

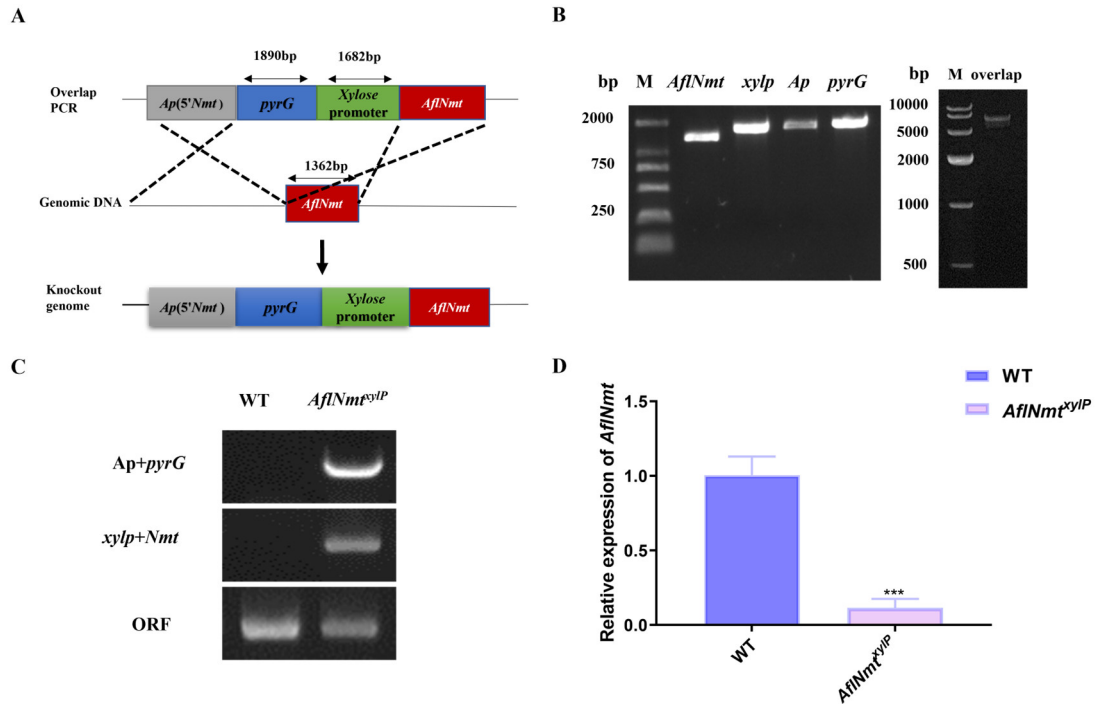


Figure S1. Construction of *AflNmt*^{xylP} strain. (A) Strategy and schematic diagram of the construction of *AflNmt*^{xylP} strain. The native promoter of *AflNmt* was replaced with the xylose-inducible promoter by using homologous recombination method in the *AflNmt*^{xylP} strain. Ap (5'*Nmt*): flanking regions upstream of *AflNmt* gene; *pyrG*: nutritional marker; *Xylose* promoter (*xylp*): xylose-inducible promoter. (B) Left panel: the PCR result of *AflNmt*, *xylp*, Ap and *pyrG* fragments. *AflNmt*: *AflNmt* gene fragments; *xylp*: xylose-inducible promoter segment; Ap: flanking regions upstream of *AflNmt* gene; *pyrG*: nutritional marker segment. Right panel: these four fragments were fused by overlapping extension PCR; M: standard marker for molecular weight. (C) The homologous recombination transformant was verified by PCR. (D) The fluorescent quantitative analysis of Nmt expression levels in WT and *AflNmt*^{xylP} strains. *** means significant difference level ($P < 0.001$) based on t tests with three biological replicates.

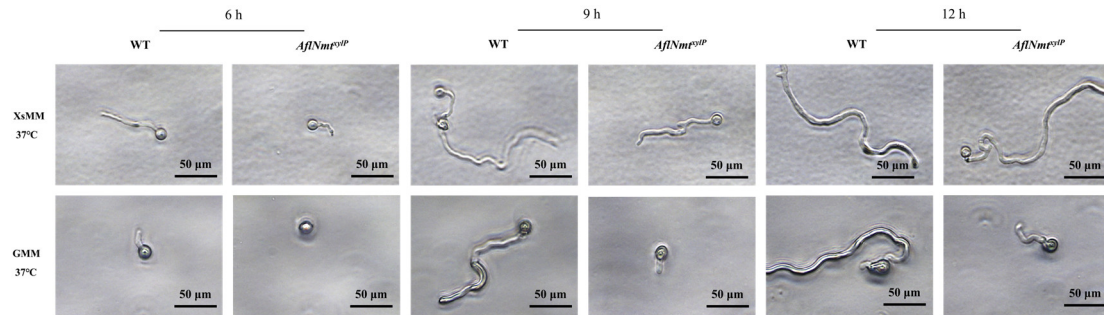


Figure S2 Microscopic observation of conidia germination for WT and *AflNmt*^{xyLP} strains. The results indicated that *AflNmt*^{xyLP} strains delayed germination at 37°C in GMM 和 XsMM liquid culture medium. The scale bar is 50 μm.

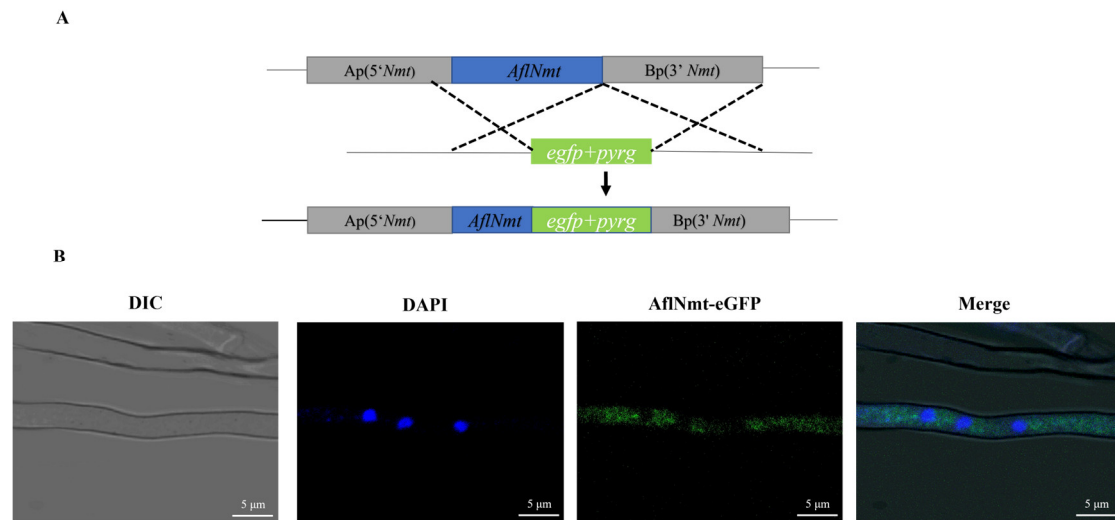


Figure S3 *AflNmt* is localized in the cytoplasm. (A) Strategy and schematic diagram of the construction of *AflNmt*-eGFP strain. Ap (5'*Nmt*): flanking regions upstream of *AflNmt* gene; egfp+pyrG: genome segment of enhanced green fluorescent protein and nutritional marker; Bp (3'*Nmt*): flanking regions downstream of *AflNmt* gene. (B) *AflNmt*-eGFP fusion protein was localized in the cytoplasm. The blue dye (DAPI) was used as a nucleus dye to locate nuclei. The scale bar is 5 μm.

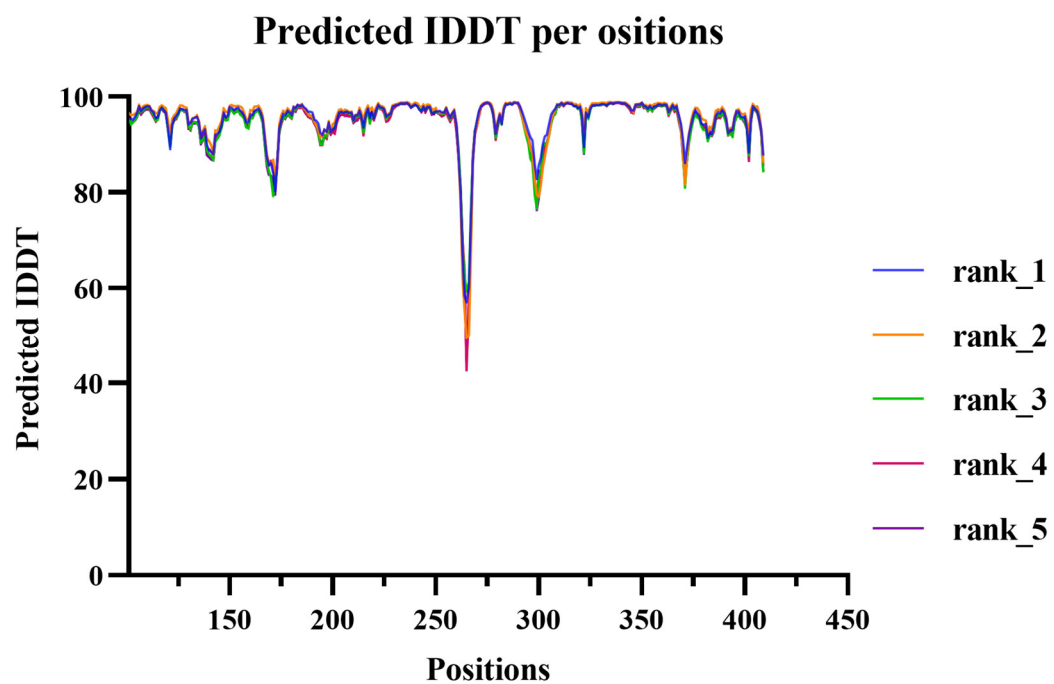


Figure S4. The value of Local Distance Difference Test (IDDT) for the five best prediction models was given by AlphaFold2.

Table S1 Strains used in this study

Strain	Genotype description	Reference
<i>A. flavus</i> CA14 PTs	$\Delta ku70$, $\Delta pyrG$	(1)
wild type (WT)	$\Delta ku70$, $\Delta pyrG::pyrG$	this study
<i>AflNmt</i> ^{xyIP}	$\Delta ku70$, $\Delta pyrG$, $\Delta AflNmt::pyrG::xyIP::AflNmt$	this study
<i>AflNmt</i> -eGFP	$\Delta ku70$, $\Delta pyrG$, $\Delta AflNmt::pyrG::AflNmt::eGFP$	this study
rAflNmt	<i>pET-32a-AflNmt</i>	this study

Table S2 Composition of culture media

Medium	Composition
XsMM	10 g/L xylose, 1 mol/L ammonium tartrate, 6 g/L NaNO ₃ , 0.52 g/L MgSO ₄ •7H ₂ O, KH ₂ PO ₄ 1.52 g/L, KCl 0.52 g/L, 1 mL/L trace elements
YGT	5 g/L yeast extract, 20 g/L glucose, and 1 mL/L trace elements
YXsT	5 g/L yeast extract, 20 g/L xylose, and 1 mL/L trace elements
GMM	10 g/L glucose, 1 mol/L ammonium tartrate, 6 g/L NaNO ₃ , 0.52 g/L MgSO ₄ •7H ₂ O, KH ₂ PO ₄ 1.52 g/L, KCl 0.52 g/L, and 1mL/L trace elements
Trace element (100ml)	2.2 g ZnSO ₄ •7H ₂ O, 1.1 g H ₃ BO ₃ , 0.5 g MnCl ₂ •4H ₂ O, 0.5 g FeSO ₄ •7H ₂ O, 0.17 g CoCl ₂ •5H ₂ O, 0.16 g CuSO ₄ •5H ₂ O, 0.005 g (NH ₄) ₆ Mo ₇ O ₂₄ •5H ₂ O, 4.45g Na ₂ EDTA

Table S3 Synthesized primers used in this study

Primers Name	Primers Sequence (5'-3')	Application
<i>nmt</i> -F1	TTGATGAAGCGGCTCGTGT	for <i>AflNmt</i> ^{xyIP} strain construction
<i>nmt</i> -R3	TGAAGAGCATTGTTTGAAGCGGGTTGA	
<i>xyIP</i> -F6	GTGCGGAAGGTTT	
<i>xyIP</i> -R7	GCATCAGTGCCTCCTCTCAGACCTCGAG	
<i>nmt</i> -F8	GTTCGACGGAAGCG	
<i>nmt</i> -R10	GTTGGTTCTTCGAGTCGATGAATG	
	CATCGACTCGAAGAACCAACATGTCGGAC	
	CCTAAGGATACCAA	
	AGGCATCGTTCATCAGGAGTAGA	

<i>nmt</i> -F2	GGTTCACCTCCCAGATACA	
<i>nmt</i> -R9	TCGTGGACGGCGTCAATAT	
<i>pyrG</i> -F4	GCCTCAAACAATGCTCTTCACCC	
<i>pyrG</i> -R5	GTCTGAGAGGAGGCACTGATGC	
<i>nmt</i> -ORF-F	CAGACACAACCCGTACCCCGT	
<i>nmt</i> -ORF-R	TCACAACATAACAACGCCAAC	
<i>nmt</i> -eGFP-F1	GTCGGACCCTAAGGATACCAA	
<i>nmt</i> -eGFP-R3	GGCTCCAGCGCCTGCACCAGCTCCCAACAT AACAACGCCAACACC	
eGFP-F4	GGAGCTGGTGCAGGCGCTGGAGCCATGGTGA GCAAGGGCGAGGA	for <i>AflNmt</i> - <i>eGFP</i> strain construction
eGFP-R5	GGGTGAAGAGCATTGTTTGAGGCTTACTTGTA CAGCTCGTCCATG	
<i>nmt</i> -eGFP -F6	GCATCAGTGCCCTCCTCTCAGACCGTCAGTCGC CGAGCAAT	
<i>nmt</i> -eGFP -R8	CGCCCTTCGTTCAAGTCAG	
<i>nmt</i> -eGFP -F2	TGGTCTATCGGTTCGGTGGG	
<i>nmt</i> -eGFP -R7	GGGAGTCGTGCCTGAACACTT	
<i>AflsumO</i> -c-qRT-F	CTTCTTTCTTGTTCTCAGCTG	
<i>AflsumO</i> -c-qRT-R	TAGGCGACCCCGTTCC	
<i>AflsumO</i> -g-qRT-F	ATCCTTTCCGTTCTCTGGC	
<i>AflsumO</i> -g-qRT-R	AGATTGTCAGCTCTGGTGC	for gene copy number identification
<i>Aflnmt</i> -c-qTR-F	CAGACACAACCCGTACCCCGT	
<i>Aflnmt</i> -c-qRT-R	TCACAACATAACAACGCCAAC	
<i>Aflnmt</i> -g-qRT-F	CGCAGGAAAGGAGCA	
<i>Aflnmt</i> -g-qRT-R	GACCGTTGATGGGAGG	
<i>brlA</i> -F	GCCTCCAGCGTCAACCTTC	
<i>brlA</i> -R	TCTCTTCAAATGCTCTTGCCTC	
<i>nsdC</i> -F	GCCAGACTTGCCAATCAC	
<i>nsdC</i> -R	CATCCACCTTGCCCTTTA	
<i>nsdD</i> -F	GGACTTGCGGGTCGTGCTA	
<i>nsdD</i> -R	AGAACGCTGGGTCTGGTGC	for cDNA- qRT-PCR
<i>aflR</i> -F	AAAGCACCCTGTCTTCCCTAAC	
<i>aflR</i> -R	GAAGAGGTGGGTCAAGTGTGTTGTAG	
<i>aflS</i> -F	CGAGTCGCTCAGGCGCTCAA	
<i>aflS</i> -R	GCTCAGACTGACCGCCGCTC	
<i>Afnmt</i> -OF	CAGACACAACCCGTACCCCGT	
<i>Afnmt</i> -OR	TCACAACATAACAACGCCAAC	
<i>actin</i> -F	ACGGTGTCGTCACAACTGG	reference gene

