



Research



Cite this article: Lin P-A, Chan W-P, Cai L, Hsiao Y, Dankowicz E, Gilbert KJ, Pierce NE, Felton G. 2025 The Salient Aroma Hypothesis: host plant specialization is linked with plant volatile availability in Lepidoptera. *Proc. R. Soc. B* **292**: 20242426.

<https://doi.org/10.1098/rspb.2024.2426>

Received: 10 October 2024

Accepted: 13 February 2025

Subject Category:

Evolution

Subject Areas:

evolution, ecology, plant science

Keywords:

volatile organic compounds, specialization, host range, diet breadth, diel activity, information availability

Author for correspondence:

Po-An Lin

e-mail: poanlin@ntu.edu.tw

[†]These authors contributed equally to the study.

Electronic supplementary material is available online at <https://doi.org/10.6084/m9.figshare.c.7684197>.

The Salient Aroma Hypothesis: host plant specialization is linked with plant volatile availability in Lepidoptera

Po-An Lin^{1,†}, Wei-Ping Chan^{3,†}, Liming Cai⁵, Yun Hsiao², Even Dankowicz³, Kadeem J. Gilbert⁶, Naomi E. Pierce⁴ and Gary Felton⁷

¹Department of Entomology, and ²Institute of Ecology and Evolutionary Biology, National Taiwan University, Taipei, Taiwan

³Department of Organismic and Evolutionary Biology, and ⁴Museum of Comparative Zoology, Harvard University, Cambridge, MA, USA

⁵Department of Integrative Biology, The University of Texas at Austin, Austin, TX, USA

⁶Department of Plant Biology, Michigan State University, W K Kellogg Biological Station, Hickory Corners, MI, USA

⁷Department of Entomology, The Pennsylvania State University, University Park, PA, USA

W-PC, 0000-0002-7132-9191; KJG, 0000-0003-0105-8020; NEP, 0000-0003-3366-1625

Host plant use in Lepidoptera has been a primary focus in studies of ecological specialization, and multiple factors are likely to be involved in shaping the evolution of diet breadth. Here, we first describe the Salient Aroma Hypothesis, suggesting that the availability of chemical information, particularly host-associated aromas, plays a critical role in shaping dietary specialization. According to the Salient Aroma Hypothesis, herbivores active during periods when chemical information is abundant, particularly during the daytime hours when plant aromas are hypothesized to be more prevalent, are more likely to evolve specialized diets. First, with meta-analysis, we show that plants release more diverse and abundant volatile compounds during daylight hours, increasing the availability of chemical information. We found that diurnal Lepidoptera tend to have specialized diets, while nocturnal species are more generalized, consistent with the prediction of the Salient Aroma Hypothesis. We further observed that morphological differences in the antennae of female Lepidoptera are correlated with variation in diet breadth and diel activity patterns, indirectly supporting the Salient Aroma Hypothesis. While multiple factors influence host plant specialization, the Salient Aroma Hypothesis offers a useful framework linking chemical information availability (e.g. plant volatiles) and ecological specialization.

1. Introduction

Ecological specialization is a widespread phenomenon in biological systems [1,2]. Herbivorous insects are notable for specializing on particular host plants, with less than 10% of herbivores feeding on more than three host plant families [3]. Most herbivores specialize on specific groups of plants [4], although exceptional cases of diet generalists also occur [5]. The relationship between lepidopteran herbivores and their host plants is one of the most thoroughly studied examples of ecological specialization, and research on this topic has spanned multiple contexts, including but not limited to coevolutionary arms races [6,7], evolutionary dynamics [8,9], genetic architecture [10], and geographic range and voltinism [11]. Despite considerable advances in our understanding of factors driving host specialization, explanations for the broad pattern of host specificity observed in phytophagous insects remain unclear [12].

Host range of most Lepidoptera is governed by adult females laying eggs on appropriate host plants, as well as the ability of larvae to overcome plant defences [13]. In seeking oviposition sites, females employ a combination of hierarchical and concurrent cues. Visual cues may initially guide them towards vegetation [14–16], while gustatory cues refine the process through the detection of specific plant chemicals [17]. Notably, chemical information, particularly olfactory cues by plant volatiles, is often used for mid-range foraging, for both floral and vegetative tissues [18–21], serving as signals for host plant selection by both butterflies [22] and moths [23,24]. The diversity and quantity of these volatile cues are important for insect herbivores to locate their host plants [25–28]. Changes in abundance and diversity of volatiles affect the host-searching efficiency and potential specialization of insect herbivores [29–31]. Recent advances in chemical ecology, emphasizing the chemical information (e.g. plant volatiles) for host-searching by insect herbivores, form the basis for the Salient Aroma Hypothesis (SAH).

The synthesis and emission of plant volatiles, both defensive and non-defensive, are tied to photosynthesis and stomatal activity, and thus imply a likely circadian pattern [32]. Plants appear to emit higher amounts and more diverse blends of volatiles during the day than at night [33–35], yet a systematic investigation across diverse plant taxa is lacking.

Like plants, Lepidoptera tend to display a strong diel activity preference usually by the time they mate and forage [36]. Since foraging behaviour requires host-associated chemical information that assists decision-making, it seems likely that the diel cycle of plant volatile emissions influences the foraging choices of ovipositing herbivores. Lepidoptera that are not strictly nocturnal, which we refer to here as light-active species, are likely to encounter more volatile cues during daylight (or photophase) hours than at night (or scotophase). The enhanced availability of chemical information presumably provides greater opportunities for more precise host searching and identification.

Here, we propose the SAH (figure 1) based on the intersection between the daily cycles of plant volatile availability, the daily activity patterns of adult Lepidoptera and their host plant choices. According to this hypothesis, salient volatile cues—those that are significant for herbivores in identifying and locating host plants—may facilitate host plant selection and specialization in Lepidoptera. Thus, the evolution of host plant specialization in Lepidoptera may be influenced by ecological factors, specifically the availability and diversity of ecologically meaningful chemical information, such as plant volatiles.

To detect plant volatiles, herbivorous insects typically possess advanced olfactory abilities [37]. Olfaction is also often correlated with both the number of sensilla [38–40] and the surface area [40,41]. For example, larger antennae have been shown to be correlated with enhanced chemical sensing ability in foraging bumble bee workers [42] and males locating females by their sex pheromones [40,43–45].

In environments where there is an opportunity to utilize specific cues, sensory traits tend to be more elaborate. This occurs whether these cues are abundant or scarce. For example, animals in environments with either high or low light availability can develop complex visual systems [46]. Although much work has been done on how the light environment influences visual traits, less is known about how the chemical environment might influence olfactory traits.

Selection favours investment in sensory organs when detection of scarce, yet present stimuli is critical for survival. For example, the well-developed eyes of nocturnal species active in dim light conditions indicate an increased investment in detecting scarce yet present cues [46], as are the antennae of male moths detecting sex pheromones [43]. In herbivores, the ability to locate hosts is crucial for survival, so the high availability of plant volatiles during the day likely drives the evolution of larger olfactory structures, especially in host-seeking females.

Although the scarcity of volatiles at night may promote the development of more refined olfactory systems to detect these cues, as in the antennae of male moths, it is also possible that scarcity of volatiles can lead to reduced investment in olfactory organs. When the opportunity to use environmental cues is absent, organisms often exhibit a reduction in the corresponding sensory traits. For instance, cave-dwelling fish may lose their eyesight entirely [47]. Similarly, moths in bat-free habitats exhibit reduced auditory sensitivity [48], highlighting how relaxed selective pressures can lead to regressive evolution of sensory traits.

Variation in the availability and diversity of volatiles across time and space is exacerbated due to the transient nature of plant volatiles, which break down quickly in the atmosphere [30,49–55]. The need for specific receptors to detect these volatiles [40,56] and the necessity of detecting specific combinations of these volatiles to make them ecologically meaningful for herbivores [26] suggest that Lepidoptera active during periods with limited plant-associated volatiles might have less opportunity to utilize these plant cues. As a consequence, they may become less chemically receptive, and this could lead to less elaborate olfactory structures compared with those in environments rich in chemical information.

Following this reasoning, we argue that volatile availability (linked to diel activity) likely influences the size and/or sensitivity of Lepidoptera antennae. Given the impracticality of investigating receptor abundance and function across hundreds of species, we focused instead on measuring antennal size as a more feasible proxy for olfactory investment [38,42,57]. We predict that specialist herbivores should have larger antennae than their generalist counterparts due to the increased ecological demand for accurately locating specific host plants. We further hypothesize that species active during the day may have larger antennae given the greater availability of chemical information in daylight. For nocturnal species, two scenarios seem possible. If volatile cues are scarce but important for survival, these species might evolve larger antennae, analogous to the evolution of complex nocturnal eyes or elaborate antennae in male moths seeking females emitting pheromones [43,45]. However, if low availability of plant volatiles significantly limits the opportunity for insects to use these cues in host seeking, this could lead to reduced antennal size, especially in females.

Our analyses apply meta-analytic, phylogenetic and morphological comparative approaches to examine potential links between: (i) diel emission patterns of plant volatiles, (ii) host plant specialization of Lepidoptera, (iii) diel activity preferences of Lepidoptera and (iv) lepidopteran antennal morphology. Specifically, we investigate whether in general, plant volatiles are more abundant and diverse during the daytime than at night. We then examine whether this pattern is correlated with key behavioural and morphological traits associated with host plant specialization in Lepidoptera by assessing the activity patterns of different species, their host breadth, as well as the relative sizes of their antennae.

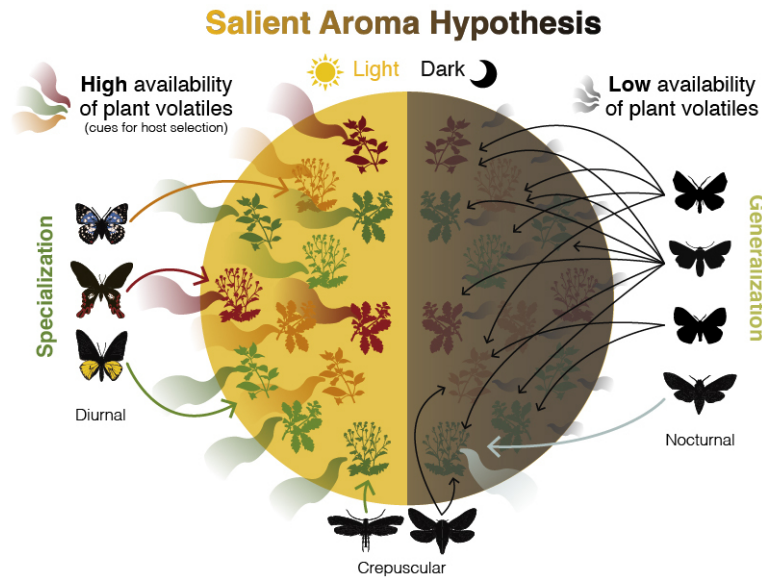


Figure 1. The Salient Aroma Hypothesis. Plants exhibit diel patterns of volatile emission, releasing more volatiles during the photophase than the scotophase, creating temporal variation in chemical information availability. This variation in chemical information may influence herbivore host specialization, with higher availability promoting specialization and lower availability favouring generalization.

2. Material and methods

(a) Literature-based characterization of plant volatile emission pattern

We searched Google Scholar for studies on diel plant volatile emissions using keywords such as ‘plant’, ‘biogenic’, ‘volatile’, ‘diel’, ‘circadian’, ‘rhythms’, ‘light or dark’ and ‘day or night’. After applying quality criteria (electronic supplementary material, note S1), we selected 115 references covering 149 cases of distinct plant species or landscapes (electronic supplementary material, table S1). Data were categorized by attributes including: (i) collection location (laboratory or landscape), (ii) plant habit (e.g. conifer, broadleaf tree, shrub, herbaceous plant), (iii) plant family and species, (iv) physical damage (undamaged or herbivory), (v) stress type (non-stressed, biotic, abiotic), (vi) volatile source (leaf, floral, landscape) and (vii) flowering time (day or night). We extracted means, standard deviations (s.d.), or standard errors (s.e.) for daytime and night-time volatile emissions and, where available, recorded the number of volatile compounds reported in the literature (electronic supplementary material, note S1).

(b) Meta-analysis of the diel volatile emission pattern

Meta-analysis synthesizes findings from multiple studies to identify patterns using effect size measures like Hedges’ g , which standardizes mean differences by variance for an unbiased effect size estimate [58,59]. We used the *metacont* function in the R package *meta* [60] to calculate Hedges’ g , quantifying differences in daytime versus night-time volatile emissions. A random-effects model (inverse variance method) accounted for variability within and between studies [61].

To assess factors influencing day–night volatile differences, we performed meta-regression, identifying sources of heterogeneity across studies [62]. This analysis covered volatiles from different sources (flowers, leaves, landscapes), plant habits (e.g. conifers, shrubs, herbaceous) and families. Small study effects were evaluated via funnel plots, trim-and-fill analysis (*trimfill* function in *meta*), and the Begg & Mazumdar Test (*metabias* function) [59], ensuring robust results (electronic supplementary material, note S2, figure S1; [60]). We also analysed day–night volatile diversity across 125 cases using a generalized linear model (package *lme4* function *glm*) with a Poisson distribution and log link function [63] predicting the number of identified compounds based on treatment (day or night).

(c) Lepidoptera phylogeny and diel activity preference

We used the most comprehensive Lepidoptera phylogeny (197 species, 84 families, 34 superfamilies) as our reference tree [64]. Diel preferences were recorded primarily from peer-reviewed sources, supplemented by online records when necessary (electronic supplementary material, table S2). Host plant data were available for 145 species and included in downstream analysis. Although diel activity is a continuous trait [65], it is typically categorized due to limited behavioural data. To account for this, we classified diel activity in two ways: a two-state model (diurnal versus nocturnal) and a four-state model (all-day, crepuscular, diurnal and nocturnal) following Kawahara *et al.* [36]. These categories apply to adult insects, particularly female host-finding behaviours, though detailed records are scarce. To avoid bias, we excluded light trap records, which may misrepresent diel activity, and did not incorporate larval diel activity preferences.

(d) Lepidoptera host plant diversity and correlated evolution of diel preference and host specificity at plant family and genus levels

For each species in the Lepidoptera phylogeny, we determined host plant preferences based mostly on peer-reviewed publications. For species where peer-reviewed publications were unavailable, we determined the host plants using field guides and databases [66].

To evaluate the correlation between Lepidoptera diel preference and host family specificity, we used a binary phylogenetic generalized linear mixed model (PGLMM), which incorporates phylogenetic relationships and random effects to account for non-independence of data points due to shared ancestry [67]. For host family specificity, species were scored as either specialist (feeding on one plant family) or generalist (feeding on multiple families) [4]. We then used the function *binaryPGLMM* from the R package *ape* [68] to test for the significance of trait correlation under the PGLMM model. We calculated phylogenetic variance (s^2) to measure trait variance attributable to shared ancestry. A significant s^2 indicates similar diel preferences among closely related species [67]. To address the issue of uneven representation across our phylogeny (further elaborated in the discussion), which is skewed towards certain lepidopteran lineages (like Papilionoidea), we implemented random sampling within our dataset to mitigate selection bias. To ensure a balanced representation and reduce potential sampling bias (electronic supplementary material, note S3, figure S2), we conducted subsampling analyses using bootstrapping with 1000 iterations with two distinct random sampling strategies. The first method equalized the number of sampled species across diel activity categories, minimizing bias related to functional comparisons. The second method balanced the representation across superfamilies, preserving phylogenetic diversity (PD) based on Lepidoptera composition [69]. Both approaches yielded results highly consistent with our main analyses (electronic supplementary material, note S3, figure S2) demonstrating the robustness of our findings and providing a framework to address taxonomic biases in similar datasets.

For an in-depth examination of host diversity and diel activity preferences among Lepidoptera, we used phylogenetic metrics to measure richness and the extent of divergence of host plants scored at genus level. Our phylogenetic assessments were anchored to a calibrated phylogeny of seed plants as outlined by Smith & Brown [70]. We pruned the tree to include two species per plant genus whose divergence time represented the crown group age of the genus. For paraphyletic genera, we used a custom Python script to identify the earliest diverging monophyletic clade to set the crown group.

Second, we used the functions *pd*, *ses.mpd* and *ses.mntd* from the R package *picante* to calculate six phylogenetic metrics to measure the richness and divergence of host plants [71]. For host plant richness, the raw number of host plant genera (electronic supplementary material, table S3) and Faith's PD were used [72]. For host plant divergence, the mean pairwise distance (MPD) and the mean nearest taxon distance (MNTD) [73] were calculated. To further assess specialization, we included the distance-based specialization index as a Z-score of MPD or MNTD, which measures deviation from a random expectation using a null model with 200 bootstrap replications [74]. These metrics provided complementary insights into host diversity and specialization across different Lepidoptera species. The complete R script used for these calculations is publicly available on GitHub (https://github.com/lmcai/day_and_night_moths).

Finally, to evaluate the significance of differences in host diversity between species exhibiting different diel activity preferences (both two-state and four-state), we conducted the PGLMM in R package *stats*. To visualize the character states across phylogeny, we reconstructed the ancestral state of diel activity (two states) and host specificity (two states) under the all-rates-different model using the *ace* function in the R package *ape* [68]. This approach enables the estimation of character state evolution while accounting for varying rates of trait change across phylogenetic branches, providing insights into how these traits evolved over time [75].

(e) Antennal size quantification

Antennal size was quantified using specimens from the Lepidoptera collection of the Museum of Comparative Zoology. We included six male and six female specimens for each species. Ten of the 197 species without adequate replicates were not included (electronic supplementary material, table S4). The dorsal and ventral sides were imaged using a high-resolution Digital Single-Lens Reflex camera (Nikon D800) for each specimen. After background removal and image segmentation in MATLAB R2019b, the estimated area of each antenna was then used as a proxy for its actual size. To take body size variation into account, we measured the length and width of the body and then normalized the antennal size by the body size, which was estimated as an ellipsoid, by taking the residuals after conducting phylogenetic GLMM [76]. This normalization process removes the body size signal while accounting for phylogenetic relationships among species (electronic supplementary material, figure S3). Both left and right antennal sizes were measured. Finally, the antennal size of each species was determined and scored as the median across replicates for each of the two sexes (electronic supplementary material, table S4).

3. Results

(a) Diel emission pattern of plant volatiles

Regardless of growth form or evolutionary history, plants emit significantly higher amounts and more diverse volatiles during the day than at night (figure 2a,b). The increase in daytime emissions is substantial (Hedges' $g = 1.66$, $Z = 8.77$, $p < 0.0001$),

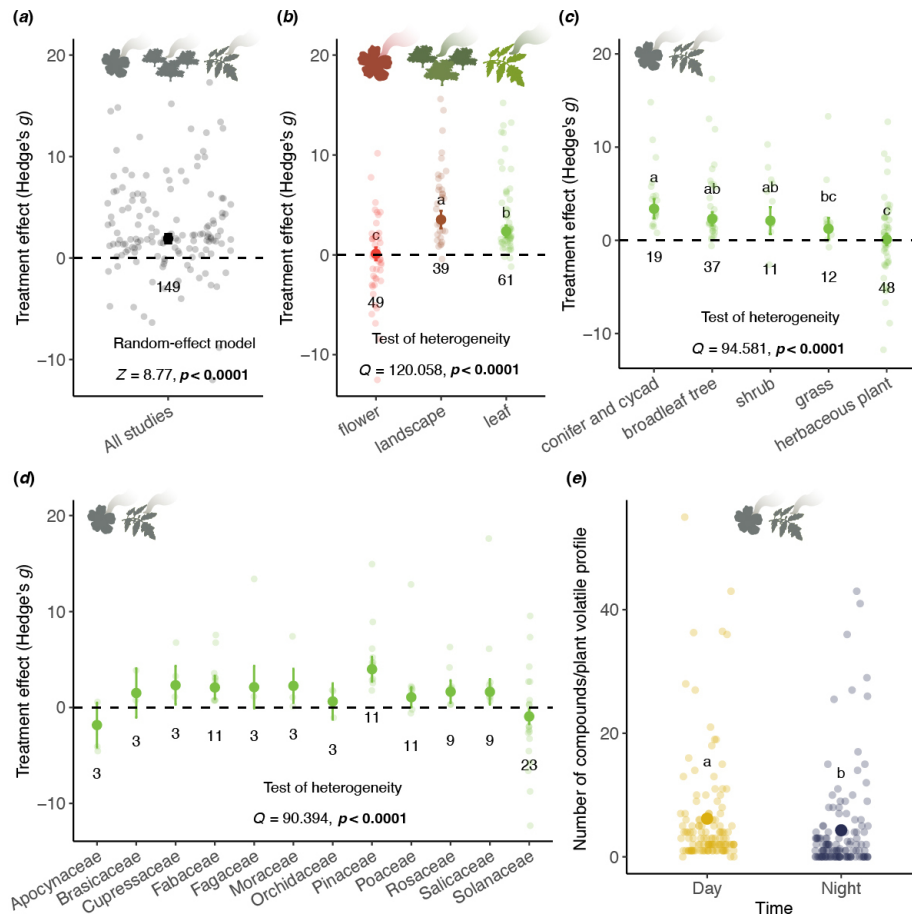


Figure 2. A survey of diel emission patterns of plant volatiles for 65 species representing 57 plant families. This result shows that plants release more volatiles during the day than at night. (a) Result of meta-analysis on diel volatile emission patterns from all studies. (b–d) Results of subgroup analyses on (b) volatile type, (c) plant habit and (d) plant family. Hedges' g indicates the standardized mean differences between daytime and night-time volatile emissions. Numbers below the circles indicate sample size. The letters above the circles indicate significant differences between subgroups. (e) Number of different compounds present in the volatile profile of different plants ($n = 125$ cases). Different letters indicate statistical significance (GLM, $p < 0.05$, Tukey HSD). Icons on top of the panels indicate volatile types.

with volatile types also varying significantly in effect size ($Q = 120.06, p < 0.0001$), indicating heterogeneity among studies [77]. Landscape-level studies report the largest day–night differences (Hedges' $g = 3.40$), with vegetative volatiles showing greater variation (Hedges' $g = 2.29$) than floral volatiles (Hedges' $g = 0.06$). Emission patterns also differ by growth habit (figure 2c, $Q = 94.58, p < 0.0001$). Woody plants, such as gymnosperms (Hedges' $g = 3.38$) and broadleaf trees (Hedges' $g = 2.30$), exhibit the highest day–night differences, followed by shrubs (Hedges' $g = 2.10$), grasses (Hedges' $g = 1.24$) and herbaceous plants (Hedges' $g = 0.09$). Significant differences also exist among plant families (figure 2d, $Q = 90.39, p < 0.0001$). While most families emit more volatiles during the day, Apocynaceae and Solanaceae emit more at night (Hedges' $g = -1.83$ and -0.94 , respectively), aligning with their recruitment of nocturnal pollinators and well-studied floral volatiles (electronic supplementary material, table S5). Night-time volatile collections contain significantly fewer compounds (figure 2e, estimate = $-0.36, p < 0.0001$), with diversity higher during the day (6.18 compounds) than at night (4.3 compounds) across 125 cases. Correlation analysis of studies with detailed compound lists ($n = 24$) confirmed a significant association between increased daytime emissions and volatile diversity (Pearson correlation = $0.593, p = 0.0022, t = 3.46, \text{d.f.} = 22$). This suggests higher volatile emissions generally correspond to greater chemical diversity.

(b) Associations between adult diel activity of Lepidoptera and host plant specificity at genus and family levels

We incorporated data for 145 out of the 197 (73.6%) species from the reference Lepidoptera phylogeny to investigate the association between diel activity and host family specificity. Our dataset included 69 light-active (7 diurnal generalists, 49 diurnal specialists, 2 crepuscular specialists and 1 crepuscular generalist, 9 all-active specialists and 1 all-active generalist) and 76 night-active species (37 nocturnal generalists, 39 nocturnal specialists). Given that the existing phylogeny is based on a restricted dataset of exemplars from only 62 out of 126 families and 31 of 46 superfamilies, we also carried out sensitivity analyses (bootstrapping with 1000 iterations) using two distinct random sampling methodologies (electronic supplementary material, note S3, figure S2).

To examine the link between lepidopteran diel activity and host plant diversity, we collected genus-level host data for 136 of 197 species (69.0%) in the higher-level Lepidoptera phylogeny (figure 3). The median number of host genera per species is 3.5, with 29.4% feeding on only one genus. Nocturnal species feed on an average of 17.12 plant genera, compared with 7.42 in diurnal species (figure 3a). Diurnal lepidopterans have significantly lower host plant PD (figure 3b, PGLMM, estimate = $-654.91, Z = -2.89, p = 0.0038$, table 1). Pairwise phylogenetic distance analyses confirm that diurnal species also have significantly lower

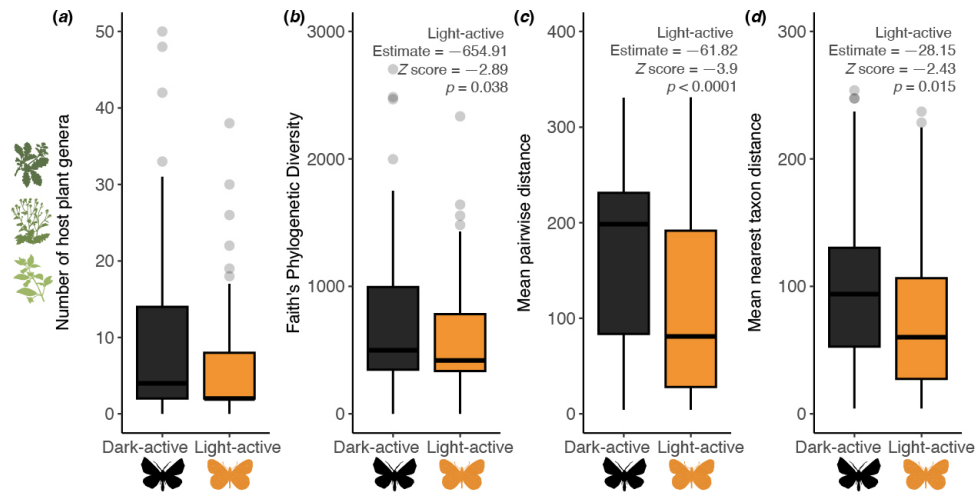


Figure 3. Host plant diversity, measured by the number of host genera consumed, varies with adult diel activity in 136 Lepidoptera species. Species that are light-active for any part of the cycle (diurnal, crepuscular or generalist) are significantly more specialized in host plant choice, both in number and PD, than nocturnal species. (a) Host plant genera per Lepidoptera species. (b) Faith's PD, reflecting evolutionary diversity of host plants, is lower in light-active species, indicating reduced phylogenetic breadth in host use. (c) MPD, measuring evolutionary divergence among host plants, is lower in light-active species, supporting their specialization. (d) MNTD, quantifying evolutionary proximity to the closest related host plant, is also lower, reinforcing their preference for closely related taxa. Statistical results are summarized in PGLMM models (table 1).

Table 1. Results of PGLMM estimates: associations between diel activity preference and host specificity (Faith's PD, MPD and MNTD). MNTD, mean nearest taxon distance; MPD, mean pairwise distance; PD, phylogenetic diversity; PGLMM, phylogenetic generalized linear mixed model.

	estimate	SE	Z-score	p-value
two-level diel activity				
PD				
intercept	1324.55	161.34	8.21	<0.0001
light-active	-654.91	226.38	-2.89	0.0038
MPD				
intercept	166.19	11.31	14.7	<0.0001
light-active	-61.82	15.86	-3.9	<0.0001
MNTD				
intercept	102.47	8.24	12.43	<0.0001
light-active	-28.15	11.56	-2.43	0.015

^a'Light-active' species are those classified as diurnal, crepuscular or all-active throughout the day and night.

Table 2. PGLMM model estimates: associations between diel activity preference and host specificity (two states). PGLMM, phylogenetic generalized linear mixed model.

	estimate	s.e.	Z-score	p-value	s ²
two-level diel activity					3.018 ($p < 0.0001$)
intercept	0.49	0.88	0.56	0.57	
light-active	1.80	0.46	3.96	<0.0001	

s² = phylogenetic signal.

host plant divergence than nocturnal species (figure 3c, PGLMM, estimate = -61.82, $Z = -3.9$, $p < 0.0001$; figure 3d, PGLMM, estimate = -28.15, $Z = -2.43$, $p = 0.015$, table 1). Expanding analyses to four diel activity states (diurnal, nocturnal, crepuscular and generalist) yielded non-significant results for PD, MPD and MNTD (electronic supplementary material, figure S5 and table S6), likely due to sample size disparities (nocturnal = 76, diurnal = 56, crepuscular = 3 and generalist = 10).

To examine the link between diel activity and host plant specificity, we classified Lepidoptera as light-active or dark-active and as specialists (feeding on a single plant family) or generalists (feeding on multiple families). Light-active species are significantly more likely to be specialists (table 2, PGLMM, estimate = 1.8, $Z = 3.96$, $p < 0.0001$), and diel activity preference shows a strong phylogenetic signal ($s^2 = 3.018$, $p < 0.0001$). Bootstrapped sensitivity analyses confirmed these results (electronic

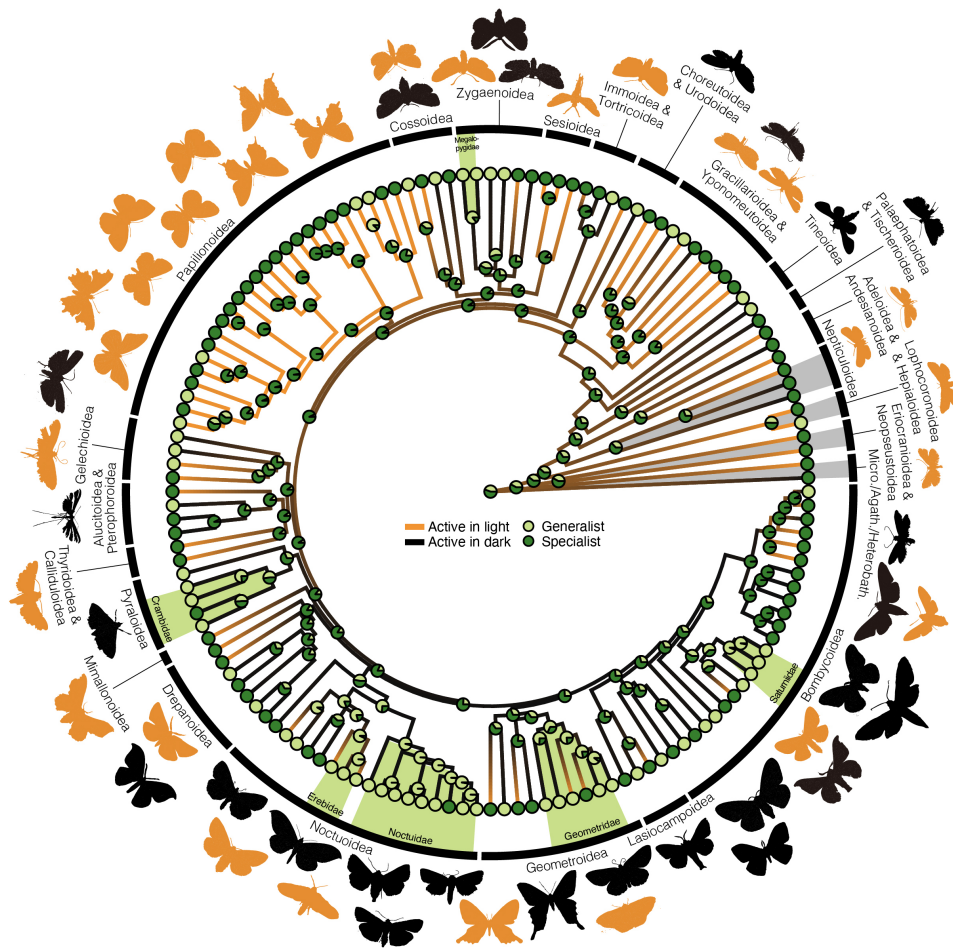


Figure 4. Ancestral state reconstruction suggests that early Lepidoptera were likely diurnally active as adults and host plant specialists as larvae. Circular diagrams at tips and nodes represent host specificity: dark green indicates specialists (single plant family), and light green indicates generalists (multiple families, posterior probability). Adult diel activity is shown by branch colour (posterior density map): yellow/black represent light- and dark-active species, respectively, with transitions marked by colour shifts. Shaded light green areas indicate ancestrally generalist clades, while shaded grey highlights early diverging lineages. Lepidoptera exhibit diverse life histories, including diurnal versus nocturnal activity and generalist versus specialist feeding. Our analysis, however, covers only a small fraction of Lepidoptera species (<0.1%).

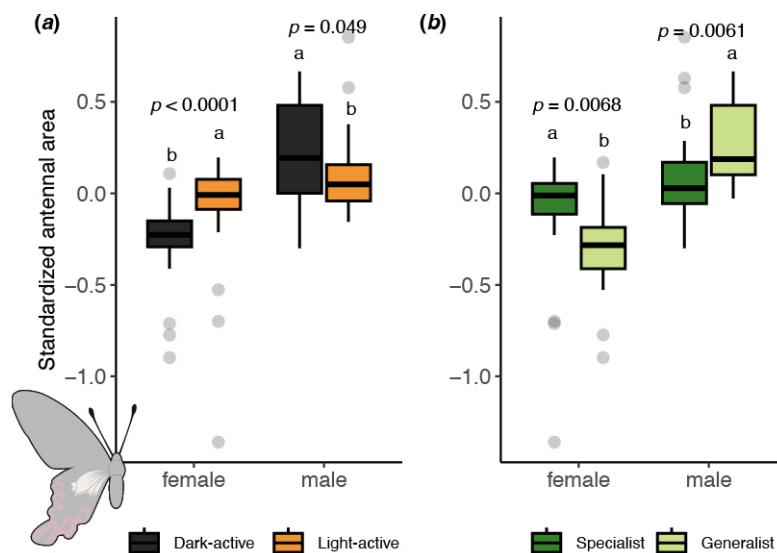


Figure 5. Antennal traits in moths and butterflies vary by diel activity and host specificity. Among Lepidoptera, females of light-active species have significantly larger relative antennae than those of dark-active species. (a) Antennal area relative to body size differs between diel activity groups ($n = 101$ light-active, $n = 86$ dark-active). (b) Relative antennal area also varies by host specificity ($n = 111$ specialists, $n = 64$ generalists). Different letters indicate statistical differences (Wilcoxon sum rank test) within each sex. Relative antennal areas account for body size and phylogenetic association.

supplementary material, figure S2, note S3), as did analyses using four diel activity states (electronic supplementary material, tables S6 and S7). Overall, our findings support the hypothesis that nocturnal Lepidoptera are more likely to have generalist diets.

Ancestral state reconstruction suggests the earliest Lepidoptera were slightly more likely to be specialists (55.42%) than generalists (44.57%) and light-active (54.27%) rather than dark-active (45.73%). Generalist diets are more common in terminal branches, while most internal nodes are inferred as specialists (67.7% on average). Notably, early-diverging ‘micromoth’ lineages (e.g. Nepticuloidea) show equivocal diet preferences (50.12–65.27% specialists) (figure 4). Clades ancestrally generalist also tend to be ancestrally nocturnal, including Actias–Attacus, Biston–Idaea, Agrotis–Amphipyra and Arctia–Amata (figure 4, electronic supplementary material, figure S4).

(c) Links between morphological traits, lepidopteran diel activity preferences and host specialization

We measured antennal and body size in 582 specimens from 94 species (electronic supplementary material, table S4) and found that antennal size varies with sex, diel activity and host diversity (figure 5). Females of light-active species have significantly larger relative antennal sizes than dark-active species (Wilcoxon test, $W = 437$, $p < 0.0001$), whereas males of light-active species have smaller relative antennae ($W = 189$, $p = 0.049$). Similarly, specialist females have significantly larger relative antennae than generalists ($W = 305$, $p = 0.006$), while specialist males have smaller relative antennae than generalists ($W = 117$, $p = 0.006$).

4. Discussion

Host plant specialization studies in the Lepidoptera have contributed much to our current understanding of coevolution and ecological specialization in general [78–82]. Numerous studies have explored the role of non-volatile plant metabolites in shaping host plant specialization patterns [12,83]. However, the potential role of plant volatiles in the evolution of host breadth remains poorly understood. Our findings suggest a potential role of plant volatile availability in influencing the evolution of host specialization of butterflies and moths. The significant correlation between lepidopteran host plant specificity and adult diel activity closely aligns with the general diel cycle of plant volatile emissions, supporting the predictions of the SAH. Specifically, we show that Lepidoptera that are active during the day (or photophase) are exposed to a greater abundance and diversity of host plant-associated volatiles. They are more likely to have specialized dietary preferences. In contrast, species active only at night (i.e. during the scotophase) encounter significantly fewer and less abundant plant volatiles. They are also less likely to specialize in particular groups of host plants.

Additionally, we found that nocturnal adult females, regardless of their degree of host specialization, tend to have smaller antennae relative to their body sizes compared with their diurnal counterparts. This suggests that the opportunity to utilize chemical information is more limited at night, leading to reduced investment in olfactory structures, which indirectly supports a key premise of the SAH. The observed differences in antennal size and diel activity patterns between sexes raise intriguing questions for future research. For instance, the smaller antennal size in nocturnal females might indicate differing sensory priorities, such as a reduced reliance on olfactory cues for activities like host location, foraging or mate finding. In contrast, the trend observed in males suggests that their increased antennal size may be influenced by different ecological pressures, such as the need to locate mates or forage efficiently. The larger antennae in generalist males may be an outcome of the scarce and unpredictable distribution of females in the environment due to the absence of a specific host plant. This unpredictability may drive a greater reliance on chemical cues for mate detection. These speculations still require further investigation, and future studies could also explore whether antennal differences correlate with other ecological or behavioural traits, such as adult foraging preferences (e.g. specialist floral visitors potentially having larger antennae). Investigating these relationships could shed light on the selective pressures driving the evolution of sensory structures in Lepidoptera.

Landscape-level studies on plant volatiles suggest that lower availability of plant volatiles at night may be a common characteristic of terrestrial ecosystems (figure 2b). However, exceptions exist at the individual plant level. While the majority of land plants exhibit increased volatile emissions during daylight hours—correlated with factors like light availability [84], photosynthesis [32], stomatal behaviour [85] and temperature [86]—certain plants instead release floral volatiles primarily at night [87]. Numerous studies, including those on model systems like *Petunia* spp. and *Nicotiana* spp., have highlighted these nocturnal emissions (figures 2b and 1c). Nocturnal pollination is a widespread phenomenon across angiosperms [88], with many night-blooming flowers emitting strong volatiles to attract nocturnal pollinators [88–91], which often feed on the same plant in their larval stages [92–94]. This suggests that plants emitting significant volatiles at night may be more noticeable (salient) to nocturnal herbivores, potentially facilitating host plant specialization among nocturnal Lepidoptera that co-occur with these plants.

An interesting observation is that these nocturnal-specialist Lepidoptera often associate with plants that release volatiles at night, such as the specialist tobacco hornworm moths, *Manduca sexta*, and their host plant tobacco [95]. In our analysis, the higher emission of volatiles at night is more commonly observed in herbaceous plants (figure 2c). Apart from the reasonable explanation that mutualism with nocturnal pollinators may be more common in herbaceous plants than other plant types, the number of studies focusing on specific herbaceous model systems may also influence these results (electronic supplementary material, table S5). For instance, *Nicotiana* spp. and *Petunia* spp. are known to have flowers that attract nocturnal pollinators [95,96] that are also specialist herbivores (e.g. tobacco hornworm, *Manduca sexta* [21]). Another well-documented case is the interaction between the evergreen shrub/tree genus *Yucca* and its specialist nocturnal herbivorous pollinator, *Tegeticula*

5. Considerations for future studies

While our findings support the SAH, several limitations must be acknowledged to refine future research. A key constraint is the absence of a well-resolved Lepidoptera phylogeny for comparative analysis [64]. Although we sampled about half of Lepidoptera families, this represents less than 0.1% of species diversity. Early branching clades like Micropterigoidea, Agathiphaogoidea and Heterobathmioidea are absent, and their life histories remain poorly known. Additionally, we undersampled speciose families such as Noctuidae, Crambidae and Pyralidae, particularly diurnal species. These gaps stem from phylogenetic and trait data constraints rather than deliberate exclusions. For instance, classifying Mimallonoidea as diurnal contradicts the common view that most mimallonids are nocturnal, underscoring the need for better sampling. Similarly, Hedylidae, a nocturnal family within diurnal Papilionoidea, was omitted due to data limitations, impeding tests of SAH predictions about nocturnality reversals and associated traits like polyphagy and antennal morphology.

Underrepresentation of clades with variable diel activity, such as Erebidae and Geometridae, limits the generalizability of our findings. Seifert *et al.* [97] found only a marginal link between diurnality and host specialization in Geometridae, aligning with SAH but highlighting the need for broader datasets and lineage-specific analyses. Furthermore, our antennal morphology analysis, constrained by uneven taxonomic coverage, relies on existing phylogenies and available specimens. These limitations emphasize the need for expanded phylogenetic and morphological data to refine future studies.

Limited taxon sampling also affects ancestral state reconstructions. Expanding coverage to include early-branching groups, Hedylidae, and diurnal species in Noctuidae and Pyralidae is critical for more reliable analyses. Future research should leverage museum specimens, enhance phylogenetic data and integrate detailed life-history information to fill these gaps. Addressing these challenges will clarify how diel activity, host specialization and antennal morphology are shaped by ecological factors like host plant volatiles, advancing our understanding of Lepidopteran specialization.

6. Conclusion

Recent advances in chemical ecology have deepened our understanding of how plant volatiles influence herbivore behaviour and evolution. The SAH offers a mechanistic framework linking plant volatile availability to host specialization in Lepidoptera. Based on plant volatile emissions, Lepidoptera diel and diet preferences, and antennal morphology, our study provides three lines of evidence supporting the SAH while offering novel insights into herbivore navigation of complex ecological landscapes. However, taxon sampling limitations warrant cautious interpretation. Despite these constraints, the SAH has significant potential for elucidating how plant volatiles drive host specialization in Lepidoptera and other herbivorous lineages.

Deviations from SAH predictions present intriguing cases for further study, helping to uncover mechanisms underlying ecological specialization and generalization. Future research should improve sampling of underrepresented Lepidoptera families and explore diel activity as a labile trait in Geometridae, Noctuidae and Pyralidae. Species-level studies will further clarify connections between diel activity, sensory adaptations and host plant specialization. Additionally, the SAH offers opportunities to investigate multi-modal plant–insect interactions, integrating sensory modalities like visual and tactile cues. Addressing current limitations and adopting a multidisciplinary approach will refine hypothesis testing, revealing new dimensions of plant–insect interactions and ecological specialization.

Ethics. We utilize the insect collection from the Harvard Museum of Comparative Zoology, and no permits are required for this work.

Data accessibility. The data supporting the findings of this study are included as supplementary information accompanying this publication.

Supplementary material is available online [98].

Declaration of AI use. We have not used AI-assisted technologies in creating this article.

Authors' contributions. P.A.L.: conceptualization, data curation, formal analysis, funding acquisition, investigation, methodology, project administration, resources, software, supervision, validation, visualization, writing—original draft, writing—review and editing; W.P.C.: data curation, formal analysis, investigation, methodology, software, supervision, validation, visualization, writing—original draft, writing—review and editing; L.C.: data curation, formal analysis, investigation, methodology, resources, software, visualization, writing—original draft, writing—review and editing; Y.H.: methodology, validation, writing—review and editing; E.D.: data curation, investigation, methodology, resources, validation; K.J.G.: conceptualization, supervision, writing—review and editing; N.E.P.: conceptualization, data curation, funding acquisition, project administration, resources, supervision, validation, writing—review and editing; G.F.: funding acquisition, project administration, resources, supervision, validation, writing—review and editing.

All authors gave final approval for publication and agreed to be held accountable for the work performed therein.

Conflict of interest declaration. We declare we have no competing interests.

Funding. This work was supported by the NSTC of Taiwan Funding [112-2313-B-002-007-MY2, NSTC 113-2313-B-002-044-MY3 and NSTC 113-2628-B-002-025 to Po-An Lin], NSF of USA [DEB-1541560 to Naomi Pierce]. Po-An Lin was supported by the Yushan Fellowship Program from the Ministry of Education of Taiwan [MOE-112 YSFAG-0003-001-P1]. Liming Cai was supported by the Stengl–Wyer postdoctoral fellowship from the University of Texas at Austin.

Acknowledgements. We thank Crystal Maier and Rachel Hawkins for assistance with specimens from the Museum of Comparative Zoology. Katherine Angier, Cong Liu, Annina Kennedy Yoon, Jomar Hinolan, Micael Gabriel Itliong and Valentina Morgan assisted with imaging specimens for analysis. We thank Dr Po-Ju Ke and Dr Hsi-Cheng Ho for their assistance in revising and proofreading the manuscript.

References

1. Futuyma DJ, Moreno G. 1988 The evolution of ecological specialization. *Annu. Rev. Ecol. Syst.* **19**, 207–233. (doi:10.1146/annurev.es.19.110188.001231)
2. Thompson JN. 2005 *The geographic mosaic of coevolution*. Chicago, IL: University of Chicago Press.
3. Bernays E, Graham M. 1988 On the evolution of host specificity in phytophagous arthropods. *Ecology* **69**, 886–892. (doi:10.2307/1941237)
4. Forister ML *et al.* 2015 The global distribution of diet breadth in insect herbivores. *Proc. Natl Acad. Sci. USA* **112**, 442–447. (doi:10.1073/pnas.1423042112)
5. Ali JG, Agrawal AA. 2012 Specialist versus generalist insect herbivores and plant defense. *Trends Plant Sci.* **17**, 293–302. (doi:10.1016/j.tplants.2012.02.006)
6. Edger PP *et al.* 2015 The butterfly plant arms-race escalated by gene and genome duplications. *Proc. Natl Acad. Sci. USA* **112**, 8362–8366. (doi:10.1073/pnas.1503926112)
7. Endara MJ, Coley PD, Ghabash G, Nicholls JA, Dexter KG, Donoso DA, Stone GN, Pennington RT, Kursar TA. 2017 Coevolutionary arms race versus host defense chase in a tropical herbivore-plant system. *Proc. Natl Acad. Sci. USA* **114**, E7499–E7505. (doi:10.1073/pnas.1707727114)
8. Wheat CW, Vogel H, Wittstock U, Braby MF, Underwood D, Mitchell-Olds T. 2007 The genetic basis of a plant-insect coevolutionary key innovation. *Proc. Natl Acad. Sci. USA* **104**, 20427–20431. (doi:10.1073/pnas.0706229104)
9. Wahlberg N, Wheat CW, Peña C. 2013 Timing and patterns in the taxonomic diversification of Lepidoptera (butterflies and moths). *PLoS One* **8**, e80875. (doi:10.1371/journal.pone.0080875)
10. Agosta SJ, Klemens JA. 2009 Resource specialization in a phytophagous insect: no evidence for genetically based performance trade-offs across hosts in the field or laboratory. *J. Evol. Biol.* **22**, 907–912. (doi:10.1111/j.1420-9101.2009.01694.x)
11. Seifert CL, Fiedler K. 2024 Macroecological patterns in European butterflies unveil strong interrelations between larval diet breadth, latitudinal range size and voltinism. *Ecography* **2024**, e07021. (doi:10.1111/ecog.07021)
12. Hardy NB, Kaczmarski C, Bird G, Normark BB. 2020 What we don't know about diet-breadth evolution in herbivorous insects. *Annu. Rev. Ecol. Syst.* **51**, 103–122. (doi:10.1146/annurev-ecolsys-011720-023322)
13. Forister ML, Dyer LA, Singer MS, Stireman JO 3rd, Lill JT. 2012 Revisiting the evolution of ecological specialization, with emphasis on insect-plant interactions. *Ecology* **93**, 981–991. (doi:10.1890/11-0650.1)
14. Rojas JC, Wyatt TD. 1999 Role of visual cues and interaction with host odour during the host-finding behaviour of the cabbage moth. *Entomol. Exp. Appl.* **91**, 59–65. (doi:10.1046/j.1570-7458.1999.00466.x)
15. Yang X, Fan F, An L, Li M, Wei G. 2019 Oviposition preferences in: the relative importance of visual and olfactory cues. *Entomol. Exp. et Appl.* **167**, 722–728. (doi:10.1111/eea.12828)
16. Nagaya H, Stewart FJ, Kinoshita M. 2021 Swallowtail butterflies use multiple visual cues to select oviposition sites. *Insects* **12**, 1047. (doi:10.3390/insects12111047)
17. Ozaki K, Ryuda M, Yamada A, Utoguchi A, Ishimoto H, Calas D, Marion-Poll F, Tanimura T, Yoshikawa H. 2011 A gustatory receptor involved in host plant recognition for oviposition of a swallowtail butterfly. *Nat. Commun.* **2**, 542. (doi:10.1038/ncomms1548)
18. Honda K, Ômura H, Hayashi N. 1998 Identification of floral volatiles from that stimulate flower-visiting by cabbage butterfly. *J. Chem. Ecol.* **24**, 2167–2180. (doi:10.1023/A:1020750029362)
19. Tang YC, Zhou CL, Chen XM, Zheng H. 2013 Visual and olfactory responses of seven butterfly species during foraging. *J. Insect Behav.* **26**, 387–401. (doi:10.1007/s10905-012-9358-0)
20. Schäpers A, Carlsson MA, Gamberale-Stille G, Janz N. 2015 The role of olfactory cues for the search behavior of a specialist and generalist butterfly. *J. Insect Behav.* **28**, 77–87. (doi:10.1007/s10905-014-9482-0)
21. Haverkamp A, Hansson BS, Baldwin IT, Knaden M, Yon F. 2018 Floral trait variations among wild tobacco populations influence the foraging behavior of Hawkmoth pollinators. *Front. Ecol. Evol.* **6**. (doi:10.3389/fevo.2018.00019)
22. Carlsson MA, Bisch-Knaden S, Schäpers A, Mozuraitis R, Hansson BS, Janz N. 2011 Odour maps in the brain of butterflies with divergent host-plant preferences. *PLoS One* **6**, e24025. (doi:10.1371/journal.pone.0024025)
23. Liu XL *et al.* 2020 The molecular basis of host selection in a crucifer-specialized moth. *Curr. Biol.* **30**, 4476–4482. (doi:10.1016/j.cub.2020.08.047)
24. Yactayo-Chang JP, Mendoza J, Willms SD, Rering CC, Beck JJ, Block AK. 2021 *Zea mays* volatiles that influence oviposition and feeding behaviors of *Spodoptera frugiperda*. *J. Chem. Ecol.* **47**, 799–809. (doi:10.1007/s10886-021-01302-w)
25. Bruce TJA, Wadhams LJ, Woodcock CM. 2005 Insect host location: a volatile situation. *Trends Plant Sci.* **10**, 269–274. (doi:10.1016/j.tplants.2005.04.003)
26. Bruce TJA, Pickett JA. 2011 Perception of plant volatile blends by herbivorous insects—finding the right mix. *Phytochemistry* **72**, 1605–1611. (doi:10.1016/j.phytochem.2011.04.011)
27. Conchou L, Lucas P, Meslin C, Proffitt M, Staudt M, Renou M. 2019 Insect odorscapes: from plant volatiles to natural olfactory scenes. *Front. Physiol.* **10**, 972. (doi:10.3389/fphys.2019.00972)
28. Shao X, Cheng K, Wang Z, Zhang Q, Yang X. 2021 Use of odor by host-finding insects: the role of real-time odor environment and odor mixing degree. *Chemoecology* **31**, 149–158. (doi:10.1007/s00049-021-00342-8)
29. Burkle LA, Runyon JB. 2016 Drought and leaf herbivory influence floral volatiles and pollinator attraction. *Glob. Chang. Biol.* **22**, 1644–1654. (doi:10.1111/gcb.13149)
30. Li T, Blande JD, Holopainen JK. 2016 Atmospheric transformation of plant volatiles disrupts host plant finding. *Sci. Rep.* **6**, 33851. (doi:10.1038/srep33851)
31. Chan JK, Parasurama S, Atlas R, Xu R, Jongebloed UA, Alexander B, Langenhan JM, Thornton JA, Riffell JA. 2024 Olfaction in the anthropocene: NO₃ negatively affects floral scent and nocturnal pollination. *Science* **383**, 607–611. (doi:10.1126/science.adi0858)
32. Niinemets U, Loreto F, Reichstein M. 2004 Physiological and physicochemical controls on foliar volatile organic compound emissions. *Trends Plant Sci.* **9**, 180–186. (doi:10.1016/j.tplants.2004.02.006)
33. Ibrahim MA, Egu MC, Kasurinen A, Yahya A, Holopainen JK. 2010 Diversity of volatile organic compound emissions from flowering and vegetative branches of yeheb, *Cordeauxia edulis* (Caesalpinaceae), a threatened evergreen desert shrub. *Flavour Fragr. J.* **25**, 83–92. (doi:10.1002/ffj.1971)
34. Yáñez-Serrano AM *et al.* 2015 Diel and seasonal changes of biogenic volatile organic compounds within and above an Amazonian rainforest. *Atmos. Chem. Phys.* **15**, 3359–3378. (doi:10.5194/acp-15-3359-2015)
35. Ramos YJ, Machado D de B, Queiroz GA de, Guimarães EF, Defaveri ACA e, Moreira D de L. 2020 Chemical composition of the essential oils of circadian rhythm and of different vegetative parts from Kunth – a medicinal plant from Brazil. *Biochem. Syst. Ecol.* **92**, 104116. (doi:10.1016/j.bse.2020.104116)
36. Kawahara AY, Plotkin D, Hamilton CA, Gough H, St Laurent R, Owens HL, Homziak NT, Barber JR. 2018 Diel behavior in moths and butterflies: a synthesis of data illuminates the evolution of temporal activity. *Org. Divers. Evol.* **18**, 13–27. (doi:10.1007/s13127-017-0350-6)

37. Hansson BS, Stensmyr MC. 2011 Evolution of insect olfaction. *Neuron* **72**, 698–711. (doi:10.1016/j.neuron.2011.11.003)
38. Chapman R. 1982 Chemoreception: the significance of receptor numbers. In *Advances in insect physiology* (eds M Berridge, J Treherne, V Wigglesworth), pp. 247–356. San Diego, CA: Academic Press. (doi:10.1016/S0065-2806(08)60155-1)
39. Wittwer B, Hefetz A, Simon T, Murphy LEK, Elgar MA, Pierce NE, Kocher SD. 2017 Solitary bees reduce investment in communication compared with their social relatives. *Proc. Natl Acad. Sci. USA* **114**, 6569–6574. (doi:10.1073/pnas.1620780114)
40. Elgar MA, Zhang D, Wang Q, Wittwer B, Thi Pham H, Johnson TL, Freelance CB, Coquilleau M. 2018 Insect antennal morphology: the evolution of diverse solutions to odorant perception. *Yale J. Biol. Med.* **91**, 457–469.
41. Wang Q, Shang Y, Hilton DS, Inthavong K, Zhang D, Elgar MA. 2018 Antennal scales improve signal detection efficiency in moths. *Proc. R. Soc. B* **285**, 20172832. (doi:10.1098/rspb.2017.2832)
42. Spaethe J, Brockmann A, Halbig C, Tautz J. 2007 Size determines antennal sensitivity and behavioral threshold to odors in bumblebee workers. *Naturwissenschaften* **94**, 733–739. (doi:10.1007/s00114-007-0251-1)
43. Symonds MR, Johnson TL, Elgar MA. 2012 Pheromone production, male abundance, body size, and the evolution of elaborate antennae in moths. *Ecol. Evol.* **2**, 227–246. (doi:10.1002/ece3.81)
44. Streinzer M, Kelber C, Pfabigan S, Kleineidam CJ, Spaethe J. 2013 Sexual dimorphism in the olfactory system of a solitary and a eusocial bee species. *J. Comp. Neurol.* **521**, 2742–2755. (doi:10.1002/cne.23312)
45. Nicolas A, Pourtois J, Karier P, Markó I, Hance T. 2020 Sexual dimorphism and sex pheromone detection in *Aphidoletes aphidimyza*. *Anim. Biol.* **70**, 13–25. (doi:10.1163/15707563-20191038)
46. Warrant EJ, Johnsen S. 2013 Vision and the light environment. *Curr. Biol.* **23**, R990–R994. (doi:10.1016/j.cub.2013.10.019)
47. Porter ML, Sumner-Rooney L. 2018 Evolution in the dark: unifying our understanding of eye loss. *Integr. Comp. Biol.* **58**, 367–371. (doi:10.1093/icb/icy082)
48. Fullard JH. 1994 Auditory changes in noctuid moths endemic to a bat-free habitat. *J. Evol. Biol.* **7**, 435–445. (doi:10.1046/j.1420-9101.1994.7040435.x)
49. Calogirou A, Larsen BR, Kotzias D. 1999 Gas-phase terpene oxidation products: a review. *Atmos. Environ.* **33**, 1423–1439. (doi:10.1016/S1352-2310(98)00277-5)
50. Atkinson R, Arey J. 2003 Gas-phase tropospheric chemistry of biogenic volatile organic compounds: a review. *Atmos. Environ.* **37**, S197–S219. (doi:10.1016/S1352-2310(03)00391-1)
51. Holopainen JK, Blande JD. 2013 Where do herbivore-induced plant volatiles go? *Front. Plant Sci.* **4**, 185. (doi:10.3389/fpls.2013.00185)
52. Blande JD, Holopainen JK, Niinemets U. 2014 Plant volatiles in polluted atmospheres: stress responses and signal degradation. *Plant Cell Environ.* **37**, 1892–1904. (doi:10.1111/pce.12352)
53. Holopainen JK, Kivimäenpää M, Nizkorodov SA. 2017 Plant-derived secondary organic material in the air and ecosystems. *Trends Plant Sci.* **22**, 744–753. (doi:10.1016/j.tplants.2017.07.004)
54. Roggatz CC, Saha M, Blanchard S, Schirmacher P, Fink P, Verheggen F, Hardege JD. 2022 Becoming nose-blind—climate change impacts on chemical communication. *Glob. Chang. Biol.* **28**, 4495–4505. (doi:10.1111/gcb.16209)
55. Touhami D, Mofikoya AO, Girling RD, Langford B, Misztal PK, Pfrang C. 2024 Atmospheric degradation of ecologically important biogenic volatiles: investigating the ozonolysis of (E)- β -ocimene, isomers of α and β -farnesene, α -terpinene and 6-methyl-5-hepten-2-one, and their gas-phase products. *J. Chem. Ecol.* **50**, 129–142. (doi:10.1007/s10886-023-01467-6)
56. Tanaka K, Uda Y, Ono Y, Nakagawa T, Suwa M, Yamaoka R, Touhara K. 2009 Highly selective tuning of a silkworm olfactory receptor to a key mulberry leaf volatile. *Curr. Biol.* **19**, 881–890. (doi:10.1016/j.cub.2009.04.035)
57. Silva IM da, Pereira K de S, Spranghers T, Zanuncio JC, Serrão JE. 2016 Antennal sensilla and sexual dimorphism of the parasitoid *Trichospilus pupivorus* (Hymenoptera: Eulophidae). *Microsc. Microanal.* **22**, 913–921. (doi:10.1017/S1431927616011314)
58. Hedges LV, Olkin I. 2014 *Statistical methods for meta-analysis*. San Diego, CA: Academic Press.
59. Schwarzer G, Carpenter R JR, Rücker G. 2015 *Meta-analysis with R*. Berlin, Germany: Springer.
60. Balduzzi S, Rücker G, Schwarzer G. 2019 How to perform a meta-analysis with R: a practical tutorial. *Evid. Based Ment. Health* **22**, 153–160. (doi:10.1136/ebmental-2019-300117)
61. Borenstein M, Hedges L, Higgins J, Rothstein H. 2021 *Introduction to meta-analysis*. Hoboken, NJ: John Wiley & Sons.
62. Thompson SG, Higgins JPT. 2002 How should meta-regression analyses be undertaken and interpreted? *Stat. Med.* **21**, 1559–1573. (doi:10.1002/sim.1187)
63. Bates D. 2014 Fitting linear mixed-effects models using lme4. *arXiv preprint*. (doi:10.18637/jss.v067.i01)
64. Kawahara AY *et al.* 2019 Phylogenomics reveals the evolutionary timing and pattern of butterflies and moths. *Proc. Natl Acad. Sci. USA* **116**, 22657–22663. (doi:10.1073/pnas.1907847116)
65. Fullard JH, Napoleone N. 2001 Diel flight periodicity and the evolution of auditory defences in the Macrolepidoptera. *Anim. Behav.* **62**, 349–368. (doi:10.1006/anbe.2001.1753)
66. Robinson G, Ackery P, Kitching I, Beccaloni G, Hernández L. 2010 HOSTS - a database of the world's Lepidopteran hostplants. London, UK: Natural History Museum. See <https://www.nhm.ac.uk/hosts>.
67. Ives AR, Garland TJ Jr. 2010 Phylogenetic logistic regression for binary dependent variables. *Syst. Biol.* **59**, 9–26. (doi:10.1093/sysbio/syp074)
68. Paradis E, Claude J, Strimmer K. 2004 APE: Analyses of Phylogenetics and Evolution in R language. *Bioinformatics* **20**, 289–290. (doi:10.1093/bioinformatics/btg412)
69. Goldstein PZ. 2017 Diversity and significance of lepidoptera: a phylogenetic perspective. In *Insect biodivers* (eds RG Footitt, PH Adler), pp. 463–495. (doi:10.1002/9781118945568)
70. Smith SA, Brown JW. 2018 Constructing a broadly inclusive seed plant phylogeny. *Am. J. Bot.* **105**, 302–314. (doi:10.1002/ajb2.1019)
71. Kembel SW, Cowan PD, Helmus MR, Cornwell WK, Morlon H, Ackerly DD, Blomberg SP, Webb CO. 2010 Picante: r tools for integrating phylogenies and ecology. *Bioinformatics* **26**, 1463–1464. (doi:10.1093/bioinformatics/btq166)
72. Faith DP. 1992 Conservation evaluation and phylogenetic diversity. *Biol. Conserv.* **61**, 1–10. (doi:10.1016/0006-3207(92)91201-3)
73. Webb CO, Ackerly DD, McPeck MA, Donoghue MJ. 2002 Phylogenies and community ecology. *Annu. Rev. Ecol. Syst.* **33**, 475–505. (doi:10.1146/annurev.ecolsys.33.010802.150448)
74. Jorge LR, Prado PI, Almeida-Neto M, Lewinsohn TM. 2014 An integrated framework to improve the concept of resource specialisation. *Ecol. Lett.* **17**, 1341–1350. (doi:10.1111/ele.12347)
75. Pagel M. 1999 Inferring the historical patterns of biological evolution. *Nature* **401**, 877–884. (doi:10.1038/44766)
76. Lynch M. 1991 Methods for the analysis of comparative data in evolutionary biology. *Evolution* **45**, 1065–1080. (doi:10.1111/j.1558-5646.1991.tb04375.x)
77. Cochran WG. 1954 The combination of estimates from different experiments. *Biometrics* **10**, 101. (doi:10.2307/3001666)
78. Ehrlich PR, Raven PH. 1964 Butterflies and plants: a study in coevolution. *Evolution* **18**, 586. (doi:10.2307/2406212)

79. Janz N, Nylin S. 2008 The oscillation hypothesis of host-plant range and speciation. In *Specialization, speciation, and radiation: the evolutionary biology of herbivorous insects*, pp. 203–215. Berkeley, CA: University of California Press. (doi:10.1525/california/9780520251328.003.0015)
80. Hardy NB, Otto SP. 2014 Specialization and generalization in the diversification of phytophagous insects: tests of the musical chairs and oscillation hypotheses. *Proc. R. Soc. B* **281**, 20132960. (doi:10.1098/rspb.2013.2960)
81. Braga MP, Guimarães PR Jr, Wheat CW, Nylin S, Janz N. 2018 Unifying host-associated diversification processes using butterfly-plant networks. *Nat. Commun.* **9**, 5155. (doi:10.1038/s41467-018-07677-x)
82. Agrawal AA, Zhang X. 2021 The evolution of coevolution in the study of species interactions. *Evolution* **75**, 1594–1606. (doi:10.1111/evo.14293)
83. Ehrlich PR, Murphy DD. 1988 Plant chemistry and host range in insect herbivores. *Ecology* **69**, 908–909. (doi:10.2307/1941244)
84. He J, Halitschke R, Schuman MC, Baldwin IT. 2021 Light dominates the diurnal emissions of herbivore-induced volatiles in wild tobacco. *BMC Plant Biol.* **21**, 401. (doi:10.1186/s12870-021-03179-z)
85. Seidl-Adams I, Richter A, Boomer KB, Yoshinaga N, Degenhardt J, Tumlinson JH. 2015 Emission of herbivore elicitor-induced sesquiterpenes is regulated by stomatal aperture in maize (*Zea mays*) seedlings. *Plant Cell Environ.* **38**, 23–34. (doi:10.1111/pce.12347)
86. Chen J, Bi H, Yu X, Fu Y, Liao W. 2019 Influence of physiological and environmental factors on the diurnal variation in emissions of biogenic volatile compounds from pinus tabuliformis. *J. Environ. Sci.* **81**, 102–118. (doi:10.1016/j.jes.2019.01.020)
87. Fenske MP, Imaizumi T. 2016 Circadian rhythms in floral scent emission. *Front. Plant Sci.* **7**, 462. (doi:10.3389/fpls.2016.00462)
88. Borges RM, Somanathan H, Kelber A. 2016 Patterns and processes in nocturnal and crepuscular pollination services. *Q. Rev. Biol.* **91**, 389–418. (doi:10.1086/689481)
89. Raguso RA. 2008 Wake up and smell the roses: the ecology and evolution of floral scent. *Annu. Rev. Ecol. Syst.* **39**, 549–569. (doi:10.1146/annurev.ecolsys.38.091206.095601)
90. Tröger A *et al.* 2021 Tetranorsesquiterpenoids as attractants of yucca moths to yucca flowers. *J. Chem. Ecol.* **47**, 1025–1041. (doi:10.1007/s10886-021-01308-4)
91. Buxton MN, Gaskett AC, Lord JM, Pattermore DE. 2022 A global review demonstrating the importance of nocturnal pollinators for crop plants. *J. Appl. Ecol.* **59**, 2890–2901. (doi:10.1111/1365-2664.14284)
92. Powell JA. 1992 Interrelationships of yuccas and yucca moths. *Trends Ecol. Evol.* **7**, 10–15. (doi:10.1016/0169-5347(92)90191-D)
93. Bronstein JL, Huxman T, Horvath B, Farabee M, Davidowitz G. 2009 Reproductive biology of *Datura wrightii*: the benefits of a herbivorous pollinator. *Ann. Bot.* **103**, 1435–1443. (doi:10.1093/aob/mcp053)
94. Balbuena MS, Broadhead GT, Dahake A, Barnett E, Vergara M, Skogen KA, Jogesh T, Raguso RA. 2022 Mutualism has its limits: consequences of asymmetric interactions between a well-defended plant and its herbivorous pollinator. *Phil. Trans. R. Soc. Lond. B Biol. Sci.* **377**, 20210166. (doi:10.1098/rstb.2021.0166)
95. Raguso RA, Levin RA, Foose SE, Holmberg MW, McDade LA. 2003 Fragrance chemistry, nocturnal rhythms and pollination 'syndromes' in *Nicotiana*. *Phytochemistry* **63**, 265–284. (doi:10.1016/s0031-9422(03)00113-4)
96. Klahre U, Gurba A, Hermann K, Saxenhofer M, Bossolini E, Guerin PM, Kuhlmeier C. 2011 Pollinator choice in *Petunia* depends on two major genetic loci for floral scent production. *Curr. Biol.* **21**, 730–739. (doi:10.1016/j.cub.2011.03.059)
97. Seifert CL, Strutzenberger P, Fiedler K. 2024 Are day-flying moths more specialized in larval dietary breadth? A test of the 'salient aroma hypothesis' in a predominantly nocturnal clade. *Evolution* **78**, 1174–1182. (doi:10.1093/evolut/qpae052)
98. Lin PA, Chan WP, Cai L, Hsiao Y, Dankowicz E, Gilbert KJ *et al.* 2025 Supplementary material from: The Salient Aroma Hypothesis: Host plant specialization is linked with plant volatile availability in Lepidoptera. Figshare. (doi:10.6084/m9.figshare.c.7684197)