

EDITORIAL

Necrosis, Apoptosis, Necroptosis, Pyroptosis: It Matters How Acinar Cells Die During Pancreatitis



Acute pancreatitis is an unpredictable disease. Most patients develop edema of the pancreas and quickly recover whatever the therapy (or lack thereof), but a minority develop severe complications with significant morbidity and mortality. When the disease is studied in experimental animal models,¹ to distinguish between the 2 forms, it appears that all processes begin in acinar cells² but then develop in different directions. The way acinar cells undergo injury and cell death seems to determine the ultimate severity. A highly regulated form of cell death is apoptosis, characterized by the activation of multiple caspases, in response to which acinar cells shrink and disintegrate into neat membrane-confined packages that are taken up by neighboring cells or macrophages. No cellular content is released and no tissue infiltration with inflammatory cells is triggered. Conversely, when acinar cells undergo necrosis, a process previously regarded as completely unregulated, their membranes disintegrate, their proteolytic enzymes and organelles are spilled into the interstitial space³ where some components act as damage-associated molecular patterns such as free adenosine triphosphate, free DNA, or released histones.⁴ Although cell death is never a good thing, a shift from necrosis to apoptosis in response to a pathologic stimulus can render experimental pancreatitis significantly less severe⁵ and thus confers a beneficial effect.

Recent studies have shown that necrosis is not necessarily haphazard or spontaneous and can be tightly regulated as well. The best investigated form of regulated necrosis was termed *necroptosis* and involves activation of the receptor-interacting protein kinases (RIP)1/RIP3/mixed lineage kinase domain-like (MLKL) pathway.⁶ This is where Louhimo et al⁷ started. They investigated 2 reductionist, isolated, acini-based models of pancreatitis, 1 using supramaximal secretagogue stimulation and the other meant to mimic gallstone-induced pancreatitis,⁸ asking what form of cell death prevails and which role necroptosis plays. By using genetic deletion of RIP3 and a potent inhibitor of the RIP1/RIP3 pathway, necrostatin,⁹ they found not only that necroptosis is the predominant form of acinar cell death in these models (rather than apoptosis), but that the prevention of necroptosis greatly affects the disease severity *in vivo*—for the better. This makes necroptosis an attractive target for the prevention of pancreatitis or at least for the reduction of its severity. In another set of experiments, Louhimo et al⁷ established that a therapy directed against necroptosis can still be effective when pancreatitis is already established, making therapy, rather than prevention, an attractive goal.

A signaling mechanism reported to be involved in necrosis¹⁰ as well as apoptosis is that involving tumor necrosis

factor (TNF) α as a ligand. He et al⁶ showed that TNF α is also the critical stimulator of the RIP1/RIP3/MLKL pathway and induces necroptosis. In pancreatic acinar cells, injury has previously been attributed to high cytosolic Ca⁺⁺ concentrations¹¹ released in response to pathologic cholecystokinin or acetylcholine concentrations. What Louhimo et al⁷ established is that necroptosis not only depends on TNF α , but also on pathologic Ca⁺⁺ signaling. Louhimo et al⁷ correctly concluded that targeting necroptosis is probably the most attractive way of reducing the severity of pancreatitis.

However, according to the data of Louhimo et al,⁷ inhibiting necroptosis still leaves 40% of cells dead, with neither necroptosis nor apoptosis being involved in their demise. Could another programmed form of necrosis called pyroptosis account for the rest? Pyroptosis, a highly inflammatory variety of cell death, involves activation of nuclear factor- κ B and the expression of components and effectors of the NACHT, LRR and PYD domains-containing protein 3-inflammasome, a cytosolic protein complex consisting of NACHT, LRR and PYD domains-containing protein 3, apoptosis-associated speck-like protein (Apoptosis-associated speck-like protein containing a CARD), and procaspase 1. It proteolytically activates pro-interleukin (IL)1 β and pro-IL18, and induces release of active IL1 β , IL18, and high-mobility group protein B1 in response to a wide range of stimuli, including extracellular adenosine triphosphate, Nicotinamide-Adenin-Dinucleotide, and saturated free fatty acids. Interestingly, when components of the inflammasome pathway are genetically deleted, the cell death rate in pancreatitis is reduced to the same extent^{4,12} as found by Louhimo et al⁷ when they prevented necroptosis. These observations must not be mutually exclusive because they were both obtained in highly reductionist models of pancreatitis. We believe they are linked and that preventing one type of regulated cell death induces activation of an alternative pathway of programmed cell death.

What remains to be investigated in our view is whether the defensive mechanism of the pancreas such as autophagy¹³ and endosomal/lysosomal degradation¹⁴ can counteract necroptosis effectively. It further needs to be determined whether the necrosis/inflammation/fibrosis sequence that mediates the progression from an isolated episode of acute pancreatitis to chronic pancreatitis¹⁵ with atrophy, exocrine insufficiency, and endocrine insufficiency also lends itself to therapeutic intervention based on preventing necroptosis. Louhimo et al⁷ have accomplished a significant step forward in this direction.

MATTHIAS SENDLER, PhD

JULIA MAYERLE, MD

MARKUS M. LERCH, MD, FRCP

Department of Medicine A

University Medicine Greifswald

Ferdinand-Sauerbruch-Strasse

Greifswald, Germany

References

1. Lerch MM, Gorelick FS. Models of acute and chronic pancreatitis. *Gastroenterology* 2013;144:1180–1193.
2. Lerch MM, Saluja AK, Dawra R, et al. Acute necrotizing pancreatitis in the opossum: earliest morphological changes involve acinar cells. *Gastroenterology* 1992;103:205–213.
3. Mayerle J, Schnekenburger J, Krüger B, et al. Extracellular cleavage of E-cadherin by leukocyte elastase during acute experimental pancreatitis in rats. *Gastroenterology* 2005;129:1251–1267.
4. Hoque R, Sohail M, Malik A, et al. TLR9 and the NLRP3 inflammasome link acinar cell death with inflammation in acute pancreatitis. *Gastroenterology* 2011;141:358–369.
5. Mareninova OA, Sung KF, Hong P, et al. Cell death in pancreatitis: caspases protect from necrotizing pancreatitis. *J Biol Chem* 2006;281:3370–3381.
6. He S, Wang L, Miao L, et al. Receptor interacting protein kinase-3 determines cellular necrotic response to TNF-alpha. *Cell* 2009;137:1100–1111.
7. Louhimo JM, Steer ML, Perides G. Necroptosis is an important severity determinant and potential therapeutic target in experimental severe pancreatitis. *Cell Mol Gastroenterol Hepatol* 2016;2:519–535.
8. Hernández CA, Lerch MM. Sphincter stenosis and gallstone migration through the biliary tract. *Lancet* 1993;341:1371–1373.
9. Degterev A, Hitomi J, Germscheid M, et al. Identification of RIP1 kinase as a specific cellular target of necrostatins. *Nat Chem Biol* 2008;4:313–321.
10. Sendler M, Dummer A, Weiss FU, et al. Tumour necrosis factor α secretion induces protease activation and acinar cell necrosis in acute experimental pancreatitis in mice. *Gut* 2013;62:430–439.
11. Krüger B, Albrecht E, Lerch MM. The role of intracellular calcium signaling in premature protease activation and the onset of pancreatitis. *Am J Pathol* 2000;157:43–50.
12. Hoque R, Mehal WZ. Inflammasomes in pancreatic physiology and disease. *Am J Physiol Gastrointest Liver Physiol* 2015;308:G643–G651.
13. Mareninova OA, Sendler M, Malla SR, et al. Lysosome associated membrane proteins maintain pancreatic acinar cell homeostasis: LAMP-2 deficient mice develop pancreatitis. *Cell Mol Gastroenterol Hepatol* 2015;1:678–694.
14. Hirano T, Saluja A, Ramarao P, et al. Apical secretion of lysosomal enzymes in rabbit pancreas occurs via a secretagogue regulated pathway and is increased after pancreatic duct obstruction. *J Clin Invest* 1991;87:865–869.
15. Gress TM, Müller-Pillasch F, Lerch MM, et al. Balance of expression of genes coding for extracellular matrix proteins and extracellular matrix degrading proteases in chronic pancreatitis. *Z Gastroenterol* 1994;32:221–225.

Correspondence

Address correspondence to: Markus M. Lerch, MD, FRCP, Department of Medicine A, University Medicine Greifswald, Ferdinand-Sauerbruch-Strasse, 17475 Greifswald, Germany. e-mail: lerch@uni-greifswald.de.

Conflicts of interest

The authors disclose no conflicts.

Most current article

© 2016 The Authors. Published by Elsevier Inc. on behalf of the AGA Institute. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).
2352-345X
<http://dx.doi.org/10.1016/j.jcmgh.2016.05.007>