## CASE REPORT



# Case report: Coxiella burnetii vascular infection and lymphoma in the Netherlands

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## Abstract

Objectives and design Non-Hodgkin lymphoma has been linked to infection with *Coxiella burnetii*, potentially through overproduction of IL-10 during infection with C. burnetii.

Materials and methods Description of a case report. Results We describe a patient with retroperitoneal non-Hodgkin lymphoma and vascular infection with C. burnetii. Immunofluorescence staining and fluorescence in situ hybridization targeting specific C. burnetii 16S rRNA were performed on the retroperitoneal lymphoma tissue sample obtained at diagnosis of NHL. Both were strongly positive for the presence of C. burnetii.

Conclusions This case provokes questions regarding a potential association between C. burnetii and NHL, and underlines the importance of further exploration of this association.

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**Keywords** *Coxiella burnetii* · Q fever · Non-Hodgkin lymphoma

# Case report

A 58-year-old male patient, with a history of a vascular bypass because of occlusion of the infrarenal aorta in September 2010 and rheumatoid arthritis for which he used etanercept weekly, underwent abdominal ultrasound during routine follow-up in September 2012. He mentioned lower abdominal pain since a few weeks, but did not report any night sweats, fever, weight loss, rash or other complaints. He did not have any contact with animals, but lived in the Netherlands in an area where Q fever had been highly epidemic between 2007 and 2010 [1]. Laboratory results on presentation are shown in Table 1. On abdominal ultrasound, bilateral hydronephrosis with extensive retroperitoneal masses was observed. A subsequent computed tomography scan and positron emission tomography (PET) scan showed extensive retroperitoneal, intra-abdominal, mediastinal, cervical, and axillary lymphadenopathy, a pulmonary mass and a lesion in the right adrenal. In addition, the abdominal aorta and left iliac artery showed increased <sup>18</sup>F-FDG uptake, which was not further analyzed. The PET-scan was made as part of the standard work-up procedure of suspected NHL, in accordance with Dutch guidelines [2, 3]. Biopsy of the retroperitoneal masses and a bone marrow biopsy revealed an Ann Arbor stage IV B cell non-Hodgkin lymphoma (NHL), with initially indefinite histopathological classification of the subtype of NHL. Re-examination of the pathology specimens at an academic hospital confirmed the presence of a B-cell NHL, with a marginal zone lymphoma as the most likely histopathological subtype. Chemo-immunotherapy



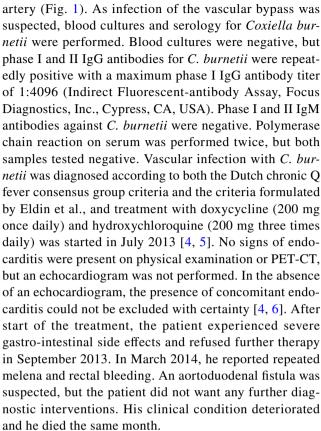
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Table 1 Laboratory results at the day of presentation

Laboratory measurement	Result	Normal range	Units of measure- ment
Hemoglobin	7.2	8.5–11.0	mmol/L
Thrombocytes	179	150-400	$10^{9}/L$
Leukocytes <sup>a</sup>	9.2	4.0-10.0	$10^{9}/L$
Creatinine <sup>a</sup>	551	60-110	μmol/L
C-reactive protein	14	0–8	mg/L
Aspartate aminotransferase <sup>a</sup>	19	0-34	U/L
Alanine aminotransferase <sup>a</sup>	13	0-44	U/L
$Gamma-glutamyltransferase^{a} \\$	<10	0-54	U/L
Alkaline phosphatase <sup>a</sup>	81	43–115	U/L
Lactate dehydrogenase <sup>a</sup>	246	0-247	U/L

 $^{\rm a}$ In the days following initial presentation, these laboratory values changed. Maximum values during admission were: leukocyte count  $35.6 \times 10^9$ /L, C-reactive protein 186 mg/L, aspartate aminotransferase 35, alanine aminotransferase 49, alkaline phosphatase 177, gamma-glutamyltransferase 146, lactate dehydrogenase 400

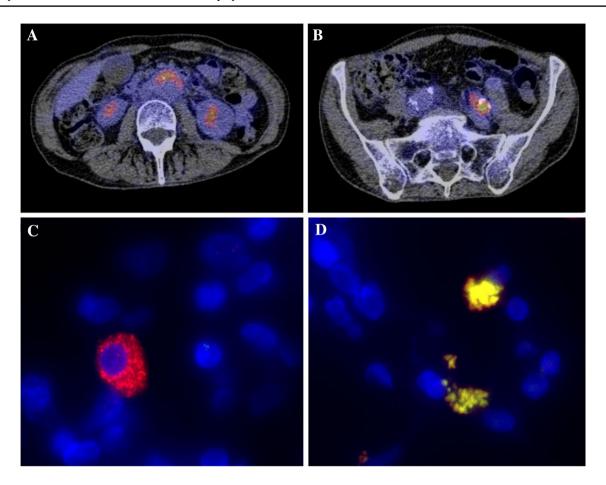
(rituximab, cyclophosphamide, doxorubicin, vincristine, and prednisone) was initiated and etanercept was discontinued. After discontinuation of etanercept, the patient experienced two flares of his rheumatoid arthritis for which he received short courses of prednisone. Followup PET scan after three cycles of chemotherapy showed regression of all lymphoma locations, with the exception of pulmonary lesion, adrenal mass, and lower cervical lymph nodes. The increased <sup>18</sup>F-FDG uptake in aorta and left iliac artery was unchanged. Because of unresponsiveness to chemotherapy of the pulmonary lesion, a second malignancy, possibly metastatic lung carcinoma, was suspected and further analysis was planned. Meanwhile, chemotherapy was discontinued, while awaiting analysis of the suspected lung lesion. Shortly after the last PET scan, the patient was diagnosed with pulmonary embolism for which heparin was started. Because of the use of anticoagulants and difficult accessibility of the pulmonary lesion, the diagnosis could not be confirmed pathologically. A follow-up PET scan was performed in February 2013, on which all NHL localizations were persistently in regression. However, the <sup>18</sup>F-FDG uptake in the aorta and left iliac artery had strongly increased and the pulmonary lesion and pathological cervical lymph nodes were unchanged. The adrenal lesion was slightly larger. Patient was referred for second opinion and after due consideration did not want any further diagnostics and treatment of his pulmonary lesion. Analysis of the increased uptake in the aorta and left iliac artery was performed in June 2013. Repeated PET scanning revealed highly increased uptake of <sup>18</sup>F-FDG of the abdominal aorta and left iliac artery, and a lesion suspicious of a small abscess near the left iliac



We performed immunofluorescence staining (IF) and fluorescence in situ hybridization (FISH) targeting specific C. burnetii 16S rRNA on the retroperitoneal lymphoma tissue sample obtained at diagnosis of NHL in 2012, which were both strongly positive, Fig. 1. We refer to a previous article for technical details on IF and FISH for C. burnetii [7]. FISH is a relatively novel diagnostic method for detection of C. burnetii, with few studies using FISH targeting C. burnetii performed. The technique was applied in one clinical study and one case-report only [7, 8]. Melenotte et al. were able to detect C. burnetii in four out of seven lymphoma biopsies with FISH. All samples had positive immunofluorescence as confirmation, but negative immunohistochemistry [7]. Both FISH and IF are highly sensitive new diagnostic techniques to detect bacteria in situ, that may be superior to immunohistochemistry.

The presence of *C. burnetii* in the NHL tissue indicates that the infection was already present at the time of diagnosis of NHL in September 2012. At the moment of presentation in September 2012, our patient did not report any weight loss or fever. Nevertheless, he developed clear signs of inflammation shortly after admission (see Table 1). A potential explanation for the absence of fever at presentation may be the fact that our patient was immunocompromised. Absence of weight loss may be due to the fact that self-reporting of symptoms is not very accurate. In the Dutch national chronic Q fever database with data of 439 patients with persistent or





**Fig. 1** Composite figure of PET-CT, immunofluorescence (IF) and fluorescence in situ hybridization (FISH). **a** Highly increased <sup>18</sup>F-FDG uptake in the aortic wall. **b** Highly increased <sup>18</sup>F-FDG uptake in the left iliac artery. **c** Microscopic image (original magnification ×100) of immunofluorescence staining (IF) of retroperitoneal lymphoma tissue of a patient with vascular chronic Q fever in which nuclei are stained *blue* (4',6-diamidino-2-phenylindole, DAPI), while perinuclear *Coxiella burnetii* is stained *red*. **d** Microscopic

image (original magnification ×100) of fluorescence in situ hybridization (FISH) of the same tissue in which nuclei are stained *blue* (4',6-diamidino-2-phenylindole, DAPI), while *C. burnetii*, organized in perinuclear vacuoles, is stained *yellow*. *Yellow* signal results of the co-localization of the universal probe EUB (*red*) and specific 16S rRNA *C. burnetii* probe (*green*). For both **c**, **d** Leica DMI6000 B microscope was used

chronic Q fever from the Netherlands, only 14% of patients had fever and only 28% of patients reported weight loss at presentation [9]. The absence of these symptoms illustrates that patients with a chronic infection can present with atypical symptoms.

It has been hypothesized that there is a causal relationship between persistent infection with *C. burnetii* and development of B-cell NHL [7]. Our patient presents similar as the index case of the series reporting a link between Q fever and B cell lymphoma: both patients had a vascular focus of infection and lymphoma located closely to the focus of infection [7]. A potential pathophysiological pathway is overproduction of IL-10 during infection with *C. burnetii*, which could play a role in the development of B-cell NHL [7, 10]. However, both diseases have a considerable diagnostic delay. It cannot be ruled out that *C. burnetii* infects monocytes and macrophages in tumorous tissue and that this patient

had developed NHL before infection. Furthermore, both diseases have common risk factors, such as immunocompromised state [11, 12]. Our patient was severely immunocompromised, which is a risk factor for development of both the lymphoma and the vascular infection with *C. burnetii*.

In this report, we describe a case of NHL after Q fever. Naturally, no hard conclusions with regard to the association between *C. burnetii* and NHL and its causality can be drawn based on one single case. However, this case provokes further questions regarding the potential association between *C. burnetii* infection and NHL and its causality and potential diagnostic tools for detection of *C. burnetii* in tissues.

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Author contribution SER: performance of IF and FISH, writing of manuscript. CM: performance of IF and FISH, writing of manuscript. MHAH: providing of histopathological material, writing of manuscript. HAMS: obtaining informed consent, providing clinical details, writing of manuscript. PTGAN: providing of histopathological material, writing of manuscript. GA: performance of IF and FISH. AIMH: writing of manuscript. JJO: supervision of SER, writing of manuscript. D. Raoult: supervision of CM, performance of IF and FISH, writing of manuscript. PCW: supervision of SER, writing of manuscript.

#### Compliance with ethical standards

**Ethical standards statement** The conduct of this study was approved by the Medical Ethical Committee of Brabant, the Netherlands, and the Science Bureau of the Jeroen Bosch Hospital, the Netherlands. Furthermore, the patients' widow granted us permission to publish this report.

Conflict of interest S. E. van Roeden: institution received research grant from foundation Q-support and Institut Mérieux, no other conflicts of interest. I confirm that we had full access to all data in the report and that we have final responsibility for the decision to submit this manuscript for the publication at Infection. No other conflicts of interest. C. Melenotte: no conflicts of interest. M. H. A. Hermans: no conflicts of interest. H. A. M. Sinnige: no conflicts of interest. P. T. G. A. Nooijen: no conflicts of interest. G. Audoly: no conflicts of interest. A. I. M. Hoepelman: institution received research grant from foundation Q-support and Institut Mérieux, no other conflicts of interest. J. J. Oosterheert: institution received research grant from foundation Q-support and Institut Mérieux, no other conflicts of interest. D. Raoult: no conflicts of interest. P. C. Wever: no conflicts of interest.

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