Contents lists available at ScienceDirect



Review Article

Non-coding RNA Research



journal homepage: www.keaipublishing.com/en/journals/non-coding-rna-research

Non-coding RNA and Drug resistance in cholangiocarcinoma



Zhaowei Wu^a, Shiming Jiang^a, Yong Chen^{a,*}

^a Hepatobiliary Surgery, The First Affiliated Hospital of Chongqing Medical University, Medical College Street, Yuzhong District, 404100, Chongqing, China

ARTICLE INFO

Cholangiocarcinoma

Drug resistance

Keywords:

ncRNA

miRNA

IncRNA

Circ RNA

ABSTRACT

Cholangiocarcinoma is a highly aggressive cancer with a dismal prognosis and limited resectability. Chemotherapy has demonstrated tremendous benefits for patients with advanced and inoperable cancer, but drug resistance poses a significant obstacle. Despite recent progress in cancer therapy, the mechanisms driving drug resistance are multifaceted and not completely comprehended. Non-coding RNA refers to RNA molecules that are endogenous and do not code for proteins. Particularly microRNAs, long non-coding RNAs, circular RNAs, are widely acknowledged to be involved in cancer initiation, proliferation, and metastasis. Recently, evidences suggests that abnormal expression of non-coding RNAs contributes to resistance to different type of cancer therapies in cholangiocarcinoma. This occurs via the rewiring of signaling pathways including the reduction of anticancer drugs, apoptosis, interaction between cholangiocarcinoma and tumor-infiltrating immune cells, and cancer stemness. Thus, our review aims to demonstrate the potential of targeting non-coding RNA to override drug resistance and summarize the molecular mechanisms of how non-coding RNA contributes to drug resistance in cholangiocarcinoma.

1. Background

Cholangiocarcinoma, or CCA, is an aggressive cancer that originates from the cells lining the bile ducts. These bile ducts are part of the biliary tree, which includes both intrahepatic and extrahepatic ducts, as well as the common bile duct, hepatic ducts, and smaller bile ducts within the liver. CCA may arise in any part of the biliary tree, which encompasses the intrahepatic, perihilar, or distal bile ducts [1]. Accumulated evidence suggests a worldwide increase in CCA incidence. This trend has been observed in various regions across the globe [2]. The prognosis for patients with CCA is exceedingly grim. One of the chief obstacles in enhancing CCA outcomes is the paucity of early warning signs, which frequently results in late-stage detection. Consequently, a sizable percentage, approximately 70-80 %, of patients are diagnosed after the cancer has progressed to an advanced stage [3]. The prognosis for CCA is extremely poor, with less than 5 % of diagnosed patients surviving for at least five years. The low survival rate underscores the disease's aggressive nature and treatment-related challenges. Anticancer drugs have made noteworthy strides in increasing overall survival rates for CCA patients. These medications, such as chemotherapy, targeted therapy drugs, and immune checkpoint inhibitors, indicate significant outcomes in expanding life expectancy and ameliorating the quality of life for some patients. However, drug resistance remains a significant challenge in the treatment of CCA. Despite the initial response to therapy, many patients eventually develop resistance to the medications, leading to disease progression and limited treatment options. Therefore, it is crucial to understand the mechanisms underlying drug resistance for developing strategies to overcome this obstacle (see Tables 1–7, Fig. 1).

Gemcitabine, cisplatin, 5-fluorouracil (5-FU), and capecitabine are commonly used chemotherapy medications for treating CCA in both advanced and post-operative settings. These medications should be considered as standard treatment options for patients with CCA. Capecitabine serves as an adjuvant therapy for postoperative patients suffering from CCA to reduce the chances of cancer recurrence [4]. In the context of advanced and unresectable CCA, the preferred initial systemic chemotherapy typically utilizes a gemcitabine and cisplatin combination [5]. Meanwhile, patients who exhibit poor mismatch repair (MMR), high microsatellite instability (MSI), and high levels of PD-L1 expression have been identified as potential candidates for benefiting from immunotherapy [6]. Targeted drugs have demonstrated promise in the treatment of CCA. Notably, Pemigatinib, Ivosidenib, and Trametinib target specific genetic alterations present in CCA tumors [6]. However, drug resistance poses a considerable obstacle in cancer treatment and can lead to cancer recurrence and refractory disease. Numerous genes linked to chemoresistance have been identified by researchers. These genes are classified into various mechanisms of chemoresistance (MOC),

* Corresponding author. *E-mail addresses:* 18875215020@163.com (Z. Wu), 1041730269@qq.com (S. Jiang), Chenyong61@126.com (Y. Chen).

https://doi.org/10.1016/j.ncrna.2023.11.003

Received 20 September 2023; Received in revised form 7 November 2023; Accepted 8 November 2023 Available online 12 November 2023 2468-0540/© 2023 The Authors – Publishing services by Elsevier B V, on behalf of KeAi Communication

^{2468-0540/© 2023} The Authors. Publishing services by Elsevier B.V. on behalf of KeAi Communications Co. Ltd. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

Table 1

Oncogenic miRNAs and gemcitabine resistance in CCA.

miRNA	Expression	Gene and Pathway	Mechanism	Reference
miR-21	up	PTEN, PI3K/ AKT	MOC-1b and MOC- 5b	[22,25]
miR-130a- 3n	up	PPARα, PPARγ	MOC-4	[26,27]
miR-200b	up	PTPN12, Src	MOC-5b	[22]
miR-210	up	HIF-3α	MOC-5b	[28,29]
miR-210	up	RECK	MOC-5b	[30]
miR-181c	up	NDRG2	MOC-5b	[31,32]
let-7a	up	NF2/Stat3	MOC-5b	[33]
miR-1249	up	FZD8	MOC-7	[34]
miR-125a-	up	-	unknown	[35]
5p				

Table 2

Tumor suppressive miRNAs and gemcitabine resistance.

miRNA	Expression	Gene and Pathway	Mechanism	Reference
miR-29b	down	PIK3R1, MMP-2	MOC-5a and	[35]
			MOC-6	
miR-221	down	PIK3R1	MOC-5a	[35]
miR-637	down	LASP1	MOC-5a	[40,41]
miR-205	down	MMP2	MOC-6	[35,42]
miR-206	down	CAF	MOC-6	[43]
miR-424-	down	BCL9L	MOC-7	[44]
5p				
miR-520c-	down	MYCN/YAP1/	MOC-7	[45]
Зp		TEAD2/SOX2		
miR-149-	down	-	MOC-7	[46]
5p				

Table 3

MiRNAs and 5-FU resistance in CCA.

miR-20a-5p up SHOC, ERK1/2 MOC-1b [49] miR-320 down Mcl-1 MOC-5a [50] miR-204 down Bcl-2 MOC-5a [50] miR-106b down Zbtb7a MOC-5a [51-53] miR-328 down GPAM MOC-5a [54] miR-300 b/c down CSCs MOC-7 [55] miR-885-5p down MTPN unknown [56]

Table 4

MiRNAs and cisplatin resistance in CCA.

miRNA	Expression	Gene and Pathway	Mechanism	Reference
miR-199a-3p	up	mTOR/MDR1	MOC-1b	[60]
miR-637	down	LASP1	MOC-5a	[40,41]
miR-520c-3p	down	MYCN	MOC-7	[45]
miR-885-5p	down	MTPN	unknown	[56]

Table 5

MiRNAs and sorafenib resistance in CCA.

miRNA	Expression	Gene and Pathway	Mechanism	Reference
miR-141	up	hOCT1	MOC-1a	[68]
miR-138	down	SOX4	MOC-1a MOC-7	[69]

including MOC1-1a: reduced drug uptake, MOC-1b: enhanced drug export, MOC-2a: reduced prodrug activation, MOC-2b: enhanced drug inactivation, MOC-3: altered drug targets, MOC-4: enhanced DNA repair, MOC-5a: pro-apoptotic, MOC-5b: pro-survival, MOC-6: changes in tumor microenvironment, MOC-7: Phenotypic transition [7].

Table 6

lncRNA	Expression	Gene and Pathway	Mechanism	Reference
FALEC	up	miR-20a-5p/SHOC2	MOC-1b	[49,76]
LINC01714	down	FOXO3	MOC-4	[77,78]
lnc-PKD2-2-	up	miR-328/GPAM	MOC-5b	[54]
3				
HOTTIP	up	miR-637/LASP1	MOC-5b	[40,41]
HOXD-AS1	up	miR-520c/MYCN	MOC-7	[45]
LINC00665	up	miR-424/BCL9L	MOC-7	[44]
DLEU1	up	miR-149-5p/YAP1/	MOC-7	[46]
		TEAD2/SOX2		
NEAT-1	up	-	unknown	[79]

Table 7				
Circ-RNAs an	d drug	resistance	in	CC/

	8			
circ-RNA	Expression	Gene and Pathway	Mechanism	Reference
cNFIB SMARCA5 HMGCS1- 016	down down up	MEK1/ERK - miR1236-3p/CD73, GAL8	MOC-3 MOC-5a MOC-6	[81] [82,83] [84]

Non-coding RNAs (ncRNAs) are a diverse collection of RNA molecules that lack the ability to encode proteins but perform crucial functions in a variety of biological processes. They can be grouped into numerous subtypes based on their length and structural characteristics, such as microRNA, small nuclear RNA, Piwi-interaction RNA, small nucleolar RNA, and long non-coding RNA [8-10]. Dysregulation of microRNAs (miRNAs) is commonly observed in cancer, and several miRNAs are upregulated in cancer cells, contributing to tumor progression [11]. In addition to up-regulated miRNAs acting as oncogenes, some miRNAs also function as tumor suppressors but are down-regulated in various tumor types. Failure to regulate key genes that control cell growth, invasion, and metastasis due to decreased expression of these tumor suppressor miRNAs could contribute to tumor progression [12,13]. Certainly, long non-coding RNAs (lncRNAs) and circular RNAs (circRNAs) are capable of functioning as oncogenes or tumor suppressors in a variety of tumor types. These non-coding RNAs possess vital regulatory roles in the development and progression of cancer [14-17]. Recently, studies have shown that miRNAs, lncRNAs, and circRNAs play critical roles in regulating proliferation, invasion, and metastasis in CCA, as well as in the development of drug resistance. These non-coding RNAs are significant players in the molecular mechanisms driving drug resistance in CCA [18-20].

In this thorough examination, we seek to investigate the processes of biogenesis and regulation for ncRNAs and their connections with drug resistance in CCA.

2. miRNA and drug resistance

Currently, gemcitabine, cisplatin, 5-fluorouracil, and sorafenib are frequently administered in clinical settings. MiRNAs, ranging in size from 17 to 25 nucleotides, are a type of small non-coding RNA molecules that crucially contribute to gene regulation at the post-transcriptional level [21]. Extensive research has shown that dysregulation of miR-NAs significantly contributes to CCA's development, proliferation, and metastasis [22,23]. Recent studies have provided compelling evidence that the dysregulation of multiple miRNAs is associated with drug resistance in CCA. These dysregulated miRNAs have been identified as key players in the acquisition and maintenance of drug resistance, contributing to treatment failure and disease progression [24].

2.1. miRNAs related gemcitabine-resistance

Gemcitabine has been widely utilized as a neoadjuvant, adjuvant,

and palliative treatment for CCA, whether as a single agent or in combination therapy [36]. However, the development of resistance to gemcitabine is a significant challenge for many patients in their treatment of CCA. Various studies have implicated several miRNAs in the development of gemcitabine resistance in CCA. These oncogenic miR-NAs modulate cellular response to gemcitabine treatment, leading to reduced drug sensitivity and resistance. The miRNAs involved in this effect include miR-21, miR-130a-3p, miR-200b, miR-210, miR-181c, miR-1249, and miR-125a-5p.

Studies have revealed significant insights into the molecular mechanisms by which particular oncogenic miRNAs contribute to gemcitabine resistance in CCA. It have been reported that miR-21 is an oncogenic microRNA. Overexpression of miR-21 in CCA leads to the downregulation of the tumor-suppressor gene, phosphatase and tensin Homolog (PTEN). Subsequently, this dysregulation leads to activation of the PI3K/AKT signaling pathway, which in turn promotes cell survival and resistance to gemcitabine-induced apoptosis [22]. Besides, PI3K/AKT pathway could induce expression of membrane transporters and decreased susceptibility to chemotherapy [25]. The study revealed that dihydromyricetin and galangin could inhibit miR-21, resulting in decreased CCA development by promoting apoptosis in CCA cells [37, 38]. MiR-130a-3p could promotes resistance to gemcitabine by suppressing the expression of PPARG, which are involved in nucleotide metabolism [26]. It was reported that PPARG have anti-tumor effects via



Fig. 1. Several miRNAs, LncRNAs, circRNAs were identified to be involved in drug resistance in CCA.

cell-cycle arrest and DNA-damage repair [27]. Interestingly, pioglitazone, which is a PPARy agonist, is capable of reducing the expression of miR-130a-3p and thereby alleviating gemcitabine resistance. Several miRNAs play a part in resistance to gemcitabine by inhibiting apoptosis, including miR-200b, miR-210, let-7a, and miR-181c. Excessive expression of miR-200b in CCA could suppress PTP non-receptor type 12 (PTPN12), a phosphatase that negatively regulates the oncogene Src. By inhibiting PTPN12 expression, miR-200b contributes to resistance to gemcitabine via reduced apoptosis in CCA [22]. MiR-210 has been associated with a poor prognosis in CCA, and inhibiting endogenous miR-210 would improve gemcitabine sensitivity via HIF-1a [28]. In CCA, it was reported that HIF-1 α promotes proliferation, invasion, and migration, while down-regulated HIF-1 α enhances apoptosis [39]. Therefore, suppressed expression of HIF-3a, a negative regulator of HIF-1 α , induced by miR-210, resulted in gemcitabine resistance. Additionally, it has been reported that miR-210 promotes CCA migration, metastasis, and resistance to gemcitabine by inhibiting the expression of RECK. MiR-181c was up-regulated in CCA and contributed to down-regulated N-myc downstream-regulated gene 2 (NDRG2), which could inhibit CCA proliferation, metastasis and gemcitabine resistance [31]. NDRG2 has been shown to enhance cisplatin-induced apoptosis via modulation of the Bak-Mcl-1 ratio [32]. Based on this, we hypothesize that miR-181c may promote gemcitabine resistance through the NDRG2/Mcl-1 axis. Let-7a suppresses expression of neurofibromatosis 2 (NF2) and enhances Stat3 phosphorylation and kinase activity, which lead to decreased apoptosis caused by gemcitabine and 5-FU treatment [33]. MiR-1249 was upregulated in gemcitabine-resistant and CD133+ CCA. It could modulate the chemotherapy-induced enrichment of CD+133 cells and thus contribute to gemcitabine resistance through frizzled class receptor 8 (FZD8), a negative regulator of the canonical Wnt pathway [34]. MiR-125a-5p was found to be up-regulated in CCA compared to corresponding normal tissue, potentially reducing sensitivity to gemcitabine. However, the literature does not discuss its mechanisms [35].

Several tumor suppressor miRNAs, including miR-29b, miR-205, miR-221, miR-637, let-7a, miR-424-5p, miR-520c-3p, miR-149-5p, and miR-328, have been identified as playing essential roles in gemcitabine resistance in CCA. Understanding the functions of these mechanisms reveals potential strategies for surmounting gemcitabine resistance in CCA. Several tumor suppressive miRNA contribute to gemcitabine resistance by modulation of apoptosis, including miR-29b, miR-221, and miR-637. MiR-29b and miR-221 could suppress phosphoinositide-3-Kinase regulatory subunit 1 (PIK3R1) expression, a PI3K pathway component, promoting apoptosis induced by gemcitabine in CCA [35]. Furthermore, targeting matrix metalloproteinase-2 (MMP-2), miR-29b, and miR-205 augments gemcitabine sensitivity in CCA. MMP-2 plays a role in cancer cell invasion and metastasis [35]. Overexpression of miR-637 heightens sensitivity to gemcitabine through inhibition of the LIM and SH3 protein 1 (LASP1), which plays a role in cancer progression and cytoskeleton organization [41]. Studies suggest that LASP1 may enhance CCA proliferation, invasion, and migration, while suppressing apoptosis. Thus, miR-637 might promotes gemcitabine sensitivity by improving apoptosis [40]. Up-regulated miR-205 has been observed to resensitize primary gemcitabine-resistant CCA cells to gemcitabine. Subsequent research revealed the role of MMP2 as the target of miR-205, involved in tumor microenvironment remodeling that contributes to gemcitabine resistance [35]. MiR-206 was found to be downregulated in intrahepatic cholangiocarcinoma (iCCA), affecting tumor proliferation, invasion, and migration. It is noteworthy that miR-206 expression in CCA decreased when co-cultured with normal fibroblast cells (NFs) which subsequently transformed into cancer-associated fibroblasts (CAFs). The CAFs potentially induced gemcitabine resistance and tumor deterioration. Moreover, the heightened expression of miR-206 in both iCCA and CAF possesses the capability to enhance sensitivity to gemcitabine, resulting in improved therapeutic outcomes [43]. MiR-424-5p, miR-520c-3p and miR-149-5p expression levels were downregulated

in CCA. They had the potential to suppress both cancer stemness and EMT, thereby reversing the desensitization of CCA to gemcitabine. Increased miR-424-5p expression heightens gemcitabine sensitivity by blocking B-cell lymphoma 9 (BCL9) expression and activating the Wnt/ β -Catenin signaling pathway [44]. MiR-520c-3p would restore CCA sensitivity to gemcitabine by suppressing MYCN [45]. This reduction in miR-149-5p' expression was found to significantly promote malignant biological behavior of CCA, mediated by the YAP1/TEAD2/SOX2 pathway [46]. Consequently, the upregulation of miR-149-5p could effectively restore CCA sensitivity to gemcitabine.

2.2. miRNAs related 5-fluorouracil (5-FU) resistance

5-FU is a common anti-metabolite drug utilized to treat numerous types of cancer, including CCA [47]. In advanced CCA, the combination of 5-FU and oxaliplatin has been used as a second-line therapy after progression on first-line cisplatin and gemcitabine, resulting in enhanced overall survival in comparison to solely managing active symptoms [48]. However, the effectiveness of 5-FU is often hindered by the development of general chemotherapy resistance. Several recent studies have revealed the role of miRNAs in mediating 5-FU resistance, with numerous miRNAs acting as potential modifiers of resistance by suppressing tumors.

MiR-20a-5p was up-regulated in CCA compared to corresponding normal tissue. It was found that overexpression of miR-20a-5p reverse 5-FU resistance induced by FALEC, a onco-lncRNA [49]. MiR-20a-5p could inhibit expression of SHOC, a gene of MAPK pathway, and p-ERK1/2, which might promote apoptosis induced by 5-FU [57]. MiR-320 and miR-204 function as a tumor suppressor in CCA. However, their expression is markedly decreased in CCA compared to adjacent normal bile duct tissue. Up-regulated miR-320 and miR-204 could facilitate apoptosis induced 5-FU by negatively regulated expression of Mcl-1 or Bcl-2 [50]. MiR-106b is noticeably downregulated in CCA and act as tumor suppressive miRNA. Recent studies indicate that it enhances 5-FU sensitivity in CCA by directing zinc finger and BTB domain-containing 7A (Zbtb7a), a transcriptional regulator involved in various cellular processes [51]. In colorectal cancer and osteosarcoma, inhibition of Zbtb7 may enhance tumor susceptibility to chemotherapeutic drugs by facilitating apoptosis [52,53]. Thus, it is speculated that miR-106 could override 5-FU resistance by modulating apoptosis. MiR-328 inhibits tumor proliferation and facilitates apoptosis induced by 5-FU by down-regulating GPAM [54]. Up-regulated miR-200 b/c exhibit the ability to hinder tumor migration and enhance the susceptibility of CCA cells to 5-FU treatment [55]. In CCA, miR-200 b/c plays a role in the expansion of CD133+ cells (cancer stem cells), thus leading to 5-FU resistance. However, miR-200b plays a contradictory role in CCA. As mentioned above, it could promote resistance to gemcitabine in CCA. MiR-885-5p exhibits tumor-suppressive effects in CCA, hindering tumor proliferation and metastasis [58]. Additionally, miR-885-5p may enhance the sensitivity of CCA cells to 5-FU by targeting myotrophin (MTPN), a protein associated with cell migration and invasion [56]. However, the mechanism underlying miR-885-5p-mediated resistance to 5-FU has yet to be elucidated.

2.3. miRNAs related cisplatin-resistance

Cisplatin is a coordination compound containing platinum that has widespread use in cancer treatment, particularly CCA [59]. The combination of cisplatin and gemcitabine is now considered the first-line standard of care for advanced CCA, demonstrating superior survival outcomes in comparison to gemcitabine monotherapy [48]. However, developing cisplatin resistance remains a crucial challenge in treating CCA. Several studies have illuminated miRNAs' role in cisplatin resistance in CCA.

MiR-199a-3p is a tumor suppressor miRNA associated with cisplatin resistance in various types of tumors, including CCA. Its role involves regulating the mTOR signaling pathway by modulating Multidrug resistance gene 1 (MDR1), a multidrug resistance protein [61-63]. Upregulation of miR-199a-3p enhances CCA cell sensitivity to cisplatin by inhibiting MDR1 expression, thereby increasing its cytotoxic impact. The results demonstrate the potential of miR-199a-3p regulation in improving CCA chemotherapy [60]. MiR-637 has been linked to drug resistance in various types of cancer [64]. As mentioned above, miR-637 is significantly downregulated in CCA, and its decrease is linked to resistance to cisplatin via LASP1. MiR-637 might promotes cisplatin sensitivity by improving apoptosis [40]. MiR-520c-3p expression levels were downregulated in CCA. They had the potential to suppress both cancer stemness and EMT, thereby reversing the desensitization of CCA to cisplatin [45]. Upregulating miR-885-5p has demonstrated the capacity to overcome cisplatin resistance in cases of CCA. Its elevation of MTPN expression heightens cisplatin sensitivity, leading to the inhibition of tumor proliferation and metastasis [56]. However, the mechanism underlying miR-885-5p-mediated resistance to 5-FU has yet to be elucidated.

2.4. miRNAs related sorafenib-resistance

Sorafenib, a multi-targeted tyrosine kinase inhibitor (TKI), has been shown to be effective in treating advanced hepatocellular carcinoma (HCC) by targeting pathways associated with angiogenesis and proliferation [65]. However, the efficacy of sorafenib in treating CCA is hindered by general resistance. A prospective study of unresectable and advanced CCA patients treated with sorafenib and best supportive care revealed a disease control rate (DCR) of 15.9 %, despite the absence of a control group for comparison [66]. In addition, patients with aberrant human organic cation transporter 1 (OCT1) variants that facilitate the intracellular accumulation of sorafenib exhibited heightened sensitivity to the drug in CCA [67]. In the context of sorafenib resistance in CCA, several studies have identified downregulation of tumor suppressor miRNAs as contributing factors. These miRNAs include miR-141, miR-330, and miR-138.

MiR-141 and miR-330 are acknowledged as tumor-suppressive miRNAs in cases of CCA. They possess the capability to hinder tumor proliferation and metastasis by regulating the Hippo pathway, which is a key mediator of organ size and cell growth [70,71]. MiR-141 and miR-330 can enhance the sensitivity of CCA cells towards sorafenib by precisely targeting the hOCT1 protein responsible for intracellular sorafenib accumulation [68]. On the contrary, miR-138 is a significant contributor to overcoming sorafenib resistance in CCA by modulation of tumor stemness. It targets the transcription factor Sex-determining region Y-related (SRY) high-mobility group box 4 (SOX4), which regulates tumor growth, metastasis, and stemness [69,72]. Downregulation of miR-138 is associated with increased proliferation, migration, and invasion in CCA [73].

3. IncRNAs and drug resistance

Long non-coding RNAs (lncRNAs) are RNA molecules exceeding 200 nucleotides and not coding for proteins. They possess significant roles in diverse cellular processes, such as regulating chromatin dynamics, gene expression, cell growth, differentiation, and development [74]. It have the ability to act as enhancer-like molecules, activate additional non-coding RNAs like miRNA and piRNA, as well as compete for RNA molecule binding [75]. Several lncRNAs have been identified as involved in drug resistance. These include HOTTIP, HOXD-AS1, LINC00665, NEAT-1, lncRNA H19, lnc-PKD2-2-3, and lncRNAMEG3, all of which have been implicated in drug resistance mechanisms. These processes include but are not limited to - cell proliferation, apoptosis, DNA repair, and drug efflux. Additionally, lncRNAs have the ability to interact with other molecules such as miRNAs and proteins and modify gene expression, thus managing cellular responses in context to drug exposure.

The expression of FALEC was significantly higher in 5-FU resistant CCA patients in comparison to normal control. It could sponge up miR-20a-5p and decrease its expression, thereby potentially enhancing resistance to 5-FU [49]. MiR-20a-5p could inhibit expression of SHOC, a gene of MAPK pathway, and p-ERK1/2, which might promote apoptosis induced by 5-FU [57]. Additionally, LINC01714 was significantly down-regulated in CCA and was observed to be correlated with a favorable prognosis. LINC01714 enhances sensitivity to gemcitabine by suppressing the phosphorylation of forkhead box O3 (FOXO3) [77]. Recent studies suggest that FOXP3 may play a role in chemo-resistance through its modulation of DNA damage repair [78]. Lnc-PKD2-2-3 and HOTTIP were found to be involved in resistance to chemotherapeutic agents by modulation of apoptosis. Lnc-PKD2-2-3 can sponge up miR-328, leading to increased expression of GPAM and restore 5-FU resistance [54]. HOTTIP can promote resistance to gemcitabine and cisplatin chemotherapy in CCA by regulating the miR-637/LASP1 pathway [41]. HOXD-AS1, LINC00665, and DLEU1 were discovered to contribute to resistance to chemotherapy by regulating the stemness of CCA. HOXD-AS1 function as a miR-520c-3p sponge, ultimately upregulating the expression of the MYCN, which contribute to gemcitabine-resistance [45]. LINC00665 promote gemcitabine resistance by acting as a sponge for miR-424-5p, leading to activation of the BCL9L/Wnt/β-Catenin pathway [44]. DLEU1 hinders stemness preservation in CCA and heightens susceptibility to gemcitabine by serving as a miR-149-5p sponge, ultimately leading to augmented YAP1 expression [46]. NEAT-1 has been linked to the promotion of gemcitabine resistance in CCA, although additional research is necessary to comprehend the exact mechanisms at play [79].

These findings illuminate the intricate regulatory functions of long non-coding RNAs in drug resistance and prognosis in CCA. Additional research is required to comprehend the fundamental mechanisms and assess their applicability as treatment targets.

4. Circ-RNAs and drug resistance

Indeed, circular RNAs (circRNAs) play a crucial role in multiple biological processes, including chemotherapy resistance in cancer. Various mechanisms have been suggested by which circRNAs can contribute to drug resistance in CCA [80]. Understanding the particular circular RNAs involved in drug resistance mechanisms in CCA and their functional roles will provide significant insights for developing new therapeutic strategies to conquer drug resistance and enhance patient outcomes.

Several circular RNAs have critically contributed to CCA, including cNFIB, SMARCA5, and HMGCS1-016. The evidence points to their significant involvement in the disease. Circular RNAs cNFIB is downregulated in CCA, functions as a tumor suppressor [81]. Downregulated cNFIB was significantly associated with aggressive characteristic and poor prognosis. High levels of cNFIB have been found to delay resistance to trametinib, a MEK inhibitor, by competitively interacting with MEK1, and thereby inhibiting the ERK signaling pathway. The expression of circ- SMARCA5 was reduced in CCA compared with normal tissue and associated with better prognosis [82]. It could restore CCA cell sensitivity to gemcitabine and cisplatin. Recent studies have indicated that SMARCA5 suppresses the expression of miR-95-3p, promoting apoptosis [85]. Perhaps SMARCA5 could overcome chemo-resistance by controlling apoptosis. HMGCS1-016 is a circ-RNA that is upregulated in CCA and is associated with adverse clinical outcomes [84]. It was found that HMGCS1-016 might suppress tumor-infiltrating CD8⁺ T and contribute to suppressive tumor microenvironment via miR1236-3p/CD73 and GAL8. In CCA, patients with high-level of HMGCS1-016 benefit less from PD-1 antibody.

5. Discussion

Growing evidence suggests that non-coding RNAs, including

miRNAs, lncRNAs, and circRNAs, are involved in drug resistance mechanisms in CCA. Understanding the underlying mechanisms presents prospects to develop treatment plans aimed at subduing drug resistance in CCA. In summary, dysregulated ncRNAs in CCA, including miRNAs, lncRNAs, and circRNAs, have been found to contribute to drug resistance.

This is achieved through modulation of reduced drug uptake (MOC1-1a), enhanced drug export (MOC-1b), altered drug targets (MOC-3), enhanced DNA repair (MOC-4), pro-apoptotic (MOC-5a), pro-survival (MOC-5b), changes in the tumor microenvironment (MOC-6), and phenotypic transition (MOC-7). Therein, miR-141 and miR-330 promote CCA sensitivity to sorafenib by targeting hOCT1 for intracellular sorafenib accumulation. Furthermore, miR-21, miR199a-3p, and FALEC were upregulated in CCA and promoted drug export that conferred chemoresistance. Several dysregulated miRNAs may contribute to drug resistance in CCA by modulating apoptosis, including miR-200b, miR-210, miR-181c, let-7a, miR-221, miR637, miR-320, miR-204, miR-106b, and miR-328. In these miRNAs, down-regulated tumor suppressive miRNAs and up-regulated oncogenic miRNAs led to primary chemoresistance. Furthermore, the lncRNA lnc-PKD2-2-3, HOTTIP, and circular RNA SMARCA5 are downregulated in CCA, reflecting drug resistance via sponging miRNAs and inhibition of apoptosis induced by chemotherapeutic agents. In addition, several dysregulated ncRNAs would promote chemo-resistance by modulating of cancer stemness, including miR-1249, miR-149-5p, miR-424-5p, miR-520c-3p, miR-200 b/c, miR138, HOXD-AS1, LINC00665 and DLEU1. In summary, this study found that the dysregulated ncRNAs are involved in primary resistance to chemotherapy. Specifically, miR-130a-3p, miR-1249, miR-125a-5p, miR-29b, miR-221, miR-205, miR-424-5p, miR-20a-5p, miR-106b, miR-199a-3p, LINC00665, FALEC, and NEAT-1 were upregulated in drug-resistant CCA cells compared to non-resistant CCA cells. These up-regulated ncRNAs are believed to contribute to secondary drug resistance in CCA. Interestingly, tumor derived extracellular vesicles could promote chemoresistance in CCA by delivering miR-210. Furthermore, dysregulated miRNAs developed drug resistance by modulating of interaction between CCA cell and tumor infiltrating immune cell. Up-regulated miR-206 in CAF and CCA cell override drug resistance. On the contrary, circHMGCS1 was negatively associated with tumor-infiltrating CD8+T cell that contributed to drug resistance in CCA. It is confusing that while we believe CCA cells with stemness traits tend to be more resistant to chemotherapy, the mechanisms of chemoresistance in cancer stem cells are still complex. Should we consider CCA stemness as a unique factor involved in chemo-resistance? It was reported that cancer stem cells develop resistance to chemotherapy by maintaining a high level of ABC transporter expression, enhancing DNA damage repair, and promoting angiogenesis [86]. Thus, we are still investigating whether dysregulated ncRNAs contribute to drug resistance by modulating the aforementioned mechanisms. The interaction between CCA and tumor-infiltrating immune cell was also promising aspect to override drug resistance. As is well-known, miRNA from chemo-resistant tumor-derived exosomes may confer resistance to chemotherapeutic agents in sensitive tumor cells. Similarly, the secretion of miRNA from CAFs is involved in the development of chemo-resistance [87]. It was reported that ncRNAs derived from chemo-resistance tumor or suppressive tumor-infiltrating immune cell developed drug resistance by modulating of apoptosis [88], ferroptosis [89], CSCs [90,91], and EMT [92]. Perhaps we can divide MOCs into two subgroups: "pre-effect" and "effect". MOC-1 and MOC-2 belong to the "pre-effect" subgroup, where effective drug concentration is suppressed. MOC-3, MOC-4, and MOC-5 belong to the "effect" subgroup, where drug-induced DNA damage and apoptosis are inhibited and drug targets are altered. We believed that these dysregulated ncRNAs led to drug resistance by modulating of interaction between CCA and tumor-infiltrating immune cell (MOC-6) and CCA stemness (MOC-7). However, it is still necessary to consider the mechanisms of drug resistance in CCA that are involved in the tumor microenvironment and

stemness of cancer.

Targeting dysregulated miRNAs in CCA represents one potential pathway to override chemo-resistance. Therapeutic interventions may involve designing strategies to induce miRNA-like function by introducing synthetic miRNA mimics or miRNA analogs that replicate the effects of tumor-suppressive miRNAs. It is also feasible to explore the restoration of tumor-suppressive miRNAs that have been downregulated or the depletion of oncogenic miRNAs that have been upregulated as potential therapeutic interventions [93]. RNA therapeutics currently were used including ASO [94], siRNA [95], shRNA [96], miRNA mimic [97], antimiRs [98], miRNA sponges [99], and miRNA masking ASOs [100]. The development of nanoparticle-based delivery systems has greatly enhanced the delivery of miRNA therapeutics and surmounted various restrictions of conventional delivery methods. In targeting miRNAs for therapeutic objectives, nanoparticles provide a number of benefits [101,102]. However, despite the increased interest and substantial research regarding lncRNAs and circRNAs, there are currently no lncRNA or circRNA-based therapeutics in clinical development for CCA or other illnesses. The translation of ncRNAs as therapeutic targets is still in its nascent phases, requiring further research and clinical trials to uncover their potential. In this review, we have highlighted the involvement of various non-coding RNAs, including miRNAs, lncRNAs, and circRNAs, in drug resistance in CCA. Understanding the underlying mechanisms by which these non-coding RNAs contribute to drug resistance can provide valuable insights for the development of novel therapeutic strategies in CCA. As our comprehension of the molecular mechanisms and biological functions of non-coding RNAs in CCA deepens, their clinical potential as therapeutic targets or diagnostic markers will likely be further investigated. Further research and precisely-designed clinical trials are vital in realizing the complete potency of miRNAs, lncRNAs, and circRNAs for optimizing prognosis and treatment outcomes in CCA patients.

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Availability of data and material

Not applicable.

Funding

This study was funded by the National Natural Science Foundation of China under grants No. 81871261.

CRediT authorship contribution statement

Zhaowei Wu: Writing – review & editing, Writing – original draft, Software, Methodology, Investigation, Formal analysis, Data curation, Conceptualization. **Shiming Jiang:** Supervision, Project administration, Investigation. **Yong Chen:** Writing – review & editing, Writing – original draft, Resources, Project administration, Funding acquisition.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgements

Not applicable.

Abbreviations

CCA	Cholangiocarcinoma
ncRNA	Non-coding RNA
lncRNA	Long non-coding RNA
circRNA	Circular RNA
snRNA	Small nuclear RNA
snoRNA	Small nucleolar RNA
MDR	Multiple drug resistance
CSC	Cancer stem cell
EMT	Epithelial-mesenchymal transition
MOC	Mechanism of chemoresistance
5-FU	5-fluorouracil
CDDP	cisplatin
PTEN	Phosphatase and Tensin Homolog
PTPN12	PTP non-receptor type 12
PPARα	Peroxisome Proliferator-Activated α-Receptor
FZD8	Frizzled 8
HIF-3α	Hypoxia-inducible factor-3α
NDRG2	N-myc downstream-regulated gene 2
PIK3R1	Phosphoinositide-3-Kinase Regulatory Subunit 1
MMP-2	Matrix metalloproteinase-2
LASP1	LIM and SH3 protein 1
STAT3	Signal transducer and activator of transcription 3
BCL9	B-cell lymphoma 9
YAP1	Yes-associated protein1
GPAM	Glycerol-3-phosphate acyltransferase mitochondrial
MCL-1	Myeloid cell leukemia sequence 1
MTPN	Myotrophin
ZBTB7A	Zinc finger and BTB domain-containing 7A
mTOR	Mechanistic target of rapamycin
MDR-1	Multidrug resistance gene
OCT1	human organic cation transporter 1
SOX4	Sex-determining region Y-related (SRY) high-mobility
	box 4

FOXO3 forkhead box O3

References

- S. Rizvi, S.A. Khan, C.L. Hallemeier, R.K. Kelley, G.J. Gores, Cholangiocarcinoma - evolving concepts and therapeutic strategies, Nat. Rev. Clin. Oncol. 15 (2) (2018) 95–111.
- [2] J.M. Banales, J.J.G. Marin, A. Lamarca, P.M. Rodrigues, S.A. Khan, L.R. Roberts, et al., Cholangiocarcinoma 2020: the next horizon in mechanisms and management, Nat. Rev. Gastroenterol. Hepatol. 17 (9) (2020) 557–588.
- [3] S. Rizvi, G.J. Gores, Pathogenesis, diagnosis, and management of cholangiocarcinoma, Gastroenterology 145 (6) (2013) 1215–1229.
- [4] J.N. Primrose, R.P. Fox, D.H. Palmer, H.Z. Malik, R. Prasad, D. Mirza, et al., Capecitabine compared with observation in resected biliary tract cancer (BILCAP): a randomised, controlled, multicentre, phase 3 study, Lancet Oncol. 20 (5) (2019) 663–673.
- [5] J. Valle, H. Wasan, D.H. Palmer, D. Cunningham, A. Anthoney, A. Maraveyas, et al., Cisplatin plus gemcitabine versus gemcitabine for biliary tract cancer, N. Engl. J. Med. 362 (14) (2010) 1273–1281.
- [6] A.E. Kam, A. Masood, R.T. Shroff, Current and emerging therapies for advanced biliary tract cancers, Lancet Gastroenterol Hepatol 6 (11) (2021) 956–969.
- [7] J.J.G. Marin, E. Lozano, E. Herraez, M. Asensio, S. Di Giacomo, M.R. Romero, et al., Chemoresistance and chemosensitization in cholangiocarcinoma, Biochim. Biophys. Acta, Mol. Basis Dis. 1864 (4 Pt B) (2018) 1444–1453.
- [8] M. Esteller, P.P. Pandolfi, The epitranscriptome of noncoding RNAs in cancer, Cancer Discov. 7 (4) (2017) 359–368.
- [9] E. Anastasiadou, L.S. Jacob, F.J. Slack, Non-coding RNA networks in cancer, Nat. Rev. Cancer 18 (1) (2018) 5–18.
- [10] F.J. Slack, A.M. Chinnaiyan, The role of non-coding RNAs in oncology, Cell 179 (5) (2019) 1033–1055.
- [11] H. Yan, P. Bu, Non-coding RNA in cancer, Essays Biochem. 65 (4) (2021) 625–639.
- [12] B. Jin, W. Wang, X.X. Meng, G. Du, J. Li, S.Z. Zhang, et al., Let-7 inhibits selfrenewal of hepatocellular cancer stem-like cells through regulating the epithelial-

mesenchymal transition and the Wnt signaling pathway, BMC Cancer 16 (1) (2016) 863.

- [13] X.X. Li, X. Di, S. Cong, Y. Wang, K. Wang, The role of let-7 and HMGA2 in the occurrence and development of lung cancer: a systematic review and metaanalysis, Eur. Rev. Med. Pharmacol. Sci. 22 (23) (2018) 8353–8366.
- [14] H. Luo, G. Zhu, J. Xu, Q. Lai, B. Yan, Y. Guo, et al., HOTTIP IncRNA promotes hematopoietic stem cell self-renewal leading to AML-like disease in mice, Cancer Cell 36 (6) (2019), 645-59.e8.
- [15] C.E. Olivero, E. Martínez-Terroba, J. Zimmer, C. Liao, E. Tesfaye, N. Hooshdaran, et al., p53 activates the long noncoding RNA Pvt1b to inhibit myc and suppress tumorigenesis, Mol. Cell. 77 (4) (2020), 761-74.e8.
- [16] S. Wang, F. Liu, H. Ma, X. Cui, S. Yang, R. Qin, circCDYL acts as a tumor suppressor in triple negative breast cancer by sponging miR-190a-3p and upregulating TP53INP1, Clin. Breast Cancer 20 (5) (2020) 422–430.
- [17] Y. Xu, K. Leng, Y. Yao, P. Kang, G. Liao, Y. Han, et al., A circular RNA, cholangiocarcinoma-associated circular RNA 1, contributes to cholangiocarcinoma progression, induces angiogenesis, and disrupts vascular endothelial barriers, Hepatology 73 (4) (2021) 1419–1435.
- [18] Z. Liang, B. Zhu, D. Meng, X. Shen, X. Li, Z. Wang, et al., Down-regulation of IncRNA-NEF indicates poor prognosis in intrahepatic cholangiocarcinoma, Biosci. Rep. 39 (5) (2019).
- [19] Y. Xu, K. Leng, Z. Li, F. Zhang, X. Zhong, P. Kang, et al., The prognostic potential and carcinogenesis of long non-coding RNA TUG1 in human cholangiocarcinoma, Oncotarget 8 (39) (2017) 65823–65835.
- [20] R. Castro-Oropeza, J. Melendez-Zajgla, V. Maldonado, K. Vazquez-Santillan, The emerging role of lncRNAs in the regulation of cancer stem cells, Cell. Oncol. 41 (6) (2018) 585–603.
- [21] D.P. Bartel, MicroRNAs: genomics, biogenesis, mechanism, and function, Cell 116 (2) (2004) 281–297.
- [22] F. Meng, R. Henson, M. Lang, H. Wehbe, S. Maheshwari, J.T. Mendell, et al., Involvement of human micro-RNA in growth and response to chemotherapy in human cholangiocarcinoma cell lines, Gastroenterology 130 (7) (2006) 2113–2129.
- [23] F.M. Selaru, A.V. Olaru, T. Kan, S. David, Y. Cheng, Y. Mori, et al., MicroRNA-21 is overexpressed in human cholangiocarcinoma and regulates programmed cell death 4 and tissue inhibitor of metalloproteinase 3, Hepatology 49 (5) (2009) 1595–1601.
- [24] B. He, Z. Zhao, Q. Cai, Y. Zhang, P. Zhang, S. Shi, et al., miRNA-based biomarkers, therapies, and resistance in Cancer, Int. J. Biol. Sci. 16 (14) (2020) 2628–2647.
- [25] R. Liu, Y. Chen, G. Liu, C. Li, Y. Song, Z. Cao, et al., PI3K/AKT pathway as a key link modulates the multidrug resistance of cancers, Cell Death Dis. 11 (9) (2020) 797.
- [26] K. Asukai, K. Kawamoto, H. Eguchi, M. Konno, A. Asai, Y. Iwagami, et al., Micro-RNA-130a-3p regulates gemcitabine resistance via PPARG in cholangiocarcinoma, Ann. Surg Oncol. 24 (8) (2017) 2344–2352.
- [27] J.A. Pedroza-Garcia, Y. Xiang, L. De Veylder, Cell cycle checkpoint control in response to DNA damage by environmental stresses, Plant J. 109 (3) (2022) 490–507.
- [28] R. Silakit, Y. Kitirat, S. Thongchot, W. Loilome, A. Techasen, P. Ungarreevittaya, et al., Potential role of HIF-1-responsive microRNA210/HIF3 axis on gemcitabine resistance in cholangiocarcinoma cells, PLoS One 13 (6) (2018), e0199827.
- [29] X. Zhang, Z. Qi, H. Yin, G. Yang, Interaction between p53 and Ras signaling controls cisplatin resistance via HDAC4- and HIF-1α-mediated regulation of apoptosis and autophagy, Theranostics 9 (4) (2019) 1096–1114.
- [30] Y. Fu, Y. Liu, K. Liu, L. Tan, Tumor cell-derived extracellular vesicles promote the growth, metastasis and chemoresistance in cholangiocarcinoma by delivering microRNA-210 to downregulate RECK, Mol. Biotechnol. 65 (7) (2023) 1151–1164.
- [31] J. Wang, C. Xie, S. Pan, Y. Liang, J. Han, Y. Lan, et al., N-myc downstreamregulated gene 2 inhibits human cholangiocarcinoma progression and is regulated by leukemia inhibitory factor/MicroRNA-181c negative feedback pathway, Hepatology 64 (5) (2016) 1606–1622.
- [32] S. Park, S.S. Oh, K.W. Lee, Y.K. Lee, N.Y. Kim, J.H. Kim, et al., NDRG2 contributes to cisplatin sensitivity through modulation of BAK-to-Mcl-1 ratio, Cell Death Dis. 9 (2) (2018) 30.
- [33] F. Meng, R. Henson, H. Wehbe-Janek, H. Smith, Y. Ueno, T. Patel, The MicroRNA let-7a modulates interleukin-6-dependent STAT-3 survival signaling in malignant human cholangiocytes, J. Biol. Chem. 282 (11) (2007) 8256–8264.
- [34] P. Carotenuto, S. Hedayat, M. Fassan, V. Cardinale, A. Lampis, V. Guzzardo, et al., Modulation of biliary cancer chemo-resistance through MicroRNA-mediated rewiring of the expansion of CD133+ cells, Hepatology 72 (3) (2020) 982–996.
- [35] K. Okamoto, K. Miyoshi, Y. Murawaki, miR-29b, miR-205 and miR-221 enhance chemosensitivity to gemcitabine in HuH28 human cholangiocarcinoma cells, PLoS One 8 (10) (2013), e77623.
- [36] Z.V. Fong, S.A. Brownlee, M. Qadan, K.K. Tanabe, The clinical management of cholangiocarcinoma in the United States and europe: a comprehensive and evidence-based comparison of guidelines, Ann. Surg Oncol. 28 (5) (2021) 2660–2674.
- [37] Y. Zou, R. Li, D. Kuang, M. Zuo, W. Li, W. Tong, et al., Galangin inhibits cholangiocarcinoma cell growth and metastasis through downregulation of MicroRNA-21 expression, BioMed Res. Int. 2020 (2020), 5846938.
- [38] L. Chen, Z.S. Yang, Y.Z. Zhou, Y. Deng, P. Jiang, S.L. Tan, Dihydromyricetin inhibits cell proliferation, migration, invasion and promotes apoptosis via regulating miR-21 in Human Cholangiocarcinoma Cells, J. Cancer 11 (19) (2020) 5689–5699.

group

- [**39**] A. Yu, L. Zhao, Q. Kang, J. Li, K. Chen, H. Fu, Transcription factor HIF1α [**67**] E. Herraez, E. Loza
- promotes proliferation, migration, and invasion of cholangiocarcinoma via long noncoding RNA H19/microRNA-612/Bcl-2 axis, Transl. Res. 224 (2020) 26–39.
 [40] H. Zhang, Z. Li, B. Chu, F. Zhang, Y. Zhang, F. Ke, et al., Upregulated LASP-1
- correlates with a malignant phenotype and its potential therapeutic role in human cholangiocarcinoma, Tumour Biol 37 (6) (2016) 8305–8315.
- [41] K. Gao, S. Chen, X. Yang, HOTTIP enhances gemcitabine and cisplatin resistance through sponging miR-637 in cholangiocarcinoma, Front. Oncol. 11 (2021), 664916.
- [42] P. Vihinen, V.M. Kähäri, Matrix metalloproteinases in cancer: prognostic markers and therapeutic targets, Int. J. Cancer 99 (2) (2002) 157–166.
- [43] R. Yang, D. Wang, S. Han, Y. Gu, Z. Li, L. Deng, et al., MiR-206 suppresses the deterioration of intrahepatic cholangiocarcinoma and promotes sensitivity to chemotherapy by inhibiting interactions with stromal CAFs, Int. J. Biol. Sci. 18 (1) (2022) 43–64.
- [44] M. Lu, X. Qin, Y. Zhou, G. Li, Z. Liu, X. Geng, et al., Long non-coding RNA LINC00665 promotes gemcitabine resistance of Cholangiocarcinoma cells via regulating EMT and stemness properties through miR-424-5p/BCL9L axis, Cell Death Dis. 12 (1) (2021) 72.
- [45] J. Li, X. Jiang, Z. Li, L. Huang, D. Ji, L. Yu, et al., SP1-induced HOXD-AS1 promotes malignant progression of cholangiocarcinoma by regulating miR-520c-3p/MYCN, Aging (Albany NY) 12 (16) (2020) 16304–16325.
- [46] J. Li, X. Jiang, Y. Xu, P. Kang, P. Huang, N. Meng, et al., YY1-induced DLEU1/ miR-149-5p promotes malignant biological behavior of cholangiocarcinoma through upregulating YAP1/TEAD2/SOX2, Int. J. Biol. Sci. 18 (11) (2022) 4301–4315.
- [47] C. Sethy, C.N. Kundu, 5-Fluorouracil (5-FU) resistance and the new strategy to enhance the sensitivity against cancer: implication of DNA repair inhibition, Biomed. Pharmacother. 137 (2021), 111285.
- [48] A. Lamarca, J. Edeline, L. Goyal, How I treat biliary tract cancer, ESMO Open 7 (1) (2022), 100378.
- [49] H. Du, S. Hou, L. Zhang, C. Liu, T. Yu, W. Zhang, LncRNA FALEC increases the proliferation, migration and drug resistance of cholangiocarcinoma through competitive regulation of miR-20a-5p/SHOC2 axis, Aging (Albany NY) 15 (9) (2023) 3759–3770.
- [50] L. Chen, H.X. Yan, W. Yang, L. Hu, L.X. Yu, Q. Liu, et al., The role of microRNA expression pattern in human intrahepatic cholangiocarcinoma, J. Hepatol. 50 (2) (2009) 358–369.
- [51] D. Jiao, Y. Yan, S. Shui, G. Wu, J. Ren, Y. Wang, et al., miR-106b regulates the 5fluorouracil resistance by targeting Zbtb7a in cholangiocarcinoma, Oncotarget 8 (32) (2017) 52913–52922.
- [52] Z. Wang, X. Zhao, W. Wang, Y. Liu, Y. Li, J. Gao, et al., ZBTB7 evokes 5-fluorouracil resistance in colorectal cancer through the NF-κB signaling pathway, Int. J. Oncol. 53 (5) (2018) 2102–2110.
- [53] L. Zhang, Y. Wang, X. Li, X. Xia, N. Li, R. He, et al., ZBTB7A enhances osteosarcoma chemoresistance by transcriptionally repressing lncRNALINC00473-IL24 activity, Neoplasia 19 (11) (2017) 908–918.
- [54] L. Zhang, D. Ma, F. Li, G. Qiu, D. Sun, Z. Zeng, Inc-PKD2-2-3/miR-328/GPAM ceRNA network induces cholangiocarcinoma proliferation, invasion and 5-FU chemoresistance, Front. Oncol. 12 (2022), 871281.
- [55] F. Peng, J. Jiang, Y. Yu, R. Tian, X. Guo, X. Li, et al., Direct targeting of SUZ12/ ROCK2 by miR-200b/c inhibits cholangiocarcinoma tumourigenesis and metastasis, Br. J. Cancer 109 (12) (2013) 3092–3104.
- [56] Y. Wang, W. Zhang, L. Chen, W. Chen, S. Xu, L. Tang, et al., The ATO/miRNA-885-5p/MTPN axis induces reversal of drug-resistance in cholangiocarcinoma, Cell. Oncol. 44 (4) (2021) 907–916.
- [57] R. Roskoski Jr., Targeting ERK1/2 protein-serine/threonine kinases in human cancers, Pharmacol. Res. 142 (2019) 151–168.
- [58] S. Lixin, S. Wei, S. Haibin, L. Qingfu, P. Tiemin, miR-885-5p inhibits proliferation and metastasis by targeting IGF2BP1 and GALNT3 in human intrahepatic cholangiocarcinoma, Mol. Carcinog. 59 (12) (2020) 1371–1381.
- [59] S. Dasari, P.B. Tchounwou, Cisplatin in cancer therapy: molecular mechanisms of action, Eur. J. Pharmacol. 740 (2014) 364–378.
- [60] Q. Li, X. Xia, J. Ji, J. Ma, L. Tao, L. Mo, et al., MiR-199a-3p enhances cisplatin sensitivity of cholangiocarcinoma cells by inhibiting mTOR signaling pathway and expression of MDR1, Oncotarget 8 (20) (2017) 33621–33630.
- [61] Y. Cui, F. Wu, D. Tian, T. Wang, T. Lu, X. Huang, et al., miR-199a-3p enhances cisplatin sensitivity of ovarian cancer cells by targeting ITGB8, Oncol. Rep. 39 (4) (2018) 1649–1657.
- [62] Y. Deng, F. Zhao, L. Hui, X. Li, D. Zhang, W. Lin, et al., Suppressing miR-199a-3p by promoter methylation contributes to tumor aggressiveness and cisplatin resistance of ovarian cancer through promoting DDR1 expression, J. Ovarian Res. 10 (1) (2017) 50.
- [63] X. Fan, S. Zhou, M. Zheng, X. Deng, Y. Yi, T. Huang, MiR-199a-3p enhances breast cancer cell sensitivity to cisplatin by downregulating TFAM (TFAM), Biomed. Pharmacother. 88 (2017) 507–514.
- [64] B. Xu, J. Guo, M. Chen, Circ_0017274 acts on miR-637/CDX2 axis to facilitate cisplatin resistance in gastric cancer, Clin. Exp. Pharmacol. Physiol. 49 (10) (2022) 1105–1115.
- [65] W. Tang, Z. Chen, W. Zhang, Y. Cheng, B. Zhang, F. Wu, et al., The mechanisms of sorafenib resistance in hepatocellular carcinoma: theoretical basis and therapeutic aspects, Signal Transduct. Targeted Ther. 5 (1) (2020) 87.
- [66] X. Luo, W. Jia, Z. Huang, X. Li, B. Xing, X. Jiang, et al., Effectiveness and safety of sorafenib in the treatment of unresectable and advanced intrahepatic cholangiocarcinoma: a pilot study, Oncotarget 8 (10) (2017) 17246–17257.

- [67] E. Herraez, E. Lozano, R.I. Macias, J. Vaquero, L. Bujanda, J.M. Banales, et al., Expression of SLC22A1 variants may affect the response of hepatocellular carcinoma and cholangiocarcinoma to sorafenib, Hepatology 58 (3) (2013) 1065–1073.
- [68] E. Lozano, R.I.R. Macias, M.J. Monte, M. Asensio, S. Del Carmen, L. Sanchez-Vicente, et al., Causes of hOCT1-dependent cholangiocarcinoma resistance to sorafenib and sensitization by tumor-selective gene therapy, Hepatology 70 (4) (2019) 1246–1261.
- [69] Y. Zheng, J. Zhang, B. Ye, miR-138 mediates sorafenib-induced cell survival and is associated with poor prognosis in cholangiocarcinoma cells, Clin. Exp. Pharmacol. Physiol. 47 (3) (2020) 459–465.
- [70] L. Xia, X. Chen, J. Yang, S. Zhu, L. Zhang, Q. Yin, et al., Long non-coding RNA-PAICC promotes the tumorigenesis of human intrahepatic cholangiocarcinoma by increasing YAP1 transcription, Front. Oncol. 10 (2020), 595533.
- [71] Y. Xin, X. He, W. Zhao, M. Zhan, Y. Li, J. Xiao, et al., LncRNA PCAT6 increased cholangiocarcinoma cell proliferation and invasion via modulating miR-330-5p, Am J Transl Res 11 (9) (2019) 6185–6195.
- [73] Q. Wang, H. Tang, S. Yin, C. Dong, Downregulation of microRNA-138 enhances the proliferation, migration and invasion of cholangiocarcinoma cells through the upregulation of RhoC/p-ERK/MMP-2/MMP-9, Oncol. Rep. 29 (5) (2013) 2046–2052.
- [74] A. Bhan, S.S. Mandal, LncRNA HOTAIR: a master regulator of chromatin dynamics and cancer, Biochim. Biophys. Acta 1856 (1) (2015) 151–164.
- [75] G. St Laurent, C. Wahlestedt, P. Kapranov, The Landscape of long noncoding RNA classification, Trends Genet. 31 (5) (2015) 239–251.
- [76] R.M. Fleeman, G. Debevec, K. Antonen, J.L. Adams, R.G. Santos, G.S. Welmaker, et al., Identification of a novel polyamine scaffold with potent efflux pump inhibition activity toward multi-drug resistant bacterial pathogens, Front. Microbiol. 9 (2018) 1301.
- [77] S. Shen, J. Wang, B. Zheng, Y. Tao, M. Li, Y. Wang, et al., LINC01714 enhances gemcitabine sensitivity by modulating FOXO3 phosphorylation in cholangiocarcinoma, Mol. Ther. Nucleic Acids 19 (2020) 446–457.
- [78] S. Yao, L.Y. Fan, E.W. Lam, The FOXO3-FOXM1 axis: a key cancer drug target and a modulator of cancer drug resistance, Semin. Cancer Biol. 50 (2018) 77–89.
- [79] M. Parasramka, I.K. Yan, X. Wang, P. Nguyen, A. Matsuda, S. Maji, et al., BAP1 dependent expression of long non-coding RNA NEAT-1 contributes to sensitivity to gemcitabine in cholangiocarcinoma, Mol. Cancer 16 (1) (2017) 22.
- [80] C. Cui, J. Yang, X. Li, D. Liu, L. Fu, X. Wang, Functions and mechanisms of circular RNAs in cancer radiotherapy and chemotherapy resistance, Mol. Cancer 19 (1) (2020) 58.
- [81] J. Du, T. Lan, H. Liao, X. Feng, X. Chen, W. Liao, et al., CircNFIB inhibits tumor growth and metastasis through suppressing MEK1/ERK signaling in intrahepatic cholangiocarcinoma, Mol. Cancer 21 (1) (2022) 18.
- [82] Q. Lu, T. Fang, Circular RNA SMARCA5 correlates with favorable clinical tumor features and prognosis, and increases chemotherapy sensitivity in intrahepatic cholangiocarcinoma, J. Clin. Lab. Anal. 34 (4) (2020), e23138.
- [83] Z. Aydin Ö, W. Vermeulen, H. Lans, ISWI chromatin remodeling complexes in the DNA damage response, Cell Cycle 13 (19) (2014) 3016–3025.
- [84] Y.P. Xu, Z.N. Dong, S.W. Wang, Y.M. Zheng, C. Zhang, Y.Q. Zhou, et al., circHMGCS1-016 reshapes immune environment by sponging miR-1236-3p to regulate CD73 and GAL-8 expression in intrahepatic cholangiocarcinoma, J. Exp. Clin. Cancer Res. 40 (1) (2021) 290.
- [85] G. Wang, X. Gao, Z. Sun, T. He, C. Huang, S. Li, et al., Circular RNA SMARCA5 inhibits cholangiocarcinoma via microRNA-95-3p/tumor necrosis factor receptor associated factor 3 axis, Anti Cancer Drugs 34 (9) (2023) 1002–1009.
- [86] N.K. Lytle, A.G. Barber, T. Reya, Stem cell fate in cancer growth, progression and therapy resistance, Nat. Rev. Cancer 18 (11) (2018) 669–680.
- [87] H. Zhang, T. Deng, R. Liu, T. Ning, H. Yang, D. Liu, et al., CAF secreted miR-522 suppresses ferroptosis and promotes acquired chemo-resistance in gastric cancer, Mol. Cancer 19 (1) (2020) 43.
- [88] X. Qin, H. Guo, X. Wang, X. Zhu, M. Yan, X. Wang, et al., Exosomal miR-196a derived from cancer-associated fibroblasts confers cisplatin resistance in head and neck cancer through targeting CDKN1B and ING5, Genome Biol. 20 (1) (2019) 12.
- [89] R. Qi, Y. Bai, K. Li, N. Liu, Y. Xu, E. Dal, et al., Cancer-associated fibroblasts suppress ferroptosis and induce gemcitabine resistance in pancreatic cancer cells by secreting exosome-derived ACSL4-targeting miRNAs, Drug Resist. Updates 68 (2023), 100960.
- [90] J. Ren, L. Ding, D. Zhang, G. Shi, Q. Xu, S. Shen, et al., Carcinoma-associated fibroblasts promote the stemness and chemoresistance of colorectal cancer by transferring exosomal lncRNA H19, Theranostics 8 (14) (2018) 3932–3948.
- [91] J. Zhuang, L. Shen, M. Li, J. Sun, J. Hao, J. Li, et al., Cancer-associated fibroblastderived miR-146a-5p generates a niche that promotes bladder cancer stemness and chemoresistance, Cancer Res. 83 (10) (2023) 1611–1627.
- [92] G. Shan, X. Zhou, J. Gu, D. Zhou, W. Cheng, H. Wu, et al., Downregulated exosomal microRNA-148b-3p in cancer associated fibroblasts enhance chemosensitivity of bladder cancer cells by downregulating the Wnt/β-catenin pathway and upregulating PTEN, Cell. Oncol. 44 (1) (2021) 45–59.
- [93] M. Winkle, S.M. El-Daly, M. Fabbri, G.A. Calin, Noncoding RNA therapeutics challenges and potential solutions, Nat. Rev. Drug Discov. 20 (8) (2021) 629–651.
- [94] S.T. Crooke, Molecular mechanisms of antisense oligonucleotides, Nucleic Acid Therapeut. 27 (2) (2017) 70–77.

- [95] S.M. Elbashir, J. Harborth, W. Lendeckel, A. Yalcin, K. Weber, T. Tuschl, Duplexes of 21-nucleotide RNAs mediate RNA interference in cultured mammalian cells, Nature 411 (6836) (2001) 494–498.
- [96] D.D. Rao, N. Senzer, Z. Wang, P. Kumar, C.M. Jay, J. Nemunaitis, Bifunctional short hairpin RNA (bi-shRNA): design and pathway to clinical application, Methods Mol. Biol. 942 (2013) 259–278.
- [97] E. van Rooij, S. Kauppinen, Development of microRNA therapeutics is coming of age, EMBO Mol. Med. 6 (7) (2014) 851–864.
- [98] J. Krützfeldt, N. Rajewsky, R. Braich, K.G. Rajeev, T. Tuschl, M. Manoharan, et al., Silencing of microRNAs in vivo with 'antagomirs', Nature 438 (7068) (2005) 685–689.
- [99] M.S. Ebert, J.R. Neilson, P.A. Sharp, MicroRNA sponges: competitive inhibitors of small RNAs in mammalian cells, Nat. Methods 4 (9) (2007) 721–726.
- [100] Z. Wang, The principles of MiRNA-masking antisense oligonucleotides technology, Methods Mol. Biol. 676 (2011) 43–49.
- [101] M.J. Mitchell, M.M. Billingsley, R.M. Haley, M.E. Wechsler, N.A. Peppas, R. Langer, Engineering precision nanoparticles for drug delivery, Nat. Rev. Drug Discov. 20 (2) (2021) 101–124.
- [102] S.W.L. Lee, C. Paoletti, M. Campisi, T. Osaki, G. Adriani, R.D. Kamm, et al., MicroRNA delivery through nanoparticles, J. Contr. Release 313 (2019) 80–95.