## Vaccine 36 (2018) 7011-7016

Contents lists available at ScienceDirect

## Vaccine

journal homepage: www.elsevier.com/locate/vaccine

# Memory inflation following adenoviral vaccination depends on IL-21

Claire L. Gordon<sup>2</sup>, Claire L. Hutchings<sup>2</sup>, Andrew J. Highton, Julia M. Colston, Nicholas M. Provine, Paul Klenerman<sup>1,\*</sup>

Peter Medawar Building for Pathogen Research, University of Oxford, Oxford OX2 3SY, UK

#### ARTICLE INFO

Article history: Received 1 June 2018 Received in revised form 7 September 2018 Accepted 25 September 2018 Available online 29 September 2018

Keywords: Adenovirus vectors Memory inflation Memory T cells

## ABSTRACT

Cytomegalovirus (CMV) and non-replicating adenoviral vectors can induce expanded, sustained effectormemory CD8<sup>+</sup> T-cell responses, termed "memory inflation". During murine CMV (MCMV) infection, CD4<sup>+</sup> Tcells maintain inflationary virus-specific CD8<sup>+</sup> T-cell responses via IL-2 but not IL-21. Adenovirus vector vaccination can induce phenotypically, functionally and transcriptionally similar inflationary responses, but it is not known how IL-21 influences the inflating memory response to adenoviral vaccination. Here, we show that IL-21 is an absolute requirement for induction and maintenance of vaccine-derived inflationary CD8<sup>+</sup> T-cell responses. These data indicate that the induction pathway of inflationary Ad-LacZ T-cells is distinct from inflationary MCMV-specific T-cells and is highly dependent on IL-21. Our observations highlight a fundamental difference in the mechanism by which adenovirus vectors and MCMV drive inflationary T-cell responses.

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## 1. Introduction

The success of novel T-cell vaccine strategies depends on the ability of the vaccine to elicit large numbers of long-lived effector memory CD8<sup>+</sup> T-cells. Cytomegalovirus (CMV) and non-replicating adenoviral vectors can induce an expanded, sustained effector-memory CD8<sup>+</sup> T-cell response to specific epitopes, termed "memory inflation", which develop in parallel with conventional non-inflating (contracting) central memory responses to many epitopes [1]. Inflationary T-cells maintain effector function, ability to proliferate and lack features of T-cell exhaustion [2], leading to interest in these cells in vaccine development. Since adenoviral vectors form the basis of many novel vaccines in infectious diseases and cancer this has high translational potential [3].

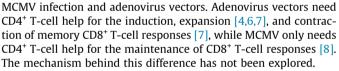
The recombinant replication-deficient human adenovirus 5 (HuAd5) vector expressing  $\beta$ -galactosidase (Ad-lacZ) induces two distinct pathways for memory – a typical contracting response to one epitope (I8V) and an inflating response to a second epitope (D8V), which is analogous to conventional (M45) and inflating (M38) responses to MCMV infection [4,5]. While inflationary responses induced by MCMV and Ad-lacZ are phenotypically and functionally similar, the requirement of CD4<sup>+</sup> T-cell help for the induction and maintenance of T-cell memory differs between

\* Corresponding author at: Peter Medawar Building for Pathogen Research, South Parks Road, Oxford OX1 3SY, UK.

E-mail address: paul.klenerman@ndm.ox.ac.uk (P. Klenerman).

https://doi.org/10.1016/j.vaccine.2018.09.061

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CD4<sup>+</sup> T-cells produce IL-2 and IL-21 and maintenance of antigen-specific CD8<sup>+</sup> T-cells in persistent viral infections (i.e. LCMV, HIV-1) has been shown to depend on these cytokines [9–11]. Studies of MCMV infection have indicated that IL-2 but not IL-21 maintains inflating virus-specific CD8<sup>+</sup> T-cell responses [8]. In comparison, studies have indicated that non-inflating adenovirus-derived CD8<sup>+</sup> T-cell memory responses are dependent on CD4<sup>+</sup> T-cell help, likely through the production of IL-2 and IL-21 [12,13], however, the dependence of adenovirus-derived inflating memory responses on IL-21 has not been examined.

Here, we show that IL-21 is absolutely required for vaccinederived inflationary CD8<sup>+</sup> T-cell responses using a mouse model of Ad-lacZ intravenous immunization. Our observations highlight a fundamental difference between adenovirus vectors and MCMV, especially around the time of memory induction.

## 2. Materials and methods

## 2.1. Mice

Experiments were performed according to UK Home Office regulations (PPL 30/3293, 30/2744, 30/2414) after approval by the University of Oxford ethical review board.  $Il21^{-/-}$  transgenic







<sup>&</sup>lt;sup>1</sup> Lead Contact.

<sup>&</sup>lt;sup>2</sup> These authors contributed equally.

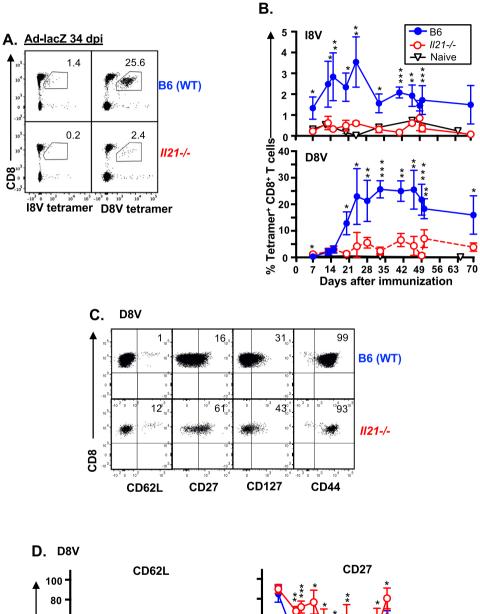
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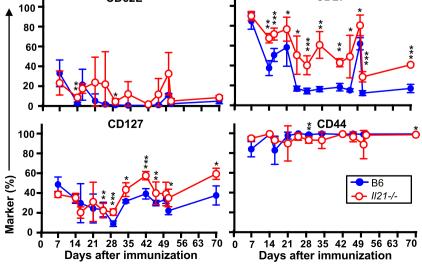
knock-in mice (Jean Langhorne, National Institute for Medical Research) and C57BL/6 WT mice (Envigo, UK) were kept under conventional conditions in individually ventilated cages and fed normal chow diet.

CD8

#### 2.2. MCMV and adenoviral vector

MCMV (Strain Smith; ATCC: VR194; from U.H. Koszinoswki, Max von Pettenkofer Institute) and recombinant adenovirus





expressing the  $\beta$ -gal protein under the control of the human CMV promoter (Ad-lacZ) was propagated as previously described [4,5]. Mice were injected intravenously (i.v.) with Ad-lacZ  $1 \times 10^9$  pfu and/or MCMV  $1 \times 10^6$  pfu. Recombinant IL-2 was obtained from eBioscience (Paisley, UK).

#### 2.3. Detection and analysis of antigen-specific T-cells

Tetramers and fluorochrome-conjugated antibodies are shown in Table S1 and S2, respectively. MHC class I monomers complexed with M38 (H-2Kb), M45 (H-2Db),  $\beta$  gal D8V (H-2Kb) and  $\beta$  gal I8V (H-2Kb) were tetramerized by addition of streptavidin-PE (BD Bioscience) or streptavidin-APC (Invitrogen). Whole blood (100 µl) was stained using a 50 µl tetramer solution at 37 °C for 20 min followed by antibody staining. Viable leukocytes were analyzed using a BD LSR II flow cytometer and FlowJo (Treestar).

#### 2.4. Statistical analysis

Statistical analysis was performed using GraphPad PRISM (La Jolla, USA). P-values were determined by two-tailed T test and corrected using Holm-Sidak for multiple comparisons.

## 3. Results

Studies indicate that IL-2 but not IL-21 is required for MCMVinduced memory inflation [8]. This is in contrast to nonconventional memory responses to adenoviral vaccination [12], and to other situations of persistent antigen presentation such as chronic viral infection, in which CD4<sup>+</sup> T-cells support CD8<sup>+</sup> T-cell responses through IL-21 secretion [10,11,14]. We assessed whether IL-21 mediates CD4<sup>+</sup> T-cell support of inflating Ad-lacZderived CD8<sup>+</sup> T-cell responses by immunizing mice lacking the IL-21 protein ( $ll21^{-/-}$ ) with Ad-lacZ. Intravenous immunization with replication-deficient Ad-lacZ induces a classical (noninflating) memory response to one epitope (I8V) and an inflating response to a second epitope (D8V) [4,5]. We found that conventional T-cell responses were completely abolished, and that inflationary T-cell responses were markedly reduced in  $ll21^{-/-}$ mice (Fig. 1A and B).

We next asked whether tissue homing, activation state or maintenance profile of inflationary Ad-lacZ-specific CD8<sup>+</sup> T-cells differed between  $II21^{-/-}$  and WT mice. We examined lymph node homing based on L-selectin (CD62L) expression, differentiation based on CD27 expression, the homeostatic cytokine IL-7 receptor CD127 (IL-7R) and the memory marker CD44. CD27 is upregulated during the first days after T-cell receptor activation and downregulated during T-cell effector differentiation [15]. CD127 expression is downregulated by activated effector T-cells, and on inflationary CD8<sup>+</sup> T-cells following response to IL-7 [4]. Overall, inflationary AdlacZ-specific CD8<sup>+</sup> T-cells in  $II21^{-/-}$  mice retained an effector memory phenotype (CD62L<sup>-</sup>, CD44<sup>+</sup>), but were less differentiated than their WT counterparts (CD27<sup>+</sup>, Fig. 1C and D). We next examined Ad-lacZ-specific T-cell responses in the tissues (spleen, liver and lung) of  $Il21^{-/-}$  and WT mice (Fig. 2). Similar to the blood, conventional memory responses were absent in the tissues of  $Il21^{-/-}$  mice, and inflationary responses in the spleen and lung were significantly reduced (Fig. 2A). Unexpectedly, in contrast to blood and other organs, the magnitude of inflating memory responses in the liver was relatively preserved in  $Il21^{-/-}$ mice although these showed reduced differentiation, as defined by downregulation of CD27 (Fig. 2B). These data suggest that – in contrast to MCMV – optimal induction, maintenance and differentiation of Ad-lacZ-derived inflating memory T-cell responses are dependent on IL-21.

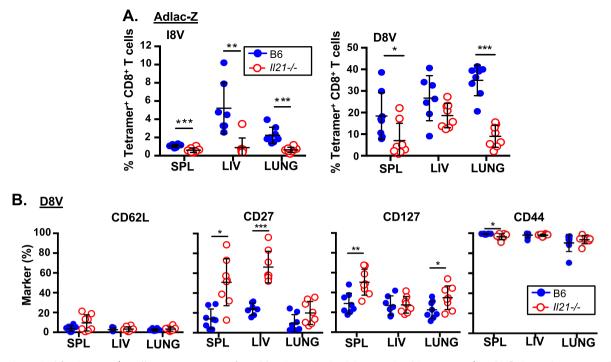
We investigated whether vaccine-derived inflationary CD8<sup>+</sup> Tcell responses could be "rescued" from their reliance on IL-21. Because IL-21 is not needed for memory inflation following MCMV infection [8], we asked whether co-infection with MCMV could provide the necessary factors to overcome the inhibition of adenovirus-vaccine-induced memory inflation observed in Il21-/mice. Simultaneous co-infection with MCMV and Ad-lacZ does not impact the magnitude of inflationary responses [5]. First, we infected WT and  $Il21^{-/-}$  mice with  $1 \times 10^6$  pfu MCMV and confirmed that IL-21 was not required for MCMV-specific inflationary (M38-tetramer<sup>+</sup>) T-cell responses (Fig. 3A). Next, we infected B6 and  $Il21^{-/-}$  mice simultaneously with  $1 \times 10^6$  pfu MCMV and  $1 \times 10^9$  pfu Ad-lacZ. We observed that MCMV did not rescue inflationary Ad-lacZ-specific T-cell responses in *Il21<sup>-/-</sup>* mice (Fig. 3B), indicating that the requirement for IL-21 is regulated by the precise microenvironment where CD8<sup>+</sup> T-cells interact with antigen presenting cells during priming. These data indicate that the induction pathway of inflationary Ad-lacZ-specific CD8<sup>+</sup> T-cells is distinct from inflationary MCMV-specific cells and is highly dependent on IL-21.

Studies have shown that IL-2, likely derived from CD4<sup>+</sup> T-cells, is important during CD8<sup>+</sup> T-cell expansion in response to adenovirus vector vaccination [13] and is needed for MCMV-induced memory inflation [8]. We examined whether IL-2 supplementation could rescue vaccine-derived inflating T-cell responses in *Il*21<sup>-/-</sup> mice (2 µg IL-2 i.p. first 5 days following immunization). Administration of IL-2 did not improve conventional or inflationary adenovirus vaccine-derived T-cell responses (Fig. 3C), although the administration of IL-2 may have been too transient to have a long-term effect. These data suggest that IL-21 and possibly IL-2, each play critical non-redundant roles.

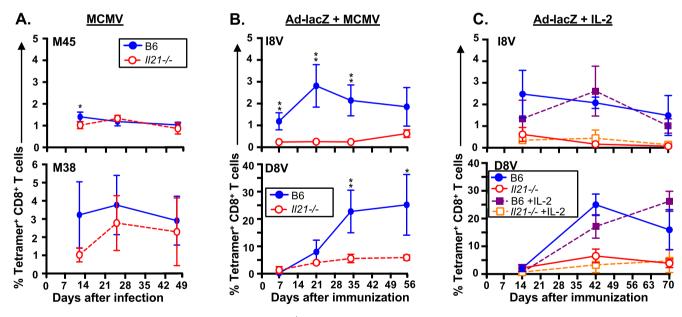
#### 4. Discussion

Understanding the induction and maintenance of memory inflation is important for the development of CD8<sup>+</sup> T-cells vaccines that aim to induce large numbers of memory CD8<sup>+</sup> T-cells capable of providing protection against complex pathogens and cancers. The requirement of CD4<sup>+</sup> T-cell help for the induction and maintenance of T-cell memory has been described to differ between MCMV infection and adenovirus vectors. MCMV-induced memory inflation requires CD4<sup>+</sup> T-cell help for maintenance and is dependent

**Fig. 1.** IL-21 is required for conventional and inflating memory responses after Ad-lacZ immunization C57BL/6 (B6, wild type) and  $ll21^{-/-}$  mice were immunized i.v. with Ad-lacZ 1 × 10<sup>9</sup> pfu and blood serially sampled. Conventional memory responses were assessed by I8V-tetramer staining, and inflationary memory responses were assessed by D8V-tetramer staining. The lower limit of detection of the tetramers is indicated by tetramer staining of naïve B6 and  $ll21^{-/-}$  mice ("Naïve", black open triangle, n = 3–4 mice) (A) Composite FACS plots (n = 3) of I8V (left) – and D8V (right) -tetramer staining of live lymphocytes in B6 (top) and  $ll21^{-/-}$  mice (bottom) 34 dpi. Mean tetramer\* CD8\* T-cells are indicated (n = 3). (B) Mean (±SD) I8V (top)- and D8V (bottom)-tetramer\* CD8\* T-cells (n = 4–5 mice (C) Mean (±SD) I8V (top)- and D8V (bottom)-tetramer\* CD8\* T-cells (n = 3–5 mice). (D and E) Expression of surface markers CD62L, CD27, CD127 and CD44 on D8V-tetramer\* CD8\* T-cells in B6 (top) and  $ll21^{-/-}$  mice (bottom) (n = 3) of CD62L, CD27, CD127 and CD44 on D8V-tetramer\* CD8\* T-cells in B6 (top) and  $ll21^{-/-}$  mice (bottom) (n = 3–5). Significant differences were determined by T test and corrected for multiple comparisons (Holm-Sidak). P = 0.05 to 0.011 (\*), p = 0.01 to 0. 001 (\*\*\*). Data is compiled from 5 independent time course experiments (3–5 mice per time point).



**Fig. 2.** IL-21 is required for tissue CD8<sup>+</sup> T-cell memory responses after Ad-lacZ immunization (A) Conventional (I8V-tetramer<sup>+</sup>) and inflationary (D8V-tetramer<sup>+</sup>) memory CD8<sup>+</sup> T-cells responses were assessed in spleen (SPL), liver (LIV) and lung from C57BL/6 (B6 WT, n = 6) and  $II21^{-/-}$  mice (n = 6) 55–93 days following Ad-lacZ immunization. (B) Expression of surface markers CD62L, CD27, CD127 and CD44 on D8V-tetramer<sup>+</sup> CD8<sup>+</sup> T-cells from SPL, LIV and lung from B6 WT and  $II21^{-/-}$  mice 55–93 dpi. Data is compiled from 2 independent experiments.



**Fig. 3.** Defective Ad-lacZ-induced inflationary memory responses in  $ll21^{-/-}$  mice cannot be rescued by co-infection with MCMV or IL-2 administration (A) C57BL/6 (B6) and  $ll21^{-/-}$  mice were infected i.v. with MCMV 1 × 10<sup>6</sup> pfu and blood serially sampled. Mean (±SD) M45 (top)- and M38 (bottom)-tetramer<sup>+</sup> CD8<sup>+</sup> T-cells (n = 4 mice per time point) are shown. (B) B6 and  $ll21^{-/-}$  mice were simultaneously infected i.v. with Ad-lacZ and MCMV and blood serially sampled. Mean (±SD) I8V (top)- and D8V (bottom)-tetramer<sup>+</sup> CD8<sup>+</sup> T-cells (n = 3-4 mice per time point) are shown. (C) B6 and  $ll21^{-/-}$  mice were immunized i.v. with Ad-lacZ and 2ug IL-2 or PBS was intra-peritoneally injected dially for 5 doses. Blood was serially sampled. Mean (±SD) I8V (top)- and D8V (bottom)-tetramer<sup>+</sup> CD8<sup>+</sup> T-cells (n = 3 mice per time point) are shown. Significant differences were determined by T test and corrected for multiple comparisons (Holm-Sidak). P = 0.05 to 0.011 (\*), p = 0.01 to 0.001 (\*\*\*).

on IL-2 but not IL-21 [8]. In contrast, we found that adenovirusinduced memory inflation is dependent on IL-21 for optimal induction, maintenance and differentiation. These data show that although MCMV infection and Ad-LacZ immunization induce inflationary CD8<sup>+</sup> T-cell responses that are functionally and phenotypically similar, there are parallel pathways of memory induction with the adenovirus vector-pathway highly dependent on IL-21. The relevant sources of IL-21 required for memory inflation in the adenovirus vaccine model (as opposed to the MCMV model) were not examined in this study. IL-21 – like IL-2 – can be derived from different cellular sources, including both CD4<sup>+</sup> and CD8<sup>+</sup> T cells [16]. In MCMV it has been shown that CD8<sup>+</sup> T cell-derived autocrine IL-2 production is critical for optimal memory inflation [17]. Further adoptive transfer or bone marrow chimera studies to define the cellular source(s) of IL-21 necessary or sufficient for memory inflation in the adenovirus model would be valuable in this context. Similarly, to what extent exogenous boosting with IL-21 could further augment T cell responses driven by adenovirus vaccines has a clear translational implication. This approach has already been taken using others vectors, with clear benefit in pre-clinical models [18] – but given the important differences seen here between vectors, this type of approach could have specific relevance in the development of adenovirus vectors now widely used in clinical vaccine studies.

The differences in the fundamental biology MCMV infection and Ad-lacZ immunization may account for differing pathways of induction of memory inflation. Ad-lacZ is a persistent replicationincompetent vector while MCMV is a persistent viral infection characterized by sporadic low-level reactivation events. It is thought that the need for CD4<sup>+</sup> T-cell help during priming occurs in systems where innate immune activation is more modest, as occurs with replication-incompetent vectors [6]. In MCMV infection innate responses are robust enough to drive the CD4<sup>+</sup>-independent expansion of inflationary CD8<sup>+</sup> T-cells (as occurs in acute LCMV infection [19]), such that CD4<sup>+</sup> T-cell-derived IL-21 is not needed [8]. In contrast, adenovirus vectors induce comparatively modest and transient innate immune responses and the induction of a robust CD8<sup>+</sup> T-cell response is more dependent on CD4<sup>+</sup> T-cell help which may require multiple signals, including IL-21. In addition, while both MCMV infection and Ad-lacZ-derived antigens persist, the nature of persistence is quite different. For adenovirus vectors, there is a set amount of antigen that is maintained at some level and then slowly declines over time [4,20]. In comparison, during MCMV infection, the virus pool is maintained - reactivation events allow viral spread to new cells with accompanying innate immune activation. We hypothesized that these differences in antigen persistence and innate immune activation relate to the differential role for specific CD4<sup>+</sup> T-cell pathways in helping CD8<sup>+</sup> T-cell responses. However, we also show that the use of MCMV as an adjuvant in the absence of IL-21 was not sufficient to drive the induction of adenovirus-specific CD8<sup>+</sup> T-cell memory inflation. Therefore, taken together our data indicates that the induction pathway of inflationary Ad-lacZ CD8<sup>+</sup> T-cells is distinct from inflationary MCMV-specific cells and is highly dependent on IL-21.

One surprising observation was the relative preservation of liver-associated inflationary populations (not seen in noninflationary pools), which showed reduced differentiation (CD27hi). In other settings IL-21 is critical for hepatic immunity (e.g. in response to HBV [21]). We hypothesize that this observation may be potentially linked to the local immune-regulatory features in the liver, for example local survival signals mediated by liver sinusoidal endothelial cells [22].

In conclusion, we demonstrate that IL-21 plays a crucial role in the development of inflationary memory T-cell responses following adenovirus-vector vaccination. These data have implications for novel vaccine strategies aiming to promote large and sustained memory T-cell responses.

#### 5. Author contributions

C.H., A.J. H and J.M.C performed the mouse experiments. C.LG, C. H., A.J.H and P.K. analyzed the data. C.L.G., N.M.P. and P.K. wrote the manuscript. All authors attest they meet the ICMJE criteria for authorship.

## Acknowledgements

This work was supported by a Wellcome Trust fellowship (WT091663MA and 109965MA) to P.K., NIH (U19AI 082630), the

Oxford Martin School, the Oxford University Returning Carers Fund (C.H.), Endeavour Postgraduate Scholarship (C.L.G.), and Oxford-UCB Postdoctoral Fellowship (N.M.P.). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript. We acknowledge the NIH Tetramer Core Facility (HHSN272201300006C) for provision of MHC class I monomers. The content is solely the responsibility of the authors and does not necessarily represent the official views of the NIH. The authors have no competing financial interests.

## **Conflicts Of Interest**

Declarations of interest: None.

## Appendix A. Supplementary material

Supplementary data to this article can be found online at https://doi.org/10.1016/j.vaccine.2018.09.061.

#### References

- Karrer U, Sierro S, Wagner M, Oxenius A, Hengel H, Koszinowski UH, et al. Memory inflation: continuous accumulation of antiviral CD8+ T cells over time. J Immunol 2003;170:2022–9.
- [2] Klenerman P, Oxenius A. T cell responses to cytomegalovirus. Nat Rev Immunol 2016;16:367–77.
- [3] Vitelli A, Folgori A, Scarselli E, Colloca S, Capone S, Nicosia A. Chimpanzee adenoviral vectors as vaccines – challenges to move the technology into the fast lane. Exp Rev Vac 2017;16:1241–52.
- [4] Bolinger B, Sims S, O'Hara G, de Lara C, Tchilian E, Firner S, et al. A new model for CD8+ T cell memory inflation based upon a recombinant adenoviral vector. J Immunol 2013;190:4162–74.
- [5] Lee LN, Bolinger B, Banki Z, de Lara C, Highton AJ, Colston JM, et al. Adenoviral vaccine induction of CD8+ T cell memory inflation: impact of co-infection and infection order. PLoS Pathog 2017;13:e1006782.
- [6] Yang TC, Millar J, Groves T, Zhou W, Grinshtein N, Parsons R, et al. On the role of CD4+ T cells in the CD8+ T-cell response elicited by recombinant adenovirus vaccines. Mol Ther 2007;15:997–1006.
- [7] Provine NM, Larocca RA, Penaloza-MacMaster P, Borducchi EN, McNally A, Parenteau LR, et al. Longitudinal requirement for CD4+ T cell help for adenovirus vector-elicited CD8+ T cell responses. J Immunol 2014;192: 5214–25.
- [8] Walton SM, Torti N, Mandaric S, Oxenius A. T-cell help permits memory CD8(+) T-cell inflation during cytomegalovirus latency. Eur J Immunol 2011;41: 2248–59.
- [9] Yi JS, Du M, Zajac AJ. A vital role for interleukin-21 in the control of a chronic viral infection. Science 2009;324:1572–6.
- [10] Elsaesser H, Sauer K, Brooks DG. IL-21 is required to control chronic viral infection. Science 2009;324:1569–72.
- [11] Frohlich A, Kisielow J, Schmitz I, Freigang S, Shamshiev AT, Weber J, et al. IL-21R on T cells is critical for sustained functionality and control of chronic viral infection. Science 2009;324:1576–80.
- [12] Barker BR, Gladstone MN, Gillard GO, Panas MW, Letvin NL. Critical role for IL-21 in both primary and memory anti-viral CD8+ T-cell responses. Eur J Immunol 2010;40:3085–96.
- [13] Provine NM, Larocca RA, Aid M, Penaloza-MacMaster P, Badamchi-Zadeh A, Borducchi EN, et al. Immediate dysfunction of vaccine-elicited CD8+ T Cells primed in the absence of CD4+ T Cells. J Immunol 2016;197:1809–22.
- [14] Yue FY, Lo C, Sakhdari A, Lee EY, Kovacs CM, Benko E, et al. HIV-specific IL-21 producing CD4+ T cells are induced in acute and chronic progressive HIV infection and are associated with relative viral control. J Immunol 2010;185:498–506.
- [15] Hintzen RQ, de Jong R, Lens SM, Brouwer M, Baars P, van Lier RA. Regulation of CD27 expression on subsets of mature T-lymphocytes. J Immunol 1993;151:2426–35.
- [16] Tian Y, Zajac AJ. IL-21 and T cell differentiation: consider the context. Trends Immunol 2016;37:557–68.
- [17] Redeker A, Welten SP, Baert MR, Vloemans SA, Tiemessen MM, Staal FJ, et al. The quantity of autocrine IL-2 governs the expansion potential of CD8+ T Cells. J Immunol 2015;195:4792–801.
- [18] Zeng R, Spolski R, Finkelstein SE, Oh S, Kovanen PE, Hinrichs CS, et al. Synergy of IL-21 and IL-15 in regulating CD8+ T cell expansion and function. J Exp Med 2005;201:139–48.
- [19] Matloubian M, Concepcion RJ, Ahmed R. CD4+ T cells are required to sustain CD8+ cytotoxic T-cell responses during chronic viral infection. J Virol 1994;68:8056–63.

- [20] Finn JD, Bassett J, Millar JB, Grinshtein N, Yang TC, Parsons R, et al. Persistence of transgene expression influences CD8+ T-cell expansion and maintenance following immunization with recombinant adenovirus. J Virol 2009;83: 12027-36.
  [21] Philipere L Could With With the Could Philadel
- [21] Publicover J, Goodsell A, Nishimura S, Vilarinho S, Wang ZE, Avanesyan L, et al. IL-21 is pivotal in determining age-dependent effectiveness of immune

responses in a mouse model of human hepatitis B. J Clin Invest 2011;121: 1154-62.

[22] Schurich A, Berg M, Stabenow D, Bottcher J, Kern M, Schild HJ, et al. Dynamic regulation of CD8 T cell tolerance induction by liver sinusoidal endothelial cells. J Immunol 2010;184:4107–14.