

# Role of PKC and Ca<sub>v</sub>1.2 in Detrusor Overactivity in a Model of Obesity Associated with Insulin Resistance in Mice

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## Abstract

Obesity/metabolic syndrome are common risk factors for overactive bladder. This study aimed to investigate the functional and molecular changes of detrusor smooth muscle (DSM) in high-fat insulin resistant obese mice, focusing on the role of protein kinase C (PKC) and Ca<sub>v</sub>1.2 in causing bladder dysfunction. Male C57BL/6 mice were fed with high-fat diet for 10 weeks. *In vitro* functional responses and cystometry, as well as PKC and Ca<sub>v</sub>1.2 expression in bladder were evaluated. Obese mice exhibited higher body weight, epididymal fat mass, fasting glucose and insulin resistance. Carbachol (0.001–100 μM), α,β-methylene ATP (1–10 μM), KCl (1–300 mM), extracellular Ca<sup>2+</sup> (0.01–100 mM) and phorbol-12,13-dibutyrate (PDBu; 0.001–3 μM) all produced greater DSM contractions in obese mice, which were fully reversed by the Ca<sub>v</sub>1.2 blocker amlodipine. Cystometry evidenced augmented frequency, non-void contractions and post-void pressure in obese mice that were also prevented by amlodipine. Metformin treatment improved the insulin sensitivity, and normalized the *in vitro* bladder hypercontractility and cystometric dysfunction in obese mice. The PKC inhibitor GF109203X (1 μM) also reduced the carbachol induced contractions. PKC protein expression was markedly higher in bladder tissues from obese mice, which was normalized by metformin treatment. The Ca<sub>v</sub>1.2 channel protein expression was not modified in any experimental group. Our findings show that Ca<sub>v</sub>1.2 blockade and improvement of insulin sensitization restores the enhanced PKC protein expression in bladder tissues and normalizes the overactive detrusor. It is likely that insulin resistance importantly contributes for the pathophysiology of this urological disorder in obese mice.

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## Introduction

Metabolic syndrome is a highly prevalent public health problem that is defined by the coexistence of central obesity, insulin resistance, glucose intolerance, dyslipidemia and arterial hypertension [1]. Metabolic syndrome increases the risk of developing type II diabetes and cardiovascular diseases [2]. Insulin resistance is the central feature of metabolic syndrome, whereby target tissues fail to respond efficiently to normal concentrations of insulin. Obesity is the main etiological factor that predisposes to insulin resistance and vascular diseases [3–5].

Along with the dramatic increase in the incidence of diabetes, obesity and metabolic syndrome, there has been a parallel increase in urological complications [6]. Obesity and metabolic syndrome have been established as common risk factors for lower urinary tract symptoms (LUTS) including overactive bladder and urinary incontinence [7–9]. Overactive bladder is a highly prevalent condition that affects millions of people worldwide with a profound effect on quality of life and considerable costs to health care systems. The third National Health and Nutritional Examination Survey (NHANES III) showed that an increase in

body mass index (BMI) in people over 25 years of age is positively correlated with LUTS, and men with a larger waist circumference (>102 cm) are more likely to exhibit LUTS [10]. However, few studies have explored the mechanisms of bladder dysfunction in metabolic syndrome / diabetes. Male Wistar rats fed with 60% fructose-enriched chow for 6 weeks became overweight, hyperinsulinemic and hyperglycemic, and exhibited unstable preicturition bladder contractions that were suggestive of detrusor overactivity [11]. Female Wistar rats treated with the same high-fructose diet for prolonged periods (24 weeks) showed no increase in body weight, although insulin resistance was observed [12]. Bladders from these animals showed markedly lower bladder contractions to electrical-field stimulation (EFS), carbachol and KCl *in vitro*, along with higher contractions to ATP. A cystometric study carried out in conscious female obese Zucker rats revealed decreased voiding frequency and non-voiding contractions in both obese non-diabetic and obese diabetic animals, suggesting that chronic obesity itself reduces the activity of the bladder, with no significant roles in voiding outcomes due to diabetes [13]. Although human obesity is mostly derived from consumption of high-fat diets combined with low expenditure of energy [14], only

a single experimental study has focused on bladder activity in high-fat diet-induced obesity. Male Sprague-Dawley rats fed with a hyperlipidemia diet for 24 weeks gained more weight and showed an increase in voiding and non-voiding bladder contractions as evaluated by cystometry, suggesting bladder overactivity [15]. Urinary bladder dysfunction during obesity has thus been poorly studied, with conflicting findings that may be due to differences in the strain and sex of test animals, and experimental paradigm.

Protein kinase C (PKC) isoforms have been established as important regulators of lipid-induced insulin resistance [16]. Defects in the diacylglycerol (DAG)-PKC pathway in skeletal muscle and adipocytes are implicated in the insulin-resistant states of obesity and type II diabetes, as observed in animals [17,18] and humans [19]. Under physiological conditions, coupling of muscarinic receptors (mAChR) to  $\text{Ca}_v1.2$  L-type  $\text{Ca}^{2+}$  channels in pig and mice bladder has recently been shown to involve an atypical signaling cascade involving associated PKC and  $\text{Ca}_v1.2$  channels [20]. In the present study we have used a model of metabolic syndrome, in which mice on a high-fat diet develop obesity and insulin resistance, to further investigate the functional and molecular changes in the urinary bladder in this condition. Since elevated  $\text{Ca}^{2+}$  entry via  $\text{Ca}_v1.2$  L-type  $\text{Ca}^{2+}$  channels plays a major role in the altered detrusor contractions observed under diabetic conditions [21], we have also focused on the role of PKC-mediated extracellular  $\text{Ca}^{2+}$  influx through  $\text{Ca}_v1.2$   $\text{Ca}^{2+}$  channels to bladder dysfunction and its relationship with insulin resistance in the high-fat fed obese mice.

## Materials and Methods

### Animals

All animal procedures and experimental protocols are in accordance with the Ethical Principles in Animal Research adopted by the Brazilian College for Animal Experimentation (COBEA) and were approved by the institutional Committee for Ethics in Animal Research/State University of Campinas (CEEA-UNICAMP, protocol number 2067-1). Four-week-old male C57BL6/J mice were provided by the Central Animal House Services of State University of Campinas (UNICAMP).

### Diet-induced Obesity and Treatments

The animals were housed three per cage on a 12 h light–dark cycle, and fed for 10 weeks with either a standard chow diet (carbohydrate: 70%; protein: 20%; fat: 10%) or a high fat diet that induces obesity (carbohydrate: 29%; protein: 16%; fat: 55%), as previously described [22,23].

Lean and obese mice were further divided into two additional groups, namely, animals treated chronically with either the L-type  $\text{Ca}^{2+}$  channel blocker amlodipine (25 mg/kg/day, given in the drinking water from the 7<sup>th</sup> to the 10<sup>th</sup> week) [24] or the insulin sensitizing agent metformin (300 mg/kg/day, given by gavage from the 8<sup>th</sup> to the 10<sup>th</sup> week) [25]. Tail-cuff pressure was evaluated in all groups, adapted to mice.

### In vivo Insulin Sensitivity

After 6 h fasting, systemic insulin sensitivity was analyzed by the Insulin Tolerance Test (ITT) which was performed. Tail blood samples were collected before (0 min) and at 5, 10, 15, 20, 25 and 30 min after an intraperitoneal injection of 1.00 U/Kg of regular insulin (Novolin R, NovoNordisk, Bagsvaerd, Denmark). Glucose concentrations were measured using a glucometer (ACCUCHEK Performa; Roche Diagnostics, Indianapolis, IN, USA) and the values were used to calculate the constant rate for blood glucose disappearance (Kitt), based on the linear regression of the naperian

logarithm of glucose concentrations obtained from 0 to 30 min of the test. Kitt was calculated using the formula  $0.693/(t1/t2) \times -1 \times 100$ .

### In vitro Functional Assays

Mice were anesthetized with isoflurane, and urinary bladders removed and sectioned horizontally at the level of the ureters. Two longitudinal detrusor smooth muscle (DSM) strips with intact urothelium were obtained from each bladder. Strips of DSM were mounted in 10 ml organ baths containing Krebs-Henseleit solution with the following composition (mM): 117 NaCl, 4.7 KCl, 2.5  $\text{CaCl}_2$ , 1.2  $\text{MgSO}_4$ , 1.2  $\text{KH}_2\text{PO}_4$ , 25  $\text{NaHCO}_3$  and 11 glucose, pH 7.4, at 37°C and bubbled with a gas mixture of 95%  $\text{O}_2$  and 5%  $\text{CO}_2$ . Changes in isometric force were recorded using a Power Lab v.7.0 system (Colorado Springs, CO, USA). The resting tension was adjusted to 0.5 g at the beginning of the experiments. The equilibration period was 60 min and the bathing medium was changed every 15 min.

Nonlinear regression analysis to determine the  $\text{pEC}_{50}$  was carried out using GraphPad Prism (GraphPad Software, San Diego, CA, USA) with the constraint that  $\Phi = 0$ . All concentration–response data were evaluated for a fit to a logistics function in the form:

$$E = E_{\max} / ([1 + (10^c / 10^x)^n] + \Phi).$$

$E$  is the maximum response produced by agonists;  $c$  is the logarithm of the  $\text{EC}_{50}$ , the concentration of drug that produces a half-maximal response;  $x$  is the logarithm of the concentration of the drug; the exponential term,  $n$ , is a curve-fitting parameter that defines the slope of the concentration–response line and  $\Phi$  is the response observed in the absence of added drug. The values of  $\text{pEC}_{50}$  data represent the mean  $\pm$  S.E. Maximal response ( $E_{\max}$ ) data were normalized to the wet weight of the respective urinary bladder strips, and the values of  $E_{\max}$  were represented by mN per milligram wet weight.

### Contractile Responses to Carbachol, $\alpha, \beta$ -methylene ATP, KCl, Extracellular $\text{CaCl}_2$ and phorbol-12,13-dibutyrate (PDBu) in Mice DSM

Cumulative concentration–response curves to the mAChR agonist carbachol (0.001–100  $\mu\text{M}$ ), the PKC activator PDBu (0.001–3  $\mu\text{M}$ ) and KCl (1–300 mM) in DSM strips were performed. Contractile response to the P2X1 purinergic agonist  $\alpha, \beta$ -methylene ATP were obtained through non-cumulative addition of three different drug concentrations (1, 3 and 10  $\mu\text{M}$ ).

Cumulative concentration–response curves to extracellular  $\text{CaCl}_2$  (0.01–30 mM) in depolarizing conditions were constructed. The strips were prepared and mounted in 10 ml organ baths containing Krebs-Henseleit  $\text{Ca}^{2+}$ -free solution containing EGTA (1 mM) to sequester  $\text{Ca}^{2+}$  ions, and cyclopiazonic acid (CPA, 1  $\mu\text{M}$ ) to deplete sarcoplasmic reticulum  $\text{Ca}^{2+}$  stores. Next, bath solution was removed and replaced by Krebs-Henseleit  $\text{Ca}^{2+}$ -free solution containing KCl (80 mM) and CPA (1  $\mu\text{M}$ ). After an equilibration period of 15 min, the cumulative curve to  $\text{CaCl}_2$  was constructed [26].

The concentration–response curves to carbachol, KCl, PDBu and extracellular  $\text{CaCl}_2$  in DSM strips obtained from both lean and obese mice were performed in the absence and in the presence of either the L-type  $\text{Ca}^{2+}$  channel blocker amlodipine (3  $\mu\text{M}$ , 30 min) or antidiabetic agent metformin (1  $\mu\text{M}$ , 30 min). Concentration–response curves to carbachol were also performed in

the presence of the PKC inhibitor GF109203X (1  $\mu$ M, 30 min). Contractile responses to PDBu was also evaluated in a  $\text{Ca}^{2+}$ -free Krebs buffer medium. Contractile responses to these agents were also evaluated in DSM strips from lean and obese mice treated chronically with amlodipine (25 mg/kg/day for 21 days) or metformin (300 mg/kg/day, 14 days).

### Electrical-field Stimulated-induced DSM Contractions

Frequency-response curves (1–32 Hz) were elicited by stimulating the tissues for 10 s with pulses of 1 ms width at 80 V, with 3 min interval between stimulations. Subsequently after 30 min incubation periods, frequency-response curves were repeated in the presence of the non-selective mAChR antagonist atropine (1  $\mu$ M, 30 min) and the P2X receptor blocker PPADS (30  $\mu$ M, 30 min), to confirm the mediation by mAChR and P2X receptor activation. Frequency-response curves were also performed in the presence of the voltage-gated sodium channel blocker tetrodotoxin (1  $\mu$ M, 30 min) to confirm the neurogenic nature of the contractions.

### Cystometry

Mice were anaesthetized with an intraperitoneal injection of urethane (1.8 g/kg). A 1 cm abdominal incision was made, the bladder was exposed and a butterfly cannula (25 G) was inserted into the bladder dome. The cannula was connected to a three-way tap, one port of which was connected to a pressure transducer and the other to the infusion pump through a catheter (PE50). Before starting the cystometry, the bladder was emptied via the third port. Continuous cystometry was carried out by infusing saline into the bladder at a rate of 0.6 ml/h. The following parameters were assessed: Threshold pressure (TP; the intravesical pressure immediately before micturition); Post-void pressure (PVP; the intravesical pressure immediately after micturition); Peak pressure (PP; the peak pressure reached during micturition); Capacity (CP; the volume of saline needed to induce the first micturition); Compliance (CO; the ratio of CP to TP); Frequency of voiding contractions (VC) and frequency of non-voiding contractions (NVCs). NVCs were defined as spontaneous bladder contractions greater than 4 mmHg from the baseline pressure that did not result in a void. Bladders from mice used in the cystometry were not used in the other experiments.

### Western Blotting Detection of PKC and $\text{Ca}_v1.2$ Calcium Channels in Urinary Bladder

Bladder tissues were homogenized in a SDS lysis buffer with a Polytron PTA 20S generator (model PT 10/35; Brinkmann Instruments, Inc., Westbury, NY) operated at maximum speed for 30 sec and centrifuged (12,000 $\times$ g, 4°C, 20 min) to remove insoluble material. Protein concentrations of the supernatants were determined by the Bradford assay, and equal amount of protein from each sample (50  $\mu$ g) was treated with Laemmli buffer containing dithiothreitol 100 mM. Samples were heated in a boiling water bath for 15 min and resolved by SDS-PAGE. Electrotransfer of proteins to nitrocellulose membrane was performed for 60 min at 15 V (constant) in a semi-dry device (Bio-Rad, Hercules, CA, USA). Nonspecific protein binding to nitrocellulose was reduced by pre-incubating the membrane overnight at 4°C in blocking buffer (0.5% non-fat dried milk, 10 mM Tris, 100 mM NaCl, and 0.02% Tween 20). Detection using specific antibodies, HRP-conjugated secondary antibodies, and luminol solution was performed, as described previously [27]. Anti-PKC (ab59363), anti-DHPR $\alpha$ 1 subunit (ab58552) antibodies were obtained from AbCam Technology (Cambridge, England,

UK) and anti GAPDH (SC25778) was from Santa Cruz Biotechnology (Santa Cruz, CA, USA). Densitometry was performed using the Scion Image software (Scion Corporation, Frederick, MD) and results represented as the ratio of the density of the PKC /  $\text{Ca}_v1.2$ - $\alpha$ 1 band to the density of the GAPDH band.

### Drugs

Urethane, metformin, carbachol, pyridoxal phosphate-6-azophenyl-2',4'-disulfonic acid (PPADS),  $\alpha,\beta$ -methylene ATP, atropine, amlodipine, tetrodotoxin, phorbol-12,13-dibutyrate (PDBu) and cyclopiazonic acid were obtained from Sigma (St. Louis, MO, USA).

### Statistical Analysis

Data are expressed as mean  $\pm$  SEM of *n* experiments. In the cumulative concentration and frequency-response curves data were expressed as mean of the contraction in mN/mg of wet strip weight  $\pm$  SEM of *n* experiments. The program Instat (GraphPad Software) was used for statistical analysis. One-way analysis of variances (ANOVA) followed by a Tukey test was used in all groups, and  $p < 0.05$  was accepted as significant.

## Results

### Body Weight, Epididymal Fat Mass and Insulin Sensitivity

Obese mice exhibited a significant increase in body weight and epididymal fat mass compared with lean mice ( $p < 0.001$ ; Table 1). Bladder weight was not significantly modified between lean and obese groups. Fasting glucose levels were increased by 70% in obese group ( $p < 0.001$ ; Table 1), whereas insulin sensitivity was markedly reduced when compared with lean mice ( $p < 0.05$ ), as evidenced by the curve representing glucose decay and Kitt values (Figure 1A and B). Tail-cuff pressure was not modified in obese compared with lean group ( $84 \pm 3.4$  and  $86 \pm 3.3$  mmHg, respectively).

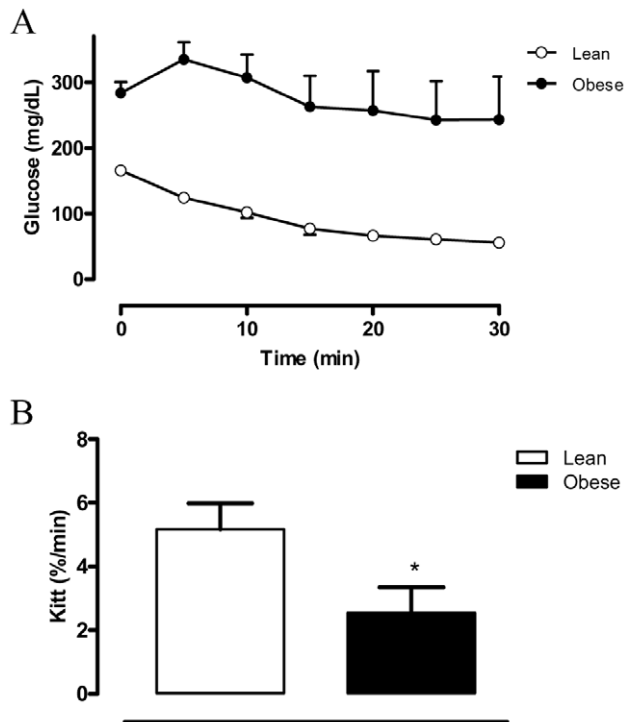
### In vitro Functional Assays

Figure 2A shows that the mAChR agonist carbachol (1 nM – 100  $\mu$ M) produced concentration-dependent contractions in strips of isolated DSM with a maximal response ( $E_{\text{max}}$ ) greater in the obese ( $p < 0.01$ ) than the lean group ( $5.02 \pm 0.89$  and  $1.83 \pm 0.36$  mN/mg, respectively;  $n = 6-7$ ). No significant differences for the  $p\text{EC}_{50}$  values for carbachol were found between the lean and obese groups ( $6.23 \pm 0.09$  and  $6.11 \pm 0.06$ , respectively).

Contractile responses to the P2X receptor agonist  $\alpha,\beta$ -methylene ATP (1 – 10  $\mu$ M) did not significantly differ between control and obese groups ( $n = 6-7$ ; Figure 2B).

Electrical-field stimulation (EFS; 1–32 Hz) produced frequency-dependent DSM contractions in both groups, which were greater in obese mice at all frequencies employed (Figure 2C;  $n = 6$ ). Pretreatment of DSM preparations with the mAChR antagonist atropine (1  $\mu$ M) together with the purinergic receptor blocker PPADS (30  $\mu$ M) markedly reduced ( $p < 0.001$ ) the EFS-induced contractions in both lean ( $E_{\text{max}}$ :  $1.57 \pm 0.25$  and  $0.37 \pm 0.09$  mN/mg for untreated and treated preparations, respectively;  $n = 6$ ) and obese mice ( $E_{\text{max}}$ :  $2.82 \pm 0.38$  and  $0.93 \pm 0.19$  mN/mg for untreated and treated preparations, respectively;  $n = 6$ ). Incubation with the voltage-gated sodium channel blocker tetrodotoxin (1  $\mu$ M) almost abolished the EFS-elicited contractions at all frequencies tested ( $n = 4$ , data not shown).

Cumulative concentration-response curves to the receptor-independent agents KCl and  $\text{CaCl}_2$  were also obtained in DSM strips from obese and lean mice (Figure 2 D and E). Potassium chloride (KCl; 1 – 300 mM;  $n = 6-7$ ) and  $\text{CaCl}_2$  (0.01 – 100 mM;



**Figure 1. Insulin tolerance test (ITT): (A) insulin sensitivity test after intraperitoneal injection of insulin (1.00 U/kg body wt) was performed on lean and obese groups.** Blood samples were collected from the tail at indicated time points and analyzed for glucose concentration; (B) the constant rate for blood glucose disappearance (Kitt), based on the linear regression of the neperian logarithm of glucose concentrations, obtained from indicated time points. Results are expressed as mean  $\pm$  SEM from 5–7 animals in each group. doi:10.1371/journal.pone.0048507.g001

$n = 5-6$ ) produced concentration-dependent DSM contractions with  $E_{max}$  significantly greater in obese mice for both agents ( $p < 0.05$ ) compared with the lean group (Figure 2D and E). No differences at the  $pEC_{50}$  levels for KCl and  $CaCl_2$  were found between lean ( $1.25 \pm 0.07$  and  $2.19 \pm 0.08$ , respectively) and obese groups ( $1.36 \pm 0.07$  and  $2.14 \pm 0.06$ , respectively).

#### Effect of $Ca_v1.2$ L-type $Ca^{2+}$ Channels Blockade by Amlodipine

Since KCl and  $CaCl_2$ -induced contractions were enhanced in strips taken from obese DSM, we hypothesized that an increase in  $Ca^{2+}$  entry through L-type voltage-operated  $Ca^{2+}$  channels is likely

to play a key role in the overactive DSM. *In vitro* incubation of DSM with the dihydropyridine calcium channel blocker amlodipine ( $3 \mu M$ ;  $n = 4-5$ ) nearly normalized the enhanced contractile responses ( $E_{max}$ ) to carbachol in obese mice, with a small but significant inhibition of contraction in the lean mice (Figure 3 A and B). Similarly to carbachol, the enhanced contractile responses to KCl (Figure 3 C and D;  $n = 5-7$ ) and  $CaCl_2$  (Figure 3 E and F;  $n = 4-5$ ) in obese mice were prevented by *in vitro* incubation with amlodipine.

In separate groups, obese and lean mice were treated orally with amlodipine ( $25 \text{ mg/kg/day}$ , 21 days), and contractile responses of DSM strips to carbachol ( $n = 6-7$ ), KCl ( $n = 5-10$ ) and  $CaCl_2$  ( $n = 5-11$ ) were obtained. Long-term amlodipine administration prevented the enhancement of contractions to carbachol (Figure 4 A and B), KCl (Figure 4 C and D) and  $CaCl_2$  (Figure 4 E and F) in obese mice. In lean group, oral amlodipine did not significantly alter the contractile responses to carbachol and KCl, but significantly reduced the  $CaCl_2$ -induced DSM contractions.

#### Role for Insulin Resistance in the Overactive DSM in Obese Mice

Chronic treatment with the anti-hyperglycemic agent metformin ( $300 \text{ mg/kg/day}$ , 14 days) did not significantly affect body weight and epididymal fat mass in obese or lean mice (Table 1,  $n = 10$ ). However, metformin treatment normalized the lower insulin sensitivity seen in obese mice, restoring the Kitt to control values ( $5.17 \pm 0.81$ ,  $4.68 \pm 1.23$ ,  $2.54 \pm 0.80$  and  $5.76 \pm 0.38$  % / min for lean untreated, lean treated, obese untreated and obese treated, respectively;  $n = 4-6$ ), as well as the fasting blood glucose values ( $166 \pm 10$  and  $162 \pm 5.2$  mg/dL for lean and obese treated, respectively;  $n = 5$ ).

Chronic metformin treatment had no significant effect on DSM contractions in lean mice (Figure 4 A, C and E). However, the increase in DSM contractions to carbachol (Figure 4B), KCl (Figure 4D) and  $CaCl_2$  (Figure 4F) in obese mice was suppressed by chronic metformin treatment ( $n = 6-11$ ). In contrast, *in vitro* incubation of DSM strips with metformin ( $1 \mu M$ , 30 min) had no effect on DSM contractions in lean or obese mice (Figure 3).

#### Cystometric Studies

During cystometry, lean mice showed regular micturition cycles with rare non-voiding contractions (Figure 5A). In contrast, obese mice exhibited an irregular micturition pattern (Figure 5B) and significant increases ( $p < 0.01$ ) in the frequency of voiding and non-voiding contractions (Figure 6 A and B;  $n = 5-7$ ). The post-void pressure (PVP), which reflects the efficiency of bladder emptying, was markedly higher ( $p < 0.01$ ) in obese mice compared with the lean group (Figure 6C). The bladder capacity, threshold pressure,

**Table 1. Body weight, epididymal fat mass, bladder weight and glucose levels in lean and high-fat fed obese mice, treated or not with metformin ( $300 \text{ mg/kg/day}$ , 14 days).**

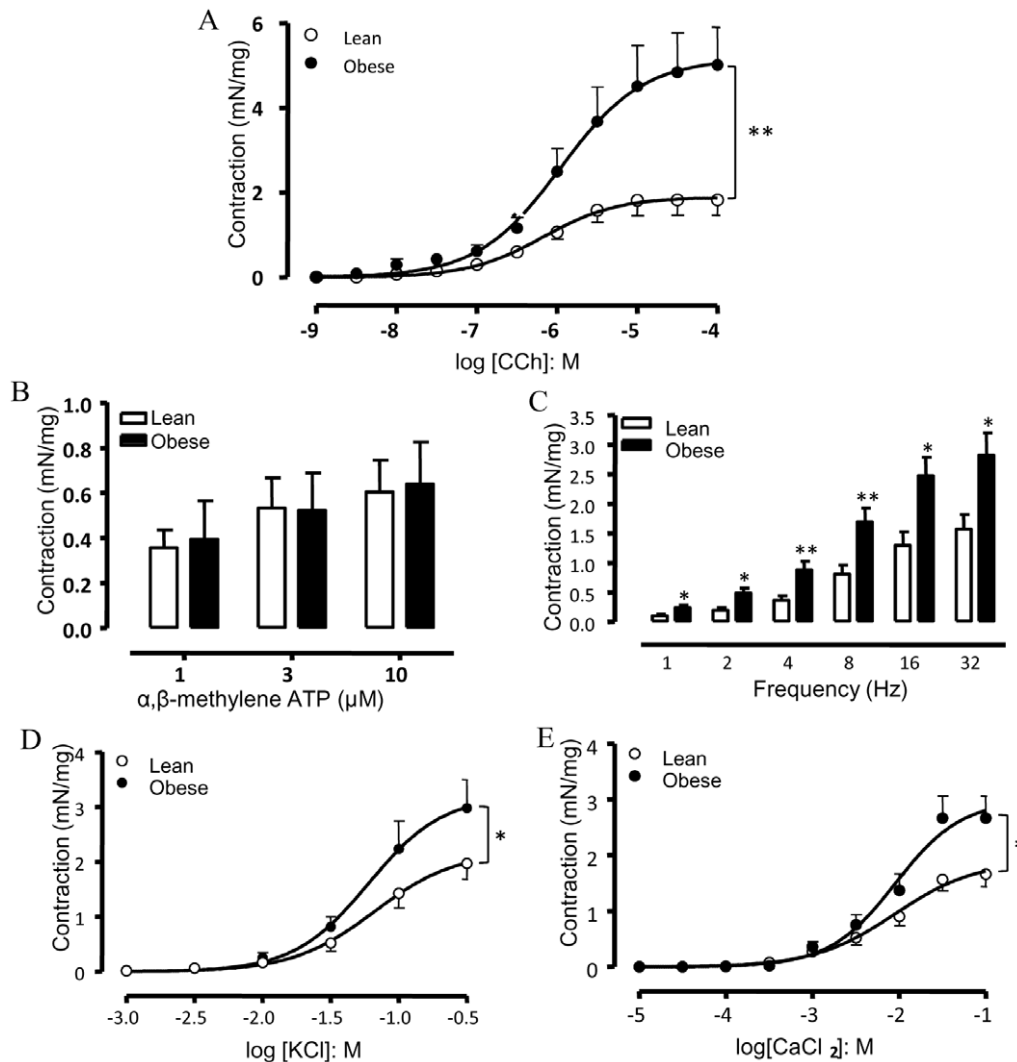
	Lean	Lean + Metformin	Obese	Obese + Metformin
Body weight (g)	$28.5 \pm 0.3$ (9)	$28.4 \pm 0.3$ (10)	$45.7 \pm 1.1^{***}$ (9)	$43.7 \pm 0.9^{***}$ (10)
Epididymal fat mass (g)	$0.25 \pm 0.01$ (9)	$0.39 \pm 0.3$ (10)	$1.71 \pm 0.08^{***}$ (9)	$1.86 \pm 0.1^{***}$ (10)
Bladder weight (mg)	$24.9 \pm 4.1$ (6)	$25.7 \pm 0.8$ (7)	$27.9 \pm 6.9$ (6)	$27.1 \pm 0.9$ (7)
Glucose (mg/dl)	$139 \pm 6.6$ (9)	$166 \pm 10$ (5)	$238 \pm 12^{***}$ (10)	$162 \pm 5.2^{\#}$ (5)

Data represent the means  $\pm$  SEM for 5–10 mice.

\*\*\* $p < 0.001$  compared with lean group;

$\#p < 0.001$  compared with respective obese group.

doi:10.1371/journal.pone.0048507.t001



**Figure 2. Detrusor smooth muscle contraction in response to the muscarinic agonist carbachol (A), the purinergic P2X agonist  $\alpha,\beta$ -methylene-ATP (B), electrical-field stimulation (C), KCl (D) and  $\text{CaCl}_2$  (E) in bladder strips from control and obese mice.** Data represent the mean  $\pm$  SEM for 6 to 7 mice each group. \*  $p < 0.05$ , \*\*  $p < 0.01$  compared with control group. doi:10.1371/journal.pone.0048507.g002

compliance and peak pressure did not significantly differ between lean and obese mice (Table 2).

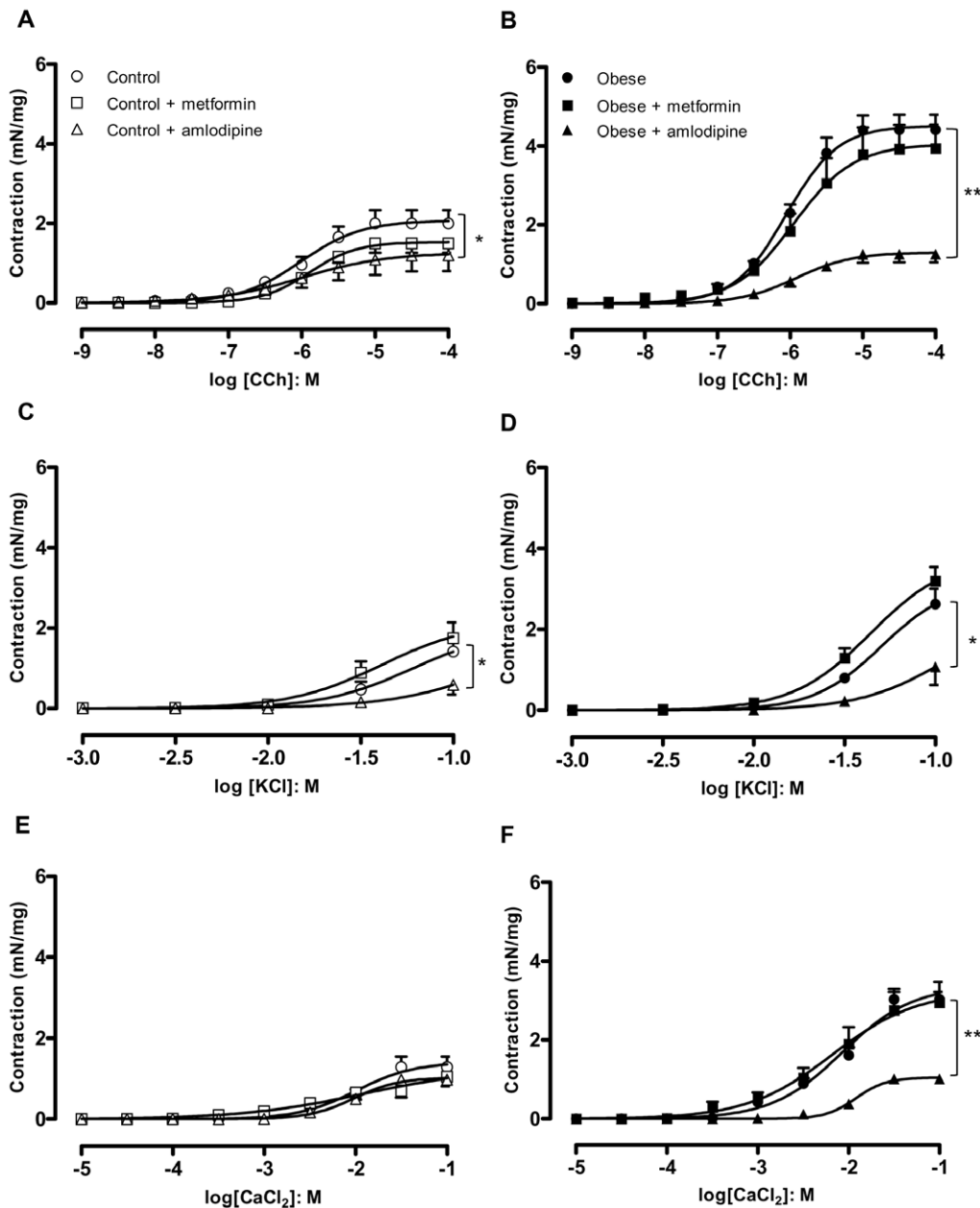
Since amlodipine normalized the overactive DSM in obese mice, we further investigated its effects on the urodynamic changes. Long-term treatment with amlodipine had no significant effects on the cystometric parameters of lean mice ( $n = 5-7$ ). However, this treatment prevented the increased frequencies of voiding and non-voiding contractions, and prevented the increased PVP seen in the obese group (Figures 5C and 6). The other cystometric parameters (capacity, threshold pressure, compliance, and peak pressure) remained unchanged in amlodipine-treated mice (Table 2).

As the improvement of insulin sensitivity by chronic treatment with metformin also normalized the overactive DSM in obese mice, we next investigated the effect of this anti-hyperglycemic agent on the cystometric alterations. The increased frequency of voiding and non-voiding contractions, as well as the increased PVP in obese mice was normalized by metformin treatment (Figures 5D and 6). Metformin treatment did not significantly affect these parameters in lean mice (Figure 6). The other

cystometric parameters remained unchanged in metformin-treated mice (Table 2).

### Role of Protein Kinase C (PKC) in the Enhanced Contractile-responses

As PKC regulates  $\text{Ca}_v1.2$  calcium channel activity in various smooth muscle cell types, including DSM [20], we investigated the role of PKC in the enhanced DSM contractions in obese mice, by using functional and molecular approaches. The PKC inhibitor GF109203X (1  $\mu\text{M}$ ) normalized the enhanced carbachol-induced contractions in obese mice (Figure 7A). In addition, the PKC activator PDBu (0.001–3  $\mu\text{M}$ ) produced concentration-dependent contractions in strips of isolated DSM that were greater in the obese than the lean group (Figure 7B;  $n = 4-7$ ;  $p < 0.01$ ). Prior incubation with amlodipine (3  $\mu\text{M}$ ) abolished the PDBu-induced DSM contractions in both obese and lean mice (Figure 7A). Similar data were obtained with chronic treatment with amlodipine ( $n = 4$ ; not shown). No contractile responses to PDBu were observed in  $\text{Ca}^{2+}$ -free medium ( $n = 3$ ; not shown).



**Figure 3. Cumulative concentration-response curves to carbachol (A and B), KCl (C and D) and CaCl<sub>2</sub> (E and F) in the presence of *in vitro* pre-incubated metformin (1  $\mu$ M) or amlodipine (3  $\mu$ M) in detrusor smooth muscle from lean and obese mice.** Data represent the mean  $\pm$  SEM for 4 to 7 mice each group. \*  $p < 0.05$ , \*\*  $p < 0.01$  compared with untreated group. doi:10.1371/journal.pone.0048507.g003

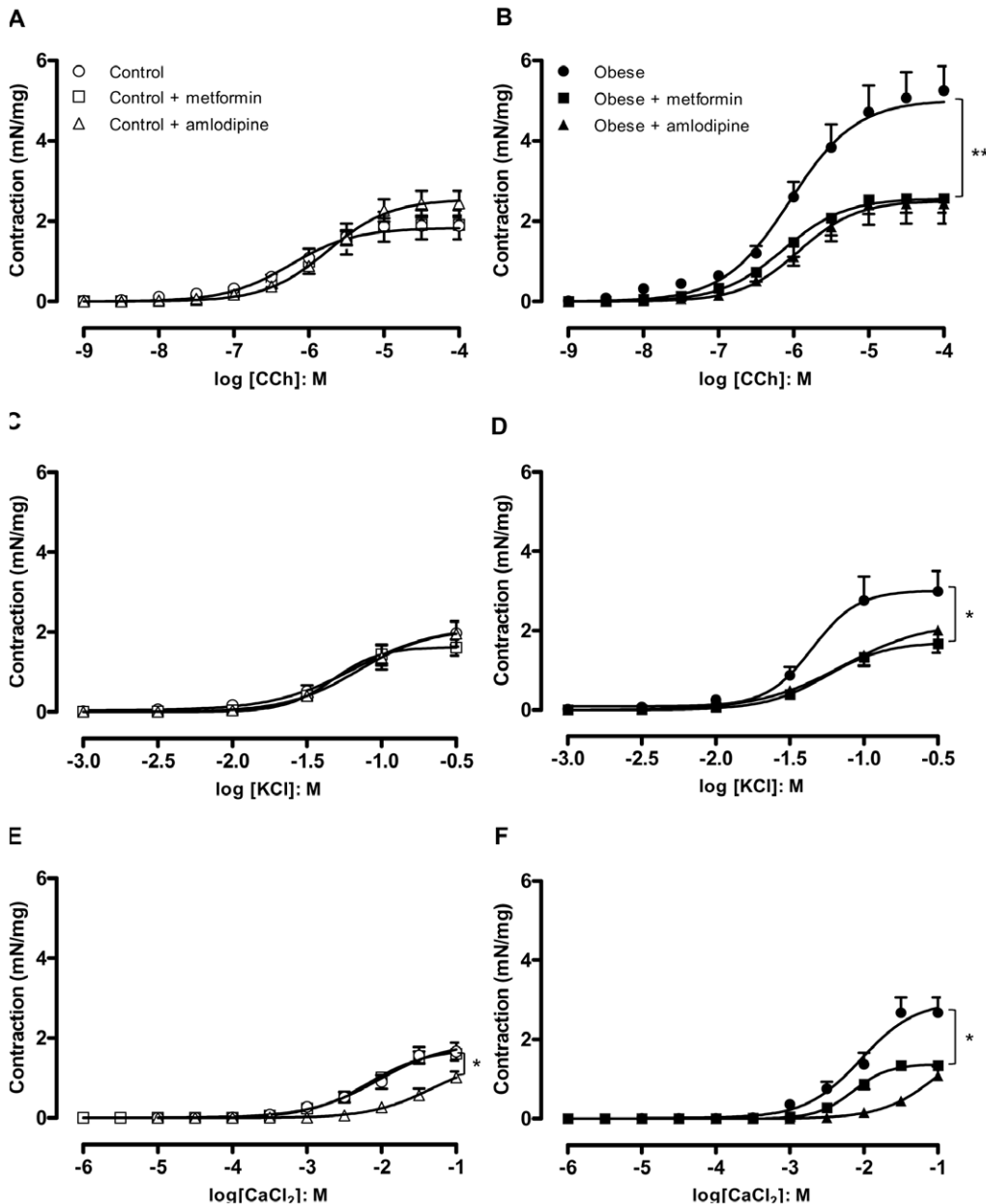
The PKC protein expression was significantly higher in bladder tissues from obese mice ( $p < 0.05$ ), and the increase in PKC expression was abrogated by amlodipine or metformin (Figure 7 C, D and E). Neither of these treatments affected the expression of PKC in the lean group. Expression of the  $\alpha 1$  subunit of Ca<sub>v</sub>1.2 was equivalent in all treatment groups (Figure 7C, D and E).

## Discussion

The present study shows that high-fat fed obese mice display overactive bladder and enhanced PKC protein expression in bladder tissues that are normalized by blockade of Ca<sub>v</sub>1.2 and improvement of insulin sensitivity. It is likely that insulin resistance

in obese mice plays a major role in the pathophysiology of this urological disorder.

The term “metabolic syndrome” describes the combination of metabolic abnormalities or risk factors for type II diabetes and cardiovascular diseases, such as central obesity, dyslipidemia, hypertension, insulin resistance and glucose intolerance [2,28]. Among these risk factors, central obesity is regarded to be the major determinant criteria for metabolic syndrome [4]. In the model of diet-induced obesity employed here, mice exhibited increased body weight, epididymal fat mass (corresponding to central obesity in human) and insulin resistance. Previous studies have also identified dyslipidemia and impaired glucose tolerance in high-fat fed mice [22,23]. These changes suggest that this mouse



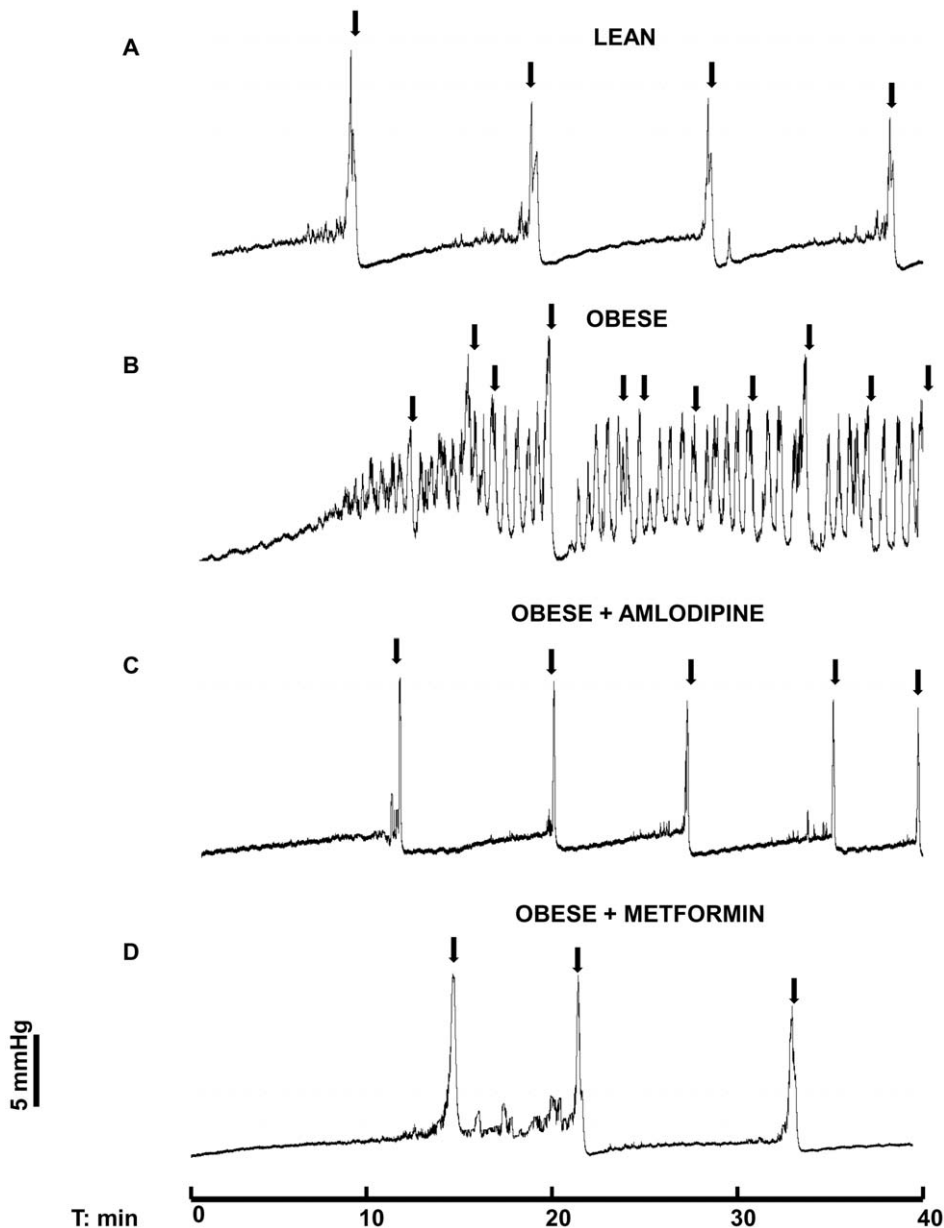
**Figure 4. Cumulative concentration-response curves to carbachol (A and B), KCl (C and D) and CaCl<sub>2</sub> (E,F) in detrusor smooth muscle from lean and obese mice chronically treated with metformin (300 mg/kg/day, 14 days) or amlodipine (25 mg/kg/day, 21 days).** Data represent the mean  $\pm$  SEM for 5 to 11 mice each group. \*  $p < 0.05$ , \*\*  $p < 0.01$  compared with untreated group. doi:10.1371/journal.pone.0048507.g004

model of metabolic syndrome closely mirrors the changes in humans with metabolic syndrome / type II diabetes. Obese mice did not show increased tail-cuff pressure, excluding arterial hypertension as a cause for the present functional bladder alterations.

Epidemiological studies support a strong causal link between obesity and the development of urinary incontinence [29]. Data from our urodynamic study revealed detrusor overactivity in obese mice, as evidenced by the higher frequencies of micturition and non-voiding contractions (NVC), as well as the increased post-voiding pressure (PVP). On the other hand, the bladder capacity, threshold pressure, compliance and peak pressure did not significantly change in the obese group. This contrasts with streptozotocin-induced type I diabetes in mice, where all of these

parameters are increased in comparison with control animals [21], possibly as a consequence of the enhanced urine output (diuresis) and the resulting bladder remodeling [30]. Obesity / metabolic syndrome markedly increase the risk of benign prostatic hyperplasia (BPH) that is in turn a risk factor for detrusor overactivity resultant from bladder outlet obstruction [31]. High fat-fed insulin-resistant Sprague Dawley rats exhibit prostate enlargement that is prevented by treatment with the anti-hyperglycemic agent pioglitazone [32]. Therefore, in our study, whether the increased PVP and poor bladder emptying efficiency seen in obese mice reflect a failure to compensate for enhanced urethral resistance due to prostate enlargement requires further studies [32].

Urinary bladder function is regulated by a complex interaction of efferent and afferent fibers from the autonomic nervous system

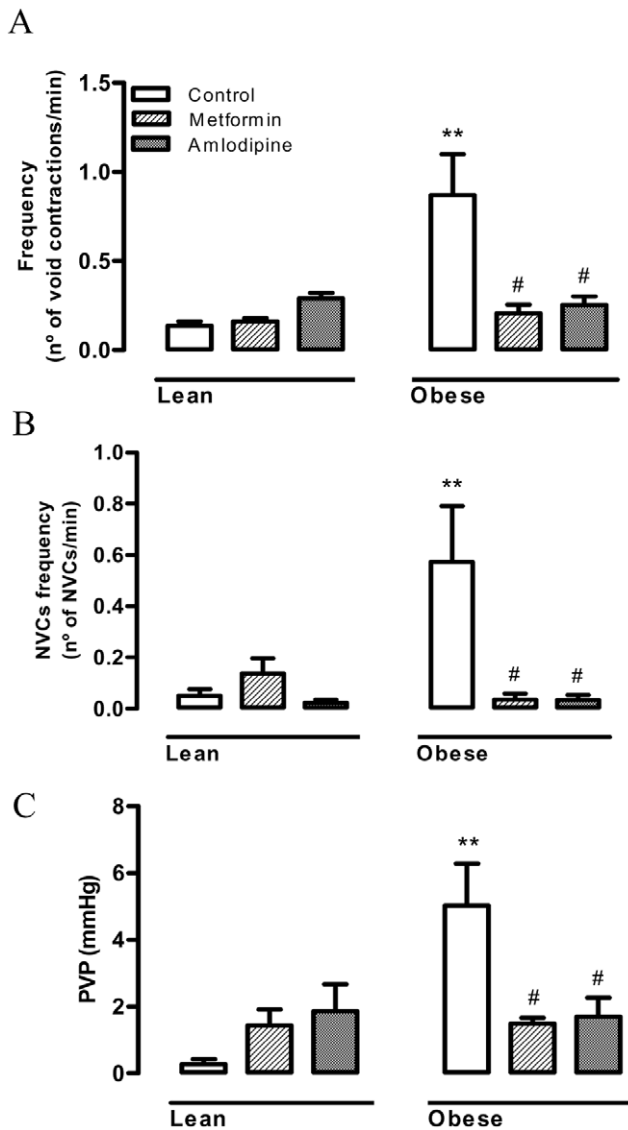


**Figure 5. Representative cystometric recordings from lean (A), obese (B), obese treated with amlodipine (C) and obese treated with metformin (D) mice.** Arrows in the cystometric trace indicate the micturition peaks.  
doi:10.1371/journal.pone.0048507.g005

and somatic innervations [33]. The neurogenic contractions of the bladder mainly reflect the release of the excitatory transmitter acetylcholine (ACh) from parasympathetic fibers. The bladder contains all mAChR subtypes, but the  $M_3$  receptor is responsible for the urinary bladder contractions [34]. The excitatory transmitter ATP, through ionotropic P2X1 receptors, also mediates part of the atropine-resistant neurogenic bladder contractions under normal and pathophysiological conditions, although to varying extents across species from rodent to man [35]. There is also a muscarinic- and purinergic-resistant neurogenic component that has not yet been fully characterized [36,37]. In our study, neurogenic- and carbachol-induced DSM contractions were greater in obese mice compared with lean mice, whereas no significant differences for the purinergic P2X1 agonist  $\alpha, \beta$ -methylene ATP were found.

Muscarinic  $M_3$  receptors interact with  $G_q$  to elicit phosphoinositide hydrolysis and generation of the second messenger inositol-1,4,5-trisphosphate ( $IP_3$ ), which activates the  $IP_3$  receptor to release  $Ca^{2+}$  from internal stores [34]. Muscarinic agonist-induced contractions have also been shown to partly depend on  $Ca^{2+}$  entry through  $Ca_v1.2$  channels, as evidenced in animal and human bladders under physiological conditions [38–42]. High levels of extracellular  $K^+$  depolarize the cell membrane and activate  $Ca_v1.2$  channels, resulting in increased inward movement of  $Ca^{2+}$ , which in turn activates contractile proteins [43]. In smooth muscle tissues, dihydropyridine  $Ca^{2+}$  channel blockers inhibit the increase in  $[Ca^{2+}]_i$  induced by high  $K^+$  or extracellular  $CaCl_2$ . Our findings that DSM strips obtained from obese mice show greater contractile responses to both receptor-dependent (carbachol) and -independent agents (KCl and extracellular





**Figure 6. Cystometric parameters in lean and obese mice treated or not with metformin (300 mg/kg/day, 14 days) or amlodipine (25 mg/kg/day, 21 days).** (A) Frequency of voiding contractions, (B) Frequency of non-voiding contractions and (C) Post-void pressure. Data represent the mean  $\pm$  SEM for 5–7 mice each group. \*\*  $p < 0.01$  compared with control group, #  $p < 0.01$  compared with untreated obese mice. doi:10.1371/journal.pone.0048507.g006

$\text{CaCl}_2$ ) strongly suggest that enhanced extracellular  $\text{Ca}^{2+}$  entry via  $\text{Ca}_v1.2$  channels plays a critical role in the overactive DSM in obese mice. This is reinforced by our data showing that pretreatment with the dihydropyridine  $\text{Ca}^{2+}$  channel blocker amlodipine fully reversed the enhanced contractile responses to carbachol, KCl and  $\text{CaCl}_2$  in obese mice. The urodynamic alterations in the obese group were also reversed by treatment of mice with amlodipine, which is consistent with the reversal of the enhanced DSM contractions *in vitro* to carbachol, KCl and extracellular  $\text{CaCl}_2$  by amlodipine.

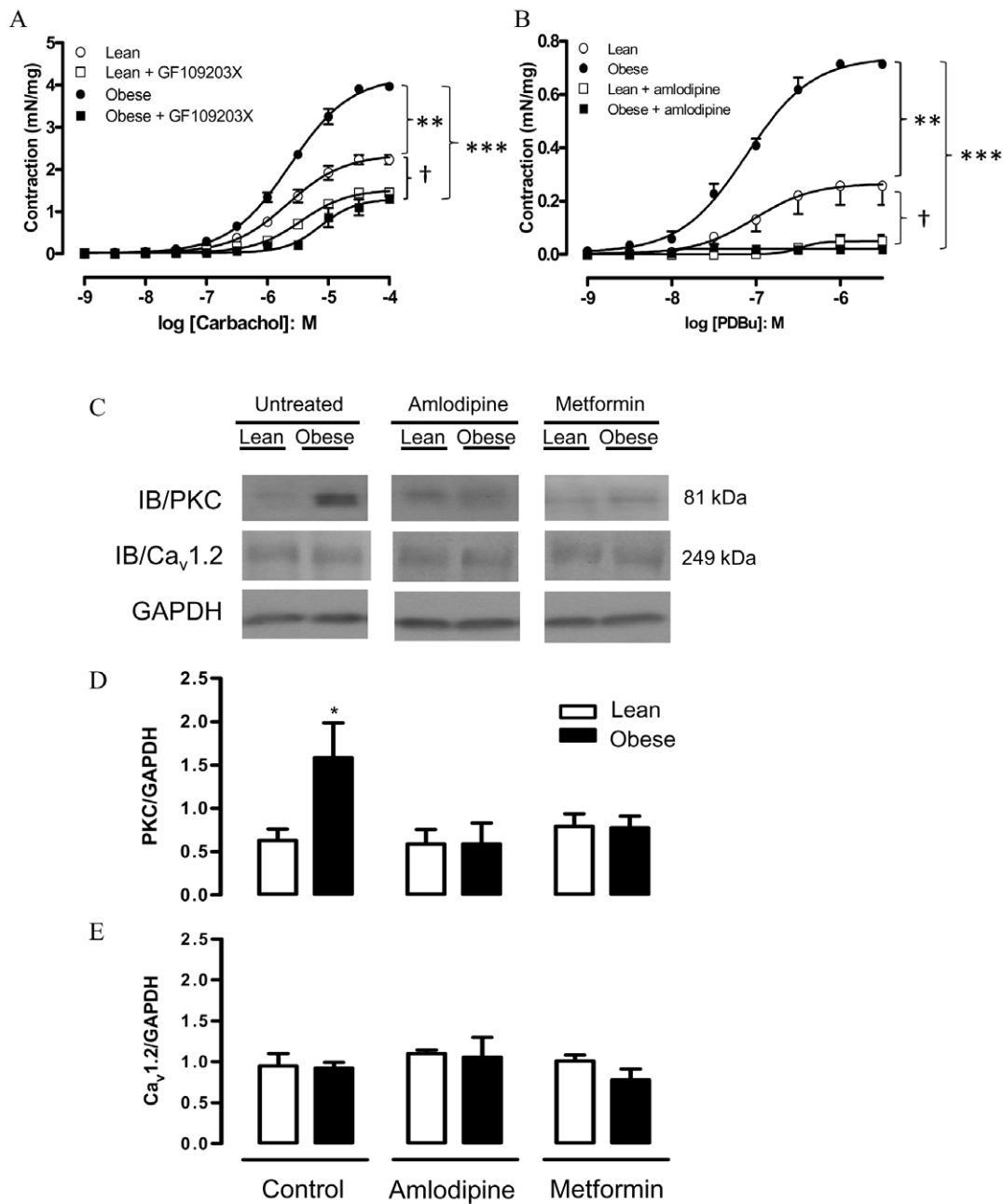
The signaling cascade for  $M_3$  receptors in smooth muscle also involves generation of DAG leading to PKC activation and inhibition of myosin light-chain (MLC) phosphatase, thus enhancing the contractile response [43]. Dysfunctions of the DAG–PKC pathway have been associated with vascular abnormalities in diabetic and insulin resistant states [44]. In the present study, we have performed concentration-response curves to carbachol in the presence of the PKC inhibitor GF109203X, and found that enhanced DSM contractions in the strips taken from obese mice were nearly normalized by this inhibitor. Additionally, the PKC activator PDBu produced concentration-dependent DSM contractions that were markedly greater in the strips taken from obese mice, an effect nearly abolished by amlodipine. This is consistent with the higher PKC protein expression in the bladder tissues of obese mice. Our data that the  $\alpha 1$  subunit of  $\text{Ca}_v1.2$  channel expression remained unchanged between groups together with functional data suggest that overactive DSM in obese mice takes place through elevated  $\text{Ca}_v1.2$  channel activity, increasing the extracellular  $\text{Ca}^{2+}$  influx. However, the lack of a direct measurement of  $\text{Ca}_v1.2$  activity or extracellular  $\text{Ca}^{2+}$  influx by electrophysiological studies in the bladder limits our comprehension about the observed phenomenon. We cannot ascertain, for instance, if changes in each of the component of the  $M_3$ /PKC/ $\text{Ca}_v1.2$  axis act independently or as a signaling pathway to determine the overactive DSM.  $\text{Ca}_v1.2$  channels have been suggested to serve as an anchoring target for PKC during translocation of this protein to the cell membrane after agonist stimulation [20]. The increased carbachol-induced bladder contractions observed under *in vitro* hyperglycemic conditions has also been associated to rho-kinase-mediated increased PKC activity [45]. Interestingly, in our study, amlodipine normalized the PKC protein levels in bladder tissue of obese mice. One may speculate that inactivation of  $\text{Ca}_v1.2$  channels results in downregulation of PKC expression, which is consistent with a previous study showing that amlodipine decreases the phosphorylation and activity of PKC in cultured human endothelial cells stimulated with the PKC activator PMA [46].

Metformin is a first-line pharmacological treatment for patients with type II diabetes mellitus because of its favorable overall

**Table 2. Values of cystometric parameters evaluated in lean and obese mice treated or not with amlodipine (25 mg/kg/day, 21 days) or metformin (300 mg/kg/day, 14 days).**

	Lean mice			Obese mice		
	Untreated	Amlodipine	Metformin	Untreated	Amlodipine	Metformin
Capacity (ml)	0.13 $\pm$ 0.01	0.15 $\pm$ 0.02	0.19 $\pm$ 0.04	0.13 $\pm$ 0.02	0.13 $\pm$ 0.03	0.16 $\pm$ 0.02
Threshold pressure (mmHg)	3.6 $\pm$ 0.7	5.3 $\pm$ 0.7	4.2 $\pm$ 0.3	4.3 $\pm$ 0.9	4.0 $\pm$ 0.6	5.4 $\pm$ 0.9
Compliance (mmHg/mL)	0.03 $\pm$ 0.004	0.03 $\pm$ 0.007	0.04 $\pm$ 0.009	0.03 $\pm$ 0.009	0.03 $\pm$ 0.007	0.036 $\pm$ 0.011
Peak pressure (mmHg)	12.2 $\pm$ 0.82	9.3 $\pm$ 1.02	13.9 $\pm$ 1.44	12.2 $\pm$ 2.3	9.9 $\pm$ 0.88	16.2 $\pm$ 1.9

Results are expressed as means  $\pm$  SEM from 4–7 animals in each group. doi:10.1371/journal.pone.0048507.t002



**Figure 7. Contractile responses to carbachol (A) and phorbol-12,13-dibutyrate (PDBu; B) in detrusor smooth muscle from lean and obese mice.** Responses to carbachol were performed in the presence of the PKC inhibitor GF109203X (1  $\mu$ M, 30 min), whereas responses to PDBu were performed in the presence of amlodipine (3  $\mu$ M). Protein expression (Western blotting) for PKC and  $\alpha$ 1 subunit of Ca<sub>v</sub>1.2 calcium channel in bladder tissues from lean and obese mice, treated or not with either amlodipine (3  $\mu$ M, 30 min) or metformin (300 mg/kg/day, 14 days), is shown in panels C, D and E. Data represent the mean  $\pm$  SEM for 4 to 7 mice each group. \*  $p < 0.05$ , \*\*  $p < 0.01$  compared with untreated lean group; \*\*\*  $p < 0.001$ , †  $p < 0.05$  compared with respective amlodipine groups.  
doi:10.1371/journal.pone.0048507.g007

profile, including its glucose-lowering ability, weight-neutral effects, and low risk of hypoglycemia. In our study, oral treatment of obese mice with metformin normalized the insulin resistance, as well as the *in vitro* and the *in vivo* (cystometry) bladder dysfunction. Moreover, metformin reversed the enhanced PKC levels in the bladder tissues from obese mice. Accordingly, a previous study has shown that PKC- $\theta$  knockout mice are protected from fat-induced insulin resistance in skeletal muscle [47]. Our findings that *in vitro* incubation of DSM strips with metformin does not alter the

overactive DSM indicate that normalization of bladder function resultant from the long-term treatment with this anti-hyperglycemic agent is secondary to the improvement of insulin sensitivity, rather than through a direct effect on the contractile machinery. Atypical and conventional PKC isoforms are classically related to mechanisms that contribute to insulin resistance in peripheral tissues responsible for glucose disposal and energy homeostasis [48]. Our present results evidence that PKC activation induced by high fat feeding plays a role in a very specific complication

associated with insulin resistance. Thus, our data suggest that PKC activity plays an important pathological role that is beyond the impairment glucose homeostasis control. In agreement with our findings, endothelial insulin resistance and vascular hyperactivity in high fat diet fed mice was recently described to be ameliorated by PKC inhibition [44,49], suggesting that the mechanism presently described is not specific for the bladder.

Obese mice treated with metformin remained overweight, suggesting that insulin resistance and subsequent hyperglycemia, rather than obesity, is the main cause of bladder overactivity. Although metformin reduces body weight and waist circumference in humans [50], no such effect has been reported in animals treated chronically with this agent [51]. Interestingly, in bladder dysfunction in female obese Zucker rats, it was the presence of

chronic obesity rather than the onset of diabetes that led to impaired bladder function [13].

In summary, the present study shows that the contractile and urodynamic alterations as well as the enhanced PKC expression in bladder tissues seen in high-fat fed obese mice are normalized by blockade of Ca<sub>v</sub>1.2 and improvement of insulin sensitivity. Up-regulation of PKC in obese mice is likely to simultaneously mediate the insulin resistance and hence overactive bladder.

## Author Contributions

Conceived and designed the experiments: LOL FZM GFA ADG EA. Performed the experiments: LOL CS MCC LL GFA. Analyzed the data: LOL FZM GFA ADG GDN EA. Contributed reagents/materials/analysis tools: GFA GDN AZ EA. Wrote the paper: LOL FZM GFA GDN AZ ADG EA.

## References

- Grundy SM, Brewer HB Jr, Cleeman JI, Smith SC Jr, Lenfant C (2004) American Heart Association; National Heart, Lung, and Blood Institute. Definition of metabolic syndrome: Report of the National Heart, Lung, and Blood Institute/American Heart Association conference on scientific issues related to definition. *Circulation* 109: 433–438.
- Opie LH (2007) Metabolic syndrome. *Circulation* 115: 32–35.
- Kahn BB, Flier JS (2000) Obesity and insulin resistance. *J Clin Invest* 106: 473–481.
- Carr DB, Utzschneider KM, Hull RL, Kodama K, Retzlaff BM, et al. (2004) Intra-abdominal fat is a major determinant of the national Cholesterol Education Program Adult Treatment Panel III Criteria for the Metabolic Syndrome. *Diabetes* 53: 2087–2094.
- Barton M, Baretella O, Meyer MR (2012) Obesity and Risk of Vascular Disease: Importance of Endothelium-Derived Vasoconstriction. *Br J Pharmacol* 165: 591–605.
- Moul S, McVary KT (2010) Lower urinary tract symptoms, obesity and the metabolic syndrome. *Curr Opin Urol* 20: 7–12.
- Kirby MG, Wagg A, Cardozo L, Chapple C, Castro-Diaz D, et al (2010) Overactive bladder: Is there a link to the metabolic syndrome in men? *NeuroUrol Urodyn* 29: 1360–1364.
- Laven BA, Orsini N, Andersson SO, Johansson JE, Gerber GS, et al. (2008) Birth weight, abdominal obesity and the risk of lower urinary tract symptoms in a population based study of Swedish men. *J Urol* 179: 1891–1895.
- Richter HE, Kenton K, Huang L, Nygaard I, Kraus S, et al. (2010) The impact of obesity on urinary incontinence symptoms, severity, urodynamic characteristics and quality of life. *J Urol* 183: 622–628.
- Rohrmann S, Smit E, Giovannucci E, Platz EA (2005) Association between markers of the metabolic syndrome and lower urinary tract symptoms in the Third National Health and Nutritional Examination Survey (NHANES III). *Int J Obes (Lond)* 29: 310–316.
- Tong YC and Cheng JT (2007) Alterations of M<sub>2,3</sub>-muscarinic receptor protein and mRNA expression in the bladder of the fructose fed obese rat. *J Urol* 178: 1537–1542.
- Lee W, Chien C, Yu H, Lee S (2008) Bladder dysfunction in rats with metabolic syndrome induced by long-term fructose feeding. *J Urol* 179: 2470–2476.
- Gasbarro G, Lin DL, Vurbic D, Quisno A, Kinley B, et al. (2010) Voiding function in obese and type 2 diabetic female rats. *Am J Physiol Renal Physiol* 298: F72–77.
- Galgani J, Ravussin E (2008) Energy metabolism, fuel selection and body weight regulation. *Int J Obes (Lond)* 32: S109–119.
- Rahman NU, Phonsombat S, Bochinski D, Carrion ED, Nunes L, et al. (2007) An animal model to study lower urinary tract and erectile dysfunction: the hyperlipidaemic rat. *BJU Int* 100: 658–663.
- Turban S, Hajdich E (2011) Protein kinase C isoforms: mediators of reactive lipid metabolites in the development of insulin resistance. *FEBS Lett* 585: 269–274.
- Yu CL, Chen Y, Cline GW, Zhang DY, Zong HH, et al. (2002) Mechanism by which fatty acids inhibit insulin activation of insulin receptor substrate-1 (IRS-1)-associated phosphatidylinositol 3-kinase activity in muscle. *J Biol Chem* 277: 50230–50236.
- Chibalin AV, Leng Y, Vieira E, Krook A, Bjornholm M, et al. (2008) Downregulation of diacylglycerol kinase delta contributes to hyperglycemia-induced insulin resistance. *Cell* 132: 375–386.
- Itani SI, Ruderman NB, Schmieder F, Boden G (2002) Lipid-induced insulin resistance in human muscle is associated with changes in diacylglycerol, protein kinase C, and I kappa B-alpha. *Diabetes* 51: 2005–2011.
- Huster M, Frei E, Hofmann F, Wegener JW (2010) A complex of Ca<sub>v</sub>(V)1.2/PKC is involved in muscarinic signaling in smooth muscle. *FASEB J* 24: 2651–2659.
- Leiria L, Mónica FZ, Carvalho F, Claudino MA, Franco-Penteado C, et al. (2011) Functional, morphological and molecular characterization of bladder dysfunction in streptozotocin-induced diabetic mice: Evidence of a role for L-type voltage-operated Ca<sup>2+</sup> channels. *Br J Pharmacol* 163: 1276–1288.
- Tsukumo DM, Carvalho-Filho MA, Carvalheira JB, Prada PO, Hirabara SM, et al. (2007) Loss-of-function mutation in Toll-like receptor 4 prevents diet-induced obesity and insulin resistance. *Diabetes* 56: 1986–1998.
- Calixto MC, Lintomen L, Schenka A, Saad MJ, Zanesco A, et al. (2010). Obesity enhances eosinophilic inflammation in a murine model of allergic asthma. *Br J Pharmacol* 159: 617–625.
- De Oliveira CF, Nathan LP, Metzke K, Moreno H Jr, De Luca IM, et al. (1999) Effect of Ca<sup>2+</sup> channel blockers on arterial hypertension and heart ischaemic lesions induced by chronic blockade of nitric oxide in the rat. *Eur J Pharmacol* 373: 195–200.
- Shore SA, Williams ES, Zhu M (2008) No effect of metformin on the innate airway hyperresponsiveness and increased responses to ozone observed in obese mice. *J Appl Physiol* 105: 1127–33.
- Lagaud GJ, Randriamboavonjy V, Roul G, Stoclet JC, Andriantsitohaina R (1999) Mechanism of Ca<sup>2+</sup> release and entry during contraction elicited by norepinephrine in rat resistance arteries. *Am J Physiol* 276: H300–308.
- Anhê GF, Hirabara SM, Turrer TC, Capero LC, Anhê FF, et al. (2007). Postpartum glycemic homeostasis in early lactating rats is accompanied by transient and specific increase of soleus insulin response through IRS2/AKT pathway. *Am J Physiol Regul Integr Comp Physiol* 292: R2225–2233.
- Hanley AJG, Wagenknecht LE, D'Agostino RB Jr, Zinman B, Haffner SM (2003) Identification of Subjects with Insulin Resistance and  $\beta$ -Cell Dysfunction Using Alternative Definitions of the Metabolic Syndrome. *Diabetes* 52: 2740–2747.
- Richter HE, Burgio KL, Brubaker L, Moalli PA, Markland AD, et al. (2005) Urinary Incontinence Treatment Network. Factors associated with incontinence frequency in a surgical cohort of stress incontinent women. *Am J Obstet Gynecol* 193: 2088–2093.
- Daneshgari F, Liu G, Birder L, Hanna-Mitchell AT, Chacko S (2009) Diabetic bladder dysfunction: current translational knowledge. *J Urol* 182: S18–26.
- Parsons JK, Sarma AV, McVary K, Wei JT (2009) Obesity and benign prostatic hyperplasia: clinical connections, emerging etiological paradigms and future directions. *J Urol* 182: S27–S31.
- Vikram A, Jena G, Ramarao P (2010) Pioglitazone attenuates prostatic enlargement in diet-induced insulin-resistant rats by altering lipid distribution and hyperinsulinaemia. *Br J Pharmacol* 161: 1708–1721.
- Michel MC, Barendrecht MM (2008) Physiological and pathological regulation of the autonomic control of urinary bladder contractility. *Pharmacol Ther* 117: 297–312.
- Abrams P, Andersson KE, Buccafusco JJ, Chapple C, de Groat WC, et al. (2006) Muscarinic receptors: their distribution and function in body systems, and the implications for treating overactive bladder. *Br J Pharmacol* 148: 565–578.
- Ford AP, Cockayne DA (2011) ATP and P2X purinoceptors in urinary tract disorders. *Handb Exp Pharmacol* 202: 485–526.
- Pinna C, Sanvito P, Puglisi L (2006) Altered neurogenic and mechanical responses to acetylcholine, ATP and substance P in detrusor from rat with outlet obstruction. *Life Sci* 79: 1301–1306.
- Kennedy C, Tasker PN, Gallacher G, Westfall TD (2007) Identification of atropine and P2X1 receptor antagonist-resistant, neurogenic contractions of the urinary bladder. *J Neurosci* 27: 845–851.
- Uchida W, Masuda N, Shirai Y, Shibasaki K, Satoh N, et al. (1994) The role of extracellular Ca<sup>2+</sup> in carbachol-induced tonic contraction of the pig detrusor smooth muscle. *Naunyn Schmiedeberg's Arch Pharmacol* 350: 398–402.
- Masters JG, Neal DE, Gillespie JI (1999) The contribution of intracellular Ca<sup>2+</sup> release to contraction in human bladder smooth muscle. *Br J Pharmacol* 127: 996–1002.
- Wegener JW, Schulla V, Lee TS, Koller A, Feil S, et al. (2004) An essential role of Cav1.2 L-type calcium channel for urinary bladder function. *FASEB J* 18: 1159–1161.

41. Wuest M, Hiller N, Braeter M, Hakenberg OW, Wirth MP, et al. (2007) Contribution of  $Ca^{2+}$  influx to carbachol-induced detrusor contraction is different in human urinary bladder compared to pig and mouse. *Eur J Pharmacol* 565: 180–189.
42. Frazier EP, Peters SLM, Braverman AS, Ruggieri MR, Michel MC (2008) Signal transduction underlying the control of urinary bladder smooth muscle tone by muscarinic receptors and  $\beta$ -adrenoceptors. *Naunyn-Schmiedeberg's Arch Pharmacol* 377: 449–462.
43. Andersson KE, Arner A (2004) Urinary bladder contraction and relaxation: physiology and pathophysiology. *Physiol Rev* 84: 935–986.
44. Naruse K, Rask-Madsen C, Takahara N, Ha SW, Suzuma K, et al. (2006) Activation of vascular protein kinase C- $\beta$  inhibits Akt-dependent endothelial nitric oxide synthase function in obesity-associated insulin resistance. *Diabetes* 55: 691–698.
45. Nobe K, Yamazaki T, Tsumita N, Hashimoto T, Honda K (2009) Glucose-dependent enhancement of diabetic bladder contraction is associated with a rho kinase-regulated protein kinase C pathway. *J Pharmacol Exp Ther* 328: 940–50.
46. Lenasi H, Kohlstedt K, Fichtschere B, Mülsch A, Busse R, et al. (2003) Amlodipine activates the endothelial nitric oxide synthase by altering phosphorylation on Ser1177 and Thr495. *Cardiovasc Res* 59: 844–853.
47. Kim JK, Fillmore JJ, Sunshine MJ, Albrecht B, Higashimori T, et al. (2004) PKC- $\theta$  knockout mice are protected from fat-induced insulin resistance. *J Clin Invest* 114: 823–827.
48. Stretton C, Evans A, Hundal HS (2010) Cellular depletion of atypical PKC $\{\lambda\}$  is associated with enhanced insulin sensitivity and glucose uptake in L6 rat skeletal muscle cells. *Am J Physiol Endocrinol Metab*. 299: E402–412.
49. Lu X, Bean JS, Kassab GS, Rekhter MD (2011) Protein kinase C inhibition ameliorates functional endothelial insulin resistance and vascular smooth muscle cell hypersensitivity to insulin in diabetic hypertensive rats. *Cardiovasc Diabetol* 10: 48–59.
50. Lee M, Aronne LJ (2007) Weight management for type 2 diabetes mellitus: global cardiovascular risk reduction. *Am J Cardiol* 99: 68B–79B.
51. Sena CM, Matafome P, Louro T, Nunes E, Fernandes R, et al. (2011) Metformin restores endothelial function in aorta of diabetic rats. *Br J Pharmacol* 163: 424–437.