

Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active. Contents lists available at ScienceDirect

Clinical Immunology

journal homepage: www.elsevier.com/locate/yclim



Micronutrients as immunomodulatory tools for COVID-19 management

Amin Gasmi^a, Torsak Tippairote^{b,c}, Pavan Kumar Mujawdiya^d, Massimiliano Peana^e, Alain Menzel^t, Maryam Dadar^g, Asma Gasmi Benahmed^h, Geir Bjørklund^{i,}

^a Société Francophone de Nutrithérapie et de Nutrigénétique Appliquée, Villeurbanne, France

^b Philosophy Program in Nutrition, Faculty of Medicine, Ramathibodi Hospital and Institute of Nutrition, Mahidol University, Bangkok, Thailand

^c Nutritional and Environmental Medicine Department, BBH Hospital, Bangkok, Thailand

^d Birla Institute of Technology and Science -Pilani, Hyderabad, India

e Department of Chemistry and Pharmacy, University of Sassari, Italy

f Laboratoires Réunis, Junglinster, Luxembourg

⁸ Razi Vaccine and Serum Research Institute, Agricultural Research, Education and Extension Organization (AREEO), Karaj, Iran

^h Académie Internationale de Médecine Dentaire Intégrative, Paris, France

ⁱ Council for Nutritional and Environmental Medicine (CONEM), Mo i Rana, Norway

ARTICLE INFO

Keywords: Immunomodulation COVID-19 SARs-CoV-2 Nutrients Vitamins Trace elements

ABSTRACT

COVID-19 rapidly turned to a global pandemic posing lethal threats to overwhelming health care capabilities, despite its relatively low mortality rate. The clinical respiratory symptoms include dry cough, fever, anosmia, breathing difficulties, and subsequent respiratory failure. No known cure is available for COVID-19. Apart from the anti-viral strategy, the supports of immune effectors and modulation of immunosuppressive mechanisms is the rationale immunomodulation approach in COVID-19 management. Diet and nutrition are essential for healthy immunity. However, a group of micronutrients plays a dominant role in immunomodulation. The deficiency of most nutrients increases the individual susceptibility to virus infection with a tendency for severe clinical presentation. Despite a shred of evidence, the supplementation of a single nutrient is not promising in the general population. Individuals at high-risk for specific nutrient deficiencies likely benefit from supplementation. The individual dietary and nutritional status assessments are critical for determining the comprehensive actions in COVID-19.

1. Introduction

The coronavirus disease 2019 (COVID-19) is a respiratory disorder that is the consequence of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection [1]. Since the first identified case in Wuhan, China, it took only three months for a global pandemic of the disease due to the highly contagious of this virus through droplets transmission [2-5].

The host responses to SARS-CoV-2 infection diverse; 78% of newly infected persons may remain asymptomatic, while 84.3%, 9.6%, and 6.1 % of clinical patients present with mild, moderate, and severe symptoms, respectively [6,7]. The viral responses of innate and adaptive immune machinery differ upon the host metabolic determinants, including age, sex, nutritional status, smoking habits, and co-existing medical conditions [6,8-12]. The findings of asymptomatic and mild clinical symptoms in younger individuals signify the role of host status in SARS-CoV-2 infection [9,13-15].

In contrast to the current anti-viral approach targeting the specific pathogen, an emerging strategy aims the host immunity activation to fight the virus [16,17]. With the expanding knowledge of cellular immunity mechanisms, the development of drugs, substances, or measures that modulate immune responses contributes to the management of many life-threatening infections [18-24]. Small clinical trials in the severe infection conditions explored the compassionate options to activate the early-responding immune effector cells through various immunoadjuvant agents, such as interleukin-7, anti-programmed death 1, interferon-y, and granulocyte-macrophage colony-stimulating factor [16,25]. The clinical management of severe infections commonly includes the modulation of immunosuppressive mechanisms, i.e., the alleviation of T-cell exhaustion, myeloid-derived suppressor cells, or regulatory T cells [16,21,24,26,27].

The balance between the immune activation and the counter-regulatory immunosuppression is crucial upon the virus-host encountering responses [28,29]. This balance determines the variation of subsequent

* Corresponding author at: Council for Nutritional and Environmental Medicine, Toften 24, 8610 Mo i Rana, Norway. E-mail address: bjorklund@conem.org (G. Bjørklund).

https://doi.org/10.1016/j.clim.2020.108545

Received 17 April 2020; Received in revised form 19 July 2020; Accepted 19 July 2020 Available online 22 July 2020



Review Article

^{1521-6616/ © 2020} Elsevier Inc. All rights reserved.



Fig. 1. The immunomodulation strategy and roles of a group of nutrients in different processes of virus-host immune responses.

clinical manifestations. Several micronutrients contribute to these immunomodulatory effects [16,30-34]. This article reviews some nutrients that can potentially modulate immunity to SARs-CoV-2 infection.

2. Nutrients and virus-host immunologic responses

Micronutrients involve in the continuum of host immune responses to the virus from the initial virus-host interaction, innate immune activation, to adaptive immune responses, as summarized in Fig. 1 [30,35]. The healthy immunity requires the synergistic contribution from multiple micronutrients, and single nutrient barely drives the whole immune machinery. However, the viral-host resistance relies on the support from a dominant group of nutrients, including vitamins A, C, D, E, B6, B12, folate, iron (Fe), zinc (Zn), copper (Zn), selenium (Se), and magnesium (Mg) [30,36–38].

The first-line defenses against the virus are the physical and biochemical barriers of the respiratory tract, which their normal epithelial differentiation and growth require vitamin A and Fe [36,37]. Vitamins A, C, D, and Zn regulate membrane fluidity, membrane integrity, gapjunction communication, and membrane repair [37,39–46]. Vitamin E mitigates the membrane lipid peroxidation from reactive oxygen species [37]. Vitamins A, D, C, and the trace elements Zn, Fe, Cu, and Se regulate the membrane-bounded antimicrobial peptide activities and mucosal-associated microbiota [30,47,48]. The mucosal migration and regulation of immune cell functions also synchronize with the integrated pathways of vitamins B6, B12, and folate [47,49].

Interferon (IFN) is a crucial anti-viral innate immune response than regulates and shapes the balance of Th1 and Th2 phenotypes in adaptive immunity [50]. IFN- λ s are key antiviral cytokines at the epithelial barriers, and induce the inflammatory response and apoptotic cell death [50,51]. Apart from that, type I IFNs increase in responses to the viral activation of the toll-like receptor 7 and the mitochondrial antiviral-signaling [52,53]. Vitamins A, C, D, C, Zn, Fe, Cu, and Se regulate IFNs production [30,36,44,45,54–58].

Upon the intrusion of SARS-CoV-2 into the airway epithelial cells, innate immune cells respond through their movement, migration, differentiation, proliferation, and activation to counteract the viral replication. The cytokines and oxidative burst induce the pro-inflammatory milieu, while the virus delays and suppress type I IFNs responses [35]. Without the optimal counter-regulatory immune reactions, the activation of the Th1/Th17 phenotypes of adaptive immunity further exacerbates the hyperinflammatory conditions and the 'cytokine storms' [27,35,59]. However, healthy immunity eventually proceeds to the production of SARS-CoV-2 specific antibodies that neutralize the virus and resolve the infection [59,60].

Vitamins A, C, D, E, B6, B12, and folate, and the trace elements Zn, Fe, Cu, Se, as well as the mineral Mg comprise a group of nutrients that support the entire continuum of virus-host immune responses. Their contributions range from the regulation of number and function of innate immune cells such as neutrophils, natural killer cells, monocytes, and macrophages [36,37,45,54,61–73], the production of pro-, and anti-inflammatory cytokines, the responses to inflammation, the oxidative burst function, the reductive-oxidative hemodynamics [36,37,45,61–64,71,72,74–85], to the responses of adaptive immunity, including differentiation, proliferation, and functions of T-cells [32,36,37,45,54,71,77,84,86–95], the interactions with the presenting viral antigens [37,54,71,73,96], and the production and development of virus-specific antibodies [36,37,45,71,73,97,98].

Despite their synergistic contributions to virus-host responses, the deficiency state of specific nutrients increases an individual susceptibility to the severe clinical manifestation of SARS-CoV-2 infection. The following sections explore the consequences of some micronutrient deficiencies and the potential effects of their supplementations to COVID-19.

3. Vitamins

3.1. Vitamin D

Vitamin D is involved in a wide range of immunomodulatory activities, including the maintenance of immune barrier integrity [40–44,47,48], the production of antimicrobial peptides [99–102], the support of monocytes, macrophages, and dendritic cells functions [36,37,62–64], the modulation of oxidative burst potential [37,62–64], the promotion of anti-inflammatory cytokine production [62,74–76], the inhibition of IFN γ [54–58], nuclear factor κ B [103], other proinflammatory cytokines [104,105], and the subsequent responses of adaptive immune cells [32,54,71,87–91].

The low level of vitamin D increases the risks, severity, morbidity,

Table 1

The immunomodulating properties, the risk for the deficiency states, and the impacts of supplementation of a group of nutrients.

Nutrients	Immunomodulating properties	Risks for deficiency states	Impacts of supplementation
Vitamin A	 maintaining the barrier integrity and normal differentiation of epithelial tissues [37,123] mucosal immune responses and acts as an anti-inflammatory agent [39,47,124,125] regulates the number and function of natural killer cells and supports the phagocytic and oxidative burst activities of macrophages [37,61] the Th1/Th2 phenotypic differentiation and development [37,86] downregulates IFN γ, interleukin 2, and tumor-necrosis factor α productions by Th1 cells, thus, maintains the normal antibody-mediated Th2 responses [36,37,45] 	 increased susceptibility to virus-induced respiratory tract infections, measles, and diarrhea [37,107,126,127] failed to mount the protective immunologic responses to the vaccine [128] 	 improves antibody titer responses to vaccines [37] Supplementation in vitamin A deficiency individuals reduced the incidence of Mycoplasma pneumoniae infection [129,130] The supplementation of vitamin A to deficient children decreased their risk of all-cause mortality and morbidity from infectious diseases. Nevertheless, vitamin A supplementation showed no benefits for pneumonia [107,131,132].
Vitamin C	 supports antibody production by B cells [37]. epithelial barrier integrity [37,45] innate immune cells activities, movement, functions, proliferation, and differentiation [36,37,45,54,61,65] antimicrobial activities; increases serum complement proteins, and the production of IFN γ [36,45] antioxidant, maintains the intracellular reductive-oxidative homeostasis [37,45,77] roles in antibody production and supports of differentiation and proliferation of T-cells 	 increased the risk and severity of several respiratory infections, including pneumonia [37,45,107,110,117] 	 shortens the symptoms of the common cold in children, reduces the incidence of pneumonia in the elderly [107,117–119] combination of vitamin C and red ginseng reduced the influenza virus-induced lung inflammation and increased the survival rate in mice [120]. high dose intravenous vitamin C shortens the recovery periods of severely ill patients with virus-induced acute respiratory distress syndrome [77,121,122]
Vitamin D	 [30,45] immune barrier integrity [40–44,47,48] production of antimicrobial peptides [99–102] support innate cells functions [36,37,62–64] modulation of oxidative burst [37,62–64] promotion of anti-inflammatory cytokines [62,74–76] inhibition IFN γ [54–58], nuclear factor κB [103], other pro-inflammatory cytokines [104,105] support adaptive immune cells [32,54,71,87–91] 	 increases risks, severity, morbidity, and mortality of several respiratory conditions, such as rhinitis, asthma, tuberculosis, chronic pulmonary disorders, viral respiratory infections, and potentially including the COVID-19 [106–110] deteriorates several pulmonary conditions [112–114] 	 reduced risk of respiratory infections [115,116] risk reduction benefit only in the vitamin D deficient individuals [61,107]
Vitamin E	 support adaptive inimitie cens (52,54,71,67791) a potent lipid-soluble antioxidant that protects the cell membranes against the oxidative damage and supports the integrity of respiratory epithelial barriers [37,133,134] enhances the natural killer cell cytotoxic activity and decreases prostaglandin E2 production by macrophages [36,37,54,61,66,78] modulates the production of IFN-γ and interleukin 2 [36,132,135] supports lymphocyte proliferation, T-cell-mediated functions, Th1 response optimization, and Th2 response suppression [36,37,61] supports the active immune synapses between Th cells require vitamin E supports [54] increases the proportion of antigen-experienced memory T-cells [96] 	• impairs the functions of both humoral and cell- mediated adaptive immunity, thus facilitates viral infection with high virulent strains, severe subsequent pathologies, and abnormal immune responses [71,132,133,136]	 improves overall immune functions, reduced respiratory tract infection incidences, severity, lower virus load in lung tissues, and increased the antibody titers, particularly in the elderly [37,107,135,137]
Zinc	 modulates the functions of approximately 2,000 enzymes and 750 transcription factors involving in various biological and physiological processes, including immunity, growth, and development [46,138] antiviral property; inhibits the RNA-dependent RNA polymerase enzyme [138,140,141] maintains the integrity of immune barriers [46] enhances the cytotoxic activity of natural killer cells and supports the cellular functions, growth, and differentiation of innate immune cells [37,54,61,67,68] involves in complement protein activities and the IFN γ production [36,45] modulation of the cytokine release and the Th17 and Th9 development [36,54,77,81–83,92,93] influences on the antioxidant proteins [37,77] supports the proliferation of Cytotoxic T cells, the differentiation, development, and activation of T-cells, the cytokine production of Th1 cells, and the development of regulatory T cells 	 increases the risk and morbidity of inflammatory disorders, infections, and viral pneumonia, particularly in the children and elderly persons [37,93,107,110,127,143,144] 	 Zinc supplementation in children reduced their susceptibilities, severity of symptoms, and the duration of common colds and viral pneumonia [77,107,138,145,146] increased serum Zn levels and the number of T-cells in nursing home elderly [147]

Table 1 (continued)

Nutrients	Immunomodulating properties	Risks for deficiency states	Impacts of supplementation
Selenium	 involves in antibody production, mainly the immunoglobulin G [37,97,98]. component of selenoproteins that are essential for the functions of the immune system and the 	 increases the risk and virulence of virus-induced pulmonary infections, particularly in infants during 	 maintaining optimal Se status protects against several viral infections [148–150]
	 reductive-oxidative homeostasis [61,148] modulates the activities of virus-induced innate and adaptive immunity through the regulation of IFN α, IFN γ, and IFN β production influences on the functions and differentiation of natural killer cells and T-cells, and the antibody production [36,45,71,84,95,149,150]. 	their first six weeks of life [107,132,150,151]	 reduced the pathogenicity of influenza virus infection in association with diets that contain both low and high Se quantities [152] enhances immune responses to the virus in deficit individuals [37,132]
Magnesium	 involves in nucleic acid metabolism, DNA replication, leukocyte activation, antigen-binding to macrophages, and apoptotic regulation [38,72,73] influences both the cell-mediated and humoral adaptive immunity [73,153] protect DNA from oxidative damages and reduce the superoxide anion production at high concentrations [72,85] 	 increases the susceptibility to recurrent upper respiratory tract infections [73,153,154] promotes chronic low-grade inflammation through the production of pro-inflammatory cytokines, acute-phase proteins, and free radicals [155] 	• Normal Mg levels maintain healthy lung structure and functions, and lower Mg levels are associated with increased respiratory complications [156,157]

and mortality of several respiratory conditions, such as rhinitis, asthma, tuberculosis, chronic pulmonary disorders, viral respiratory infections, and possibly also the COVID-19 [106–110]. The potential role of vitamin D in the modulation of immune response to viral respiratory tract infection (ALRI) has been evidenced in a study involving a young patient with individual genetic polymorphisms of vitamin D receptors [111]. Vitamin D influences lung structures, size, volume, and functions. Vitamin D deficiency thus worsens several pulmonary conditions [112–114].

A recent meta-analysis reported the associations of individuals with adequate vitamin D levels or daily oral supplementation with vitamin D and the reduced risk of respiratory infections [115,116]. The previous studies also suggested this risk reduction benefit of the supplementation, but only in the vitamin D deficient individuals [61,107]. With this information, vitamin D supplementation is a potential preventive strategy of COVID-19 in the individual with an established deficient state or has a high risk of vitamin D deficiency.

3.2. Vitamin C

Vitamin C supports the epithelial barrier integrity through its contributions to the collagen synthesis, keratinocyte differentiation, fibroblast migration, and proliferation [37,45]. Innate immune cells require vitamin C to maintain their activities, movement, functions, proliferation, and differentiation [36,37,45,54,61,65]. Vitamin C promotes the antimicrobial activities, increases serum complement proteins, and stimulated the production of IFN- γ [36,45]. Vitamin C is a powerful antioxidant, thus, maintains the intracellular reductive-oxidative homeostasis during the active immune responses [37,45,77]. It also plays roles in antibody production from plasma cells together with the supports of differentiation and proliferation of T-cells, particularly the cytotoxic T-cells [36,45].

Vitamin C deficiency increased the risk and severity of several respiratory infections, including pneumonia [37,45,107,110,117]. Despite many conflicting and inconclusive pieces of evidence, the oral supplementation of vitamin C potentially shortens the symptoms of the common cold in children. It also reduces the incidence of pneumonia in the elderly [107,117–119]. The combination of vitamin C and red ginseng reduced the influenza virus-induced lung inflammation and increased the survival rate in mice [120]. The treatment with high dose intravenous vitamin C shortens the recovery periods of severely ill patients with virus-induced acute respiratory distress syndrome [77,121,122]. Concerning its affordability, availability, and safety, vitamin C is still a functional option to consider in the management of COVID-19.

3.3. Vitamin A

Vitamin A is an essential micronutrient for maintaining the barrier integrity and normal differentiation of epithelial tissues [37,123]. It supports the mucosal immune responses and acts as an anti-in-flammatory agent [39,47,124,125]. Vitamin A regulates the number and function of natural killer cells and supports the phagocytic and oxidative burst activities of macrophages [37,61]. The Th1/Th2 phenotypic differentiation and development of T-cells require vitamin A [37,86]. It downregulates IFN- γ , interleukin 2, and tumor-necrosis factor α productions by Th1 cells, thus, maintains the normal antibody-mediated Th2 responses [36,37,45]. Vitamin A also supports antibody production by B cells [37].

Vitamin A deficiency is a common risk factor for the increased susceptibility to virus-induced respiratory tract infections, measles, and diarrhea [37,107,126,127]. Young cows with vitamin A deficiency failed to mount the protective immunologic responses to the BRSV-NP vaccine (amphiphilic polyanhydride nanoparticle-based vaccine encapsulating the fusion and attachment proteins from bovine respiratory syncytial virus), with the subsequent lung infections after challenging by the virus [128]. Vitamin A supplementation improves antibody titer responses to vaccines [37]. Supplementation of vitamin A to deficient individuals reduces the incidence of Mycoplasma pneumoniae infection, which is a common post-viral secondary bacterial infection in COVID-19 [129,130]. The supplementation in vitamin A deficit children, 6month to 5-year of age, decreased their risk of all-cause mortality and morbidity from infectious diseases. Nevertheless, vitamin A supplementation showed no benefits for pneumonia [107,131,132]. Concerning the potential adverse effects of vitamin A, the supplementation is sensible in the COVID-19 management of undernourished individuals or those with the evidence of vitamin A deficiency [132].

3.4. Vitamin E

Vitamin E is a potent lipid-soluble antioxidant that protects cell membranes against oxidative damage and supports the integrity of respiratory epithelial barriers [37,133,134]. It enhances the natural killer cell cytotoxic activity and decreases prostaglandin E2 production by macrophages [36,37,54,61,66,78]. Vitamin E modulates the production of IFN- γ and interleukin 2 [36,132,135]. It supports lymphocyte proliferation, T-cell-mediated functions, Th1 response optimization, and Th2 response suppression [36,37,61]. The active immune synapses between Th cells require vitamin E supports [54]. Vitamin E also increases the proportion of antigen-experienced memory T-cells [96]. Vitamin E deficiency is rare in humans. The deficit state impairs the functions of both humoral and cell-mediated adaptive immunity, thus facilitates the viral infection with high virulent strains, severe subsequent pathologies, and abnormal immune responses [71,132,133,136]. Vitamin E supplementation improves overall immune functions, reduces respiratory tract infection incidences, severity, lowers virus load in lung tissues, and increases the antibody titers, particularly in the elderly [37,107,135,137]. Malnourished individuals should benefit from the inclusion of vitamin E supplementation in COVID-19 management.

4. Essential trace elements and magnesium

4.1. Zinc

Zinc is an essential trace element that modulates the functions of approximately 2,000 enzymes and 750 transcription factors involved in various biological and physiological processes, including immunity, growth, and development [46,138,139]. Zinc also possesses a variety of direct and indirect antiviral properties. For instance, the pyrrolidine dithiocarbamate - a Zn ionophore - inhibits the RNA-dependent RNA polymerase enzyme that promotes SARS-CoV-2 replication [138,140,141]. Zinc maintains the integrity of immune barriers through its cofactor function in metalloenzymes [46,142]. It enhances the cytotoxic activity of natural killer cells and supports the cellular functions, growth, and differentiation of innate immune cells [37,54,61,67,68]. Zinc involves in complement protein activities and the IFN- γ production [36,45]. It has anti-inflammatory properties by the modulation of the cytokine release and the Th17 and Th9 development [36,54,77,81-83,92,93]. Zinc also exerts anti-oxidant effects through its influences on antioxidant proteins [37,77]. It supports the proliferation of cytotoxic T cells, the differentiation, development, and activation of T-cells, the cytokine production of Th1 cells, and the development of regulatory T cells [36,54,77,92-94]. Zinc is involved in antibody production, mainly the immunoglobulin G antibody [37,97,98].

Zinc deficiency increases the risk and morbidity of inflammatory disorders, infections, and viral pneumonia, particularly in children and the elderly [37,93,107,110,127,143,144]. Supplementation of Zn in children reduced their susceptibilities, severity of symptoms, and the duration of common colds and viral pneumonia [77,107,138,145,146]. Zinc supplementation in nursing home elderly increased their serum Zn levels and their number of T-cells [147]. Despite the few shreds of confirming evidence, Zn supplementation can benefit in the management of COVID-19, particularly in high-risk individuals for Zn deficiency.

4.2. Selenium

Selenium is a trace component of selenoproteins that are essential for the functions of the immune system and the reductive-oxidative homeostasis [61,148]. It modulates the activities of virus-induced innate and adaptive immunity through the regulation of IFN- α , IFN- γ , and IFN- β production, the influences on the functions and differentiation of natural killer cells and T-cells, and the antibody production [36,45,71,84,95,149,150].

Selenium deficiency increases the risk and virulence of virus-induced pulmonary infections through the aberrant immune responses and excessive cytokines production, particularly in infants during their first six weeks of life [107,132,150,151]. At the same time, the maintaining of optimal Se status through an adequate diet protects against several viral infections [148–150]. Dietary selenium supplementation potentiates innate antiviral immune responses reducing, for instance, the pathogenicity of avian influenza virus infection [152]. Consequently, Se supplementation in deficit individuals distinctively enhances the immune responses to the virus [37,132]. Selenium supplementation is the rationale management of COVID-19 in susceptible hosts.

4.3. Magnesium

Magnesium is a crucial mineral for healthy physiologic functions, including bioenergetics, immune responses, and acid-base balance; it is involved in nucleic acid metabolism, DNA replication, leukocyte activation, antigen-binding to macrophages, and apoptotic regulation [38,72,73]. Magnesium influences both the cell-mediated and humoral adaptive immunity [73,153]. It can protect DNA from oxidative damages and reduce the superoxide anion production at high concentrations [72,85]. Magnesium deficiency increases the susceptibility to recurrent upper respiratory tract infections [73,153,154]. A deficiency of Mg promotes chronic low-grade inflammation through the production of pro-inflammatory cytokines, acute-phase proteins, and free radicals [155]. Normal Mg levels maintain healthy lung structure and functions, while its lower levels are often associated with increased respiratory complications [156,157]. To date, no available study explores the impact of Mg supplementation on the clinical virus-induced respiratory infection.

5. Other potential immunomodulators for the COVID-19 management

5.1. N-acetylcysteine

N-acetylcysteine (NAC) is a precursor of glutathione that is a thiol reducing agent with antioxidant and anti-inflammatory properties. NAC reduces the elasticity and viscosity of mucus and improves the clearance of pulmonary secretions. NAC reduces oxidative stress and inflammation in chronic obstructive pulmonary disease patients. With the exposure to the influenza virus, NAC inhibits the production of TNF- α in alveolar macrophages, the expression of intercellular adhesion molecule 1 in respiratory epithelial cells, and increases the heme oxygenase 1 level in cells [158–160]. The combination of NAC and glutathione reduced the antigen levels of human immunosuppressive virus 1 and their reverse transcriptase activities in a cell line study [161]. A murine model study reported the synergistic actions of NAC and Oseltamivir combination in survival rate improvement- up to 100%- from the lethal strain of influenza infection [162].

The clinical application of NAC in patients with community-acquired pneumonia reported the reduction of oxidative stress and inflammation, as shown by the improved levels of TNF- α and malondialdehyde [163]. The long-term administration of NAC in elderly persons reduced the severity and duration of influenza-like symptoms [164]. Concerning the safety profile of NAC, it can be a sensible option to include in COVID-19 management despite a few pieces of clinical evidence.

5.2. Polyphenolic compounds

Polyphenolic compounds are a major class of phytonutrients with several biological and pharmacological properties, including antioxidant, anti-inflammatory, antibacterial, and antiviral potentials [165]. The span of the antiviral property of polyphenols involves the viruses from the *Coronaviriade* family. Resveratrol inhibits the Middle East Respiratory Syndrome coronavirus *in vitro* [166]. Anti-viral face masks and the cleaning wipes have their fiber filter surface grafted with the polyphenol catechin [167,168].

A recent computerized virtual screening of molecular structures identified six polyphenol molecules, i.e., sanguiin, theaflavin gallate, theaflavin digallate, kaempferol, punicalagin, and protocatechuic acid, that potentially target the main protease of SARS-CoV-2 [169]. Stilbene and flavonoid derivatives, such as herbacetin, isobavachalcone, quercetin $3-\beta$ -d-glucoside, and helichrysetin, inhibit 3C-like protease

[170,171]. Resveratrol inhibits the virus nucleocapsid protein synthesis, thus suppress viral replication [166]. Quercetin inhibits the helicase, an enzyme for viral replication [172]. Other polyphenols such as delphinidin and epigallocatechin gallate inhibit the entry attachment of the virus [173,174]. These polyphenols and flavonoids have potential effects on COVID-19 management. However, these pieces of evidence are all from *in vitro* studies; to date, there is no available *in vivo* study. It is then premature to conclude their clinical applications.

6. Concluding remarks

Due to the great impact on medical services and the massive demand for health care. COVID-19 rapidly turned into a global pandemic. posing a lethal threat to the population despite its low mortality rate. The clinical respiratory symptoms include dry cough, fever, anosmia, breathing difficulties, and subsequent respiratory failure. No known cure is available for COVID-19. Apart from the anti-viral strategy, the supports of immune effectors and modulation of immunosuppression is the rationale immunomodulation approach in COVID-19 management. Diet and nutrition are essential for healthy immunity, but a group of micronutrients somehow plays a dominant role in immunomodulation [175]. This paper reviews the mechanisms, the effects of their deficiency states, and the potential impacts of their supplementations in COVID-19, as summarized in Table 1. The deficiency states of most nutrients increase the individual susceptibility to virus infection with a tendency for severe clinical presentation. Despite a shred of evidence, the supplementation of a single nutrient is not promising in the general population. The high-risk individual of a specific micronutrient deficiency is likely to benefit from the supplementation. The individual dietary and nutritional status assessments are critical for determining the comprehensive actions in COVID-19.

References

- D.N. Valencia, Brief review on COVID-19: the 2020 pandemic caused by SARS-CoV-2, Lancet 12 (2020), https://doi.org/10.7759/cureus.7386 e7386.
- [2] L. Lin, L. Lu, W. Cao, T. Li, Hypothesis for potential pathogenesis of SARS-CoV-2 infection-a review of immune changes in patients with viral pneumonia, Emerg. Microb. Infect. 9 (2020) 727–732, https://doi.org/10.1080/22221751.2020. 1746199.
- [3] C. Huang, Y. Wang, X. Li, L. Ren, J. Zhao, Y. Hu, L. Zhang, G. Fan, J. Xu, X. Gu, Z. Cheng, T. Yu, J. Xia, Y. Wei, W. Wu, X. Xie, W. Yin, H. Li, M. Liu, Y. Xiao, H. Gao, L. Guo, J. Xie, G. Wang, R. Jiang, Z. Gao, Q. Jin, J. Wang, B. Cao, Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China, Lancet 395 (2020) 497–506, https://doi.org/10.1016/S0140-6736(20)30183-5.
- [4] F.A. Rabi, M.S. Al Zoubi, G.A. Kasasbeh, D.M. Salameh, A.D. Al-Nasser, SARS-CoV-2 and coronavirus disease 2019: what we know so far, Pathogens 9 (2020) 231.
- [5] K. Dhama, K. Sharun, R. Tiwari, M. Dadar, Y.S. Malik, K.P. Singh, W. Chaicumpa, COVID-19, an emerging coronavirus infection: advances and prospects in designing and developing vaccines, immunotherapeutics, and therapeutics, Hum. Vaccin. Immunother. (2020) 1–7.
- [6] W.-j. Guan, Z.-y. Ni, Y. Hu, W.-h. Liang, C.-q. Ou, J.-x. He, L. Liu, H. Shan, C.-l. Lei, D.S.C. Hui, B. Du, L.-j. Li, G. Zeng, K.-Y. Yuen, R.-c. Chen, C.-l. Tang, T. Wang, P.-y. Chen, J. Xiang, S.-y. Li, J.-l. Wang, Z.-j. Liang, Y.-x. Peng, L. Wei, Y. Liu, Y.-h. Hu, P. Peng, J.-m. Wang, J.-y. Liu, Z. Chen, G. Li, Z.-j. Zheng, S.-q. Qiu, J. Luo, C.-j. Ye, S.-y. Zhu, N.-s. Zhong, Clinical characteristics of coronavirus disease 2019 in China, N. Engl. J. Med. (2020), https://doi.org/10.1056/NEJMoa2002032.
- M. Day, Covid-19: four fifths of cases are asymptomatic, China figures indicate, BMJ 369 (2020) m1375, https://doi.org/10.1136/bmj.m1375.
- [8] S.R. Bornstein, R. Dalan, D. Hopkins, G. Mingrone, B.O. Boehm, Endocrine and metabolic link to coronavirus infection, Nat. Rev. Endocrinol. (2020), https://doi. org/10.1038/s41574-020-0353-9.
- [9] C.M. Hedrich, COVID-19 considerations for the paediatric rheumatologist, Clin. Immunol. 214 (2020) 108420, https://doi.org/10.1016/j.clim.2020.108420.
- [10] K. Yuki, M. Fujiogi, S. Koutsogiannaki, COVID-19 pathophysiology: a review, Clin. Immunol. 215 (2020) 108427, https://doi.org/10.1016/j.clim.2020.108427.
- [11] F. Ciceri, A. Castagna, P. Rovere-Querini, F. De Cobelli, A. Ruggeri, L. Galli, C. Conte, R. De Lorenzo, A. Poli, A. Ambrosio, C. Signorelli, E. Bossi, M. Fazio, C. Tresoldi, S. Colombo, G. Monti, E. Fominskiy, S. Franchini, M. Spessot, C. Martinenghi, M. Carlucci, L. Beretta, A.M. Scandroglio, M. Clementi, M. Locatelli, M. Tresoldi, P. Scarpellini, G. Martino, E. Bosi, L. Dagna, A. Lazzarin, G. Landoni, A. Zangrillo, Early predictors of clinical outcomes of COVID-19 outbreak in Milan, Italy, Clin. Immunol. 217 (2020) 108509, https://doi.org/10. 1016/j.clim.2020.108509.
- [12] J.M. Urra, C.M. Cabrera, L. Porras, I. Rodenas, Selective CD8 cell reduction by

SARS-CoV-2 is associated with a worse prognosis and systemic inflammation in COVID-19 patients, Clin. Immunol. 217 (2020) 108486, https://doi.org/10. 1016/j.clim.2020.108486.

- [13] Y. Wang, Y. Liu, L. Liu, X. Wang, N. Luo, L. Ling, Clinical outcome of 55 asymptomatic cases at the time of hospital admission infected with SARS-Coronavirus-2 in Shenzhen, China, J. Infect. Dis. (2020), https://doi.org/10.1093/infdis/jiaa119.
- [14] Z. Hu, C. Song, C. Xu, G. Jin, Y. Chen, X. Xu, H. Ma, W. Chen, Y. Lin, Y. Zheng, J. Wang, Z. Hu, Y. Yi, H. Shen, Clinical characteristics of 24 asymptomatic infections with COVID-19 screened among close contacts in Nanjing, China, Sci. China Life Sci. (2020), https://doi.org/10.1007/s11427-020-1661-4.
- [15] M. Kloc, R.M. Ghobrial, E. Kuchar, S. Lewicki, J.Z. Kubiak, Development of child immunity in the context of COVID-19 pandemic, Clin. Immunol. 217 (2020) 108510, https://doi.org/10.1016/j.clim.2020.108510.
- [16] R.S. Hotchkiss, S.M. Opal, Activating immunity to fight a foe a new path, N. Engl. J. Med. 382 (2020) 1270–1272, https://doi.org/10.1056/ NEJMcibr1917242.
- [17] S. Felsenstein, J.A. Herbert, P.S. McNamara, C.M. Hedrich, COVID-19: Immunology and treatment options, Clin. Immunol. 215 (2020) 108448, https:// doi.org/10.1016/j.clim.2020.108448.
- [18] M.C. Amezcua Vesely, P. Pallis, P. Bielecki, J.S. Low, J. Zhao, C.C.D. Harman, L. Kroehling, R. Jackson, W. Bailis, P. Licona-Limón, H. Xu, N. Iijima, P.S. Pillai, D.H. Kaplan, C.T. Weaver, Y. Kluger, M.S. Kowalczyk, A. Iwasaki, J.P. Pereira, E. Esplugues, N. Gagliani, R.A. Flavell, Effector T(H)17 Cells Give Rise to Long-Lived T(RM) Cells that Are Essential for an Immediate Response against Bacterial Infection, Cell 178 (2019) 1176–1188 e1115 https://doi.org/10.1016/j.cell.2019. 07.032.
- [19] R. Zander, D. Schauder, G. Xin, C. Nguyen, X. Wu, A. Zajac, W. Cui, CD4(+) T cell help is required for the formation of a cytolytic CD8(+) T cell subset that protects against chronic infection and cancer, Immunity 51 (2019) 1028–1042.e1024, https://doi.org/10.1016/j.immuni.2019.10.009.
- [20] S.M. Opal, Non-antibiotic treatments for bacterial diseases in an era of progressive antibiotic resistance, Criti. Care (London, England) 20 (2016) 397, https://doi. org/10.1186/s13054-016-1549-1.
- [21] P. Conti, G. Ronconi, A. Caraffa, C.E. Gallenga, R. Ross, I. Frydas, S.K. Kritas, Induction of pro-inflammatory cytokines (IL-1 and IL-6) and lung inflammation by Coronavirus-19 (COVI-19 or SARS-CoV-2): anti-inflammatory strategies, J. Biol. Regul. Homeost. Agents 34 (2020), https://doi.org/10.23812/conti-e.
- [22] B. Vellingiri, K. Jayaramayya, M. Iyer, A. Narayanasamy, V. Govindasamy, B. Giridharan, S. Ganesan, A. Venugopal, D. Venkatesan, H. Ganesan, K. Rajagopalan, P. Rahman, S.G. Cho, N.S. Kumar, M.D. Subramaniam, COVID-19: A promising cure for the global panic, Sci. Total Environ. 725 (2020) 138277, , https://doi.org/10.1016/j.scitotenv.2020.138277.
- [23] L. Lin, S. Luo, R. Qin, M. Yang, X. Wang, Q. Yang, Y. Zhang, Q. Wang, R. Zhu, H. Fan, H. Wang, Y. Hu, L. Wang, D. Hu, Long-term infection of SARS-CoV-2 changed the body's immune status, Clin. Immunol. (2020) 108524, https://doi. org/10.1016/j.clim.2020.108524.
- [24] W. Zhang, Y. Zhao, F. Zhang, Q. Wang, T. Li, Z. Liu, J. Wang, Y. Qin, X. Zhang, X. Yan, X. Zeng, S. Zhang, The use of anti-inflammatory drugs in the treatment of people with severe coronavirus disease 2019 (COVID-19): the perspectives of clinical immunologists from China, Clin. Immunol. 214 (2020) 108393, https:// doi.org/10.1016/j.clim.2020.108393.
- [25] B. Francois, R. Jeannet, T. Daix, A.H. Walton, M.S. Shotwell, J. Unsinger, G. Monneret, T. Rimmelé, T. Blood, M. Morre, A. Gregoire, G.A. Mayo, J. Blood, S.K. Durum, E.R. Sherwood, R.S. Hotchkiss, Interleukin-7 restores lymphocytes in septic shock: the IRIS-7 randomized clinical trial, JCI Insight 3 (2018) e98960, , https://doi.org/10.1172/jci.insight.98960.
- [26] R.S. Hotchkiss, L.L. Moldawer, S.M. Opal, K. Reinhart, I.R. Turnbull, J.L. Vincent, Sepsis and septic shock, Nat. Rev. Dis. Primers 2 (2016) 16045, https://doi.org/ 10.1038/nrdp.2016.45.
- [27] P. Mehta, D.F. McAuley, M. Brown, E. Sanchez, R.S. Tattersall, J.J. Manson, COVID-19: consider cytokine storm syndromes and immunosuppression, Lancet 395 (2020) 1033–1034, https://doi.org/10.1016/S0140-6736(20)30628-0.
- [28] B.A. Kane, K.J. Bryant, H.P. McNeil, N.T. Tedla, Termination of immune activation: an essential component of healthy host immune responses, J. Innate Immun. 6 (2014) 727–738, https://doi.org/10.1159/000363449.
- [29] Y.S. Malik, S. Sircar, S. Bhat, K. Sharun, K. Dhama, M. Dadar, R. Tiwari, W. Chaicumpa, Emerging novel coronavirus (2019-nCoV)—current scenario, evolutionary perspective based on genome analysis and recent developments, Vet. Q. 40 (2020) 68–76.
- [30] A.F. Gombart, A. Pierre, S. Maggini, A review of micronutrients and the immune system-working in harmony to reduce the risk of infection, Nutrients 12 (2020) 236, https://doi.org/10.3390/nu12010236.
- [31] A. Gasmi, S. Noor, T. Tippairote, M. Dadar, A. Menzel, G. Bjorklund, Individual risk management strategy and potential therapeutic options for the COVID-19 pandemic, Clin. Immunol. 215 (2020) 108409, https://doi.org/10.1016/j.clim. 2020.108409.
- [32] F. Sassi, C. Tamone, P. D'Amelio, Vitamin D: nutrient, hormone, and immunomodulator, Nutrients 10 (2018), https://doi.org/10.3390/nu10111656.
- [33] J. Sun, P. de Vos, Editorial: immunomodulatory functions of nutritional ingredients in health and disease, Front. Immunol. 10 (2019), https://doi.org/10. 3389/fimmu.2019.00050.
- [34] T. Goswami, R. Bhar, S.E. Jadhav, S. Joardar, G.C. Ram, Role of dietary zinc as a nutritional immunomodulator, Asian Australas. J. Anim. Sci. 18 (2005), https:// doi.org/10.5713/ajas.2005.439.
- [35] E. Prompetchara, C. Ketloy, T. Palaga, Immune responses in COVID-19 and potential vaccines: Lessons learned from SARS and MERS epidemic, Asian Pac. J.

Allergy Immunol. 38 (2020) 1-9, https://doi.org/10.12932/ap-200220-0772.

- [36] B. Haryanto, T. Suksmasari, E. Wintergerst, S. Maggini, Bayer, Multivitamin supplementation supports immune function and ameliorates conditions triggered by reduced air quality, Vitam. Min. 4 (2015), https://doi.org/10.4172/2376-1318. 1000128.
- [37] S. Maggini, S. Beveridge, P.J.P. Sorbara, G. Senatore, Feeding the immune system: the role of micronutrients in restoring resistance to infections, CAB reviews: perspectives in agriculture, Vet. Sci. Nutr. Nat. Resour. 3 (2008) 1–21, https://doi. org/10.1079/PAVSNNR20083098.
- [38] M.A. Zoroddu, J. Aaseth, G. Crisponi, S. Medici, M. Peana, V.M. Nurchi, The essential metals for humans: a brief overview, J. Inorg. Biochem. 195 (2019) 120–129, https://doi.org/10.1016/j.jinorgbio.2019.03.013.
- [39] B.P. Chew, J.S. Park, Carotenoid action on the immune response, J. Nutr. Biochem. 134 (2004) 257S–261S, https://doi.org/10.1093/jn/134.1.257S.
- [40] A. Clairmont, D. Tessman, A. Stock, S. Nicolai, W. Stahl, H. Sies, Induction of gap junctional intercellular communication by vitamin D in human skin fibroblasts is dependent on the nuclear Induction of gap junctional intercellular communication by vitamin D in human skin fibroblasts is dependent on the nuclear vitamin D receptor, Carcinogenesis 17 (1996) 1389–1391, https://doi.org/10.1093/carcin/ 17.6.1389.
- [41] R. Gniadecki, B. Gajkowska, M. Hansen, 1,25-dihydroxyvitamin D3 stimulates the assembly of adherens junctions in keratinocytes: involvement of protein kinase C, Endocrinology 138 (1997) 2241–2248, https://doi.org/10.1210/endo.138.6. 5156.
- [42] H.G. Pálmer, J.M. González-Sancho, J. Espada, M.T. Berciano, I. Puig, J. Baulida, M. Quintanilla, A. Cano, A.G. de Herreros, M. Lafarga, A. Muñoz, Vitamin D(3) promotes the differentiation of colon carcinoma cells by the induction of E-cadherin and the inhibition of beta-catenin signaling, J. Cell Biol. 154 (2001) 369–387, https://doi.org/10.1083/jcb.200102028.
- [43] M. Mihajlovic, M. Fedecostante, M.J. Oost, S.K.P. Steenhuis, E.G.W.M. Lentjes, I. Maitimu-Smeele, M.J. Janssen, L.B. Hilbrands, R. Masereeuw, Role of vitamin D in maintaining renal epithelial barrier function in uremic conditions, Int. J. Mol. Sci. 18 (2017) 2531, https://doi.org/10.3390/ijms18122531.
- [44] Z. Yin, V. Pintea, Y. Lin, B.D. Hammock, M.A. Watsky, Vitamin D enhances corneal epithelial barrier function, Invest. Ophthalmol. Vis. Sci. 52 (2011) 7359–7364, https://doi.org/10.1167/iovs.11-7605.
- [45] A.C. Carr, S. Maggini, Vitamin C and immune function, Nutrients 9 (2017), https://doi.org/10.3390/nu9111211.
- [46] P.-H. Lin, M. Sermersheim, H. Li, P.H.U. Lee, S.M. Steinberg, J. Ma, Zinc in wound healing modulation, Nutrients 10 (2017) 16, https://doi.org/10.3390/ nu10010016.
- [47] H.K. Biesalski, Nutrition meets the microbiome: micronutrients and the microbiota, Ann. N. Y. Acad. Sci. 1372 (2016) 53–64, https://doi.org/10.1111/nyas. 13145.
- [48] A. Clark, N. Mach, Role of vitamin D in the hygiene hypothesis: the interplay between vitamin D, vitamin D receptors, gut microbiota, and immune response, Front. Immunol. 7 (2016) 627, https://doi.org/10.3389/fimmu.2016.00627.
- [49] K. Yoshii, K. Hosomi, K. Sawane, J. Kunisawa, Metabolism of dietary and microbial vitamin B family in the regulation of host immunity, Front. Nutr. 6 (2019) 48, https://doi.org/10.3389/fnut.2019.00048.
- [50] J. Zhou, Y. Wang, Q. Chang, P. Ma, Y. Hu, X. Cao, Type III interferons in viral infection and antiviral immunity, Cell. Physiol. Biochem. 51 (2018) 173–185, https://doi.org/10.1159/000495172.
- [51] S.H. Lee, J.Y. Kwon, S.-Y. Kim, K. Jung, M.-L. Cho, Interferon-gamma regulates inflammatory cell death by targeting necroptosis in experimental autoimmune arthritis, Sci. Rep. 7 (2017) 10133, https://doi.org/10.1038/s41598-017-09767-0.
- [52] M.F. McCarty, J.J. DiNicolantonio, Nutraceuticals have potential for boosting the type 1 interferon response to RNA viruses including influenza and coronavirus, Prog. Cardiovasc. Dis. (2020), https://doi.org/10.1016/j.pcad.2020.02.007.
- [53] B. Huang, J. Li, X. Zhang, Q. Zhao, M. Lu, Y. Lv, RIG-1 and MDA-5 signaling pathways contribute to IFN-β production and viral replication in porcine circovirus virus type 2-infected PK-15 cells in vitro, Vet. Microbiol. 211 (2017) 36–42, https://doi.org/10.1016/j.vetmic.2017.09.022.
- [54] D. Wu, E.D. Lewis, M. Pae, S.N. Meydani, Nutritional modulation of immune function: analysis of evidence, mechanisms, and clinical relevance, Front. Immunol. 9 (2018) 3160, https://doi.org/10.3389/fimmu.2018.03160.
- [55] T. Matsui, R. Takahashi, Y. Nakao, T. Koizumi, Y. Katakami, K. Mihara, T. Sugiyama, T. Fujita, 1,25-Dihydroxyvitamin D3-regulated expression of genes involved in human T-lymphocyte proliferation and differentiation, Cancer Res. 46 (1986) 5827–5831.
- [56] H. Reichel, H.P. Koeffler, A. Tobler, A.W. Norman, 1 alpha,25-Dihydroxyvitamin D3 inhibits gamma-interferon synthesis by normal human peripheral blood lymphocytes, Proceedings of the National Academy of Sciences of the United States of America, 84 1987, pp. 3385–3389, https://doi.org/10.1073/pnas.84.10.3385.
- [57] W.F. Rigby, S. Denome, M.W. Fanger, Regulation of lymphokine production and human T lymphocyte activation by 1,25-dihydroxyvitamin D3. Specific inhibition at the level of messenger RNA, J. Clin. Invest. 79 (1987) 1659–1664, https://doi. org/10.1172/jci113004.
- [58] M. Inoue, T. Matsui, A. Nishibu, Y. Nihei, K. Iwatsuki, F. Kaneko, Regulatory effects of 1alpha,25-dihydroxyvitamin D3 on inflammatory responses in psoriasis, Eur. J. Dermatol. 8 (1998) 16–20.
- [59] Y. Shi, Y. Wang, C. Shao, J. Huang, J. Gan, X. Huang, E. Bucci, M. Piacentini, G. Ippolito, G. Melino, COVID-19 infection: the perspectives on immune responses, Cell Death Differ. (2020), https://doi.org/10.1038/s41418-020-0530-3.
- [60] B. Shanmugaraj, K. Siriwattananon, K. Wangkanont, W. Phoolcharoen,

Perspectives on monoclonal antibody therapy as potential therapeutic intervention for Coronavirus disease-19 (COVID-19), Asian Pac. J. Allergy Immunol. 38 (2020) 10–18, https://doi.org/10.12932/ap-200220-0773.

- [61] S. Maggini, A. Pierre, P.C. Calder, Immune function and micronutrient requirements change over the life course, Nutrients 10 (2018) 1531, https://doi.org/10. 3390/nu10101531.
- [62] K. Wishart, Increased Micronutrient Requirements during Physiologically Demanding Situations: Review of the Current Evidence, (2017), https://doi.org/ 10.4172/2376-1318.1000166.
- [63] L.M. Sly, M. Lopez, W.M. Nauseef, N.E. Reiner, 1alpha,25-Dihydroxyvitamin D3induced monocyte antimycobacterial activity is regulated by phosphatidylinositol 3-kinase and mediated by the NADPH-dependent phagocyte oxidase, J. Biol. Chem. 276 (2001) 35482–35493, https://doi.org/10.1074/jbc.M102876200.
- [64] H. Tanaka, K.A. Hruska, Y. Seino, J.D. Malone, Y. Nishii, S.L. Teitelbaum, Disassociation of the macrophage-maturational effects of vitamin D from re spiratory burst priming, J. Biol. Chem. 266 (1991) 10888–10892.
- [65] S.M. Bozonet, A.C. Carr, The role of physiological vitamin C concentrations on key functions of neutrophils isolated from healthy individuals, Nutrients 11 (2019) 1363, https://doi.org/10.3390/nu11061363.
- [66] D. Wu, S.N. Meydani, Age-associated changes in immune function: impact of vitamin E intervention and the underlying mechanisms, Endocr Metab Immune Disord Drug Targets 14 (2014) 283–289, https://doi.org/10.2174/ 1871530314666140922143950.
- [67] H. Gao, W. Dai, L. Zhao, J. Min, F. Wang, The role of zinc and zinc homeostasis in macrophage function, J Immunol Res 2018 (2018) 6872621, https://doi.org/10. 1155/2018/6872621.
- [68] A. Sheikh, S. Shamsuzzaman, S.M. Ahmad, D. Nasrin, S. Nahar, M.M. Alam, A. Al Tarique, Y.A. Begum, S.S. Qadri, M.I. Chowdhury, A. Saha, C.P. Larson, F. Qadri, Zinc influences innate immune responses in children with enterotoxigenic Escherichia coli-induced diarrhea, J. Nutr. Biochem. 140 (2010) 1049–1056, https://doi.org/10.3945/jn.109.111492.
- [69] R. Agoro, M. Taleb, V.F.J. Quesniaux, C. Mura, Cell iron status influences macrophage polarization, PLoS One 13 (2018) e0196921, https://doi.org/10.1371/ journal.pone.0196921.
- [70] A.N. Besold, E.M. Culbertson, V.C. Culotta, The Yin and Yang of copper during infection, J. Biol. Inorg. Chem. 21 (2016) 137–144, https://doi.org/10.1007/ s00775-016-1335-1.
- [71] F. Saeed, M. Nadeem, R.S. Ahmed, M. Tahir Nadeem, M.S. Arshad, A. Ullah, Studying the impact of nutritional immunology underlying the modulation of immune responses by nutritional compounds – a review, Food Agric. Immunol. 27 (2016) 205–229, https://doi.org/10.1080/09540105.2015.1079600.
- [72] J. Petrović, D. Stanić, G. Dmitrašinović, B. Plećaš-Solarović, S. Ignjatović, B. Batinić, D. Popović, V. Pešić, Magnesium supplementation diminishes peripheral blood lymphocyte DNA oxidative damage in athletes and sedentary young man, Oxidative Med. Cell. Longev. (2016) (2016) 2019643, https://doi.org/10. 1155/2016/2019643.
- [73] M.J. Laires, C. Monteiro, Exercise, magnesium and immune function, Magnes. Res. 21 (2008) 92–96.
- [74] Z. Lin, W. Li, The roles of vitamin D and its analogs in inflammatory diseases, Curr. Top. Med. Chem. 16 (2016) 1242–1261, https://doi.org/10.2174/ 1568026615660150915111557
- [75] Y. Zhang, D.Y.M. Leung, B.N. Richers, Y. Liu, L.K. Remigio, D.W. Riches, E. Goleva, Vitamin D inhibits monocyte/macrophage proinflammatory cytokine production by targeting MAPK phosphatase-1, J. Immunol. 188 (2012) 2127–2135, https://doi.org/10.4049/jimmunol.1102412.
- [76] I. Topilski, L. Flaishon, Y. Naveh, A. Harmelin, Y. Levo, I. Shachar, The anti-inflammatory effects of 1,25-dihydroxyvitamin D3 on Th2 cells in vivo are due in part to the control of integrin-mediated T lymphocyte homing, Eur. J. Immunol. 34 (2004) 1068–1076, https://doi.org/10.1002/eji.200324532.
- [77] E.S. Wintergerst, S. Maggini, D.H. Hornig, Immune-enhancing role of vitamin C and zinc and effect on clinical conditions, Ann. Nutr. Metab. 50 (2006) 85–94, https://doi.org/10.1159/000090495.
- [78] G.Y. Lee, S.N. Han, The role of vitamin E in immunity, Nutrients 10 (2018) 1614, https://doi.org/10.3390/nu10111614.
- [79] L. Sakakeeny, R. Roubenoff, M. Obin, J.D. Fontes, E.J. Benjamin, Y. Bujanover, P.F. Jacques, J. Selhub, Plasma pyridoxal-5-phosphate is inversely associated with systemic markers of inflammation in a population of U.S. adults, J. Nutr. Biochem. 142 (2012) 1280–1285, https://doi.org/10.3945/jn.111.153056.
- [80] P.M. Ueland, A. McCann, Ø. Midttun, A. Ulvik, Inflammation, vitamin B6 and related pathways, Mol. Asp. Med. 53 (2017) 10–27, https://doi.org/10.1016/j. mam.2016.08.001.
- [81] M. Jarosz, M. Olbert, G. Wyszogrodzka, K. Młyniec, T. Librowski, Antioxidant and anti-inflammatory effects of zinc. Zinc-dependent NF-κB signaling, Inflammopharmacology 25 (2017) 11–24, https://doi.org/10.1007/s10787-017-0309-4.
- [82] M. Foster, S. Samman, Zinc and regulation of inflammatory cytokines: implications for cardiometabolic disease, Nutrients 4 (2012) 676–694, https://doi.org/10. 3390/nu4070676.
- [83] I. Wessels, L. Rink, Micronutrients in autoimmune diseases: possible therapeutic benefits of zinc and vitamin D, J. Nutr. Biochem. 77 (2020) 108240, https://doi. org/10.1016/j.jnutbio.2019.108240.
- [84] P.T. Alpert, The role of vitamins and minerals on the immune system, Home Health Care Manag. Pract. 29 (2017) 199–202, https://doi.org/10.1177/ 1084822317713300.
- [85] F.I. Bussière, A. Mazur, J.L. Fauquert, A. Labbe, Y. Rayssiguier, A. Tridon, High magnesium concentration in vitro decreases human leukocyte activation, Magnes

Res. 15 (2002) 43-48.

- [86] A.C. Ross, Vitamin A and retinoic acid in T cell-related immunity, Am. J. Clin. Nutr. 96 (2012) 1166s–1172s, https://doi.org/10.3945/ajcn.112.034637.
- [87] H. Sigmundsdottir, J. Pan, G.F. Debes, C. Alt, A. Habtezion, D. Soler, E.C. Butcher, DCs metabolize sunlight-induced vitamin D3 to 'program' T cell attraction to the epidermal chemokine CCL27, Nat. Immunol. 8 (2007) 285–293, https://doi.org/ 10.1038/ni1433.
- [88] M.T. Cantorna, L. Snyder, Y.-D. Lin, L. Yang, Vitamin D and 1,25(OH)2D regulation of T cells, Nutrients 7 (2015) 3011–3021, https://doi.org/10.3390/ nu7043011.
- [89] G. Penna, L. Adorini, 1 Alpha,25-dihydroxyvitamin D3 inhibits differentiation, maturation, activation, and survival of dendritic cells leading to impaired alloreactive T cell activation, J. Immunol. 164 (2000) 2405–2411, https://doi.org/10. 4049/immunol.164.5.2405.
- [90] L. Piemonti, P. Monti, M. Sironi, P. Fraticelli, B.E. Leone, E. Dal Cin, P. Allavena, V. Di Carlo, Vitamin D3 affects differentiation, maturation, and function of human monocyte-derived dendritic cells, J. Immunol. 164 (2000) 4443–4451, https:// doi.org/10.4049/jimmunol.164.9.4443.
- [91] M. Bscheider, E.C. Butcher, Vitamin D immunoregulation through dendritic cells, Immunology 148 (2016) 227–236, https://doi.org/10.1111/imm.12610.
- [92] C. Kitabayashi, T. Fukada, M. Kanamoto, W. Ohashi, S. Hojyo, T. Atsumi, N. Ueda, I. Azuma, H. Hirota, M. Murakami, T. Hirano, Zinc suppresses Th17 development via inhibition of STAT3 activation, Int. Immunol. 22 (2010) 375–386, https://doi. org/10.1093/intimm/dxq017.
- [93] M. Maywald, F. Wang, L. Rink, Zinc supplementation plays a crucial role in T helper 9 differentiation in allogeneic immune reactions and non-activated T cells, J. Trace Elem. Med. Biol. 50 (2018) 482–488, https://doi.org/10.1016/j.jtemb. 2018.02.004.
- [94] E.S. Wintergerst, S. Maggini, D.H. Hornig, Contribution of selected vitamins and trace elements to immune function, Ann. Nutr. Metab. 51 (2007) 301–323, https://doi.org/10.1159/000107673.
- [95] B.E. Hurwitz, J.R. Klaus, M.M. Llabre, A. Gonzalez, P.J. Lawrence, K.J. Maher, J.M. Greeson, M.K. Baum, G. Shor-Posner, J.S. Skyler, N. Schneiderman, Suppression of human immunodeficiency virus type 1 viral load with selenium supplementation: a randomized controlled trial, Arch. Intern. Med. 167 (2007) 148–154, https://doi.org/10.1001/archinte.167.2.148.
- [96] S.N. Han, O. Adolfsson, C.K. Lee, T.A. Prolla, J. Ordovas, S.N. Meydani, Vitamin E and gene expression in immune cells, Ann. N. Y. Acad. Sci. 1031 (2004) 96–101, https://doi.org/10.1196/annals.1331.010.
- [97] A.H. Shankar, A.S. Prasad, Zinc and immune function: the biological basis of altered resistance to infection, Am. J. Clin. Nutr. 68 (1998) 447s-463s, https://doi. org/10.1093/ajcn/68.2.447S.
- [98] L. Rink, H. Kirchner, Zinc-altered immune function and cytokine production, J. Nutr. 130 (2000) 1407s–1411s, https://doi.org/10.1093/jn/130.5.1407S.
- [99] T.T. Wang, F.P. Nestel, V. Bourdeau, Y. Nagai, Q. Wang, J. Liao, L. Tavera-Mendoza, R. Lin, J.W. Hanrahan, S. Mader, J.H. White, Cutting edge: 1,25-dihydroxyvitamin D3 is a direct inducer of antimicrobial peptide gene expression, J. Immunol. 173 (2004) 2909–2912, https://doi.org/10.4049/jimmunol.173.5. 2909.
- [100] A.F. Gombart, N. Borregaard, H.P. Koeffler, Human cathelicidin antimicrobial peptide (CAMP) gene is a direct target of the vitamin D receptor and is strongly upregulated in myeloid cells by 1,25-dihydroxyvitamin D3, FASEB J. (2005) 1067–1077, https://doi.org/10.1096/fj.04-3284com.
- [101] G. Weber, J.D. Heilborn, C.I. Chamorro Jimenez, A. Hammarsjo, H. Törmä, M. Stahle, Vitamin D induces the antimicrobial protein hCAP18 in human skin, J. Invest. Dermatol. 124 (2005) 1080–1082, https://doi.org/10.1111/j.0022-202X. 2005.23687.x.
- [102] A.F. Gombart, The vitamin D-antimicrobial peptide pathway and its role in protection against infection, Future Microbiol. 4 (2009) 1151–1165, https://doi.org/ 10.2217/fmb.09.87.
- [103] S. Hansdottir, M.M. Monick, N. Lovan, L. Powers, A. Gerke, G.W. Hunninghake, Vitamin D decreases respiratory syncytial virus induction of NF-kappaB-linked chemokines and cytokines in airway epithelium while maintaining the antiviral state, J. Immunol. 184 (2010) 965–974, https://doi.org/10.4049/jimmunol. 0902840.
- [104] D.A. Hughes, R. Norton, Vitamin D and respiratory health, Clin. Exp. Immunol. 158 (2009) 20–25, https://doi.org/10.1111/j.1365-2249.2009.04001.x.
- [105] S. Hansdottir, M.M. Monick, S.L. Hinde, N. Lovan, D.C. Look, G.W. Hunninghake, Respiratory epithelial cells convert inactive vitamin D to its active form: potential effects on host defense, J. Immunol. 181 (2008) 7090–7099, https://doi.org/10. 4049/jimmunol.181.10.7090.
- [106] E. Laird, R.A. Kenny, Vitamin D deficiency in Ireland –implications for COVID-19. Results from the Irish Longitudinal Study on Ageing (TILDA), Irish Longitud. Study Age. (2020), https://doi.org/10.38018/TildaRe.2020-05.
- [107] S. Prentice, They are what you eat: can nutritional factors during gestation and early infancy modulate the neonatal immune response? Front. Immunol. 8 (2017) 1641, https://doi.org/10.3389/fimmu.2017.01641.
- [108] J.J. Cannell, R. Vieth, J.C. Umhau, M.F. Holick, W.B. Grant, S. Madronich, C.F. Garland, E. Giovannucci, Epidemic influenza and vitamin D, Epidemiol. Infect. 134 (2006) 1129–1140, https://doi.org/10.1017/s0950268806007173
- [109] K.R. Jat, Vitamin D deficiency and lower respiratory tract infections in children: a systematic review and meta-analysis of observational studies, Trop. Dr. 47 (2017) 77–84, https://doi.org/10.1177/0049475516644141.
- [110] M. Caplan, P. Calder, S. Prescott (Eds.), Scientific Review: The Role of Nutrients in Immune Function of Infants and Young Children Emerging Evidence for Longchain Polyunsaturated Fatty Acids, 2007.

- [111] D.E. Roth, A.B. Jones, C. Prosser, J.L. Robinson, S. Vohra, Vitamin D receptor polymorphisms and the risk of acute lower respiratory tract infection in early childhood, J. Infect. Dis. 197 (2008) 676–680, https://doi.org/10.1086/527488.
- [112] G.R. Zosky, L.J. Berry, J.G. Elliot, A.L. James, S. Gorman, P.H. Hart, Vitamin D deficiency causes deficits in lung function and alters lung structure, Am. J. Respir. Crit. Care Med. 183 (2011) 1336–1343, https://doi.org/10.1164/rccm.201010-15960C.
- [113] S. Beyhan-Sagmen, O. Baykan, B. Balcan, B. Ceyhan, Association between severe vitamin D deficiency, lung function and asthma control, Arch. Bronconeumol. 53 (2017) 186–191.
- [114] A. Bener, M.S. Ehlayel, H.Z. Bener, Q. Hamid, The impact of Vitamin D deficiency on asthma, allergic rhinitis and wheezing in children: an emerging public health problem, J. Fam. Community Med. 21 (2014) 154–161, https://doi.org/10.4103/ 2230-8229.142967.
- [115] M.E. Hejazi, F. Modarresi-Ghazani, T. Entezari-Maleki, A review of Vitamin D effects on common respiratory diseases: asthma, chronic obstructive pulmonary disease, and tuberculosis, J. Res. Pharm. Pract. 5 (2016) 7–15, https://doi.org/10. 4103/2279-042X.176542.
- [116] A.R. Martineau, D.A. Jolliffe, R.L. Hooper, L. Greenberg, J.F. Aloia, P. Bergman, G. Dubnov-Raz, S. Esposito, D. Ganmaa, A.A. Ginde, E.C. Goodall, C.C. Grant, C.J. Griffiths, W. Janssens, I. Laaksi, S. Manaseki-Holland, D. Mauger, D.R. Murdoch, R. Neale, J.R. Rees, S. Simpson Jr., I. Stelmach, G.T. Kumar, M. Urashima, C.A. Camargo Jr., Vitamin D supplementation to prevent acute respiratory tract infections: systematic review and meta-analysis of individual participant data, BMJ 356 (2017), https://doi.org/10.1136/bmj.i6583 i6583.
- [117] H. Hemilä, Vitamin C and infections, Nutrients 9 (2017) 339, https://doi.org/10. 3390/nu9040339.
- [118] H. Hemila, P. Louhiala, Vitamin C for preventing and treating pneumonia, Cochrane Database Syst. Rev. (2013) CD005532, https://doi.org/10.1002/ 14651858.CD005532.pub3.
- [119] Y. Cai, Y.F. Li, L.P. Tang, B. Tsoi, M. Chen, H. Chen, X.M. Chen, R.R. Tan, H. Kurihara, R.R. He, A new mechanism of vitamin C effects on A/FM/1/ 47(H1N1) virus-induced pneumonia in restraint-stressed mice, Biomed. Res. Int. 2015 (2015) 675149, https://doi.org/10.1155/2015/675149.
- [120] H. Kim, M. Jang, Y. Kim, J. Choi, J. Jeon, J. Kim, Y.I. Hwang, J.S. Kang, W.J. Lee, Red ginseng and vitamin C increase immune cell activity and decrease lung inflammation induced by influenza A virus/H1N1 infection, J. Pharm. Pharmacol. 68 (2016) 406–420, https://doi.org/10.1111/jphp.12529.
- [121] H. Hemilä, E. Chalker, Vitamin C can shorten the length of stay in the ICU: a metaanalysis, Nutrients 11 (2019), https://doi.org/10.3390/nu11040708.
- [122] A.A. Fowler III, C. Kim, L. Lepler, R. Malhotra, O. Debesa, R. Natarajan, B.J. Fisher, A. Syed, C. DeWilde, A. Priday, V. Kasirajan, Intravenous vitamin C as adjunctive therapy for enterovirus/rhinovirus induced acute respiratory distress syndrome, World J. Crit. Care Med. 6 (2017) 85–90, https://doi.org/10.5492/wjccm.v6. i1.85.
- [123] Z. Huang, Y. Liu, G. Qi, D. Brand, S.G. Zheng, Role of vitamin A in the immune system, J. Clin. Med. 7 (2018), https://doi.org/10.3390/jcm7090258.
- [124] M. Levy, C.A. Thaiss, E. Elinav, Metabolites: messengers between the microbiota and the immune system, Genes Dev. 30 (2016) 1589–1597, https://doi.org/10. 1101/gad.284091.116.
- [125] S. Sirisinha, The pleiotropic role of vitamin A in regulating mucosal immunity, Asian Pac. J. Allergy Immunol. 33 (2015) 71–89.
- [126] P.C. Calder, Feeding the immune system, Proc. Nutr. Soc. 72 (2013) 299–309, https://doi.org/10.1017/s0029665113001286.
- [127] A.C. Ross, Modern Nutrition in Health and Disease, Wolters Kluwer Health/ Lippincott Williams & Wilkins, Philadelphia, 2014.
- [128] J.L. McGill, S.M. Kelly, M. Guerra-Maupome, E. Winkley, J. Henningson, B. Narasimhan, R.E. Sacco, Vitamin A deficiency impairs the immune response to intranasal vaccination and RSV infection in neonatal calves, Sci. Rep. 9 (2019) 15157, https://doi.org/10.1038/s41598-019-51684-x.
- [129] Y. Xing, K. Sheng, X. Xiao, J. Li, H. Wei, L. Liu, W. Zhou, X. Tong, Vitamin A deficiency is associated with severe *Mycoplasma pneumoniae* pneumonia in children, Ann. Transl. Med. 8 (2020) 120, https://doi.org/10.21037/atm.2020.02.33.
- [130] B.E. Fan, K.G.E. Lim, C.L. Chong, S.S.W. Chan, K.H. Ong, P. Kuperan, COVID-19 and mycoplasma pneumoniae coinfection, Am. J. Hematol. (2020), https://doi. org/10.1002/ajh.25785.
- [131] S.R. D, Micronutrient deficiencies during the weaning period and the first years of life, in: J.M. Pettifor, S. Zlotkin (Eds.), Nestlé Nutrition Workshop Series Pediatric Program, Nestec Ltd., Basel, 2004, pp. 137–152, https://doi.org/10.1159/ 000080608.
- [132] M.I. Center, L.P. Institute (Ed.), Immunity In Brief, 2017.
- [133] M.G. Traber, J. Atkinson, Vitamin E, antioxidant and nothing more, Free Radic. Biol. Med. 43 (2007) 4–15, https://doi.org/10.1016/j.freeradbiomed.2007.03. 024.
- [134] M. Mileva, A.S. Galabov, Vitamin E and Influenza Virus Infection, (2018).
- [135] S.N. Han, D. Wu, W.K. Ha, A. Beharka, D.E. Smith, B.S. Bender, S.N. Meydani, Vitamin E supplementation increases T helper 1 cytokine production in old mice infected with influenza virus, Immunology 100 (2000) 487–493, https://doi.org/ 10.1046/j.1365-2567.2000.00070.x.
- [136] M.A. Beck, Selenium and vitamin E status: impact on viral pathogenicity, J. Nutr. 137 (2007) 1338–1340, https://doi.org/10.1093/jn/137.5.1338.
- [137] M. De la Fuente, A. Hernanz, N. Guayerbas, M. Victor, F. Arnalich, Vitamin E ingestion improves several immune functions in elderly men and women, Free Radic. Res. 42 (2008) 272–280, https://doi.org/10.1080/10715760801898838.
- [138] S.A. Read, S. Obeid, C. Ahlenstiel, G. Ahlenstiel, The role of zinc in antiviral immunity, Adv. Nutr. 10 (2019) 696–710, https://doi.org/10.1093/advances/

nmz013.

- [139] C.T. Chasapis, P.A. Ntoupa, C.A. Spiliopoulou, M.E. Stefanidou, Recent aspects of the effects of zinc on human health, Arch. Toxicol. 94 (2020) 1443–1460, https:// doi.org/10.1007/s00204-020-02702-9.
- [140] R.O. Suara, J.E. Crowe Jr., Effect of zinc salts on respiratory syncytial virus replication, Antimicrob. Agents Chemother. 48 (2004) 783–790, https://doi.org/10. 1128/aac.48.3.783-790.2004.
- [141] H. Ghaffari, A. Tavakoli, A. Moradi, A. Tabarraei, F. Bokharaei-Salim, M. Zahmatkeshan, M. Farahmand, D. Javanmard, S.J. Kiani, M. Esghaei, V. Pirhajati-Mahabadi, S.H. Monavari, A. Ataei-Pirkooh, Inhibition of H1N1 influenza virus infection by zinc oxide nanoparticles: another emerging application of nanomedicine, J. Biomed. Sci. 26 (2019) 70, https://doi.org/10.1186/s12929-019-0563-4.
- [142] C.T. Chasapis, P.A. Ntoupa, C.A. Spiliopoulou, M.E. Stefanidou, Recent aspects of the effects of zinc on human health, Arch. Toxicol. 94 (2020) 1443–1460, https:// doi.org/10.1007/s00204-020-02702-9.
- [143] W. Savino, M. Dardenne, Nutritional imbalances and infections affect the thymus: consequences on T-cell-mediated immune responses, Proc. Nutr. Soc. 69 (2010) 636–643, https://doi.org/10.1017/s0029665110002545.
- [144] B. Sandström, A. Cederblad, B.S. Lindblad, B. Lönnerdal, Acrodermatitis enteropathica, zinc metabolism, copper status, and immune function, Arch. Pediatr. Adolesc. Med. 148 (1994) 980–985, https://doi.org/10.1001/archpedi.1994. 02170090094017.
- [145] D. Hulisz, Efficacy of zinc against common cold viruses: an overview, J. Am. Pharm. Assoc. 44 (2004) (2003) 594–603, https://doi.org/10.1331/1544-3191. 44.5.594.hulisz.
- [146] H. Hemilä, Zinc lozenges may shorten the duration of colds: a systematic review, Open Respir. Med. J. 5 (2011) 51–58, https://doi.org/10.2174/ 1874306401105010051.
- [147] J.B. Barnett, M.C. Dao, D.H. Hamer, R. Kandel, G. Brandeis, D. Wu, G.E. Dallal, P.F. Jacques, R. Schreiber, E. Kong, S.N. Meydani, Effect of zinc supplementation on serum zinc concentration and T cell proliferation in nursing home elderly: a randomized, double-blind, placebo-controlled trial, Am. J. Clin. Nutr. 103 (2016) 942–951, https://doi.org/10.3945/ajcn.115.115188.
- [148] M.P. Rayman, The importance of selenium to human health, Lancet 356 (2000) 233-241, https://doi.org/10.1016/S0140-6736(00)02490-9.
- [149] H. Steinbrenner, S. Al-Quraishy, M.A. Dkhil, F. Wunderlich, H. Sies, Dietary selenium in adjuvant therapy of viral and bacterial infections, Adv. Nutr. 6 (2015) 73–82, https://doi.org/10.3945/an.114.007575.
- [150] H.K. Nelson, Q. Shi, P. Van Dael, E.J. Schiffrin, S. Blum, D. Barclay, O.A. Levander, M.A. Beck, Host nutritional selenium status as a driving force for influenza virus mutations, FASEB J. 15 (2001) 1846–1848, https://doi.org/10.1096/fj.01-0115fie.
- [151] O.M. Guillin, C. Vindry, T. Ohlmann, L. Chavatte, Selenium, selenoproteins and viral infection, Nutrients 11 (2019), https://doi.org/10.3390/nu11092101.
- [152] B. Shojadoost, R.R. Kulkarni, A. Yitbarek, A. Laursen, K. Taha-Abdelaziz, T. Negash Alkie, N. Barjesteh, W.M. Quinteiro-Filho, T.K. Smith, S. Sharif, Dietary selenium supplementation enhances antiviral immunity in chickens challenged with low pathogenic avian influenza virus subtype H9N2, Vet. Immunol. Immunopathol. 207 (2019) 62–68, https://doi.org/10.1016/j.vetimm.2018.12. 002.
- [153] B. Chaigne-Delalande, F.Y. Li, G.M. O'Connor, M.J. Lukacs, P. Jiang, L. Zheng, A. Shatzer, M. Biancalana, S. Pittaluga, H.F. Matthews, T.J. Jancel, J.J. Bleesing, R.A. Marsh, T.W. Kuijpers, K.E. Nichols, C.L. Lucas, S. Nagpal, H. Mehmet, H.C. Su, J.I. Cohen, G. Uzel, M.J. Lenardo, Mg2+ regulates cytotoxic functions of NK and CD8 T cells in chronic EBV infection through NKG2D, Science 341 (2013) 186–191, https://doi.org/10.1126/science.1240094.
- [154] S. Johnson, The multifaceted and widespread pathology of magnesium deficiency, Med. Hypotheses 56 (2001) 163–170, https://doi.org/10.1054/mehy.2000.1133.
- [155] F.H. Nielsen, Magnesium deficiency and increased inflammation: current perspectives, J. Inflamm. Res. 11 (2018) 25–34, https://doi.org/10.2147/JIR. S136742.
- [156] R.A. Landon, E.A. Young, Role of magnesium in regulation of lung function, J. Am. Diet. Assoc. 93 (1993) 674–677, https://doi.org/10.1016/0002-8223(93) 91675-g.
- [157] R. Janssen, Magnesium to counteract elastin degradation and vascular calcification in chronic obstructive pulmonary disease, Med. Hypotheses 107 (2017)

74-77, https://doi.org/10.1016/j.mehy.2017.08.014.

- [158] A. Mahmoud Abd, L. El Hafiz, H. Mohammed El Wakeel, A.E.R. Mourad Mohammed El Hady, High dose N-acetyl cysteine improves inflammatory response and outcome in patients with COPD exacerbations, Egypt. J. Chest Dis. Tuberc. 62 (2013) 51–57, https://doi.org/10.1016/j.ejcdt.2013.02.012.
- [159] C.A. Dick, D.M. Brown, K. Donaldson, V. Stone, The role of free radicals in the toxic and inflammatory effects of four different ultrafine particle types, Inhal. Toxicol. 15 (2003) 39–52, https://doi.org/10.1080/08958370304454.
- [160] M. Mata, I. Sarrion, M. Armengot, C. Carda, I. Martinez, J.A. Melero, J. Cortijo, Respiratory syncytial virus inhibits ciliagenesis in differentiated normal human bronchial epithelial cells: effectiveness of N-acetylcysteine, PLoS One 7 (2012) e48037, https://doi.org/10.1371/journal.pone.0048037.
- [161] W.Z. Ho, S.D. Douglas, Glutathione and N-acetylcysteine suppression of human immunodeficiency virus replication in human monocyte/macrophages in vitro, AIDS Res. Hum. Retrovir. 8 (1992) 1249–1253, https://doi.org/10.1089/aid. 1992.8.1249.
- [162] A. Garozzo, G. Tempera, D. Ungheri, R. Timpanaro, A. Castro, N-acetylcysteine synergizes with oseltamivir in protecting mice from lethal influenza infection, Int. J. Immunopathol. Pharmacol. 20 (2007) 349–354, https://doi.org/10.1177/ 039463200702000215.
- [163] Q. Zhang, Y. Ju, Y. Ma, T. Wang, N-acetylcysteine improves oxidative stress and inflammatory response in patients with community acquired pneumonia: A randomized controlled trial, Medicine (Baltimore) 97 (2018) e13087, https://doi. org/10.1097/MD.00000000013087.
- [164] S. De Flora, C. Grassi, L. Carati, Attenuation of influenza-like symptomatology and improvement of cell-mediated immunity with long-term N-acetylcysteine treatment, Eur. Respir. J. 10 (1997) 1535–1541, https://doi.org/10.1183/09031936. 97.10071535.
- [165] H. Cory, S. Passarelli, J. Szeto, M. Tamez, J. Mattei, The role of polyphenols in human health and food systems: a mini-review, Front. Nutr. 5 (2018) 87, https:// doi.org/10.3389/fnut.2018.00087.
- [166] S.C. Lin, C.T. Ho, W.H. Chuo, S. Li, T.T. Wang, C.C. Lin, Effective inhibition of MERS-CoV infection by resveratrol, BMC Infect. Dis. 17 (2017) 144, https://doi. org/10.1186/s12879-017-2253-8.
- [167] M. Catel-Ferreira, H. Tnani, C. Hellio, P. Cosette, L. Lebrun, Antiviral effects of polyphenols: Development of bio-based cleaning wipes and filters, J. Virol. Methods 212 (2015) 1–7, https://doi.org/10.1016/j.jviromet.2014.10.008.
- [168] V. Balachandar, I. Mahalaxmi, J. Kaavya, G. Vivekanandhan, S. Ajithkumar, N. Arul, G. Singaravelu, N. Senthil Kumar, S. Mohana Dev, COVID-19: emerging protective measures, Eur. Rev. Med. Pharmacol. Sci. 24 (2020) 3422–3425, https://doi.org/10.26355/eurrev_202003_20713.
- [169] B. Sonam, G. Sabeena, L.F. Arnica, S. Shaminder, Battle Against Coronavirus: Repurposing Old Friends (Food Borne Polyphenols) for New Enemy (COVID-19), (2020).
- [170] Y.Q. Li, Z.L. Li, W.J. Zhao, R.X. Wen, Q.W. Meng, Y. Zeng, Synthesis of stilbene derivatives with inhibition of SARS coronavirus replication, Eur. J. Med. Chem. 41 (2006) 1084–1089, https://doi.org/10.1016/j.ejmech.2006.03.024.
- [171] S. Jo, H. Kim, S. Kim, D.H. Shin, M.-S. Kim, Characteristics of flavonoids as potent MERS-CoV 3C-like protease inhibitors, Chem. Biol. Drug Des. 94 (2019) 2023–2030, https://doi.org/10.1111/cbdd.13604.
- [172] H.R. Park, H. Yoon, M.K. Kim, S.D. Lee, Y. Chong, Synthesis and antiviral evaluation of 7-O-arylmethylquercetin derivatives against SARS-associated coronavirus (SCV) and hepatitis C virus (HCV), Arch. Pharm. Res. 35 (2012) 77–85, https://doi.org/10.1007/s12272-012-0108-9.
- [173] A. Vazquez-Calvo, N. Jimenez de Oya, M.A. Martin-Acebes, E. Garcia-Moruno, J.C. Saiz, antiviral properties of the natural polyphenols delphinidin and epigallocatechin gallate against the flaviviruses West Nile virus, zika virus, and dengue virus, Front. Microbiol. 8 (2017) 1314, https://doi.org/10.3389/fmicb. 2017.01314.
- [174] M. Sokmen, M. Angelova, E. Krumova, S. Pashova, S. Ivancheva, A. Sokmen, J. Serkedjieva, In vitro antioxidant activity of polyphenol extracts with antiviral properties from *Geranium sanguineum* L, Life Sci. 76 (2005) 2981–2993, https:// doi.org/10.1016/j.lfs.2004.11.020.
- [175] M. Iyer, K. Jayaramayya, M.D. Subramaniam, S.B. Lee, A.A. Dayem, S.G. Cho, B. Vellingiri, COVID-19: an update on diagnostic and therapeutic approaches, BMB Rep. 53 (2020) 191–205.