



Case Report Kinetics of Bilirubin and Ammonia Elimination during Hemadsorption Therapy in Secondary Sclerosing Cholangitis Following ECMO Therapy and Severe COVID-19

Désirée Tampe ¹, Peter Korsten ¹, Sebastian C. B. Bremer ², Martin S. Winkler ³ and Björn Tampe ^{1,*}

- ¹ Department of Nephrology and Rheumatology, University Medical Center Göttingen, 37075 Göttingen, Germany; desiree.tampe@med.uni-goettingen.de (D.T.); peter.korsten@med.uni-goettingen.de (P.K.)
- ² Department of Gastroenterology, Gastrointestinal Oncology and Endocrinology, University Medical Center Göttingen, 37075 Göttingen, Germany; sebastian.bremer@med.uni-goettingen.de

Abstract: In critically ill patients, liver dysfunction often results in coagulopathy and encephalopathy

- ³ Department of Anesthesiology, Emergency and Intensive Care Medicine, University Medical Center Göttingen, 37075 Göttingen, Germany; martin.winkler@med.uni-goettingen.de
- * Correspondence: bjoern.tampe@med.uni-goettingen.de; Tel.: +49-551-39-10575

check for updates

Citation: Tampe, D.; Korsten, P.; Bremer, S.C.B.; Winkler, M.S.; Tampe, B. Kinetics of Bilirubin and Ammonia Elimination during Hemadsorption Therapy in Secondary Sclerosing Cholangitis Following ECMO Therapy and Severe COVID-19. *Biomedicines* 2021, 9, 1841. https:// doi.org/10.3390/biomedicines9121841

Academic Editor: Henricus A. M. Mutsaers

Received: 20 October 2021 Accepted: 4 December 2021 Published: 5 December 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). and is associated with high mortality. Extracorporeal clearance of hepatotoxic metabolites, including bilirubin and ammonia, aims to attenuate further hepatocyte damage and liver injury, resulting in decreased mortality. The efficacy of hemadsorption combined with conventional hemodialysis to eliminate bilirubin and ammonia to support the liver's excretory function in acute liver injury has been described previously. However, the optimal use of liver support systems in chronic liver dysfunction due to secondary sclerosing cholangitis in critically ill patients (SSC-CIP) has not been defined yet. We herein describe the kinetics of successful bilirubin and ammonia elimination by hemadsorption in a patient with SSC-CIP after extracorporeal membrane oxygenation (ECMO) therapy for severe acute respiratory distress syndrome (ARDS) in a patient with coronavirus disease 2019 (COVID-19). During the course of the disease, the patient developed laboratory signs of liver injury during ECMO therapy before clinically detectable jaundice or elevated bilirubin levels. A diagnosis of SSC-CIP was confirmed by endoscopic retrograde cholangiopancreatography (ERCP) based on intraductal filling defects in the intrahepatic bile ducts due to biliary casts. The patient showed stable elevations of bilirubin and ammonia levels thereafter, but presented with progressive nausea, vomiting, weakness, and exhaustion. Based on these laboratory findings, hemadsorption was combined with hemodialysis treatment and successfully eliminated bilirubin and ammonia. Moreover, direct comparison revealed that ammonia is more efficiently eliminated by hemadsorption than bilirubin levels. Clinical symptoms of nausea, vomiting, weakness, and exhaustion improved. In summary, bilirubin and ammonia were successfully eliminated by hemadsorption combined with hemodialysis treatment in SSC-CIP following ECMO therapy and severe COVID-19. This observation is particularly relevant since it has been reported that a considerable subset of critically ill patients with COVID-19 suffer from liver dysfunction associated with high mortality.

Keywords: bilirubin elimination; ammonia elimination; hemadsorption; CytoSorb; secondary sclerosing cholangitis; SSC-CIP; COVID-19; extracorporeal membrane oxygenation; intensive care medicine

1. Introduction

In critically ill patients, liver dysfunction potentially results in coagulopathy and encephalopathy and is associated with high mortality [1,2]. Liver dysfunction can either occur because of primary liver diseases or result from secondary causes. Reasons for primary liver dysfunction are hepatotoxic agents, including drugs or viral infection, leading to acute liver dysfunction and potential recovery from injury [3,4]. More frequently, liver dysfunction is observed secondary to cholestasis, hypoxic liver injury, sepsis, or

cardiogenic shock with acute and possibly persistent liver dysfunction in critically ill patients [5,6]. Among them, secondary sclerosing cholangitis (SSC) has been observed after hypoxic liver injury, but also in critically ill patients (SSC-CIP) requiring extracorporeal membrane oxygenation (ECMO) therapy [7]. SSC-CIP and impaired liver clearance cause accumulation of toxic metabolites (including elevated levels of bilirubin and ammonia) with sustained synthetic liver function [7]. In critically ill patients, elevated bilirubin levels are associated with higher mortality [8,9].

Furthermore, accumulation of ammonia can cause cerebral edema with a risk of persistent cerebral injury [10]. Treatment regimens of liver dysfunction in critically ill patients include prevention of ongoing liver injury, ultimately leading to the requirement for liver transplantation or death [11,12]. In addition, liver support systems are used as supportive therapy. Extracorporeal clearance of hepatotoxic metabolites, including bilirubin and ammonia, aims to attenuate further hepatocyte damage and liver injury, potentially resulting in decreased mortality [13]. Liver support systems include therapeutic plasma exchange, which has been shown to improve outcomes but may lead to hypotension and increased bleeding risk [14–17]. Another approach is albumin dialysis, as described with molecular adsorbent recirculating system (MARS) or advanced organ support (ADVOS) [18-20]. ADVOS is an advanced hemodialysis system combining organ support for the liver and kidneys. Although case studies have shown the efficacy of the ADVOS system in critically ill patients with acute liver dysfunction, it has not yet been established in daily intensive care practice [21,22]. Finally, the efficacy of hemadsorption in combination with conventional hemodialysis to eliminate bilirubin and ammonia to support the liver's excretory function in acute liver injury has been described previously [23-26].

However, the role of liver support systems in chronic liver dysfunction, including SSC, has not yet been defined. We herein describe the kinetics of successful bilirubin and ammonia elimination by hemadsorption in a patient with SSC-CIP following ECMO therapy and coronavirus disease 2019 (COVID-19).

2. Case Description

A 61-year-old woman with confirmed COVID-19 was admitted to a different hospital with a productive cough experienced for a few days. She required invasive assisted ventilation shortly after admission. Her medical history included rheumatoid arthritis, obstructive sleep apnea, and arterial hypertension. Because of progressive severe acute respiratory distress syndrome (ARDS), the patient was transferred to our tertiary care hospital for kinetic therapy (prone positioning for at least 12 h per day) and continuous renal replacement therapy (CRRT) due to oliguric acute kidney injury (AKI) (Figure 1A). Because of progressive hypoxemia, venovenous ECMO therapy was initiated 13 days after admission to our hospital (Figure 1A). Subsequently, nasopharyngeal swabs and tracheal aspirates tested negative for SARS-CoV-2. After tracheotomy and weaning, ECMO therapy and invasive assisted ventilation were no longer required, but the patient still needed intermittent renal replacement therapy (IRRT) (Figure 1A).

During the course of the disease, the patient developed laboratory signs of liver injury during ECMO therapy before the clinical appearance of jaundice with elevated bilirubin levels, but sustained synthetic liver function reflected by the international normalized ratio (INR) and serum albumin measurements (Figure 1B–E). A diagnosis of SSC-CIP was confirmed by endoscopic retrograde cholangiopancreatography (ERCP), showing intraductal filling defects in the intrahepatic bile ducts due to biliary casts. In addition, the patient received drugs that have previously been associated with SSC, including amoxicillin-clavulanate, and ketamine sedation [27–29]. Plasma levels of bilirubin and ammonia gradually increased after that, with stable liver synthesis reflected by normal values of the international normalized ratio (INR) without substituting coagulation factors (Figure 1D,E).



Figure 1. Timeline of COVID-19 disease course. (**A**) Timeline of treatment regimens after admission, kinetic therapy included prone positioning for at least 12 h per day. (**B**–**E**) Time course of plasma AST, ALT, GGT, AP, INR, bilirubin, and albumin. Abbreviations: ALT, alanine transaminase; AP, alkaline phosphatase; AST, aspartate transaminase; COVID-19, coronavirus disease 2019; GGT, gamma-glutamyl transferase; INR, international normalized ratio.

Nevertheless, the patient developed progressive nausea, vomiting, weakness, and exhaustion as the disease progressed. Hepatic encephalopathy was treated with lactulose

and rifaximin, but clinical symptoms worsened (Figure 2A). Based on these observations, hemadsorption using the CytoSorb hemoperfusion device (CytoSorbents Europe, Berlin, Germany) was used in combination with IRRT (6 treatments within 7 days and 8–12 h per session). In this patient with SSC following ECMO therapy and severe COVID-19, hemadsorption successfully eliminated bilirubin, ammonia, and C-reactive protein (CRP) levels, while serum albumin levels remained stable (Figure 2A). Direct comparison revealed that bilirubin was less efficiently eliminated by hemadsorption by 33% and 56% after 2 and 6 treatments, respectively (Figure 2B) compared to ammonia by 65% and 74% after 2 and 6 treatments, respectively (Figure 2C). During successful elimination of bilirubin and ammonia, clinical symptoms of nausea, vomiting, weakness, and exhaustion improved. Following a continuation of IRRT but termination of hemadsorption therapy, bilirubin (32.2 mg/dL) and ammonia levels (208 g/dL) increased again, worsening clinical symptoms within 20 days. The patient suddenly died due to cardiac arrhythmia before liver transplantation evaluation was initiated.



Figure 2. Timeline of hemadsorption therapy. (**A**) Arrowheads indicate the time points of hemadsorption. Plasma levels of bilirubin, albumin (upper panel), ammonia (middle panel), CRP, and WBC count (lower panel) are shown. (**B**) Levels of bilirubin before (95 days after admission) and after the initiation of hemadsorption therapy (106 and 110 days after admission). (**C**) Levels of ammonia before (95 days after admission) and after the initiation of hemadsorption therapy (106 and 110 days after admission). (**C**) Levels of and 110 days after admission). Abbreviations: CRP, C-reactive protein; WBC, white blood cell.

3. Discussion

During acute liver dysfunction, a range of toxic metabolites accumulate in the blood. These metabolites include hydrophobic, albumin-bound molecules such as unconjugated bilirubin, bile acids, phenols, aromatic amino acids, and fatty acids, in addition to watersoluble compounds such as ammonia and circulating cytokines. Water-soluble metabolites, including ammonia, are in principle removed by hemodialysis but require high cut-off membranes for sufficient clearance [30–35]. In contrast, protein-bound liver toxins, including bilirubin, are not effectively removed by hemodialysis alone [36,37]. The aim of adsorption technologies as liver support system is the removal of such albumin-bound molecules. In in vitro studies, bilirubin is effectively removed by hemadsorption with a minimal loss of albumin itself [38,39]. Therefore, combining hemadsorption with hemodialysis is an attractive approach for eliminating protein-bound and water-soluble metabolites in patients with severe liver dysfunction. Various blood purification systems are available as liver support therapies to eliminate toxic metabolites in acute liver dysfunction [39]. In particular, methods of albumin dialysis have been frequently used for the clearance of protein-bound liver toxins [19,20]. Beside this, these systems are complicated to use, expensive, and have limited value in daily intensive care practice [21,22]. In addition, a survival benefit has not been demonstrated in randomized clinical trials [40,41]. However, there is evidence that the use of liver support systems improves clinical symptoms of liver dysfunction and might reduce mortality not attributed to a particular liver support system [13].

The application of hemadsorption is best studied in the context of sepsis. The inflammatory state associated with sepsis leads to proinflammatory cytokine release into the systemic circulation with deleterious effects and high mortality [42–44]. These observations resulted in efforts to attenuate this inflammatory response by extracorporeal cytokine removal using CRRT [45]. However, no randomized controlled trials could demonstrate a survival benefit of using CRRT for cytokine removal, including the use of high-volume procedures [46–48]. In contrast to CRRT, multiple experimental studies using animal models of sepsis have demonstrated the efficacy of hemadsorption for reducing various circulating cytokines and chemokines (including CRP) associated with attenuation of organ injury and improved survival [49–53]. According to the manufacturer's data, hemadsorbers have a surface area of about 45,000 m² and eliminate molecules up to 55 kDa in size. In contrast to soluble cytokines and chemokines in circulation, most liver toxins are bound to plasma albumin (39). Protein-bound bilirubin and most bile acids are below 55 kDa in size and can, therefore, principally be eliminated through hemadsorption [26,39]. However, water-soluble metabolites, including ammonia with 17 kDa, are only removed by high-flux hemodialysis [30–35]. While the effectiveness of additional hemadsorption to eliminate protein-bound bilirubin and water-soluble ammonia has been described previously in critically ill patients with acute liver failure, treatment efficacy in SSC-CIP has not yet been reported [54–56]. The pathogenesis of SSC-CIP remains elusive, but current concepts suggest that the primary insult is hypoperfusion of the biliary vasculature. The intrahepatic cholangiocyte epithelium is supplied by the peribiliary vascular plexus that arises from the hepatic arteries. This contrasts with the liver parenchyma supplied by a dual blood source arising from the hepatic arteries and portal system. Therefore, the biliary vasculature is thought to be more susceptible to hypoperfusion and consecutive ischemia [57]. SSC-CIP has previously been observed after ECMO therapy, including in cases of COVID-19, and a direct viral tropism with detection of SARS-CoV-2 RNA and associated nucleocapsid protein in cholangiocytes and bile ducts has been proposed [58,59]. Finally, endotheliitis resulting in hypercoagulability in the peribiliary vascular plexus may also aggravate ischemia of the biliary tract.

We herein expand current knowledge and describe a case of SSC-CIP following ECMO therapy and severe COVID-19 treated by hemadsorption. The main limitation is a case description, and further clinical research is required to strengthen our observations. Moreover, hemadsorption has already been described in the context of acute liver failure [54–56]. This is particularly relevant since it has been reported that about one-third of patients critically ill with COVID-19 suffer from liver dysfunction associated with high mortality [60]. Moreover, this is the first report of SSC-CIP treated with hemadsorption, especially relevant since SSC-CIP affects a considerable number of COVID-19 patients requiring ECMO therapy [58]. By combining hemadsorption with hemodialysis treatment, successful bilirubin and ammonia elimination were observed. More importantly, our patient showed that extracorporeal clearance of bilirubin and ammonia were associated with improved clinical symptoms, including nausea, vomiting, weakness, and exhaustion.

4. Conclusions

In conclusion, the kinetics of bilirubin and ammonia during hemadsorption therapy confirmed efficient removal in this case of SSC following ECMO therapy and severe COVID-19, associated with regredient clinical symptoms of chronic liver dysfunction. Furthermore, direct comparison revealed that ammonia is more efficiently eliminated by hemadsorption than bilirubin levels.

Author Contributions: D.T. and B.T. conceived the case report, collected and analyzed data, and co-wrote the first draft. S.C.B.B., M.S.W. and B.T. were directly involved in the patient's treatment. P.K., S.C.B.B. and M.S.W. edited the manuscript. All authors have read and agreed to the published version of the manuscript.

Funding: We acknowledge support from the Open Access Publication Funds of the Georg August University Göttingen.

Institutional Review Board Statement: The study was conducted according to the guidelines of the Declaration of Helsinki, and approval by the Institutional Review Board was not required for reporting individual case reports. Written informed consent to publish this report was obtained directly from the patient.

Data Availability Statement: Deidentified data are available upon reasonable request from the corresponding author.

Conflicts of Interest: The authors declare no conflict of interest. The funders had no role in the design of the study, in the collection, analyses, or interpretation of data, in the writing of the manuscript, or in the decision to publish the results.

References

- Reuben, A.; Tillman, H.; Fontana, R.J.; Davern, T.; McGuire, B.; Stravitz, R.T.; Durkalski, V.; Larson, A.M.; Liou, I.; Fix, O.; et al. Outcomes in adults with acute liver failure between 1998 and 2013: An observational cohort study. *Ann. Intern. Med.* 2016, 164, 724–732. [CrossRef]
- Huang, H.; Li, H.; Chen, S.; Zhou, X.; Dai, X.; Wu, J.; Zhang, J.; Shao, L.; Yan, R.; Wang, M.; et al. Prevalence and characteristics of hypoxic hepatitis in COVID-19 patients in the intensive care unit: A first retrospective study. *Front. Med.* 2020, 7, 607206. [CrossRef]
- 3. Stravitz, R.T.; Lee, W.M. Acute liver failure. Lancet 2019, 394, 869-881. [CrossRef]
- 4. Real, M.; Barnhill, M.S.; Higley, C.; Rosenberg, J.; Lewis, J.H. Drug-induced liver injury: Highlights of the recent literature. *Drug Saf.* **2019**, *42*, 365–387. [CrossRef]
- 5. Horvatits, T.; Trauner, M.; Fuhrmann, V. Hypoxic liver injury and cholestasis in critically ill patients. *Curr. Opin. Crit. Care.* 2013, 19, 128–132. [CrossRef]
- 6. Lelubre, C.; Vincent, J.L. Mechanisms and treatment of organ failure in sepsis. Nat. Rev. Nephrol. 2018, 14, 417–427. [CrossRef]
- Tunney, R.; Scott, J.; Rudralingam, V.; Liong, S.; Athwal, V.S. Secondary sclerosing cholangitis following extracorporeal membrane oxygenation for acute respiratory distress in polytrauma. *Clin. Case Rep.* 2018, *6*, 1849–1853. [CrossRef]
- 8. Vitek, L.; Hubacek, J.A.; Pajak, A.; Dorynska, A.; Kozela, M.; Eremiasova, L.; Danzig, V.; Stefler, D.; Bobak, M. Association between plasma bilirubin and mortality. *Ann. Hepatol.* **2019**, *18*, 379–385. [CrossRef]
- Kramer, L.; Jordan, B.; Druml, W.; Bauer, P.; Metnitz, P.G. Austrian epidemiologic study on intensive care ASG. Incidence and prognosis of early hepatic dysfunction in critically ill patients—A prospective multicenter study. *Crit. Care Med.* 2007, 35, 1099–1104. [CrossRef]
- 10. Kerbert, A.J.; Engelmann, C.; Jalan, R. Neurocritical care management of hepatic encephalopathy and coma in liver failure. *Semin. Respir. Crit. Care Med.* **2018**, *39*, 523–537. [CrossRef]

- 11. Olivo, R.; Guarrera, J.V.; Pyrsopoulos, N.T. Liver transplantation for acute liver failure. *Clin. Liver Dis.* **2018**, 22, 409–417. [CrossRef]
- 12. Khan, R.; Koppe, S. Modern management of acute liver failure. Gastroenterol. Clin. North Am. 2018, 47, 313–326. [CrossRef]
- 13. Alshamsi, F.; Alshammari, K.; Belley-Cote, E.; Dionne, J.; Albrahim, T.; Albudoor, B.; Ismail, M.; Al-judaibi, B.; Baw, B.; Subramanian, R.M.; et al. Extracorporeal liver support in patients with liver failure: A systematic review and meta-analysis of randomized trials. *Intensive Care Med.* **2020**, *46*, 1–16. [CrossRef] [PubMed]
- 14. Larsen, F.S.; Schmidt, L.E.; Bernsmeier, C.; Rasmussen, A.; Isoniemi, H.; Patel, V.C.; Triantafyllou, E.; Bernal, W.; Auzinger, G.; Shawcross, D.; et al. High-volume plasma exchange in patients with acute liver failure: An open randomised controlled trial. *J. Hepatol.* **2016**, *64*, 69–78. [CrossRef] [PubMed]
- 15. Yuan, S.; Qian, Y.; Tan, D.; Mo, D.; Li, X. Therapeutic plasma exchange: A prospective randomized trial to evaluate 2 strategies in patients with liver failure. *Transfus. Apher. Sci.* 2018, *57*, 253–258. [CrossRef]
- 16. Stahl, K.; Hadem, J.; Schneider, A.; Manns, M.P.; Wiesner, O.; Schmidt, B.M.W.; Hoeper, M.M.; Busch, M.; David, S. Therapeutic plasma exchange in acute liver failure. *J. Clin. Apher.* **2019**, *34*, 589–597. [CrossRef]
- 17. Stahl, K.; Busch, M.; Fuge, J.; Schneider, A.; Manns, M.P.; Seeliger, B.; Schmidt, J.J.; Wiesner, O.; Schmidt, B.M.W.; Taubert, R.; et al. Therapeutic plasma exchange in acute on chronic liver failure. *J. Clin. Apher.* **2020**, *35*, 316–327. [CrossRef]
- 18. Tsipotis, E.; Shuja, A.; Jaber, B.L. Albumin dialysis for liver failure: A systematic review. *Adv. Chronic Kidney Dis.* **2015**, *22*, 382–390. [CrossRef]
- Falkensteiner, C.; Kortgen, A.; Leonhardt, J.; Bauer, M.; Sponholz, C. Comparison of albumin dialysis devices molecular adsorbent recirculating system and advanced organ support system in critically ill patients with liver failure—A retrospective analysis. *Ther. Apher. Dial.* 2021, 25, 225–236. [CrossRef]
- Fuhrmann, V.; Perez Ruiz de Garibay, A.; Faltlhauser, A.; Tyczynski, B.; Jarczak, D.; Lutz, J.; Weinamnn-Menke, J.; Kribben, A.; Kluge, S. Registry on extracorporeal multiple organ support with the advanced organ support (ADVOS) system: 2-year interim analysis. *Medicine* 2021, 100, e24653. [CrossRef]
- Fuhrmann, V.; Weber, T.; Roedl, K.; Motaabbed, J.; Tariparast, A.; Jarczak, D.; Perez Ruiz de Garibay, A.; Kluwe, J.; Boenisch, O.; Herkner, H.; et al. Advanced organ support (ADVOS) in the critically ill: First clinical experience in patients with multiple organ failure. *Ann. Intensive Care* 2020, 10, 96. [CrossRef]
- 22. Huber, W.; Henschel, B.; Schmid, R.; Al-Chalabi, A. First clinical experience in 14 patients treated with ADVOS: A study on feasibility, safety and efficacy of a new type of albumin dialysis. *BMC Gastroenterol.* **2017**, *17*, 32. [CrossRef]
- 23. Acar, U.; Gokkaya, Z.; Akbulut, A.; Ferah, O.; Yenidunya, O.; Acik, M.E.; Tokat, Y.; Yentür, E. Impact of cytokine adsorption treatment in liver failure. *Transplant. Proc.* **2019**, *51*, 2420–2424. [CrossRef]
- 24. Dhokia, V.D.; Madhavan, D.; Austin, A.; Morris, C.G. Novel use of CytoSorb haemadsorption to provide biochemical control in liver impairment. *J. Intensive Care Soc.* 2019, 20, 174–181. [CrossRef] [PubMed]
- Piwowarczyk, P.; Kutnik, P.; Potrec-Studzinska, B.; Sysiak-Slawecka, J.; Rypulak, E.; Borys, M.; Czczuwar, M. Hemoadsorption in isolated conjugated hyperbilirubinemia after extracorporeal membrane oxygenation support. Cholestasis of sepsis: A case report and review of the literature on differential causes of jaundice in ICU patient. *Int. J. Artif. Organs.* 2019, 42, 263–268. [CrossRef]
- Calabro, M.G.; Febres, D.; Recca, G.; Lembo, R.; Fominskiy, E.; Scandroglio, A.M.; Zangrillo, A.; Pappalardo, F. Blood purification with CytoSorb in critically ill patients: Single-center preliminary experience. *Artif. Organs.* 2019, 43, 189–194. [CrossRef]
- 27. Brooling, J.; Leal, R. Secondary sclerosing cholangitis: A review of recent literature. *Curr. Gastroenterol. Rep.* **2017**, *19*, 44. [CrossRef]
- Visentin, M.; Lenggenhager, D.; Gai, Z.; Kullak-Ublick, G.A. Drug-induced bile duct injury. *Biochim. Biophys. Acta Mol. Basis Dis.* 2018, 1864, 1498–1506. [CrossRef] [PubMed]
- 29. Lo, R.S.; Krishnamoorthy, R.; Freeman, J.G.; Austin, A.S. Cholestasis and biliary dilatation associated with chronic ketamine abuse: A case series. *Singapore Med. J.* 2011, *52*, e52–e55. [PubMed]
- 30. Wakim-Fleming, J. Hepatic encephalopathy: Suspect it early in patients with cirrhosis. *Cleve Clin. J. Med.* **2011**, *78*, 597–605. [CrossRef]
- 31. Bernal, W.; Wendon, J. Acute liver failure. N. Engl. J Med. 2013, 369, 2525–2534. [CrossRef] [PubMed]
- 32. Bernal, W.; Lee, W.M.; Wendon, J.; Larsen, F.S.; Williams, R. Acute liver failure: A curable disease by 2024? *J. Hepatol.* 2015, 62 (Suppl. S1), S112–S120. [CrossRef]
- 33. Slack, A.J.; Auzinger, G.; Willars, C.; Dew, T.; Musto, R.; Corsilli, D.; Sherwood, R.; Wendon, J.A.; Bernal, W. Ammonia clearance with haemofiltration in adults with liver disease. *Liver Int.* **2014**, *34*, 42–48. [CrossRef]
- 34. Cardoso, F.S.; Gottfried, M.; Tujios, S.; Olson, J.C.; Karvellas, C.J.; Group USALFS. Continuous renal replacement therapy is associated with reduced serum ammonia levels and mortality in acute liver failure. *Hepatology* **2018**, *67*, 711–720. [CrossRef]
- 35. Boer, D.P.; Mourik, S.L.; van den Hoogen, M.W.F.; Langendonk, J.G.; de Geus, H.R.H. Successful treatment of severe hyperammonaemia with ultra-high dose continuous veno-venous haemodiafiltration. *Blood Purif.* **2019**, *48*, 283–285. [CrossRef]
- Mikhalovsky, S.V. Emerging technologies in extracorporeal treatment: Focus on adsorption. *Perfusion* 2003, 18 (Suppl. S1), 47–54. [CrossRef] [PubMed]
- 37. Weber, C.; Linsberger, I.; Rafiee-Tehrani, M.; Falkenhagen, D. Permeability and adsorption capacity of dialysis membranes to lipid A. *Int. J. Artif. Organs.* **1997**, *20*, 144–152. [CrossRef]

- Gemelli, C.; Cuoghi, A.; Magnani, S.; Atti, M.; Ricci, D.; Siniscalchi, A.; Mancini, E.; Faenza, S. Removal of Bilirubin with a new adsorbent system: In vitro kinetics. *Blood Purif.* 2019, 47, 10–15. [CrossRef]
- Dominik, A.; Stange, J. Similarities, differences, and potential synergies in the mechanism of action of albumin dialysis using the MARS albumin dialysis device and the CytoSorb hemoperfusion device in the treatment of liver failure. *Blood Purif.* 2021, 50, 119–128. [CrossRef]
- 40. Niewinski, G.; Raszeja-Wyszomirska, J.; Hrenczuk, M.; Rozga, A.; Malkowski, P.; Rozga, J. Intermittent high-flux albumin dialysis with continuous venovenous hemodialysis for acute-on-chronic liver failure and acute kidney injury. *Artif. Organs.* **2020**, *44*, 91–99. [CrossRef] [PubMed]
- 41. Gerth, H.U.; Pohlen, M.; Tholking, G.; Pavenstadt, H.; Brand, M.; Wilms, C.; Hüsing-Kabar, A.; Görlich, D.; Kabar, I.; Schmidt, H.H.J. Molecular adsorbent recirculating system (MARS) in acute liver injury and graft dysfunction: Results from a case-control study. *PLoS ONE* **2017**, *12*, e0175529. [CrossRef]
- 42. Gruda, M.C.; Ruggeberg, K.G.; O'Sullivan, P.; Guliashvili, T.; Scheirer, A.R.; Golobish, T.D.; Capponi, V.J.; Chan, P.P. Broad adsorption of sepsis-related PAMP and DAMP molecules, mycotoxins, and cytokines from whole blood using CytoSorb(R) sorbent porous polymer beads. *PLoS ONE* **2018**, *13*, e0191676. [CrossRef]
- 43. Monneret, G.; Finck, M.E.; Venet, F.; Debard, A.L.; Bohe, J.; Bienvenu, J.; Lepape, A. The anti-inflammatory response dominates after septic shock: Association of low monocyte HLA-DR expression and high interleukin-10 concentration. *Immunol. Lett.* **2004**, *95*, 193–198. [CrossRef] [PubMed]
- Frencken, J.F.; van Vught, L.A.; Peelen, L.M.; Ong, D.S.Y.; Klein Klouwenberg, P.M.C.; Horn, J.; Bonten, M.J.M.; van der Poll, T.; Cremer, O.L. An unbalanced inflammatory cytokine response is not associated with mortality following sepsis: A prospective cohort study. *Crit. Care Med.* 2017, 45, e493–e499. [CrossRef] [PubMed]
- 45. De Vriese, A.S.; Colardyn, F.A.; Philippe, J.J.; Vanholder, R.C.; De Sutter, J.H.; Lameire, N.H. Cytokine removal during continuous hemofiltration in septic patients. *J. Am. Soc. Nephrol.* **1999**, *10*, 846–853. [CrossRef] [PubMed]
- 46. Lukaszewicz, A.C.; Payen, D. Purification methods: A way to treat severe acute inflammation related to sepsis? *Crit. Care.* **2013**, 17, 159. [CrossRef]
- 47. Cole, L.; Bellomo, R.; Journois, D.; Davenport, P.; Baldwin, I.; Tipping, P. High-volume haemofiltration in human septic shock. *Intensive Care Med.* **2001**, *27*, 978–986. [CrossRef] [PubMed]
- Honore, P.M.; Jamez, J.; Wauthier, M.; Lee, P.A.; Dugernier, T.; Pirenne, B.; Hanique, G.; Matson, J.R. Prospective evaluation of short-term, high-volume isovolemic hemofiltration on the hemodynamic course and outcome in patients with intractable circulatory failure resulting from septic shock. *Crit. Care Med.* 2000, *28*, 3581–3587. [CrossRef] [PubMed]
- 49. Kellum, J.A.; Song, M.; Venkataraman, R. Hemoadsorption removes tumor necrosis factor, interleukin-6, and interleukin-10, reduces nuclear factor-kappaB DNA binding, and improves short-term survival in lethal endotoxemia. *Crit. Care Med.* **2004**, *32*, 801–805. [CrossRef]
- Namas, R.A.; Namas, R.; Lagoa, C.; Barclay, D.; Mi, Q.; Zamora, R.; Peng, Z.; Wen, X.; Fedorchak, M.V.; Valenti, I.E.; et al. Hemoadsorption reprograms inflammation in experimental gram-negative septic peritonitis: Insights from in vivo and in silico studies. *Mol. Med.* 2012, *18*, 1366–1374. [CrossRef] [PubMed]
- 51. Peng, Z.Y.; Carter, M.J.; Kellum, J.A. Effects of hemoadsorption on cytokine removal and short-term survival in septic rats. *Crit. Care Med.* **2008**, *36*, 1573–1577. [CrossRef] [PubMed]
- 52. Peng, Z.Y.; Wang, H.Z.; Carter, M.J.; Dileo, M.V.; Bishop, J.V.; Zhou, F.H.; Wen, X.-Y.; Rimmelé, T.; Singbartl, K.; Federspiel, W.J.; et al. Acute removal of common sepsis mediators does not explain the effects of extracorporeal blood purification in experimental sepsis. *Kidney Int.* **2012**, *81*, 363–369. [CrossRef]
- 53. Akil, A.; Ziegeler, S.; Reichelt, J.; Rehers, S.; Abdalla, O.; Semik, M.; Fischer, S. Combined use of CytoSorb and ECMO in patients with severe pneumogenic sepsis. *Thorac. Cardiovasc. Surg.* **2021**, *69*, 246–251. [CrossRef]
- 54. Singh, A.; Mehta, Y.; Trehan, N. Bilirubin removal using CytoSorb filter in a cardiac surgical patient. *J. Cardiothorac. Vasc. Anesth.* **2019**, *33*, 881–883. [CrossRef]
- Scharf, C.; Liebchen, U.; Paal, M.; Becker-Pennrich, A.; Irlbeck, M.; Zoller, M.; Schroeder, I. Successful elimination of bilirubin in critically ill patients with acute liver dysfunction using a cytokine adsorber and albumin dialysis: A pilot study. *Sci. Rep.* 2021, *11*, 10190. [CrossRef]
- 56. Buttner, S.; Patyna, S.; Koch, B.; Finkelmeier, F.; Geiger, H.; Sarrazin, C.; Farnik, H. Application of hemoadsorption in a case of liver cirrhosis and alcohol-related steatohepatitis with preexisting hepatitis C infection. *Blood Purif.* **2017**, *44*, 30–31. [CrossRef]
- Leonhardt, S.; Veltzke-Schlieker, W.; Adler, A.; Schott, E.; Eurich, D.; Faber, W.; Neuhaus, P.; Seehofer, D. Secondary sclerosing cholangitis in critically ill patients: Clinical presentation, cholangiographic features, natural history, and outcome: A series of 16 cases. *Medicine* 2015, 94, e2188. [CrossRef] [PubMed]
- Meersseman, P.; Blondeel, J.; De Vlieger, G.; van der Merwe, S.; Monbaliu, D. Collaborators leuven liver transplant p. Secondary sclerosing cholangitis: An emerging complication in critically ill COVID-19 patients. *Intensive Care Med.* 2021, 47, 1037–1040. [CrossRef]

- 59. Kaltschmidt, B.; Fitzek, A.D.E.; Schaedler, J.; Forster, C.; Kaltschmidt, C.; Hansen, T.; Steinfurth, F.; Windmöller, B.A.; Pilger, C.; Kong, C.; et al. Hepatic vasculopathy and regenerative responses of the liver in fatal Cases of COVID-19. *Clin. Gastroenterol. Hepatol.* **2021**, *19*, 1726–1729.e3. [CrossRef]
- 60. Roedl, K.; Jarczak, D.; Drolz, A.; Wichmann, D.; Boenisch, O.; de Heer, G.; Burdelski, C.; Frings, D.; Sensen, B.; Nierhaus, A.; et al. Severe liver dysfunction complicating course of COVID-19 in the critically ill: Multifactorial cause or direct viral effect? *Ann. Intensive Care* **2021**, *11*, 44. [CrossRef]