

## Draft Genome Sequence of *Kitasatospora cheerisanensis* KCTC 2395, Which Produces Plecomacrolide against Phytopathogenic Fungi

Jae Yoon Hwang,<sup>a</sup> Soo Hee Kim,<sup>a</sup> Hye Ryeung Oh,<sup>a</sup> Yong-Joon Cho,<sup>b</sup> Jongsik Chun,<sup>b</sup> Young Ryun Chung,<sup>c</sup> Doo Hyun Nam<sup>a</sup>

College of Pharmacy, Yeungnam University, Gyongsan, South Korea<sup>a</sup>; Chunlab, Inc., Seoul National University, Seoul, South Korea<sup>b</sup>; Department of Microbiology, Gyeongsang National University, Jinju, South Korea<sup>c</sup>

*Kitasatospora cheerisanensis* KCTC 2395, which produces antifungal metabolites with bafilomycin derivatives, including bafilomycin C1-amide, was isolated from a soil sample at Mt. Jiri, South Korea. Here, we report its draft genome sequence, which contains 8.04 Mb with 73.6% G+C content and 7,810 protein-coding genes.

Received 28 May 2014 Accepted 6 June 2014 Published 19 June 2014

Citation Hwang JY, Kim SH, Oh HR, Cho Y-J, Chun J, Chung YR, Nam DH. 2014. Draft genome sequence of *Kitasatospora cheerisanensis* KCTC 2395, which produces plecomacrolide against phytopathogenic fungi. Genome Announc. 2(3):e00604-14. doi:10.1128/genomeA.00604-14.

Copyright © 2014 Hwang et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 3.0 Unported license

Address correspondence to Doo Hyun Nam, dhnam@ynu.ac.kr.

The genus *Kitasatospora* for actinomycete strains was first suggested by Omura et al. (1). Members of this genus possess different contents of *meso*-diaminopimelic acid and galactose in whole-cell lysate from the closely related members of the *Streptomyces* genus. After a period of debate, the genus *Kitasatospora* was revived by Zhang et al. (2), based on the distinct phylogenetic clades in the 16S rRNA genes as well as a 16S-23S rRNA gene spacer. More recently, the gene for the RNA polymerase  $\beta$  subunit, in addition to the 16S rRNA gene sequence, was employed for the phylogenetic classification of *Kitasatospora* was reported by Ichikawa et al. (4), who determined the complete genome sequence of *Kitasatospora setae* NBRC 14216, which produces bafilomycin B1 (setamycin), belonging to the plecomacrolide group.

*K. cheerisanensis* YC75 (KCTC 2395) was isolated from a soil sample at Cheeri-San (Mt. Jiri) in the process of screening biological control agents for a phytopathogenic fungus (5). Later, the antifungal metabolites produced by this strain were confirmed as bafilomycin derivatives, including bafilomycin C1-amide (6).

The genome sequence of *K. cheerisanensis* was obtained with a combination of an Illumina GAIIx 100-bp paired-end library (971.18× coverage), a Roche 454 Titanium 8-kb paired-end library (9.12× coverage), a PacBio 5-kb library (29.68× coverage), and a PacBio 10-kb library (84.06× coverage). The procedure for library construction was performed according to the manufacturers' instructions.

Illumina and PacBio sequencing data were assembled with CLC Genomic Workbench 6.5 (CLCbio, Denmark) and PacBio SMRT Analysis 2.0 using the HGAP2 protocol (Pacific Biosciences, USA). Resulting contigs were scaffolded using GS Assembler 2.6 (Roche Diagnostics, CT). The final assembly provided a total of 5 scaffolds containing 178 contigs. The draft genome of *K. cheerisanensis* consists of 8,035,179 bp, with a 73.6% G+C content. A total of 7,810 coding sequences (CDSs) with 9 rRNA operons and 72 tRNA genes were predicted.

The CDSs were predicted using Glimmer 3.02 (7), and tRNA and rRNA were searched using tRNAscan-SE and HMMER with

ezTaxon-e database bacterial rRNA profiles (8–10). The annotation of each CDS was made by homology search against NCBI reference sequence (RefSeq), Clusters of Orthologous Groups (COG), SEED, CatFam, SMART 6.2, PRINTS 42.0, TIGRFAM 13.0, Pfam 27.0, and InterPro 44.0 databases (11–18).

Several gene clusters for the biosynthesis of secondary metabolites were found in the genome, including type I polyketide synthase (PKS) gene clusters (KCH\_04080 to KCH\_04120 for the biosynthesis of the bafilomycin backbone and KCH\_45030 to KCH\_45040), a type II PKS gene cluster (KCH\_73510 to KCH\_73550), nonribosomal peptide synthetase (NRPS) gene clusters (KCH\_06280 to KCH\_06390, KCH\_45400 to KCH\_45410, KCH\_61460 to KCH\_61470, and KCH\_70790 to KCH\_70800), and PKS/NRPS hybrid gene clusters (KCH\_67350 to KCH\_67370 and KCH\_74020 to KCH\_74040).

Genes for resistance to  $\beta$ -lactam antibiotics, including AmpC  $\beta$ -lactamase (KCH\_10470 and KCH\_10480) and metallo- $\beta$ -lactamase (KCH\_19220 and KCH\_36670), were identified. The other putative resistance genes for aminoglycoside antibiotics (KCH\_00980 and KCH\_00990) and chloramphenicol (KCH\_72240 and KCH\_73860) were also found in this draft genome.

**Nucleotide sequence accession numbers.** This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession number JNBY00000000. The version described in this paper is version JNBY01000000.

## ACKNOWLEDGMENT

This work was supported by the General Researcher Program grant NRF-2010-0005904 from the Korea Research Fund.

## REFERENCES

- 1. Omura S, Takahashi Y, Iwai Y, Tanaka H. 1982. *Kitasatosporia*, a new genus of the order *Actinomycetales*. J. Antibiot. 35:1013–1019. http://dx.doi.org/10.7164/antibiotics.35.1013.
- Zhang Z, Wang Y, Ruan J. 1997. A proposal to revive the genus *Kitasatospora* (Omura, Takahashi, Iwai, and Tanaka 1982). Int. J. Syst. Bacteriol. 47:1048–1054. http://dx.doi.org/10.1099/00207713-47-4-1048.
- 3. Kim BJ, Kim CJ, Chun J, Koh YH, Lee SH, Hyun JW, Cha CY, Kook

YH. 2004. Phylogenetic analysis of the genera *Streptomyces* and *Kitasatospora* based on partial RNA polymerase  $\beta$ -subunit gene (*rpoB*) sequences. Int. J. Syst. Evol. Microbiol. 54:593–598. http://dx.doi.org/10.1099/ ijs.0.02941-0.

- 4. Ichikawa N, Oguchi A, Ikeda H, Ishikawa J, Kitani S, Watanabe Y, Nakamura S, Katano Y, Kishi E, Sasagawa M, Ankai A, Fukui S, Hashimoto Y, Kamata S, Otoguro M, Tanikawa S, Nihira T, Horinouchi S, Ohnishi Y, Hayakawa M, Kuzuyama T, Arisawa A, Nomoto F, Miura H, Takahashi Y, Fujita N. 2010. Genome sequence of *Kitasatospora setae* NBRC 14216<sup>T</sup>: an evolutionary snapshot of the family *Streptomycetaceae*. DNA Res. 17:393–406. http://dx.doi.org/10.1093/dnares/dsq026.
- Chung YR, Sung KC, Mo HK, Son DY, Nam JS, Chun J, Bae KS. 1999. *Kitasatospora cheerisanensis* sp. nov., a new species of the genus *Kitasato-spora* that produces an antifungal agent. Int. J. Syst. Bacteriol. 49(Pt 2): 753–758. http://dx.doi.org/10.1099/00207713-49-2-753.
- Moon SS, Hwang WH, Chung YR, Shin J. 2003. New cytotoxic bafilomycin C1-amide produced by *Kitasatospora cheerisanensis*. J. Antibiot. 56:856-861. http://dx.doi.org/10.7164/antibiotics.56.856.
- Delcher AL, Bratke KA, Powers EC, Salzberg SL. 2007. Identifying bacterial genes and endosymbiont DNA with Glimmer. Bioinformatics 23:673–679. http://dx.doi.org/10.1093/bioinformatics/btm009.
- 8. Eddy SR. 1998. Profile hidden Markov models. Bioinformatics 14: 755–763. http://dx.doi.org/10.1093/bioinformatics/14.9.755.
- Kim OS, Cho YJ, Lee K, Yoon SH, Kim M, Na H, Park SC, Jeon YS, Lee JH, Yi H, Won S, Chun J. 2012. Introducing EzTaxon-e: a prokaryotic 16S rRNA gene sequence database with phylotypes that represent uncultured species. Int. J. Syst. Evol. Microbiol. 62:716–721. http://dx.doi.org/ 10.1099/ijs.0.038075-0.
- Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. Nucleic Acids Res. 25: 955–964. http://dx.doi.org/10.1093/nar/25.5.0955.
- Finn RD, Mistry J, Tate J, Coggill P, Heger A, Pollington JE, Gavin OL, Gunasekaran P, Ceric G, Forslund K, Holm L, Sonnhammer EL, Eddy SR, Bateman A. 2010. The Pfam protein families database. Nucleic Acids Res. 38:D211–D222. http://dx.doi.org/10.1093/nar/gkp985.

- 12. Hunter S, Jones P, Mitchell A, Apweiler R, Attwood TK, Bateman A, Bernard T, Binns D, Bork P, Burge S, de Castro E, Coggill P, Corbett M, Das U, Daugherty L, Duquenne L, Finn RD, Fraser M, Gough J, Haft D, Hulo N, Kahn D, Kelly E, Letunic I, Lonsdale D, Lopez R, Madera M, Maslen J, McAnulla C, McDowall J, McMenamin C, Mi H, Mutowo-Muellenet P, Mulder N, Natale D, Orengo C, Pesseat S, Punta M, Quinn AF, Rivoire C, Sangrador-Vegas A, Selengut JD, Sigrist CJ, Scheremetjew M, Tate J, Thimmajanarthanan M, Thomas PD, Wu CH, Yeats C, Yong SY. 2012. Interpro in 2011: new developments in the family and domain prediction database. Nucleic Acids Res. 40:D306–D312. http://dx.doi.org/10.1093/nar/gkr948.
- 13. Overbeek R, Begley T, Butler RM, Choudhuri JV, Chuang HY, Cohoon M, de Crécy-Lagard V, Diaz N, Disz T, Edwards R, Fonstein M, Frank ED, Gerdes S, Glass EM, Goesmann A, Hanson A, Iwata-Reuyl D, Jensen R, Jamshidi N, Krause L, Kubal M, Larsen N, Linke B, McHardy AC, Meyer F, Neuweger H, Olsen G, Olson R, Osterman A, Portnoy V, Pusch GD, Rodionov DA, Rückert C, Steiner J, Stevens R, Thiele I, Vassieva O, Ye Y, Zagnitko O, Vonstein V. 2005. The subsystems approach to genome annotation and its use in the project to annotate 1000 genomes. Nucleic Acids Res. 33:5691–5702. http://dx.doi.org/10.1093/nar/gki866.
- Pruitt KD, Tatusova T, Klimke W, Maglott DR. 2009. NCBI reference sequences: current status, policy and new initiatives. Nucleic Acids Res. 37:D32–D36. http://dx.doi.org/10.1093/nar/gkn721.
- Tatusov RL, Galperin MY, Natale DA, Koonin EV. 2000. The COG database: a tool for genome-scale analysis of protein functions and evolution. Nucleic Acids Res. 28:33–36. http://dx.doi.org/10.1093/nar/28.1.33.
- Yu C, Zavaljevski N, Desai V, Reifman J. 2009. Genome-wide enzyme annotation with precision control: catalytic families (CatFam) databases. Proteins 74:449–460. http://dx.doi.org/10.1002/prot.22167.
- 17. Letunic I, Doerks T, Bork P. 2012. SMART 7: recent updates to the protein domain annotation resource. Nucleic Acids Res. 40:D302–D305. http://dx.doi.org/10.1093/nar/gkr931.
- Attwood TK, Beck ME, Bleasby AJ, Parry-Smith DJ. 1994. PRINTS—a database of protein motif fingerprints. Nucleic Acids Res. 22:3590–3596.