



# **Dysregulation of Liver Regeneration by Hepatitis B Virus Infection: Impact on Development of Hepatocellular Carcinoma**

Eun-Sook Park<sup>1,†</sup>, Mehrangiz Dezhbord<sup>2,†</sup>, Ah Ram Lee<sup>2</sup>, Bo Bae Park<sup>1</sup> and Kyun-Hwan Kim<sup>2,\*</sup>

- <sup>1</sup> Institute of Biomedical Science and Technology, School of Medicine, Konkuk University, Seoul 05029, Korea; espark97@gmail.com (E.-S.P.); bbpark1@gmail.com (B.B.P.)
- <sup>2</sup> Department of Precision Medicine, School of Medicine, Sungkyunkwan University, Suwon 16419, Korea; m.dezhbord@yonsei.ac.kr (M.D.); ahram2g@naver.com (A.R.L.)
- \* Correspondence: khkim10@skku.edu; Tel.: +82-31-299-6126
- + These authors contributed equally to this work.

**Simple Summary:** Liver regeneration after injury or partial removal is a complicated and critical process that can be affected by several factors such as hepatitis B virus (HBV) or intracellular signaling pathways. Understanding regeneration repressive mechanisms provides insight to encounter these inhibitory factors and prevent further complications including liver cancer. Here we summarized pathways of liver regeneration and pointed out in detail, the different mechanisms that may dysregulate this process during HBV infection. In addition, other intracellular factors and pathways related to the regulation of liver regeneration have been recapped. The overall information provided here can be beneficial for understanding the method of preventing and treatment of liver failure due to the defective regeneration.

Abstract: The liver is unique in its ability to regenerate in response to damage. The complex process of liver regeneration consists of multiple interactive pathways. About 2 billion people worldwide have been infected with hepatitis B virus (HBV), and HBV causes 686,000 deaths each year due to its complications. Long-term infection with HBV, which causes chronic inflammation, leads to serious liver-related diseases, including cirrhosis and hepatocellular carcinoma. HBV infection has been reported to interfere with the critical mechanisms required for liver regeneration. In this review, the studies on liver tissue characteristics and liver regeneration mechanisms are summarized. Moreover, the inhibitory mechanisms of HBV infection in liver regeneration are investigated. Finally, the association between interrupted liver regeneration and hepatocarcinogenesis, which are both triggered by HBV infection, is outlined. Understanding the fundamental and complex liver regeneration process is expected to provide significant therapeutic advantages for HBV-associated hepatocellular carcinoma.

Keywords: liver; liver regeneration; hepatitis B virus; HBV-related hepatocellular carcinoma

## 1. Introduction

Infection with hepatitis B virus (HBV) results in a variety of liver diseases, including acute or chronic inflammation, cirrhosis, and hepatocellular carcinoma (HCC), leading to liver injury [1–3].

The liver is known as the only organ that is capable of completely regenerating itself after partial hepatectomy or after being damaged by various stimuli such as toxins and pathogens. The study of liver regeneration started with a model of partial hepatectomy in rodents in 1931 [4], and research on molecular mechanisms occurring during liver regeneration has been actively conducted until recently.

The hepatocyte loss caused by long-term liver damage is compensated by activating liver regeneration signals that trigger the very complex and highly organized hepatic regeneration process which occurs through the following three steps. Many growth factors



Citation: Park, E.-S.; Dezhbord, M.; Lee, A.R.; Park, B.B.; Kim, K.-H. Dysregulation of Liver Regeneration by Hepatitis B Virus Infection: Impact on Development of Hepatocellular Carcinoma. *Cancers* **2022**, *14*, 3566. https://doi.org/10.3390/ cancers14153566

Academic Editor: Marcello Persico

Received: 29 June 2022 Accepted: 21 July 2022 Published: 22 July 2022

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). and cytokines are involved in each tightly controlled phase [5,6]. Regeneration begins when cytokines such as interleukin-6 (IL-6) are secreted from Kupffer cells in the liver. Next, hepatocyte division is activated by growth factors such as hepatocyte growth factor (HGF) and transforming growth factor-alpha (TGF- $\alpha$ ). The final stage of regeneration is regulated by cytokines of the transforming growth factor-beta (TGF- $\beta$ ) family [5–11].

Inadequate liver regeneration causes chronic and rapid turnover of hepatocytes, which can be an important factor in liver dysfunction and eventually development of the HCC [1,2]. Balanced liver regeneration is essential for liver homeostasis. Therefore, disruption of this process by viral infection may be a major cause of various virus-mediated liver diseases, including liver cancer. In this review, we summarize the dysregulation of liver regeneration by HBV and its effect on liver regeneration in the development of HBV-related HCC.

#### 2. Pathway of Liver Regeneration

Successful liver regeneration is the restoration of the liver size and number of hepatocytes to the same level as prior to the injury. This characteristic of liver regeneration is very important for maintaining liver function homeostasis. Liver regeneration is an organized response that is induced by specific external stimuli and accompanies by successive changes in gene expression, growth factor production, and morphological structure [6].

Experimentally, the liver regeneration can be induced by various methods. Several models have been proposed for the study of liver regeneration. The most preferred method is two-thirds partial hepatectomy, which is widely used for studying liver regeneration in experimental animals [6,12]. Therefore, here, we mainly discuss the liver regeneration triggered after partial hepatectomy.

The liver regeneration process can generally be divided into three stages, namely initiation, proliferation, and termination, with various molecules participating in these three stages [7].

In the liver tissue of adults, division of hepatocytes occurs infrequent [3]. Nevertheless, following partial removal of the liver tissue, almost all the hepatocytes start to enter the replicative phase immediately [13]. Most hepatocytes are in a quiescent state and require priming before replication. Cytokines are involved in the initiation of liver regeneration. Tumor necrosis factor alpha (TNF- $\alpha$ ) and IL-6 are the most important initiating factors in regenerating liver. After partial hepatectomy, TNF binds to the TNF receptor 1 on Kupffer cells, leading to the activation of NF-kB and production of IL-6 [14]. IL-6 activates the signal transducer and activator of transcription 3 (STAT3) and extracellular signalrelated kinase 1 and 2 (ERK1/2) pathways on hepatocytes [15]. The addition of anti-TNF antibodies after partial hepatectomy significantly inhibits IL-6 production and also inhibits liver regeneration in a rat model [16]. The IL-6 knockout mouse demonstrated impaired liver regeneration with liver necrosis and failure [17]. It has been recently demonstrated that the STAT3 activity in mice is inhibited by CREB/ATF bZIP transcription factor CREBZF, which results in repression of liver regeneration. Regarding the critical role of STAT3 in promoting hepatic regeneration, controlled impeding of the CREBZF in hepatocytes may improve the liver regeneration process [18].

The epidermal growth factor (EGF) receptor ligand family and hepatocyte growth factors (HGFs) are important growth factors that are involved in the proliferation step and are responsible for cell-cycle progression [19]. HGF and its receptor-encoding gene, *c-met*, are necessary for liver regeneration and repair [20]. HGF/c-met signaling contributes to the activation of ERK1/2 [21], which induces liver cell reproduction. The EGF receptor ligand family contains EGF, TGF- $\alpha$ , heparin-binding EGF-like growth factor (HBEGF), and amphiregulin (AR). The Brunner's gland is responsible for the production of the EGF, and, therefore, its extirpation decreases liver regeneration. Additionally, the EGF antiserum reduced liver regeneration significantly [22]. Following hepatocyte reproduction, the TGF- $\alpha$  is produced and contributes to the constant proliferation of liver cells [23].

The final stage of regeneration is regulated by cytokines of the TGF- $\beta$  family. TGF- $\beta$  suppresses cell growth and terminates liver regeneration. During the proliferation step, increased expression level of Smad7 by HGF and EGF receptors has been shown to inhibit TGF- $\beta$ , which functions as an antiproliferative factor. At the termination step, however, the TGF- $\beta$  production is increased from kupffer cells and upregulated stellate cells [24,25]. In addition to TGF- $\beta$ , the suppressors of cytokine signaling (SOCS) block hepatocyte regeneration by downregulating the STAT3 and IL-6 signaling pathway. In another study, the mitogen-activated protein kinase kinase 4 (MKK4) function was examined in mouse models, and MKK4 was proven to work as a strong inhibitor of liver regeneration [26]. The absence of liver regeneration terminators can have detrimental consequences due to the hepatocyte hyperproliferation and liver injury.

## 3. HBV Infection and Liver Regeneration

HBV is a major pathogen that chronically infects more than 300 million people globally. Chronic HBV infection is an important cause of death through induction of cirrhosis and hepatocellular carcinoma [1–3]. Chronic inflammation of the liver due to the host immune response to the virus is a major contributor to the pathogenesis of HCC [27,28]. HBV X protein (HBx), a pleiotropic protein encoded by HBV, plays a crucial role in viral liver pathogenesis [29] and HBV replication [30–32]. HBx can also transactivate host genes by binding to many transcription factors (TFs), including AP-1 and AP-2 [33], ATF/CREB [34], C/EBP [35], NF-AT [36], p53 [37,38], HIF-1 $\alpha$  [39], HNF1 [40], SMAD4 [41], STAT3, and NF- $\kappa$ B [42]. A number of studies have shown that HBx is involved in gene activation [43–45], apoptosis [46,47], and cell cycle control [48,49].

Studies have been conducted to understand the mechanism of regeneration by analyzing gene expression occurring in the regenerative liver. For instance, a large-scale gene expression analysis was performed to reveal the expression pattern throughout the course of liver regeneration in mice [50–53].

#### 3.1. HBx Effect on Liver Regeneration by Cell Cycle Regulation

Various studies have reported that HBV infection can impair the liver regeneration [54–60] (Table 1 and Figure 1). Intriguingly, HBx expression has paracrine inhibitory effect on liver cell proliferation in HBx-transgenic mice [54]. High mortality, low liver mass recovery, and impaired liver regeneration were observed in HBx-transgenic mice after partial hepatectomy. HBx protein blocks cell-cycle progression by preventing G1/S transition of the hepatocyte, and it causes both failure of liver functionality and cell death in the regenerating liver of the HBx-transgenic mice [55]. Equally important, HBx is capable of modulating significant cell-cycle regulators, such as p21 and proliferating cell nuclear antigen (PCNA), in order to disrupt normal cell-cycle checkpoints, resulting in premature entry to S phase and impeded liver regeneration [57].

Viral Protein	Model	Injury	Effect	Mechanism	Reference
HBx	HBx-Tg mice	PHx	Inhibition of liver regeneration	Paracrine inhibitory effect	[54]
HBx	HBx-Tg mice	PHx	Failure of liver functionality and cell death	Blocking of G1/S transition	[55]
-	HBV-Tg mice	PHx	Impairment of Liver regeneration	Activated NKT cells	[56]
HBx	HBx-Tg mice	PHx	Normal liver regeneration	Premature cell cycle	[57]
HBx	Hydrodynamic Injection of viral DNA	PHx	Inhibition of liver regeneration	Epigenetic regulation of uPA	[58]
HBx	HBx-Tg mice	PHx	Delayed liver regeneration	IL6 overexpression	[59]
-	HBV-Tg mice	CCL <sub>4</sub>	Delayed liver regeneration	Retention of insulin receptor	[60]

Table 1. Studies on the effect of hepatitis B virus infection on liver regeneration using mouse models.

-, not identified; Tg, transgenic; PHx, partial hepatectomy.



**Figure 1.** Inhibition of the hepatic regenerative pathway by HBV infection. PHx, partial hepatectomy. This illustration was created with BioRender.com.

## 3.2. HBx Effect on Liver Regeneration by Regulating Cellular Factors

In another investigation performed on the HBV-transgenic mice, at the initial steps of the liver regeneration, the natural killer T (NKT) cell proliferation and CD69 marker protein expression at their surface were observed. This was along with elevated levels of the interferon gamma (IFN-) production. The HBx-induced high levels of these factors were proved to be the major inhibitor of the liver generation [56].

The urokinase-type plasminogen activator (uPA) upregulates liver regeneration via activation of HGF in the extracellular matrix scaffold of the hepatocytes [61]. As a consequence, mice that are deficient in uPA or plasminogen are markedly impaired in liver regeneration [61,62]. Previously, we have reported that the HBx protein inhibits liver regeneration via downregulation of uPA expression through epigenetic regulation by way of hypermethylation of its promoter [58].

## 3.3. HBx Effect on Liver Regeneration by Modulating Cell Signaling Pathways 3.3.1. IL-6-STAT3 Signaling Pathway

Despite what is known about the role of IL-6 in stimulating hepatocytes to grow after partial hepatectomy, the actual function of the IL-6/STAT3 pathway in tumorigenesis and liver cancer is elusive. Several studies have reported that HBx-induced IL-6 overexpression is involved in delayed liver regeneration [59]. The HBx-induced higher secretion of IL-6 was associated with an elevation of STAT3 phosphorylation, SOCS3 accumulation, and a decrease in ERK1/2 phosphorylation in the livers of HBx-transgenic mice [59].

#### 3.3.2. Lipid Metabolism Pathway

Cell membrane phospholipids have a critical function in lipid metabolism and liver regeneration [63]. It has been noted that elevated fat levels from adipose tissue at early stages of liver regeneration may facilitate hepatocyte proliferation. Meanwhile, it has been reported that a major regulator of glucose and lipid metabolism named hepatic pyruvate dehydrogenase kinase 4 (PDK4) level was enhanced after two-thirds partial hepatectomy [64].

The effect of HBx on the gene expression profile in the regenerated liver after partial hepatectomy of HBx-transgenic mice has been analyzed [65]. Impaired DNA synthesis and reduction in liver mass in HBx-transgenic mice have been shown to be associated with a weaker expression of genes involved in the cholesterol and nonsterol isoprenoid pathways, as well as serum amyloid A [65]. Moreover, using MALDI imaging MS analysis, we have previously reported that phosphatidylcholine biosynthesis in the regenerating liver is altered by HBV infection, causing a regeneration defect [63].

## 3.3.3. Insulin Receptor Pathway

Finally, insulin receptor (IR)-dependent signaling represents a key factor in controlling liver regeneration [66,67]. Although HBV induces expression of the insulin receptor, liver regeneration is delayed in HBV-transgenic mice due to the intracellular holding of the insulin receptor [60]. Since the glucose is essential for cell growth, the hepatocyte proliferation rate in the liver of these mice is significantly lower than normal and is coupled with elevated fibrosis markers.

## 4. Liver Regeneration and HCC

HCC is the most common primary liver cancer. The chronic hepatitis B virus infection, which is characterized by cycles of cell death and regeneration, is one of the most important causes of HCC globally [68]. Chronic inflammation and cytokines are key determinants in the development of fibrosis and liver cell proliferation. HBx expression has been reported to contribute to liver disease and HCC development by affecting the regeneration of the liver [57], as hepatocarcinogenesis was accelerated in HBx-transgenic mice after partial hepatectomy [69].

Although partial hepatectomy is considered as a potential treatment for HCC [70], the long-term survival of patients with HBV-associated HCC after partial hepatectomy remains challenging. The 5-year recurrence rate after surgery, which is as high as 70%, implies the low survival rate of patients [71–73]. The high de novo recurrence of HBV-related HCC after partial hepatectomy results from induction of the sequential changes of liver-regeneration-related SOCS family proteins, growth factors, and transcription factors, which may promote growth on the precancerous remnant liver [69].

In a study using liver explant from four patients with HBV-associated acute liver failure, Nissim and colleagues reported the upregulation of transcripts related to liver regeneration, including stem/progenitor markers, fibrogenesis molecules, and cell-proliferation and -tumorigenesis-related factors [74]. The authors suggested that the increased gene expression which is associated with extensive fibrogenesis is part of the repair process of damaged liver tissue, although they could not conclusively demonstrate whether the regenerative gene expression profile would have resulted in an actual successful liver regeneration due to the devastating consequences of the disease progression. However, it is important to note that, in the subpopulation of patients with submassive hepatic necrosis, hepatic stem/progenitor gene signature is accompanied with growth/proliferation and tumorigenesis-related gene expression profiles. These results denote the interesting inter-correlation among HBV pathogenesis, liver regeneration, and liver cancer. Regardless, whether non-massive chronic liver damage after HBV infection may induce dysregulated regenerative responses, along with manifestation and/or aggravation of hepatocellular cancer, remains to be determined.

## 4.1. ALR

The regulators of damage response and hepatocyte regeneration have been implicated in the manifestation and pathophysiology of HCC. For example, human augmenter of liver regeneration (ALR), a member of the ALR/Erv1 protein family with FAD-linked sulfhydryl oxidase activity catalyzing disulfide bond formation, is a hepatotropic factor and is involved in the modulation of hepatocytes' proliferation and liver regeneration [75–80]. ALR has been shown to provide antimetastatic potential in HCC [81]. HepG2 cells expressing cytosolic 15 kDa ALR isoform (HepG2-ALR) showed reduced cell motility and epithelialto-mesenchymal transition (EMT), as evidenced by the changes in EMT marker expression. Similarly, the decreased size of the tumor mass was observed in HepG2-ALR tumors grown in nude mice. A negative correlation of angioinvasion and tumor grading against the level of ALR, which seems to be elevated in patients with cirrhosis, cholangiocellular carcinoma (CCC), and HCC [82], has been observed in HCC patients. On the contrary, the deletion of ALR expression in liver-specific fashion exacerbates the development of HCC in mice [83]. Interestingly, ALR expression is higher in hepatoma cells and primary human hepatocytes infected with HBV compared with hepatitis C virus (HCV). Because HBV regulates the expression of ALR through activation of the nuclear factor erythroid 2 (Nrf2), it has been proposed that HBV-induced oxidative stress may be linked to the hepatic regeneration through the modulation of ALR to ensure survival of damaged cells [84]. In this regard, it is noteworthy to see that the serum ALR level of patients with HBV-infected acute-onchronic liver failure was higher in the survival group than in the dead group, suggesting the essential role of ALR in situations where HBV induces pathological changes in liver that make it possible to use ALR as a prognosis marker in such cases [85].

## 4.2. SOCS

SOCS is a family of genes involved in the inhibition of JAK-STAT signaling pathway, which is involved in physiological and pathological regulation of cytokine and growth factor responses, as well as tumorigenesis. Among these, SOCS3 has been implicated to inhibit cytokine activation in the early stages of liver regeneration [86]. Hepatocyte-specific knockout of SOCS3 expression (Socs3 h-KO) induced the augmentation of DNA replication and increase in liver weight after partial hepatectomy, which might be induced by the extended phosphorylation of STAT3 and activation of ERK1/2 after partial hepatectomy. The carcinogenesis was increased in Socs3 h-KO mice, suggesting the essential role of SOCS3 as a tumor suppressor [87,88]. In HBV-infected hepatocytes, cytokine signaling pathways such as the STAT/SOCS-signaling pathway are dysregulated both in vivo and in vitro, and this is correlated with the severity of pathological inflammatory changes in the liver. It has been suggested that HBV-induced interference of STAT/SOCS signaling may serve as key molecular mechanism underlying viral pathogenesis and malignant liver tissue transformation [89]. SOCS3 hypermethylation—hence, the lower expression level of SOCS3—was significantly correlated with a poor clinical outcome of HCC patients [90]. Mechanistically, HBV induces the production of mitochondrial ROS, and the increased ROS can result in Snail-mediated epigenetic silencing of SOCS3 expression. It is proposed that the reduced expression of SOCS3 results in sustained activation of the IL-6/STAT3 pathway, which could be essential to hepatocarcinogenesis [91]. Similarly, a recent meta-analysis work demonstrated that the SOCS3 hypermethylation is associated with HBV-related HCC [92], and this may validate the possibility of SOCS3 expression or methylation status of the SOCS3 gene as a prognosis marker for HCC associated with HBV infection [93].

## 4.3. Cytokines

Several cytokines, including IL-6, TNF- $\alpha$ , and TGF- $\beta$ , are critically involved in the modulation of liver regeneration process and carcinogenesis, as well. After partial hepatectomy, mice lacking type I TNF receptor (TNFR-1) showed massive impairment in DNA synthesis and activation of signaling pathways such as NF- $\kappa$ B, AP-1, and STAT3 [94]. The authors argued that the altered response is mediated by the dysregulation of IL-6 and STAT-3 signaling pathways, particularly due to the reversal of TNFR-1 KO phenotypes by the administration of IL-6 [94]. On the other hand, the excessive TNF signal has been reported to play a detrimental role in the liver regeneration processes. In Timp3 knockout mice, TNF- $\alpha$  converting enzyme activity was increased, thereby providing excessive release of TNF and augmentation of TNF signaling in the liver, which induced hepatitis-like inflammatory reaction and regeneration failure [95]. These phenomena are attributed to higher level of cell death induced by the increased production of TNF [95]. Recently, a concentration-dependent role for TNF- $\alpha$  in the modulation of hepatocyte proliferation and liver regeneration has been suggested. Zhao et al. reported that low TNF- $\alpha$  concentration stimulates hepatocyte proliferation and protects the liver from damage, as evidenced by the increased levels of ALT and AST in the plasma [96]. Liver damage was intensified with increasing concentrations of TNF- $\alpha$ , which might be regulated by a mechanosensitive transcriptional regulator, Yes-associated protein (Yap) activity. In other words, the low TNF- $\alpha$  concentration activated Yap1 nuclear translocation and the proliferation of hepatocytes, while a higher concentration of TNF- $\alpha$  inactivates Yap1, promoting hepatic cell death [96]. The increased plasma level of TNF- $\alpha$  and other cytokines has been reported after HBV infection, and it has been suggested that HBV (HBx) enhanced the TNF- $\alpha$ -induced activation of NF- $\kappa$ B in hepatocytes, resulting in a cooperative increase in gene expression, including those involved in the modulation of metabolic pathways [97]. Dysregulation of immune responses and hepatic proliferation may link to further damage and pathologic conversion of liver diseases from chronic hepatitis to HCC [98]. Chronic HBV infection perturbs immune responses in infected individuals [99] and may induce immune-mediated liver injury accompanied by the elevations of TNF- $\alpha$  and IL-1 $\beta$  levels [97], which also underlie HCC progression due to accelerated hepatocyte turnover rates and the accumulation of deleterious mutations [100].

## 4.4. Growth Factors

The main growth factors involved in the liver regeneration are HGF, EGF, IL-6, and TGF- $\beta$  [101]. Due to the need of hepatocyte proliferation at the early stage of liver regeneration, the anti-proliferative effects of TGF- $\beta$  are generally considered to be dispensable [102]. However, TGF-B plays a crucial role during the later stage by promoting EMT, inflammation, restoration of the original tissue architecture, and cell–cell interactions [103,104]. Therefore, the balance and kinetics of growth factors and TGF- $\beta$  signaling during liver regeneration process should be delicately regulated [105]. Interestingly, TGF modulates uPA/PAI-1 expression, and uPA regulates TGF- $\beta$  and its signaling pathway, suggesting the importance of the delicate interplay among the extracellular matrix, growth factors, uPA/PAI-1, and TGF- $\beta$ . It has been suggested that TGF- $\beta$  plays an essential role in the regulation of pathological consequences of HBV infection and subsequent HCC and cirrhosis of the liver [106]. This might be through the regulation of Th17 and T regulatory lymphocytes development and function by TGF-beta. Recent studies also provided evidence that TGF-β directly modulates hepatocyte function during HBV infection. Although some studies reported the anti-HBV action of TGF- $\beta$  [107], it has also been suggested that HBV infection upregulated the expression of TGF- $\beta$  [108–111], which might be related to the inhibition of immune responses, as well as the development of fibrosis and intra hepatic cholangiocarcinoma [110,111]. The mechanisms whereby TGF- $\beta$  regulates liver pathogenesis during chronic HBV infection includes but is not restricted to the regulation of extracellular matrix system [112], regulation of immune modulators such as Th17 and regulatory T cells (Tregs) [113,114], production of cytokines and chemokines [115], regulation of miRNA signaling [116], and cell proliferation [117].

Another essential regulator of liver regeneration is HGF, a peptide with pleotropic cytokinelike functions. HGF and its target protein c-Met modulate JAK/STAT3, PI3K/Akt/NF- $\kappa$ B, and Ras/Raf signaling pathways in many types of epithelial cells, one of which is the liver. The GHGF/c-Met signaling pathways are critically involved in physiological processes such as cell proliferation, growth, and survival. The role of HGF in liver fibrosis, regeneration, and inflammation, as well as carcinogenesis, has been a focus for many review articles [118,119]. After partial hepatectomy, the level of HGF in the plasma increases 10 to 20 times [120], initiating the delicate process of liver regeneration ending with TGF-beta-induced rebuild up of the extracellular matrix and inhibition of HGF activity [121,122]. In HBV infected patients, the concentration of growth factors, including HGF, is increased [123,124], and this is correlated with the severity of the liver pathology [123]. In general, the increased HGF level is believed to offer survival opportunity with mechanisms involving modulation of the activity of PI3K kinase/Akt, STAT3 and cdc42-Associated Kinase 1 [124,125], as well as indirect survival promotion effects through the regulation of extracellular matrix organization [124]. Aberrant growth factor signal has been associated with the hepatocellular carcinogenesis. One of the suggested mechanisms is the overactivation of c-Met and related signals, such as the Akt pathways, resulting in increased proliferation of hepatic tissue, enhanced angiogenesis, and modulation of autophagy [126–128]. A recent review highlighted the importance of balance between liver regeneration signal and carcinogenesis [129]. It is reasonable to assume that HBV infection might modulate both regeneration and carcinogenesis processes through the regulation of growth factors and related signaling pathways.

One of the key factors regulating liver regeneration and carcinogenesis is extracellular matrix proteins and factors regulating the attachment and liberation of the interacting molecules, such as uPA. It has been suggested that uPA plays a crucial role in the processing and release of HGF from the hepatic extracellular matrix stores [130], and it has been suggested that the modulatory role of uPA and the plasminogen system on liver regeneration is independent of the proliferation process [131]. Signals from immune modulators and danger or pathogen sensors such as cytokines and TLRs have been implicated in the modulation of uPA expression [130]. We have previously reported that HBx can modulate the expression of uPA by regulating methylation status on its promoter region via the mechanism involving the recruitment of DNA methyltransferase (DNMT) 3A2 [58], which underlies the observed liver regeneration inhibitory effects of HBV infection. Considering the fact that uPA plays an important role in tumor invasion and progression [132], regulating uPA level and activity via HBV infection has a significant impact on the liver regeneration process and hepatocellular carcinogenesis.

## 4.5. HSPC

In the liver regeneration process, not only the mature hepatocyte proliferation, but also the cell division and differentiation of hepatic stem/progenitor cell (HSPC), plays an important role depending on the context of liver damage. In general, HSPC activation is more important in the situation where the liver loss is massive and hepatic proliferation is minimal by chronic liver diseases [133]. Interestingly, a genetic analysis of HBV-infected patients showed altered expression profiles of genes and pathways related to HSPC modulation, along with overt cell proliferation and tumorigenesis signals [74]. In a HBsAg HBV-transgenic mouse model, activation of the hepatic stem cells' compartment was observed during hepatocarcinogenesis, suggesting the essential role of modulating HSPC activation by chronic HBV infection for the carcinogenesis [134]. The critical time window and extent of HBV infection for the modulation of mature hepatocyte proliferation and HSPC activation need to be determined further in the future and may provide insights into the correlation among HBV, liver regeneration, and carcinogenesis, as well as the means to treat the pathological outcomes.

## 5. Conclusions and Perspectives

HBV is one of the important and relevant risk factors for HCC. Since the liver has a strong regenerative ability, surgical resection of liver cancer or transplantation of a healthy liver is one of the treatment methods for HBV-induced liver cancer. However, the long-term survival of patients with HBV-associated HCC after partial hepatectomy is still limited. It is also not yet known how HBV infection affects liver resection or regeneration of transplanted livers. What is known is that HBV causes the disturbance of liver regeneration and promotes hepatocarcinogenesis by regulating the signal proteins involved in the liver regeneration. Although there is no direct study on the induction of liver cancer due to abnormal liver regeneration caused by HBV, the related studies strongly suggest the possibility of a relationship between the two events (Figure 2). Therefore, for the effective treatment of HBV-induced liver cancer, it is very important to study the mechanisms involved in the modulation of liver regeneration by HBV infection. A greater understanding of the regenerative process has significant benefits in treating liver failure and is expected to better explain the pathogenesis of liver cancer.



**Figure 2.** Schematic diagram of HBV-related liver regeneration and HCC. The illustration represents the increased incidence of liver cancer due to the abnormal liver regeneration caused by HBV infection. This illustration was created with BioRender.com (accessed on 18 June 2022).

**Author Contributions:** Conceptualization, E.-S.P. and K.-H.K.; writing—original draft preparation, E.-S.P. and M.D.; writing—review and editing, E.-S.P., B.B.P., A.R.L. and K.-H.K.; funding acquisition, E.-S.P. and K.-H.K. All authors have read and agreed to the published version of the manuscript.

**Funding:** This study was supported by the National Research Foundation of Korea (NRF) grants funded by the Korea government (NRF-2019R1A2B5B01069635 and 2022R1A2C2002809 (E.S.P.); NRF-2020R1A2C3010511 and 2021M3A9I2080488 (K.H.K.)). This research was also supported by a grant of the Korea Health Technology R&D Project through the KHIDI, funded by the Ministry of Health and Welfare, Republic of Korea (grant number: HV22C0000).

Conflicts of Interest: The authors declare no conflict of interest.

## References

- 1. Feitelson, M.A. Hepatitis B virus in hepatocarcinogenesis. J. Cell Physiol. 1999, 181, 188–202. [CrossRef]
- 2. Brechot, C.; Gozuacik, D.; Murakami, Y.; Paterlini-Brechot, P. Molecular bases for the development of hepatitis B virus (HBV)related hepatocellular carcinoma (HCC). *Semin. Cancer Biol.* **2000**, *10*, 211–231. [CrossRef]
- 3. Seeger, C.; Mason, W.S. Hepatitis B virus biology. Microbiol. Mol. Biol. Rev. 2000, 64, 51–68. [CrossRef]
- 4. Higgins, G.M. Experimental pathology of the liver, I: Restoration of the liver of the white rat following partial surgical removal. *Arch. Pathol.* **1931**, *12*, 186–202.
- 5. Fausto, N.; Laird, A.D.; Webber, E.M. Liver regeneration. 2. Role of growth factors and cytokines in hepatic regeneration. *FASEB J.* **1995**, *9*, 1527–1536. [CrossRef]
- 6. Michalopoulos, G.K.; DeFrances, M.C. Liver regeneration. *Science* **1997**, 276, 60–66. [CrossRef]
- 7. Fausto, N.; Campbell, J.S.; Riehle, K.J. Liver regeneration. *Hepatology* 2006, 43, S45–S53. [CrossRef]
- 8. Michalopoulos, G.K. Liver regeneration: Molecular mechanisms of growth control. FASEB J. 1990, 4, 176–187. [CrossRef]
- 9. Taub, R. Liver regeneration: From myth to mechanism. Nat. Rev. Mol. Cell Biol. 2004, 5, 836–847. [CrossRef]
- 10. Michalopoulos, G.K. Liver regeneration. J. Cell Physiol. 2007, 213, 286–300. [CrossRef]
- 11. Fausto, N.; Campbell, J.S.; Riehle, K.J. Liver regeneration. J. Hepatol. 2012, 57, 692–694. [CrossRef]
- 12. Fausto, N. Liver regeneration. J. Hepatol. 2000, 32, 19–31. [CrossRef]
- 13. Fausto, N.; Riehle, K.J. Mechanisms of liver regeneration and their clinical implications. *J. Hepatobiliary Pancreat. Surg.* 2005, 12, 181–189. [CrossRef]
- 14. FitzGerald, M.J.; Webber, E.M.; Donovan, J.R.; Fausto, N. Rapid DNA binding by nuclear factor kappa B in hepatocytes at the start of liver regeneration. *Cell Growth Differ*. **1995**, *6*, 417–427.
- 15. Cressman, D.E.; Diamond, R.H.; Taub, R. Rapid activation of the Stat3 transcription complex in liver regeneration. *Hepatology* **1995**, *21*, 1443–1449. [CrossRef]
- 16. Akerman, P.; Cote, P.; Yang, S.Q.; McClain, C.; Nelson, S.; Bagby, G.J.; Diehl, A.M. Antibodies to tumor necrosis factor-alpha inhibit liver regeneration after partial hepatectomy. *Am. J. Physiol.* **1992**, *263*, G579–G585. [CrossRef]
- 17. Cressman, D.E.; Greenbaum, L.E.; DeAngelis, R.A.; Ciliberto, G.; Furth, E.E.; Poli, V.; Taub, R. Liver failure and defective hepatocyte regeneration in interleukin-6-deficient mice. *Science* **1996**, 274, 1379–1383. [CrossRef]
- 18. Hu, Z.; Han, Y.; Liu, Y.; Zhao, Z.; Ma, F.; Cui, A.; Zhang, F.; Liu, Z.; Xue, Y.; Bai, J.; et al. CREBZF as a Key Regulator of STAT3 Pathway in the Control of Liver Regeneration in Mice. *Hepatology* **2020**, *71*, 1421–1436. [CrossRef]
- 19. Michalopoulos, G.K.; Khan, Z. Liver regeneration, growth factors, and amphiregulin. *Gastroenterology* **2005**, *128*, 503–506. [CrossRef]
- 20. Huh, C.G.; Factor, V.M.; Sánchez, A.; Uchida, K.; Conner, E.A.; Thorgeirsson, S.S. Hepatocyte growth factor/c-met signaling pathway is required for efficient liver regeneration and repair. *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 4477–4482. [CrossRef]
- Borowiak, M.; Garratt, A.N.; Wüstefeld, T.; Strehle, M.; Trautwein, C.; Birchmeier, C. Met provides essential signals for liver regeneration. *Proc. Natl. Acad. Sci. USA* 2004, 101, 10608–10613. [CrossRef]
- Skov Olsen, P.; Boesby, S.; Kirkegaard, P.; Therkelsen, K.; Almdal, T.; Poulsen, S.S.; Nexø, E. Influence of epidermal growth factor on liver regeneration after partial hepatectomy in rats. *Hepatology* 1988, 8, 992–996. [CrossRef]
- 23. Webber, E.M.; Wu, J.C.; Wang, L.; Merlino, G.; Fausto, N. Overexpression of transforming growth factor-alpha causes liver enlargement and increased hepatocyte proliferation in transgenic mice. *Am. J. Pathol.* **1994**, *145*, 398–408.
- 24. Derynck, R.; Zhang, Y.E. Smad-dependent and Smad-independent pathways in TGF-beta family signalling. *Nature* **2003**, 425, 577–584. [CrossRef]
- Michalopoulos, G.K.; Bowen, W.C.; Mulè, K.; Stolz, D.B. Histological organization in hepatocyte organoid cultures. *Am. J. Pathol.* 2001, 159, 1877–1887. [CrossRef]
- 26. Wuestefeld, T.; Pesic, M.; Rudalska, R.; Dauch, D.; Longerich, T.; Kang, T.W.; Yevsa, T.; Heinzmann, F.; Hoenicke, L.; Hohmeyer, A.; et al. A Direct in vivo RNAi screen identifies MKK4 as a key regulator of liver regeneration. *Cell* **2013**, *153*, 389–401. [CrossRef]
- 27. Chisari, F.V. Rous-Whipple Award Lecture. Viruses, immunity, and cancer: Lessons from hepatitis B. *Am. J. Pathol.* **2000**, *156*, 1117–1132. [CrossRef]
- Guidotti, L.G.; Rochford, R.; Chung, J.; Shapiro, M.; Purcell, R.; Chisari, F.V. Viral clearance without destruction of infected cells during acute HBV infection. *Science* 1999, 284, 825–829. [CrossRef]
- 29. Ali, A.; Abdel-Hafiz, H.; Suhail, M.; Al-Mars, A.; Zakaria, M.K.; Fatima, K.; Ahmad, S.; Azhar, E.; Chaudhary, A.; Qadri, I. Hepatitis B virus, HBx mutants and their role in hepatocellular carcinoma. *World J. Gastroenterol.* **2014**, *20*, 10238–10248. [CrossRef]
- 30. Xu, Z.; Yen, T.S.; Wu, L.; Madden, C.R.; Tan, W.; Slagle, B.L.; Ou, J.H. Enhancement of hepatitis B virus replication by its X protein in transgenic mice. *J. Virol.* 2002, *76*, 2579–2584. [CrossRef]
- Tang, H.; Delgermaa, L.; Huang, F.; Oishi, N.; Liu, L.; He, F.; Zhao, L.; Murakami, S. The transcriptional transactivation function of HBx protein is important for its augmentation role in hepatitis B virus replication. *J. Virol.* 2005, 79, 5548–5556. [CrossRef] [PubMed]
- 32. Keasler, V.V.; Hodgson, A.J.; Madden, C.R.; Slagle, B.L. Enhancement of hepatitis B virus replication by the regulatory X protein in vitro and in vivo. *J. Virol.* 2007, *81*, 2656–2662. [CrossRef]

- 33. Seto, E.; Mitchell, P.J.; Yen, T.S. Transactivation by the hepatitis B virus X protein depends on AP-2 and other transcription factors. *Nature* **1990**, *344*, 72–74. [CrossRef] [PubMed]
- Maguire, H.F.; Hoeffler, J.P.; Siddiqui, A. HBV X protein alters the DNA binding specificity of CREB and ATF-2 by protein-protein interactions. *Science* 1991, 252, 842–844. [CrossRef]
- 35. Unger, T.; Shaul, Y. The X protein of the hepatitis B virus acts as a transcription factor when targeted to its responsive element. *EMBO J.* **1990**, *9*, 1889–1895. [CrossRef] [PubMed]
- 36. Lara-Pezzi, E.; Armesilla, A.L.; Majano, P.L.; Redondo, J.M.; López-Cabrera, M. The hepatitis B virus X protein activates nuclear factor of activated T cells (NF-AT) by a cyclosporin A-sensitive pathway. *EMBO J.* **1998**, *17*, 7066–7077. [CrossRef] [PubMed]
- Wang, X.W.; Forrester, K.; Yeh, H.; Feitelson, M.A.; Gu, J.R.; Harris, C.C. Hepatitis B virus X protein inhibits p53 sequence-specific DNA binding, transcriptional activity, and association with transcription factor ERCC3. *Proc. Natl. Acad. Sci. USA* 1994, 91, 2230–2234. [CrossRef]
- Truant, R.; Antunovic, J.; Greenblatt, J.; Prives, C.; Cromlish, J.A. Direct interaction of the hepatitis B virus HBx protein with p53 leads to inhibition by HBx of p53 response element-directed transactivation. *J. Virol.* 1995, 69, 1851–1859. [CrossRef]
- Yoo, Y.G.; Na, T.Y.; Seo, H.W.; Seong, J.K.; Park, C.K.; Shin, Y.K.; Lee, M.O. Hepatitis B virus X protein induces the expression of MTA1 and HDAC1, which enhances hypoxia signaling in hepatocellular carcinoma cells. *Oncogene* 2008, 27, 3405–3413. [CrossRef]
- Li, J.; Xu, Z.; Zheng, Y.; Johnson, D.L.; Ou, J.H. Regulation of hepatocyte nuclear factor 1 activity by wild-type and mutant hepatitis B virus X proteins. J. Virol. 2002, 76, 5875–5881. [CrossRef]
- Lee, D.K.; Park, S.H.; Yi, Y.; Choi, S.G.; Lee, C.; Parks, W.T.; Cho, H.; de Caestecker, M.P.; Shaul, Y.; Roberts, A.B.; et al. The hepatitis B virus encoded oncoprotein pX amplifies TGF-beta family signaling through direct interaction with Smad4: Potential mechanism of hepatitis B virus-induced liver fibrosis. *Genes Dev.* 2001, 15, 455–466. [CrossRef] [PubMed]
- 42. Waris, G.; Huh, K.W.; Siddiqui, A. Mitochondrially associated hepatitis B virus X protein constitutively activates transcription factors STAT-3 and NF-kappa B via oxidative stress. *Mol. Cell Biol.* 2001, *21*, 7721–7730. [CrossRef] [PubMed]
- 43. Lucito, R.; Schneider, R.J. Hepatitis B virus X protein activates transcription factor NF-kappa B without a requirement for protein kinase C. J. Virol. **1992**, *66*, 983–991. [CrossRef]
- 44. Su, F.; Theodosis, C.N.; Schneider, R.J. Role of NF-kappaB and myc proteins in apoptosis induced by hepatitis B virus HBx protein. *J. Virol.* **2001**, *75*, 215–225. [CrossRef]
- 45. Tanaka, Y.; Kanai, F.; Ichimura, T.; Tateishi, K.; Asaoka, Y.; Guleng, B.; Jazag, A.; Ohta, M.; Imamura, J.; Ikenoue, T.; et al. The hepatitis B virus X protein enhances AP-1 activation through interaction with Jab1. *Oncogene* **2006**, *25*, 633–642. [CrossRef]
- 46. Chirillo, P.; Pagano, S.; Natoli, G.; Puri, P.L.; Burgio, V.L.; Balsano, C.; Levrero, M. The hepatitis B virus X gene induces p53-mediated programmed cell death. *Proc. Natl. Acad. Sci. USA* **1997**, *94*, 8162–8167. [CrossRef] [PubMed]
- Kim, K.H.; Seong, B.L. Pro-apoptotic function of HBV X protein is mediated by interaction with c-FLIP and enhancement of death-inducing signal. *EMBO J.* 2003, 22, 2104–2116. [CrossRef] [PubMed]
- 48. Koike, K.; Moriya, K.; Yotsuyanagi, H.; Iino, S.; Kurokawa, K. Induction of cell cycle progression by hepatitis B virus HBx gene expression in quiescent mouse fibroblasts. *J. Clin. Investig.* **1994**, *94*, 44–49. [CrossRef] [PubMed]
- Benn, J.; Schneider, R.J. Hepatitis B virus HBx protein deregulates cell cycle checkpoint controls. *Proc. Natl. Acad. Sci. USA* 1995, 92, 11215–11219. [CrossRef] [PubMed]
- Arai, M.; Yokosuka, O.; Chiba, T.; Imazeki, F.; Kato, M.; Hashida, J.; Ueda, Y.; Sugano, S.; Hashimoto, K.; Saisho, H.; et al. Gene expression profiling reveals the mechanism and pathophysiology of mouse liver regeneration. *J. Biol. Chem.* 2003, 278, 29813–29818. [CrossRef]
- 51. Fukuhara, Y.; Hirasawa, A.; Li, X.K.; Kawasaki, M.; Fujino, M.; Funeshima, N.; Katsuma, S.; Shiojima, S.; Yamada, M.; Okuyama, T.; et al. Gene expression profile in the regenerating rat liver after partial hepatectomy. *J. Hepatol.* **2003**, *38*, 784–792. [CrossRef]
- Togo, S.; Makino, H.; Kobayashi, T.; Morita, T.; Shimizu, T.; Kubota, T.; Ichikawa, Y.; Ishikawa, T.; Okazaki, Y.; Hayashizaki, Y.; et al. Mechanism of liver regeneration after partial hepatectomy using mouse cDNA microarray. *J. Hepatol.* 2004, 40, 464–471. [CrossRef] [PubMed]
- 53. Su, A.I.; Guidotti, L.G.; Pezacki, J.P.; Chisari, F.V.; Schultz, P.G. Gene expression during the priming phase of liver regeneration after partial hepatectomy in mice. *Proc. Natl. Acad. Sci. USA* **2002**, *99*, 11181–11186. [CrossRef] [PubMed]
- 54. Tralhao, J.G.; Roudier, J.; Morosan, S.; Giannini, C.; Tu, H.; Goulenok, C.; Carnot, F.; Zavala, F.; Joulin, V.; Kremsdorf, D.; et al. Paracrine in vivo inhibitory effects of hepatitis B virus X protein (HBx) on liver cell proliferation: An alternative mechanism of HBx-related pathogenesis. *Proc. Natl. Acad. Sci. USA* 2002, *99*, 6991–6996. [CrossRef] [PubMed]
- Wu, B.K.; Li, C.C.; Chen, H.J.; Chang, J.L.; Jeng, K.S.; Chou, C.K.; Hsu, M.T.; Tsai, T.F. Blocking of G1/S transition and cell death in the regenerating liver of Hepatitis B virus X protein transgenic mice. *Biochem. Biophys. Res. Commun.* 2006, 340, 916–928. [CrossRef] [PubMed]
- Dong, Z.; Zhang, J.; Sun, R.; Wei, H.; Tian, Z. Impairment of liver regeneration correlates with activated hepatic NKT cells in HBV transgenic mice. *Hepatology* 2007, 45, 1400–1412. [CrossRef]
- 57. Hodgson, A.J.; Keasler, V.V.; Slagle, B.L. Premature cell cycle entry induced by hepatitis B virus regulatory HBx protein during compensatory liver regeneration. *Cancer Res.* 2008, *68*, 10341–10348. [CrossRef]
- Park, E.S.; Park, Y.K.; Shin, C.Y.; Park, S.H.; Ahn, S.H.; Kim, D.H.; Lim, K.H.; Kwon, S.Y.; Kim, K.P.; Yang, S.I.; et al. Hepatitis B virus inhibits liver regeneration via epigenetic regulation of urokinase-type plasminogen activator. *Hepatology* 2013, 58, 762–776. [CrossRef]

- Quétier, I.; Brezillon, N.; Duriez, M.; Massinet, H.; Giang, E.; Ahodantin, J.; Lamant, C.; Brunelle, M.N.; Soussan, P.; Kremsdorf, D. Hepatitis B virus HBx protein impairs liver regeneration through enhanced expression of IL-6 in transgenic mice. *J. Hepatol.* 2013, 59, 285–291. [CrossRef]
- 60. Barthel, S.R.; Medvedev, R.; Heinrich, T.; Büchner, S.M.; Kettern, N.; Hildt, E. Hepatitis B virus inhibits insulin receptor signaling and impairs liver regeneration via intracellular retention of the insulin receptor. *Cell Mol. Life Sci.* 2016, 73, 4121–4140. [CrossRef]
- 61. Shimizu, M.; Hara, A.; Okuno, M.; Matsuno, H.; Okada, K.; Ueshima, S.; Matsuo, O.; Niwa, M.; Akita, K.; Yamada, Y.; et al. Mechanism of retarded liver regeneration in plasminogen activator-deficient mice: Impaired activation of hepatocyte growth factor after Fas-mediated massive hepatic apoptosis. *Hepatology* **2001**, *33*, 569–576. [CrossRef] [PubMed]
- 62. Roselli, H.T.; Su, M.; Washington, K.; Kerins, D.M.; Vaughan, D.E.; Russell, W.E. Liver regeneration is transiently impaired in urokinase-deficient mice. *Am. J. Physiol.* **1998**, 275, G1472–G1479. [CrossRef] [PubMed]
- 63. Park, E.S.; Lee, J.H.; Hong, J.H.; Park, Y.K.; Lee, J.W.; Lee, W.J.; Lee, J.W.; Kim, K.P.; Kim, K.H. Phosphatidylcholine alteration identified using MALDI imaging MS in HBV-infected mouse livers and virus-mediated regeneration defects. *PLoS ONE* **2014**, *9*, e103955. [CrossRef] [PubMed]
- 64. Zhao, Y.; Tran, M.; Wang, L.; Shin, D.J.; Wu, J. PDK4-Deficiency Reprograms Intrahepatic Glucose and Lipid Metabolism to Facilitate Liver Regeneration in Mice. *Hepatol. Commun.* **2020**, *4*, 504–517. [CrossRef] [PubMed]
- Sidorkiewicz, M.; Jais, J.P.; Tralhao, G.; Morosan, S.; Giannini, C.; Brezillon, N.; Soussan, P.; Delpuech, O.; Kremsdorf, D. Gene modulation associated with inhibition of liver regeneration in hepatitis B virus X transgenic mice. *World J. Gastroenterol.* 2008, 14, 574–581. [CrossRef]
- 66. Beyer, T.A.; Werner, S. The cytoprotective Nrf2 transcription factor controls insulin receptor signaling in the regenerating liver. *Cell Cycle* **2008**, *7*, 874–878. [CrossRef]
- 67. Beyer, T.A.; Xu, W.; Teupser, D.; auf dem Keller, U.; Bugnon, P.; Hildt, E.; Thiery, J.; Kan, Y.W.; Werner, S. Impaired liver regeneration in Nrf2 knockout mice: Role of ROS-mediated insulin/IGF-1 resistance. *EMBO J.* **2008**, 27, 212–223. [CrossRef]
- 68. Chemin, I.; Zoulim, F. Hepatitis B virus induced hepatocellular carcinoma. *Cancer Lett.* 2009, 286, 52–59. [CrossRef]
- 69. Teng, C.F.; Chang, H.Y.; Tsai, H.W.; Hsieh, W.C.; Kuo, Y.H.; Su, I.J.; Lin, Y.J. Liver regeneration accelerates hepatitis B virus-related tumorigenesis of hepatocellular carcinoma. *Mol. Oncol.* **2018**, *12*, 1175–1187. [CrossRef]
- 70. Kishi, Y.; Hasegawa, K.; Sugawara, Y.; Kokudo, N. Hepatocellular carcinoma: Current management and future developmentimproved outcomes with surgical resection. *Int. J. Hepatol.* **2011**, 2011, 728103. [CrossRef]
- Grazi, G.L.; Ercolani, G.; Pierangeli, F.; Del Gaudio, M.; Cescon, M.; Cavallari, A.; Mazziotti, A. Improved results of liver resection for hepatocellular carcinoma on cirrhosis give the procedure added value. *Ann. Surg.* 2001, 234, 71–78. [CrossRef] [PubMed]
- 72. Nakashima, Y.; Nakashima, O.; Tanaka, M.; Okuda, K.; Nakashima, M.; Kojiro, M. Portal vein invasion and intrahepatic micrometastasis in small hepatocellular carcinoma by gross type. *Hepatol. Res.* 2003, *26*, 142–147. [CrossRef]
- 73. Tang, Z.Y.; Ye, S.L.; Liu, Y.K.; Qin, L.X.; Sun, H.C.; Ye, Q.H.; Wang, L.; Zhou, J.; Qiu, S.J.; Li, Y.; et al. A decade's studies on metastasis of hepatocellular carcinoma. *J. Cancer Res. Clin. Oncol.* **2004**, *130*, 187–196. [CrossRef] [PubMed]
- 74. Nissim, O.; Melis, M.; Diaz, G.; Kleiner, D.E.; Tice, A.; Fantola, G.; Zamboni, F.; Mishra, L.; Farci, P. Liver regeneration signature in hepatitis B virus (HBV)-associated acute liver failure identified by gene expression profiling. *PLoS ONE* 2012, 7, e49611. [CrossRef] [PubMed]
- Dayoub, R.; Thasler, W.E.; Bosserhoff, A.K.; Singer, T.; Jauch, K.W.; Schlitt, H.J.; Weiss, T.S. Regulation of polyamine synthesis in human hepatocytes by hepatotrophic factor augmenter of liver regeneration. *Biochem. Biophys. Res. Commun.* 2006, 345, 181–187. [CrossRef] [PubMed]
- 76. Thasler, W.E.; Dayoub, R.; Mühlbauer, M.; Hellerbrand, C.; Singer, T.; Gräbe, A.; Jauch, K.W.; Schlitt, H.J.; Weiss, T.S. Repression of cytochrome P450 activity in human hepatocytes in vitro by a novel hepatotrophic factor, augmenter of liver regeneration. *J. Pharmacol. Exp. Ther.* 2006, 316, 822–829. [CrossRef]
- 77. Pawlowski, R.; Jura, J. ALR and liver regeneration. Mol. Cell Biochem. 2006, 288, 159–169. [CrossRef]
- Gatzidou, E.; Kouraklis, G.; Theocharis, S. Insights on augmenter of liver regeneration cloning and function. World J. Gastroenterol. 2006, 12, 4951–4958. [CrossRef]
- Senkevich, T.G.; White, C.L.; Koonin, E.V.; Moss, B. A viral member of the ERV1/ALR protein family participates in a cytoplasmic pathway of disulfide bond formation. *Proc. Natl. Acad. Sci. USA* 2000, 97, 12068–12073. [CrossRef]
- Senkevich, T.G.; White, C.L.; Koonin, E.V.; Moss, B. Complete pathway for protein disulfide bond formation encoded by poxviruses. *Proc. Natl. Acad. Sci. USA* 2002, 99, 6667–6672. [CrossRef]
- 81. Dayoub, R.; Wagner, H.; Bataille, F.; Stöltzing, O.; Spruss, T.; Buechler, C.; Schlitt, H.J.; Weiss, T.S. Liver regeneration associated protein (ALR) exhibits antimetastatic potential in hepatocellular carcinoma. *Mol. Med.* **2011**, *17*, 221–228. [CrossRef]
- Yu, H.Y.; Zhu, M.H.; Xiang, D.R.; Li, J.; Sheng, J.F. High expression of 23 kDa protein of augmenter of liver regeneration (ALR) in human hepatocellular carcinoma. *Onco Targets Ther.* 2014, 7, 887–893. [CrossRef] [PubMed]
- Gandhi, C.R.; Chaillet, J.R.; Nalesnik, M.A.; Kumar, S.; Dangi, A.; Demetris, A.J.; Ferrell, R.; Wu, T.; Divanovic, S.; Stankeiwicz, T.; et al. Liver-specific deletion of augmenter of liver regeneration accelerates development of steatohepatitis and hepatocellular carcinoma in mice. *Gastroenterology* 2015, 148, 379–391.e374. [CrossRef] [PubMed]
- Dayoub, R.; Vogel, A.; Schuett, J.; Lupke, M.; Spieker, S.M.; Kettern, N.; Hildt, E.; Melter, M.; Weiss, T.S. Nrf2 activates augmenter of liver regeneration (ALR) via antioxidant response element and links oxidative stress to liver regeneration. *Mol. Med.* 2013, 19, 237–244. [CrossRef]

- Hongbo, S.; Yu, C.; Ming, K.; Honglin, S.; Ping, H.Y.; Ping, D.Z. Augmenter of liver regeneration may be a candidate for prognosis of HBV related acute-on-chronic liver failure as a regenerative marker. *Hepatogastroenterology* 2012, *59*, 1933–1938. [CrossRef] [PubMed]
- 86. Campbell, J.S.; Prichard, L.; Schaper, F.; Schmitz, J.; Stephenson-Famy, A.; Rosenfeld, M.E.; Argast, G.M.; Heinrich, P.C.; Fausto, N. Expression of suppressors of cytokine signaling during liver regeneration. *J. Clin. Investig.* **2001**, *107*, 1285–1292. [CrossRef]
- 87. Riehle, K.J.; Campbell, J.S.; McMahan, R.S.; Johnson, M.M.; Beyer, R.P.; Bammler, T.K.; Fausto, N. Regulation of liver regeneration and hepatocarcinogenesis by suppressor of cytokine signaling 3. *J. Exp. Med.* **2008**, 205, 91–103. [CrossRef] [PubMed]
- Liu, Z.K.; Li, C.; Zhang, R.Y.; Wei, D.; Shang, Y.K.; Yong, Y.L.; Kong, L.M.; Zheng, N.S.; Liu, K.; Lu, M.; et al. EYA2 suppresses the progression of hepatocellular carcinoma via SOCS3-mediated blockade of JAK/STAT signaling. *Mol. Cancer* 2021, 20, 79. [CrossRef]
- 89. Koeberlein, B.; zur Hausen, A.; Bektas, N.; Zentgraf, H.; Chin, R.; Nguyen, L.T.; Kandolf, R.; Torresi, J.; Bock, C.T. Hepatitis B virus overexpresses suppressor of cytokine signaling-3 (SOCS3) thereby contributing to severity of inflammation in the liver. *Virus Res.* **2010**, *148*, 51–59. [CrossRef]
- Zhang, X.; You, Q.; Zhang, X.; Chen, X. SOCS3 Methylation Predicts a Poor Prognosis in HBV Infection-Related Hepatocellular Carcinoma. Int. J. Mol. Sci. 2015, 16, 22662–22675. [CrossRef]
- Yuan, K.; Lei, Y.; Chen, H.N.; Chen, Y.; Zhang, T.; Li, K.; Xie, N.; Wang, K.; Feng, X.; Pu, Q.; et al. HBV-induced ROS accumulation promotes hepatocarcinogenesis through Snail-mediated epigenetic silencing of SOCS3. *Cell Death Differ.* 2016, 23, 616–627. [CrossRef] [PubMed]
- 92. Zheng, H.; Yan, Y.; Cheng, J.; Yu, S.; Wang, Y. Association between SOCS3 hypermethylation and HBV-related hepatocellular carcinoma and effect of sex and age: A meta-analysis. *Medicine* **2021**, *100*, e27604. [CrossRef] [PubMed]
- Hoan, N.X.; Van Tong, H.; Giang, D.P.; Cuong, B.K.; Toan, N.L.; Wedemeyer, H.; Bock, C.T.; Kremsner, P.G.; Song, L.H.; Velavan, T.P. SOCS3 genetic variants and promoter hypermethylation in patients with chronic hepatitis B. *Oncotarget* 2017, *8*, 17127–17139. [CrossRef] [PubMed]
- 94. Yamada, Y.; Kirillova, I.; Peschon, J.J.; Fausto, N. Initiation of liver growth by tumor necrosis factor: Deficient liver regeneration in mice lacking type I tumor necrosis factor receptor. *Proc. Natl. Acad. Sci. USA* **1997**, *94*, 1441–1446. [CrossRef]
- 95. Mohammed, F.F.; Smookler, D.S.; Taylor, S.E.; Fingleton, B.; Kassiri, Z.; Sanchez, O.H.; English, J.L.; Matrisian, L.M.; Au, B.; Yeh, W.C.; et al. Abnormal TNF activity in Timp3-/- mice leads to chronic hepatic inflammation and failure of liver regeneration. *Nat. Genet.* 2004, *36*, 969–977. [CrossRef]
- 96. Zhao, S.; Jiang, J.; Jing, Y.; Liu, W.; Yang, X.; Hou, X.; Gao, L.; Wei, L. The concentration of tumor necrosis factor-α determines its protective or damaging effect on liver injury by regulating Yap activity. *Cell Death Dis.* **2020**, *11*, 70. [CrossRef]
- 97. Wu, S.; Kanda, T.; Nakamoto, S.; Jiang, X.; Nakamura, M.; Sasaki, R.; Haga, Y.; Shirasawa, H.; Yokosuka, O. Cooperative effects of hepatitis B virus and TNF may play important roles in the activation of metabolic pathways through the activation of NF-κB. *Int. J. Mol. Med.* **2016**, *38*, 475–481. [CrossRef]
- 98. Kanda, T.; Goto, T.; Hirotsu, Y.; Moriyama, M.; Omata, M. Molecular Mechanisms Driving Progression of Liver Cirrhosis towards Hepatocellular Carcinoma in Chronic Hepatitis B and C Infections: A Review. *Int. J. Mol. Sci.* **2019**, *20*, 1358. [CrossRef]
- 99. Kennedy, P.T.F.; Litwin, S.; Dolman, G.E.; Bertoletti, A.; Mason, W.S. Immune Tolerant Chronic Hepatitis B: The Unrecognized Risks. *Viruses* 2017, 9, 96. [CrossRef]
- Alqahtani, A.; Khan, Z.; Alloghbi, A.; Said Ahmed, T.S.; Ashraf, M.; Hammouda, D.M. Hepatocellular Carcinoma: Molecular Mechanisms and Targeted Therapies. *Medicina* 2019, 55, 526. [CrossRef]
- Böhm, F.; Köhler, U.A.; Speicher, T.; Werner, S. Regulation of liver regeneration by growth factors and cytokines. *EMBO Mol. Med.* 2010, 2, 294–305. [CrossRef] [PubMed]
- 102. Romero-Gallo, J.; Sozmen, E.G.; Chytil, A.; Russell, W.E.; Whitehead, R.; Parks, W.T.; Holdren, M.S.; Her, M.F.; Gautam, S.; Magnuson, M.; et al. Inactivation of TGF-beta signaling in hepatocytes results in an increased proliferative response after partial hepatectomy. *Oncogene* 2005, 24, 3028–3041. [CrossRef]
- Streuli, C.H.; Schmidhauser, C.; Kobrin, M.; Bissell, M.J.; Derynck, R. Extracellular matrix regulates expression of the TGF-beta 1 gene. J. Cell Biol. 1993, 120, 253–260. [CrossRef] [PubMed]
- Margadant, C.; Sonnenberg, A. Integrin-TGF-beta crosstalk in fibrosis, cancer and wound healing. *EMBO Rep.* 2010, 11, 97–105. [CrossRef] [PubMed]
- 105. Karkampouna, S.; Ten Dijke, P.; Dooley, S.; Julio, M.K. TGFβ signaling in liver regeneration. *Curr. Pharm. Des.* 2012, 18, 4103–4113. [CrossRef] [PubMed]
- 106. Karimi-Googheri, M.; Daneshvar, H.; Nosratabadi, R.; Zare-Bidaki, M.; Hassanshahi, G.; Ebrahim, M.; Arababadi, M.K.; Kennedy, D. Important roles played by TGF-β in hepatitis B infection. J. Med. Virol. 2014, 86, 102–108. [CrossRef] [PubMed]
- 107. Park, S.O.; Kumar, M.; Gupta, S. TGF-β and iron differently alter HBV replication in human hepatocytes through TGF-β/BMP signaling and cellular microRNA expression. *PLoS ONE* **2012**, *7*, e39276. [CrossRef]
- Akcam, F.Z.; Tigli, A.; Kaya, O.; Ciris, M.; Vural, H. Cytokine levels and histopathology in chronic hepatitis B and chronic hepatitis C. J. Interferon Cytokine Res. 2012, 32, 570–574. [CrossRef]
- 109. El-Tayeh, S.F.; Hussein, T.D.; El-Houseini, M.E.; Amer, M.A.; El-Sherbini, M.; Elshemey, W.M. Serological biomarkers of hepatocellular carcinoma in Egyptian patients. *Dis. Markers* 2012, *32*, 255–263. [CrossRef]

- Liu, W.; Chen, J.R.; Hsu, C.H.; Li, Y.H.; Chen, Y.M.; Lin, C.Y.; Huang, S.J.; Chang, Z.K.; Chen, Y.C.; Lin, C.H.; et al. A zebrafish model of intrahepatic cholangiocarcinoma by dual expression of hepatitis B virus X and hepatitis C virus core protein in liver. *Hepatology* 2012, 56, 2268–2276. [CrossRef]
- 111. Trehanpati, N.; Shrivastav, S.; Shivakumar, B.; Khosla, R.; Bhardwaj, S.; Chaturvedi, J.; Sukriti; Kumar, B.; Bose, S.; Mani Tripathi, D.; et al. Analysis of Notch and TGF-β Signaling Expression in Different Stages of Disease Progression During Hepatitis B Virus Infection. *Clin. Transl. Gastroenterol.* 2012, *3*, e23. [CrossRef] [PubMed]
- 112. Tache, D.; Bogdan, F.; Pisoschi, C.; Baniţă, M.; Stănciulescu, C.; Fusaru, A.M.; Comănescu, V. Evidence for the involvement of TGF-β1-CTGF axis in liver fibrogenesis secondary to hepatic viral infection. *Rom. J. Morphol. Embryol.* 2011, 52, 409–412. [PubMed]
- Murawski, M.R.; Litherland, S.A.; Clare-Salzler, M.J.; Davoodi-Semiromi, A. Upregulation of Foxp3 expression in mouse and human Treg is IL-2/STAT5 dependent: Implications for the NOD STAT5B mutation in diabetes pathogenesis. *Ann. N. Y. Acad. Sci.* 2006, 1079, 198–204. [CrossRef] [PubMed]
- 114. Qin, H.; Wang, L.; Feng, T.; Elson, C.O.; Niyongere, S.A.; Lee, S.J.; Reynolds, S.L.; Weaver, C.T.; Roarty, K.; Serra, R.; et al. TGF-beta promotes Th17 cell development through inhibition of SOCS3. *J. Immunol.* **2009**, *183*, 97–105. [CrossRef]
- 115. Yang, P.; Li, Q.J.; Feng, Y.; Zhang, Y.; Markowitz, G.J.; Ning, S.; Deng, Y.; Zhao, J.; Jiang, S.; Yuan, Y.; et al. TGF-β-miR-34a-CCL22 signaling-induced Treg cell recruitment promotes venous metastases of HBV-positive hepatocellular carcinoma. *Cancer Cell* 2012, 22, 291–303. [CrossRef]
- 116. Li, W.; Yu, X.; Chen, X.; Wang, Z.; Yin, M.; Zhao, Z.; Zhu, C. HBV induces liver fibrosis via the TGF-β1/miR-21-5p pathway. *Exp. Ther. Med.* **2021**, *21*, 169. [CrossRef]
- 117. Dong, K.S.; Chen, Y.; Yang, G.; Liao, Z.B.; Zhang, H.W.; Liang, H.F.; Chen, X.P.; Dong, H.H. TGF-β1 accelerates the hepatitis B virus X-induced malignant transformation of hepatic progenitor cells by upregulating miR-199a-3p. *Oncogene* 2020, 39, 1807–1820. [CrossRef]
- Zhao, Y.; Ye, W.; Wang, Y.D.; Chen, W.D. HGF/c-Met: A Key Promoter in Liver Regeneration. Front. Pharmacol. 2022, 13, 808855.
  [CrossRef]
- 119. Hoffmann, K.; Nagel, A.J.; Tanabe, K.; Fuchs, J.; Dehlke, K.; Ghamarnejad, O.; Lemekhova, A.; Mehrabi, A. Markers of liver regeneration-the role of growth factors and cytokines: A systematic review. *BMC Surg.* **2020**, *20*, 31. [CrossRef]
- 120. Patijn, G.A.; Lieber, A.; Schowalter, D.B.; Schwall, R.; Kay, M.A. Hepatocyte growth factor induces hepatocyte proliferation in vivo and allows for efficient retroviral-mediated gene transfer in mice. *Hepatology* **1998**, *28*, 707–716. [CrossRef]
- 121. Kogure, K.; Zhang, Y.Q.; Maeshima, A.; Suzuki, K.; Kuwano, H.; Kojima, I. The role of activin and transforming growth factor-beta in the regulation of organ mass in the rat liver. *Hepatology* **2000**, *31*, 916–921. [CrossRef] [PubMed]
- 122. Ichikawa, T.; Zhang, Y.Q.; Kogure, K.; Hasegawa, Y.; Takagi, H.; Mori, M.; Kojima, I. Transforming growth factor beta and activin tonically inhibit DNA synthesis in the rat liver. *Hepatology* **2001**, *34*, 918–925. [CrossRef] [PubMed]
- Ozden, M.; Kalkan, A.; Demirdag, K.; Denk, A.; Kilic, S.S. Hepatocyte growth factor (HGF) in patients with hepatitis B and meningitis. J. Infect. 2004, 49, 229–235. [CrossRef] [PubMed]
- 124. Barreiros, A.P.; Sprinzl, M.; Rosset, S.; Höhler, T.; Otto, G.; Theobald, M.; Galle, P.R.; Strand, D.; Strand, S. EGF and HGF levels are increased during active HBV infection and enhance survival signaling through extracellular matrix interactions in primary human hepatocytes. *Int. J. Cancer* 2009, 124, 120–129. [CrossRef]
- 125. Lee, H.W.; Choi, Y.; Lee, A.R.; Yoon, C.H.; Kim, K.H.; Choi, B.S.; Park, Y.K. Hepatocyte Growth Factor-Dependent Antiviral Activity of Activated cdc42-Associated Kinase 1 against Hepatitis B Virus. *Front. Microbiol.* **2021**, *12*, 800935. [CrossRef]
- 126. Horiguchi, N.; Takayama, H.; Toyoda, M.; Otsuka, T.; Fukusato, T.; Merlino, G.; Takagi, H.; Mori, M. Hepatocyte growth factor promotes hepatocarcinogenesis through c-Met autocrine activation and enhanced angiogenesis in transgenic mice treated with diethylnitrosamine. Oncogene 2002, 21, 1791–1799. [CrossRef]
- 127. Ogunwobi, O.O.; Liu, C. Hepatocyte growth factor upregulation promotes carcinogenesis and epithelial-mesenchymal transition in hepatocellular carcinoma via Akt and COX-2 pathways. *Clin. Exp. Metastasis* **2011**, *28*, 721–731. [CrossRef]
- 128. Huang, X.; Gan, G.; Wang, X.; Xu, T.; Xie, W. The HGF-MET axis coordinates liver cancer metabolism and autophagy for chemotherapeutic resistance. *Autophagy* 2019, *15*, 1258–1279. [CrossRef]
- 129. Álvarez-Mercado, A.I.; Caballeria-Casals, A.; Rojano-Alfonso, C.; Chávez-Reyes, J.; Micó-Carnero, M.; Sanchez-Gonzalez, A.; Casillas-Ramírez, A.; Gracia-Sancho, J.; Peralta, C. Insights into Growth Factors in Liver Carcinogenesis and Regeneration: An Ongoing Debate on Minimizing Cancer Recurrence after Liver Resection. *Biomedicines* 2021, 9, 1158. [CrossRef]
- Stöß, C.; Laschinger, M.; Wang, B.; Lu, M.; Altmayr, F.; Hartmann, D.; Hüser, N.; Holzmann, B. TLR3 promotes hepatocyte proliferation after partial hepatectomy by stimulating uPA expression and the release of tissue-bound HGF. *FASEB J.* 2020, 34, 10387–10397. [CrossRef]
- 131. Miura, A.; Ishiguro, K.; Koizumi, K.; Yaita, Y.; Ozaki-Masuzawa, Y.; Hosono, T.; Seki, T. Effects of pharmacological inhibition of plasminogen binding on liver regeneration in rats. *Biosci. Biotechnol. Biochem.* **2017**, *81*, 2105–2111. [CrossRef] [PubMed]
- 132. Tsai, M.C.; Yen, Y.H.; Chang, K.C.; Hung, C.H.; Chen, C.H.; Lin, M.T.; Hu, T.H. Elevated levels of serum urokinase plasminogen activator predict poor prognosis in hepatocellular carcinoma after resection. *BMC Cancer* **2019**, *19*, 1169. [CrossRef] [PubMed]

- Roskams, T.A.; Libbrecht, L.; Desmet, V.J. Progenitor cells in diseased human liver. Semin. Liver Dis. 2003, 23, 385–396. [CrossRef]
  [PubMed]
- 134. Anfuso, B.; El-Khobar, K.E.; Ie, S.I.; Avellini, C.; Radillo, O.; Raseni, A.; Tiribelli, C.; Sukowati, C.H.C. Activation of hepatic stem cells compartment during hepatocarcinogenesis in a HBsAg HBV-transgenic mouse model. *Sci. Rep.* 2018, *8*, 13168. [CrossRef]