



www.bioinformation.net **Volume 15(11)** 

### **Research Article**

# Molecular docking analysis of *timepidium* with Acetylcholine and *lumacaftor* with GABA(A) activator

### Warda Ali, Nisar A. Shar\*

Department of Biomedical Engineering, NED University of Engineering & Technology Karachi, Pakistan; Nisar A. Shar – E-mail: nisarshar@neduet.edu.pk; \*Corresponding author

Received November 20, 2019; Revised December 10, 2019; Accepted December 11, 2019; Published December 16, 2019

DOI: 10.6026/97320630015824

### Abstract:

Epilepsy is a chronic disorder characterized by disturbed tissue related molecular activity within the brain irrespective of age. The cause is very difficult to understand towards a suitable treatment. However, its symptoms like seizures are treated and suppressed by known medications. Moreover, the condition is linked with neuro-transmitters such as GABA (gamma amino butyric acid) and acetylcholine. Therefore, it is of interest to design and develop inhibitors for these targets. Hence, we describe the molecular binding features of timepidium with acetylcholine and lumacaftor with GABA(A) activator using molecular docking based geometric optimization and screening analysis for further consideration.

Keywords: Epilepsy, repurposing, docking, scoring, electrostatic interaction, seizures.

### Background:

Epilepsy is a neurological condition in which a person experiences recurrent and unprovoked seizures within a day [1]. There are different causes of Epilepsy. The most common cause of Epilepsy is disturbance in the activity of Acetylcholine and GABA. In normal state, GABA act as inhibitory neurotransmitter and Acetylcholine is excitatory neurotransmitter. They regulate cortical function of brain including attention, learning, memory, sleep-wake alternation, and are implicated in neurodegenerative diseases [2]. It implies that both GABA and Acetylcholine are important for normal cognitive functions [3]. These neurotransmitters also play significant role in sleep deprivation [4], direct coding in retina [5] and in age related hearing loss [6]. In epileptic patients, the activity of GABA is suppressed whereas the activity of Acetylcholine is greatly increased [7]. However, in case of schizophrenia, the levels of GABA are increased. It has been found that the compounds derived from plants are used to inhibit the level of GABA [8]. Different drugs are known to activate the selected neurotransmitters like Emamectin and Ivermectin. Emamectin directly activates acetylcholine and GABA receptors whereas Ivermectin activates GABA (A) receptor only **[9].** Herbal compounds are also known to activate GABA receptor such as Rosmarinic Acid and Kaempferol **[10].** 

The treatment of epilepsy after occurrence of first seizure is a controversial issue because the underlying mechanisms of brain damage and processes that lead to the development of epileptic conditions are still unknown. However, many successful antiepileptic drugs AED's have been developed to control seizures; which is one of the most common conditions of epilepsy. These drugs mainly include brivaracetam [11], topiramate [12], phensuximide [13] and fingolimod [14]. AED's stop seizures in approximately 70% of people by controlling chemical activity in brain but they do not cure epilepsy. A study was conducted to check the drug resistance in epileptic patients. If drugs are not effective then seizure activity may be treated either by ketogenic diet [15] or by surgery [16]. It investigated the use of complementary and alternative medicine (CAM) among epileptic patients. It also analyzed the impact of CAM on AED's. The results

ISSN 0973-2063 (online) 0973-8894 (print) Bioinformation 15(11): 832-837 (2019)



showed that there is less association between AED's and use of CAM [17].

### Methodology:

### Protein target and ligand structures:

The first step was extraction of three-dimensional structures of drugs and proteins. The 3D structures of GABA activators and the program database files (PDB) of Acetylcholine inhibitors were downloaded from Drug Bank Database for docking. The protein ID's for the chosen proteins were obtained from Uniprot. These IDs were then used as an input to download PDB structure of Acetylcholine and GABA receptors from protein data bank. Different receptor chains of Acetylcholine and GABA were analyzed. However, on the basis of their functional properties, six receptor chains of Acetylcholine and four chains of GABA were selected. 45 drugs were randomly selected for Acetylcholine while 47 were selected for GABA. Acetylcholine and GABA recognized some of these drugs while others were unrecognized.

#### **Electrostatic interactions calculation:**

The next step was calculation of electrostatic interactions. SCORE, is used to calculate the electrostatic interactions between the protein as receptor/target and drug. These electrostatic interactions were calculated between randomly selected recognized and unrecognized drugs and target proteins i.e., extracted protein chains of Acetylcholine and GABA. For Acetylcholine, one drug is interacted with three chains (out of six chains) whereas in case of GABA three drugs are interacted with three chains (out of four chains). These interactions are shown in **Table 1**.

### Molecular docking analysis:

Selecting the highly negative interactions between receptor and ligand using publically available Docking Server and Hex software performed the molecular docking. Chimera was used to visualize results of docking between protein chains and drugs. When drug binds to its target, it releases binding free energy. The binding free energies of the ligand and proteins were computed by using the compute energy tool of the Swiss PDB viewer. The docking server was then used to validate the post docking results. Motifs and domains of the receptor protein were then obtained using SCANPROSITE and ProDom. Examining motifs and domains of the considered protein then did a comparison of the active sites. The residues that lie between the sequence of the motifs and chains were considered as best docked results.



**Figure 1:** The results of docking between extracted proteins and their targets. Figure 1(a) depicts the Solid model result of Docked Acetylcholine Receptor Alpha unit 2 with Timepidium drug along with an interactive ribbon docked model showing hydrogen bonds. Figure 1(b) depicts the Solid model result of docked Gamma Amino butyric Acid (GABA) Receptor (4MQE) with drug Lumacaftor from CHIMERA. Figure 1(c) further shows two hydrogen bonds as interactive ribbon docked models. It is evident that the bonds in 1(a) are formed between Glutamine residue of Chain-A and Oxygen atoms of the drug with a distance of 2.264 Angstrom whereas in 1(c) the bonds with Aspartate and Lysine residues of Chain A with a distance of 2.255 Angstrom and 2.282 Angstrom respectively. All figures follow a common legend, blue represents the drug chains, red represents Chain-A of Acetylcholine/Chain B of GABA receptor and the orange red represent Chain E of Acetylcholine Receptor/Chain-A of GABA receptor.



Table 1: Total interactions of recognized and not recognized drugs with Acetylcholine and GABA receptor chain to show the electrostatic interactions among drugs and receptor chains

|   | Total                 | Negative Interaction Value Distribution |                                       |   |  |  |   |   |   |
|---|-----------------------|---|---------------------------------------|---|--|--|---|---|---|
|   | Number<br>of<br>Drugs | Number of<br>Positive<br>Interactions   | Number of<br>Negative<br>Interactions | Drug<br>interacting<br>with all six<br>chains | Drug<br>Interacting<br>with five<br>Chains | Drug<br>Interacting<br>with four<br>Chains | Drug<br>Interacting<br>with three<br>Chains | Drug<br>Interacting<br>with two<br>Chains | Drug<br>Interacting<br>with one<br>Chains |
| Drugs<br>Recognized<br>By acetyl<br>choline     | 24                    | 91                                      | 53                                    | 3   | 2  | 2  | 1   | 4   | 6   |
| Drugs Not<br>Recognized<br>By acetyl<br>choline | 21                    | 107                                     | 19                                    | 0   | 0  | 0  | 1   | 6   | 4   |
| Drugs<br>Recognized<br>By GABA                  | 20                    | 52                                      | 28                                    | -   | -  | 0  | 2   | 7   | 8   |
| Drugs Not<br>Recognized<br>By GABA              | 27                    | 66                                      | 42                                    | -   | -  | 3  | 3   | 9   | 3   |

Table 2: Drugs docked with Acetylcholine and GABA receptor chains and their binding energy values derived after scoring.

Selected and Docked drugs for Acetyl Choline Chains

|               |               | Distance                  | Electrostatic      |
|---------------|---------------|---------------------------|--------------------|
|               |               | Diugs                     | Interactions       |
|               | CHRNA 2. 5FJV | Timepidium                | -113.6015 Kcal/mol |
|               | CHRNA4.6CN    | Timepidium                | -254.0600 Kcal/mol |
|               | CHRNA.2LLY    | Timepidium                | -198.3092 Kcal/mol |
| Acotylcholino | CHRNA.2LLY    | Emetonium iodide          | -132.9165 Kcal/mol |
| Chaine        | CHRNB5.KXI    | Imipramine oxide          | -216.1342 Kcal/mol |
| Chanis        | CHRNA4.6CN    | Bifemelane                | -17.4069 Kcal/mol  |
|               | CHRNB2.KSR    | 3,4-Dihydroxybenzoic Acid | -158.9715 Kcal/mol |
| GABA Chain    | CHRNA.2LLY    | Tretamine                 | -114.1014 Kcal/mol |
|               | 4MQE          | Dactinomycin              | -62.2064 Kcal/mol  |
|               | 4MQE          | Lumacaftor                | -122.4524 Kcal/mol |
|               | 4MQF          | Dalfopristin              | -89.5999 Kcal/mol  |

### Table 3: Hydrogen and polar bonds formed between the Acetylcholine receptor and Timepidium drug and GABA receptor and Lumacaftor drug.

| Interaction                                | Hydrogen Bond | Polar Bond   |
|--|---------------|--------------|
|  | N1 - GLU328   | O3 - SER84   |
| Acetylcholine receptor and Timepidium Drug |               | N1 - LYS332  |
|  |               | O1 - LYS332  |
|  | O9 - SER 84   | O7 - ASP81   |
|  | O8 - ASN 323  | N4 - SER84   |
| CARA recorder and Lumacofter Drug          |               | N1 - ASN323  |
| GABA receptor and Lumacattor Drug          |               | H1 - ASN323  |
|  |               | N3 - ASN 323 |
|  |               | N6 - LYS332  |

| Table 4. Binding free Energy value computation for protein-figand interactions. |           |       |        |         |          |            |               |              |
|---|-----------|-------|--------|---------|----------|------------|---------------|--------------|
|   | Residue   | bonds | Angles | Torsion | Improper | Non bonded | Electrostatic | Total KJ/mol |
| 4MQE  | Asp A 100 | 2.267 | 4.954  | 4.939   | 0.522    | -37.12     | -0.03         | -32.469      |
| 4MQE  | Lys A 110 | 5.933 | 5.013  | 7.861   | 0.008    | -48.26     | -5.97         | -35.405      |
| 5FJV  | Glu A 128 | 3.592 | 4.308  | 11.824  | 0.059    | -6.42      | 0.57          | 13.932       |
| 5FJV  | Glu B 128 | 4.401 | 5.236  | 8.342   | 1.663    | -31.14     | 1.72          | -9.774       |



#### Table 5: Comparison of target sites of acetylcholine and GABA receptor proteins through motifs and domains by Scan Prosite and ProDom web server

| Examine Target Sites Of Protein |                       |  |   |  |  |  |
|---------------------------------|-----------------------|--|---|--|--|--|
| Motifs                          | Domains               | 2D Drug Protein Docking Plot<br>(Predicted active sites) | Active Sites From Literature Review<br>(Uniprot 2019) |  |  |  |
| 6CNK                            |                       |  |   |  |  |  |
| 155 to 169                      | 265 to 524            | 122,178,179,219,222,226                                  | Not available   |  |  |  |
| 5FJV                            |                       |  |   |  |  |  |
| 161 to 175                      | 62 to 240, 243 to 620 | 84,136,141,146,147,148,212,332,328                       | Not available   |  |  |  |
| 5KXI                            |                       |  |   |  |  |  |
| 133 to 144                      | 36 to 214, 217 to 344 | 136.138.147.148.212                                      | 84, 136, 144  |  |  |  |
| 135 to 149                      | 00 10 211, 21, 10 011 | 100,100,111,110,212                                      | 01,100,111  |  |  |  |
| 2LLY                            |                       |  |   |  |  |  |
| No Hits                         | 7 to 144              | Absent   | 84, 136, 144  |  |  |  |
| 2KSR                            |                       |  |   |  |  |  |
| No Hits                         | No Hit                | Absent   | 84, 136, 144  |  |  |  |
| 4MQE                            |                       |  |   |  |  |  |
| No Hits                         | 37 to 230, 260 to 423 | 81,84,319,322,323,324,328,332,354                        | 247, 270, 287, 367, 395, 466                          |  |  |  |
| 4MQF                            |                       |  |   |  |  |  |
| No Hits                         | 37 to 230, 260 to 423 | 81,84,324,328,332  | 247, 270, 287, 367, 395, 466                          |  |  |  |

#### **Results and Discussion:**

The results of scoring show positive and negative electrostatic interactions between drugs and their targets. The negative electrostatic interaction indicates more possibility of drug binding with protein. Table 2 indicates that 21 drugs, which are not recognized for Acetylcholine, interacted with all chains of Acetylcholine. It was found that out of these 21 drugs, the bestinteracted drug is Timepidium, which gives highly negative electrostatic interaction. However, in case of GABA, 27 drugs that were not recognized by GABA interacted with the chains of GABA. Only 3 drugs show interactions with 3 chains of GABA. The drug Lumacaftor was selected from them as it gave highly negative interaction with the chains of GABA. Hydrogen bonds are weak interactions and important for stabilizing the protein structure in open conformational environment with ligand [18]. Among all highly negative electrostatic interaction of drugs and proteins, hydrogen bonds were observed only in two selected drugs i.e., Timepidium and Lumacaftor.

The results of docking are shown in **Figure 1.** In a number of docking results, *Timepidium* has produced the best result of docking with Acetylcholine receptor 5FJV instead of 6CN. Figure 2a shows that there is only one hydrogen bond between Glutamine residue of Chain A of 5FJV and Oxygen atom of the drug. The distance of this hydrogen bond was found to be 2.264 Angstrom. It indicates best interaction among all docked results. The drug *Lumacaftor* is found to be well docked with the binding site GABA receptor chain (4MQE). It was found that this interaction was strong forming two hydrogen bonds. The distance of one hydrogen bond between Aspartate residue of Chain A and Oxygen atom of the drug was 2.255 Angstrom whereas other hydrogen

bond between Lysine of Chain A and Oxygen atom of Drug had a distance of 2.282 Angstrom.

The results of docking indicate maximum hydrogen and polar bonds between acetylcholine chain with *Timepidium* and GABA with *Lumacaftor*. The hydrogen and polar bonds formed between the receptor-ligand complexes of Acetycholine receptor and *Timepidium* are shown in **Table 3**. The different types of bond linkages indicate the best-docked results of GABA receptor and Lumacaftor. Table 3 also gives Hydrogen and polar bonds between the oxygen atom of *Lumacaftor* and the GABA chains. Thus, the presence of hydrogen and polar bonds validate the acquired results. **Table 4** is showing binding free energies of drug and their targets. The negative value of the binding energy of the proteinligand complex is preferred for binding of ligand with its desired protein. It was observed that 4MQE gave highly negative binding free energy with Lysine A whereas 5FJV showed negative interaction with Glutamine B.

**Table 5** indicates the predicted binding sites of the chains of the studied proteins. Comparing predicted binding sites of their Motifs and Domains then checked the presence of active sites in Acetylcholine and GABA receptor binding sites. The results indicate that all the binding sites of Acetylcholine receptor chain (5FJV) and GABA receptor chain (4MQE) are present in their corresponding Domains. However, in other chains of GABA and Acetylcholine, the binding sites were not matched in motifs and domains.

### **Conclusion:**

Epilepsy is known to be linked with neuro-transmitters such as GABA (gamma amino butyric acid) and acetylcholine. Therefore, it



is of interest to design and develop inhibitors for these targets. However, it is known that lumacaftor has been used to treat Cystic fibrosis (CF) **[19]** and timepidium bromide to treat abdominal diseases **[20]**. Hence, it is of importance to evaluate and describe the molecular binding features of timepidium with acetylcholine and lumacaftor with the GABA(A) activator using molecular docking based geometric optimization and screening analysis for further consideration in this context.

### Acknowledgements:

NAS and WA acknowledge funding from Higher Education Commission (HEC) Pakistan and Ministry of Planning Development and Reforms under National Center in Big Data and Cloud computing at Exascale Open Data Analytics Lab (Genomics Lab) NED University of Engineering & Technology.

### **References:**

- [1] Fisher RS & Bonner AM. *The Neurodiagnostic Journal*. 2018 58:1 [PMID: 29562876]
- [2] Takács VT *et al. Nature Commuication.* 2018 9:2848 [PMID: 30030438].
- [3] Granger AJ *et al. Neuropharmacology.* 2016 **100:**40-46 [PMID: 26220313].
- [4] Toossi H *et al. Brain Structure and Function.* 2017 **222**:3163 [PMID: 28299422].
- [5] Sethuramanujam S *et al. Neuron.* 2016 **90:1**243. [PMID: 27238865].

- [6] Tang X et al. Neuroscience. 2014 259:184. [PMID: 24316061].
- [7] Hsien Yi Chen *et al. British journal of clinical pharmacology.* 2016 81:412. [PMID: 26174744]
- [8] Mohammed Marunnan Sahila *et al. Bioinformation.* 2015. 280. [PMID: 26229288].
- [9] Xiaojun Xu et al. Biochemical and biophysical research communications. 2016 473:795. [PMID: 27049309]
- [10] Bresnahan R *et al. The cochrance database of systematic reviews.*2019 **10:**CD004154. [PMID: 31638272].
- [11] Abhishek N et al. Journal of Applied Pharmaceutical Science. 2018 38.
- [12] Bresnahan R *et al. The cochrance database of systematic reviews.* 2019 [PMID: 31642054].
- [13] D'Andrea Meira *et al. Frontiers in neuroscience.* 2019 [PMID: 30760973].
- [14] Farrukh MJ *et al. Patient Preference and Adherence*. 2018 2111. [PMC6188960].
- [15] Wang S *et al. Biosci Rep.* 2019 **39:**BSR20191247 [PMID: 31427480].
- [16] Pitsch J et al. Mol Neurobiol. 2019 56:1825 [PMID: 29934763].
- [17] Corpet Fet al. Nucleic Acids Research. 1998 26:323 [PMID: 15608179].
- [18] Patil R, et al. Plos One 2010 5: e12029. [PMID: 20808434]
- [19] Loukou I *et al. Journal of cystic fibrosis.* 2019 1569. [PMID: 31676345]
- [20] Irié T *et al. Pathology international.* 2004 54:850 [PMID: 15533228]

### Edited by P Kangueane

Citation: Ali & Shar, Bioinformation 15(11): 832-837 (2019)

License statement: This is an Open Access article which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly credited. This is distributed under the terms of the Creative Commons Attribution License



Agro Informatics Society BIOMEDICAL Biomedical Informatics Society since 2005 BIOINFORMATION Discovery at the interface of physical and biological sciences indexed in Pub Med EBSCO EMERGING WEB OF SCIENCE SOURCES CITATION Web of Science Group COI Crossref ResearchGate publons