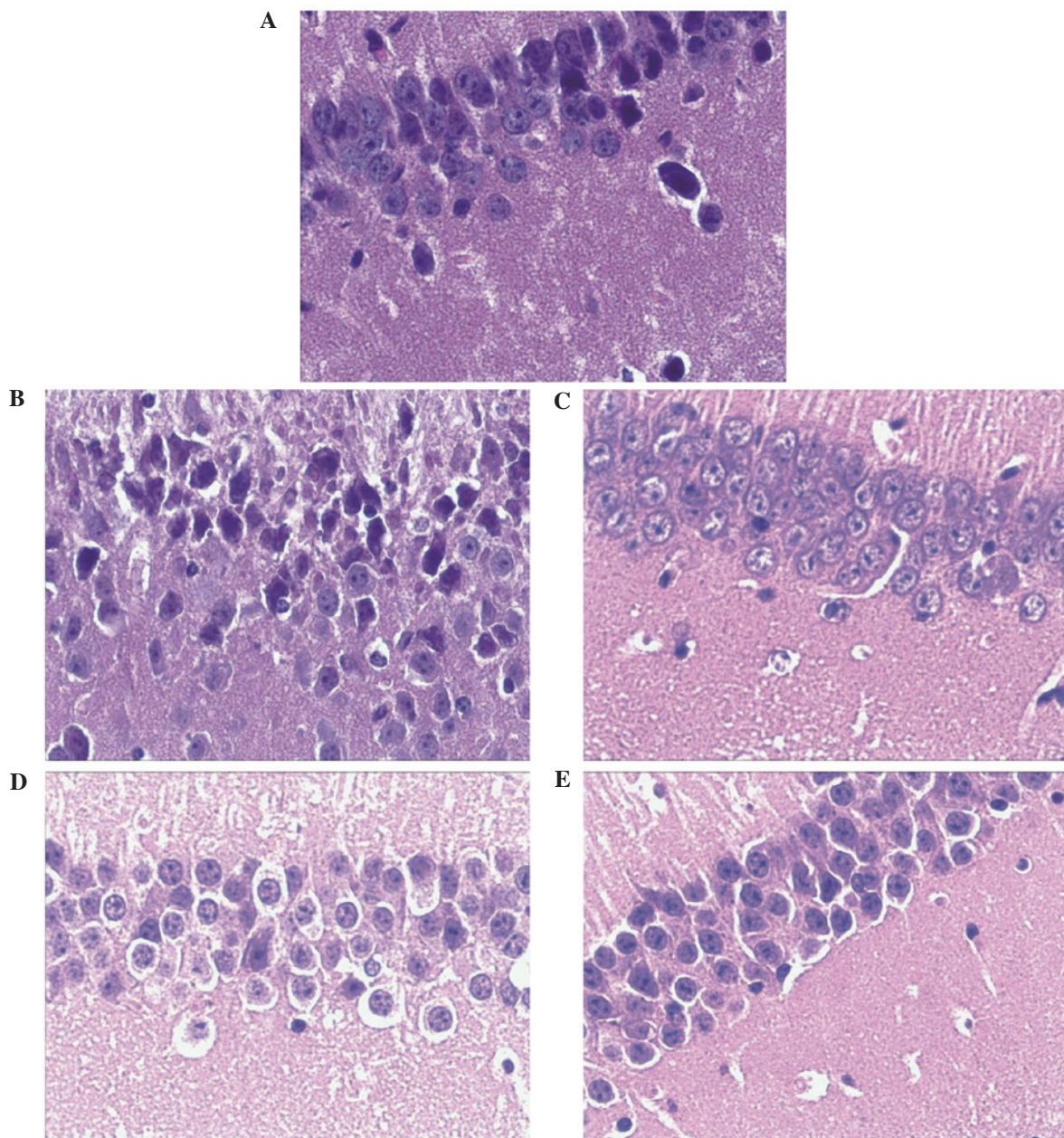


Figure 7. Hematoxylin and eosin staining (magnification, x400) of the (A) control; (B) APP/PS1 model; (C) APP/PS1 plus donepezil; (D) APP/PS1 plus low dose GSPA (50 mg/kg/day); and (E) APP/PS1 plus high dose GSPA (100 mg/kg/day) groups. APP, amyloid precursor protein; PS1, presenilin-1; GSPA, grape seed proanthocyanidin.

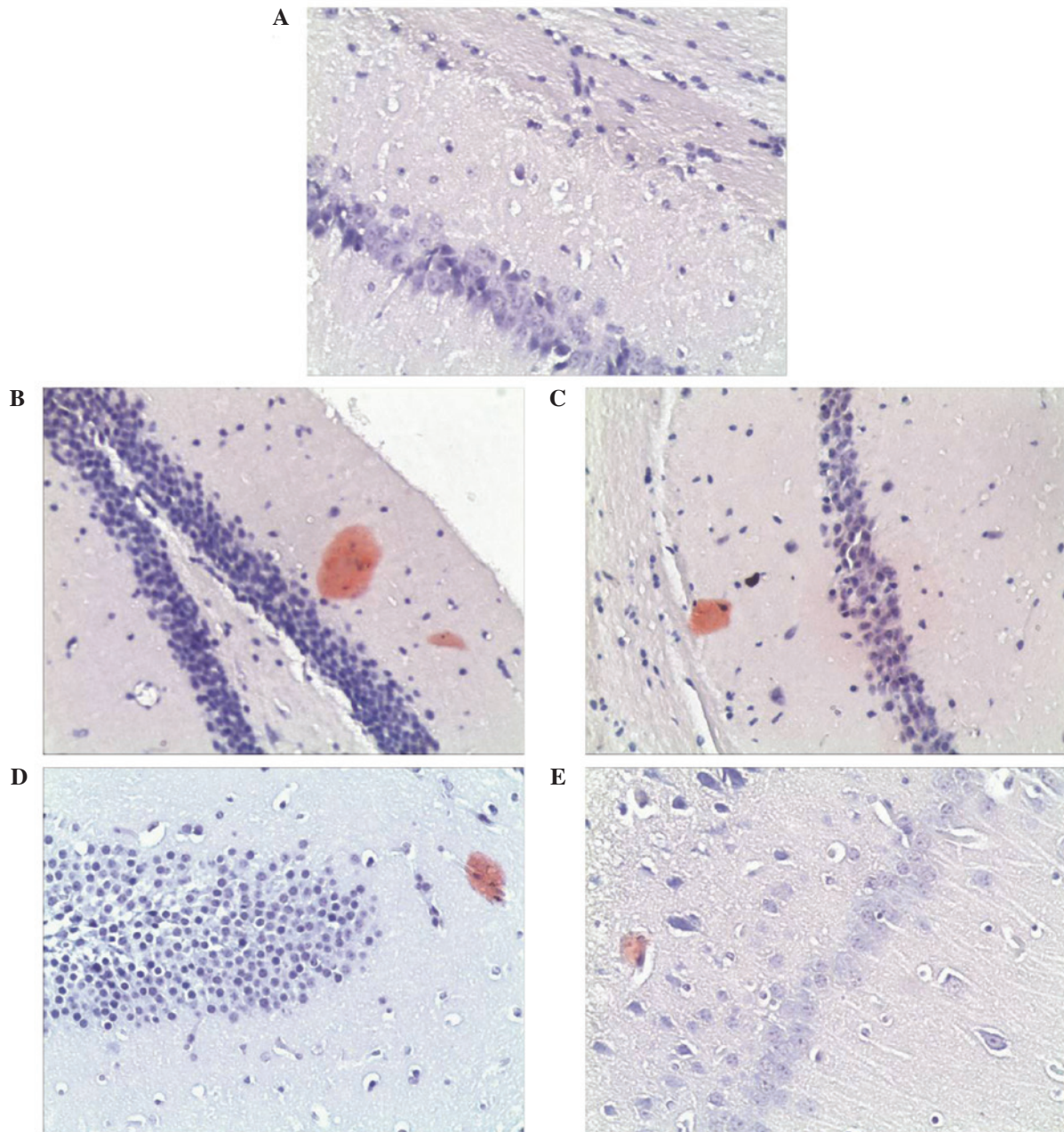


Figure 8. Congo red staining (magnification, x200) of the (A) control; (B) APP/PS1 model; (C) APP/PS1 plus donepezil; (D) APP/PS1 plus low dose GSPA (50 mg/kg/day); and (E) APP/PS1 plus high dose GSPA (100 mg/kg/day) groups. APP, amyloid precursor protein; PS1, presenilin-1; GSPA, grape seed proanthocyanidin.

Oxidative stress has an important role in the pathophysiology of AD. Previous studies have indicated that oxidative stress leads to apoptosis and excessive ROS facilitates neuronal apoptosis in A β -induced neuronal cell death (36,37). Moreover, it has been demonstrated that the overproduction of A β leads to A β -associated free-radical production and cell death (37,38). These findings indicate that A β induces oxidative stress and vice versa, which leads to more oxidative damage, including membrane damage and reduced cell viability. When cell membranes are damaged, LDH levels increase. The present study demonstrated that GSPA is able to reverse A β 25-35-induced LDH leakage and decreased PC12 cell viability (39). These results demonstrated that GSPA is capable of scavenging oxygen

radicals. Therefore, the protective effect of GSPA against A β 25-35 induced oxidative stress in PC12 cells may be due to enhanced anti-oxidant capacity and reduced accumulation of intracellular ROS.

Cell apoptosis has a central role in the pathogenesis of AD. Oxidative stress induces apoptotic cell death via the activation of caspase 3 (40). In the present study, Annexin V-FITC and PI double staining was used to investigate basal apoptosis rates. PC12 cells treated with extracellular A β 25-35 at concentrations of 20 μ M exhibited significantly increased levels of apoptosis, as compared with the control cells. These data suggested that A β 25-35 may increase caspase 3 expression levels and induce oxidative stress, leading to apoptosis. However, treatment with 50 or 100 μ g/ml GSPA significantly decreased cell apoptosis.

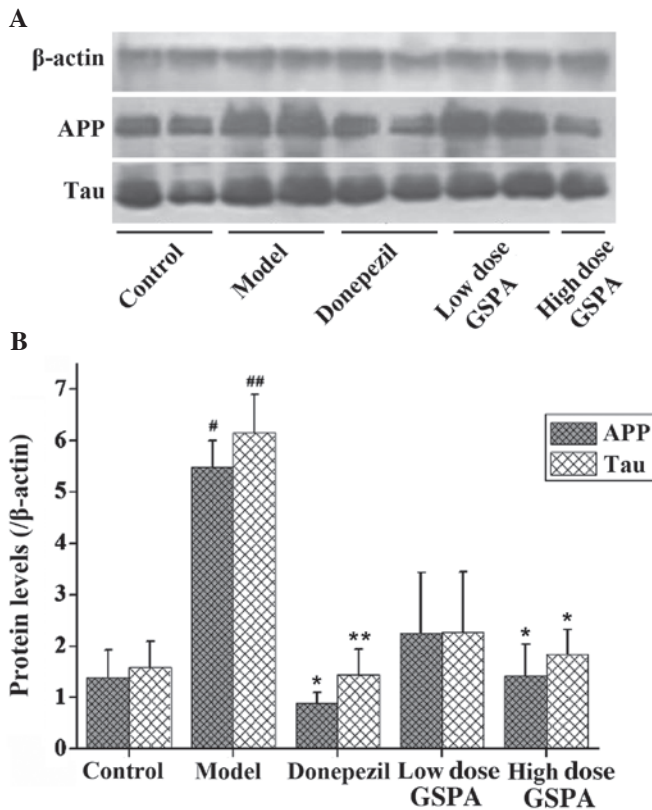


Figure 9. (A) Western blot analysis of APP and Tau protein among the control, APP/PS1 model, APP/PS1 plus donepezil and APP/PS1 plus low (50 mg/kg/day) and high (100 mg/kg/day) dose GSPA groups, respectively. (B) Quantitative comparison of the protein levels of APP and Tau among the group, relative to β -actin. Data are expressed as the mean \pm standard error of the mean (n=6) mice. [#]P<0.05 and ^{##}P<0.01, as compared with the control group; ^{*}P<0.05 and ^{**}P<0.01, as compared with the APP/PS1 model group.

This result may be associated with decreasing levels of caspase 3 and oxidative stress.

A significant reduction of the mitochondrial Ψ_m can trigger apoptosis through the release of caspase 3. Previous studies have demonstrated that A β ₂₅₋₃₅ leads to oxidative stress and mitochondrial damage due to the loss of mitochondrial Ψ_m (41,42). Consistent with a previous study (43), the results of the present study demonstrated a significant decrease in mitochondrial Ψ_m following stimulation with extracellular A β ₂₅₋₃₅, as compared with the control cells, using rhodamine 123 fluorescence. Pretreatment of PC12 cells with GSPA reversed the mitochondrial Ψ_m depolarization induced by A β ₂₅₋₃₅, which suggested that GSPA may prevent cell death by blocking the activation of the mitochondrial apoptosis pathway.

In the present study, the significant cytotoxic effect of A β ₂₅₋₃₅ on PC12 cells was demonstrated by the CCK-8 assay, and Annexin V-FITC and PI double staining with flow cytometry. It was hypothesized that activation of caspase 3 induced apoptosis. This result may be associated with oxidative stress and stress-related damage to the mitochondria or its membranes. Notably, GSPA may reverse the A β ₂₅₋₃₅-induced increase of ROS and decrease of mitochondrial Ψ_m , which ultimately protected the cells from apoptosis.

The results of the present study also demonstrated that oral administration of GSPA significantly reduced the

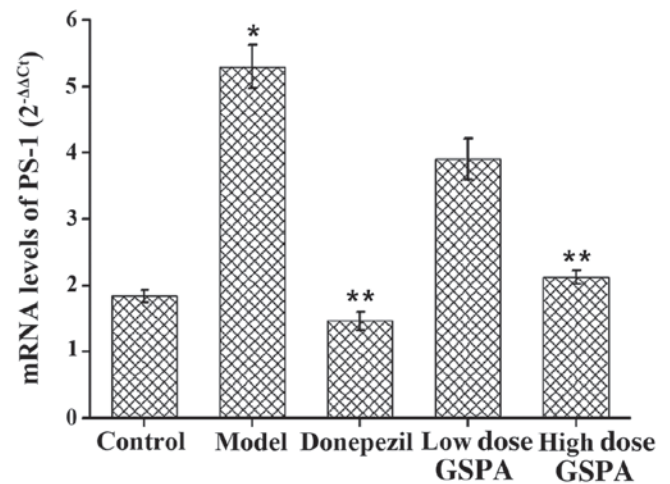


Figure 10. Reverse transcription-quantitative polymerase chain reaction analysis. Comparison of PS-1 mRNA expression levels among the control, APP/PS1 model, APP/PS1 plus donepezil and APP/PS1 plus low (50 mg/kg/day) and high (100 mg/kg/day) dose GSPA groups, respectively. Data are expressed as the mean \pm standard error of the mean (n=6). ^{*}P<0.01, as compared with the control group; ^{**}P<0.01, as compared with the APP/PS1 model group. APP, amyloid precursor protein; PS1, presenilin-1; GSPA, grape seed proanthocyanidin.

accumulation of A β peptides, alleviated the hyperphosphorylation of tau protein and improved learning behavior and memory in APP/PS1 double transgenic mice. The mechanisms underlying these phenomenon may involve the downregulation of A β accumulation and the disruption of tau protein hyperphosphorylation in the brain.

APP/PS1 mice exhibited a significant loss of learning, cognition and memory behavior in the present study, as compared with the control mice. These results are consistent with previous studies, which have demonstrated APP/PS1 mice with early onset brain amyloidosis, and synaptic changes in the hippocampus and brain cortex (44,45). The MWM test is a well-known behavioral task which is used to assess oriented navigation and spatial probe memory capacity (46). During the spatial probe phase of the MWM test in the present study, the GSPA treatment group exhibited significant decreased escape latency periods and an increased number of platform crossing incidences, as compared with APP/PS1 model group. These data demonstrated that the GSPA-treated APP/PS1 mice exhibited an enhanced capacity for learning, cognition and memory. We hypothesize that this effect may be induced by a reduction in A β in the brain, as excessive toxic A β has been demonstrated to induce the formation of A β plaques (47). HE and Congo red staining was used to detect amyloidosis in the hippocampi of the mice. The APP/PS1 model group exhibited a significant increase in the number of A β plaques in the hippocampus, as compared with the control group. Notably, GSPA treatment significantly ameliorated these A β plaques.

The results of the present study have demonstrated that mutations in the APP and PS genes increase oxidative stress in the neurons of APP/PS1 mice, and oxidative stress is mediated by A β . The A β protein is critical to the pathogenesis of AD (48,49); however, the precise mechanism underlying the pathogenesis of AD is yet to be fully elucidated. Therefore,

one of the aims of preventive or early treatment of AD is to decrease oxidative stress. A β peptides are produced from the APP, which are cleaved by β - and γ - secretases into amino acid peptides within the cerebral cortex and hippocampus (3). γ -secretase is an intramembrane aspartyl protease that is critically involved in AD via the proteolysis of APP, which generates the pathogenic and amyloid plaque-forming A β 1-42 peptide (50). PS-1 is a member of a multi-subunit intramembranous protein complex with γ -secretase; thus, a reduction in PS-1 mRNA expression levels directly suppresses the activation of γ -secretase. PS-1 also stimulates tau protein aggregation into abnormally hyperphosphorylated tau via A β (51). Therefore, it was hypothesized that restraining PS-1 mRNA and APP protein expression levels via decreased oxidative stress may be a viable strategy for AD therapy. In the present study, western blot and RT-qPCR analyses were used to confirm that treatment with GSPA significantly decreased PS-1 mRNA expression levels. The effects of GSPA on PS-1 mRNA expression and γ -secretase activation may interrupt the generation of the A β peptide. Moreover, the results of the present study demonstrated that oral treatment with GSPA significantly reduced the accumulation of APP and insoluble tau in the brains of APP/PS1 mice. GSPA may function as a strong antioxidant by decreasing the levels of APP and tau protein. This phenomenon may be associated with: i) the strong antioxidant action of GSPA suppressing the activation of γ - secretases to reduce A β production; or ii) GSPA directly interfering with the aggregation of A β or dissociated aggregation of A β into oligomers, which may reduce oxidative stress and restore the antioxidant balance.

In conclusion, the present preclinical study demonstrated that GSPA treatment attenuates APP production or accumulation, interrupts the aggregation of neurotoxic A β and decreases hyperphosphorylated tau deposition. Notably, GSPA treatment improved the cognition and memory capacity of a APP/PS1 double transgenic mouse model. The results of the present *in vitro* biochemistry studies and *in vivo* preclinical studies suggested that GSPA administration may benefit AD via two, non-exclusive mechanisms: i) enhanced antioxidant function and ii) downregulation of caspase 3-mediated aggregation of A β in response to oxidative stress. The results of the present study provide support for continuing the development of GSPA for the treatment and/or prevention of AD.

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