

Supplementary file

Applied Microbiology and biotechnology

**Effects of *hap2* deletion on *mnp/vp* transcription in *Pleurotus ostreatus* grown on
lignocellulosic substrates**

**Keita Kayama, Takehito Nakazawa*, Iori Yamaguchi, Moriyuki Kawauchi, Masahiro
Sakamoto, Yoichi Honda**

Graduate School of Agriculture, Kyoto University, Sakyo-ku, Kyoto 606-8502, Japan

*Corresponding author: Takehito Nakazawa

Graduate School of Agriculture, Kyoto University, Oiwakecho, Kitashirakawa,

Sakyo-ku, Kyoto 606-8502, Japan

Tel.: +81 75 753 6465

Fax: +81 75 753 6471

E-mail: nakazawa.takehito.8u@kyoto-u.ac.jp

Running Title: Functional analysis of *P. ostreatus* hap2

Table S1 The composition of lignocellulose-based media used in this study.

Media	Composition
BWS-I	1.9 g extracted ^a beech wood sawdust, 0.1 g extracted ^a wheat bran, and 6.0 ml water
BWS-II	1.9 g size-fractionated beech wood sawdust (250–500 µm), 0.1 g wheat bran, and 6.0 ml water
Avicel-I	1.9 g Avicel ^b , 0.1 g extracted ^a wheat bran, and 6.0 ml water
Holocellulose-I	1.9 g holocellulose ^c , 0.1 g extracted ^a wheat bran, and 6.0 ml water

^aSolvent extraction was performed as described by Nakazawa et al. (2023a).

^bAvicel is a crystalline cellulose with an average degree of 200.

^cHolocellulose was prepared by the Jayme-Wise method (Green 1963) as described by Nakazawa et al. (2023a).

Table S2 Primers used for GST-pulldown, *hap2* deletion, and EMSA

Primer	Sequence (5'→3')
KYK136	GTGATTGGCCGGCATCAGCG
KYK137	CGCTGATGCCGGCCAATCAC
M13R	ACAATTTACACAGGAAACAGCTATGACC
Selex_R	GCACGATGACAGCATACTCTAAG
Selex_Rdm_comp	NNNNNNNNNNNNNNNNNNNNNGCTTAGAGTATGCTGTCATCGT GC
TN48	TGGCTGTTGACTTATCGTGGCG
TN378	TCACAAGTTAACAGCCACGGATTAAGC
TN400	TCCCAGTCACGACGTCAGGCAAAACCCAGATACGC
TN991	TCGGGGTCCTGCATGCTAGCG
TN992	ACGTCGTGACTGGGATCTATGGGGGCCTCGTCTGG
TN993	CCTGTGTGAAATTGTATCGACCCTACTGCCCCGCC
TN994	TCCCACCGCCGGACGTTTAC
TN995	ATCGACAGGGGCTGACCTCTTCTG
TN996	AGCGCGACAGCAATGTAGGG
TN997	AGCAGTTCCACCCGCATCAG
TN998	TGGCTGCATGTTTATTGGTGG
TN1097	CGGCCGCATCGTGACCCAGACGAGGCCCCCATAGAC
TN1098	GCAGATCGTCAGTCAAGTAGGGTCGATGCCCGCTG
TN1523	CATATGTATATCTCCTTATTGTTTC
TN1524	CATACCAAGCGGGCCCTGAAAC
TN1526	TAATGACGTACCATGGTAAAGTCTACACC
TN1528	GGAGATATACATATGATGTCCGATAGCCTCCAG
TN1529	CATGGTACGTCATTATTCTTCATCTAACGACTCTAGC
TN1531	GGAGATATACATATGTCGTCCAGCAAGCCTTTCGTC
TN1532	CATGGTACGTCATTAAAGAAGGGATGTTATTCCGTTCTG
TNS12	GTGATTGGCCGGCATCAGCGCCTGTAGCGTGTTCCGGCAACTT
TNS13	GTGAAGCGCCGGCATCAGCGCCTGTAGCGTGTTCCGGCAACTT

60

61

Table S3 Primers used for the real-time PCR and digital PCR.

Target gene (Protein ID ^a)	Sequence (5' - 3')	Amplification efficiency (%)
<i>β-tub</i> ^b	GTGCGTAAGGAAGCTGAGGG TGTGGCATTGTACGGCTCAAC	98
<i>vp1</i> ^b	TTGTTGGCTAGAGACCCCCAGA CAAGTGGGCCGCTCCGAC	83.5
<i>vp2</i> ^b	GCCTTTCGATAGCGTGGATAAG GGTCCCTTGCAACATTGTCTC	95.1
<i>vp3</i> ^b	CAGGCCGTCAATGCCGTA CGTAAGGACGGAACCATCA	104.7
<i>mnp1</i> ^b	GTTCGCTCGAGACGATAGGACAT GGGGAGATGTGGTTTGGTTACA	95.2
<i>mnp2</i> ^b	ATTGCTAATGAAAGGCTCATGGTT GGTGGCAGCACAAGCAG	102.9
<i>mnp3</i> ^b	CCTCCTGACTTTGGCATCTCA TGTCGTCGATTCCACCGTTG	82.5
<i>mnp4</i> ^b	CTTCATCTCCAACCCCAACTC GCTTTGCGGCAGGATG	107.1
<i>mnp5</i> ^b	TTCCCAGGTACTGCCGGA CCAAAGACAGTTTCAACATCGC	117.6
<i>mnp6</i> ^b	CTTCGACTCTACTCCCAACAGC GAGGCGTGGTCGGTGAT	94

62

^aA genomic fragment containing the genes that corresponds to each Protein ID were from the

63

genome database of strain PC9 (JGI *Pleurotus ostreatus* PC9 v1.0,

64

https://genome.jgi.doe.gov/PleosPC9_1/PleosPC9_1.home.html).

65

^bPrimers designed in Salame et al. (2012b)

66

67

68

69

70

71

72

73

74
75
76
77
78
79
80
81
82
83
84
85
86
87
88
89
90
91
92
93
94
95
96
97

Table S4 Probes used for digital PCR.

Gene ^a	Sequence (5' - 3') ^b	PCR condition ^c
<i>β-tubulin</i>	TGCTGGTATGGGTACACTCCTGAT	A
<i>vp1</i>	CAGTCCATGGTTAACAACCAGC	A
<i>vp2</i>	ACCGTTGAAGTCGTCTGGCTGC	A
<i>vp3</i>	AGCGAATAGGCGCAATGCATTTC	A
<i>mnp1</i>	ACCTTCAGCAGAACCTCTTCGACG	A
<i>mnp2</i>	CAGGACGCCATGATCGATTG	A
<i>mnp3</i>	AGCCACCCTTGACAAAGTCC	B
<i>mnp4</i>	CTCTTTGATGGTGCCGAGTGTGG	A
<i>mnp5</i>	AACCCAAAGAAGCTGATTGACTGC	B
<i>mnp6</i>	CGAGGAGAGATGCGGCTTCAGTC	A

^a Probes for the indicated genes were originally designated by Nakazawa et al. (2023b).
^b The synthesized oligonucleotides are modified with 6-FAM (5'), ZEM quencher (internal) and Iowa Black Dark quencher (3').
^c The PCR condition was as follows: one cycle at 95°C (10 min) for initial denaturation, 40 cycles at 95°C (30 sec) and 58°C (A) or 61°C (B) (60 sec), followed by denature step at 95°C (10 min). Ramp Rate was 2°C/sec.

98

99

Table S5 Promoter analysis of *G. subvermispora*

Gene ^a	protein ID ^b	TATAA-box ^{c, d}	CCAAT-box ^{c, e, f}
<i>gp</i>	112162	-	-
<i>lip1</i>	99382	-	160(-)
<i>lip2</i>	118677	67	167(-)
<i>mnp1</i>	116608	90	149(-)
<i>mnp2</i>	50297	95	-
<i>mnp3</i>	139965	98, 89	148(-)
<i>mnp4</i>	94398	83	163(-)
<i>mnp5</i>	49863	93	175(-)
<i>mnp6</i>	50686	95	297(+)
<i>mnp7</i>	105539	75	230(-), 146(-), 106(-)
<i>mnp8</i>	114036	91	161(-)
<i>mnp9</i>	114076	-	-
<i>mnp10</i>	117436	94	238(+), 172(-), 46(+)
<i>mnp11</i>	143390	78	149(-)
<i>mnp12</i>	157986	-	-
<i>mnp13</i>	124076	76	186(-)
<i>lcs1</i>	118801	-	-
<i>lcs2</i>	88089	-	-
<i>lcs3</i>	108852	-	75(-)
<i>lcs4</i>	137686	76	-
<i>lcs5</i>	115068	83	201(+)
<i>lcs6</i>	115063	74	243(+)
<i>lcs7</i>	84170	82	240(+)

^{a, b} Gene names and protein ID are based on the genome database of *Gelatoportia subvermispora*

B (<https://mycocosm.jgi.doe.gov/Cersu1/Cersu1.home.html>)

^c The distances of the TATAA-box and CCAAT-box from the start codon are shown.

^d Only TATAA-boxes located within 100 bp from the start codon are shown.

^e CCAAT-boxes located within 300 bp from the start codon are shown.

^f (+) indicates the presence of CCAAT on the coding strand, while (-) on the template strand.

107

108

Table S6 Promoter analysis of *P. chrysosporium*

Gene ^a	protein ID ^b	TATAA-box ^{c, d}	CCAAT-box ^{c, e, f}
<i>lipA</i>	2989894	81	180(-)
<i>lipB</i>	1716776	77	258(+)
<i>lipC</i>	3032409	81	-
<i>lipD</i>	1386770	88	192(-)
<i>lipE</i>	1716042	77	-
<i>lipF</i>	2910310	74	166(-)
<i>lipG</i>	2918435	79	-
<i>lipH</i>	2918661	71	-
<i>lipI</i>	3032425	84	188(-)
<i>lipJ</i>	3043032	71	193(-)
<i>mnp1</i>	2971944	90	191(-), 205(-)
<i>mnp2</i>	3589	87	236(-)
<i>mnp3</i>	2896529	-	-
<i>mnp4</i>	8191	93	194(-), 208(-)
<i>mnp5</i>	2907883	-	19(+), 153(-)

^{a, b} Gene names and protein ID are based on the genome database of *Phanerochaete*

chrysosporium RP-78 (<https://mycocosm.jgi.doe.gov/Phchr2/Phchr2.home.html>)

^c The distances of the TATAA-box and CCAAT-box from the start codon are shown.

^d Only TATAA-boxes located within 100 bp from the start codon are shown.

^e CCAAT-boxes located within 300 bp from the start codon are shown.

^f (+) indicates the presence of CCAAT on the coding strand, while (-) on the template strand.

115

Table S7 Homology search against CBC (CCAAT-binding complex) components of *Aspergillus nidulans*.

Query (<i>Aspergillus nidulans</i>)		1st Hit (<i>Pleurotus ostreatus</i>)				2nd Hit (<i>Pleurotus ostreatus</i>)			
Protein name	NCBI Accession no.	NCBI Accession no.	Query cover	Percent Identification	E-value	NCBI Accession no.	Query cover	Percent Identification	E-value
HapB	XP_050467982.1	XP_036630745.1	26%	62.63%	3.00E-28	-	-	-	-
HapC	XP_661638.1	XP_036626823.1	42%	73.12%	2.00E-47	XP_036633301.1	48%	33.96%	6.00E-17
HapE	XP_664096.2	XP_036633731.1	39%	65.71%	7.00E-47	-	-	-	-

Table S8 Reciprocal Blastp for identification of CBC (CCAAT-binding complex) components.

Query (<i>Pleurotus ostreatus</i>)		1st Hit (<i>Aspergillus nidulans</i>)				2nd Hit (<i>Aspergillus nidulans</i>)			
Protein name	NCBI Accession no.	NCBI Accession no.	Query cover	Percent Identification	E-value	NCBI Accession no.	Query cover	Percent Identification	E-value
Putative Hap2	XP_036630745.1	XP_050467982.1	22%	64.21%	3.00E-28	-	-	-	-
Putative Hap3	XP_036626823.1	XP_661638.1	61%	71.13%	2.00E-47	XP_050468969.1	47%	38.46%	9.00E-12
Putative Hap5	XP_036633731.1	XP_664096.2	56%	65.71%	6.00E-47	-	-	-	-

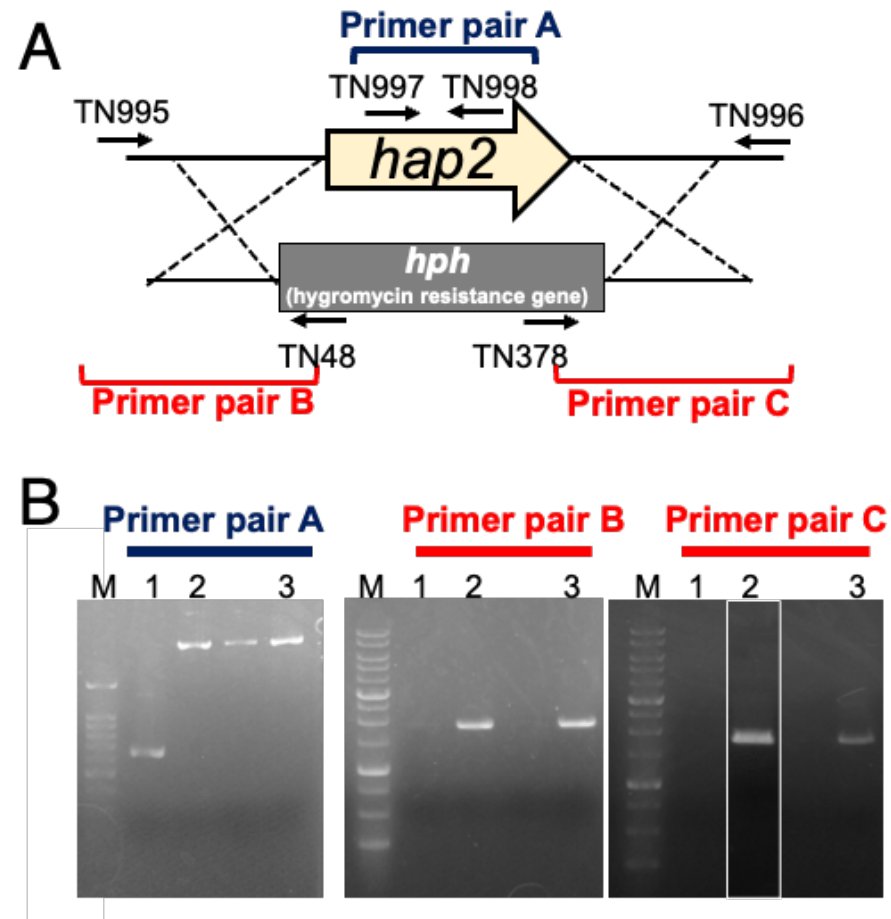


Fig. S1 Generation of the *hap2* deletants ($\Delta hap2\#1$ and $\Delta hap2\#2$) derived from 20b using homologous recombination. (A) A diagram of the genomic locus of *Pleurotus ostreatus hap2*. Black arrows indicate the primers used for the polymerase chain reaction (PCR) amplification. (B) Genomic PCR experiments confirming the *hap2* deletion in the deletants. Primers used in each PCR experiment are shown in (A).

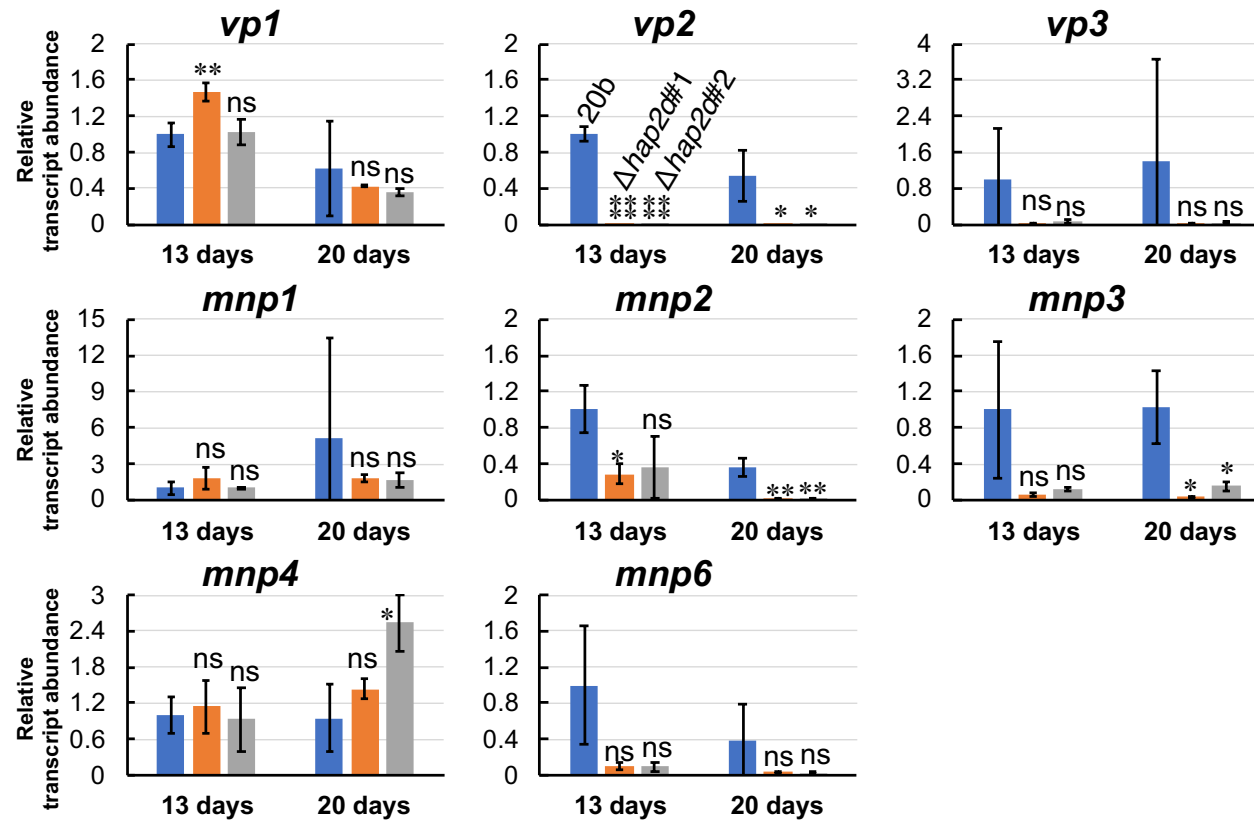


Fig. S2 Relative transcriptional expression levels of the eight *mnp/vp* genes in the *hap2* deletants grown on Holocellulose-I at 13- and 20-day culture periods ($n = 3$). Expression levels were standardized by β -tubulin, and the expression level of 20b at 13 days is shown as 1. Graphs indicate mean values and bars indicate standard deviations. Statistical significance tests between the indicated two strains at the respective culture periods were performed using a two-tailed equal variance *t*-test. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$. “ns” indicates statistical non-significance ($p > 0.05$).

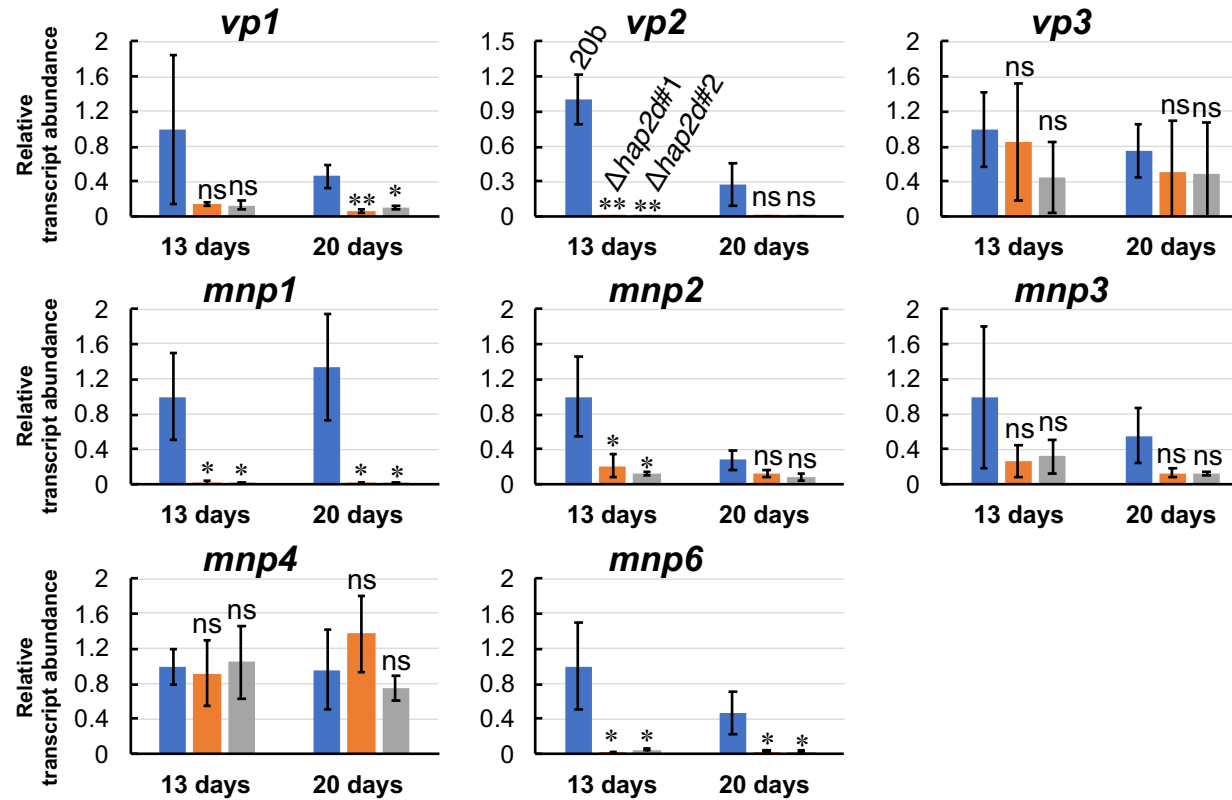


Fig. S3 Relative transcriptional expression levels of the eight *mnp/vp* genes in the *hap2* deletants grown on Avicel-I at 13- and 20-day culture periods (n = 3). Expression levels were standardized by β -*tubulin*, and the expression level of 20b at 13 days is shown as 1. Graphs indicate mean values and bars indicate standard deviations. Statistical significance tests between the indicated two strains at the respective culture periods were performed using a two-tailed equal variance *t*-test. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$. “ns” indicates statistical non-significance ($p > 0.05$).

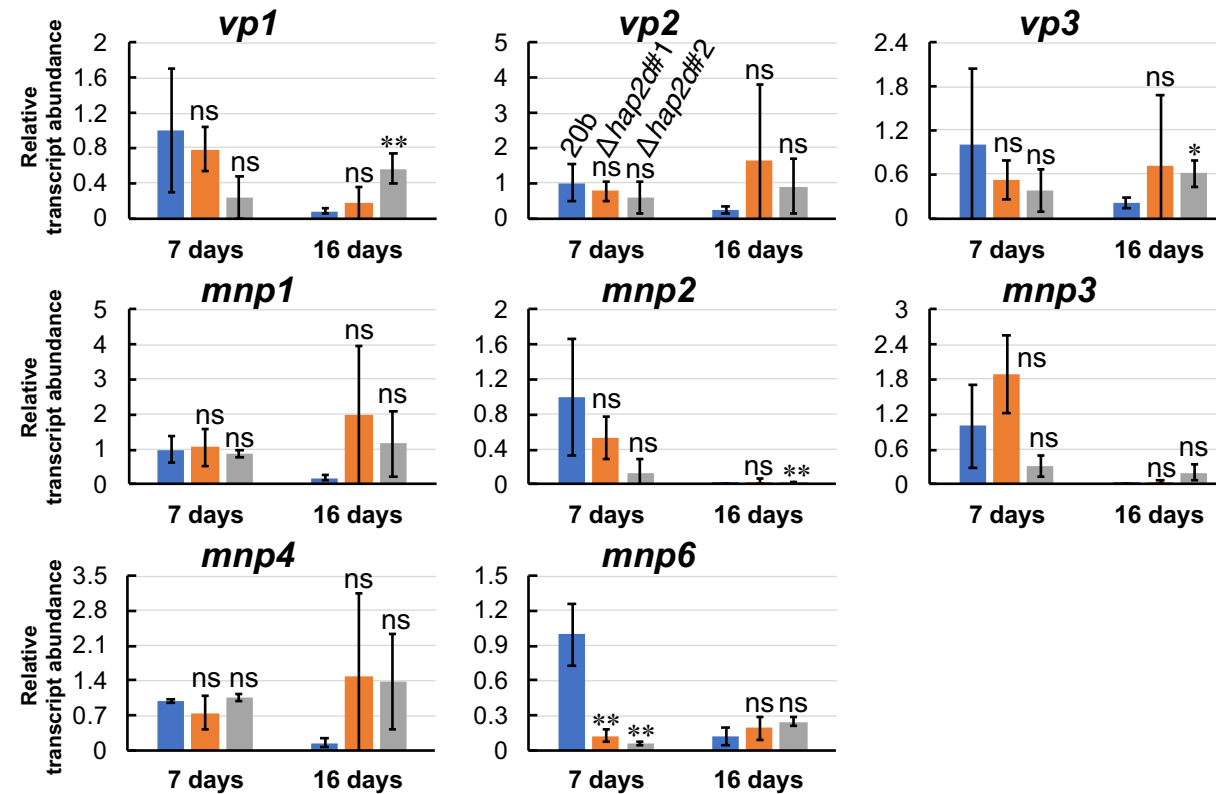


Fig. S4 Relative transcriptional expression levels of the eight *mnp/vp* genes in the *hap2* deletants grown on YMG agar plates at 7- and 16-day culture periods (n = 3). Expression levels were standardized by β -tubulin, and the expression level of 20b at 7 days is shown as 1. Graphs indicate mean values and bars indicate standard deviations. Statistical significance tests between the indicated two strains at the respective culture periods were performed using a two-tailed equal variance *t*-test. **p* < 0.05, ***p* < 0.01, ****p* < 0.001. “ns” indicates statistical non-significance (*p* > 0.05).