

# Genes in S and T Subgenomes Are Responsible for Hybrid Lethality in Interspecific Hybrids between *Nicotiana tabacum* and *Nicotiana occidentalis*

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### **Abstract**

**Background:** Many species of *Nicotiana* section *Suaveolentes* produce inviable  $F_1$  hybrids after crossing with *Nicotiana* tabacum (genome constitution SSTT), a phenomenon that is often called hybrid lethality. Through crosses with monosomic lines of *N. tabacum* lacking a Q chromosome, we previously determined that hybrid lethality is caused by interaction between gene(s) on the Q chromosome belonging to the S subgenome of *N. tabacum* and gene(s) in *Suaveolentes* species. Here, we examined if hybrid seedlings from the cross *N. occidentalis* (section *Suaveolentes*)  $\times N$ . tabacum are inviable despite a lack of the Q chromosome.

**Methodology/Principal Findings:** Hybrid lethality in the cross of *N. occidentalis* × *N. tabacum* was characterized by shoots with fading color. This symptom differed from what has been previously observed in lethal crosses between many species in section *Suaveolentes* and *N. tabacum*. In crosses of monosomic *N. tabacum* plants lacking the Q chromosome with *N. occidentalis*, hybrid lethality was observed in hybrid seedlings either lacking or possessing the Q chromosome. *N. occidentalis* was then crossed with two progenitors of *N. tabacum*, *N. sylvestris* (SS) and *N. tomentosiformis* (TT), to reveal which subgenome of *N. tabacum* contains gene(s) responsible for hybrid lethality. Hybrid seedlings from the crosses *N. occidentalis* × *N. tomentosiformis* and *N. occidentalis* × *N. sylvestris* were inviable.

**Conclusions/Significance:** Although the specific symptoms of hybrid lethality in the cross *N. occidentalis* × *N. tabacum* were similar to those appearing in hybrids from the cross *N. occidentalis* × *N. tomentosiformis*, genes in both the S and T subgenomes of *N. tabacum* appear responsible for hybrid lethality in crosses with *N. occidentalis*.

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# Introduction

Reproductive isolation involves various types of prezygotic and postzygotic barriers in plants [1]. Although these barriers contribute to the formation of species, they are obstacles for plant breeders, especially in breeding programs involving wide hybridization. Hybrid plants from normal parents often show weak growth or death before maturity. Several terms are used to describe this phenomenon, including hybrid lethality, hybrid weakness, hybrid necrosis, and hybrid inviability. Hybrid lethality is a type of postzygotic barrier and is observed in certain cross combinations in numerous plant species, including *Nicotiana* sp. [2], *Oryza* sp. [3,4], *Phaseolus vulgaris* [5], *Lactuca* sp. [6], and *Arabidopsis thaliana* [7].

The genus *Nicotiana* comprises 76 species that are predominantly distributed in the Americas and Australia [8]. This complex genus has attracted the attention of many researchers investigating evolutionary processes. The most well-studied and characterized species in this genus is cultivated tobacco, *Nicotiana tabacum* 

(2n = 48, SSTT), which belongs to section *Nicotiana*. *N. tabacum* is a natural allotetraploid (amphidiploid) that originated by interspecific hybridization of *N. sylvestris* (2n = 24, SS) with *N. tomentosiformis* (2n = 24, TT) and subsequent chromosome doubling [9–16]. Each chromosome of *N. tabacum* is lettered alphabetically (A–Z, excluding X and Y); chromosomes A–L and M–Z belong to the T and S subgenomes, respectively. A complete set of 24 monosomic lines of *N. tabacum* (Haplo-A–Z), which lack a certain chromosome, has been established in the genetic background of 'Red Russian' and is useful for locating genes on specific chromosomes [17–19]. Monosomic analyses have identified one *N. tabacum* chromosome containing a gene or genes that trigger hybrid lethality [20–22].

Hybrid lethality in the genus *Nicotiana* is currently classified into four types based on surface symptoms, as follows: Type I, browning of shoot apex and root tips; Type II, browning of hypocotyl and roots; Type III, yellowing of true leaves; and Type IV, formation of multiple shoots [2]. Although several methods to overcome hybrid lethality have been reported in the genus

Nicotiana, their effectiveness is dependent on the type of hybrid lethality encountered. For example, hybrid lethality of Types I, II, and III is temperature sensitive; i.e., hybrid lethality is observed at 28°C, but not at elevated temperatures of approximately 36°C. In contrast, Type IV lethality is not suppressed at elevated temperatures [2].

Our previous studies indicate that many species in Nicotiana section Suaveolentes produce inviable hybrids after crosses with N. tabacum [20-22]. Section Suaveolentes includes 26 species, most of which are endemic to Australia, although three species are found in other locations. For example, N. debnevi is distributed in Eastern Australia, New Caledonia, and Lord Howe Island, while N. fragrans is distributed in New Caledonia, and the Loyalty, Tonga, and Marquesas Islands [23,24]. One species, N. africana, is distributed in Namibia (South-West Africa), and represents the only Nicotiana species discovered in Africa to date [25]. Species in section Suaveolentes are geographically isolated from the majority of species in other sections, which are distributed in the Americas.

All species in section Suaveolentes are allotetraploids and comprise an almost complete an euploid series of n = 16-24, with only n = 17yet to be identified. Section Suaveolentes is considered to have originated from a single polyploid event approximately 10 mya, followed by speciation to produce the species known today [26]. This hypothesis is supported by recent studies indicating that section Suaveolentes is a monophyletic group based on analyses of internal transcribed spacer (ITS) regions [14], plastid genes [27], and nuclear-encoded chloroplast-expressed glutamine synthetase (ncpGS) [16]. Furthermore, progenitors of this section have been identified based on sequence analyses of ncpGS; the maternal progenitor is N. sylvestris and the paternal progenitor is section Trigonophyllae [16]. One group of Suaveolentes species is genetically distant from N. tabacum, even though they share the same maternal progenitor, N. sylvestris [14,16,27]. Although Suaveolentes species are valuable as sources of disease resistance [28–30] and cytoplasmic male sterility [31,32], the occurrence of hybrid lethality represents a barrier for introducing desirable characteristics into N. tabacum by interspecific crosses.

After crosses with N. tabacum, nine species in section Suaveolentes, namely N. africana, N. debneyi, N. excelsior, N. goodspeedii, N. gossei, N. maritima, N. megalosiphon, N. suaveolens, and N. velutina, produce inviable hybrids, whereas only N. benthamiana and N. fragrans generate 100% viable hybrids [20-22,33]. Hybrid lethality observed in all crosses at 28°C is Type II and is suppressed at temperatures of 34–36°C. We also previously conducted genetic analyses of hybrid lethality, which revealed that the Q chromosome in the S subgenome of N. tabacum encodes one or more genes leading to hybrid lethality in crosses using the nine species of section Suaveolentes [20-22]. Based on the existence of a single origin of section Suaveolentes, we proposed that many species in section Suaveolentes share the same gene(s) triggering hybrid lethality by interaction with gene(s) on the Q chromosome [22].

In the present study, we examined hybrid lethality in crosses between  $\mathcal{N}$ . occidentalis, which belongs to section Suaveolentes, and  $\mathcal{N}$ . tabacum. N. occidentalis is closely related to other Australian species in section Suaveolentes and is distant from N. tabacum based on recently constructed phylogenetic trees [14,16,27]. Although Ternovskii et al. [34] reported that hybrid seedlings from the cross  $\mathcal{N}$ . occidentalis  $Q \times \mathcal{N}$ . tabacum  $\mathcal{O}$  die without forming roots at the cotyledonary stage, the nature of the lethality was incompletely described. Here, based on phenotypic and genetic analyses, we demonstrate that  $\mathcal{N}$ . occidentalis exhibits a distinct type of hybrid lethality after crosses with  $\mathcal{N}$ . tabacum than that observed with the nine Suaveolentes species examined in our previous studies.

# Results

# Temperature-sensitive lethality in hybrid seedlings of N. occidentalis Q×N. tabacum ♂

Reciprocal crosses between N. occidentalis and N. tabacum (cultivars 'Red Russian' and 'Samsun NN') were carried out using conventional cross-pollination (Table 1). When  $\mathcal{N}$  occidentalis was used as the paternal parent, flowers of N. tabacum dropped approximately 7 days after pollination. The ovaries and ovules of these flowers did not enlarge, suggesting that fertilization did not occur. In contrast, when N occidentalis was used as the maternal parent, seeds that germinated well at 28°C under in-vitro conditions were obtained.

Hybrid seedlings from the cross N. occidentalis  $Q \times N$ . tabacum Owere grown in vitro at 24, 28, and 34°C (Table 2). At 24°C, hybrid seedlings grew normally and developed cotyledons and roots by approximately 3 days after germination (DAG), although root growth stopped at this stage (Figure 1A). The color of cotyledons and the hypocotyl began to fade approximately 8 DAG (Figure 1B, C), with hybrid seedlings dying at about 30 DAG (Figure 1D). A characteristic symptom of hybrid lethality was the fading of shoot color. As this symptom was distinct from those of the four known types of Nicotiana hybrid lethality [2], the type of lethality observed here was designated as Type V.

Hybrid seedlings also displayed Type V lethality at 28°C, although the lethal symptoms appeared later (approximately 30 DAG) than for seedlings grown at 24°C, with the seedlings surviving for a few months (Table 2). In contrast, hybrid seedlings grown at 34°C did not show lethal symptoms for at least 60 DAG (Table 2). When these hybrid seedlings were transferred to 28°C, they ceased growing and died 40-70 days after the transfer. The three parental species, N. occidentalis, 'Red Russian', and 'Samsun NN', grew normally without the appearance of lethal symptoms for at least 60 DAG at 24, 28, and 34°C (Table 2, Figure 1E-J).

# Hybrid seedlings die even when they lack the Q chromosome

N. tabacum monosomic plants lacking the O chromosome, namely  $F_1$  progeny derived from the cross N. tabacum Haplo-Q Q×'Samsun NN' or [35], were crossed with N. occidentalis to determine whether the Q chromosome is responsible for hybrid lethality. Monosomic plants were used as maternal parents, as the transmission of the monosomic condition through pollen seldom occurs in  $\mathcal{N}$ . tabacum [36]. When  $\mathcal{N}$ . tabacum was pollinated with  $\mathcal{N}$ . occidentalis, fertilization did not occur, as described above. Therefore, test-tube pollination and ovule culture were performed to overcome this crossing barrier.

Twenty-nine placentas of monosomic plants were pollinated in vitro, resulting in 445 enlarged ovules that were cultured at 28°C. The 30 hybrid seedlings obtained from the cultured ovules were further grown at 34°C and then assessed for the presence or absence of the Q chromosome using three Q-chromosome-specific sequence-tagged-site (STS) markers: QCS2, QCS3, and QCS4 [20,35]. Five of the hybrid seedlings exhibited abnormal morphology and lacked cotyledons and leaves. Because genomic DNA could not be isolated from these five seedlings, they were excluded from further analyses.

Using the Q-chromosome-specific STS markers, the remaining 25 hybrid seedlings were divided into two types: 11 hybrid seedlings possessing the Q chromosome and 14 hybrid seedlings lacking the Q chromosome (Table 3). Both types of hybrid seedlings were cultured for 30 DAG and then transferred from 34 to 28°C. After 40–70 days, both types of seedlings showed lethal symptoms and died (Figure 1K, L). Although a characteristic

Table 1. Conventional crossings between N. occidentalis and N. tabacum.

Cross combination	No. of flowers pollinated	No. of capsules obtained	No. of seeds sown	No. of hybrids obtained
N. occidentalis Q×'Red Russian' マ	20	19	340	305
'Red Russian' ♀×N. occidentalis ♂	21	0	-	-
N. occidentalis ♀×'Samsun NN' ♂	20	12	259	215
'Samsun NN' ♀×N. occidentalis ♂	20	0	_	-
N. occidentalis	-	-	140	136
'Red Russian'	-	-	140	137
'Samsun NN'	-	-	140	129

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symptom of hybrid lethality is the fading of shoot color, as reported above, no difference in this symptom was observed between the two types of hybrid seedlings. These results indicated that lack of the Q chromosome does not suppress hybrid lethality in  $\mathcal{N}$ . tabacum  $Q \times \mathcal{N}$ . occidentalis  $\mathcal{O}$  progeny.

# Causal genes for hybrid lethality exist in both the S and T subgenomes of *N. tabacum*

Two progenitors of  $\mathcal{N}$ . tabacum,  $\mathcal{N}$ . sylvestris and  $\mathcal{N}$ . tomentosiformis, were crossed with  $\mathcal{N}$ . occidentalis to reveal which subgenome of  $\mathcal{N}$ . tabacum encodes gene(s) responsible for hybrid lethality. Conventional cross-pollination was successful when  $\mathcal{N}$ . occidentalis was used as the maternal parent (Table 4). Seeds from two crosses, as well as those of  $\mathcal{N}$ . sylvestris and  $\mathcal{N}$ . tomentosiformis, germinated well at 28°C under in-vitro conditions. Hybrid seedlings and the parental species were grown at 24 or 28°C in vitro. At these two temperatures, all hybrid seedlings from both crosses died, whereas the parental species grew normally (Table 5, Figure 2). Lethal symptoms in hybrid seedlings of  $\mathcal{N}$ . occidentalis  $\mathcal{Q} \times \mathcal{N}$ . sylvestris  $\mathcal{O}$  grown at 24°C appeared approximately 18 DAG as browning of the hypocotyl

(Figure 2B). A few days later, chlorosis of leaves was observed. We identified this type of lethality as Type II. For hybrid seedlings of  $\mathcal{N}$ . occidentalis  $\mathcal{Q} \times \mathcal{N}$ . tomentosiformis  $\mathcal{O}$ , lethal symptoms at 24°C appeared as fading of shoot color at about 5 DAG and thus, the observed lethality was identified as Type V (Figure 2E). Within each cross, the same characteristic symptoms appeared at 24 and 28°C (Table 5).

#### Discussion

Possible involvement of a dual lethal system causing hybrid lethality in crosses between *N. occidentalis* and *N. tabacum* 

Hybrid seedlings of  $\mathcal{N}$ . occidentalis  $Q \times \mathcal{N}$ . tabacum  $\mathcal{O}$  and  $\mathcal{N}$ . occidentalis  $Q \times \mathcal{N}$ . tomentosiformis  $\mathcal{O}$  exhibited hybrid lethality that was characterized by the fading of shoot color. As this characteristic was distinct from those of the four known types of lethality in  $\mathcal{N}$ icotiana [2], we designated this type of hybrid lethality as Type V. Notably, Type V lethality is also temperature sensitive, similar to Types I, II, and III.

**Table 2.** Viability of hybrid seedlings generated from crosses between N. occidentalis and N. tabacum at different temperatures.

	Temperature (°C)	No. of hybrids			
Cross combination		Total	Viable	Inviable	Lethality type <sup>a</sup>
N. occidentalis Ç×'Red Russian' ♂	24	80	0	80	V
	28	205	0	205	V
	34	20	20	0	-
N. occidentalis ♀×'Samsun NN' ♂	24	74	0	74	V
	28	121	0	121	V
	34	20	20	0	-
N. occidentalis	24	57	57	0	-
	28	59	59	0	-
	34	20	20	0	_
'Red Russian'	24	57	57	0	-
	28	60	60	0	_
	34	20	20	0	-
'Samsun NN'	24	56	56	0	_
	28	53	53	0	-
	34	20	20	0	-

<sup>a</sup>Type V, fading of shoot color. doi:10.1371/journal.pone.0036204.t002



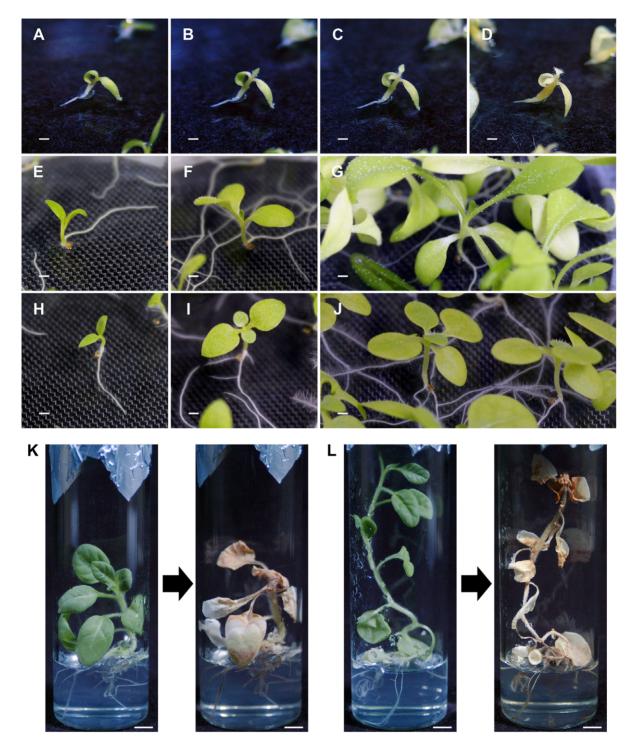


Figure 1. Lethal symptoms observed in hybrid seedlings from crosses between *N. occidentalis* and *N. tabacum*. (A–D) Hybrid seedlings from the cross *N. occidentalis*  $Q \times N$ . tabacum 'Red Russian' O at  $24^{\circ}$ C. Although hybrid seedlings at 3 days after germination (DAG) exhibited apparently normal growth, root growth nearly stopped (A). The color of cotyledons and hypocotyl faded beginning from 8 DAG (B), continued to advance at 10 DAG (C), with the seedlings turning completely brown and dying by 30 DAG (D). (E–G) *N. occidentalis* grown at  $24^{\circ}$ C. *N. occidentalis* grown at  $24^{\circ}$ C. N. occidentalis grown at 3 (E), 10 (F), or 30 DAG (G). (H–J) 'Red Russian' grown at  $24^{\circ}$ C. 'Red Russian' also grew normally and did not display lethal symptoms at 3 (H), 10 (I), or 30 DAG (J). (K, L) Hybrid seedlings from the cross (N. tabacum Haplo- $Q \times N$ . tabacum 'Samsun NN')  $Q \times N$ . occidentalis O. Hybrid seedlings possessing (K) and lacking (L) the Q chromosome died after being transferred from  $34^{\circ}$ C (left panels) to  $24^{\circ}$ C (right panels). Scale bars = 1 mm (A–J) and 5 mm (K, L). doi:10.1371/journal.pone.0036204.g001

The type of lethality observed in the cross  $\mathcal{N}$ . occidentalis  $Q \times \mathcal{N}$ . tabacum  $\mathcal{O}$  (Type V) was different from that (Type II) typically observed in crosses between species of section Suaveolentes and  $\mathcal{N}$ .

tabacum [2,22]. In addition to lethality type, the cause of hybrid lethality in  $\mathcal{N}$  occidentalis  $Q \times \mathcal{N}$ . tabacum  $\mathcal{O}$  was also unique. As mentioned, we previously found that one or more genes on the Q

**Table 3.** Relationship between the Q chromosome and hybrid lethality in crosses between N. tabacum and N. occidentalis.

		No. of hybrids		
Cross combination	STS markers <sup>a</sup>	Total	Viable	Inviable
(Haplo-Q×'Samsun NN') ♀×N. occidentalis ♂	+	11	0	11
	-	14	0	14

a'+' indicates that Q-chromosome-specific STS markers were detected and '-' indicates that they were not. doi:10.1371/journal.pone.0036204.t003

chromosome in the S subgenome of N. tabacum were responsible for hybrid lethality in crosses of nine species in section Suaveolentes [20–22]. However, here, hybrid seedlings of N. tabacum  $Q \times N$ . occidentalis of were inviable, even when they lacked the Q chromosome. Surprisingly, N. occidentalis yielded inviable hybrids after crossing with both progenitors of N. tabacum. Characteristic symptoms of hybrid lethality in the cross  $\mathcal{N}$ . occidentalis  $Q \times \mathcal{N}$ . tabacum  $\circ$  were similar to those appearing in the cross  $\mathcal{N}$ . occidentalis QXN. tomentosiformis O, strongly suggesting the involvement of gene(s) in the T subgenome. Nevertheless, we could not rule out the possibility that gene(s) in the S subgenome are also responsible for hybrid lethality in the cross  $\mathcal{N}$ . occidentalis  $Q \times \mathcal{N}$ . tabacum  $\mathcal{O}$ . As lethal symptoms in the cross N. occidentalis  $Q \times N$ . tomentosiformis Oappeared faster than in the cross  $\mathcal{N}$ . occidentalis  $Q \times \mathcal{N}$ . sylvestris  $\mathcal{O}$ , the phenotype of Type V lethality might be preferentially expressed in the cross  $\mathcal{N}$ . occidentalis  $Q \times \mathcal{N}$ . tabacum  $\mathcal{O}$ . If this supposition is true, the hybrid lethality observed in the cross between N. occidentalis and  $\mathcal{N}$ . tabacum is the first reported for a dual lethal system in the same cross-combination. Furthermore, the causal chromosome in the S subgenome of N. tabacum may be the Q chromosome, because hybrid lethality in the cross  $\mathcal{N}$ . occidentalis  $Q \times \mathcal{N}$ . sylvestris  $\mathcal{O}$  was Type II, which is often observed in crosses between species of section Suaveolentes and N. tabacum [2,22]. Although further evidence is needed, we conclude that genes from both the S and T subgenomes of N. tabacum are responsible for hybrid lethality in the cross between N. occidentalis and N. tabacum.

The viability and reproductive fitness of hybrids are often compromised when they are derived from parents that differ in ploidy levels [37]. Such dosage-sensitive incompatibility in interploidy crosses arises in the absence of any allelic diversity and is under genetic control [37,38]. Crosses between N. occidentalis and the two progenitors of N. tabacum were both interspecific and interploidal, because N. occidentalis is an allopolyploid, whereas N. sylvestris and N. tomentosiformis are diploids. Thus, it is difficult to determine whether hybrid lethality arose due to ploidy differences between the parental species. However, as we observed the identical hybrid lethality phenotypes in interploidy and same

ploidy crosses, ploidy differences are not likely to have contributed to the observed hybrid lethality in crosses between N. occidentalis and the two progenitors of N. tabacum.

# Unilateral incompatibility in crosses between *N. tabacum* and *Suaveolentes* species

Unilateral incompatibility or unilateral incongruity (UI) is often observed in interspecific crosses in *Nicotiana*. In crosses between  $\mathcal{N}$ . tabacum and Australian species of section Suaveolentes, including N. occidentalis, pollen of Suaveolentes species is often compatible with the N. tabacum pistil, but the reciprocal cross is incompatible. UI in Nicotiana arises from: (1) self-incompatibility [39], (2) differences in pistil length and pollen tube growth rate [40,41], and (3) morphological abnormalities of pollen tubes, such as winding, swelling, and irregular callose deposition [42,43]. Self-incompatibility is unlikely to be a factor in UI for crosses between N. tabacum and Australian species of section Suaveolentes, because both N. tabacum and Suaveolentes species are self-compatible. In crosses using N. tabacum as the female parent, pollen from six short-pistil species, N. benthamiana, N. debneyi, N. goodspeedii, N. maritima, N. occidentalis, and N. velutina, did not produce seeds, while pollen from three long-pistil species, N. excelsior, N. gossei, and N. megalosiphon, occasionally produced seeds (pollen from N. tabacum is compatible with pistils of all of these short- and long-pistil species) [21,22,33]. Therefore, pistil length and pollen tube growth rate might be involved in UI, although this is unlikely to be the only factor, as we observed that many flowers of long-pistil species dropped after pollination with N. tabacum pollen. Although morphological abnormalities of pollen tubes are observed in other crosses within the genus Nicotiana, we have no data on pollen tubes for the crosses between N. tabacum and Suaveolentes species.

# Evolutionary process of hybrid lethality in section *Suaveolentes*

Of the 12 Suaveolentes species used in the present and previous studies [20–22,33], only three species (N. occidentalis, N. benthamiana,

Table 4. Conventional crossing between N. occidentalis and two progenitors of N. tabacum.

Cross combination	No. of flowers pollinated	No. of capsules obtained	No. of seeds sown	No. of hybrids obtained	
N. occidentalis ♀×N. sylvestris ♂	20	19	160	119	
N. sylvestris Q×N. occidentalis ♂	20	0	-	_	
N. occidentalis Q×N. tomentosiformis ♂	17	10	157	140	
N. tomentosiformis ♀×N. occidentalis ♂	20	0	-	-	
N. sylvestris	-	-	120	117	
N. tomentosiformis	_	_	120	114	

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**Table 5.** Viability of hybrid seedlings generated from crosses between *N. occidentalis* and two progenitors of *N. tabacum* at different temperatures.

		No. of hybrids			
Cross combination	Temperature (°C)	Total	Viable	Inviable	Lethality type <sup>a</sup>
N. occidentalis Q×N. sylvestris ♂	24	58	0	58	II
	28	61	0	61	II
N. occidentalis Q×N. tomentosiformis ♂	24	71	0	71	V
	28	69	0	69	V
N. sylvestris	24	59	59	0	-
	28	58	58	0	-
N. tomentosiformis	24	58	58	0	_
	28	56	56	0	-

<sup>a</sup>Type II, browning of hypocotyl and roots; Type V, fading of shoot color. doi:10.1371/journal.pone.0036204.t005

and  $\mathcal{N}$ . fragrans) gave results different from those of the remaining 9 species. N. benthamiana and N. fragrans were the only species that produced viable hybrids after crosses with  $\mathcal{N}$ . tabacum. This result is surprising, as section Suaveolentes presumably originated from a single polyploid event [26]. In a previous study, we concluded that the common gene(s) that trigger hybrid lethality by interaction with gene(s) on the Q chromosome should be widely distributed in species of section Suaveolentes [22]. Based on this conclusion, we suspect that the progenitor(s), namely  $\mathcal{N}$ . sylvestris and/or section Trigonophyllae, which were involved in the formation of section Suaveolentes, might have possessed the gene(s) triggering hybrid lethality, whereas gene loss might have occurred during diploidization of the genome on polyploid establishment or during the subsequent divergence of N. benthamiana and N. fragrans [22]. However, it is also possible that the progenitors do not possess causal gene(s) for hybrid lethality in crosses with  $\mathcal{N}$ . tabacum.

Information on phylogeny is critical to estimate the evolutionary order and timing of causal genetic changes underlying reproductive isolation [44]. In three phylogenetic trees respectively constructed by analyses of sequence data for ITS regions, plastid genes, and ncpGS, section Suaveolentes formed a monophyletic group with the African species, N. africana, as a sister to the remaining species of section Suaveolentes [14,16,27]. More recently, Marks et al. [45] reported the results of phylogenetic analyses of section Suaveolentes based on the combined data set of morphological characters, chromosome number, and nodes that received strong bootstrap support in trees constructed from individual analyses of sequences of ITS regions, plastid genes, and ncpGS. In their tree, which was constructed using N. africana as the functional outgroup, the South Pacific species N. fragrans was the basal lineage in the Australian and South Pacific clade of section Suaveolentes. In addition, the Australian species N. debneyi (New Caledonia, Lord Howe Island, and Eastern Australia) was a sister species to all other endemic Australian species, including N. occidentalis and N. benthamiana.

Taking into account the results of recent phylogenetic analyses [45] and the crosses with *N. tabacum* performed here, we propose a new model for the evolutionary process leading to hybrid lethality in section *Suaveolentes* (Figure 3). It has been reported that the allotetraploid ancestor of the section, of which the maternal and paternal progenitors are *N. sylvestris* and section *Trigonophyllae*, respectively, occurred in South America [16,27]. In our proposed model, we considered that the allotetraploid ancestor did not cause hybrid lethality after crosses with *N. tabacum*. The appearance of

Type II lethality in the lineages leading to N. africana and the Australian clade possibly occurred after the divergence of  $\mathcal{N}$ . africana, N. fragrans, and the remaining species in section Suaveolentes (Figure 3). However, we cannot deny the possibility that Type II lethality appeared in the allotetraploid descendent after the divergence of  $\mathcal{N}$ . fragrans, and the allotetraploid descendent subsequently dispersed to Africa and Australia. Type II lethality may be caused by interaction with the Hla1-1 or other alleles found in the HLA1 locus, which was originally identified in N. debneyi [33], and gene(s) on the Q chromosome of N. tabacum. In Australia, an explosive radiation of taxa occurred that was largely accompanied by dysploid reductions, likely due to chromosomal fusions [27]. During the process, additional genetic changes that reinforced postzygotic isolation with N. tabacum accumulated in the lineage leading to  $\mathcal{N}$ . occidentalis, leading to the appearance of Type V lethality (Figure 3). Conversely, loss of Type II lethality gave rise to the lineage leading to N. benthamiana.

Although the considerable amount of accumulated phylogenetic data for section *Suaveolentes* has allowed us to propose an evolutionary model of hybrid lethality, the present data are insufficient to definitively determine the origin and interspecies relationships of this section. The accumulation of additional phylogenetic data and a greater understanding of the mechanisms underlying hybrid lethality will allow further clarification of how hybrid lethality has evolved through speciation in section *Suaveolentes*.

#### **Materials and Methods**

### Plant materials

 $\mathcal{N}.\ tabacum\ (2n=48,\ SSTT)\ 'Red\ Russian'\ and 'Samsun\ NN',\ \mathcal{N}.\ sylvestris\ (2n=24,\ SS),\ and\ \mathcal{N}.\ tomentosiformis\ (2n=24,\ TT)\ were\ used\ as\ parents\ for\ reciprocal\ crosses\ with\ \mathcal{N}.\ occidentalis\ (2n=42).$   $\mathcal{N}.\ tabacum\ monosomic\ plants\ (2n=47)\ were\ also\ used\ for\ crossing\ with\ \mathcal{N}.\ occidentalis.$  The monosomic plants were identified from  $F_1$  plants from the cross of  $\mathcal{N}.\ tabacum\ Haplo-Q\ Q\times 'Samsun\ NN'\ 'O'\ using\ Q-chromosome-specific\ STS\ marker\ QCS1\ [35].\ All\ plants\ were\ cultivated\ in\ a\ greenhouse.$ 

## Interspecific crosses

Conventional crossing and sowing were carried out as follows: flowers of plants used as maternal parents were emasculated one day before anthesis and pollinated with the pollen of paternal parent plants.  $F_1$  seeds were soaked in a 0.5% gibberellic acid

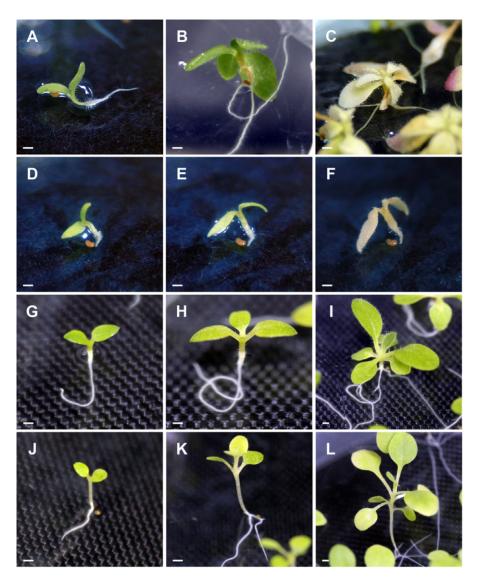


Figure 2. Lethal symptoms observed in hybrid seedlings from crosses between *N. occidentalis* and *N. tabacum* progenitors. (A–C) Hybrid seedlings from the cross *N. occidentalis*  $Q \times N$ . sylvestris Q at Q C. The seedlings showed apparently normal growth at 3 days after germination (DAG) (A). The hypocotyl turned brown at 18 DAG (B), and the seedlings turned completely brown and died at 61 DAG (C). (D–F) Hybrid seedlings from the cross *N. occidentalis*  $Q \times N$ . tomentosiformis Q at Q at Q at Q through the seedlings exhibited apparently normal growth, root growth nearly stopped at 2 DAG (D), and the color of cotyledons and hypocotyl faded at 5 DAG (E). The seedlings turned completely brown and died at 16 DAG (F). (G–I) *N. sylvestris* grown at Q C. *N. sylvestris* grew normally and did not show lethal symptoms at 3 (G), 10 (H), or 30 DAG (I). (J–L) *N. tomentosiformis* grown at Q C. *N. tomentosiformis* also grew normally and did not display any lethal symptoms at 3 (J), 10 (K), or 30 DAG (L). Scale bars = 1 mm. doi:10.1371/journal.pone.0036204.q002

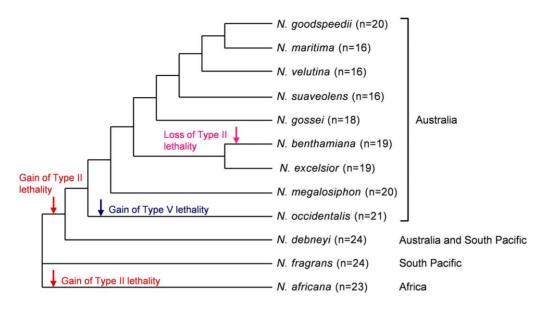
(GA<sub>3</sub>) solution for 30 min and sterilized with 5% sodium hypochlorite for 15 min. The sterilized seeds were sown in Petri dishes (60-mm diameter, 17-mm depth) containing 8 ml of 1/2 MS medium [46] supplemented with 1% sucrose and 0.2% Gelrite (pH 5.8), and then cultured at 28°C under continuous illumination (approximately 150  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup>).

Test-tube pollination in combination with ovule culture was performed as previously described [47] to obtain hybrid seedlings between  $\mathcal{N}$ . tabacum monosomic plants and  $\mathcal{N}$ . occidentalis. Anthers of  $\mathcal{N}$ . occidentalis plants used as paternal parents were aseptically excised from still-closed flowers and stimulated to dehisce in an incubator held at 28°C. Flowers of monosomic plants used as maternal parents were emasculated one day before anthesis. On the next day, flowers of monosomic plants were collected and their corolla, sepals, and styles were removed. The ovaries were surface-

sterilized with 70% ethanol for 30 s followed by a 5% sodium hypochlorite solution for 5 min. The ovary walls were peeled back to expose the placentas with intact ovules and the ovaries were then placed in Petri dishes containing 8 ml of medium supplemented with 3% sucrose and 0.8% agar (pH 5.8). Pollen of  $\mathcal{N}$  occidentalis was spread on the surface of the placentas, which were then maintained at 28°C under continuous illumination. Fertilized and enlarged ovules were excised from placentas 10 to 14 days after pollination and cultured in Petri dishes containing 8 ml of 1/2 MS medium supplemented with 3% sucrose and 0.8% agar (pH 5.8) at 28°C under continuous illumination.

### Cultivation of hybrid seedlings

Hybrid seedlings and parental species were cultured at 24 or 28°C under continuous illumination. Several seedlings were



(adapted from Marks et al., 2011)

**Figure 3. Schematic phylogenetic tree showing gain and loss of hybrid lethality in section** *Suaveolentes.* The schematic phylogenetic tree including 12 *Suaveolentes* species used in our studies on hybrid lethality was drawn based on tree by Marks et al. [45]. Gains of Type II lethality were established in the lineage leading to *N. africana* and the lineage leading to Australian clade. Subsequently, gain of Type V lethality was established in the lineage leading to *N. occidentalis.* In the lineage leading to *N. benthamiana*, loss of Type II lethality was established. doi:10.1371/journal.pone.0036204.q003

transferred to flat-bottomed test tubes (25-mm diameter, 100-mm length) that contained 10 ml of 1/2 MS medium supplemented with 1% sucrose and 0.2% Gelrite (pH 5.8) immediately after germination and were cultured at 24, 28, or 34°C under continuous illumination. Seedlings cultured at 34°C for 30–60 DAG were then further cultured at 28°C under continuous illumination.

### PCR analysis

Total DNA was extracted from the leaves of each plant using a cetyltrimethylammonium bromide-based method [48]. Four Q-chromosome-specific STS markers, QCS1, QCS2, QCS3, and QCS4 [20,35], were detected by PCR as follows. Reaction mixtures consisted of 1× ThermoPol reaction buffer (New England Biolabs, Tokyo, Japan), 0.2 mM each dNTP, 0.2 μM each primer, 20 ng template DNA, and 0.5 U *Taq* DNA polymerase (New England Biolabs) in a total volume of 20 μl. PCR amplification was performed using a PC-818A Program Temp Control System (Astec, Fukuoka, Japan) programmed for 3 min at 94°C for initial denaturation, followed by 35 cycles of

30 s at  $94^{\circ}\text{C}$ , 30 s at  $60^{\circ}\text{C}$ , and 30--90 s at  $72^{\circ}\text{C}$ , with a final 5-min extension at  $72^{\circ}\text{C}$ . PCR products were separated by electrophoresis in 1.5% agarose gels with TBE buffer and were then visualized by staining with ethicium bromide.

### **Ethics**

The ethics committee of Osaka Prefecture University specifically waived the need for consent.

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# **Author Contributions**

Conceived and designed the experiments: TT WM. Performed the experiments: TT. Analyzed the data: TT WM. Contributed reagents/materials/analysis tools: TT. Wrote the paper: TT.

# References

- Rieseberg LH, Blackman BK (2010) Speciation genes in plants. Ann Bot 106: 439–455.
- Yamada T, Marubashi W, Niwa M (1999) Detection of four lethality types in interspecific crosses among Nicotiana species through the use of three rescue methods for lethality. Breed Sci 49: 203–210.
- Kuboyama T, Saito T, Matsumoto T, Wu J, Kanamori H, et al. (2009) Fine mapping of HWC2, a complementary hybrid weakness gene, and haplotype analysis around the locus in rice. Rice 2: 93–103.
- Ichitani K, Taura S, Tezuka T, Okiyama Y, Kuboyama T (2011) Chromosomal location of HWA1 and HWA2, complementary hybrid weakness genes in rice. Rice 4: 29–38.
- Hannah MA, Krämer KM, Geffroy V, Kopka J, Blair MW, et al. (2007) Hybrid weakness controlled by the dosage-dependent lethal (DL) gene system in common bean (*Phaseolus vulgaris*) is caused by a shoot-derived inhibitory signal leading to salicylic acid-associated root death. New Phytol 176: 537–549.
- Jeuken MJW, Zhang NW, McHale LK, Pelgrom K, den Boer E, et al. (2009) Rin4 causes hybrid necrosis and race-specific resistance in an interspecific lettuce hybrid. Plant Cell 21: 3368–3378.
- Bomblies K, Lempe J, Epple P, Warthmann N, Lanz C, et al. (2007) Autoimmune response as a mechanism for a Dobzhansky-Muller-type incompatibility syndrome in plants. PLoS Biol 5: e236.
- Knapp S, Chase MW, Clarkson JJ (2004) Nomenclatural changes and a new sectional classification in Nicotiana (Solanaceae). Taxon 53: 73–82.
- Gray JC, Kung SD, Wildman SG, Sheen SJ (1974) Origin of Nicotiana tabacum L. detected by polypeptide composition of Fraction I protein. Nature 252: 226–227.
- Sheen SJ (1972) Isozymic evidence bearing on the origin of Nicotiana tabacum L. Evolution 26: 143–154.
- Kenton A, Parokonny AS, Gleba YY, Bennett MD (1993) Characterization of the Nicotiana tabacum L. genome by molecular cytogenetics. Mol Gen Genet 240: 159–169.

- Lim KY, Matyášek R, Lichtenstein CP, Leitch AR (2000) Molecular cytogenetic analyses and phylogenetic studies in the *Nicotiana* section Tomentosae. Chromosoma 109: 245–258.
- Murad L, Lim KY, Christopodulou V, Matyasek R, Lichtenstein CP, et al. (2002) The origin of tobacco's T genome is traced to a particular lineage within Nicotiana tomentosiformis (Solanaceae). Am J Bot 89: 921–928.
- Chase MW, Knapp S, Cox AV, Clarkson JJ, Butsko Y, et al. (2003) Molecular systematics, GISH and the origin of hybrid taxa in Nicotiana (Solanaceae). Ann Bot 92: 107–127.
- Murad L, Bielawski JP, Matyasek R, Kovarík A, Nichols RA, et al. (2004) The origin and evolution of geminivirus-related DNA sequences in *Nicotiana*. Heredity 92: 352–358.
- Clarkson JJ, Kelly LJ, Leitch AR, Knapp S, Chase MW (2010) Nuclear glutamine synthetase evolution in Nicotiana: phylogenetics and the origins of allotetraploid and homoploid (diploid) hybrids. Mol Phylogenet Evol 55: 90\_119
- Clausen RE, Cameron DR (1944) Inheritance in Nicotiana tabacum. XVIII. Monosomic analysis. Genetics 29: 447–477.
- Monosomic analysis. Genetics 29: 447–477.

  18. Cameron DR (1959) The monosomics of *Nicotiana tabacum*. Tob Sci 3: 164–166.
- Kubo T, Sato M, Tomita H, Kawashima N (1982) Identification of the chromosome carrying the gene for cis-abienol production by the use of monosomics in Nicotiana tabacum L. Tob Sci 26: 126–128.
- Tezuka T, Marubashi W (2006) Hybrid lethality in interspecific hybrids between Nicotiana tahacum and N. suaveoleus: evidence that the Q chromosome causes hybrid lethality based on Q-chromosome-specific DNA markers. Theor Appl Genet 112: 1179–1178.
- Tezuka T, Kuboyama T, Matsuda T, Marubashi W (2007) Possible involvement of genes on the Q chromosome of Nicotiana tabacum in expression of hybrid lethality and programmed cell death during interspecific hybridization to Nicotiana debneyi. Planta 226: 753–764.
- Tezuka T, Kuboyama T, Matsuda T, Marubashi W (2010) Seven of eight species in Nicotiana section Suaveolentes have common factors leading to hybrid lethality in crosses with Nicotiana tabacum. Ann Bot 106: 267–276.
- Goodspeed TH (1954) The genus Nicotiana. Chronica Botanica Company, Waltham, Massachusetts.
- Ladiges PY, Marks CE, Nelson G (2011) Biogeography of Nicotiana section Suaveolentes (Solanaceae) reveals geographical tracks in arid Australia. J Biogeogr 38: 2066–2077.
- Merxmüller H, Buttler KP (1975) Nicotiana in der afrikanischen Namib-ein pflanzengeographisches und phylogenetisches Rätsel. Mitt Bot Staatssamml München 12: 91–104.
- Leitch IJ, Hanson L, Lim KY, Kovarik A, Chase MW, et al. (2008) The ups and downs of genome size evolution in polyploid species of *Nicotiana* (Solanaceae). Ann Bot 101: 805–814.
- Clarkson JJ, Knapp S, Garcia VF, Olmstead RG, Leitch AR, et al. (2004) Phylogenetic relationships in Nicotiana (Solanaceae) inferred from multiple plastid DNA regions. Mol Phylogenet Evol 33: 75–90.
- Burk LG, Heggestad HE (1966) The genus Nicotiana: a source of resistance to diseases of cultivated tobacco. Econom Bot 20: 76–88.
- Li B-C, Bass WT, Cornelius PL (2006) Resistance to tobacco black shank in Nicotiana species. Crop Sci 46: 554

  –560.

- Bai D, Reeleder R, Brandle JE (1995) Identification of two RAPD markers tightly linked with the Nicotiana debneyi gene for resistance to black root rot of tobacco. Theor Appl Genet 91: 1184–1189.
- Nikova VM, Zagorska NA (1990) Overcoming hybrid incompatibility between Nicotiana africana Merxm. and N. tabacum and development of cytoplasmically male sterile tobacco forms. Plant Cell Tiss Org Cult 23: 71–75.
- Nikova VM, Zagorska NA, Pundeva RS (1991) Development of four tobacco cytoplasmic male sterile sources using in vitro techniques. Plant Cell Tiss Org Cult 27: 289–295.
- Iizuka T, Kuboyama T, Marubashi W, Oda M, Tezuka T (2012) Nicotiana debneyi
  has a single dominant gene causing hybrid lethality in crosses with N. tabacum.
  Euphytica, In press. 10.1007/s10681-011-0570-3 [doi].
- Ternovskii MF, Butenko RG, Moiseeva ME (1972) The use of tissue culture to overcome the barrier of incompatibility between species and sterility of interspecies hybrids. Sov Genet 8: 27–33. Translated from Genetika 8: 38–45.
- Tezuka T, Onosato K, Hijishita S, Marubashi W (2004) Development of Qchromosome-specific DNA markers in tobacco and their use for identification of a tobacco monosomic line. Plant Cell Physiol 45: 1863–1869.
- Olmo HP (1935) Genetical studies of monosomic types of Nicotiana tabacum. Genetics 20: 286–300.
- Martienssen RA (2010) Heterochromatin, small RNA and post-fertilization dysgenesis in allopolyploid and interploid hybrids of *Arabidopsis*. New Phytol 186: 46–53.
- Dilkes BP, Spielman M, Weizbauer R, Watson B, Burkart-Waco D, et al. (2008)
   The maternally expressed WRKY transcription factor TTG2 controls lethality in interploidy crosses of Arabidopsis. PLoS Biol 6: 2707–2720.
- Hancock CN, Kondo K, Beecher B, McClure B (2003) The S-locus and unilateral incompatibility. Philos Trans R Soc B-Biol Sci 358: 1133–1140.
- Lee CB, Page LE, McClure BA, Holtsford TP (2008) Post-pollination hybridization barriers in Nicotiana section Alatae. Sex Plant Reprod 21: 183–195.
- Figueroa-Castro DM, Holtsford TP (2009) Post-pollination mechanisms in Nicotiana longiflora and N. plumbaginifolia: pollen tube growth rate, offspring paternity and hybridization. Sex Plant Reprod 22: 187–196.
- Kuboyama T, Chung CS, Takeda G (1994) The diversity of interspecific pollenpistil incongruity in Nicotiana. Sex Plant Reprod 7: 250–258.
- Kuboyama T, Takeda G (2000) Genomic factors responsible for abnormal morphology of pollen tubes in the interspecific cross Nicotiana tabacum×N. rustica. Sex Plant Reprod 12: 333–337.
- Moyle LC, Payseur BA (2009) Reproductive isolation grows on trees. Trends Ecol Evol 24: 591–598.
- Marks CE, Newbigin E, Ladiges PY (2011) Comparative morphology and phylogeny of Nicotiana section Suaveolentes (Solanaceae) in Australia and the South Pacific. Aust Syst Bot 24: 61–86.
- Murashige T, Skoog F (1962) A revised medium for rapid growth and bio assays with tobacco tissue cultures. Physiol Plant 15: 473

  –497.
- Tezuka T, Marubashi W (2004) Apoptotic cell death observed during the expression of hybrid lethality in interspecific hybrids between Nicotiana tabacum and N. suaveolens. Breed Sci 54: 59–66.
- Murray MG, Thompson WF (1980) Rapid isolation of high molecular weight plant DNA. Nucleic Acids Res 8: 4321–4325.