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Review article

Oxidative stress and inflammation contribute to traffic noise-induced vascular and cerebral dysfunction via uncoupling of nitric oxide synthases

Andreas Daiber^{a,b,*}, Swenja Kröller-Schön^a, Matthias Oelze^a, Omar Hahad^{a,b}, Huige Li^c, Rainer Schulz^d, Sebastian Steven^a, Thomas Münzel^{a,b,*}

^a Center for Cardiology, Molecular Cardiology, University Medical Center, Mainz, Germany

^b Partner Site Rhine-Main, German Center for Cardiovascular Research (DZHK), Langenbeckstr. 1, 55131, Mainz, Germany

^c Department of Pharmacology, University Medical Center, Mainz, Germany

^d Institute of Physiology, Justus-Liebig University, Giessen, Germany

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ABSTRACT

Environmental pollution and non-chemical stressors such as mental stress or traffic noise exposure are increasingly accepted as health risk factors with substantial contribution to chronic noncommunicable diseases (e.g. cardiovascular, metabolic and mental). Whereas the mechanisms of air pollution-mediated adverse health effects are well characterized, the mechanisms of traffic noise exposure are not completely understood, despite convincing clinical and epidemiological evidence for a significant contribution of environmental noise to overall mortality and disability. The initial mechanism of noise-induced cardiovascular, metabolic and mental disease is well defined by the "noise reaction model" and consists of neuronal activation involving the hypothalamicpituitary-adrenal (HPA) axis as well as the sympathetic nervous system, followed by a classical stress response via cortisol and catecholamines. Stress pathways are initiated by noise-induced annoyance and sleep deprivation/fragmentation. This review highlights the down-stream pathophysiology of noise-induced mental stress, which is based on an induction of inflammation and oxidative stress. We highlight the sources of reactive oxygen species (ROS) involved and the known targets for noise-induced oxidative damage. Part of the review emphasizes noise-triggered uncoupling/dysregulation of endothelial and neuronal nitric oxide synthase (eNOS and nNOS) and its central role for vascular dysfunction.

1. Introduction

Environmental stressors such as air pollution, noise, mental stress or climate changes are increasingly recognized as important determinants of our health. According to health experts, environmental exposure to stressors (the "exposome") even outcompetes genetic health determinants and joins classical health risk factors such as obesity and diabetes in their health impact [1,2]. As concluded by the Lancet Commission on pollution and health "Pollution is the largest environmental cause of disease and premature death in the world today. Diseases caused by pollution were responsible for an estimated 9 million premature deaths in 2015 - 16% of all deaths worldwide - three times more deaths than from AIDS, tuberculosis, and malaria combined" [3]. These data are also supported by recent World Health Organization (WHO) reports

[4,5]. The numbers are even more dramatic for the global burden of non-communicable chronic diseases since all forms of pollution cause an estimated 268 million disability adjusted life years annually (consisting of 254 million years of life lost and 14 million years lived with disability) [6]. With respect to disability adjusted life years in Europe, noise ranks second to air pollution in the list of environmental triggers, and outcompetes lead, ozone and dioxins [7]. In spite of this growing evidence, noise is not mentioned in any global health action plans, with only occupational noise being mentioned as a significant environmental health risk factor in the Global Burden of Disease (GBD) Study [8]. Traffic noise exposure is a clearly underestimated environmental risk factor, and contributes substantially to the development of cardiometabolic complications such as ischemic heart disease, hypertension, heart failure, and thereby also to global mortality and disability

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Abbreviations: BH4, tetrahydrobiopterin; HPA, hypothalamic-pituitary-adrenal; IL, interleukin; L-NAME, L-N^G-nitro-arginine methyl ester; NOS, nitric oxide synthase; NOX, NADPH oxidase; PKC, protein kinase C; RAAS, renin-angiotensin-aldosterone system; ROS, reactive oxygen species; SOD, superoxide dismutase

^{*} Corresponding author. Universitätsmedizin der Johannes Gutenberg-Universität Zentrum für Kardiologie 1 – Labor für Molekulare Kardiologie, Geb. 605 – Raum 3.262, Langenbeckstr. 1, 55131, Mainz, Germany.

E-mail addresses: daiber@uni-mainz.de (A. Daiber), tmuenzel@uni-mainz.de (T. Münzel).

adjusted life years [9–11]. This review provides mechanistic insights on noise-induced mental stress reactions, inflammation, oxidative stress and neuronal/vascular damage with an emphasis on the uncoupling of nitric oxide synthases (NOS).

2. Noise research and noise reaction model

The majority of noise research relates to the direct adverse effects on the auditory system, which is documented noise-induced hearing loss, and is not further discussed in this review. A mechanistic similarity between noise-induced hearing loss (typically observed upon noise exposure to sound pressure levels $\geq 100 \text{ dB}(\text{A})^1$) and the non-auditory effects of noise (typically studied with sound pressure levels $\leq 80 \text{ dB}$ (A)) is that both harms are accompanied by increased production of reactive oxygen species (ROS) [12]. Thus, hearing loss was improved by antioxidant treatment in experimental models [13] and noise-induced non-auditory effects such as endothelial dysfunction were improved by vitamin C infusion in healthy subjects [14].

The non-auditory effects of noise rely mainly on a neuronal component and the individual perception of noise exposure, and their health consequences were reviewed in a monograph "Effects of Noise in Man" by Karl D. Kryter in 1970 [15]. This monograph discusses the effects of noise on work performance, sleep, pain sensitivity, vision and blood circulation. Kryter postulated that noise initiates secondary reactions by stimulation of the neuronal system including the autonomic/ reticular nervous systems and (sub)cortical brain centers. A detailed overview on historic development of noise research can be found in Ref. [12].

The modern noise reaction model for non-auditory noise effects was established by Babisch, which comprises an "indirect pathway" based on the disturbance of sleep, communication and activity that is mainly triggered by low level noise exposure (even by sound pressure levels of 50-60 dB(A)) thereby causing changes of emotional and cognitive parameters and annoyance, followed by the adverse health effects described above [16,17]. The contribution of noise-induced activation of the hypothalamic-pituitary-adrenal (HPA) axis as well as the sympathetic nervous system response to these stress reactions leads to a chronic release of stress hormones such as cortisol and catecholamines as well as the subsequent induction of inflammation (with increases in interleukin [IL]-6, IL-1 β and proinflammatory monocytes) [18,19] and oxidative stress [14,20,21] (Figs. 1 and 2). Noise also triggers anxiety disorders and depression [22] as well as annoyance and atrial fibrillation [23]. The severe adverse health impact of noise-induced mental stress is documented by cases of stress-induced cardiomyopathy (so called Takotsubo Syndrome or broken heart disease) [24], which has been described as a consequence of a nighttime aircraft noise exposure [25]. The increase of glucocorticoid levels by mental stress also leads to endothelial dysfunction characterized by reduced vascular endothelial nitric oxide synthase (eNOS) function and nitric oxide ('NO) bioavailability [26] as well as increased sensitivity of the vasculature to vasoconstriction action of the sympathetic nervous system [27].

Some molecular links between mental stress induced by noise exposure (HPA axis, sympathetic nervous system and renin-angiotensinaldosterone system (RAAS) activation) and the induction of vascular

and cerebral inflammation as well as oxidative stress are illustrated in Fig. 1 (also reviewed in Ref. [19,20]). Severe life stress, including sleep deprivation as induced by noise exposure during sleep, leads to cerebral oxidative stress, which is mediated by angiotensin-II signaling and NADPH oxidase isoform 2 (NOX-2) activation and microvascular as well as neuronal inflammation [28]. Increased circulating levels of angiotensin-II is an important stress biomarker in noise exposed animals [21,29]. Mental stress induced by sleep deprivation and frequently interrupted sleep impairs the cerebral redox balance and leads to abnormal (manic-like) behavior and memory impairment, which is improved by treatment with the antioxidant quercetin or TEMPOL [28,30,31]. Oxidative stress in turn activates the sympathetic nervous system in human [32] and animal studies [33,34]. In addition, administration of catecholamines induces oxidative stress in rats, probably by serving as substrates for monoamine oxidases (MAO) [35] or activation of astrocytes, microglia and NOX-2 [36]. Inhibition of NADPH oxidase (several NOX isoforms) activity in hypertensive mice (RAAS activation model) decreases blood pressure and circulating concentrations of angiotensin-II and noradrenaline [37], whereas AT1receptor blockers and angiotensin-converting enzyme inhibitors decrease cardiovascular oxidative stress [38,39], clearly indicating an interaction of the RAAS and ROS levels.

3. Traffic noise exposure activates ROS sources and causes oxidative damage

3.1. Noise-triggered inflammation and activation of immune cells – role of phagocytic NADPH oxidase (NOX-2)

Low levels of road traffic noise cause immunological alterations (e.g. higher IL-12 levels and lower natural killer cell populations/activity) that are associated with increased circulating cortisol levels, noise sensitivity and the health status of affected individuals [40]. Traffic noise also increases high-sensitivity C-reactive protein (hsCRP) levels [41]. Train noise induces plasma proteome changes in healthy subjects leading to a pro-inflammatory, pro-oxidant and pro-atherothrombotic phenotype [42]. These data are in good harmony with the established correlation between acute psychological stress conditions, hormone levels, recruitment/activation of immune cells and impaired cardiovascular function in humans, where the highest cardiovascular event rates were associated with most pronounced immunological changes [43,44]. Noise exposure augments neuroinflammation and Alzheimer's disease pathology in rodent studies [45], which is in keeping with our recent observations in mice indicating that low level noise exposure (mean sound pressure level 73 dB(A)) increases circulating cytokines, aortic inducible nitric oxide synthase (iNOS)/monocyte chemoattractant protein-1 (MCP-1, CCL2) mRNA levels and vascular infiltration of immune cells [21,46]; this was accompanied by neuroinflammation characterized by astrocyte activation and higher cerebral CD68,² IL-6 and iNOS levels [46] (Fig. 2).

In addition, the activation of immune cells by noise as measured by an oxidative burst (a parameter for leukocyte NOX activity) was more pronounced [46] and there was a greater abundance of NOX-2 in the vasculature of noise-exposed mice [21], leading to increased vascular and cerebral ROS formation as well as vascular (endothelial) dysfunction. The concept that ROS from inflammatory cells are responsible for noise-induced oxidative stress is further supported by a marked reduction of noise-induced systemic oxidative stress (e.g. aortic superoxide formation as measured by HPLC analysis of 2-hydroxyethidium)

¹ The sound pressure level is measured in the dimension Decibel (dB) on a logarithmic scale. It is interconvertible to the sound pressure that is measured in the dimension Pascal (Pa). A whisper has a sound pressure of 0.000063 Pa, which is equivalent to a sound pressure level of 10, whereas the traffic on a highway may reach a sound pressure of 0.2 Pa, which is equivalent to a sound pressure level of 80 dB. The sound pressure level is expressed by either dB or dB (A), where dB(A) means Decibel A-weighted and accounts for the different frequency sensitivity of the human ear (e.g. reduced sensitivity for low audio frequencies). The Decibel A-weighted value compensates for this variation in the human hearing sensitivity and most commercial acoustic measurement devices express the sound pressure level as dB(A).

² CD68 (Cluster of Differentiation 68, macrosialin) is a protein that is highly expressed by cells in the monocyte lineage (e.g. monocytic phagocytes, osteoclasts), by circulating macrophages, and by tissue macrophages (e.g. Kupffer cells, microglia). CD68 is involved in cell adhesion and recruitment and activation of macrophages for phagocytosis or removal of cell debris.



Fig. 1. Noise exposure induces neuronal activation. First line neuronal events in response to noise exposure are sleep disturbance (when exposed during the sleep phase) and stress response reactions via activation of the hypothalamic-pituitary-adrenal (HPA) axis and the sympathetic nervous system. This leads to release of stress hormones (glucocorticoids and catecholamines) and secondary activation of the cerebral (and systemic) renin-angiotensin-aldosterone system (RAAS) as well as endothelin-1 expression. These potent triggers of inflammation and oxidative stress will cause activation of NOX-2 via protein kinase C (PKC) and p47^{phox} phosphorylation in the brain, expression of markers of inflammation, increased lipid peroxidation, down-regulation of neuronal nitric oxide synthase (nNOS) and loss of antioxidant genes such as catalase (Cat) and forkhead box O3 (Foxo3) transcription factor. All of these changes induce a neuroinflammatory phenotype with increased cerebral oxidative stress. These stress hormones and vasoconstrictors lead to similar adverse changes in the cardiovascular (and pulmonary) system, finally resulting in cardiometabolic disease. The HPA axis, sympathetic nervous system, RAAS, endothelin-1 expression and neuroinflammation are

redox regulated and vice versa can induce oxidative stress via NOX-2 activation and other sources. AT-II, angiotensin-II; CRH, corticotrophin-releasing hormone; ACTH, adrenocorticotrophic hormone; CVD, cardiovascular disease.



Fig. 2. Noise-induced neuronal activation induces cardiovascular ROS sources and oxidative damage as well as inflammation (summary of animal and human data). The stress hormones and vasoconstrictors have direct effects on the vasculature, the heart and immune cells via their receptors. Noise-induced sleep deprivation causes dysregulation of the circadian clock (e.g. changes in expression of period and cryptochrome genes), in part by adverse redox regulation of clock genes, which negatively affects inflammation and redox balance. In the heart, noise upregulated markers of oxidative stress (3-NT and 8-OH-dG), eNOS and NOX-2 activity, whereas noise down-regulated the antioxidant factor Foxo3 and the structural component connexin43 (Cx43), all of which indicated cardiac remodeling and fibrosis. In endothelial cells isolated from mouse lungs (MLEC), antioxidant defense genes (Hmox1 and Pgc-1a) as well as Nox1 and Nos3 (eNOS) were upregulated in the noise group suggesting counter-regulatory mechanisms. In the aorta, noise increased oxidative protein damage (3-NT and 4-HNE), markers of inflammation (e.g. iNOS, Cd68, Mcp1 and Vcam1) and NOX-2 activity. While eNOS expression was upregulated in aortic tissue of noise-exposed animals, 'NO bioavailability was decreased as well as phosphorylation of the vasodi-

lator-stimulated phosphoprotein (P-VASP), which is a read-out of the 'NO/cyclic guanosine-3',5'-monophosphate (cGMP)/protein kinase G (PKG) signaling cascade. Also aortic oxidative stress was directly measured and immune cell infiltration (mainly CD11b⁺ cells, not shown in the scheme) was quantified - both were increased upon noise exposure. Circulating markers of oxidative stress (MDA, 3-NT, 8-isoP) and inflammation (IL-6, IL-1β, IL-12, hsCRP) were increased upon noise exposure, whereas levels of reduced glutathione (GSH) were decreased. Circulating white blood cells had more oxidative DNA damage (8-OH-dG), compensatory increase in glutathione peroxidase 1 (GPx-1) and displayed more pronounced oxidative burst in noise exposed animals or subjects. Mitochondria of noise exposed animals showed increased swelling of the membranes, cristolysis and mtDNA damage, impaired function of the mitochondrial permeability transition pore (mPTP) and calcium handling as well as monoamine oxidase (MAO) activation by high noradrenaline (NA) levels due to noise exposure. Direct impact of hormones besides vasoconstriction is mentioned in the text boxes. The adverse redox changes and inflammatory signaling by noise promote the development of cardiometabolic disease. Pharmacological activation of FOXO3 and genetic Nox2 deletion largely prevent noise-induced adverse health effects. AT-II, angiotensin-II; ET-1, endothelin-1; 3-NT, 3-nitrotyrosine; 4-HNE, 4-hydroxynonenal; MDA, malondialdehyde; 8-isoP, 8-isoprostane; 8-OH-dG, 8-hydroxy-2/deoxyguanosine; hsCRP, high sensitivity Creactive protein.

by genetic deletion of the *Nox2* gene ($Nox2^{-/-}$) or pharmacological inhibition (GSK2795039) of the NOX-2 protein [46]. In the brain of around-the-clock noise exposed mice, protein kinase C (PKC) activity (as measured by higher phospho-MARCKS levels) and PKC-dependent

p47^{phox} phosphorylation at serine 328 were increased, which is a marker for enhanced translocation of cytosolic p47^{phox} to the membrane reflecting increased NOX-2 activity [46] (Fig. 1). The activation of PKC is most likely due to higher circulating (and cerebral) levels of

angiotensin-II in response to noise, leading to enhanced formation of diacylglycerol (upon activation of the angiotensin II type 1 (AT1)-receptor), a potent PKC activator [47]. In addition, PKC can be activated by oxidative modifications of the zinc-sulfur complex in the phorbol ester/diacylglycerol binding domain [47]. Importantly, inflammatory pathways and immune cell activation are triggered by oxidative stress (e.g. by redox changes in central players of inflammatory cascades such as NOD-, LRR- and pyrin domain-containing protein 3 (NLRP3) and high mobility group box 1 (HMGB1) protein) as extensively reviewed previously [48–50].

Aircraft noise increases the expression of endothelin-1 (ET-1) in the aorta of mice and enhances sensitivity to ET-1 mediated vasoconstriction [21,46] (Fig. 2). Of note, ET-1, in addition to being a potent vasoconstrictor, also increases NOX-2 activity [51,52]. Likewise, oxidative stress and NOX-2-derived ROS activate the ET-1 promoter to increase its expression [53,54], so initiating a self-promoting crosstalk between NOX-2 and endothelin-1 [55]. ET-1 also interacts with the RAAS, as demonstrated by increased vascular expression of ET-1 in angiotensin-II induced hypertensive rats [56] or by reduction of blood pressure and circulating angiotensin-II levels in bosentan (ET_{A/B} receptor blocker)-treated animals with fructose-dependent hypertension [57]. Increased ET-1 signaling is also proposed to contribute to the unique susceptibility of Alport (Col4a3^{-/-}) mice to noise toxicity, a model of glomerular disease associated with hearing loss [58].

3.2. Noise triggers ROS production by mitochondria or vascular NOX isoforms

Mitochondria which represent a central pharmacological target for the therapy of (ischemic) heart disease [59,60], are also ROS-producing organelles generating interest in noise research. Rats exposed to lowfrequency noise (\geq 90 dB(A), < 500 Hz) have enlarged cardiac mitochondria and reduced connexin 43 content, indicating mitochondrial damage, and cardiac fibrosis [61] (Fig. 2). Importantly, mitochondria also contain connexin 43 [62] and changes in mitochondrial connexin 43 content affects ROS formation [63]. Moreover, loud noise (100 dB (A)) induces mitochondrial DNA damage, swelling of mitochondrial membranes, matrix dilution and cristolysis, which may be associated with higher noradrenalin levels, MAO activity and disturbed mitophagy [64], all of which may contribute to a dysregulation of mitochondrial permeability transition and calcium handling [65]. Other specific mitochondrial ROS producing systems, e.g. p66^{shc} [66,67] or the mitochondrial NOS system [68], have so far not been studied with respect to noise-induced oxidative stress. Our recent studies also reported activation of cerebral mitochondrial ROS formation on day 4 of aroundthe-clock noise that persisted upon deletion of Nox2 [46], indicating additional mechanisms beyond the crosstalk between NOX-2- and mitochondria-derived ROS as reported in various disease conditions [55]. Sleep phase noise exposure also induced more pronounced mitochondrial ROS formation than awake phase noise (measured by mitoSOX fluorescence microtopography) [46]. Dysfunctional/damaged mitochondria provide one explanation for the substantial impact of traffic noise exposure on cardiovascular disease in general and ischemic heart disease in particular [10], as (oxidative) damage of mitochondria plays a central role in cardiovascular health and disease [69]. Additional ROS sources include NOX-1 and xanthine oxidase. In isolated lung endothelial cells and in the brain, aircraft noise increased Nox1 mRNA [21,46]. The protective effects of allopurinol on noise (125 dB(A), 2-3 kHz)-induced hearing loss also suggests a partial contribution of xanthine oxidase to overall ROS generation in noise-exposed animals [70].

3.3. Noise-triggers oxidative damage of biological structures and redoxregulation of biological pathways

Rats exposed to high levels of white noise (80-100 dB(A), 8-16 kHz)

have higher malondialdehyde levels, lower superoxide dismutase (SOD) activity and increased plasma nitrite/nitrate levels (probably from iNOS) that was associated with higher plasma concentrations of corticosterone, catecholamines and endothelin-1 [71]. Cardiomyocytes isolated from rat hearts upon exposure to white noise (> 100 dB(A)) have oxidative damage, including DNA damage, providing an explanation for cardiovascular dysfunction in response to noise [72]. White noise exposure of rats for up to 30 days (100 dB(A) for 4h/d) also induced cognitive impairment that was associated with upregulation of SOD and more lipid peroxidation in the brain as well as higher plasma corticosterone levels associated with lower glutathione concentrations [73] (Fig. 1). Exposure of mice for 4 weeks to white noise (90 dB(A) for 5h/d) increased circulating stress hormone levels and 8-hvdroxy-2'deoxyguanosine (a marker of oxidative stress induced DNA damage) [74]. Other related studies and reports of noise-induced imbalance of redox state in developing animals have been reviewed, although it should be noted that most of these studies used extremely high sound pressure levels of more than 95 dB(A) [75]. Noise exposure of humans (46 dB(A) mean sound pressure levels) caused oxidative damage of biomolecules used as redox biomarkers (including 3-nitrotyrosine and 8-isoprostane), which were substantially increased in the plasma [46]. In addition, factory workers exposed to loud noise (at least 85 dB(A)) had markedly greater oxidative DNA damage in peripheral blood mononuclear cells, higher GPx-1 levels and higher blood pressures compared to office workers with exposure to lower noise levels (40-50 dB(A)) [76] (Fig. 2). As summarized previously, noise-induced oxidative stress is mainly mediated via redox-sensitive kinases (e.g. c-Jun N-terminal kinase) that can be prevented by antioxidant therapies (e.g. natural antioxidants, synthetic ROS scavengers, cardiovascular drugs or genetic Nrf2 activation) and can be mitigated by physical exercise [77]. Of note, noise can also induce epigenetic changes as shown by substantial redox-triggered alterations of microRNA networks in the setting of hearing loss [78].

Previous studies by our group demonstrated that aircraft noise exposure (mean sound pressure level 73 dB(A) for 4d) increased oxidative stress, as indicated by higher levels of oxidative damage markers such as 4-hydroxynonenal-positive proteins in aortic tissue [21] and malondialdehyde-positive proteins in plasma as well as 3-nitrotyrosinepositive proteins in plasma and heart tissue [21,46]. We also observed oxidative DNA damage (8-hydroxy-2'deoxyguanosine) in aorta of aircraft noise-exposed mice (Kvandova et al., in press in Free Radical Research). Oxidative stress in turn provoked upregulation of antioxidant defense systems including the stress-response protein heme oxygenase-1 or the protective peroxisome proliferator-activated receptor gamma coactivator 1α (PGC- 1α) in isolated lung endothelial cells [21], which may have protective effects on eNOS coupling state (Table 1) and prevent noise-induced endothelial dysfunction. Using next generation sequencing we also demonstrated a down-regulation of genes encoding for antioxidant systems (e.g. intracellular superoxide dismutase [Sod1], forkhead-box-protein O3 [Foxo3]), whereas pro-apoptotic/cell death factors were upregulated (e.g. caspases, Fas, p38) [21]. Next generation sequencing also revealed dysregulation of the circadian clock system as central clock genes were significantly up- or down-regulated (e.g. Per1, Bmal1 (Arntl), Cry1, Npas2 (paralog of Clock) and Rora [46], which may be related to impaired FOXO3 signaling and adverse redox regulation of cysteine residues in central circadian clock proteins (e.g. cryptochrome) by noise-induced oxidative stress as reviewed in detail [64]. A disturbance of the circadian rhythm in response to "nighttime noise" is associated with multiple adverse health effects, such as cardiometabolic disease [64], characterized by increased plasma glucose and leptin levels in noise-exposed mice [46]. The HPA axis activity and thus cortisol levels are largely controlled by the circadian clock [18]. Noise-dependent dysregulation of gene expression measured by next generation sequencing was confirmed for selected genes by RT-PCR [46]. Impaired FOXO3 signaling is likely a key mechanism in our noise animal model since the activation of FOXO3 by the calcium antagonist bepridil

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eNOS uncoupling by S-glutathionylation in disease models (literature before 2012 reviewed in Ref. [47,104,105]).

Adverse regulation and uncoupling of eNOS (leading to superoxide formation) by S-glutathionylation at cysteine residues Cys689 and Cys908 in the reductase domain.	[86,106]
eNOS uncoupling by S-glutathionylation was detected in mice with lung injury after lipopolysaccharide challenges.	[107]
eNOS uncoupling by S-glutathionylation and endothelial dysfunction in hypertensive mice was prevented by pharmacological (SfA and CsA) or genetic ($CypD^{-/-}$) inhibition of mitochondrial permeability transition pore opening.	[108]
eNOS uncoupling by S-glutathionylation was found in mice with atherosclerosis induced by carbamylated low-density lipoprotein treatment.	[109]
eNOS uncoupling by S-glutathionylation was aggravated in mice with cardiac pressure-overload, whereas N-acetylcysteine treatment improved the phenotype.	[110]
eNOS uncoupling by S-glutathionylation and endothelial dysfunction in mice was aggravated by aging and genetic glutathione peroxidase 1 deficiency ($Gpx1^{-/-}$).	[111]
Pressure overload induced by transverse aortic constriction was associated with eNOS uncoupling by S-glutathionylation in mice, which was prevented by physical	[112]
exercise.	
Pharmacological induction of eNOS uncoupling by S-glutathionylation and endothelial dysfunction by isosorbide-5-mononitrate treatment of mice.	[113]
eNOS uncoupling by S-glutathionylation was associated with erectile dysfunction in rats with type 2 diabetes mellitus.	[114]
eNOS uncoupling by S-glutathionylation was aggravated by doxorubicin treatment leading to cardiomyopathy in mice and was normalized by antioxidant therapy with folic acid.	[115]
Exposure of mice to aircraft noise caused eNOS uncoupling by S-glutathionylation, endothelial dysfunction and hypertension.	[21,46]
A knock-in mouse with C101A-eNOS mutant (impaired dimer formation due to disrupted Zn(Cys) ₄ complex) displayed eNOS S-glutathionylation and higher superoxide formation rate and tyrosine nitration levels.	[116]
Human aortic endothelial cells showed pronounced eNOS S-glutathionylation upon challenges with ultrafine particles (air pollution).	[117]
Cultured endothelial cells had increased eNOS S-glutathionylation and decreased BH4 levels in response to ischemia/reperfusion.	[118]
eNOS S-glutathionylation can be reversed by glutaredoxin-1 making this eNOS dysregulatory mechanism a reversible redox regulatory pathway.	[119]
eNOS S-glutathionylation and a role for glutaredoxin-1 were demonstrated in mice with necrotizing enterocolitis.	[120,121]
Experimental fibrosis is linked to altered glutathione synthesis and enhanced eNOS S-glutathionylation.	[122]
eNOS S-glutathionylation and BH4 deficiency are obviously interconnected and predict eNOS uncoupling.	[123]
eNOS S-glutathionylation is correlated with nNOS activity.	[124]
Placental eNOS S-glutathionylation is a hallmark of preeclampsia.	[125]
Kruppel-like factor 2 (KLF2) protects endothelial function by activation of the Nrf2/HO-1 pathway and normalizing eNOS S-glutathionylation and BH4/BH2 ratio in cultured endothelial cells upon ischemia/reperfusion.	[126]
Treatment of hypertensive mice with the glucagon-like peptide analog liraglutide normalized oxidative stress, endothelial function and eNOS S-glutathionylation.	[127]

significantly improved several key parameters such as endothelial dysfunction and vascular/cerebral oxidative stress [46]. White noise exposure at similar sound pressure levels as compared to aircraft noise (73 dB(A)) did not cause any of the above described adverse vascular/ cerebral effects produced by noise, clearly identifying that it is not only the loudness of the noise but rather also the sound pattern/characteristics such as irregular breaks, the frequency of the noise, and the crescendo or descrescendo nature determine the detrimental effects of noise [21,46]. Of importance is our recent demonstration of increased oxidative stress in the vasculature and the plasma within 24h at low noise exposure levels that is very different from older reports, where oxidative damage was established by exposure to very loud white noise (90 dB(A) for 5h/d for 4 weeks) [74], which is far greater than the sound levels used for aircraft noise in our recent studies [21,46].

4. NOS uncoupling by traffic noise exposure and cardiovascular/ cerebral consequences

4.1. Noise-mediated impairment of vascular (endothelial) function

Early evidence for the systemic effects of noise exposure on the circulation was based on blood pressure increases in noise-exposed monkeys [79] and rats [80]. Of note, these increases in blood pressure were not associated with hearing loss. Exposure of rats to loud white noise for up to 4 weeks (100 dB(A) for 4h/d) caused endothelial dysfunction (measured with acetylcholine-dependent relaxation) and increased sensitivity to vasoconstriction by serotonin [81,82]. These observations in animals were extended to human studies which revealed that even exposure to low-level traffic noise (mean sound pressure levels of 46 (aircraft) or 54 (train) dB(A)) for one night induced endothelial dysfunction (measured by flow-mediated dilation, FMD) in healthy subjects or patients with established coronary artery disease. Impaired FMD was associated with impaired sleep quality and/or increased stress hormones [14,83], markers of inflammation/thrombosis [42] and oxidative stress (3-nitrotyrosine, 8-isoprostane and interleukins in plasma samples of exposed subjects) [46]. According to our observations, exposure of mice to aircraft noise induced endothelial dysfunction, increased blood pressure [21,46], increased circulating levels of neurohormones and induced a supersensitivity to vasoconstrictors such as endothelin-1 and noradrenalin [46].

4.2. Noise-mediated uncoupling/dysregulation of eNOS

As a potential explanation of noise-induced endothelial dysfunction, we observed diminished vascular 'NO bioavailability in the aorta of noise-exposed mice (measured by electron paramagnetic resonance [EPR] spectroscopy) [21]. This is likely a consequence of a more pronounced oxidative break-down of 'NO by a rapid reaction with superoxide to form a highly reactive intermediate peroxynitrite [84], which in turn may be responsible for a dysfunctional eNOS [55,85] (Fig. 3). Higher rates of formation of ROS (including superoxide) by aircraft noise exposure were discussed above, but eNOS S-glutathionylation represents another mechanism for uncoupling of the eNOS protein [21,46]. S-glutathionylation of eNOS suggests an altered electron flow in the reductase domain and the transfer of these electrons to molecular oxygen and thus superoxide formation [86], a pathomechanism established for numerous diseases (see Table 1 and reviewed in Ref. [87]). Other mechanisms of eNOS dysregulation that were previously not reported for noise exposure but that are conceivable include: PKC or protein tyrosine kinase 2 (PYK-2) dependent adverse phosphorylation at Thr495 and Tyr657, respectively, or accumulation of asymmetric dimethylarginine, oxidative disruption of the zinc-sulfur-complex at the dimer binding interface and oxidative tetrahydrobiopterin (BH4) depletion (all explained in detail in Refs. [47,87] and summarized in Table 2). Another method to detect eNOS-dependent endothelial ROS formation in aortic tissue of noise-exposed mice is the quantification of endothelial dihydroethidium by cryo-staining [21,46]. Inhibition of this signal by the eNOS inhibitor L-N^G-nitro-arginine methyl ester (L-NAME) indicates substantial uncoupling of eNOS protein [88]. In contrast, identical sound pressure levels of white noise increased rather than decreased endothelial dihydroethidium fluorescence signal in the presence of L-NAME, pointing to a functional (coupled) eNOS protein [21]. Importantly, aircraft noise increased eNOS mRNA in isolated lung endothelial cells and eNOS protein in aorta suggesting a compensatory upregulation in response to the oxidative uncoupling/dysregulation of eNOS protein [21].



Fig. 3. Noise exposure causes eNOS and nNOS uncoupling. Superoxide formation induced by noise (e.g. NOX-2 activation) causes oxidative break-down of 'NO leading to peroxynitrite formation and providing an explanation for increased protein tyrosine nitration (also iNOS-derived 'NO contributes) as well as impairment of 'NO/cGMP signaling. Despite upregulation of eNOS and the tetrahydrobiopterin (BH4)-generating enzymes GTP-cyclohydrolase 1 (GCH-1) and dihydrofolate reductase (DHFR), diminished 'NO bioavailability was observed in aorta of noise-exposed mice. BH4 is an essential cofactor of eNOS (but also of iNOS) and oxidative depletion by ROS (e.g. peroxynitrite) to dihydrobiopterin (BH2) causes uncoupling of all NOS isoforms. BH4 levels were not measured upon noise exposure but upregulation of GCH-1 and DHFR obviously cannot compensate for the loss of function of eNOS in this setting. Therefore, eNOS was found to be dysfunctional or uncoupled, which was supported by S-glutathionylation (GSS-modification) by either ROS/ GSH or S-nitrosoglutathione (GSNO) reaction. eNOS S-glutathionylation is an accepted marker of eNOS uncoupling (see Table 1) and uncoupled eNOS after noise exposure was detected at the molecular level by endothelial ROS formation that was sensitive to inhibition by the NOS inhibitor L-NAME. Although

not measured in tissues of noise-exposed animals so far, the inactivating phosphorylation of eNOS at threonine 495 or tyrosine 657 (see Table 2), mediated by redoxactivated protein kinase C (PKC) and protein tyrosine kinase 2 (PYK-2) respectively, would be conceivable under noise-induced oxidative stress conditions. In the brain, oxidative stress induction (e.g. NOX-2 activation) by noise caused nNOS uncoupling as envisaged by cerebral ROS formation that was sensitive to inhibition by the selective nNOS inhibitor ARL-17477 as well as phosphorylation at serine 847 (previously reported for uncoupled nNOS). Phospho-Ser847 in nNOS is introduced by calcium/calmodulin-dependent protein kinase (CaMKII) that is activated by ROS. The adverse redox regulation of eNOS and nNOS by noise induced oxidative stress promote the development of cardiometabolic disease and cognitive impairment that were previously reported for noise exposure for clinical/epidemiological studies.

The enzymes responsible for the synthesis and recycling of BH4 are GTP-cyclohydrolase-1 and dihydrofolate reductase [89]. In the setting of increased oxidative stress, oxidative conversion of BH4 to the 'BH3 radical and finally to dihydrobiopterin (BH2) leads to depletion of this essential eNOS cofactor and causes eNOS dysfunction [89-91]. The observation that aircraft noise increases the expression of GTP-cyclohydrolase-1 and dihydrofolate reductase points to a compensatory (however futile) upregulation of these enzymes to overcome eNOS uncoupling and endothelial dysfunction [21] (Fig. 3). Although redoxdependent proteasomal degradation of GTP-cyclohydrolase-1 and dihydrofolate reductase have been reported [92], the upregulation of GTP-cyclohydrolase-1 and dihydrofolate reductase under inflammatory conditions can also known the cofactor BH4 for iNOS activity [93]. The consequences of eNOS uncoupling in response to noise are decreased 'NO bioavailability (due to eNOS dysfunction/uncoupling as well as direct oxidative break-down of 'NO by enhanced superoxide formation) and increased vasoconstrictor sensitivity of the vasculature to noradrenaline, endothelin-1 and angiotensin-II, all of which contributes to endothelial dysfunction. Of note, exposure to "white noise" with identical mean sound pressure levels did not induce these adverse vascular effects, indicating that it may not only be the loudness (quantity) of a specific noise but rather also its characteristics (quality such as frequencies, pattern) that determine its harmful effects [21].

4.3. Noise-mediated uncoupling/dysregulation of nNOS

Aircraft noise caused nNOS downregulation and uncoupling in the brain of mice as evidenced by increased phosphorylation at serine 847 of the enzyme [46] (Fig. 3); this phosphorylation site was previously shown to cause uncoupling of nNOS [94]. In addition, ROS formation in brain tissue sections from noise-exposed mice was prevented by the highly specific nNOS inhibitor ARL-17477, implicating nNOS-derived cerebral ROS formation (nNOS uncoupling) [46]. Thus, downregulation and uncoupling of the nNOS protein leads to a cerebral deficit of the neurotransmitter 'NO, resulting in enhanced neuroinflammation, downregulation of *Foxo3* and increased cerebral oxidative stress [46].

Table 2

eNOS uncoupling/dysregulation by other mechanisms (reviewed in detail in Ref. [47,87]).

Oxidative depletion of tetrahydrobiopterin (BH4) is the most prominent and well accepted mechanism of eNOS uncoupling and was reported for hypertension,	[90,128–132]
diabetes and atherosclerosis.	
GTP-cyclohydrolase-1 (GCH-1), the major enzymatic source for BH4, is an important regulator of eNOS activity and endothelial function. eNOS/GCH-1 ratio	[131,133]
determines proper NO synthesis.	
Dihydrofolate reductase (DHFR) represents the "salvage pathway" and is responsible for recycling of oxidized BH2 back to BH4.	[134,135]
Oxidative disruption of the zinc-sulfur-complex (ZnCys ₄) in the binding region of the eNOS dimer is another mechanism of eNOS uncoupling and described in	[136–138]
diabetic animals and men. This oxidative damage of eNOS is most-likely mediated by HOCl and peroxynitrite.	
Phosphorylation at Thr495 and Tyr657 by PKC and protein tyrosine kinase 2 (PYK-2), respectively, cause at least dysfunction of eNOS and were observed in	[111,139–141]
models of hypertension, aging and nitrate tolerance.	
The formation and degradation of the potent endogenous eNOS inhibitor (and maybe uncoupling agent), asymmetric dimethylarginine (ADMA), is regulated by	[105,142,143]
protein arginine methyltransferases and by dimethylarginine dimethylaminohydrolases (DDAH) that are both redox-regulated	
1-arginine deficiency may contribute to uncoupling of eNOS as shown by a case report on a patient with impaired ADMA due to genetic inefficiency of y ⁺ LAT-1	[144,145]
(amino acid transporter)-dependent ADMA export from endothelial cells. This patient had severe angina pectoris and substantial eNOS uncoupling that was	
improved by high dose L-arginine therapy driving the ADMA export via the cationic amino acid transporter.	

Cerebral nNOS down-/dysregulation and concomitant NOX-2 upregulation can also account for the impaired cognitive development (including memory and learning) of noise exposed children [95], which is also supported by the established role of NOX-2 dependent ROS formation for learning and memory impairment in female adult rats [96]. Redox-regulation of adverse (uncoupling) nNOS phosphorylation at serine 847 by the redox sensitivity of calcium/calmodulin-dependent protein kinase [94] provides another important mechanism for NOX-2 dependent cognitive impairment.

5. Conclusions and outlook

Exposure to aircraft noise in humans and experimental animals activates the sympathetic and HPA systems, leading to increases in stress hormones such as catecholamines, angiotensin-II, endothelin-1 and cortisol. These changes contribute to the direct activation of phagocytic NADPH oxidase, causing oxidative stress and thus oxidative depletion of 'NO, and eNOS uncoupling e.g. by S-glutathionylation [21,46]. Likewise, noise-induced cerebral oxidative stress causes nNOS downregulation and uncoupling the consequences of which include immune cell activation/infiltration (potentiating the oxidative damage in cardiovascular/cerebral tissues) and enhanced vasoconstriction. Moreover, decreased nNOS activity contributes to changes of gene expression towards pro-inflammatory and pro-oxidative conditions (including impaired circadian clock or dysregulated FOXO signaling) [46]. These vascular and cerebral (oxidative) alterations caused by aircraft noise-exposure of mice are mostly due to eNOS and nNOS dysregulation acting as central players that contribute to the development of cardiometabolic disorders such as arterial hypertension and diabetes mellitus [21,46] and which are also confirmed by human studies [9,97]. The key role of oxidative stress for the harms to health caused by noise is further supported by the demonstration of impaired antioxidant defenses and the prevention of noise inflicted damage by treatment with exogenous antioxidants or the activation of endogenous antioxidant pathways [75]. Taken together, the major pathomechanisms of noise exposure are the induction of endothelial dysfunction, oxidative stress and inflammation (e.g. by activation of the NADPH oxidase and by eNOS uncoupling).

These mechanisms are strikingly similar to those reported for classical cardiovascular risk factors such as diabetes [98], hypertension [99] and hypercholesterolemia [100]. It remains to be established whether treatment with AT1 receptor blockers, ACE-inhibitors, antidiabetic agents and also statins could prevent noise-induced adverse cardiovascular health effects, as these cardiovascular drugs possess potent antioxidant and anti-inflammatory pleiotropic properties [101,102], and thereby could interfere with the major pathomechanisms of noise-induced health harms. Confirmation of established cardiovascular drugs as efficient preventive measures against noise-inflicted damage would be important, since patients in need of those drugs (e.g. those with preestablished cardiovascular diseases) may represent a population of high noise-dependent vulnerability [83]. Other populations of high noise-dependent vulnerability may be children at developmental stage, who showed cognitive impairment in response to noise [95], as well as elderly, who showed an increased risk of metabolic syndrome in response to noise [103].

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Declaration of competing interest

The authors declare that they have no conflicts of interest with the contents of this article.

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References

- S.M. Rappaport, Genetic factors are not the major causes of chronic diseases, PloS One 11 (2016) e0154387, https://doi.org/10.1371/journal.pone.0154387.
- [2] K. Sainani, Taking on the exposome bringing bioinformatics tools to the environmental side of the health equation, Biomed. Comput. Rev. 2016 (2016) 14–21 Fall.
- [3] P.J. Landrigan, R. Fuller, N.J.R. Acosta, O. Adeyi, R. Arnold, N.N. Basu, A.B. Balde, R. Bertollini, S. Bose-O'Reilly, J.I. Boufford, P.N. Breysse, T. Chiles, C. Mahidol, A.M. Coll-Seck, M.L. Cropper, J. Fobil, V. Fuster, M. Greenstone, A. Haines, D. Hanrahan, D. Hunter, M. Khare, A. Krupnick, B. Lanphear, B. Lohani, K. Martin, K.V. Mathiasen, M.A. McTeer, C.J.L. Murray, J.D. Ndahimananjara, F. Perera, J. Potocnik, A.S. Preker, J. Ramesh, J. Rockstrom, C. Salinas, L.D. Samson, K. Sandilya, P.D. Sly, K.R. Smith, A. Steiner, R.B. Stewart, W.A. Suk, O.C.P. van Schayck, G.N. Yadama, K. Yumkella, M. Zhong, The Lancet Commission on pollution and health, Lancet 391 (2018) 462–512, https://doi.org/10.1016/S0140-6736(17)32345-0.
- WHO, Ambient air pollution: a global assessment of exposure and burden of disease, http://apps.who.int/iris/bitstream/10665/250141/1/9789241511353-eng.pdf?ua = 1, (2016).
- [5] WHO report, Preventing disease through healthy environments, https://www. who.int/quantifying_ehimpacts/publications/preventing-disease/en/, (2016).
- [6] G.B.D. DALYS, H. Collaborators, Global, regional, and national disability-adjusted life-years (DALYS) for 315 diseases and injuries and healthy life expectancy (HALE), 1990-2015: a systematic analysis for the Global Burden of Disease Study 2015, Lancet 388 (2016) 1603–1658, https://doi.org/10.1016/S0140-6736(16) 31460-X.
- S.A. Stansfeld, Noise effects on health in the context of air pollution exposure, Int. J. Environ. Res. Publ. Health 12 (2015) 12735–12760, https://doi.org/10.3390/ ijerph121012735.
- [8] G.D.D.R.F. Collaborators, Global, regional, and national comparative risk assessment of 84 behavioural, environmental and occupational, and metabolic risks or clusters of risks, 1990-2016: a systematic analysis for the Global Burden of Disease Study 2016, Lancet 390 (2017) 1345–1422, https://doi.org/10.1016/S0140-6736(17)32366-8.
- [9] T. Munzel, F.P. Schmidt, S. Steven, J. Herzog, A. Daiber, M. Sorensen, Environmental noise and the cardiovascular system, J. Am. Coll. Cardiol. 71 (2018) 688–697, https://doi.org/10.1016/j.jacc.2017.12.015.
- [10] E.V. Kempen, M. Casas, G. Pershagen, M. Foraster, WHO environmental noise guidelines for the European region: a systematic review on environmental noise and cardiovascular and metabolic effects: a summary, Int. J. Environ. Res. Publ. Health 15 (2018) 379, https://doi.org/10.3390/ijerph15020379.
- [11] D. Vienneau, L. Perez, C. Schindler, C. Lieb, H. Sommer, N. Probst-Hensch, N. Kunzli, M. Roosli, Years of life lost and morbidity cases attributable to transportation noise and air pollution: a comparative health risk assessment for Switzerland in 2010, Int. J. Hyg Environ. Health 218 (2015) 514–521, https://doi. org/10.1016/j.ijheh.2015.05.003.
- [12] T. Munzel, M. Sorensen, F. Schmidt, E. Schmidt, S. Steven, S. Kroller-Schon, A. Daiber, The adverse effects of environmental noise exposure on oxidative stress and cardiovascular risk, Antioxidants Redox Signal. 28 (2018) 873–908, https:// doi.org/10.1089/ars.2017.7118.
- [13] A.R. Fetoni, F. Paciello, R. Rolesi, S.L. Eramo, C. Mancuso, D. Troiani, G. Paludetti, Rosmarinic acid up-regulates the noise-activated Nrf2/HO-1 pathway and protects against noise-induced injury in rat cochlea, Free Radic. Biol. Med. 85 (2015) 269–281, https://doi.org/10.1016/j.freeradbiomed.2015.04.021.
- [14] F.P. Schmidt, M. Basner, G. Kroger, S. Weck, B. Schnorbus, A. Muttray, M. Sariyar, H. Binder, T. Gori, A. Warnholtz, T. Munzel, Effect of nighttime aircraft noise exposure on endothelial function and stress hormone release in healthy adults, Eur. Heart J. 34 (2013) 3508–3514a, https://doi.org/10.1093/eurheartj/eht269.
- [15] K.D. Kryter, Effects of Noise on Man, Academic Press, 1970.[16] W. Babisch, The noise/stress concept, risk assessment and research needs, Noise
- Health 4 (2002) 1–11.[17] W. Babisch, Stress hormones in the research on cardiovascular effects of noise, Noise Health 5 (2003) 1–11.
- [18] T. Meyer, P.H. Wirtz, Mechanisms of mitochondrial redox signaling in psychosocial stress-responsive systems: new insights into an old story, Antioxidants Redox Signal. 28 (2018) 760–772, https://doi.org/10.1089/ars.2017.7186.
- [19] N. Xia, H. Li Loneliness, Social isolation, and cardiovascular health, Antioxidants Redox Signal. 28 (2018) 837–851, https://doi.org/10.1089/ars.2017.7312.

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- [20] A. Daiber, S. Kroller-Schon, K. Frenis, M. Oelze, S. Kalinovic, K. Vujacic-Mirski, M. Kuntic, M.T. Bayo Jimenez, J. Helmstadter, S. Steven, B. Korac, T. Munzel, Environmental noise induces the release of stress hormones and inflammatory signaling molecules leading to oxidative stress and vascular dysfunction-Signatures of the internal exposome, Biofactors 45 (2019) 495–506, https://doi. org/10.1002/biof.1506.
- [21] T. Munzel, A. Daiber, S. Steven, L.P. Tran, E. Ullmann, S. Kossmann, F.P. Schmidt, M. Oelze, N. Xia, H. Li, A. Pinto, P. Wild, K. Pies, E.R. Schmidt, S. Rapp, S. Kroller-Schon, Effects of noise on vascular function, oxidative stress, and inflammation: mechanistic insight from studies in mice, Eur. Heart J. 38 (2017) 2838–2849, https://doi.org/10.1093/eurheartj/ehx081.
- [22] M.E. Beutel, C. Junger, E.M. Klein, P. Wild, K. Lackner, M. Blettner, H. Binder, M. Michal, J. Wiltink, E. Brahler, T. Munzel, Noise annoyance is associated with depression and anxiety in the general population- the contribution of aircraft noise, PloS One 11 (2016) e0155357, https://doi.org/10.1371/journal.pone. 0155357.
- [23] O. Hahad, M. Beutel, T. Gori, A. Schulz, M. Blettner, N. Pfeiffer, T. Rostock, K. Lackner, M. Sorensen, J.H. Prochaska, P.S. Wild, T. Munzel, Annoyance to different noise sources is associated with atrial fibrillation in the Gutenberg Health Study, Int. J. Cardiol. 264 (2018) 79–84, https://doi.org/10.1016/j.ijcard.2018. 03.126.
- [24] C. Sinning, T. Keller, N. Abegunewardene, K.F. Kreitner, T. Munzel, S. Blankenberg, Tako-Tsubo syndrome: dying of a broken heart? Clin. Res. Cardiol. 99 (2010) 771–780, https://doi.org/10.1007/s00392-010-0224-9.
- [25] T. Munzel, M. Knorr, F. Schmidt, S. von Bardeleben, T. Gori, E. Schulz, Airborne disease: a case of a Takotsubo cardiomyopathie as a consequence of nighttime aircraft noise exposure, Eur. Heart J. 37 (2016) 2844, https://doi.org/10.1093/ eurhearti/ehw314.
- [26] T. Wallerath, K. Witte, S.C. Schafer, P.M. Schwarz, W. Prellwitz, P. Wohlfart, H. Kleinert, H.A. Lehr, B. Lemmer, U. Forstermann, Down-regulation of the expression of endothelial NO synthase is likely to contribute to glucocorticoidmediated hypertension, Proc. Natl. Acad. Sci. U. S. A. 96 (1999) 13357–13362.
- [27] S. Yang, L. Zhang, Glucocorticoids and vascular reactivity, Curr. Vasc. Pharmacol. 2 (2004) 1–12.
- [28] S. Schiavone, V. Jaquet, L. Trabace, K.H. Krause, Severe life stress and oxidative stress in the brain: from animal models to human pathology, Antioxidants Redox Signal. 18 (2013) 1475–1490, https://doi.org/10.1089/ars.2012.4720.
- [29] J.W. Wright, H.A. Dengerink, J.M. Miller, P.C. Goodwin, Potential role of angiotensin II in noise-induced increases in inner ear blood flow, Hear. Res. 17 (1985) 41–46, https://doi.org/10.1016/0378-5955(85)90128-5.
- [30] L.K. Kanazawa, D.D. Vecchia, E.M. Wendler, P.A. Hocayen, F.A. Dos Reis Livero, M.C. Stipp, I.M. Barcaro, A. Acco, R. Andreatini, Quercetin reduces manic-like behavior and brain oxidative stress induced by paradoxical sleep deprivation in mice, Free Radic. Biol. Med. 99 (2016) 79–86, https://doi.org/10.1016/j. freeradbiomed.2016.07.027.
- [31] K.H. Alzoubi, O.F. Khabour, A.S. Albawaana, F.H. Alhashimi, R.Y. Athamneh, Tempol prevents chronic sleep-deprivation induced memory impairment, Brain Res. Bull. 120 (2016) 144–150, https://doi.org/10.1016/j.brainresbull.2015.11. 017.
- [32] R.M. Bruno, E. Daghini, L. Ghiadoni, I. Sudano, I. Rugani, M. Varanini, C. Passino, M. Emdin, S. Taddei, Effect of acute administration of vitamin C on muscle sympathetic activity, cardiac sympathovagal balance, and baroreflex sensitivity in hypertensive patients, Am. J. Clin. Nutr. 96 (2012) 302–308, https://doi.org/10. 3945/ajcn.112.035022.
- [33] S. Ye, H. Zhong, S. Yanamadala, V.M. Campese, Oxidative stress mediates the stimulation of sympathetic nerve activity in the phenol renal injury model of hypertension, Hypertension 48 (2006) 309–315, https://doi.org/10.1161/01. HYP.0000231307.69761.2e.
- [34] H.E. Lob, P.J. Marvar, T.J. Guzik, S. Sharma, L.A. McCann, C. Weyand, F.J. Gordon, D.G. Harrison, Induction of hypertension and peripheral inflammation by reduction of extracellular superoxide dismutase in the central nervous system, Hypertension 55 (2010) 277–283, https://doi.org/10.1161/ HYPERTENSIONAHA.109.142646 276pp. following 283.
- [35] M. Neri, D. Cerretani, A.I. Fiaschi, P.F. Laghi, P.E. Lazzerini, A.B. Maffione, L. Micheli, G. Bruni, C. Nencini, G. Giorgi, S. D'Errico, C. Fiore, C. Pomara, I. Riezzo, E. Turillazzi, V. Fineschi, Correlation between cardiac oxidative stress and myocardial pathology due to acute and chronic norepinephrine administration in rats, J. Cell Mol. Med. 11 (2007) 156–170, https://doi.org/10.1111/j. 1582-4934.2007.00009.x.
- [36] H.Y. Lee, J.S. Lee, H.G. Kim, W.Y. Kim, S.B. Lee, Y.H. Choi, C.G. Son, The ethanol extract of Aquilariae Lignum ameliorates hippocampal oxidative stress in a repeated restraint stress mouse model, BMC Compl. Alternative Med. 17 (2017) 397, https://doi.org/10.1186/s12906-017-1902-1.
- [37] M.T. Grande, G. Pascual, A.S. Riolobos, M. Clemente-Lorenzo, B. Bardaji, L. Barreiro, O. Tornavaca, A. Meseguer, J.M. Lopez-Novoa, Increased oxidative stress, the renin-angiotensin system, and sympathetic overactivation induce hypertension in kidney androgen-regulated protein transgenic mice, Free Radic. Biol. Med. 51 (2011) 1831–1841, https://doi.org/10.1016/j.freeradbiomed.2011.08. 014.
- [38] S. Rajagopalan, S. Kurz, T. Munzel, M. Tarpey, B.A. Freeman, K.K. Griendling, D.G. Harrison, Angiotensin II-mediated hypertension in the rat increases vascular superoxide production via membrane NADH/NADPH oxidase activation. Contribution to alterations of vasomotor tone, J. Clin. Invest. 97 (1996) 1916–1923, https://doi.org/10.1172/JCI118623.
- [39] H. Mollnau, M. Oelze, M. August, M. Wendt, A. Daiber, E. Schulz, S. Baldus, A.L. Kleschyov, A. Materne, P. Wenzel, U. Hink, G. Nickenig, I. Fleming,

T. Munzel, Mechanisms of increased vascular superoxide production in an experimental model of idiopathic dilated cardiomyopathy, Arterioscler. Thromb. Vasc. Biol. 25 (2005) 2554–2559.

- [40] A. Kim, J.H. Sung, J.H. Bang, S.W. Cho, J. Lee, C.S. Sim, Effects of self-reported sensitivity and road-traffic noise levels on the immune system, PloS One 12 (2017) e0187084, , https://doi.org/10.1371/journal.pone.0187084.
- [41] Y. Cai, A.L. Hansell, M. Blangiardo, P.R. Burton, K. de Hoogh BioShaRe, D. Doiron, I. Fortier, J. Gulliver, K. Hveem, S. Mbatchou, D.W. Morley, R.P. Stolk, W.L. Zijlema, P. Elliott, S. Hodgson, Long-term exposure to road traffic noise, ambient air pollution, and cardiovascular risk factors in the HUNT and lifelines cohorts, Eur. Heart J. 38 (2017) 2290–2296, https://doi.org/10.1093/eurheartj/ ehx263.
- [42] J. Herzog, F.P. Schmidt, O. Hahad, S.H. Mahmoudpour, A.K. Mangold, P. Garcia Andreo, J. Prochaska, T. Koeck, P.S. Wild, M. Sorensen, A. Daiber, T. Munzel, Acute exposure to nocturnal train noise induces endothelial dysfunction and prothromboinflammatory changes of the plasma proteome in healthy subjects, Basic Res. Cardiol. 114 (2019) 46, https://doi.org/10.1007/s00395-019-0753-y.
- [43] D. Atanackovic, M.C. Brunner-Weinzierl, H. Kroger, S. Serke, H.C. Deter, Acute psychological stress simultaneously alters hormone levels, recruitment of lymphocyte subsets, and production of reactive oxygen species, Immunol. Invest. 31 (2002) 73–91.
- [44] T.B. Herbert, S. Cohen, A.L. Marsland, E.A. Bachen, B.S. Rabin, M.F. Muldoon, S.B. Manuck, Cardiovascular reactivity and the course of immune response to an acute psychological stressor, Psychosom. Med. 56 (1994) 337–344.
- [45] B. Cui, K. Li, Z. Gai, X. She, N. Zhang, C. Xu, X. Chen, G. An, Q. Ma, R. Wang, Chronic noise exposure acts cumulatively to exacerbate Alzheimer's disease-like amyloid-beta pathology and neuroinflammation in the rat Hippocampus, Sci. Rep. 5 (2015) 12943, https://doi.org/10.1038/srep12943.
- [46] S. Kroller-Schon, A. Daiber, S. Steven, M. Oelze, K. Frenis, S. Kalinovic, A. Heimann, F.P. Schmidt, A. Pinto, M. Kvandova, K. Vujacic-Mirski, K. Filippou, M. Dudek, M. Bosmann, M. Klein, T. Bopp, O. Hahad, P.S. Wild, K. Frauenknecht, A. Methner, E.R. Schmidt, S. Rapp, H. Mollnau, T. Munzel, Crucial role for Nox2 and sleep deprivation in aircraft noise-induced vascular and cerebral oxidative stress, inflammation, and gene regulation, Eur. Heart J. 39 (2018) 3528–3539, https://doi.org/10.1093/eurheartj/ehy333.
- [47] E. Schulz, P. Wenzel, T. Munzel, A. Daiber, Mitochondrial redox signaling: interaction of mitochondrial reactive oxygen species with other sources of oxidative stress, Antioxidants Redox Signal. 20 (2014) 308–324, https://doi.org/10.1089/ ars.2012.4609.
- [48] P. Wenzel, S. Kossmann, T. Munzel, A. Daiber, Redox regulation of cardiovascular inflammation - immunomodulatory function of mitochondrial and Nox-derived reactive oxygen and nitrogen species, Free Radic. Biol. Med. 109 (2017) 48–60, https://doi.org/10.1016/j.freeradbiomed.2017.01.027.
- [49] B. Brune, N. Dehne, N. Grossmann, M. Jung, D. Namgaladze, T. Schmid, A. von Knethen, A. Weigert, Redox control of inflammation in macrophages, Antioxidants Redox Signal. 19 (2013) 595–637, https://doi.org/10.1089/ars.2012.4785.
- [50] Y. Lei, K. Wang, L. Deng, Y. Chen, E.C. Nice, C. Huang, Redox regulation of inflammation: old elements, a new story, Med. Res. Rev. 35 (2015) 306–340, https://doi.org/10.1002/med.21330.
- [51] D.D. Chen, Y.G. Dong, H. Yuan, A.F. Chen, Endothelin 1 activation of endothelin A receptor/NADPH oxidase pathway and diminished antioxidants critically contribute to endothelial progenitor cell reduction and dysfunction in salt-sensitive hypertension, Hypertension 59 (2012) 1037–1043, https://doi.org/10.1161/ HYPERTENSIONAHA.111.183368.
- [52] N. Duerrschmidt, N. Wippich, W. Goettsch, H.J. Broemme, H. Morawietz, Endothelin-1 induces NAD(P)H oxidase in human endothelial cells, Biochem. Biophys. Res. Commun. 269 (2000) 713–717.
- [53] J. Kahler, A. Ewert, J. Weckmuller, S. Stobbe, C. Mittmann, R. Koster, M. Paul, T. Meinertz, T. Munzel, Oxidative stress increases endothelin-1 synthesis in human coronary artery smooth muscle cells, J. Cardiovasc. Pharmacol. 38 (2001) 49–57.
- [54] J. Kahler, S. Mendel, J. Weckmuller, H.D. Orzechowski, C. Mittmann, R. Koster, M. Paul, T. Meinertz, T. Munzel, Oxidative stress increases synthesis of big endothelin-1 by activation of the endothelin-1 promoter, J. Mol. Cell. Cardiol. 32 (2000) 1429–1437, https://doi.org/10.1006/jmcc.2000.1178.
- [55] A. Daiber, F. Di Lisa, M. Oelze, S. Kroller-Schon, S. Steven, E. Schulz, T. Munzel, Crosstalk of mitochondria with NADPH oxidase via reactive oxygen and nitrogen species signalling and its role for vascular function, Br. J. Pharmacol. 174 (2017) 1670–1689, https://doi.org/10.1111/bph.13403.
- [56] S. Rajagopalan, J.B. Laursen, A. Borthayre, S. Kurz, J. Keiser, S. Haleen, A. Giaid, D.G. Harrison, Role for endothelin-1 in angiotensin II-mediated hypertension, Hypertension 30 (1997) 29–34.
- [57] L.T. Tran, K.M. MacLeod, J.H. McNeill, Endothelin-1 modulates angiotensin II in the development of hypertension in fructose-fed rats, Mol. Cell. Biochem. 325 (2009) 89–97, https://doi.org/10.1007/s11010-008-0023-z.
- [58] D.T. Meehan, D. Delimont, B. Dufek, M. Zallocchi, G. Phillips, M.A. Gratton, D. Cosgrove, Endothelin-1 mediated induction of extracellular matrix genes in strial marginal cells underlies strial pathology in Alport mice, Hear. Res. 341 (2016) 100–108, https://doi.org/10.1016/j.heares.2016.08.003.
- [59] K. Boengler, G. Lochnit, R. Schulz, Mitochondria "THE" target of myocardial conditioning, Am. J. Physiol. Heart Circ. Physiol. 315 (2018) H1215–H1231, https://doi.org/10.1152/ajpheart.00124.2018.
- [60] S.M. Davidson, P. Ferdinandy, I. Andreadou, H.E. Botker, G. Heusch, B. Ibanez, M. Ovize, R. Schulz, D.M. Yellon, D.J. Hausenloy, D. Garcia-Dorado, C.C. Action, Multitarget strategies to reduce myocardial ischemia/reperfusion injury: JACC review topic of the week, J. Am. Coll. Cardiol. 73 (2019) 89–99, https://doi.org/ 10.1016/j.jacc.2018.09.086.

- [61] E. Antunes, G. Borrecho, P. Oliveira, A.P. Alves de Matos, J. Brito, A. Aguas, J. Martins dos Santos, Effects of low-frequency noise on cardiac collagen and cardiomyocyte ultrastructure: an immunohistochemical and electron microscopy study, Int. J. Clin. Exp. Pathol. 6 (2013) 2333–2341.
- [62] K. Boengler, S. Stahlhofen, A. van de Sand, P. Gres, M. Ruiz-Meana, D. Garcia-Dorado, G. Heusch, R. Schulz, Presence of connexin 43 in subsarcolemmal, but not in interfibrillar cardiomyocyte mitochondria, Basic Res. Cardiol. 104 (2009) 141–147, https://doi.org/10.1007/s00395-009-0007-5.
- [63] F.R. Heinzel, Y. Luo, X. Li, K. Boengler, A. Buechert, D. Garcia-Dorado, F. Di Lisa, R. Schulz, G. Heusch, Impairment of diazoxide-induced formation of reactive oxygen species and loss of cardioprotection in connexin 43 deficient mice, Circ. Res. 97 (2005) 583–586, https://doi.org/10.1161/01.RES.0000181171.65293.65.
- [64] H. Li, A.B. Kilgallen, T. Munzel, E. Wolf, S. Lecour, R. Schulz, A. Daiber, L.W. Van Laake, Influence of mental stress and environmental toxins on circadian clocks implications for redox regulation of the heart and cardioprotection, Br. J. Pharmacol. (2019), https://doi.org/10.1111/bph.14949.
- [65] F. Salvetti, B. Chelli, M. Gesi, A. Pellegrini, G. Giannaccini, A. Lucacchini, C. Martini, Effect of noise exposure on rat cardiac peripheral benzodiazepine receptors, Life Sci. 66 (2000) 1165–1175, https://doi.org/10.1016/s0024-3205(00) 00422-7.
- [66] F. Di Lisa, M. Giorgio, P. Ferdinandy, R. Schulz, New aspects of p66Shc in ischaemia reperfusion injury and other cardiovascular diseases, Br. J. Pharmacol. 174 (2017) 1690–1703, https://doi.org/10.1111/bph.13478.
- [67] K. Boengler, J. Bornbaum, K.D. Schluter, R. Schulz, P66shc and its role in ischemic cardiovascular diseases, Basic Res. Cardiol. 114 (2019) 29, https://doi.org/10. 1007/s00395-019-0738-x.
- [68] M. Kirca, P. Kleinbongard, D. Soetkamp, J. Heger, C. Csonka, P. Ferdinandy, R. Schulz, Interaction between connexin 43 and nitric oxide synthase in mice heart mitochondria, J. Cell Mol. Med. 19 (2015) 815–825, https://doi.org/10.1111/ jcmm.12499.
- [69] K. Boengler, R. Schulz, Connexin 43 and mitochondria in cardiovascular health and disease, Adv. Exp. Med. Biol. 982 (2017) 227–246, https://doi.org/10.1007/ 978-3-319-55330-6_12.
- [70] E. Cassandro, L. Sequino, P. Mondola, G. Attanasio, M. Barbara, R. Filipo, Effect of superoxide dismutase and allopurinol on impulse noise-exposed Guinea pigselectrophysiological and biochemical study, Acta Otolaryngol. 123 (2003) 802–807.
- [71] M.A. Said, O.A. El-Gohary, Effect of noise stress on cardiovascular system in adult male albino rat: implication of stress hormones, endothelial dysfunction and oxidative stress, Gen. Physiol. Biophys. 35 (2016) 371–377, https://doi.org/10.4149/ gpb_2016003.
- [72] P. Lenzi, G. Frenzilli, M. Gesi, M. Ferrucci, G. Lazzeri, F. Fornai, M. Nigro, DNA damage associated with ultrastructural alterations in rat myocardium after loud noise exposure, Environ. Health Perspect. 111 (2003) 467–471, https://doi.org/ 10.1289/ehp.5847.
- [73] S. Manikandan, M.K. Padma, R. Srikumar, N. Jeya Parthasarathy, A. Muthuvel, R. Sheela Devi, Effects of chronic noise stress on spatial memory of rats in relation to neuronal dendritic alteration and free radical-imbalance in hippocampus and medial prefrontal cortex, Neurosci. Lett. 399 (2006) 17–22, https://doi.org/10. 1016/j.neulet.2006.01.037.
- [74] K.C. Zheng, M. Ariizumi, Modulations of immune functions and oxidative status induced by noise stress, J. Occup. Health 49 (2007) 32–38, https://doi.org/10. 1539/joh.49.32.
- [75] S.J. Molina, M. Miceli, L.R. Guelman, Noise exposure and oxidative balance in auditory and extra-auditory structures in adult and developing animals. Pharmacological approaches aimed to minimize its effects, Pharmacol. Res. 109 (2016) 86–91, https://doi.org/10.1016/j.phrs.2015.11.022.
- [76] M. Bagheri Hosseinabadi, N. Khanjani, T. Munzel, A. Daiber, M. Yaghmorloo, Chronic occupational noise exposure: effects on DNA damage, blood pressure, and serum biochemistry, Mutat. Res. 841 (2019) 17–22, https://doi.org/10.1016/j. mrgentox.2019.04.006.
- [77] S. Golbidi, H. Li, I. Laher, Oxidative stress: a unifying mechanism for cell damage induced by noise, (Water-Pipe) smoking, and emotional stress-therapeutic strategies targeting redox imbalance, Antioxidants Redox Signal. 28 (2018) 741–759, https://doi.org/10.1089/ars.2017.7257.
- [78] V. Miguel, J.Y. Cui, L. Daimiel, C. Espinosa-Diez, C. Fernandez-Hernando, T.J. Kavanagh, S. Lamas, The role of MicroRNAs in environmental risk factors, noise-induced hearing loss, and mental stress, Antioxidants Redox Signal. 28 (2018) 773–796, https://doi.org/10.1089/ars.2017.7175.
- [79] E.A. Peterson, J.S. Augenstein, D.C. Tanis, D.G. Augenstein, Noise raises blood pressure without impairing auditory sensitivity, Science 211 (1981) 1450–1452.
- [80] B.M. Altura, B.T. Altura, A. Gebrewold, H. Ising, T. Gunther, Noise-induced hypertension and magnesium in rats: relationship to microcirculation and calcium, J. Appl. Physiol. 72 (1992) 194–202.
- [81] C.C. Wu, S.J. Chen, M.H. Yen, Effects of noise on blood pressure and vascular reactivities, Clin. Exp. Pharmacol. Physiol. 19 (1992) 833–838.
- [82] C.C. Wu, S.J. Chen, M.H. Yen, Attenuation of endothelium-dependent relaxation in mesenteric artery during noise-induced hypertension, J. Biomed. Sci. 1 (1994) 49–53.
- [83] F. Schmidt, K. Kolle, K. Kreuder, B. Schnorbus, P. Wild, M. Hechtner, H. Binder, T. Gori, T. Munzel, Nighttime aircraft noise impairs endothelial function and increases blood pressure in patients with or at high risk for coronary artery disease, Clin. Res. Cardiol. 104 (2015) 23–30, https://doi.org/10.1007/s00392-014-0751-x.
- [84] R.J. Gryglewski, R.M. Palmer, S. Moncada, Superoxide anion is involved in the breakdown of endothelium-derived vascular relaxing factor, Nature 320 (1986)

454-456.

- [85] T. Munzel, A. Daiber, V. Ullrich, A. Mulsch, Vascular consequences of endothelial nitric oxide synthase uncoupling for the activity and expression of the soluble guanylyl cyclase and the cGMP-dependent protein kinase, Arterioscler. Thromb. Vasc. Biol. 25 (2005) 1551–1557, https://doi.org/10.1161/01.ATV.0000168896. 64927.bb.
- [86] C.A. Chen, T.Y. Wang, S. Varadharaj, L.A. Reyes, C. Hemann, M.A. Talukder, Y.R. Chen, L.J. Druhan, J.L. Zweier, S-glutathionylation uncouples eNOS and regulates its cellular and vascular function, Nature 468 (2010) 1115–1118, https://doi.org/10.1038/nature09599.
- [87] A. Daiber, N. Xia, S. Steven, M. Oelze, A. Hanf, S. Kroller-Schon, T. Munzel, H. Li, New therapeutic implications of endothelial nitric oxide synthase (eNOS) function/dysfunction in cardiovascular disease, Int. J. Mol. Sci. 20 (2019) 187, https:// doi.org/10.3390/ijms20010187.
- [88] A. Daiber, M. Oelze, S. Steven, S. Kroller-Schon, T. Munzel, Taking up the cudgels for the traditional reactive oxygen and nitrogen species detection assays and their use in the cardiovascular system, Redox Biol. 12 (2017) 35–49, https://doi.org/ 10.1016/j.redox.2017.02.001.
- [89] E. Schulz, T. Jansen, P. Wenzel, A. Daiber, T. Munzel, Nitric oxide, tetrahydrobiopterin, oxidative stress, and endothelial dysfunction in hypertension, Antioxidants Redox Signal. 10 (2008) 1115–1126, https://doi.org/10.1089/ars. 2007.1989.
- [90] U. Landmesser, S. Dikalov, S.R. Price, L. McCann, T. Fukai, S.M. Holland, W.E. Mitch, D.G. Harrison, Oxidation of tetrahydrobiopterin leads to uncoupling of endothelial cell nitric oxide synthase in hypertension, J. Clin. Invest. 111 (2003) 1201–1209.
- [91] J.K. Bendall, G. Douglas, E. McNeill, K.M. Channon, M.J. Crabtree, Tetrahydrobiopterin in cardiovascular health and disease, Antioxidants Redox Signal. 20 (2014) 3040–3077, https://doi.org/10.1089/ars.2013.5566.
- [92] T. Munzel, A. Daiber, Redox regulation of dihydrofolate reductase: friend or troublemaker? Arterioscler. Thromb. Vasc. Biol. 35 (2015) 2261–2262, https:// doi.org/10.1161/ATVBAHA.115.306556.
- [93] S. Kossmann, H. Hu, S. Steven, T. Schonfelder, D. Fraccarollo, Y. Mikhed, M. Brahler, M. Knorr, M. Brandt, S.H. Karbach, C. Becker, M. Oelze, J. Bauersachs, J. Widder, T. Munzel, A. Daiber, P. Wenzel, Inflammatory monocytes determine endothelial nitric-oxide synthase uncoupling and nitro-oxidative stress induced by angiotensin II, J. Biol. Chem. 289 (2014) 27540–27550, https://doi.org/10.1074/ jbc.M114.604231.
- [94] S. Kasamatsu, Y. Watanabe, T. Sawa, T. Akaike, H. Ihara, Redox signal regulation via nNOS phosphorylation at Ser847 in PC12 cells and rat cerebellar granule neurons, Biochem. J. 459 (2014) 251–263, https://doi.org/10.1042/BJ20131262.
- [95] S.A. Stansfeld, B. Berglund, C. Clark, I. Lopez-Barrio, P. Fischer, E. Ohrstrom, M.M. Haines, J. Head, S. Hygge, I. van Kamp, B.F. Berry, R.s. team, Aircraft and road traffic noise and children's cognition and health: a cross-national study, Lancet 365 (2005) 1942–1949, https://doi.org/10.1016/S0140-6736(05) 66660-3.
- [96] H. Kan, W. Hu, Y. Wang, W. Wu, Y. Yin, Y. Liang, C. Wang, D. Huang, W. Li, NADPH oxidase-derived production of reactive oxygen species is involved in learning and memory impairments in 16-month-old female rats, Mol. Med. Rep. 12 (2015) 4546–4553, https://doi.org/10.3892/mmr.2015.3894.
- [97] T. Munzel, S. Kroeller-Schon, M. Oelze, T. Gori, F.P. Schmidt, S. Steven, O. Hahad, M. Roosli, J.M. Wunderli, A. Daiber, M. Sorensen, Adverse cardiovascular effects of traffic noise with a focus on nighttime noise and the new WHO noise guidelines, Annu. Rev. Publ. Health (2020), https://doi.org/10.1146/annurev-publhealth-081519-062400.
- [98] U. Hink, H. Li, H. Mollnau, M. Oelze, E. Matheis, M. Hartmann, M. Skatchkov, F. Thaiss, R.A. Stahl, A. Warnholtz, T. Meinertz, K. Griendling, D.G. Harrison, U. Forstermann, T. Munzel, Mechanisms underlying endothelial dysfunction in diabetes mellitus, Circ. Res. 88 (2001) E14–E22.
- [99] H. Mollnau, M. Wendt, K. Szocs, B. Lassegue, E. Schulz, M. Oelze, H. Li, M. Bodenschatz, M. August, A.L. Kleschyov, N. Tsilimingas, U. Walter, U. Forstermann, T. Meinertz, K. Griendling, T. Munzel, Effects of angiotensin II infusion on the expression and function of NAD(P)H oxidase and components of nitric oxide/cGMP signaling, Circ. Res. 90 (2002) E58–E65, https://doi.org/10. 1161/01.RES.0000012569.55432.02.
- [100] M. Oelze, H. Mollnau, N. Hoffmann, A. Warnholtz, M. Bodenschatz, A. Smolenski, U. Walter, M. Skatchkov, T. Meinertz, T. Munzel, Vasodilator-stimulated phosphoprotein serine 239 phosphorylation as a sensitive monitor of defective nitric Oxide/cGMP signaling and endothelial dysfunction, Circ. Res. 87 (2000) 999–1005.
- [101] S. Steven, T. Munzel, A. Daiber, Exploiting the pleiotropic antioxidant effects of established drugs in cardiovascular disease, Int. J. Mol. Sci. 16 (2015) 18185–18223, https://doi.org/10.3390/ijms160818185.
- [102] S. Steven, K. Frenis, M. Oelze, S. Kalinovic, M. Kuntic, M.T. Bayo Jimenez, K. Vujacic-Mirski, J. Helmstadter, S. Kroller-Schon, T. Munzel, A. Daiber, Vascular inflammation and oxidative stress: major triggers for cardiovascular disease, Oxid. Med. Cell. Longev. 2019 (2019) 7092151, https://doi.org/10.1155/2019/ 7092151.
- [103] Y. Yu, K. Paul, O.A. Arah, E.R. Mayeda, J. Wu, E. Lee, I.F. Shih, J. Su, M. Jerrett, M. Haan, B. Ritz, Air pollution, noise exposure, and metabolic syndrome - a cohort study in elderly Mexican-Americans in Sacramento area, Environ. Int. 134 (2020) 105269, https://doi.org/10.1016/j.envint.2019.105269.
- [104] A. Daiber, M. Oelze, S. Daub, S. Steven, A. Schuff, S. Kroller-Schon, M. Hausding, P. Wenzel, E. Schulz, T. Gori, T. Munzel, Vascular redox signaling, redox switches in endothelial nitric oxide synthase and endothelial dysfunction, in: I. Laher (Ed.), Systems Biology of Free Radicals and Antioxidants, Springer-Verlag, Berlin

Heidelberg, 2014, pp. 1177-1211.

- [105] S. Karbach, P. Wenzel, A. Waisman, T. Munzel, A. Daiber, eNOS uncoupling in cardiovascular diseases-the role of oxidative stress and inflammation, Curr. Pharmaceut. Des. 20 (2014) 3579–3594.
- [106] J.L. Zweier, C.A. Chen, L.J. Druhan, S-glutathionylation reshapes our understanding of endothelial nitric oxide synthase uncoupling and nitric oxide/reactive oxygen species-mediated signaling, Antioxidants Redox Signal. 14 (2011) 1769–1775, https://doi.org/10.1089/ars.2011.3904.
- [107] F. Wu, W.S. Szczepaniak, S. Shiva, H. Liu, Y. Wang, L. Wang, Y. Wang, E.E. Kelley, A.F. Chen, M.T. Gladwin, B.J. McVerry, Nox2-dependent glutathionylation of endothelial NOS leads to uncoupled superoxide production and endothelial barrier dysfunction in acute lung injury, Am. J. Physiol. Lung Cell Mol. Physiol. 307 (2014) L987–L997, https://doi.org/10.1152/ajplung.00063.2014.
- [108] S. Kroller-Schon, S. Steven, S. Kossmann, A. Scholz, S. Daub, M. Oelze, N. Xia, M. Hausding, Y. Mikhed, E. Zinssius, M. Mader, P. Stamm, N. Treiber, K. Scharffetter-Kochanek, H. Li, E. Schulz, P. Wenzel, T. Munzel, A. Daiber, Molecular mechanisms of the crosstalk between mitochondria and NADPH oxidase through reactive oxygen species-studies in white blood cells and in animal models, Antioxidants Redox Signal. 20 (2014) 247–266, https://doi.org/10.1089/ars. 2012.4953.
- [109] T. Speer, F.O. Owala, E.W. Holy, S. Zewinger, F.L. Frenzel, B.E. Stahli, M. Razavi, S. Triem, H. Cvija, L. Rohrer, S. Seiler, G.H. Heine, V. Jankowski, J. Jankowski, G.G. Camici, A. Akhmedov, D. Fliser, T.F. Luscher, F.C. Tanner, Carbamylated lowdensity lipoprotein induces endothelial dysfunction, Eur. Heart J. 35 (2014) 3021–3032, https://doi.org/10.1093/eurhearti/ehu111.
- [110] E.D. van Deel, Y. Octavia, M. de Boer, R.P. Juni, D. Tempel, R. van Haperen, R. de Crom, A.L. Moens, D. Merkus, D.J. Duncker, Normal and high eNOS levels are detrimental in both mild and severe cardiac pressure-overload, J. Mol. Cell. Cardiol. 88 (2015) 145–154, https://doi.org/10.1016/j.vjmcc.2015.10.001.
- [111] M. Oelze, S. Kroller-Schon, S. Steven, E. Lubos, C. Doppler, M. Hausding, S. Tobias, C. Brochhausen, H. Li, M. Torzewski, P. Wenzel, M. Bachschmid, K.J. Lackner, E. Schulz, T. Munzel, A. Daiber, Glutathione peroxidase-1 deficiency potentiates dysregulatory modifications of endothelial nitric oxide synthase and vascular dysfunction in aging, Hypertension 63 (2014) 390–396, https://doi.org/10.1161/ HYPERTENSIONAHA.113.01602.
- [112] E.D. van Deel, Y. Octavia, M.C. de Waard, M. de Boer, D.J. Duncker, Exercise training has contrasting effects in myocardial infarction and pressure overload due to divergent endothelial nitric oxide synthase regulation, Int. J. Mol. Sci. 19 (2018), https://doi.org/10.3390/ijms19071968.
- [113] M. Oelze, M. Knorr, S. Kroller-Schon, S. Kossmann, A. Gottschlich, R. Rummler, A. Schuff, S. Daub, C. Doppler, H. Kleinert, T. Gori, A. Daiber, T. Munzel, Chronic therapy with isosorbide-5-mononitrate causes endothelial dysfunction, oxidative stress, and a marked increase in vascular endothelin-1 expression, Eur. Heart J. 34 (2013) 3206–3216, https://doi.org/10.1093/eurheartj/ehs100.
- [114] B. Musicki, J.L. Hannan, G. Lagoda, T.J. Bivalacqua, A.L. Burnett, Mechanistic link between erectile dysfunction and systemic endothelial dysfunction in type 2 diabetic rats, Andrology 4 (2016) 977–983, https://doi.org/10.1111/andr.12218.
- [115] Y. Octavia, G. Kararigas, M. de Boer, I. Chrifi, R. Kietadisorn, M. Swinnen, H. Duimel, F.K. Verheyen, M.M. Brandt, D. Fliegner, C. Cheng, S. Janssens, D.J. Duncker, A.L. Moens, Folic acid reduces doxorubicin-induced cardiomyopathy by modulating endothelial nitric oxide synthase, J. Cell Mol. Med. 21 (2017) 3277–3287, https://doi.org/10.1111/jcmm.13231.
- [116] T. Suvorava, S. Pick, G. Kojda, Selective impairment of blood pressure reduction by endothelial nitric oxide synthase dimer destabilization in mice, J. Hypertens. 35 (2017) 76–88, https://doi.org/10.1097/HJH.000000000001127.
- [117] Y. Du, M. Navab, M. Shen, J. Hill, P. Pakbin, C. Sioutas, T.K. Hsiai, R. Li, Ambient ultrafine particles reduce endothelial nitric oxide production via S-glutathionylation of eNOS, Biochem. Biophys. Res. Commun. 436 (2013) 462–466, https://doi. org/10.1016/j.bbrc.2013.05.127.
- [118] F. De Pascali, C. Hemann, K. Samons, C.A. Chen, J.L. Zweier, Hypoxia and reoxygenation induce endothelial nitric oxide synthase uncoupling in endothelial cells through tetrahydrobiopterin depletion and S-glutathionylation, Biochemistry 53 (2014) 3679–3688, https://doi.org/10.1021/bi500076r.
- [119] C.A. Chen, F. De Pascali, A. Basye, C. Hemann, J.L. Zweier, Redox modulation of endothelial nitric oxide synthase by glutaredoxin-1 through reversible oxidative post-translational modification, Biochemistry 52 (2013) 6712–6723, https://doi. org/10.1021/bi400404s.
- [120] Q. Shang, L. Bao, H. Guo, F. Hao, Q. Luo, J. Chen, C. Guo, Contribution of glutaredoxin-1 to S-glutathionylation of endothelial nitric oxide synthase for mesenteric nitric oxide generation in experimental necrotizing enterocolitis, Transl. Res. 188 (2017) 92–105, https://doi.org/10.1016/j.trsl.2016.01.004.
- [121] X. Li, X. Li, Q. Shang, Z. Gao, F. Hao, H. Guo, C. Guo, Fecal microbiota transplantation (FMT) could reverse the severity of experimental necrotizing enterocolitis (NEC) via oxidative stress modulation, Free Radic. Biol. Med. 108 (2017) 32–43, https://doi.org/10.1016/j.freeradbiomed.2017.03.011.
- [122] C. Espinosa-Diez, V. Miguel, S. Vallejo, F.J. Sanchez, E. Sandoval, E. Blanco, P. Cannata, C. Peiro, C.F. Sanchez-Ferrer, S. Lamas, Role of glutathione biosynthesis in endothelial dysfunction and fibrosis, Redox Biol. 14 (2018) 88–99, https://doi.org/10.1016/j.redox.2017.08.019.
- [123] M.J. Crabtree, R. Brixey, H. Batchelor, A.B. Hale, K.M. Channon, Integrated redox sensor and effector functions for tetrahydrobiopterin- and glutathionylation-dependent endothelial nitric-oxide synthase uncoupling, J. Biol. Chem. 288 (2013) 561–569, https://doi.org/10.1074/jbc.M112.415992.
- [124] W.O. Idigo, S. Reilly, M.H. Zhang, Y.H. Zhang, R. Jayaram, R. Carnicer, M.J. Crabtree, J.L. Balligand, B. Casadei, Regulation of endothelial nitric-oxide synthase (NOS) S-glutathionylation by neuronal NOS: evidence of a functional

interaction between myocardial constitutive NOS isoforms, J. Biol. Chem. 287 (2012) 43665–43673, https://doi.org/10.1074/jbc.M112.412031.

- [125] P. Guerby, A. Swiader, N. Auge, O. Parant, C. Vayssiere, K. Uchida, R. Salvayre, A. Negre-Salvayre, High glutathionylation of placental endothelial nitric oxide synthase in preeclampsia, Redox Biol. 22 (2019) 101126, https://doi.org/10. 1016/j.redox.2019.101126.
- [126] W. Wu, P. Geng, J. Zhu, J. Li, L. Zhang, W. Chen, D. Zhang, Y. Lu, X. Xu, KLF2 regulates eNOS uncoupling via Nrf2/HO-1 in endothelial cells under hypoxia and reoxygenation, Chem. Biol. Interact. 305 (2019) 105–111, https://doi.org/10. 1016/j.cbi.2019.03.010.
- [127] J. Helmstadter, K. Frenis, K. Filippou, A. Grill, M. Dib, S. Kalinovic, F. Pawelke, K. Kus, S. Kroller-Schon, M. Oelze, S. Chlopicki, D. Schuppan, P. Wenzel, W. Ruf, D.J. Drucker, T. Munzel, A. Daiber, S. Steven, Endothelial GLP-1 (Glucagon-Like peptide-1) receptor mediates cardiovascular protection by liraglutide in mice with experimental arterial hypertension, Arterioscler. Thromb. Vasc. Biol. 40 (2020) 145–158, https://doi.org/10.1161/atv.0000615456.97862.30.
- [128] J. Vasquez-Vivar, B. Kalyanaraman, P. Martasek, N. Hogg, B.S. Masters, H. Karoui, P. Tordo, K.A. Pritchard Jr., Superoxide generation by endothelial nitric oxide synthase: the influence of cofactors, Proc. Natl. Acad. Sci. U. S. A. 95 (1998) 9220–9225.
- [129] T.J. Guzik, S. Mussa, D. Gastaldi, J. Sadowski, C. Ratnatunga, R. Pillai, K.M. Channon, Mechanisms of increased vascular superoxide production in human diabetes mellitus: role of NAD(P)H oxidase and endothelial nitric oxide synthase, Circulation 105 (2002) 1656–1662.
- [130] N.J. Alp, S. Mussa, J. Khoo, S. Cai, T. Guzik, A. Jefferson, N. Goh, K.A. Rockett, K.M. Channon, Tetrahydrobiopterin-dependent preservation of nitric oxidemediated endothelial function in diabetes by targeted transgenic GTP-cyclohydrolase I overexpression, J. Clin. Invest. 112 (2003) 725–735, https://doi.org/10. 1172/JCI17786.
- [131] J.K. Bendall, N.J. Alp, N. Warrick, S. Cai, D. Adlam, K. Rockett, M. Yokoyama, S. Kawashima, K.M. Channon, Stoichiometric relationships between endothelial tetrahydrobiopterin, endothelial NO synthase (eNOS) activity, and eNOS coupling in vivo: insights from transgenic mice with endothelial-targeted GTP cyclohydrolase 1 and eNOS overexpression, Circ. Res. 97 (2005) 864–871.
- [132] J.B. Laursen, M. Somers, S. Kurz, L. McCann, A. Warnholtz, B.A. Freeman, M. Tarpey, T. Fukai, D.G. Harrison, Endothelial regulation of vasomotion in apoEdeficient mice: implications for interactions between peroxynitrite and tetrahydrobiopterin, Circulation 103 (2001) 1282–1288.
- [133] N.J. Alp, K.M. Channon, Regulation of endothelial nitric oxide synthase by tetrahydrobiopterin in vascular disease, Arterioscler. Thromb. Vasc. Biol. 24 (2004) 413–420.
- [134] K. Chalupsky, H. Cai, Endothelial dihydrofolate reductase: critical for nitric oxide bioavailability and role in angiotensin II uncoupling of endothelial nitric oxide synthase, Proc. Natl. Acad. Sci. U. S. A. 102 (2005) 9056–9061, https://doi.org/ 10.1073/pnas.0409594102.
- [135] I.A. Ionova, J. Vasquez-Vivar, J. Whitsett, A. Herrnreiter, M. Medhora, B.C. Cooley, G.M. Pieper, Deficient BH4 production via de novo and salvage pathways regulates NO responses to cytokines in adult cardiac myocytes, Am. J. Physiol. Heart Circ. Physiol. 295 (2008) H2178–H2187, https://doi.org/10.1152/ ajpheart.00748.2008.
- [136] M.H. Zou, C. Shi, R.A. Cohen, Oxidation of the zinc-thiolate complex and uncoupling of endothelial nitric oxide synthase by peroxynitrite, J. Clin. Invest. 109 (2002) 817–826, https://doi.org/10.1172/JCI14442.
- [137] A. Solini, C. Rossi, E. Duranti, S. Taddei, A. Natali, A. Virdis, Saxagliptin prevents vascular remodeling and oxidative stress in db/db mice. Role of endothelial nitric oxide synthase uncoupling and cyclooxygenase, Vasc. Pharmacol. 76 (2016) 62–71, https://doi.org/10.1016/j.vph.2015.10.002.
- [138] B. Musicki, A.L. Burnett, Constitutive NOS uncoupling and NADPH oxidase upregulation in the penis of type 2 diabetic men with erectile dysfunction, Andrology 5 (2017) 294–298, https://doi.org/10.1111/andr.12313.
- [139] A.E. Loot, J.G. Schreiber, B. Fisslthaler, I. Fleming, Angiotensin II impairs endothelial function via tyrosine phosphorylation of the endothelial nitric oxide synthase, J. Exp. Med. 206 (2009) 2889–2896, https://doi.org/10.1084/jem. 20090449.
- [140] M. Knorr, M. Hausding, S. Kroller-Schuhmacher, S. Steven, M. Oelze, T. Heeren, A. Scholz, T. Gori, P. Wenzel, E. Schulz, A. Daiber, T. Munzel, Nitroglycerin-induced endothelial dysfunction and tolerance involve adverse phosphorylation and S-Glutathionylation of endothelial nitric oxide synthase: beneficial effects of therapy with the ATI receptor blocker telmisartan, Arterioscler. Thromb. Vasc. Biol. 31 (2011) 2223–2231, https://doi.org/10.1161/ATVBAHA.111.232058.
- [141] I. Fleming, B. Fisslthaler, S. Dimmeler, B.E. Kemp, R. Busse, Phosphorylation of Thr(495) regulates Ca(2+)/calmodulin-dependent endothelial nitric oxide synthase activity, Circ. Res. 88 (2001) E68–E75.
- [142] R.H. Boger, Association of asymmetric dimethylarginine and endothelial dysfunction, Clin. Chem. Lab. Med. : CCLM / FESCC 41 (2003) 1467–1472, https:// doi.org/10.1515/CCLM.2003.225.
- [143] K. Sydow, T. Munzel, ADMA and oxidative stress, Atherosclerosis Suppl. 4 (2003) 41–51.
- [144] S.M. Bode-Boger, F. Scalera, L.J. Ignarro, The L-arginine paradox: importance of the L-arginine/asymmetrical dimethylarginine ratio, Pharmacol. Ther. 114 (2007) 295–306, https://doi.org/10.1016/j.pharmthera.2007.03.002.
- [145] E.I. Closs, M.A. Ostad, A. Simon, A. Warnholtz, A. Jabs, A. Habermeier, A. Daiber, U. Forstermann, T. Munzel, Impairment of the extrusion transporter for asymmetric dimethyl-I-arginine: a novel mechanism underlying vasospastic angina, Biochem. Biophys. Res. Commun. (2012), https://doi.org/10.1016/j.bbrc.2012. 05.044.