AZGP1 in POMC neuron modulates energy homeostasis and metabolism through leptin-mediated STAT3 phosphorylation

Authors

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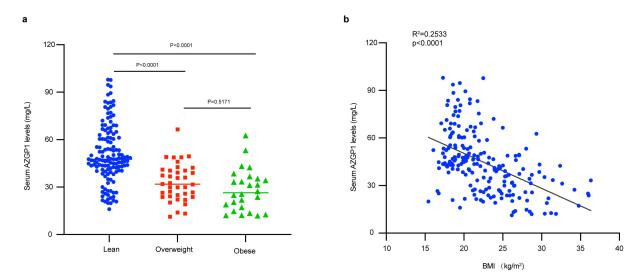
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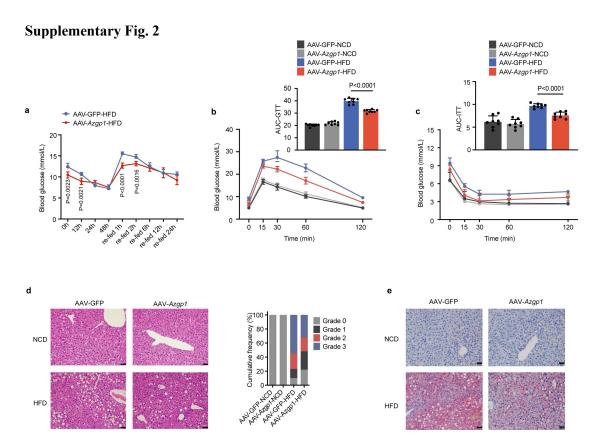
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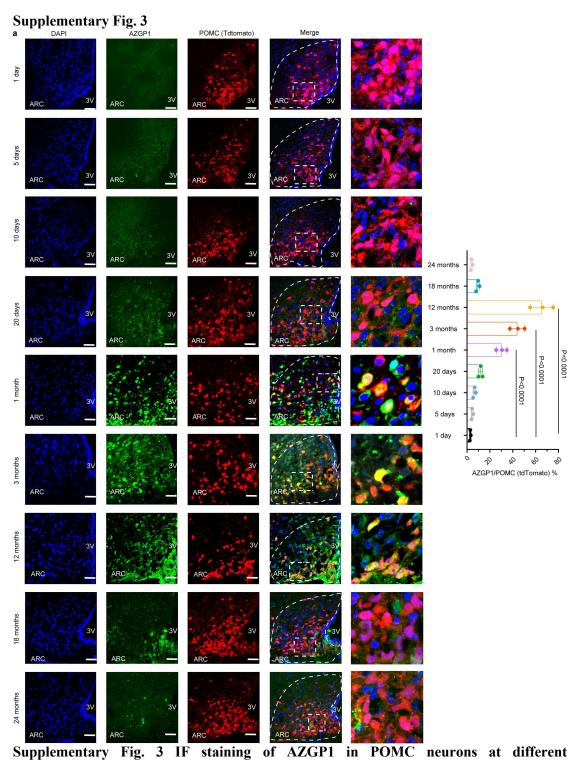
Supplementary Figures and Figure Legends



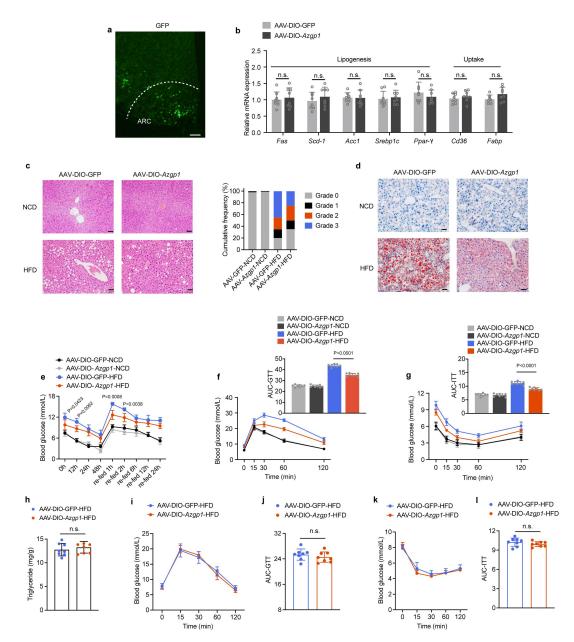
Supplementary Fig. 1 Serum AZGP1 levels are elevated in obese individuals and are associated with BMI. a Serum AZGP1 levels in lean individuals (BMI 18.5-23.9 kg/m², n = 135 independent individuals), overweight individuals (BMI 24-27.9 kg/m², n = 38 independent individuals) and obese individuals (BMI ≥ 28 kg/m², n = 26 independent individuals). b Correlation between AZGP1 levels and BMI. BMI, body mass index. The data are expressed as the mean \pm SD. One-way ANOVA followed by Tukey's test (a); pearson correlation analysis (b). Source data are provided as a Source Data file.



Supplementary Fig. 2 Effects of AZGP1 overexpression in the hypothalamus on glucose/lipid metabolism and insulin sensitivity. Eight-week-old male WT mice were injected with AAV9-Azgp1/GFP into the MBH and were fed a NCD or HFD for 12 weeks as described in the Methods. a Fasting and refeeding blood glucose levels in HFD-fed mice (n = 8 mice). b, c Blood glucose levels and the AUC of the GTT (b) and ITT (c) curves in NCD-and HFD-fed mice (n = 8 mice). d Representative H&E staining of the liver in NCD- and HFD-fed mice and quantitation of the grade of steatosis (n = 6 mice; scale bars: 50 μ m). e Oil Red O staining of the liver in NCD- and HFD-fed mice (n = 6 mice; scale bars: 50 μ m). GTT, glucose tolerance test; ITT, insulin tolerance test; AUC, area under the curve. The data are expressed as the mean \pm SEM. Statistical significance was calculated using an unpaired two-tailed t-test (a) and two-way ANOVA followed by Bonferroni's post hoc tests (b-c). Source data are provided as a Source Data file.



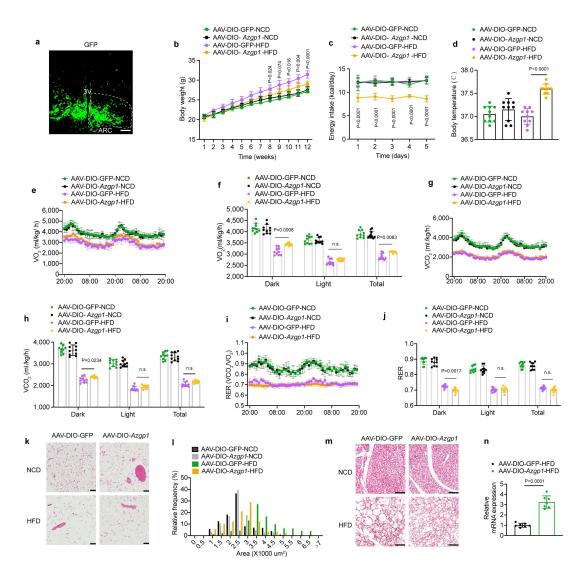
developmental stages in mice. (n = 3 mice); scale bars: 50 μ m. ARC, arcuate nucleus; 3V, third cerebral ventricle. The data are expressed as the mean \pm SEM. One-way ANOVA followed by Tukey's test (a). Source data are provided as a Source Data file.



Supplementary Fig. 4 Effects of AZGP1 overexpression in POMC neurons on glucose/

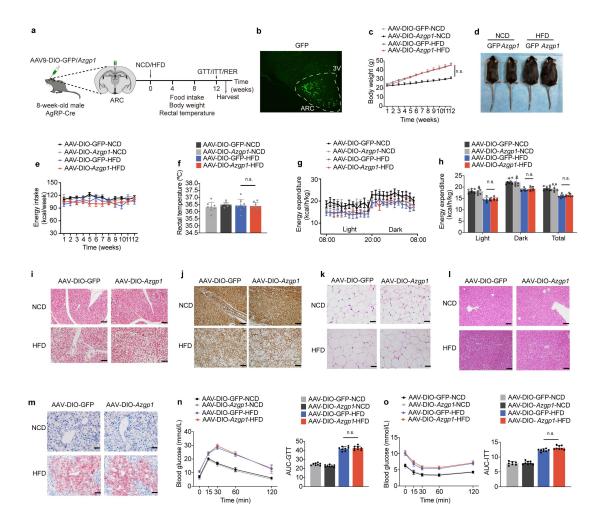
lipid metabolism. Eight-week-old male POMC-Cre mice received bilateral injections of AAV9-DIO-*Azgp1*/GFP into the MBH and were fed a NCD or HFD for 12 weeks as described in the Methods. **a** Representative IF image showing GFP expression in the mouse hypothalamus (n = 3 mice). **b** Lipid metabolism-related gene expression in eWAT in HFD-fed mice (n = 8 mice). **c** Representative H&E staining of liver sections and quantitation of the grade of steatosis (n = 5 mice; scale bars: 50 μm). **d** Oil Red O staining of representative liver sections (n = 5 mice; scale bars: 50 μm). **e** Fasting and refeeding blood glucose levels (n = 9

mice). **f, g** Blood glucose levels and the AUC of the GTT (**f**) and the ITT curves (n = 8 mice). **h-l** Eight-week-old male POMC-Cre mice were given injections of AAV9-DIO-Azgp1/GFP into the bilateral MBH and were fed a HFD for 4 weeks. (**h**) TG levels in the liver (n = 8 mice). (**i-l**) Blood glucose levels and the AUC of the GTT and ITT curves (n = 8 mice). ARC, arcuate nucleus; VMH, ventromedial nucleus; 3V, third cerebral ventricle; GTT, glucose tolerance test; ITT, insulin tolerance test; AUC, area under the curve. The data are expressed as the mean \pm SEM. Two-tailed Student's t tests were used in (**b**), (**h-l**), and two-way ANOVA with Bonferroni post-hoc tests were used in (**e-g**). Source data are provided as a Source Data file. (n.s. not significant.).



Supplementary Fig. 5 Specific overexpression of AZGP1 in POMC neurons increases energy expenditure and ameliorates metabolic disorders in HFD-fed female mice. Eight-week-old female POMC-Cre mice were injected with AAV9-DIO-*Azgp1*/GFP into the bilateral MBH and were fed a NCD or HFD for 12 weeks as described in the Methods. a Representative IF image showing GFP expression in the ARC. b Body weight curve (n = 8 mice). c Energy intake (n = 10 mice). d Rectal temperature (n = 10 mice). e, f Forty-eight hours oxygen consumption (n = 10 mice). g, h Forty-eight hours carbon dioxide consumption (n = 10 mice). i, j Respiratory exchange ratio (RER, VCO₂/VO₂) (n = 10 mice). k Representative H&E staining image of eWAT (n = 5 mice; scale bar: 50 μm). l Cross-sectional area of eWAT quantified by ImageJ (n = 5 mice). m Representative H&E staining image of

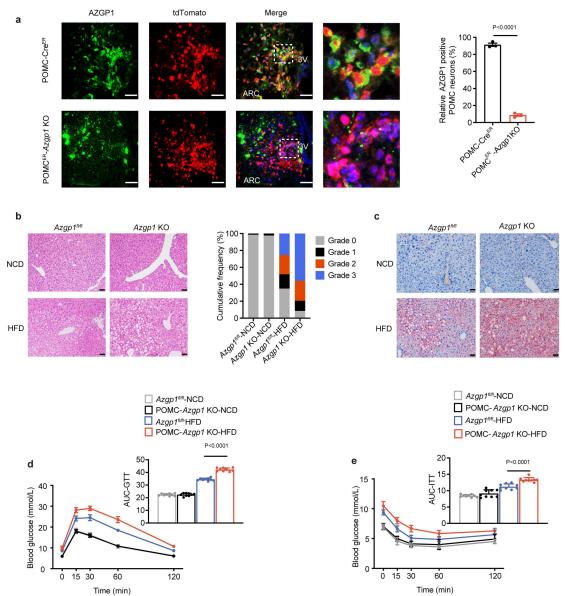
BAT (n = 5 mice; scale bar: 100 μ m). **n** The mRNA expression of *Ucp1* (n = 7 mice). ARC, arcuate nucleus; 3V, third cerebral ventricle. The data are expressed as the mean \pm SEM. Two-way ANOVA followed by Holm-Šídák multiple comparisons test (**b**, **c**; **f**, **h**, **j**); One-way ANOVA followed by Tukey's test (**d**); Two-tailed Student's t test (**n**). Source data are provided as a Source Data file. (n.s. not significant.).



Supplementary Fig. 6 Overexpression of AZGP1 in AgRP neurons did not affect energy

balance. a Schematic diagram of the experimental procedure. b IF staining image showing GFP expression in the ARC. c Body weight curve (n = 8 mice). d Representative photograph of mice. e Energy intake (n = 7 mice). f Rectal temperature (n = 8 mice). g, h Energy expenditure (n = 7 mice). i H&E staining of BAT (n = 5 mice; scale bars: 100 μm). j UCP1 immunostaining of BAT (n = 5 mice; scale bars: 100 μm). k H&E staining of eWAT (n = 5 mice; scale bars: 50 μm). l H&E staining of the liver (n = 5 mice; scale bars: 50 μm). m Oil Red O staining of the liver (n = 5 mice; scale bars: 50 μm). n, o Blood glucose levels and the AUC of the GTT (n) and ITT (o) curves (n = 8 mice). ARC, arcuate nucleus; 3V, third cerebral ventricle; GTT, glucose tolerance test; ITT, insulin tolerance test; AUC, area under the curve. The data are expressed as the mean ± SEM; one-way ANOVA followed by Tukey's

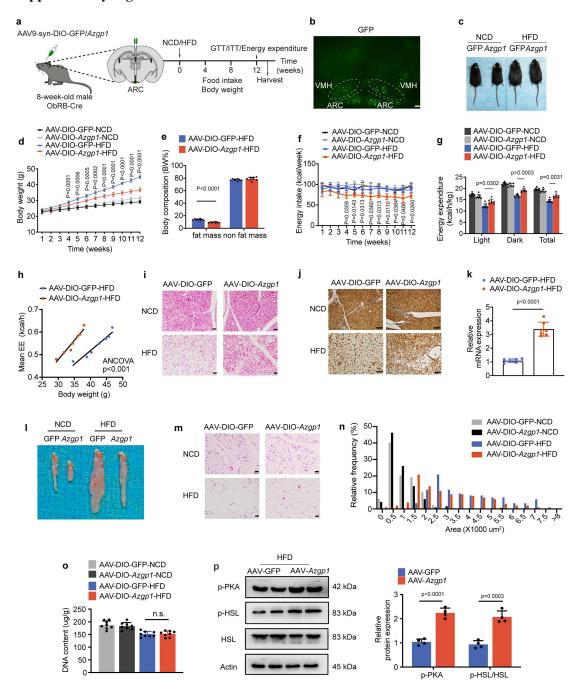
test (f), two-way ANOVA followed by Bonferroni's post hoc tests (c, e; h; n; o). Source data are provided as a Source Data file. (n.s. not significant.).



Supplementary Fig. 7 Effects of Azgp1 ablation in POMC neurons on lipid metabolism.

a IF staining of AZGP1 in the ARC POMC neurons (tdTomato) of POMC-*Azgp1* KO or POMC-Cre^{ER} mice (n = 3 mice; scale bars: 50 µm). **b**, **c** POMC-*Azgp1* KO and *Azgp1*^{fl/fl} mice were fed a NCD or HFD for 12 weeks. H&E staining of liver sections and quantitation of the grade of steatosis (**b**) (n = 5 mice; scale bars: 50 µm). **c** Oil Red O staining of liver sections (n = 5 mice; scale bars: 50 µm). **d**, **e** Blood glucose levels and the AUC of the GTT (**d**) and the ITT (**e**) curves (n = 9 mice). ARC, arcuate nucleus; 3V, third cerebral ventricle; GTT, glucose tolerance test; ITT, insulin tolerance test; AUC, area under the curve. The data are expressed as the mean \pm SEM. Two-tailed Student's t tests were used in (**a**) and two-way ANOVA with

Bonferroni post-hoc tests were used in (d), (e). Source data are provided as a Source Data file.



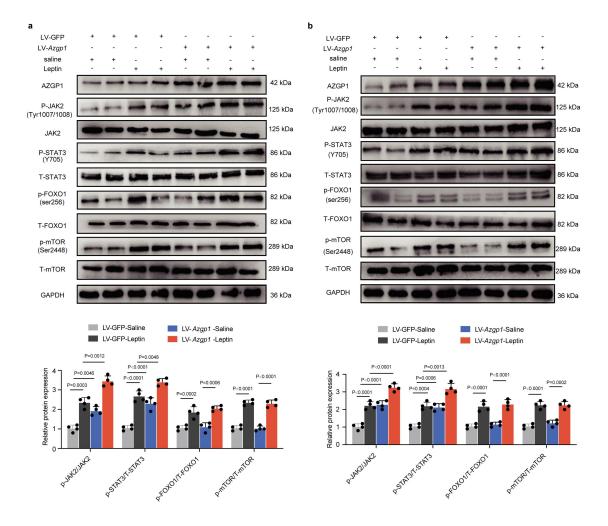
Supplementary Fig. 8 Overexpression of AZGP1 in ObRb neurons promotes energy expenditure in HFD-fed mice. a Schematic diagram of the experimental procedure. **b** IF staining of GFP expression in the ARC and VMH (n = 3 mice; scale bars: 200 μm). **c** A representative photograph of mice. **d** Body weight curve (n = 7 mice). **e** Body composition (n = 8 mice). **f** Energy intake (n = 5 mice). **g** Energy expenditure (n = 7 mice). **h** ANCOVA of energy expenditure versus body weight in HFD-fed mice (n = 7 mice). **i** Representative H&E staining images of BAT (n = 5 mice; scale bars: 50 μm). **j** UCP1 immunostaining in BAT (n =

5 mice; scale bars, 100 μm). **k** *Ucp1* mRNA expression in BAT (n = 7 mice). **l** Representative image of eWAT depots. **m** Representative H&E staining image of eWAT (n = 5 mice; scale bars: 50 μm). **n** Cross-sectional area of eWAT quantified by ImageJ (n = 5 mice; scale bars: 50 μm). **o** DNA content in eWAT (n = 8 mice). **p** Western blot analysis of p-PKA and p-HSL/HSL expression in eWAT of HFD-fed mice and densitometric quantification (n = 4 mice). ARC, arcuate nucleus; VMH, ventromedial nucleus. The data are expressed as the mean ± SEM. Two-tailed Student's t tests were used in (**e**, **k**, **o**, **p**), one-way ANCOVA using body weight as covariate (**h**) and two-way ANOVA with Bonferroni post-hoc tests were used in (**d**, **f**, **g**,). Source data are provided as a Source Data file. (n.s. not significant.).

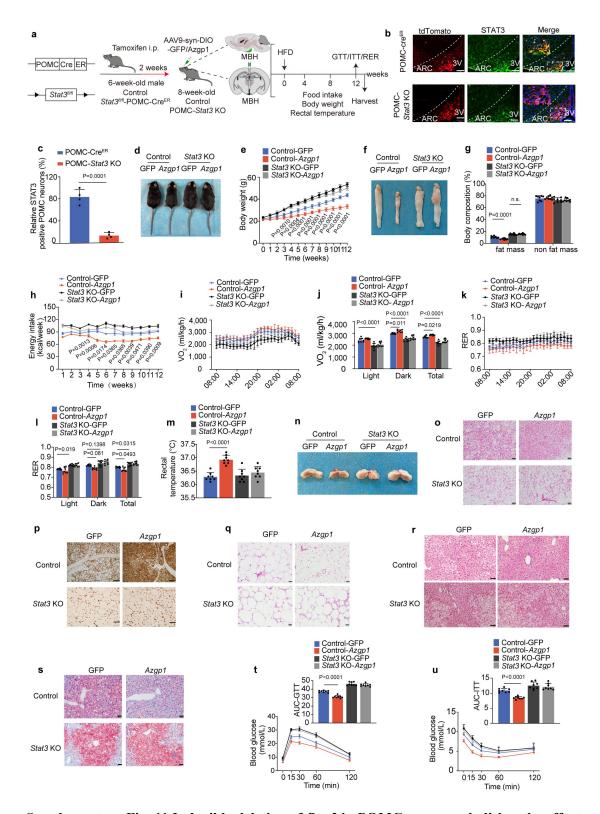
Supplementary Fig. 9 ■ AAV-DIO-GFP-NCD ■ AAV-DIO-Azgp1-NCD ■ AAV-DIO-GFP-HFD ■ AAV-DIO-Azgp1-HFD AAV-DIO-GFP-NCD AAV-DIO-*Azgp1*-NCD AAV-DIO-GFP-HFD AAV-DIO-*Azgp1*-HFD ■Grade 0 ■ Grade 2 ■Grade 1 ■ Grade 3 AAV-DIO-GFP AAV-DIO-Azap1 AAV-DIO-GFP AAV-DIO-Azap1 Blood glucose (mmol/L) 모 60 60 Time (min) 120 0 15 30 DAPI AZGP1 Merae DAPI AZGP1 cFos Merge AAV-GFP AAV-GFP aCSF aCSF AAV-Azgp1 AAV-Azgp1 AAV-GFP AAV-GFP Leptin Leptin AAV-Azgp1 AAV-Azgp1 AZGP1/POMC colocalization (No. of cells) 0 0 0 0 0 0 0 of cells) AAV/GEP+aCSE AAV-GFP+aCSF AAV-Azgp1+aCSF AAV-GFP+Leptin g 50

AAV-Azgp1+aCSF AAV-GFP+Leptin AAV-Azgp1+Leptin

Supplementary Fig. 9 Effects of AZGP1 overexpression in ObRb neurons on glucose/ lipid metabolism and the excitability of neurons. Eight-week-old male ObRb-Cre mice were injected with AAV9-DIO-Azgp1/GFP into the MBH and fed a NCD or HFD for 12 weeks. a Representative H&E staining of liver sections and quantitation of the grade of steatosis (n = 5 mice; scale bars: 50 μ m). **b** Oil Red O staining of the liver sections (n = 5 mice; scale bars: 50 μm). c, d Blood glucose levels and the AUC of the GTT (c) and ITT (d) curves (n = 9 mice). e, f IF staining for Azgp1 and POMC expression (e), and cFos (f) in the ARC of HFD-fed mice with or without MBH leptin (n = 3 mice; Scale bars: 50µm). ARC, arcuate nucleus; 3V, third cerebral ventricle; GTT, glucose tolerance test; ITT, insulin tolerance test; AUC, area under the curve. The data are expressed as the mean ± SEM. Two-way ANOVA with Bonferroni post-hoc tests were used in (c-f). Source data are provided as a Source Data file.

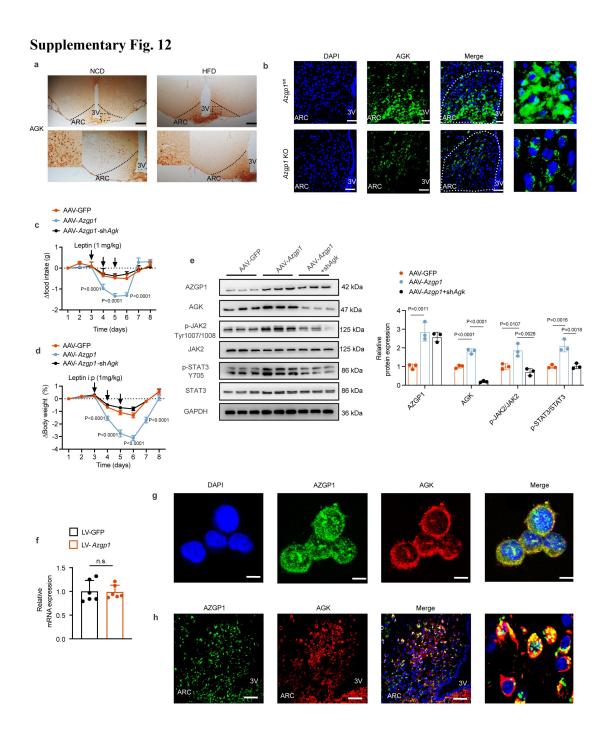


Supplementary Fig. 10 Overexpression of AZGP1 increases leptin-mediated STAT3 phosphorylation *in vitro.* GT1-7 and N2A cells were transfected with LV-*Azgp1*/GFP and treated with or without leptin as described in the Methods section. **a, b** Western blot analysis of the expression of metabolism-related signaling pathway components in GT1-7 (**a**) and N2A (**b**) cells (n = 4 independent cell experiments). The data are expressed as the mean ± SEM. Two-way ANOVA with Bonferroni post-hoc tests were used in (**a, b**). Source data are provided as a Source Data file.



Supplementary Fig. 11 Inducible deletion of *Stat3* in POMC neurons abolishes the effect of central AZGP1 on energy metabolism. a Schematic diagram of the experimental procedure. b IF images showing STAT3 in POMC neurons (tdTomato) in the ARC (n = 4

mice; scale bars: 100 μm). **c** Densitometric quantification of STAT3 expression in **(b)**. **d** Representative photograph of mice. **e** Body weight curve (n = 8 mice). **f** Representative images of eWAT depots. **g** Body composition (n = 8 mice). **h** Energy intake (n = 8 mice). **i**, **j** Twenty-four hours oxygen consumption (VO₂) (n = 8 mice). **k**, **l** Respiratory exchange ratio (RER: VCO₂/VO₂) (n = 8 mice). **m** Rectal temperature (n = 8 mice). **n** Representative images of BAT depots. **o** H&E staining of BAT (n = 5 mice; scale bars: 50 μm). **p** UCP1 immunostaining of BAT (n = 5 mice; Scale bars: 100 μm). **q** H&E staining of eWAT (n = 5 mice; scale bars: 50 μm). **r** H&E staining of the liver (n = 5 mice; scale bars: 100 μm). **s** Oil Red O staining of the liver (n = 5 mice; scale bars: 50 μm). **t**, **u** Blood glucose levels and the AUC of the GTT (t) and ITT (**u**) curves (n = 8 mice). 3V, third cerebral ventricle; ARC, arcuate nucleus; GTT, glucose tolerance test; ITT, insulin tolerance test; AUC, area under the curve. The data are expressed as the mean ± SEM. Two-tailed Student's t tests were used in (**c**); and two-way ANOVA with Bonferroni post-hoc tests were used in (**e**, **g**, **h**, **j** l, **t**, **u**). one-way ANOVA followed by Tukey's test (**m**). Source data are provided as a Source Data file.



Supplementary Fig. 12 AGK expression is downregulated by HFD in the hypothalamus and is associated with AZGP1. a IHC staining of AGK expression in the hypothalamic ARC of NCD- or HFD-fed WT mice (n = 3 mice; Scale bars: $100\mu m$). b IF staining showing AGK expression in the hypothalamic ARC of HFD-fed $Azgp1^{fl/fl}$ and POMC-Azgp1 KO mice (n = 3 mice; Scale bars: $100\mu m$). c-e Eight-week-old male WT mice received MBH injection of AAV9-Azgp1/ GFP or AVV9-Azgp1+AVV9-shAgk and were fed a HFD for 4 weeks. For the

leptin sensitivity experiment, mice were intraperitoneally injected with leptin. **c** Changes in food intake (n = 10 mice). **d** Change in body weight (n = 10 mice). **e** Western blots showing AZGP1, AGK, t-JAK2/p-JAK2 (Tyr1007/1008), and t-STAT3/p-STAT3 (Y705) expression in the hypothalamus (n = 3 mice). **f** GT1-7 cells were transfected with or without LV-*Azgp1* as indicated in the Methods. AGK mRNA expression was measured by RT-PCR (n = 6 independent cell experiments). **g** IF staining showing the co-location of AZGP1 and AGK in N2A cells (n = 3 independent cell experiments; Scale bars: 100μm). **h** IF staining showing the colocation of AZGP1 and AGK in the MBH of WT mice (n = 3 independent cell experiments; Scale bars: 100μm). 3V, third cerebral ventricle; ARC, arcuate nucleus; Data were expressed as the mean ± SEM. Two-way ANOVA with Bonferroni post-hoc tests were used in (**c**, **d**); one-way ANOVA followed by Tukey's test (**e**); Two-tailed Student's t tests were used in (**f**). Source data are provided as a Source Data file. (n.s. not significant.).

Supplemental table

Table S1 Anthropometric characteristics and metabolic indicators in the study population.

Variable	Lean	Overweight	Obesity
N (male/female)	135 (70/65)	38 (20/18)	26 (13/13)
Age (years)	25.5 ± 2.9	26.7 ± 4.0	26.54 ± 5.35
BMI (kg/m^2)	20.0 ± 1.9	$25.8 \pm 1.0^{\ (p < 0.0001)}$	$30.9 \pm 2.6~^{(p < 0.0001)}$
WHR	0.80 ± 0.05	$0.85 \pm 0.05~^{(p < 0.0001)}$	$0.90 \pm 0.05~^{(p < 0.0001)}$
FAT (%)	26.3 ± 4.9	$37.6 \pm 3.8 \ ^{(p < 0.0001)}$	$46.3 \pm 5.2 \ ^{(p < 0.0001)}$
TG (mmol/L)	0.97 ± 0.45	$1.74 \pm 1.06 \ ^{(p < 0.0001)}$	$1.65 \pm 0.91 \ ^{(p < 0.0001)}$
TC (mmol/L)	4.02 ± 1.04	4.47 ± 1.01	4.42 ± 0.98
HDL-C (mmol/L)	1.29 ± 0.59	1.29 ± 0.37	1.29 ± 0.65
LDL-C (mmol/L)	2.24 ± 0.83	$2.66 \pm 0.91 \ ^{(p=0.0104)}$	$2.66 \pm 0.47^{\ (p=0.0332)}$
FFA (mmol/L)	0.56 ± 0.26	0.61 ± 0.19	0.61 ± 0.20
FBG (mmol/L)	4.58 ± 0.51	$4.90 \pm 0.57 \ ^{(p=0.0035)}$	$4.95 \pm 0.70 \ ^{(p=0.0038)}$
0.5h-BG (mmol/L)	7.65 ± 1.60	$8.54 \pm 1.88 \ ^{(p=0.0070)}$	$9.00 \pm 1.47 \ ^{(p=0.0003)}$
1h-BG (mmol/L)	6.68 ± 1.98	$8.21 \pm 2.47 \ ^{(p=0.0003)}$	$9.90 \pm 2.41 \ ^{(p < 0.0001)}$
2h-BG (mmol/L)	5.79 ± 1.45	$7.22 \pm 2.30 \ ^{(p < 0.0001)}$	$9.32 \pm 2.45 \ ^{(p < 0.0001)}$
FIns (mU/L)	8.73 ± 4.52	$16.9 \pm 11.6^{\ (p < 0.0001)}$	$27.2 \pm 20.9 \ ^{(p < 0.0001)}$
0.5h-Ins (mU/L)	90.0 ± 63.2	$130.3 \pm 90.3 \ ^{(p=0.0025)}$	$124.8 \pm 41.4 \; ^{(p=0.0319)}$
1h-Ins (mU/L)	76.3 ± 55.1	$136.8 \pm 84.4 ^{(p < 0.0001)}$	$161.7 \pm 79.0~^{(p < 0.0001)}$
2h-Ins (mU/L)	59.6 ± 46.8	$130.3 \pm 94.2 \ ^{(p < 0.0001)}$	$182.7 \pm 91.2 \ ^{(p < 0.0001)}$
HbA1c (%)	5.23 ± 0.28	5.30 ± 0.33	$5.45 \pm 0.40 \; ^{(p=0.0020)}$

BMI, body mass index; FFA, free fatty acid; FBG, fasting blood glucose; 2h-BG, 2h-blood glucose after oral glucose tolerance test; FIns, Fasting insulin levels; 2h-Ins, 2h-insulin after oral glucose tolerance test; HDL-C, high-density lipoprotein cholesterol; LDL-C, low-density lipoprotein cholesterol; TC, total cholesterol; TG, triglyceride; WHR, waist-hip ratio. Values are given as means \pm SD. The statistical signifificance of differences were assessed by one-way ANOVA, followed by Dunnett multiple comparison test.

Table S2 The antibodies used for IF and IHC

Antibody	Source	Dilution
Anti-POMC, rabbit monoclonal	#23499, Cell Signaling Technology	1:500
Anti-AgRP, rabbit monoclonal	ab254558, Abcam	1:500
Anti-AZGP1, mouse monoclonal	sc-271957, Santa Cruz Biotechnology	1:200
Anti-cFos, mouse monoclonal	ab208942, Abcam	1:1000
Anti-AGK, rabbit polyclonal	ab137616, Abcam	1:500
Anti-NeuN, rabbit polyclonal	26975-1-AP, Proteintech	1:500
Anti-GFAP, rabbit polyclonal	16825-1-AP, Proteintech	1:500
Anti-STAT3, rabbit monoclonal	# 12640, Cell Signaling Technology	1:500
Anti-p-STAT3, rabbit monoclonal	# 9145, Cell Signaling Technology	1:500
Goat-anti-mouse IgG, polyclonal	A32723 or A32727, Invitrogen	1:1000
Goat-anti-rabbit IgG, polyclonal	A32732 or A-11034, Invitrogen	1:1000

Table S3 The antibodies used for Western blots

Antibody	Source	Dilution
Anti-AZGP1, rabbit polyclonal	orb354031, Biorbyt Ltd	1:1000
Anti-AGK, rabbit polyclonal	ab137616, Abcam	1:1000
Anti-t-STAT3, mouse monoclonal	#9139s, Cell Signaling Technology	1:1000
Anti-p-STAT3, rabbit monoclonal Anti-t-JAK2, rabbit monoclonal	#9145s, Cell Signaling Technology #3230s, Cell Signaling Technology	1:1000 1:1000
Anti-p-JAK2, rabbit polyclonal	#3771s, Cell Signaling Technology	1:1000
Anti-t-FOXO1, rabbit monoclonal	#2880s, Cell Signaling Technology	1:1000
Anti-p-FOXO1, rabbit monoclonal	#84192s, Cell Signaling Technology	1:1000
Anti-T-mTOR, rabbit monoclonal	#2983s, Cell Signaling Technology	1:1000
Anti-p-mTOR, rabbit monoclonal	#5536s, Cell Signaling Technology	1:1000
Anti-HSL, rabbit monoclonal	#18381, Cell Signaling Technology	1:1000
Anti-p-HSL, mouse polyclonal	#4137, Cell Signaling Technology	1:1000
Anti-p-PKA, rabbit monoclonal	#9624, Cell Signaling Technology	1:1000
Anti-Ub, Rabbit polyclonal	#10201-2-AP, Proteintech	1:1000
Anti-Flag, Rabbit polyclonal	#20543-1-AP, Proteintech	1:2000
Anti-Myc, Rabbit polyclonal	#16286-1-AP, Proteintech	1:2000
Anti-HA, Rabbit polyclonal	#51064-2-AP, Proteintech	1:2000
Anti-GAPDH, Rabbit polyclonal	#5174, Cell Signaling Technology	1:1000
Goat-anti-Rabbit IgG, polyclonal	#SA00001-2, Proteintech	1:8000
Goat-anti-Mouse IgG, polyclonal	#SA00001-1, Proteintech	1:8000

Table S4 The primers used for RT-PCR

Gene Name	Sequence
Pomc	R: CATTAGGCTTGGAGCAGGTC
	F: TCTTGATGATGGCGTTCTTG
Agrp	R: GGCCTCAAGAAGACAACTGC
	F: GCAAAAGGCATTGAAGAAGC
Npy	R: AAGAGCCTGGTCAAGTTCTG
1.	F: TAGGAGTAGTGCCCAAATGC
Agk	R: GCCGTCTCCTCCAGCAACAATG
8	F: AAGAAGAGCAGCCTGTCAAGAAGC
β-actin	R: GGCATAAACGCAGAGCATTCCTG
p aciii	F: CAGTGTCCATCCTCTGAGTAGC
Fas	F: CTCATCCACTCAGGTTCAG
	R: AGGTATGCTCGCTTCTCT
Scd-1	F: TGATGTTCCAGAGGAGGTA
	R: CCAGAGTGTATCGCAAGAA
Acc	F: AGCAGTTACACCACATACAT
	R: GTCATCACCATCTTCATTACC
Srebp1c	F: GCTTCTCTGCTTCTCT
	R: GCTGTAGGATGGTGAGTG
Ppar-γ	F: CCGAAGAACCATCCGATT
	R: CGCAGATCAGCAGACTC
Cd36	F: GGTCCTTACACATACAGAGT
	R: CTACAGCCAGATTCAGAACT
Fabp	F: TTCTCAGCCAGCCAGTT
_	R: CATCTCCTCGTAAGCCATT
Ppar-α	F: TGCCTTAGAACTGGATGAC
	R: ATCTGGATGGTTGCTCTG
Cptla	F: AGCCAGACGAAGAACATC
771	R: CCTTGACCATAGCCATCC
Ucpl	F: GAAACACCTGCCTCTCTCGG R: GCATTCTGACCTTCACGACCT
Adrβ3	F: TAGCCATCAAACCTGTTGAGC
	R: GGCCTCTCTAGTTCCCAG
Th	F: CTCTCCTCGAATACCACAGCC
	R: CCAAGGTTCATTGGACGGC