



## REVIEW ARTICLE

# Importance of Bmal1 in Alzheimer's disease and associated aging-related diseases: Mechanisms and interventions

Rongping Fan<sup>1,2</sup> | Xuemin Peng<sup>1,2</sup> | Lei Xie<sup>1,2</sup> | Kun Dong<sup>1,2</sup> | Delin Ma<sup>1,2</sup> | Weijie Xu<sup>1,2</sup> | Xiaoli Shi<sup>1,2</sup> | Shujun Zhang<sup>1,2</sup> | Juan Chen<sup>3</sup> | Xuefeng Yu<sup>1,2</sup> | Yan Yang<sup>1,2</sup>

<sup>1</sup>Department of Endocrinology, Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, China

<sup>2</sup>Branch of National Clinical Research Center for Metabolic Diseases, Wuhan, China

<sup>3</sup>Department of Neurosurgery, Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, China

## Correspondence

Yan Yang, Department of Endocrinology, Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan 430030, China.  
Email: [yangyan6910@163.com](mailto:yangyan6910@163.com)

## Funding information

Jie Chu Jing Ying foundation, Grant/Award Number: 2018076; National Nature Science Foundation of China, Grant/Award Number: 81974114

## Abstract

With the aging world population, the prevalence of aging-related disorders is on the rise. Diseases such as Alzheimer's, type 2 diabetes mellitus (T2DM), Parkinson's, atherosclerosis, hypertension, and osteoarthritis are age-related, and most of these diseases are comorbidities or risk factors for AD; however, our understandings of molecular events that regulate the occurrence of these diseases are still not fully understood. Brain and muscle Arnt-like protein-1 (Bmal1) is an irreplaceable clock gene that governs multiple important physiological processes. Continuous research of Bmal1 in AD and associated aging-related diseases is ongoing, and this review picks relevant studies on a detailed account of its role and mechanisms in these diseases. Oxidative stress and inflammation turned out to be common mechanisms by which Bmal1 deficiency promotes AD and associated aging-related diseases, and other Bmal1-dependent mechanisms remain to be identified. Promising therapeutic strategies involved in the regulation of Bmal1 are provided, including melatonin, natural compounds, metformin, d-Ser2-oxyntomodulin, and other interventions, such as exercise, time-restricted feeding, and adiponectin. The establishment of the signaling pathway network for Bmal1 in aging-related diseases will lead to advances in the comprehension of the molecular and cellular mechanisms, shedding light on novel treatments for aging-related diseases and promoting aging-associated brain health.

**Abbreviations:** AD, Alzheimer's disease; ADAMTS, a disintegrin and metalloproteinase with thrombospondin motif; AMPK, adenosine 5'-monophosphate (AMP)-activated protein kinase; AS, atherosclerosis; BBB, blood-brain barrier; bHLH-PAS, helix-loop-helix/Per-ARNT-SIM; BMAL1, brain and muscle Arnt-like protein-1; BMP, bone morphogenetic protein; CA, cornu ammonis; cGAS-STING, cyclic GMP-AMP synthase (cGAS) and stimulator of interferon genes; CLOCK, circadian locomotor output cycles kaput; CRY, cryptochrome; EndMT, endothelial-to-mesenchymal transition; ER, endoplasmic reticulum; ERK, extracellular signal-regulated kinase; FoxOs, forkhead box O; GABA, gamma-aminobutyric acid; GLP-1R/GCGR, glucagon-like peptide-1 receptor/glucagon receptor; GSIS, glucose-stimulated insulin secretion; GST, glutathione transferase; GTP, guanosine triphosphate; HAECs, human aortic endothelial cells; HIF1 $\alpha$ , hypoxia-inducible factor 1 $\alpha$ ; IAPP, islet amyloid polypeptide; iBRB, inner blood-retina barrier; IL, interleukin; MAPK, mitogen-activated protein kinases; MMPs, matrix metalloproteinases; MuSC, muscle stem cell; NAD (+), nicotinamide adenine dinucleotide oxidase; NAMPT, nicotinamide phosphoribosyltransferase; NFATC, nuclear factor of activated T cells; NF- $\kappa$ B, nuclear factor-kappaB; NMN, nicotinamide mononucleotide; NR, nicotinamide riboside; NRF2, nuclear factor erythroid 2-related factor 2; OA, osteoarthritis; Oxy, D-Ser2-oxyntomodulin; PD, Parkinson's disease; PER, period; PGC-1 $\alpha$ , peroxisome proliferator-activated receptor-gamma coactivator-1 $\alpha$ ; PI3K/AKT/GSK3 $\beta$ , phosphatidylinositol-3-kinase/protein Kinase B/glycogen synthase kinase-3 $\beta$ ; P. gingivalis, Porphyromonas gingivalis; REV-ERB, reverse erythroblastosis virus; RPE, retinal pigment epithelium; ROR $\alpha$ , retinoid-related orphan receptor- $\alpha$ ; ROS, reactive oxygen species; SCN, suprachiasmatic nucleus; SIRT1, Sirtuin1; SMAD, Drosophila similar to mothers against decapentaplegic; STAT, signal transducer and activator of transcription; TGF- $\beta$ , transforming growth factor-beta; TMJ-OA, temporomandibular joint osteoarthritis; TRF, time-restricted feeding; TTFL, transcriptional-translational feedback loop; T2DM, type 2 diabetes mellitus; VEGF, vascular endothelial-derived growth factor; WAT, white adipose tissue.

This is an open access article under the terms of the [Creative Commons Attribution](https://creativecommons.org/licenses/by/4.0/) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2022 The Authors. *Aging Cell* published by Anatomical Society and John Wiley & Sons Ltd.



## KEYWORDS

Alzheimer disease, ARNTL transcription factors, atherosclerosis, diabetes mellitus, type 2, osteoarthritis, Parkinson disease

## 1 | INTRODUCTION

Aging, a complex biological process, is characterized by decreased physiological function with increased age; often aging is associated with increased susceptibility to diseases, including type 2 diabetes mellitus (T2DM), cardiovascular and musculoskeletal diseases, and neurological diseases such as Alzheimer's disease (AD) and Parkinson's disease (PD) (Zhuang et al., 2019). AD is the foremost cause of impaired cognition in the elderly and has become the fifth leading cause of death worldwide (Cortes-Canteli & Iadecola, 2020). Studies have found that many aging-related diseases such as PD (Cibulka et al., 2022), diabetes mellitus, atherosclerosis (AS) (Xie, Shi, et al., 2020), hypertension (Shih et al., 2018), osteoarthritis (OA) (Innes & Sambamoorthi, 2020), and age-related macular degeneration (AMD) (Wen et al., 2021) are closely related to AD, which may be comorbidities or risk factors for AD. This important commonality is why AD and these diseases are explored in the present study. It is known that PD and AD are common neurologic diseases of old age (Fang et al., 2018). Diabetes mellitus, hypertension, and OA have been reported to be common comorbidities in AD (Kao et al., 2021; Wang, Wu, et al., 2018). AS (Xie, Shi, et al., 2020), and AMD (Wen et al., 2021) had an increased risk of their condition advancing to AD. According to the World Health Organization (WHO) estimates, by 2050, the proportion of the global population with age > 60 years will ascend to 22% (Mahjoob & Stochaj, 2021). Emerging evidence now suggests an increased trend of patients with AD and associated aging-related diseases. For example, in the United States, the number of patients aged  $\geq 65$  years with AD and related dementias is growing and is projected to reach 13.9 million by 2060 (Matthews et al., 2019). The number of adults aged  $\geq 65$  and 65–99 years with diabetes mellitus is expected to increase to 0.253 and 1.42 billion, respectively, over the next 23 years globally (Bellary et al., 2021); nearly half of these adults are expected with T2DM. These epidemiological studies indicate a serious threat to global health with reduced quality of life of older adults amidst increased risk of AD and associated aging-related diseases.

Brain and muscle Arnt-like protein-1 (Bmal1), one of the families of basic helix–loop–helix/Per-ARNT-SIM (bHLH-PAS) domain-containing transcription factors (Majumdar et al., 2017), is the core regulator of the circadian clock, driving rhythmic expression of circadian clock genes. Bmal1 is essential for regulating the circadian rhythms and maintaining the physiological functions of cells and organs. Evidence indicates that the deletion of *Bmal1* can accelerate aging. For example, symptoms of premature aging were observed in *Bmal1* deletion animal models, such as organ atrophy, sarcopenia, and cataract; these animals also had shorter lifespans (Kondratov et al., 2006). During natural aging, *Bmal1* expression attenuation was noticed in animal models (Duncan et al., 2013). These studies show

that the decreased expression of Bmal1 is closely associated with aging. Alterations of Bmal1 during aging, along with other systemic stimulators such as hormones and the changed microenvironment of tissues, may have differential impacts on the brain and peripheral tissues, promoting the progression of aging-related diseases. The underlying mechanisms of the possible role of Bmal1 dysfunction in AD and associated aging-related diseases remain unclear; increased understanding of effects of abnormal Bmal1 expression may give rise to the evolvement of new diagnostic and therapeutic approaches for better management of these diseases.

To provide an integrated picture of the role and mechanism of Bmal1 in AD and associated aging-related diseases, we performed a comprehensive search in PubMed and Google Scholar for relevant studies. Based on existing evidence from the *in vivo*, *in vitro*, and clinical studies, we tried to summarize Bmal1-dependent mechanisms in AD and associated aging-related diseases and pay attention to therapeutic interventions involved in the regulation of Bmal1.

## 2 | THE BIOLOGICAL FUNCTION OF Bmal1

*Bmal1*, also called MOP3, is a core driver of the circadian clock in mammals and is considered the only irreplaceable clock gene that regulates rhythmic behaviors (Bunger et al., 2000; Welz et al., 2019). The molecular mechanism, that drives nearly 24 h autonomous circadian oscillations, involves the transcriptional–translational feedback loop (TTFL). Here, Bmal1 and circadian locomotor output cycles kaput (CLOCK) heterodimers form the positive limb that binds to the E-box motifs and drives the expression of the period (PER1/2/3), cryptochrome (CRY1/2), reverse erythroblastosis virus  $\alpha$  (REV-ERB $\alpha$ ), and retinoid-related orphan receptor- $\alpha$  (ROR $\alpha$ ); subsequently, PER and CRY, these two proteins interact and form heterodimers in the cytoplasm, which then translocate to the nucleus to suppress the expression of the positive limb (Richards & Gumz, 2013). Further, REV-ERB $\alpha$  and ROR $\alpha$ , which form additional feedback loops, facilitate and restrain the expression of Bmal1, respectively (Peng et al., 2022). In mammals, the molecular clock based on circadian rhythmicity exists in nearly all fully differentiated cells (Reinke & Asher, 2019). The master clock, situated in the suprachiasmatic nucleus (SCN) of the hypothalamus, receives an immediate projection from the retina via light–dark cues from the environment (Reinke & Asher, 2019). The SCN synchronizes peripheral clocks located in the non-SCN brain areas and peripheral tissues such as muscle, liver, adipose tissue, pancreas, and gut through neural, endocrine, temperature, and behavioral signals (Stenvers et al., 2019). The circadian system regulates various physiological processes, including food-intake behavior, rest–activity cycle, sleep–wake cycle, and glucose

metabolism (Stenvers et al., 2019). Bmal1 is not only an important regulator of the circadian system but also involves in preserving redox homeostasis (Chhunchha et al., 2020; Xie, Tang, et al., 2020). Furthermore, Bmal1 plays a crucial role in regulating inflammatory responses (Liu et al., 2020), insulin sensitivity (Shi et al., 2013), and mitochondrial functions (E. Li, Li, et al., 2020). Bmal1 deletion abolishes 24h activity patterns (Ray et al., 2020), leading to circadian rhythm disorders and aging-related diseases, such as glycolipid metabolism disorders including T2DM (Marcheva et al., 2010), and neurodegenerative diseases (Musiek & Holtzman, 2016), as presented in Figure 1. Besides, Bmal1 alterations among patients with aging-related diseases are summarized in Table 1.

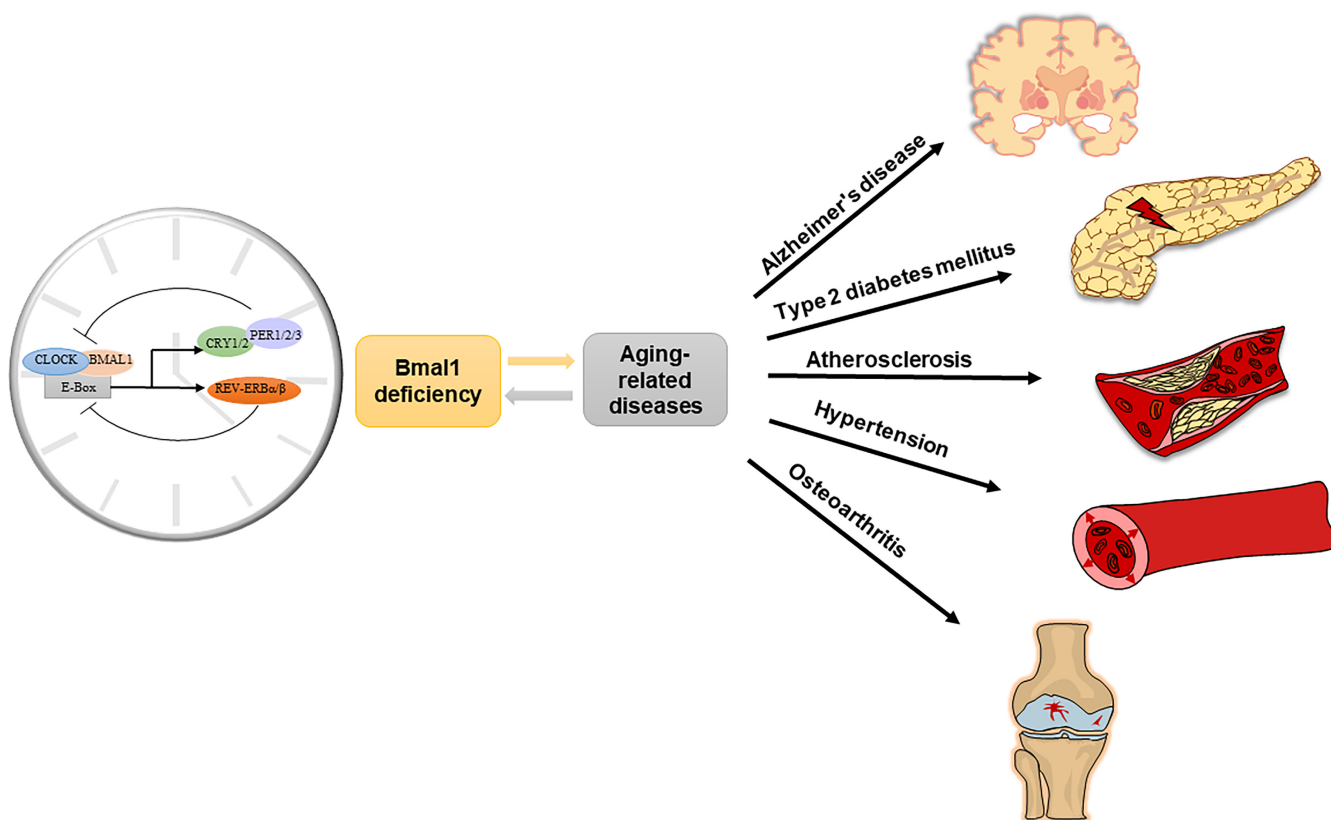
### 3 | THE ROLE AND MECHANISM OF Bmal1 IN AD

AD is the most prevalent aging-associated neurodegenerative disorder globally. It is characterized by the accumulation of extracellular  $\beta$ -amyloid ( $A\beta$ ) plaques and intracellular hyperphosphorylated tau protein, forming neurofibrillary tangles in the brain. These factors promote the progressive destruction of dendritic spines, synapses,

and loss of nerve cells, impaired neurotransmission, progressive isolation of remaining nerve cells, and eventually brain atrophy (Scheltens et al., 2016). The loss of the normal circadian rhythm is a common symptom of AD and is characterized by the increased sleep and awakening during the day and night, respectively (Mattis & Sehgal, 2016). AD-related circadian dysfunction has been widely explored by disrupting Bmal1 to understand the related mechanism, suggesting an important regulatory role of Bmal1 in AD. The contribution of Bmal1 deficiency to the neurodegeneration associated with AD and other aging-related dementia is depicted in Figure 2.

#### 3.1 | Abnormal expression of Bmal1 promoted $A\beta$ accumulation and hyperphosphorylation of tau protein

Many studies have shown that the Bmal1 function loss and  $A\beta$  deposition are mutually promoting scenarios.  $A\beta$  can incur Bmal1 degradation in AD mouse models (Song et al., 2015). Further, the aberrant expression of Bmal1 mRNA and protein in the mouse hippocampus can be induced by  $A\beta$  (Wang, Lv, et al., 2018), while it has also been reported that the Bmal1 loss can in turn accelerates the amyloid



**FIGURE 1** Bmal1 deficiency promotes Alzheimer's disease and associated aging-related diseases. The mammalian circadian clock exists in nearly all fully differentiated cells, which consists of a transcription-translation feedback loop (TTFL). The CLOCK-BMAL1 transcriptional activators bind to the E-box, driving the expression of CRY-PER and REV-ERBs transcriptional repressors. BMAL1 is in an irreplaceable position in TTFL, governing multiple important physiological processes. Bmal1 expression attenuation was observed during aging, and Bmal1 deficiency is an important contributor to aging-related diseases such as Alzheimer's disease, T2DM, atherosclerosis, hypertension, and osteoarthritis.



plaques accumulation (Kress et al., 2018). These studies indicated that the altered *Bmal1* levels might be the cause or a consequence of AD pathology. The effect of *Bmal1* deletion on A $\beta$  dynamics and amyloid plaque deposition depends on the scope of the deletion; the global deletion including the SCN generates an aberrant diurnal fluctuation of A $\beta$  in the hippocampal interstitial fluid and speeds up the amyloid plaque accumulation, whereas targeted *Bmal1* deletion in the hippocampus does not alter interstitial fluid A $\beta$  rhythms (Kress et al., 2018). Recently, it has also been found that the loss of *Bmal1* from astrocytes does not interpret the increased plaque burden in whole-brain *Bmal1*-knockout mice, suggesting that the status of astrocyte activation in AD brains has complex effects; further studies are needed to explore its non-A $\beta$ -associated contribution in AD (McKee et al., 2022). Interestingly, the inhibition of REV-ERBs can increase the transcription of *BMAL1* and enhance microglial A $\beta$  phagocytic activity in AD animal models, which may be a strategy to clear and reduce the amyloid plaque deposition (Lee et al., 2020). In addition, deficiency of *Bmal1* may lead to sleep disorders (Akladios et al., 2018; Qiu et al., 2019), preventing the removal of A $\beta$  and increasing the inflammatory cytokines and formation of A $\beta$  plaques (Ettcheto et al., 2019). Furthermore, *Bmal1* deletion induced pericyte dysfunction and decreased the integrity of the blood–brain barrier (BBB) in an age-dependent manner, which promoted the decreased cerebral blood flow rate and accumulation of blood-derived neurotoxins (Nakazato et al., 2017). Additionally, the link between *Bmal1* and tau protein has been confirmed in several studies. Disturbances in normal circadian rhythms also contribute to more disruptive daily oscillations of *Bmal1* and other clock genes and aggravate the hyperphosphorylation levels of soluble tau protein. These observations provide a link between *Bmal1* expression rhythms and AD pathology (Huang et al., 2021). Abnormal tau phosphorylation correlates with alterations in *Bmal1* protein levels (Niu et al., 2021), while in patients with AD, methylation of *Bmal1*, indicating *Bmal1* activity reduction, was correlated with tau pathology assessed by CERAD score, night wake, and alterations in cognitive function of some dimensions (Hulme et al., 2020).

### 3.2 | *Bmal1* deficiency aggravated neuropathology and synaptic degeneration

Astrocytes are the most abundant cell type in the central nervous system (CNS), having close structural and functional interactions with neurons and synapses (Dall rac et al., 2018). Further, reactive astrogliosis is a hallmark of AD (Reichenbach et al., 2019). Since it is difficult to select regulatory elements (to target *Bmal1* for deletion) in all the astrocytes in vivo using the available tools, the deletion of *Bmal1* in astrocytes in many studies refers to either the *Bmal1* deficiency in astrocytes specific to certain brain regions such as the SCN, or the astrocytes in the brain of *Bmal1* global knockout mice (Barca-Mayo et al., 2017). Mounting evidence shows that the influence of *Bmal1* deficiency on astrocytes is closely related to pathological factors that promote AD. In an in vitro study, *Bmal1*<sup>-/-</sup> astrocytes showed

a changed morphology (shorter actin filaments), lower expression of cortactin, and lower levels of Rho-GTP, impairing the actin cytoskeleton dynamics and formation of the distal astrocyte processes which led to an adverse effect on the synaptic integrity (Ali et al., 2020). In vivo, very fine astrocytic processes were lacking, and the synaptic coverage of astrocytes in CA3 was reduced in *Bmal1*<sup>-/-</sup> mice (Ali et al., 2020). These altered synaptic functions are related to cognitive deficits caused by chronodisruption. In addition, the deletion of *Bmal1* in astrocytes affects neurons; astrocytic *Bmal1* is essential to prevent the accumulation of extracellular gamma-aminobutyric acid (GABA) that mediates astrocyte-to-neuron communication (McKee et al., 2022). A study found that the administration of GABA receptor antagonists restored the cognitive functions of *Bmal1*cKO mice, demonstrating that the *Bmal1* deficiency in astrocytes may be associated with significant inhibition of learning and memory-related circuits through altered GABA levels (Barca-Mayo et al., 2017). Moreover, astrocyte-specific *Bmal1* deletion promotes neuronal death and astrogliosis and induces inflammatory gene expression in vitro (Lananna et al., 2018). Consistently, inflammatory gene expression and astrocyte activation are also induced in vivo (Lananna et al., 2018). Additionally, when *Bmal1* is abolished in both neurons and astrocytes, these mice show much more remarkable astrogliosis in the whole brain. This particular study also demonstrated that *Bmal1* regulates astrogliosis mainly via a cell-autonomous mechanism mediated by altered glutathione transferase-mediated protein glutathionylation (Lananna et al., 2018). Musiek et al. found that when *Bmal1* was partially knockdown in primary neurons, these cells exhibited spontaneous neurodegeneration, revealing another cellular mechanism by which diminished *Bmal1* expression contributed to neurodegenerative diseases (Musiek et al., 2013); further, *Bmal1* deletion contributes to the impaired structure and dysfunction of synapses, as well as damaged cortical functional connectivity in brain regions that are seriously influenced in animal models of brain aging and AD (Musiek et al., 2013).

### 3.3 | Signaling pathway for *Bmal1* deficiency in AD

Nicotinamide phosphoribosyltransferase (NAMPT) expression is controlled by the *BMAL1*/*CLOCK* complex (Ramsey et al., 2009); therefore, the inhibition of nicotinamide adenine dinucleotide (NAD<sup>+</sup>) biosynthesis is closely associated with *Bmal1* deficiency. Sirtuin1 (SIRT1) is an NAD<sup>+</sup>-dependent enzyme that regulates important physiological process, such as glucose metabolism, mitochondrial homeostasis (Bonkowski & Sinclair, 2016), and immune responses (Chang & Guarente, 2014). It has been reported that the expression of SIRT1 diminishes with the aging (Chen, Zhou, et al., 2020). The correlation of SIRT1 with the molecules participating in various signaling networks (NF- $\kappa$ B, P53, PGC1 $\alpha$ , and FoxOs) has been comprehensively reviewed in the literature (Chen, Zhou, et al., 2020; Gomes et al., 2018). SIRT1 is a positive regulator of *Bmal1* and *clock* genes in SCN (Chang & Guarente, 2013). The results of multiple evidence have indicated that the NAD<sup>+</sup>/SIRT1



TABLE 1 Bmal1 alterations among patients with aging-related diseases

First Author, Year	Participants	age	Cells or tissues	Bmal1 alterations
Hulme et al. (2020)	96 brains of donors who were participants of a large prospective cognitive aging cohort known as The University of Manchester Longitudinal Study of Cognition in Normal Healthy Old Age cohort (UMLCHA)	Age at death = $88.6 \pm 5.75$ years	Frontal cortex tissues	The positive correlation between DNA methylation at the BMAL1 gene with Braak stage and CERAD score stages, suggests a reduced activity of BMAL1 with increased AD pathology, specifically tau and neurofibrillary tangles
Chen, Peng et al. (2015)	AD patients ( $n = 296$ ) and control subjects ( $n = 423$ )	AD: $77.84 \pm 3.97$ years (range: 65 to 85 years) Control: $76.92 \pm 3.76$ years (range: 65 to 85 years)	Whole blood drawn from the antecubital vein	AD patients showed a higher prevalence of T carriers in BMAL1 rs.2278749 T/C (30.91 vs. 24.82%, $p < 0.0001$ ) than was observed in controls
Cermakian et al. (2011)	AD patients ( $N = 26$ , 13 females, 13 males) and controls ( $N = 30$ , 6 females, 24 males)	AD: $79.69 \pm 1.64$ Control: $74.43 \pm 2.16$ years	99 samples (29 BNST, 39 cingulate cortex, 31 pineal gland) from 56 donors (26 AD patients, 30 controls)	In AD patients, the phase of the 24-h harmonic of BMAL1 expression in the pineal and the phases of the 24-h rhythms of BMAL1 expression in the BNST were significantly different from controls
Yoo et al. (2020)	A total of 12 paraffin embedded brain tissues from 3 donors with AD. A total of 4 paraffin embedded adult normal brain tissues	NA	AD: frontal cortex, occipital cortex, temporal cortex and parietal cortex Control: frontal cortex, occipital cortex, temporal cortex and parietal cortex	The protein levels of BMAL1 were significantly elevated in impaired astrocytes of the cerebral cortex from patients with AD (every individual patient)
Ando et al. (2009)	Patients with type 2 diabetes ( $n = 12$ ) and healthy individual ( $n = 14$ )	T2DM: $58 \pm 6$ years Control: $59 \pm 6$ years	Peripheral leucocytes	T2DM patients expressed significantly lower transcript levels of BMAL1. BMAL1 mRNA levels were inversely correlated with HbA1c levels ( $r = -0.47$ , $p < 0.05$ )
Pappa et al. (2013)	Gestational diabetes mellitus (GDM) ( $n = 185$ ) and pregnant women with normal glucose tolerance ( $n = 161$ )	GDM: $33.63 \pm 5.14$ years Control: $30.71 \pm 9.56$ years	Peripheral blood leukocytes from 20 GDM and 20 control. Patients are selected according to their polymorphism of Bmal1 gene	Bmal1 is a crucial susceptibility gene for GDM in Greek women. The expression levels of BMAL1 mRNA were significantly lower in GDM patients than in controls
Gunton et al. (2005)	Type 2 diabetic subjects ( $n = 5$ ) and normoglycemic controls ( $n = 7$ )	Average age was 47 years in both groups. The mean duration of T2DM was $5.8 \pm 2.1$ years	Diabetic human pancreatic islets	Bmal1 expression was significantly downregulated
Yu et al. (2019)	T2D patients ( $n = 36$ ) and non-diabetic volunteers ( $n = 14$ )	T2D: $49.47 \pm 7.88$ years Control: $45.93 \pm 7.16$ years	Peripheral blood leucocytes	The BMAL1 mRNA levels were decreased in the diabetic patients. In addition, HbA1c levels were negatively correlated with BMAL1 mRNA levels

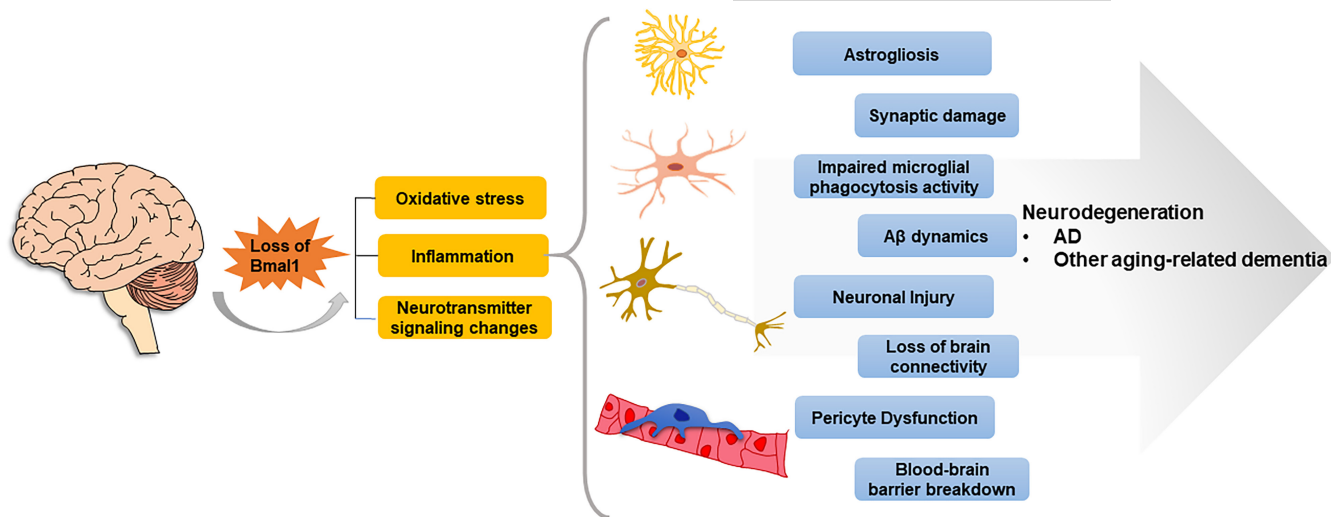
(Continues)



TABLE 1 (Continued)

First Author, Year	Participants	age	Cells or tissues	Bmal1 alterations
Cai et al. (2010)	PD patients (n = 17) and age-matched controls (n = 16)	PD: 62.2 ± 2.8 years Control: 57.8 ± 1.1 years	Peripheral leukocytes. Blood samples were collected during 21:00–09:00	Expression of BMAL1 was reduced in PD. The expression levels of BMAL1 in PD patients were correlated with their disease severity and sleep quality
Li et al. (2021)	PD patients (n = 326) and healthy controls (n = 314)	PD: 67.43 ± 9.71 years Control: 66.03 ± 9.24 years	Peripheral blood mononuclear cells (PBMCs)	BMAL1 expression was significantly decreased in PD. The severity of pRBD, the severity of daytime sleepiness, and the self-reported sleep quality were inversely associated with the expression levels of the BMAL1 gene
Breen et al. (2014)	Parkinson disease cohort (N = 239), subgroup of these patients (n = 30) and healthy age- and sex-matched controls (n = 15)	PD: age at diagnosis = 68 ± 9 years	Peripheral blood	PD patients had a lack of time-dependent variation in Bmal1 expression
Akagi et al. (2017)	OA patients (n = 14) and normal controls (n = 14)	OA: 10 females (mean age = 69 years, range: 61–82) and 6 males (mean age = 71 years, range: 66–84) Control: 5 females (mean age = 39 years range: 26–57) and 18 males (mean age = 30, range: 18–44)	Human knee cartilage	BMAL1 mRNA and protein levels were significantly reduced in OA compared with normal cartilage
Dudek et al. (2016)	Non-OA or mild OA samples (n = 6), grade 0/1; grade 2/3, moderate OA samples (n = 6); and grade 4, OA (n = 6)	Age range, 45–60 years	Human cartilage	The number of BMAL1-positive chondrocytes and the protein levels of BMAL1 were progressively reduced in OA cartilage with increasing severity compared with the numbers detected in non-OA human tissue
Wu et al. (2019)	Atherosclerosis patients (n = 31) and healthy controls (n = 15)	More than 40 years, from both sexes	Plasma from blood specimens	The expression of Bmal1 mRNA was decreased in the plasma of patients with atherosclerosis

Abbreviations: AD, Alzheimer's disease; BMAL1, brain and muscle Arnt-like protein-1; BNST, bed nucleus of the stria terminalis; CERAD, Consortium to Establish a Registry for Alzheimer's Disease; GDM, gestational diabetes mellitus; OA, osteoarthritis; PD, Parkinson's disease; pRBD, rapid eye movement sleep behavior disorder; T2D, type 2 diabetes.



**FIGURE 2** Loss of *Bmal1* contributes to the neurodegeneration associated with Alzheimer's disease and other aging-related dementia. The loss of *Bmal1* in the brain has adverse effects on the astrocytes, microglia cells, neurons, and pericyte cells, leading to an increased level of oxidative stress and inflammation, as well as neurotransmitter signaling changes. The above alterations may play a contributory role in astrogliosis, impaired microglial phagocytosis activity, neuronal injury, and pericyte dysfunction, further leading to synaptic damage,  $A\beta$  dynamics, loss of brain connectivity, and disruption of the blood–brain barrier, which constitutes the pathological basis of Alzheimer's disease and other aging-related dementias.

dysfunction contributes to AD (Julien et al., 2009; Koo et al., 2017; Lautrup et al., 2019). Mechanistic studies of the aging brain in PD also showed the involvement of the *BMAL1/SIRT1* pathway (Wang, Lv, et al., 2018). Furthermore,  $NAD^+$  depletion-mediated activation of cyclic GMP-AMP synthase and stimulator of interferon genes (cGAS-STING) have been reported to play an indispensable role in neuroinflammation and cellular senescence in AD (Hou, Wei, et al., 2021). However, the evidence of pathological progression of AD caused by  $NAD^+$  decline caused directly by *Bmal1* deficiency is not available, and further studies are needed to reveal the underlying mechanism. Thus, we describe this indirect link in light gray in Figure 3.

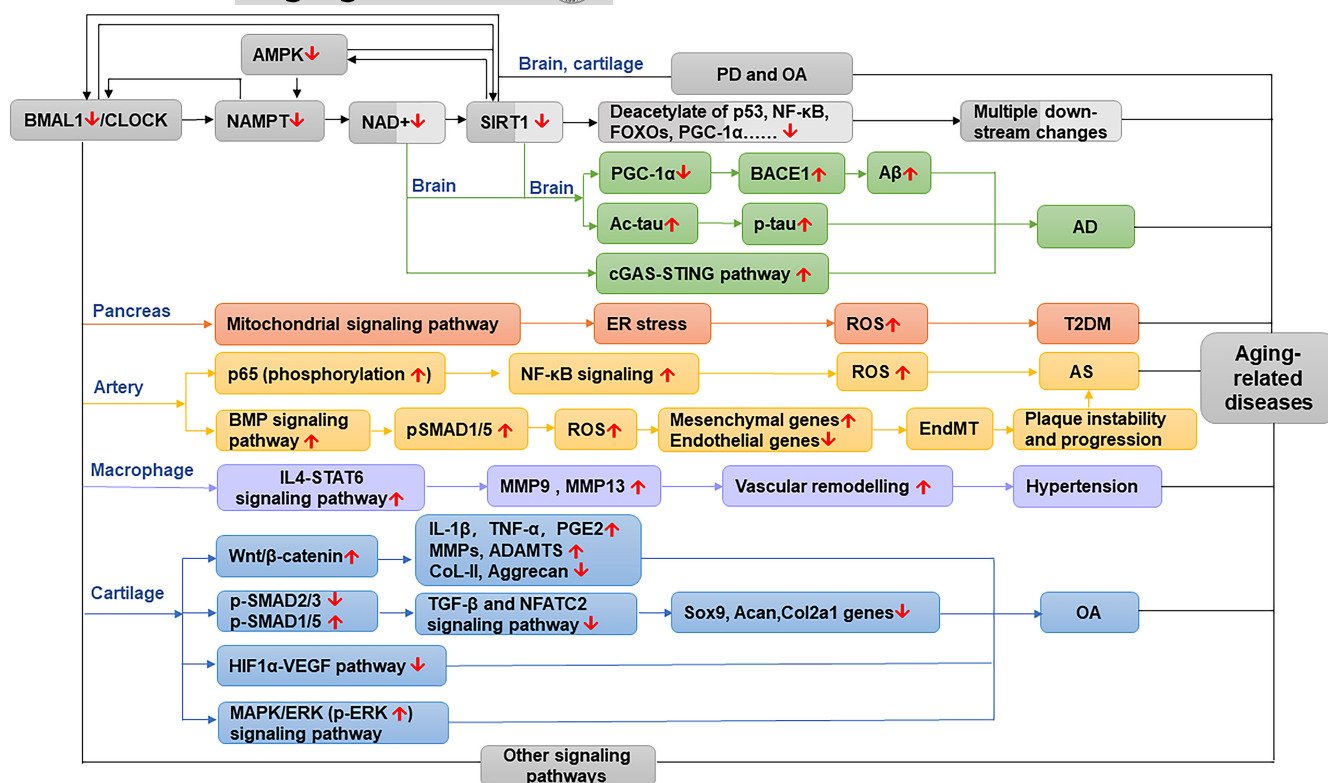
## 4 | THE ROLE AND MECHANISM OF *Bmal1* IN AD-ASSOCIATED AGING-RELATED DISEASES

### 4.1 | The role and mechanism of *Bmal1* in T2DM

T2DM is a prevalent chronic metabolic disease characterized by hyperglycemia, relative insulin deficiency, insulin resistance, and pancreatic beta-cell dysfunction (Yaribeygi et al., 2020). *Bmal1* appears to be highly relevant to the pancreatic islet's function, regulation of insulin secretion, and glucose metabolism. For instance, it was suggested that the expression of *Bmal1* in beta cells is a feature of postnatal islet cell maturation and contributes to the establishment of circadian control of glucose-stimulated insulin secretion (GSIS) (Rakshit et al., 2018). Mice adopt a high-fat diet undergo compensatory increased beta-cell mass to preserve normal beta-cell function and glucose homeostasis (Rakshit et al., 2016); however, this

response is diminished in mice with beta-cell-specific deletion of *Bmal1*; increased apoptosis was observed additionally, suggesting a regulating role of *Bmal1* in beta-cell compensatory proliferation and function (Rakshit et al., 2016). Furthermore, *Bmal1* in the hypothalamic paraventricular nucleus was significant in regulating insulin secretion and glucose tolerance by regulating the expression and release of vasopressin, a humoral factor that stimulates insulin secretion (Nakata et al., 2021).

Disruption of *Bmal1* in beta cells plays a notable role in the onset of diabetes. Marcheva et al. suggested that pancreas-specific *Bmal1* KO mice displayed reduced glucose tolerance and lessened pancreatic islets size, proliferation, and insulin release that worsened with age (Marcheva et al., 2010). Lee et al. found that deletion of *Bmal1* in beta cells results in cell dysfunction and diabetes; this was owing to the increased reactive oxygen species (ROS) accumulation, consequent mitochondrial uncoupling, and decreased antioxidant regulatory factor (Lee et al., 2018, 2013). Another study found that the loss of *Bmal1* can impair beta-cell function through the mitochondrial signaling pathway, as disruptive mitochondrial morphologies such as swelling, fracture, and disappearance of mitochondrial cristae can be observed in *Bmal1*-deficient beta cells (Ye et al., 2020). Importantly, beta-cell-specific *Bmal1* overexpression at the mRNA and protein levels enhances islet's circadian clock amplitude as well as GSIS, meanwhile, preventing obesity-induced glucose intolerance and reducing the cellular oxidative and endoplasmic reticulum (ER) stress (Rakshit & Matveyenko, 2021). These findings indicate that the downregulation of *Bmal1* may be related to T2DM by promoting mitochondrial dysfunction and oxidative stress in beta cells. Genetic variations in *Bmal1* have also been linked with susceptibility to T2DM in humans (Pappa et al., 2013; Woon et al., 2007). Consistently, patients



**FIGURE 3** Signaling pathways related to *Bmal1* deficiency in AD and associated aging-related diseases. Dark gray represents the direct link from *Bmal1* to  $\text{NAD}^+$  and SIRT1. Light gray represents the indirect link between *Bmal1* to  $\text{NAD}^+$  and SIRT1. Direct evidence suggested that the SIRT1-BMAL1 pathway was involved in PD and OA, and indirect evidence indicated that the SIRT1-BMAL1 pathway may involve in other aging-related diseases such as AD. *Bmal1* deficiency in the pancreas contributed to T2DM via accumulated ROS mediated by the mitochondrial signaling pathway. In the artery, ROS accumulation was aggravated by *Bmal1* deficiency through NF- $\kappa$ B signaling and BMP-mediated signaling, leading to the progression of AS. *Bmal1* loss in macrophages enhanced hypertensive vascular remodeling in hypertension through the IL4-STAT6 signaling pathway. *Bmal1* deficiency in cartilage exacerbated OA through interaction with multiple signaling pathways, which included the Wnt/ $\beta$ -catenin signaling pathway, TGF- $\beta$  and NFATC2 signaling pathway, HIF1 $\alpha$ -VEGF pathway, and MAPK/ERK signaling pathway. There may exist other potential *Bmal1*-dependent signaling pathways involved in aging-related diseases.

with T2DM and animal models of T2DM show disrupted *Bmal1* expression in several tissues. For example, downregulated *Bmal1* expression was found in diabetic human islets and peripheral blood leukocytes from T2DM patients (Gunton et al., 2005; Yu et al., 2019). Decreased *Bmal1* expression in T2DM rats was associated with suppressed bone marrow mesenchymal stem cell osteogenesis (Li et al., 2017). Dampened oscillations of *Bmal1* were found in diabetic mice aortas (Su et al., 2008). Decreased level of *Bmal1* protein was found in white adipose tissue (WAT) of db/db mice (Caton et al., 2011). Overall, the findings have revealed that *Bmal1* expression is impaired in T2DM.

## 4.2 | PD

PD is the second most common age-related neurodegenerative disorder and is manifested through the motor and non-motor symptoms (Wang, Lv, et al., 2018). Many clinical manifestations of PD suggest circadian rhythm dysfunction, such as disturbances of rest and activity cycles, aberrant blood pressure fluctuation patterns, and an abnormal rhythm of melatonin secretion (Wang,

Lv, et al., 2018). These observations indicate that disrupted circadian rhythm is one of the most common non-motor symptoms in PD. The clinicopathological features of PD are characterized by the dopaminergic neuronal loss in the substantia nigra pars compacta and the formation of Lewy bodies and neuritis (Gómez-Benito et al., 2020). Evidence in human studies and animal models showed that decreased level of *Bmal1* is involved in PD. Relatively lower *Bmal1* levels were found in leukocytes of PD patients, which may be a reflection of the severity of PD (Cai et al., 2010). *Bmal1* expression levels in the peripheral blood mononuclear cells and plasma melatonin levels were decreased in patients with PD (Li et al., 2021). Moreover, a lack of time-dependent variation in *Bmal1* expression was observed (Breen et al., 2014). One potential explanation for the alteration of *Bmal1* expression is that dopamine can regulate the activity of the BMAL1/CLOCK complex, and hence, a lack of dopamine in PD affects this central component of the molecular clock (Breen et al., 2014). After melatonin treatment, the levels of *Bmal1* were increased in PD patients (Delgado-Lara et al., 2020), reinforcing a close link between *Bmal1* and PD. In a rat model of PD, the expression of *Bmal1* in the striatum was significantly lower than that in the sham group at





some timepoints (Yang et al., 2021). Further, in another rat model of PD induced by lipopolysaccharide combined with rotenone, the mRNA and protein expressions of Bmal1 also decreased (Li et al., 2019).

Additionally, Bmal1 was shown to be involved in regulating PD pathogenesis. In the 1-methyl-4-phenyl-1,2,4,5-tetrahydropyridine-induced PD mouse model, inactivation of Bmal1 can lead to distinct motor dysfunction, dopaminergic neuronal injury in the substantia nigra pars compacta, deficiency in dopamine transmitters, and aggravation of the neuroinflammatory response, suggesting that Bmal1 may exert a beneficial effect on survival of dopaminergic neurons by regulating the microglia-mediated neuroinflammation response (Liu et al., 2020). The increase in oxidative stress (and loss of anti-oxidative defense) is critically related to the onset of PD (Johnson et al., 2012); treatment with 6-hydroxydopamine, in an animal model of PD, caused a reduction and elevation in the levels of SIRT1 and acetylated Bmal1, respectively, causing abnormal antioxidative activity. These findings suggest that the SIRT1-BMAL1 pathway is involved in the regulation of abnormal antioxidative responses in PD (Wang, Lv, et al., 2018).

#### 4.3 | Atherosclerosis

Several studies indicate a direct link between Bmal1 and atherosclerosis (AS). For instance, Bmal1 downregulation is involved in *Porphyromonas gingivalis* (*P. gingivalis*) induced atherosclerosis by exacerbating the development of oxidative stress (Xie, Tang, et al., 2020). In the indirect co-culture model of *P. gingivalis* and human aortic endothelial cells, Bmal1 upregulation inhibited monocyte recruitment, reduced the levels of pro-inflammatory cytokines, and decreased cell apoptosis, demonstrating the antagonistic role of Bmal1 in oxidative stress (Xie, Tang, et al., 2020). The loss of Bmal1 in macrophages could give rise to increased trafficking of Ly6chi monocytes to atherosclerotic lesions, which resulted in an increase in macrophage content and lesion size in the carotid arteries and promoted atherosclerosis (Huo et al., 2017). Bmal1 deficiency in the human carotid aggravated intracellular ROS accumulation and endothelial-to-mesenchymal transition through the bone morphogenetic protein-mediated signaling, demonstrating the central role of Bmal1 loss in atherosclerosis progression (Zhu et al., 2018). Liang et al. found that the upregulated expression of microRNA-155 (miR-155) was positively associated with the downregulation of Bmal1 in the aorta, leading to an increased atherosclerotic plaque area and weakened aortic diastolic function, and these changes were reversed after downregulation of miR-155 or an increase in Bmal1 (Liang et al., 2020). Pan et al. found that both the global and hepatic Bmal1 deficiency enhances atherosclerosis in Apoe<sup>-/-</sup> mice and demonstrated that Bmal1 is an anti-atherogenic transcription factor for its important contribution to lipoprotein and cholesterol metabolism (Pan et al., 2016). These studies supported that Bmal1 deficiency plays an important role in AS.

#### 4.4 | Hypertension

The clock gene Bmal1 is reported to be highly correlated with the etiology of hypertension. Analysis of single-nucleotide polymorphisms (SNPs) has elucidated that BMAL1 is associated with susceptibility to hypertension (Woon et al., 2007). In a genetic animal model of hypertension, the spontaneously hypertensive (SHR) rats, the expression of Bmal1 was decreased (Tharmalingam et al., 2020). A significant rhythm imbalance of the Bmal1 mRNA levels was found in SHR liver tissues (Hou, Zhang, et al., 2021). Hypertension in db/db mice is associated with attenuated Bmal1 oscillations in the vasculature (Su et al., 2008). Recently, Huo et al. suggested that BMAL1 deletion has a tonic effect on vascular remodeling, leading to the promoted blood pressure increase (Huo et al., 2021). They indicated that loss of Bmal1 enhanced the phosphorylation and nuclear translocation of STAT6 triggered by IL4, which promoted the activation of STAT6 and its target gene transcription, leading to increased expression of MMP9 and MMP13 in the vascular wall, contributing to vascular remodeling (Huo et al., 2021).

#### 4.5 | Osteoarthritis

Osteoarthritis (OA) is the most widespread degenerative joint disease and is characterized by cartilage degeneration in articulating joints, with a higher risk in older people. The OA disease affects >30 million people in the United States (Alliston et al., 2018). Bmal1 is essential for the development of hard tissues such as bones, cartilage, and teeth (Chen, Tang, et al., 2020). A growing body of studies suggest that *Bmal1* loss can inhibit osteogenesis and promote osteoclastogenesis, playing a significant role in the pathogenesis of OA. Akagi et al. found that Bmal1 is reduced in OA cartilage, and alterations in Bmal1 expression affect transforming growth factor-beta (TGF- $\beta$ ) downstream gene expression in chondrocytes, which may accelerate cartilage injury through inflammation-related pathways (Akagi et al., 2017). Consistently, Dudek et al. found that the expression of Bmal1 was reduced in the articular cartilage of human knees with OA (Dudek et al., 2016). In the *Bmal1*-cKO mouse model used in their study, targeted *Bmal1* ablation abolished circadian rhythm as well as resulted in progressive degeneration of articular cartilage, which may be related to the disturbance of cartilage homeostasis caused by Bmal1 dysregulation; reduced TGF- $\beta$  and NFATC2 pathway signaling was identified in *Bmal1*-cKO chondrocyte (Dudek et al., 2016). It is known that the increase in matrix metalloproteinases (MMPs) and a disintegrin and metalloproteinase with thrombospondin motif (ADAMTS) downstream induced by the extracellular signal-regulated kinase (ERK) lead to cartilage decomposition (Ma et al., 2014). Chen et al. observed that Bmal1 expression was decreased in rats with circadian rhythm disruption, along with increased P-ERK/MMPs/ADAMTS expression (Chen, Zhao, et al., 2020). IL-6-induced MMP3/13 and ADAMTS5 upregulation was retracted by Bmal1



overexpression in vitro; temporomandibular joint osteoarthritis (TMJ-OA) was reversed in *Bmal1*-overexpressing rats, suggesting that *Bmal1* is involved in the regulation of OA through the mitogen-activated protein kinase (MAPK)/ERK signaling pathway (Chen, Zhao, et al., 2020). Evidence in vivo and in vitro found that the deletion of *Bmal1* could lead to inhibited chondrocyte proliferation and elevated apoptosis, along with decreased expression of hypoxia-inducible factor 1 $\alpha$  (HIF1 $\alpha$ ) and vascular endothelial-derived growth factor (VEGF); consistently, the recovery of *Bmal1* expression improved the function of chondrocytes (Ma et al., 2019). Moreover, *Bmal1* takes part in the regulation of cartilage homeostasis through SIRT1, which is affected by the level of NAD<sup>+</sup> oxidase (Yang et al., 2016). *Bmal1* also participates in the regulation of matrix synthesis and catabolic metabolism in cartilage and chondrocytes through the Wnt/ $\beta$ -catenin signaling pathway (Song, Ma, et al., 2021). These findings suggest that *Bmal1* deletion of chondrocytes results in disruptive critical pathways that impair cartilage homeostasis and exacerbate a catabolic state, thus resulting in cartilage degradation.

#### 4.6 | Age-related macular degeneration

Age-related macular degeneration (AMD) is a leading cause of visual impairment and severe irreversible blindness and is classified as early-stage (medium-sized drusen and retinal pigmentary changes) to late-stage (neovascular and atrophic) in clinical settings (Mitchell et al., 2018). In patients older than 75 years, the risk of developing early AMD and late AMD is 25% and 8%, respectively, and the number of cases is expected to grow as the population ages (Stahl, 2020). By 2040, the global number of people suffering from AMD is expected to be nearly 300 million (Mitchell et al., 2018). The inner blood–retina barrier (iBRB) is exceedingly dynamic, and claudin-5, a tight junction protein, is abundantly expressed. Persistent suppression of claudin-5 induces remarkable retinal pigment epithelium (RPE) cell atrophy (Hudson et al., 2019). Hudson et al. found that disruption of *Bmal1* expression results in dysregulated claudin-5 cycling and adverse effects on the integrity of endothelial cells, suggesting that the *Bmal1* dysfunction may strongly implicate in drusen accumulation and subsequent RPE atrophy (Hudson et al., 2019). Therefore, it is necessary to explore a more comprehensive and in-depth mechanism of *Bmal1* in AMD.

## 5 | SUMMARY AND DISCUSSION OF THE ROLE AND MECHANISM OF *Bmal1* IN AD AND ASSOCIATED AGING-RELATED DISEASES

The above studies explored the role of *Bmal1* in AD and associated aging-related diseases, which suggested the dysfunction of *Bmal1* in many important organs: the brain, pancreas, blood vessels, and articular cartilage. Some ideas can be extracted from these studies.

First, oxidative stress may be the common trigger when *Bmal1* is lost in these organs, driving the aging-related loss of function in organs; and inflammation may be another important trigger. This idea was consistent with the main idea of another study discussing oxidative stress in aging and related chronic diseases (Mas-Bargues et al., 2022). Here are some issues worth considering. At the organ-tissue level, it would be interesting to know whether *Bmal1* loss in one organ affects other organs and whether the start of oxidative stress in one organ may be the trigger of oxidative stress for another organ, contributing to the organ-organ communication involved in the development of the aging-related diseases and cognition dysfunction. On a cellular level, it remains unclear whether *Bmal1* deficiency in different organs shares a common signal pathway before the initiation of the oxidative stress-related signal pathway.

Second, evidence showed that *Bmal1* was associated with other ischemia-related diseases such as myocardial infarction (Liu, Xiao, et al., 2021), stroke (Beker et al., 2018), and peripheral vascular disease (Xu et al., 2021), which are highly prevalent in aging populations (Hadjipanayi & Schilling, 2013); and some of these diseases have a close relationship with hypertension and atherosclerosis and may promote AD (Vijayan & Reddy, 2016; Zhang & Luo, 2020). There is an interesting link between vascular damage and *Bmal1* deficiency when putting these diseases together, suggesting that a mechanism related to vascular pathology may be a common trigger. Cumulative evidence has demonstrated that the lack of *Bmal1* would impair angiogenesis, and endothelial cell functions (Xu et al., 2021), accelerating microvascular and macrovascular damage (Lee et al., 2021). Endothelial dysfunction from the *Bmal1*-knockout mice was associated with enhanced superoxide levels and endothelial NO synthase uncoupling in blood vessels (Anea et al., 2012). In part, the mechanisms underlying these impairments may involve oxidative stress and that the *Bmal1* exerts on key pathways in endothelial cell signaling.

Third, most studies suggested that decreased level of *Bmal1* was a contributor to aging-related disease; there are also a few studies linking elevated *Bmal1* to certain diseases. For instance, the elevated level of *Bmal1* was also found in the pineal of streptozotocin-induced type 1 diabetes (Peschke et al., 2008), astrocytes of the cerebral cortex in AD (Yoo et al., 2020), tissue samples of follicular and papillary thyroid carcinoma (Mannic et al., 2013), indicating that *Bmal1* may exhibit a tissue-specific expression representing an additional regulation. This specific role in AD and associated aging-related disease and the potential mechanisms remain to be further clarified. Besides, in addition to contributing to the development of aging-related diseases, the deletion of *Bmal1* in specific tissue may offer new treatment strategies for some diseases. For example, microglia-specific knockdown of *Bmal1* improved memory in the animal model, and the authors proposed that microglial *Bmal1* may be a potential therapeutic target for metabolic and cognitive disorders related to psychiatric disease (Wang et al., 2021).

The present study identified important role and mechanism of *Bmal1* in AD and associated aging-related diseases such as PD, T2DM, hypertension, AS, OA, and AMD, suggesting that *Bmal1* deficiency was implicated in the etiology of these diseases. These

AD-associated diseases can also promote the initiation or progression of AD through numerous pathways, and the pathology of one of the diseases may be a risk factor for another. Thus, it is interesting to explore the role of Bmal1 in this relationship. Unfortunately, there was rare evidence supporting the role of Bmal1 in the contributing effects of AD-associated diseases on AD. For example, the deposition of islet amyloid polypeptide (IAPP) in the pancreas is one of the pathological hallmarks of T2DM. IAPP was detected in brain tissues and was associated with cognitive decline and AD development; its interaction with A $\beta$  contributed to a synergistic toxic activity (Zhang & Song, 2017). Besides, IAPP was a crucial mediator of tau pathology in AD (Zhang et al., 2022). However, we found no evidence about the role of Bmal1 in the above pathological links, which suggested that the role of Bmal1 in the mechanisms connecting these aging-related diseases to AD is poorly understood (Figure 4).

## 6 | THERAPEUTIC PERSPECTIVES

Therapeutics tailored to regulate molecular Bmal1 could be the key to preventing the progression of aging-related diseases. Several relevant treatments targeting Bmal1 in aging-related diseases are discussed in this section and summarized in Table 2. Oxidative stress is a consensus mechanism of aging, and the antioxidant N-acetyl-L-cysteine has been shown to ameliorate signs of premature aging related to Bmal1 deficiency (Kondratov et al., 2009). Antioxidants that boost nuclear factor erythroid 2-related factor 2 (NRF2) activity are available to rescue the excessive pro-oxidant and pro-inflammatory phenotype of *Bmal1*<sup>-/-</sup> macrophages (Early et al., 2018). Therefore, treatments involving antioxidative effects may have anti-aging effects, and we focused on those involved in the regulation of Bmal1.

### 6.1 | Melatonin

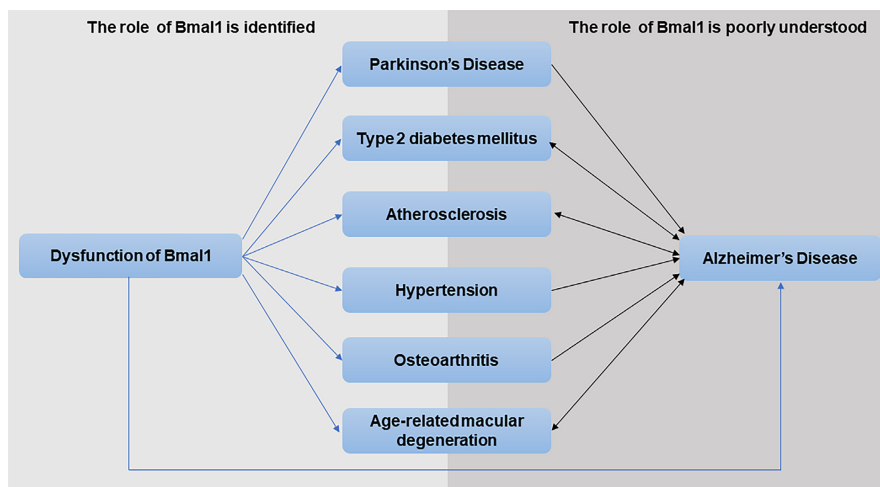
Melatonin is an amine hormone and an effective endogenous antioxidant (Tan et al., 2015), synthesized and secreted primarily by the pineal gland under normal light/dark conditions at night.

Numerous studies have suggested that melatonin plays an important role in regulating Bmal1 and exhibits beneficial physiological influences in the intervention of aging-associated diseases such as AD (Li, Zhang, et al., 2020), PD (Delgado-Lara et al., 2020), T2DM (Abdulwahab et al., 2021), AS (Xie, Tang, et al., 2020), and OA (Hosseinzadeh et al., 2016). Melatonin strengthens the expression of Bmal1 protein and regulates the expression of Bmal1 via PI3K/AKT signaling (Beker et al., 2019). A study showed that melatonin treatment increased Bmal1 expression in the hippocampus, induced rapid remodeling of hippocampal neurons, and altered the synaptic structure in the CA1 and dentate gyrus regions, suggesting that melatonin can regulate the structural changes in hippocampal neurons through the circadian clock molecule Bmal1 (Ikkeno & Nelson, 2015). Intermittent administration of melatonin can reset the daily pattern of *Bmal1* expression (Furtado et al., 2020). Furthermore, endogenous and exogenous melatonin ameliorates diabetes and related metabolic dysfunction by regulating insulin secretion and protecting against ROS (Espino et al., 2019). A study reported that melatonin increased the expression of SIRT1-BMAL1 pathway-related proteins in a dose-dependent manner and protected against cerebral ischemia-reperfusion-induced brain damage in diabetic mice (Liu, Cao, et al., 2021). The above evidence reveals that melatonin may be a prospective therapeutic agent for aging-related diseases.

### 6.2 | Natural compounds

Nobiletin, a citrus flavonoid, was shown to improve cognitive deficits and pathological features and motor and cognitive deficits in AD and PD, respectively, in animal model studies (Nakajima & Ohizumi, 2019). Nobiletin increases diurnal changes in Bmal1 expression and decreases A $\beta$  plaques in the cortex of APP/PS1 mice (Kim et al., 2021). A previous study showed that nobiletin directly binds and activates RORs, while the RORs enhance the activity of nobiletin on Bmal1 transcription. It was also postulated that nobiletin is a natural compound correlated with the enhancement of molecular clock and counteracts metabolic syndrome in a clock-dependent manner (He et al., 2016). Nobiletin can promote GSIS and

**FIGURE 4** The role of Bmal1 in the mechanisms connecting aging-related diseases to AD is poorly understood. The role of Bmal1 in AD, PD, T2DM, hypertension, AS, OA and AMD is identified (blue lines). These AD-associated diseases can also promote AD or have close interaction with AD through numerous pathways (black lines), and the role of Bmal1 in the mechanisms connecting these aging-related diseases to AD is poorly understood.





prevent ER stress-induced beta-cell apoptosis, thereby exhibiting its anti-diabetic effect (Kaneko et al., 2020). The anti-diabetic and anti-inflammatory effects of nobiletin were also determined in an in vitro human model (Nguyen-Ngo et al., 2020).

Tea polyphenols contain powerful antioxidant properties and ameliorate mitochondrial dysfunction in a Bmal1-dependent manner, playing an important role in the recovery of neurodegenerative diseases (Qi et al., 2017). Evidence has shown that tea polyphenols also improve glucose metabolism (Egbuna et al., 2021) and memory decline in aging models of rats (Song, Zhang, et al., 2021). Another natural product, LBP-4a prepared from *Lycium barbarum*, can improve hyperglycemia and insulin resistance by regulating biological rhythms, which are related to the increased expression of Bmal1 in pancreatic islet cells to improve impaired pancreatic islets (Zhao et al., 2016).

### 6.3 | Metformin and D-Ser2-oxymodulin

In db/db mice, decreased AMP-activated protein kinase (AMPK) activity, NAMPT expression, and SIRT1 expression in WAT were associated with suppressed expression of CLOCK and Bmal1 (Caton et al., 2011). Metformin treatment restored AMPK levels together with evidently increased expression of NAMPT, SIRT1, CLOCK, and Bmal1, suggesting that metformin may regulate glycolipid metabolism partly through AMPK-NAMPT-SIRT1-mediated alterations in the clock components (Caton et al., 2011). Metformin protects against the principal inwardly rectifying potassium-conducting channel (Kir4.1) in Müller cells; since, Bmal1 plays an intermediary role in the AMPK-mediated increase in Kir4.1 protein expression, which makes Bmal1 supposed to be a potential therapeutic target to prevent Müller cell dysfunction in diabetic retinopathy (Alex et al., 2020). Although several findings support the therapeutic potential of metformin for age-related diseases, including AD (Farr et al., 2019), PD (Mor et al., 2020), AS (Wu et al., 2021), and OA (Feng et al., 2020), involvement of Bmal1 is still poorly understood.

D-Ser2-oxymodulin (Oxy), a new glucagon-like peptide-1 receptor/glucagon receptor (GLP-1R/GCGR) dual receptor agonist, can rescue A $\beta$ 31-35-induced circadian rhythm disturbance and increase the expression of Bmal1 (Wang, Zhao, et al., 2020). It has also been illustrated that Oxy acts a part in the improvement of synaptic plasticity, reduction of A $\beta$ , and restoration of phosphatidylinositol-3-kinase/protein kinase B/glycogen synthase kinase-3 $\beta$  (PI3K/AKT/GSK3 $\beta$ ) cell signaling in the hippocampus, and hence, it may be used for the treatment of AD (Wang, Han, et al., 2020).

### 6.4 | Other interventions

Bmal1 gene expression in skeletal muscle was found to elevate after nearly three months of exercise intervention, along with enhanced

peripheral insulin sensitivity and reduced BMI and body weight, demonstrating that metabolic benefits from long-term exercise are partly mediated by the clock molecule Bmal1 (Erickson et al., 2020). A normal chow-fed, ad libitum, circadian mutant mice (including mice with liver-specific KO of *Bmal1*) developed some aspects of metabolic dysfunction that were exacerbated with age, while the time-restricted feeding (TRF) prevented the whole-body fat accumulation and serum hyperlipidemia, suggesting that TRF contributes to sustaining metabolic health in the condition of circadian rhythm/molecular clock defects such as *Bmal1* deletion (Chaix et al., 2019). Another study found that TRF was related to increased Bmal1 mRNA levels in the liver and improved metabolic health (Regmi et al., 2021). Adiponectin, an adipocyte-derived hormone, ameliorates the aberrant expression of the Bmal1 mRNA/protein induced by A $\beta$ 31-35 via inhibition of GSK3 $\beta$  activity, and exogenous administration of adiponectin may be a promising treatment for AD (Yuan et al., 2021). NAD<sup>+</sup> deficiency in diverse tissues with age contributes to the progress of multiple age-related diseases such as T2DM, AD, vascular dysfunction (de Picciotto et al., 2016), and cerebral ischemia. Accumulating evidence suggests that supplementation with nicotinamide mononucleotide (NMN) or nicotinamide riboside (NR) to increase NAD<sup>+</sup> levels is a promising therapy for diverse aging-related diseases (Hong et al., 2020). Recently, Zhu et al. found that increased cytosolic NAD<sup>+</sup> could restore hypoxic cell proliferation and myofiber formation in Bmal1-deficient myoblasts (Zhu et al., 2022). Hence, exercise, TRF, adiponectin, and supplementation with NMN and NR might also be promising treatment strategies for aging-related diseases that are dependent on the modulation of Bmal1.

## 7 | CONCLUSIONS AND FUTURE DIRECTIONS

Mounting evidence supports the existence of a significant interaction between Bmal1 deficiency/abnormality and aging-related diseases. Some mechanisms related to Bmal1 deficiency in AD and associated aging-related diseases (Figures 2 and 3) and interventions have been reviewed in the present study. Future studies are required to further understand the physiological intricacies of Bmal1 in the context of aging, with the following main to-be-explored aspects: (1) the signaling pathway network of Bmal1 mechanisms in aging-related diseases needs to be established, (2) several studies have shown that Bmal1 deficiency is involved in the occurrence and development of many other diseases not discussed in this review such as dilated cardiomyopathy (Lefta et al., 2012; Li, Li, et al., 2020) and cancer (Stokes et al., 2021; Wang et al., 2019). Therefore, the relationship of Bmal1 with other key diseases remains to be explored, (3) third, the effects of different Bmal1-modulating interventions on aging-related diseases have been analyzed in animal and cell culture model studies. Patients-oriented studies are lacking, and hence, interventions with sufficient evidence of efficacy should be selected to confirm their therapeutic potential using randomized controlled trials.



TABLE 2 Therapeutic strategies targeting Bmal1 in aging-related diseases in different models

First Author, Year	Design and subjects	Age <sup>a</sup>	Treatment	Bmal1 alterations	Results
Ikeno and Nelson (2015)	Siberian hamsters	16–18 w	Melatonin, 20 µg, subcutaneous injections 4h before killed	Increased	Remodeling of hippocampal neurons, altered the synaptic structure in the CA1 region and the dentate gyrus region
Beker et al. (2019)	N2A cells mimic focal cerebral ischemia in vitro	NA	Melatonin (MEL), 1 µM	Increased	Increased cellular survival under normoxic conditions and after oxygen–glucose deprivation
Delgado-Lara et al. (2020)	A double-blind, cross-over, placebo-controlled RCT, 26 patients with stage 1–3 PD	Median age = 55.5 years	First group: MEL (25 mg, twice a day for 3 months)- 4 days of washout period -Placebo (25 mg, twice a day for 3 months) Second group: Placebo- washout period - MEL. At the same way	The levels of Bmal1 genes increased from 0.56 to 2.2	The global perception of sleep comfort was increased
Kondratov et al. (2009)	Bmal1 <sup>-/-</sup> mice	4–45 w	Antioxidant N-acetyl-L-cysteine (NAC), starting from prenatal development, ending in spontaneous death	NA	The age-dependent weight loss (40 w) was delayed; the incidence of cataracts (phenotype of premature aging) was decreased, and the lifespan was extended
Kim et al. (2021)	APP/PS1 mice, 3 months	19–22 months	Regular diets containing equivalent macronutrients with Purina 5053 with or without 0.1% nobiletin (NOB), for 16–18 months	The mRNA levels of Bmal1 at ZT6 was higher compared with ZT18	The amounts of full-length APP proteins (APP-FL) were significantly reduced by NOB at ZT6, but not at ZT18. The expression of clock-controlled metabolic genes involved in insulin signaling and mitochondrial function were enhanced. The Aβ plaques in the cortex were reduced
He et al. (2016)	C57BL/6J mice, 6 week. HFD for 10 weeks. <i>Clock</i> <sup>Δ19/Δ19</sup> C57BL/6J mice. db/db mice. <i>db/dbClock</i> <sup>Δ19/Δ19</sup> mutant mice	16 w	Nobiletin (200 mg/kg body weight) via oral gavage every other day, in the time window of ZT8–10, for 10 weeks	Increased Bmal1 promoter-driven luciferase reporter expression with wild-type, but not mutant	NOB improved glucose and lipid homeostasis in diet-induced obesity (DIO) mice and <i>db/db</i> mice but not in <i>Clock</i> <sup>Δ19/Δ19</sup> C57BL/6J mice, and improved glucose homeostasis in <i>db/db</i> <i>Clock</i> <sup>Δ19/Δ19</sup> mutant mice
Qi et al. (2017)	Human neuroblastoma SH-SY5Y cell line	5–8 generations	Pretreated with tea polyphenols (TP, purity of > 98%) (40 µg/ml) for 12h and then with H2O2 (100 µM) for 12h after a washing with PBS	TP functions in a Bmal1-dependent manner	Result1: TP suppressed H2O2-triggered apoptosis and loss of cell viability and ameliorated oxidative stress and mitochondrial dysfunction. Result2: The neuroprotective effects of TP were abolished by silencing Bmal1 expression with siRNA

(Continues)



TABLE 2 (Continued)

First Author, Year	Design and subjects	Age <sup>a</sup>	Treatment	Bmal1 alterations	Results
Quezada-Fernández et al. (2019)	A double-blind, placebo-controlled RCT, 20 patients with T2DM, mean age = 53.2 years	Green tea extract (n = 10, mean age = 50.2), placebo (n = 141, mean age = 56.1)	400 mg of decaffeinated green tea extract (polyphenols ≥90%) or placebo for 12 weeks	NA	cAlx75 (75 bpm heart rate adjusted central augmentation index) was decreased. Arterial stiffness was improved
Zhao et al. (2016)	T2DM rats (high-sucrose-fat diets for 4 weeks, then intraperitoneally injected with 0.5% STZ solution at the dose of 50 mg/kg weights)	>8 w	LBP-4a treatment group at high dose (10 mg/kg-d), and the LBP-4a treatment group at low dose (5 mg/kg-d), intragastric administration for 4 weeks	Increased BMAL1 expression in pancreatic islet cells in high dose group	Blood glucose and insulin were decreased in both high dose and low dose groups. Melatonin was increased in the high dose group. Disruption of pancreatic architecture was meliorated in both high dose (more obvious) and low dose groups
Caton et al. (2011)	Db/db, Db/+ mice, 7 w	8 w	<b>Metformin</b> , (250 mg/kg/day for 7 days), by oral gavage	Increased Bmal1 mRNA expression by 2.1-fold	The BMAL1 protein expression was increased in tWAT of db/db mice. The fasting glucose and fasting insulin were decreased
Alex et al. (2020)	Db/db, Db/+ mice, 8–10 w	10–12 w	<b>Metformin</b> (164 ± 40 mg/kg/day for 14 days)	Increased BMAL1 protein	Body mass and Glycated hemoglobin were decreased. The retinal Kir4.1 level in Müller cells was increased
Wang, Zhao, et al. (2020)	C57BL/6 mice received Aβ31–35 via hippocampal injection, 9–11 w	9–11 w	<b>D-Ser2-Oxynomodulin</b> (Oxy) (30 nmol/kg), via hippocampal injection, 15 min before Aβ31–35 was injected	The rhythmic mRNA and protein expression of Bmal1 in the hippocampus were improved	The Aβ31–35-induced disruption of the circadian rhythm (The sleep–wake cycle, free-running period, locomotor activity, and ratio of the subjective night activity/total activity) was recovered. The abnormal expression of Bmal1 and Per2 was improved in the mice hippocampus and HT22 cells
Erickson et al. (2020)	26 adults with prediabetes and obesity provided with isoenergetic diets	Mean age = 66 years	12-week exercise intervention (5 days per week at 85% of heart rate max on a treadmill for 60 min)	BMAL1 gene expression in skeletal muscle was increased	Body weight, BMI, and total abdominal adiposity were decreased. Exercise capacity and peripheral insulin sensitivity were improved. The fold change in BMAL1 was positively correlated with insulin sensitivity and negatively correlated with BMI



TABLE 2 (Continued)

First Author, Year	Design and subjects	Age <sup>a</sup>	Treatment	Bmal1 alterations	Results
Chaix et al. (2019)	Liver-specific Bmal1 knockout mice, 12 w	24 w	12 weeks on the time-restricted feeding (TRF)	NA	TRF prevented whole-body fat accumulation and serum hyperlipidemia
Regmi et al. (2021)	C57BL/6J mice, 8 w, high-fat diet ad libitum for 4 w	20 w	A further 8 weeks on TRFe (initiated at lights off) or TRFd (initiated at 4-h after lights off)	The amplitude of Bmal1 mRNA levels in liver was increased	TRF reduced weight and fat mass (greater reduction in TRFe), improved glucose tolerance, and protected mice from high-fat diet-induced hepatosteatosis (no difference in TRFe vs TRFd)

Abbreviations: A $\beta$ , amyloid  $\beta$ -protein; APP, amyloid precursor protein; BMAL1, brain and muscle Arnt-like protein-1; CA, cornu ammonis; DIO, diet-induced obesity; MEL, melatonin; NAC, antioxidant N-acetyl-L-cysteine; NOB, nobiletin; N2a, neuroblastoma cell lines; Oxy, D-Ser2-oxymodulin; PD, Parkinson's disease; RCT, randomized controlled trial; STZ, streptozotocin; TP, tea polyphenols; TRF, time-restricted feeding; T2DM, type 2 diabetes mellitus; WAT, white adipose tissue; ZT, Zeitgeber time.

<sup>a</sup>The age of mice was sacrificed, or the time point of relative parameters w collected.

## ACKNOWLEDGMENTS

Special thanks to Hao Duan for his assistance with some image drawings.

## AUTHOR CONTRIBUTIONS

RF and YY drafted the manuscript. XP, LX, KD, DM, WX, XS, SZ, JC, XY, and YY revised the manuscript. All authors have read and agreed to the published version of the manuscript.

## FUNDING INFORMATION

This work was funded by grants from the National Nature Science Foundation of China (Grant number: 81974114) and the Jie Chu Jing Ying foundation (grant number 2018076).

## CONFLICT OF INTEREST

The authors have no conflicts of interest to disclose.

## DATA AVAILABILITY STATEMENT

No data were created or analyzed in this study. Data sharing is not applicable to this review article.

## ORCID

Rongping Fan <https://orcid.org/0000-0003-3587-3365>

Yan Yang <https://orcid.org/0000-0001-8146-8040>

## REFERENCES

- Abdulwahab, D. A., El-Missiry, M. A., Shabana, S., Othman, A. I., & Amer, M. E. (2021). Melatonin protects the heart and pancreas by improving glucose homeostasis, oxidative stress, inflammation and apoptosis in T2DM-induced rats. *Heliyon*, 7(3), e06474. <https://doi.org/10.1016/j.heliyon.2021.e06474>
- Akagi, R., Akatsu, Y., Fisch, K. M., Alvarez-Garcia, O., Teramura, T., Muramatsu, Y., Saito, M., Sasho, T., Su, A. I., & Lotz, M. K. (2017). Dysregulated circadian rhythm pathway in human osteoarthritis: NR1D1 and BMAL1 suppression alters TGF- $\beta$  signaling in chondrocytes. *Osteoarthritis and Cartilage*, 25(6), 943–951. <https://doi.org/10.1016/j.joca.2016.11.007>
- Akladios, A., Azzam, S., Hu, Y., & Feng, P. (2018). Bmal1 knockdown suppresses wake and increases immobility without altering orexin A, corticotrophin-releasing hormone, or glutamate decarboxylase. *CNS Neuroscience & Therapeutics*, 24(6), 549–563. <https://doi.org/10.1111/cns.12815>
- Alex, A., Luo, Q., Mathew, D., Di, R., & Bhatwadekar, A. D. (2020). Metformin corrects abnormal circadian rhythm and Kir4.1 channels in diabetes. *Investigative Ophthalmology & Visual Science*, 61(6), 46. <https://doi.org/10.1167/iov.61.6.46>
- Ali, A. A. H., Schwarz-Herzke, B., Rollenhagen, A., Anstötz, M., Holub, M., Lübke, J., Rose, C. R., Schnittler, H. J., & von Gall, C. (2020). Bmal1-deficiency affects glial synaptic coverage of the hippocampal mossy fiber synapse and the actin cytoskeleton in astrocytes. *Glia*, 68(5), 947–962. <https://doi.org/10.1002/glia.23754>
- Alliston, T., Hernandez, C. J., Findlay, D. M., Felson, D. T., & Kennedy, O. D. (2018). Bone marrow lesions in osteoarthritis: What lies beneath. *Journal of Orthopaedic Research*, 36(7), 1818–1825. <https://doi.org/10.1002/jor.23844>
- Ando, H., Takamura, T., Matsuzawa-Nagata, N., Shima, K. R., Eto, T., Misu, H., Shiramoto, M., Tsuru, T., Irie, S., Fujimura, A., & Kaneko, S. (2009). Clock gene expression in peripheral leucocytes of patients



- with type 2 diabetes. *Diabetologia*, 52(2), 329–335. <https://doi.org/10.1007/s00125-008-1194-6>
- Anea, C. B., Cheng, B., Sharma, S., Kumar, S., Caldwell, R. W., Yao, L., Ali, M. I., Merloiu, A. M., Stepp, D. W., Black, S. M., Fulton, D. J., & Rudic, R. D. (2012). Increased superoxide and endothelial NO synthase uncoupling in blood vessels of Bmal1-knockout mice. *Circulation Research*, 111(9), 1157–1165. <https://doi.org/10.1161/CIRCRESAHA.111.261750>
- Barca-Mayo, O., Pons-Espinal, M., Follert, P., Armirotti, A., Berdondini, L., & De Pietri Tonelli, D. (2017). Astrocyte deletion of Bmal1 alters daily locomotor activity and cognitive functions via GABA signaling. *Nature Communications*, 8, 14336. <https://doi.org/10.1038/ncomms14336>
- Beker, M. C., Caglayan, B., Caglayan, A. B., Kelestemur, T., Yalcin, E., Caglayan, A., Kilic, E., Baykal, A. T., Reiter, R. J., & Kilic, E. (2019). Interaction of melatonin and Bmal1 in the regulation of PI3K/AKT pathway components and cellular survival. *Scientific Reports*, 9(1), 19082. <https://doi.org/10.1038/s41598-019-55663-0>
- Beker, M. C., Caglayan, B., Yalcin, E., Caglayan, A. B., Turkseven, S., Gurel, B., Kelestemur, T., Sertel, E., Sahin, Z., Kutlu, S., Kilic, U., Baykal, A. T., & Kilic, E. (2018). Time-of-day dependent neuronal injury after ischemic stroke: Implication of circadian clock transcriptional factor Bmal1 and survival kinase AKT. *Molecular Neurobiology*, 55(3), 2565–2576. <https://doi.org/10.1007/s12035-017-0524-4>
- Bellary, S., Kyrou, I., Brown, J. E., & Bailey, C. J. (2021). Type 2 diabetes mellitus in older adults: Clinical considerations and management. *Nature Reviews Endocrinology*, 17(9), 534–548. <https://doi.org/10.1038/s41574-021-00512-2>
- Bonkowski, M. S., & Sinclair, D. A. (2016). Slowing ageing by design: The rise of NAD and sirtuin-activating compounds. *Nature Reviews Molecular Cell Biology*, 17(11), 679–690. <https://doi.org/10.1038/nrm.2016.93>
- Breen, D. P., Vuono, R., Nawarathna, U., Fisher, K., Shneerson, J. M., Reddy, A. B., & Barker, R. A. (2014). Sleep and circadian rhythm regulation in early Parkinson disease. *JAMA Neurology*, 71(5), 589–595. <https://doi.org/10.1001/jamaneurol.2014.65>
- Bunger, M. K., Wilsbacher, L. D., Moran, S. M., Clendenen, C., Radcliffe, L. A., Hogenesch, J. B., Simon, M. C., Takahashi, J. S., & Bradfield, C. A. (2000). Mop3 is an essential component of the master circadian pacemaker in mammals. *Cell*, 103(7), 1009–1017.
- Cai, Y., Liu, S., Sothorn, R. B., Xu, S., & Chan, P. (2010). Expression of clock genes Per1 and Bmal1 in total leukocytes in health and Parkinson's disease. *European Journal of Neurology*, 17(4), 550–554. <https://doi.org/10.1111/j.1468-1331.2009.02848.x>
- Caton, P. W., Kieswich, J., Yaqoob, M. M., Holness, M. J., & Sugden, M. C. (2011). Metformin opposes impaired AMPK and SIRT1 function and deleterious changes in core clock protein expression in white adipose tissue of genetically-obese db/db mice. *Diabetes, Obesity & Metabolism*, 13(12), 1097–1104. <https://doi.org/10.1111/j.1463-1326.2011.01466.x>
- Cermakian, N., Lamont, E. W., Boudreau, P., & Boivin, D. B. (2011). Circadian clock gene expression in brain regions of Alzheimer's disease patients and control subjects. *Journal of biological rhythms*, 26(2), 160–170. <https://doi.org/10.1177/0748730410395732>
- Chaix, A., Lin, T., Le, H. D., Chang, M. W., & Panda, S. (2019). Time-restricted feeding prevents obesity and metabolic syndrome in mice lacking a circadian clock. *Cell Metabolism*, 29(2), 303–319.e4. <https://doi.org/10.1016/j.cmet.2018.08.004>
- Chang, H.-C., & Guarente, L. (2013). SIRT1 mediates central circadian control in the SCN by a mechanism that decays with aging. *Cell*, 153(7), 1448–1460. <https://doi.org/10.1016/j.cell.2013.05.027>
- Chang, H.-C., & Guarente, L. (2014). SIRT1 and other sirtuins in metabolism. *Trends in Endocrinology and Metabolism*, 25(3), 138–145. <https://doi.org/10.1016/j.tem.2013.12.001>
- Chen, Q., Peng, X. D., Huang, C. Q., Hu, X. Y., & Zhang, X. M. (2015). Association between ARNTL (BMAL1) rs2278749 polymorphism T>C and susceptibility to Alzheimer disease in a Chinese population. *Genetics and molecular research: GMR*, 14(4), 18515–18522. <https://doi.org/10.4238/2015.December.23.39>
- Chen, G., Tang, Q., Yu, S., Xie, Y., Sun, J., Li, S., & Chen, L. (2020). The biological function of BMAL1 in skeleton development and disorders. *Life Sciences*, 253, 117636. <https://doi.org/10.1016/j.lfs.2020.117636>
- Chen, C., Zhou, M., Ge, Y., & Wang, X. (2020). SIRT1 and aging related signaling pathways. *Mechanisms of Ageing and Development*, 187, 111215. <https://doi.org/10.1016/j.mad.2020.111215>
- Chen, G., Zhao, H., Ma, S., Chen, L., Wu, G., Zhu, Y., Zhu, J., Ma, C., & Zhao, H. (2020). Circadian rhythm protein bmal1 modulates cartilage gene expression in temporomandibular joint osteoarthritis the MAPK/ERK pathway. *Frontiers in Pharmacology*, 11, 527744. <https://doi.org/10.3389/fphar.2020.527744>
- Chhunchha, B., Kubo, E., & Singh, D. P. (2020). Clock protein Bmal1 and Nrf2 cooperatively control aging or oxidative response and redox homeostasis by regulating rhythmic expression of Prdx6. *Cell*, 9(8), 1861. <https://doi.org/10.3390/cells9081861>
- Cibulka, M., Brodnanova, M., Grendar, M., Necpal, J., Benetin, J., Han, V., Kurca, E., Nosal, V., Skorvanek, M., Vesely, B., Stanclova, A., Lasabova, Z., Pös, Z., Szemes, T., Stuchlik, S., Grofik, M., & Kolisek, M. (2022). Alzheimer's disease-associated SNP rs708727 in may increase risk for Parkinson's disease: Report from enlarged Slovak Study. *International Journal of Molecular Sciences*, 23(3), 1604. <https://doi.org/10.3390/ijms23031604>
- Cortes-Canteli, M., & Iadecola, C. (2020). Alzheimer's disease and vascular aging: JACC focus seminar. *Journal of the American College of Cardiology*, 75(8), 942–951. <https://doi.org/10.1016/j.jacc.2019.10.062>
- Dall'érac, G., Zapata, J., & Rouach, N. (2018). Versatile control of synaptic circuits by astrocytes: Where, when and how? *Nature Reviews Neuroscience*, 19(12), 729–743. <https://doi.org/10.1038/s41583-018-0080-6>
- de Picciotto, N. E., Gano, L. B., Johnson, L. C., Martens, C. R., Sindler, A. L., Mills, K. F., Imai, S., & Seals, D. R. (2016). Nicotinamide mononucleotide supplementation reverses vascular dysfunction and oxidative stress with aging in mice. *Aging Cell*, 15(3), 522–530. <https://doi.org/10.1111/acer.12461>
- Delgado-Lara, D. L., González-Enríquez, G. V., Torres-Mendoza, B. M., González-Usigli, H., Cárdenas-Bedoya, J., Macías-Islas, M. A., de la Rosa, A. C., Jiménez-Delgado, A., Pacheco-Moisés, F., Cruz-Serrano, J. A., & Ortiz, G. G. (2020). Effect of melatonin administration on the PER1 and BMAL1 clock genes in patients with Parkinson's disease. *Biomedicine & Pharmacotherapy = Biomedicine & Pharmacotherapie*, 129, 110485. <https://doi.org/10.1016/j.biopha.2020.110485>
- Dudek, M., Gossan, N., Yang, N., Im, H.-J., Ruckshanthi, J. P. D., Yoshitane, H., Li, X., Jin, D., Wang, P., Boudiffa, M., Bellantuono, I., Fukada, Y., Boot-Handford, R. P., & Meng, Q. J. (2016). The chondrocyte clock gene Bmal1 controls cartilage homeostasis and integrity. *The Journal of Clinical Investigation*, 126(1), 365–376. <https://doi.org/10.1172/JCI82755>
- Duncan, M. J., Prochot, J. R., Cook, D. H., Tyler Smith, J., & Franklin, K. M. (2013). Influence of aging on Bmal1 and Per2 expression in extra-SCN oscillators in hamster brain. *Brain Research*, 1491, 44–53. <https://doi.org/10.1016/j.brainres.2012.11.008>
- Early, J. O., Menon, D., Wyse, C. A., Cervantes-Silva, M. P., Zaslon, Z., Carroll, R. G., Palsson-McDermott, E., Angiari, S., Ryan, D. G., Corcoran, S. E., Timmons, G., Geiger, S. S., Fitzpatrick, D. J., O'Connell, D., Xavier, R. J., Hokamp, K., O'Neill, L. A. J., & Curtis, A. M. (2018). Circadian clock protein BMAL1 regulates IL-1 $\beta$  in macrophages via NRF2. *Proceedings of the National Academy of Sciences of the United States of America*, 115(36), E8460–E8468. <https://doi.org/10.1073/pnas.1800431115>





- Egbuna, C., Awuchi, C. G., Kushwaha, G., Rudrapal, M., Patrick-Iwuanyanwu, K. C., Singh, O., Odoh, U. E., Khan, J., Jeevanandam, J., Kumarasamy, S., Chukwube, V. O., Narayanan, M., Palai, S., Gáman, M. A., Uche, C. Z., Ogaji, D. S., Ezeofor, N. J., Mteawa, A. G., Patrick-Iwuanyanwu, C. C., ... Chikwendu, C. J. (2021). Bioactive compounds effective against type 2 diabetes mellitus: A systematic review. *Current Topics in Medicinal Chemistry*, 21(12), 1067–1095. <https://doi.org/10.2174/1568026621666210509161059>
- Erickson, M. L., Zhang, H., Mey, J. T., & Kirwan, J. P. (2020). Exercise training impacts skeletal muscle clock machinery in prediabetes. *Medicine and Science in Sports and Exercise*, 52(10), 2078–2085. <https://doi.org/10.1249/MSS.0000000000002368>
- Espino, J., Rodríguez, A. B., & Pariente, J. A. (2019). Melatonin and oxidative stress in the diabetic state: Clinical implications and potential therapeutic applications. *Current Medicinal Chemistry*, 26(22), 4178–4190. <https://doi.org/10.2174/0929867325666180410094149>
- Ettcheto, M., Olloquequi, J., Sánchez-López, E., Busquets, O., Cano, A., Manzine, P. R., Beas-Zarate, C., Castro-Torres, R. D., García, M. L., Bulló, M., Auladell, C., Folch, J., & Camins, A. (2019). Benzodiazepines and related drugs as a risk factor in Alzheimer's disease dementia. *Frontiers in Aging Neuroscience*, 11, 344. <https://doi.org/10.3389/fnagi.2019.00344>
- Fang, L., Tang, B.-S., Fan, K., Wan, C.-M., Yan, X.-X., & Guo, J.-F. (2018). Alzheimer's disease susceptibility genes modify the risk of Parkinson disease and Parkinson's disease-associated cognitive impairment. *Neuroscience Letters*, 677, 55–59. <https://doi.org/10.1016/j.neulet.2018.04.042>
- Farr, S. A., Roesler, E., Niehoff, M. L., Roby, D. A., McKee, A., & Morley, J. E. (2019). Metformin improves learning and memory in the SAMP8 mouse model of Alzheimer's disease. *Journal of Alzheimer's Disease*, 68(4), 1699–1710. <https://doi.org/10.3233/JAD-181240>
- Feng, X., Pan, J., Li, J., Zeng, C., Qi, W., Shao, Y., Liu, X., Liu, L., Xiao, G., Zhang, H., Bai, X., & Cai, D. (2020). Metformin attenuates cartilage degeneration in an experimental osteoarthritis model by regulating AMPK/mTOR. *Aging*, 12(2), 1087–1103. <https://doi.org/10.18632/aging.102635>
- Furtado, A., Astaburuaga, R., Costa, A., Duarte, A. C., Gonçalves, I., Cipolla-Neto, J., Lemos, M. C., Carro, E., Relógio, A., Santos, C. R. A., & Quintela, T. (2020). The rhythmicity of clock genes is disrupted in the choroid plexus of the APP/PS1 mouse model of Alzheimer's disease. *Journal of Alzheimer's Disease*, 77(2), 795–806. <https://doi.org/10.3233/JAD-200331>
- Gomes, B. A. Q., Silva, J. P. B., Romero, C. F. R., dos Santos, S., Rodrigues, C. A., Gonçalves, P. R., Sakai, J. T., Mendes, P. F. S., Varela, E. L. P., & Monteiro, M. C. (2018). Neuroprotective mechanisms of resveratrol in Alzheimer's disease: Role of SIRT1. *Oxidative Medicine and Cellular Longevity*, 2018, 8152373. <https://doi.org/10.1155/2018/8152373>
- Gómez-Benito, M., Granado, N., García-Sanz, P., Michel, A., Dumoulin, M., & Moratalla, R. (2020). Modeling Parkinson's disease with the alpha-synuclein protein. *Frontiers in Pharmacology*, 11, 356. <https://doi.org/10.3389/fphar.2020.00356>
- Gunton, J. E., Kulkarni, R. N., Yim, S., Okada, T., Hawthorne, W. J., Tseng, Y.-H., Roberson, R. S., Ricordi, C., O'Connell, P. J., Gonzalez, F. J., & Kahn, C. R. (2005). Loss of ARNT/HIF1beta mediates altered gene expression and pancreatic-islet dysfunction in human type 2 diabetes. *Cell*, 122(3), 337–349.
- Hadjipanayi, E., & Schilling, A. F. (2013). Hypoxia-based strategies for angiogenic induction: The dawn of a new era for ischemia therapy and tissue regeneration. *Organogenesis*, 9(4), 261–272. <https://doi.org/10.4161/org.25970>
- He, B., Nohara, K., Park, N., Park, Y.-S., Guillory, B., Zhao, Z., Garcia, J. M., Koike, N., Lee, C. C., Takahashi, J. S., Yoo, S. H., & Chen, Z. (2016). The small molecule nobiletin targets the molecular oscillator to enhance circadian rhythms and protect against metabolic syndrome. *Cell Metabolism*, 23(4), 610–621. <https://doi.org/10.1016/j.cmet.2016.03.007>
- Hong, W., Mo, F., Zhang, Z., Huang, M., & Wei, X. (2020). Nicotinamide mononucleotide: A promising molecule for therapy of diverse diseases by targeting NAD<sup>+</sup> metabolism. *Frontiers in Cell and Developmental Biology*, 8, 246. <https://doi.org/10.3389/fcell.2020.00246>
- Hosseinzadeh, A., Kamrava, S. K., Joghataei, M. T., Darabi, R., Shakeri-Zadeh, A., Shahriari, M., Reiter, R. J., Ghaznavi, H., & Mehrzadi, S. (2016). Apoptosis signaling pathways in osteoarthritis and possible protective role of melatonin. *Journal of Pineal Research*, 61(4), 411–425. <https://doi.org/10.1111/jpi.12362>
- Hou, Q., Zhang, S., Li, Y., Wang, H., Zhang, D., Qi, D., Li, Y., & Jiang, H. (2021). New insights on association between circadian rhythm and lipid metabolism in spontaneously hypertensive rats. *Life Sciences*, 271, 119145. <https://doi.org/10.1016/j.lfs.2021.119145>
- Hou, Y., Wei, Y., Lautrup, S., Yang, B., Wang, Y., Cordonnier, S., Mattson, M. P., Croteau, D. L., & Bohr, V. A. (2021). NAD supplementation reduces neuroinflammation and cell senescence in a transgenic mouse model of Alzheimer's disease via cGAS-STING. *Proceedings of the National Academy of Sciences of the United States of America*, 118(37), e2011226118. <https://doi.org/10.1073/pnas.2011226118>
- Huang, J., Peng, X., Fan, R., Dong, K., Shi, X., Zhang, S., Yu, X., & Yang, Y. (2021). Disruption of circadian clocks promotes progression of Alzheimer's disease in diabetic mice. *Molecular Neurobiology*, 58(9), 4404–4412. <https://doi.org/10.1007/s12035-021-02425-7>
- Hudson, N., Celkova, L., Hopkins, A., Greene, C., Storti, F., Ozaki, E., Fahey, E., Theodoropoulou, S., Kenna, P. F., Humphries, M. M., Curtis, A. M., Demmons, E., Browne, A., Liddie, S., Lawrence, M. S., Grimm, C., Cahill, M. T., Humphries, P., Doyle, S. L., & Campbell, M. (2019). Dysregulated claudin-5 cycling in the inner retina causes retinal pigment epithelial cell atrophy. *JCI Insight*, 4(15), e130273. <https://doi.org/10.1172/jci.insight.130273>
- Hulme, B., Didikoglu, A., Bradburn, S., Robinson, A., Canal, M., Payton, A., Pendleton, N., & Murgatroyd, C. (2020). Epigenetic regulation of BMAL1 with sleep disturbances and Alzheimer's disease. *Journal of Alzheimer's Disease*, 77(4), 1783–1792. <https://doi.org/10.3233/JAD-200634>
- Huo, M., Cao, X., Zhang, H., Lau, C. W., Hong, H., Chen, F. M., Huang, Y., Chawla, A., & Tian, X. Y. (2021). Loss of myeloid Bmal1 exacerbates hypertensive vascular remodelling through interaction with STAT6 in mice. *Cardiovascular Research*, cvab336. <https://doi.org/10.1093/cvr/cvab336>
- Huo, M., Huang, Y., Qu, D., Zhang, H., Wong, W. T., Chawla, A., Huang, Y., & Tian, X. Y. (2017). Myeloid deletion increases monocyte recruitment and worsens atherosclerosis. *FASEB Journal*, 31(3), 1097–1106. <https://doi.org/10.1096/fj.201601030R>
- Ikeno, T., & Nelson, R. J. (2015). Acute melatonin treatment alters dendritic morphology and circadian clock gene expression in the hippocampus of Siberian hamsters. *Hippocampus*, 25(2), 142–148. <https://doi.org/10.1002/hipo.22358>
- Innes, K. E., & Sambamoorthi, U. (2020). The association of osteoarthritis and related pain burden to incident Alzheimer's disease and related dementias: A retrospective cohort study of U.S. medicare beneficiaries. *Journal of Alzheimer's Disease*, 75(3), 789–805. <https://doi.org/10.3233/JAD-191311>
- Johnson, W. M., Wilson-Delfosse, A. L., & Mieyal, J. J. (2012). Dysregulation of glutathione homeostasis in neurodegenerative diseases. *Nutrients*, 4(10), 1399–1440. <https://doi.org/10.3390/nu4101399>
- Julien, C., Tremblay, C., Emond, V., Lebbadi, M., Salem, N., Bennett, D. A., & Calon, F. (2009). Sirtuin 1 reduction parallels the accumulation of tau in Alzheimer disease. *Journal of Neuropathology and Experimental Neurology*, 68(1), 48–58. <https://doi.org/10.1097/NEN.0b013e3181922348>



- Kaneko, Y. K., Kan, T., & Ishikawa, T. (2020). Citrus flavonoids as a target for the prevention of pancreatic  $\beta$ -cells dysfunction in diabetes. *Nihon yakurigaku zasshi*, 155(4), 209–213. <https://doi.org/10.1254/fpj.20024>
- Kao, S.-L., Wang, J.-H., Chen, S.-C., Li, Y.-Y., Yang, Y.-L., & Lo, R. Y. (2021). Impact of comorbidity burden on cognitive decline: A prospective cohort study of older adults with dementia. *Dementia and Geriatric Cognitive Disorders*, 50(1), 43–50. <https://doi.org/10.1159/000514651>
- Kim, E., Nohara, K., Wirianto, M., Escobedo, G., Jr., Lim, J. Y., Morales, R., Yoo, S. H., & Chen, Z. (2021). Effects of the clock modulator nobilletin on circadian rhythms and pathophysiology in female mice of an Alzheimer's disease model. *Biomolecules*, 11(7), 1004. <https://doi.org/10.3390/biom11071004>
- Kondratov, R. V., Kondratova, A. A., Gorbacheva, V. Y., Vykhovanets, O. V., & Antoch, M. P. (2006). Early aging and age-related pathologies in mice deficient in BMAL1, the core component of the circadian clock. *Genes & Development*, 20(14), 1868–1873.
- Kondratov, R. V., Vykhovanets, O., Kondratova, A. A., & Antoch, M. P. (2009). Antioxidant N-acetyl-L-cysteine ameliorates symptoms of premature aging associated with the deficiency of the circadian protein BMAL1. *Aging*, 1(12), 979–987.
- Koo, J.-H., Kang, E.-B., Oh, Y.-S., Yang, D.-S., & Cho, J.-Y. (2017). Treadmill exercise decreases amyloid- $\beta$  burden possibly via activation of SIRT-1 signaling in a mouse model of Alzheimer's disease. *Experimental Neurology*, 288, 142–152. <https://doi.org/10.1016/j.expneurol.2016.11.014>
- Kress, G. J., Liao, F., Dimitry, J., Cedeno, M. R., FitzGerald, G. A., Holtzman, D. M., & Musiek, E. S. (2018). Regulation of amyloid- $\beta$  dynamics and pathology by the circadian clock. *The Journal of Experimental Medicine*, 215(4), 1059–1068. <https://doi.org/10.1084/jem.20172347>
- Lananna, B. V., Nadarajah, C. J., Izumo, M., Cedeño, M. R., Xiong, D. D., Dimitry, J., Tso, C. F., McKee, C., Griffin, P., Sheehan, P. W., Haspel, J. A., Barres, B. A., Liddelow, S. A., Takahashi, J. S., Karatsoreos, I. N., & Musiek, E. S. (2018). Cell-autonomous regulation of astrocyte activation by the circadian clock protein BMAL1. *Cell Reports*, 25(1), 1–9.e5. <https://doi.org/10.1016/j.celrep.2018.09.015>
- Lautrup, S., Sinclair, D. A., Mattson, M. P., & Fang, E. F. (2019). NAD in brain aging and neurodegenerative disorders. *Cell Metabolism*, 30(4), 630–655. <https://doi.org/10.1016/j.cmet.2019.09.001>
- Lee, J., Kim, D. E., Griffin, P., Sheehan, P. W., Kim, D.-H., Musiek, E. S., & Yoon, S.-Y. (2020). Inhibition of REV-ERBs stimulates microglial amyloid-beta clearance and reduces amyloid plaque deposition in the 5XFAD mouse model of Alzheimer's disease. *Aging Cell*, 19(2), e13078. <https://doi.org/10.1111/accel.13078>
- Lee, J., Ma, K., Moulik, M., & Yechoor, V. (2018). Untimely oxidative stress in  $\beta$ -cells leads to diabetes—Role of circadian clock in  $\beta$ -cell function. *Free Radical Biology & Medicine*, 119, 69–74. <https://doi.org/10.1016/j.freeradbiomed.2018.02.022>
- Lee, J., Moulik, M., Fang, Z., Saha, P., Zou, F., Xu, Y., Nelson, D. L., Ma, K., Moore, D. D., & Yechoor, V. K. (2013). Bmal1 and  $\beta$ -cell clock are required for adaptation to circadian disruption, and their loss of function leads to oxidative stress-induced  $\beta$ -cell failure in mice. *Molecular and Cellular Biology*, 33(11), 2327–2338. <https://doi.org/10.1128/MCB.01421-12>
- Lee, S., Nam, H. G., & Kim, Y. (2021). The core circadian component, Bmal1, is maintained in the pineal gland of old killifish brain. *iScience*, 24(1), 101905. <https://doi.org/10.1016/j.isci.2020.101905>
- Lefta, M., Campbell, K. S., Feng, H.-Z., Jin, J.-P., & Esser, K. A. (2012). Development of dilated cardiomyopathy in Bmal1-deficient mice. *American Journal of Physiology. Heart and Circulatory Physiology*, 303(4), H475–H485. <https://doi.org/10.1152/ajpheart.00238.2012>
- Li, E., Li, X., Huang, J., Xu, C., Liang, Q., Ren, K., Bai, A., Lu, C., Qian, R., & Sun, N. (2020). BMAL1 regulates mitochondrial fission and mitophagy through mitochondrial protein BNIP3 and is critical in the development of dilated cardiomyopathy. *Protein & Cell*, 11(9), 661–679. <https://doi.org/10.1007/s13238-020-00713-x>
- Li, H., Song, S., Wang, Y., Huang, C., Zhang, F., Liu, J., & Hong, J.-S. (2019). Low-grade inflammation aggravates rotenone neurotoxicity and disrupts circadian clock gene expression in rats. *Neurotoxicity Research*, 35(2), 421–431. <https://doi.org/10.1007/s12640-018-9968-1>
- Li, T., Cheng, C., Jia, C., Leng, Y., Qian, J., Yu, H., Liu, Y., Wang, N., Yang, Y., al-Nusaif, M., & Le, W. (2021). Peripheral clock system abnormalities in patients with Parkinson's disease. *Frontiers in Aging Neuroscience*, 13, 736026. <https://doi.org/10.3389/fnagi.2021.736026>
- Li, X., Liu, N., Wang, Y., Liu, J., Shi, H., Qu, Z., du, T., Guo, B., & Gu, B. (2017). Brain and muscle aryl hydrocarbon receptor nuclear translocator-like protein-1 cooperates with glycogen synthase kinase-3 $\beta$  to regulate osteogenesis of bone-marrow mesenchymal stem cells in type 2 diabetes. *Molecular and Cellular Endocrinology*, 440, 93–105. <https://doi.org/10.1016/j.mce.2016.10.001>
- Li, Y., Zhang, J., Wan, J., Liu, A., & Sun, J. (2020). Melatonin regulates A $\beta$  production/clearance balance and A $\beta$  neurotoxicity: A potential therapeutic molecule for Alzheimer's disease. *Biomedicine & Pharmacotherapy = Biomedecine & Pharmacotherapie*, 132, 110887. <https://doi.org/10.1016/j.biopha.2020.110887>
- Liang, S., Hu, J., Zhang, A., Li, F., & Li, X. (2020). miR-155 induces endothelial cell apoptosis and inflammatory response in atherosclerosis by regulating Bmal1. *Experimental and Therapeutic Medicine*, 20(6), 128. <https://doi.org/10.3892/etm.2020.9259>
- Liu, L., Cao, Q., Gao, W., Li, B.-Y., Zeng, C., Xia, Z., & Zhao, B. (2021). Melatonin ameliorates cerebral ischemia-reperfusion injury in diabetic mice by enhancing autophagy via the SIRT1-BMAL1 pathway. *FASEB Journal*, 35(12), e22040. <https://doi.org/10.1096/fj.202002718RR>
- Liu, W.-W., Wei, S.-Z., Huang, G.-D., Liu, L.-B., Gu, C., Shen, Y., Wang, X. H., Xia, S. T., Xie, A. M., Hu, L. F., Wang, F., & Liu, C. F. (2020). BMAL1 regulation of microglia-mediated neuroinflammation in MPTP-induced Parkinson's disease mouse model. *FASEB Journal*, 34(5), 6570–6581. <https://doi.org/10.1096/fj.201901565RR>
- Liu, X., Xiao, W., Jiang, Y., Zou, L., Chen, F., Xiao, W., Zhang, X., Cao, Y., Xu, L., & Zhu, Y. (2021). Bmal1 regulates the redox rhythm of HSPB1, and homooxidized HSPB1 attenuates the oxidative stress injury of cardiomyocytes. *Oxidative Medicine and Cellular Longevity*, 2021, 5542815. <https://doi.org/10.1155/2021/5542815>
- Ma, C., Wu, G., Wang, Z., Wang, P., Wu, L., Zhu, G., & Zhao, H. (2014). Effects of chronic sleep deprivation on the extracellular signal-regulated kinase pathway in the temporomandibular joint of rats. *PLoS One*, 9(9), e107544. <https://doi.org/10.1371/journal.pone.0107544>
- Ma, Z., Jin, X., Qian, Z., Li, F., Xu, M., Zhang, Y., Kang, X., Li, H., Gao, X., Zhao, L., Zhang, Z., Zhang, Y., Wu, S., & Sun, H. (2019). Deletion of clock gene Bmal1 impaired the chondrocyte function due to disruption of the HIF1 $\alpha$ -VEGF signaling pathway. *Cell Cycle*, 18(13), 1473–1489. <https://doi.org/10.1080/15384101.2019.1620572>
- Mahjoob, M., & Stochaj, U. (2021). Curcumin nanoformulations to combat aging-related diseases. *Ageing Research Reviews*, 69, 101364. <https://doi.org/10.1016/j.arr.2021.101364>
- Majumdar, T., Dhar, J., Patel, S., Kondratov, R., & Barik, S. (2017). Circadian transcription factor BMAL1 regulates innate immunity against select RNA viruses. *Innate Immunity*, 23(2), 147–154. <https://doi.org/10.1177/1753425916681075>
- Mannic, T., Meyer, P., Triponez, F., Pusztaszeri, M., le Martelot, G., Mariani, O., Schmitter, D., Sage, D., Philippe, J., & Dibner, C. (2013). Circadian clock characteristics are altered in human thyroid malignant nodules. *The Journal of Clinical Endocrinology and Metabolism*, 98(11), 4446–4456. <https://doi.org/10.1210/jc.2013-2568>
- Marcheva, B., Ramsey, K. M., Buhr, E. D., Kobayashi, Y., Su, H., Ko, C. H., Ivanova, G., Omura, C., Mo, S., Vitaterna, M. H., Lopez, J. P.,



- Philipson, L. H., Bradfield, C. A., Crosby, S. D., JeBailey, L., Wang, X., Takahashi, J. S., & Bass, J. (2010). Disruption of the clock components CLOCK and BMAL1 leads to hypoinsulinaemia and diabetes. *Nature*, 466(7306), 627–631. <https://doi.org/10.1038/nature09253>
- Mas-Bargues, C., Alique, M., Barrús-Ortiz, M. T., Borrás, C., & Rodrigues-Diez, R. (2022). Special issue "Oxidative stress in aging and associated chronic diseases". *Antioxidants*, 11(4), 701. <https://doi.org/10.3390/antiox11040701>
- Matthews, K. A., Xu, W., Gaglioti, A. H., Holt, J. B., Croft, J. B., Mack, D., & McGuire, L. C. (2019). Racial and ethnic estimates of Alzheimer's disease and related dementias in the United States (2015–2060) in adults aged  $\geq 65$  years. *Alzheimer's & Dementia*, 15(1), 17–24. <https://doi.org/10.1016/j.jalz.2018.06.3063>
- Mattis, J., & Sehgal, A. (2016). Circadian rhythms, sleep, and disorders of aging. *Trends in Endocrinology and Metabolism*, 27(4), 192–203. <https://doi.org/10.1016/j.tem.2016.02.003>
- McKee, C. A., Lee, J., Cai, Y., Saito, T., Saido, T., & Musiek, E. S. (2022). Astrocytes deficient in circadian clock gene *Bmal1* show enhanced activation responses to amyloid-beta pathology without changing plaque burden. *Scientific Reports*, 12(1), 1796. <https://doi.org/10.1038/s41598-022-05862-z>
- Mitchell, P., Liew, G., Gopinath, B., & Wong, T. Y. (2018). Age-related macular degeneration. *Lancet*, 392(10153), 1147–1159. [https://doi.org/10.1016/S0140-6736\(18\)31550-2](https://doi.org/10.1016/S0140-6736(18)31550-2)
- Mor, D. E., Sohrabi, S., Kaletsky, R., Keyes, W., Tartici, A., Kalia, V., Miller, G. W., & Murphy, C. T. (2020). Metformin rescues Parkinson's disease phenotypes caused by hyperactive mitochondria. *Proceedings of the National Academy of Sciences of the United States of America*, 117(42), 26438–26447. <https://doi.org/10.1073/pnas.2009838117>
- Musiek, E. S., & Holtzman, D. M. (2016). Mechanisms linking circadian clocks, sleep, and neurodegeneration. *Science*, 354(6315), 1004–1008.
- Musiek, E. S., Lim, M. M., Yang, G., Bauer, A. Q., Qi, L., Lee, Y., Roh, J. H., Ortiz-Gonzalez, X., Dearborn, J. T., Culver, J. P., Herzog, E. D., Hogenesch, J. B., Wozniak, D. F., Dikranian, K., Giasson, B. I., Weaver, D. R., Holtzman, D. M., & Fitzgerald, G. A. (2013). Circadian clock proteins regulate neuronal redox homeostasis and neurodegeneration. *The Journal of Clinical Investigation*, 123(12), 5389–5400. <https://doi.org/10.1172/JCI70317>
- Nakajima, A., & Ohizumi, Y. (2019). Potential benefits of nobiletin, a citrus flavonoid, against Alzheimer's disease and Parkinson's disease. *International Journal of Molecular Sciences*, 20(14), 3380. <https://doi.org/10.3390/ijms20143380>
- Nakata, M., Kumari, P., Kita, R., Katsui, N., Takeuchi, Y., Kawaguchi, T., Yamazaki, T., Zhang, B., Shimba, S., & Yada, T. (2021). Circadian clock component BMAL1 in the paraventricular nucleus regulates glucose metabolism. *Nutrients*, 13(12), 4487. <https://doi.org/10.3390/nu13124487>
- Nakazato, R., Kawabe, K., Yamada, D., Ikeno, S., Mieda, M., Shimba, S., Hinoi, E., Yoneda, Y., & Takarada, T. (2017). Disruption of *Bmal1* impairs blood-brain barrier integrity via pericyte dysfunction. *The Journal of Neuroscience*, 37(42), 10052–10062. <https://doi.org/10.1523/JNEUROSCI.3639-16.2017>
- Nguyen-Ngo, C., Salomon, C., Quak, S., Lai, A., Willcox, J. C., & Lappas, M. (2020). Nobiletin exerts anti-diabetic and anti-inflammatory effects in an in vitro human model and in vivo murine model of gestational diabetes. *Clinical Science*, 134(6), 571–592. <https://doi.org/10.1042/CS20191099>
- Niu, L., Zhang, F., Xu, X., Yang, Y., Li, S., Liu, H., & Le, W. (2021). Chronic sleep deprivation altered the expression of circadian clock genes and aggravated Alzheimer's disease neuropathology. *Brain Pathology*, 32(3), e13028. <https://doi.org/10.1111/bpa.13028>
- Pan, X., Bradfield, C. A., & Hussain, M. M. (2016). Global and hepatocyte-specific ablation of *Bmal1* induces hyperlipidaemia and enhances atherosclerosis. *Nature Communications*, 7, 13011. <https://doi.org/10.1038/ncomms13011>
- Pappa, K. I., Gazouli, M., Anastasiou, E., Iliodromiti, Z., Antsaklis, A., & Anagnou, N. P. (2013). The major circadian pacemaker ARNT-like protein-1 (BMAL1) is associated with susceptibility to gestational diabetes mellitus. *Diabetes Research and Clinical Practice*, 99(2), 151–157. <https://doi.org/10.1016/j.diabres.2012.10.015>
- Peng, X., Fan, R., Xie, L., Shi, X., Dong, K., Zhang, S., Tao, J., Xu, W., Ma, D., Chen, J., & Yang, Y. (2022). A growing link between circadian rhythms, type 2 diabetes mellitus and Alzheimer's disease. *International Journal of Molecular Sciences*, 23(1), 504. <https://doi.org/10.3390/ijms23010504>
- Peschke, E., Wolgast, S., Bazwinsky, I., Pönicke, K., & Muhlbauer, E. (2008). Increased melatonin synthesis in pineal glands of rats in streptozotocin induced type 1 diabetes. *Journal of Pineal Research*, 45(4), 439–448. <https://doi.org/10.1111/j.1600-079X.2008.00612.x>
- Qi, G., Mi, Y., Fan, R., Zhao, B., Ren, B., & Liu, X. (2017). Tea polyphenols ameliorates neural redox imbalance and mitochondrial dysfunction via mechanisms linking the key circadian regular *Bmal1*. *Food and Chemical Toxicology*, 110, 189–199. <https://doi.org/10.1016/j.fct.2017.10.031>
- Qiu, P., Jiang, J., Liu, Z., Cai, Y., Huang, T., Wang, Y., Liu, Q., Nie, Y., Liu, F., Cheng, J., Li, Q., Tang, Y. C., Poo, M. M., Sun, Q., & Chang, H. C. (2019). BMAL1 knockout macaque monkeys display reduced sleep and psychiatric disorders. *National Science Review*, 6(1), 87–100. <https://doi.org/10.1093/nsr/nwz002>
- Quezada-Fernández, P., Trujillo-Quiros, J., Pascoe-González, S., Trujillo-Rangel, W. A., Cardona-Müller, D., Ramos-Becerra, C. G., Barocio-Pantoja, M., Rodríguez-de la Cerda, M., Néri Sánchez-Rodríguez, E., Cardona-Muñoz, E. G., García-Benavides, L., & Grover-Páez, F. (2019). Effect of green tea extract on arterial stiffness, lipid profile and sRAGE in patients with type 2 diabetes mellitus: a randomised, double-blind, placebo-controlled trial. *Int J Food Sci Nutr*, 70(8), 977–985. <https://doi.org/10.1080/09637486.2019>
- Rakshit, K., Hsu, T. W., & Matveyenko, A. V. (2016). *Bmal1* is required for beta cell compensatory expansion, survival and metabolic adaptation to diet-induced obesity in mice. *Diabetologia*, 59(4), 734–743. <https://doi.org/10.1007/s00125-015-3859-2>
- Rakshit, K., & Matveyenko, A. V. (2021). Induction of core circadian clock transcription factor *bmal1* enhances  $\beta$ -cell function and protects against obesity-induced glucose intolerance. *Diabetes*, 70(1), 143–154. <https://doi.org/10.2337/db20-0192>
- Rakshit, K., Qian, J., Gaonkar, K. S., Dhawan, S., Colwell, C. S., & Matveyenko, A. V. (2018). Postnatal ontogenesis of the islet circadian clock plays a contributory role in  $\beta$ -cell maturation process. *Diabetes*, 67(5), 911–922. <https://doi.org/10.2337/db17-0850>
- Ramsey, K. M., Yoshino, J., Brace, C. S., Abrassart, D., Kobayashi, Y., Marcheva, B., Hong, H. K., Chong, J. L., Buhr, E. D., Lee, C., Takahashi, J. S., Imai, S., & Bass, J. (2009). Circadian clock feedback cycle through NAMPT-mediated NAD<sup>+</sup> biosynthesis. *Science*, 324(5927), 651–654. <https://doi.org/10.1126/science.1171641>
- Ray, S., Valekunja, U. K., Stangherlin, A., Howell, S. A., Snijders, A. P., Damodaran, G., & Reddy, A. B. (2020). Circadian rhythms in the absence of the clock gene. *Science*, 367(6479), 800–806. <https://doi.org/10.1126/science.aaw7365>
- Regmi, P., Chaudhary, R., Page, A. J., Hutchison, A. T., Vincent, A. D., Liu, B., & Heilbronn, L. (2021). Early or delayed time-restricted feeding prevents metabolic impact of obesity in mice. *The Journal of Endocrinology*, 248(1), 75–86. <https://doi.org/10.1530/JOE-20-0404>
- Reichenbach, N., Delekate, A., Plescher, M., Schmitt, F., Krauss, S., Blank, N., Halle, A., & Petzold, G. C. (2019). Inhibition of Stat3-mediated astroglial ameliorates pathology in an Alzheimer's disease model. *EMBO Molecular Medicine*, 11(2), e9665. <https://doi.org/10.15252/emmm.201809665>



- Reinke, H., & Asher, G. (2019). Crosstalk between metabolism and circadian clocks. *Nature Reviews Molecular Cell Biology*, 20(4), 227–241. <https://doi.org/10.1038/s41580-018-0096-9>
- Richards, J., & Gumz, M. L. (2013). Mechanism of the circadian clock in physiology. *American Journal of Physiology Regulatory, Integrative and Comparative Physiology*, 304(12), R1053–R1064. <https://doi.org/10.1152/ajpregu.00066.2013>
- Scheltens, P., Blennow, K., Breteler, M. M., de Strooper, B., Frisoni, G. B., Salloway, S., & Van der Flier, W. M. (2016). Alzheimer's disease. *Lancet*, 388(10043), 505–517. [https://doi.org/10.1016/s0140-6736\(15\)01124-1](https://doi.org/10.1016/s0140-6736(15)01124-1)
- Shi, S.-q., Ansari, T. S., McGuinness, O. P., Wasserman, D. H., & Johnson, C. H. (2013). Circadian disruption leads to insulin resistance and obesity. *Current Biology*, 23(5), 372–381. <https://doi.org/10.1016/j.cub.2013.01.048>
- Shih, Y.-H., Wu, S.-Y., Yu, M., Huang, S.-H., Lee, C.-W., Jiang, M.-J., & Kuo, Y.-M. (2018). Hypertension accelerates Alzheimer's disease-related pathologies in pigs and 3xTg mice. *Frontiers in Aging Neuroscience*, 10, 73. <https://doi.org/10.3389/fnagi.2018.00073>
- Song, C., Zhang, Y., Cheng, L., Shi, M., Li, X., Zhang, L., & Zhao, H. (2021). Tea polyphenols ameliorates memory decline in aging model rats by inhibiting brain TLR4/NF- $\kappa$ B inflammatory signaling pathway caused by intestinal flora dysbiosis. *Experimental Gerontology*, 153, 111476. <https://doi.org/10.1016/j.exger.2021.111476>
- Song, H., Moon, M., Choe, H. K., Han, D.-H., Jang, C., Kim, A., Cho, S., Kim, K., & Mook-Jung, I. (2015). A $\beta$ -induced degradation of BMAL1 and CBP leads to circadian rhythm disruption in Alzheimer's disease. *Molecular Neurodegeneration*, 10, 13. <https://doi.org/10.1186/s13024-015-0007-x>
- Song, X., Ma, T., Hu, H., Zhao, M., Bai, H., Wang, X., Liu, L., Li, T., Sheng, X., Xu, X., Zhang, X., & Gao, L. (2021). Chronic Circadian rhythm disturbance accelerates knee cartilage degeneration in rats accompanied by the activation of the canonical Wnt/ $\beta$ -catenin signaling pathway. *Frontiers in Pharmacology*, 12, 760988. <https://doi.org/10.3389/fphar.2021.760988>
- Stahl, A. (2020). The diagnosis and treatment of age-related macular degeneration. *Deutsches Arzteblatt International*, 117(29–30), 513–520. <https://doi.org/10.3238/arztebl.2020.0513>
- Stenvers, D. J., Scheer, F. A. J. L., Schrauwen, P., la Fleur, S. E., & Kalsbeek, A. (2019). Circadian clocks and insulin resistance. *Nature Reviews Endocrinology*, 15(2), 75–89. <https://doi.org/10.1038/s41574-018-0122-1>
- Stokes, K., Nunes, M., Trombley, C., Flôres, D. E. F. L., Wu, G., Taleb, Z., Alkhateeb, A., Banskota, S., Harris, C., Love, O. P., Khan, W. I., Rueda, L., Hogenesch, J. B., & Karpowicz, P. (2021). The circadian clock gene, *Bmal1*, regulates intestinal stem cell signaling and represses tumor initiation. *Cellular and Molecular Gastroenterology and Hepatology*, 12(5), 1847–1872.e0. <https://doi.org/10.1016/j.jcmgh.2021.08.001>
- Su, W., Guo, Z., Randall, D. C., Cassis, L., Brown, D. R., & Gong, M. C. (2008). Hypertension and disrupted blood pressure circadian rhythm in type 2 diabetic db/db mice. *American Journal of Physiology - Heart and Circulatory Physiology*, 295(4), H1634–H1641. <https://doi.org/10.1152/ajpheart.00257.2008>
- Tan, D.-X., Manchester, L. C., Esteban-Zubero, E., Zhou, Z., & Reiter, R. J. (2015). Melatonin as a potent and inducible endogenous antioxidant: Synthesis and metabolism. *Molecules*, 20(10), 18886–18906. <https://doi.org/10.3390/molecules201018886>
- Tharmalingam, S., Khurana, S., Murray, A., Lamothe, J., & Tai, T. C. (2020). Whole transcriptome analysis of adrenal glands from prenatal glucocorticoid programmed hypertensive rodents. *Scientific Reports*, 10(1), 18755. <https://doi.org/10.1038/s41598-020-75652-y>
- Vijayan, M., & Reddy, P. H. (2016). Stroke, vascular dementia, and Alzheimer's disease: Molecular links. *Journal of Alzheimer's Disease*, 54(2), 427–443. <https://doi.org/10.3233/JAD-160527>
- Wang, J., Li, S., Li, X., Li, B., Li, Y., Xia, K., Yang, Y., Aman, S., Wang, M., & Wu, H. (2019). Circadian protein BMAL1 promotes breast cancer cell invasion and metastasis by up-regulating matrix metalloproteinase9 expression. *Cancer Cell International*, 19, 182. <https://doi.org/10.1186/s12935-019-0902-2>
- Wang, J.-H., Wu, Y.-J., Tee, B. L., & Lo, R. Y. (2018). Medical comorbidity in Alzheimer's disease: A nested case-control study. *Journal of Alzheimer's Disease*, 63(2), 773–781. <https://doi.org/10.3233/JAD-170786>
- Wang, L., Zhao, J., Wang, C.-T., Hou, X.-H., Ning, N., Sun, C., Guo, S., Yuan, Y., Li, L., Hölscher, C., & Wang, X. H. (2020). D-Ser2-oxyntomodulin ameliorated A $\beta$ 31–35-induced circadian rhythm disorder in mice. *CNS Neuroscience & Therapeutics*, 26(3), 343–354. <https://doi.org/10.1111/cns.13211>
- Wang, X.-L., Koojiman, S., Gao, Y., Tzeplaef, L., Cosquer, B., Milanova, I., Wolff, S. E. C., Korpel, N., Champy, M. F., Petit-Demoulière, B., Goncalves da Cruz, I., Sorg-Guss, T., Rensen, P. C. N., Cassel, J. C., Kalsbeek, A., Boutillier, A. L., & Yi, C. X. (2021). Microglia-specific knock-down of *Bmal1* improves memory and protects mice from high fat diet-induced obesity. *Molecular Psychiatry*, 26(11), 6336–6349. <https://doi.org/10.1038/s41380-021-01169-z>
- Wang, Y., Lv, D., Liu, W., Li, S., Chen, J., Shen, Y., Wang, F., Hu, L. F., & Liu, C. F. (2018). Disruption of the circadian clock alters antioxidative defense via the SIRT1-BMAL1 pathway in 6-OHDA-induced models of Parkinson's disease. *Oxidative Medicine and Cellular Longevity*, 2018, 4854732. <https://doi.org/10.1155/2018/4854732>
- Wang, Z.-J., Han, Y.-F., Zhao, F., Yang, G.-Z., Yuan, L., Cai, H.-Y., Yang, J. T., Holscher, C., Qi, J. S., & Wu, M. N. (2020). A dual GLP-1 and Gcg receptor agonist rescues spatial memory and synaptic plasticity in APP/PS1 transgenic mice. *Hormones and Behavior*, 118, 104640. <https://doi.org/10.1016/j.yhbeh.2019.104640>
- Welz, P.-S., Zinna, V. M., Symeonidi, A., Koronowski, K. B., Kinouchi, K., Smith, J. G., Guillén, I. M., Castellanos, A., Furrow, S., Aragón, F., Crainiciuc, G., Prats, N., Caballero, J. M., Hidalgo, A., Sassone-Corsi, P., & Benitah, S. A. (2019). BMAL1-driven tissue clocks respond independently to light to maintain homeostasis. *Cell*, 177(6), 1436–1447.e12. <https://doi.org/10.1016/j.cell.2019.05.009>
- Wen, L.-Y., Wan, L., Lai, J.-N., Chen, C. S., Chen, J. J.-Y., Wu, M.-Y., Hu, K. C., Chiu, L. T., Tien, P. T., & Lin, H. J. (2021). Increased risk of Alzheimer's disease among patients with age-related macular degeneration: A nationwide population-based study. *PLoS One*, 16(5), e0250440. <https://doi.org/10.1371/journal.pone.0250440>
- Woon, P. Y., Kaisaki, P. J., Bragança, J., Bihoreau, M.-T., Levy, J. C., Farrall, M., & Gauguier, D. (2007). Aryl hydrocarbon receptor nuclear translocator-like (*BMAL1*) is associated with susceptibility to hypertension and type 2 diabetes. *Proceedings of the National Academy of Sciences of the United States of America*, 104(36), 14412–14417.
- Wu, X., Chen, L., Zeb, F., Li, C., Jiang, P., Chen, A., Xu, C., Haq, I. U., & Feng, Q. (2019). Clock-*Bmal1* mediates MMP9 induction in acrolein-promoted atherosclerosis associated with gut microbiota regulation. *Environ Pollut*, 252(Pt B), 1455–1463. <https://doi.org/10.1016/j.envpol.2019.06.042>
- Wu, H., Feng, K., Zhang, C., Zhang, H., Zhang, J., Hua, Y., Dong, Z., Zhu, Y., Yang, S., & Ma, C. (2021). Metformin attenuates atherosclerosis and plaque vulnerability by upregulating KLF2-mediated autophagy in apoE mice. *Biochemical and Biophysical Research Communications*, 557, 334–341. <https://doi.org/10.1016/j.bbrc.2021.04.029>
- Xie, B., Shi, X., Xing, Y., & Tang, Y. (2020). Association between atherosclerosis and Alzheimer's disease: A systematic review and meta-analysis. *Brain and Behavior*, 10(4), e01601. <https://doi.org/10.1002/brb3.1601>
- Xie, M., Tang, Q., Nie, J., Zhang, C., Zhou, X., Yu, S., Sun, J., Cheng, X., Dong, N., Hu, Y., & Chen, L. (2020). BMAL1-downregulation aggravates -induced atherosclerosis by encouraging oxidative stress. *Circulation Research*, 126(6), e15–e29. <https://doi.org/10.1161/CIRCRESAHA.119.315502>
- Xu, L., Liu, Y., Cheng, Q., Shen, Y., Yuan, Y., Jiang, X., Li, X., Guo, D., Jiang, J., & Lin, C. (2021). *Bmal1* downregulation worsens critical



- limb ischemia by promoting inflammation and impairing angiogenesis. *Frontiers in Cardiovascular Medicine*, 8, 712903. <https://doi.org/10.3389/fcvm.2021.712903>
- Yang, S., Wan, Y., Wu, N., Song, L., Liu, Z., Zhao, J., Liu, Y., Liu, Z., & Gan, J. (2021). L-3,4-dihydroxyphenylalanine recovers circadian rhythm disturbances in the rat models of Parkinson's disease by regulating the D1R-ERK1/2-mTOR pathway. *Frontiers in Aging Neuroscience*, 13, 719885. <https://doi.org/10.3389/fnagi.2021.719885>
- Yang, W., Kang, X., Liu, J., Li, H., Ma, Z., Jin, X., Qian, Z., Xie, T., Qin, N., Feng, D., Pan, W., Chen, Q., Sun, H., & Wu, S. (2016). Clock gene *Bmal1* modulates human cartilage gene expression by cross-talk with Sirt1. *Endocrinology*, 157(8), 3096–3107. <https://doi.org/10.1210/en.2015-2042>
- Yaribeygi, H., Sathyapalan, T., Atkin, S. L., & Sahebkar, A. (2020). Molecular mechanisms linking oxidative stress and diabetes mellitus. *Oxidative Medicine and Cellular Longevity*, 2020, 8609213. <https://doi.org/10.1155/2020/8609213>
- Ye, L., Wu, H., & Xu, W. (2020). Deletion of *Bmal1* impairs pancreatic  $\beta$ -cell function via mitochondrial signaling pathway. *BioMed Research International*, 2020, 9803024. <https://doi.org/10.1155/2020/9803024>
- Yoo, I. D., Park, M. W., Cha, H. W., Yoon, S., Boonpraman, N., Yi, S. S., & Moon, J.-S. (2020). Elevated CLOCK and BMAL1 contribute to the impairment of aerobic glycolysis from astrocytes in Alzheimer's disease. *International Journal of Molecular Sciences*, 21(21), 7862. <https://doi.org/10.3390/ijms21217862>
- Yu, R., Tian, L., Ding, Y., Gao, Y., Li, D., & Tang, Y. (2019). Correlation between inflammatory markers and impaired circadian clock gene expression in type 2 diabetes mellitus. *Diabetes Research and Clinical Practice*, 156, 107831. <https://doi.org/10.1016/j.diabres.2019.107831>
- Yuan, Y., Li, C., Guo, S., Sun, C., Ning, N., Hao, H., Wang, X., Bian, Y., Liu, H., & Wang, X. (2021). Adiponectin improves amyloid- $\beta$  31-35-induced circadian rhythm disorder in mice. *Journal of Cellular and Molecular Medicine*, 25(20), 9851–9862. <https://doi.org/10.1111/jcmm.16932>
- Zhang, G., Meng, L., Wang, Z., Peng, Q., Chen, G., Xiong, J., & Zhang, Z. (2022). Islet amyloid polypeptide cross-seeds tau and drives the neurofibrillary pathology in Alzheimer's disease. *Molecular Neurodegeneration*, 17(1), 12. <https://doi.org/10.1186/s13024-022-00518-y>
- Zhang, W., & Luo, P. (2020). Myocardial infarction predisposes neurodegenerative diseases. *Journal of Alzheimer's Disease*, 74(2), 579–587. <https://doi.org/10.3233/JAD-191225>
- Zhang, Y., & Song, W. (2017). Islet amyloid polypeptide: Another key molecule in Alzheimer's pathogenesis? *Progress in Neurobiology*, 153, 100–120. <https://doi.org/10.1016/j.pneurobio.2017.03.001>
- Zhao, R., Gao, X., Zhang, T., & Li, X. (2016). Effects of polysaccharide on type 2 diabetes mellitus rats by regulating biological rhythms. *Iranian Journal of Basic Medical Sciences*, 19(9), 1024–1030.
- Zhu, M., Tang, H., Tang, X., Ma, X., Guo, D., & Chen, F. (2018). BMAL1 suppresses ROS-induced endothelial-to-mesenchymal transition and atherosclerosis plaque progression via BMP signaling. *American Journal of Translational Research*, 10(10), 3150–3161.
- Zhu, P., Hamlisch, N. X., Thakkar, A. V., Steffek, A. W. T., Rendleman, E. J., Khan, N. H., Waldeck, N. J., DeVilbiss, A., Martin-Sandoval, M. S., Mathews, T. P., Chandel, N. S., & Peek, C. B. (2022). BMAL1 drives muscle repair through control of hypoxic NAD regeneration in satellite cells. *Genes & Development*, 36, 149–166. <https://doi.org/10.1101/gad.349066.121>
- Zhuang, W., Yue, L., Dang, X., Chen, F., Gong, Y., Lin, X., & Luo, Y. (2019). Rosenroot (*Rhodiola*): Potential applications in aging-related diseases. *Aging and Disease*, 10(1), 134–146. <https://doi.org/10.14336/AD.2018.0511>

**How to cite this article:** Fan, R., Peng, X., Xie, L., Dong, K., Ma, D., Xu, W., Shi, X., Zhang, S., Chen, J., Yu, X., & Yang, Y. (2022). Importance of *Bmal1* in Alzheimer's disease and associated aging-related diseases: Mechanisms and interventions. *Aging Cell*, 21, e13704. <https://doi.org/10.1111/acer.13704>