

RESEARCH ARTICLE

Characterization of a Single-Stranded DNA-Binding-Like Protein from *Nanoarchaeum equitans*—A Nucleic Acid Binding Protein with Broad Substrate Specificity

Marcin Olszewski^{1*}, Jan Balsewicz¹, Marta Nowak¹, Natalia Maciejewska¹, Anna Cyranka-Czaja², Beata Zalewska-Piątek¹, Rafał Piątek¹, Józef Kur¹

1 Gdańsk University of Technology, Chemical Faculty, Department of Molecular Biotechnology and Microbiology, 80–233, Gdańsk, Poland, **2** University of Wrocław, Faculty of Biotechnology, Department of Protein Engineering, 50–138, Wrocław, Poland

* molszewski@pg.gda.pl



OPEN ACCESS

Citation: Olszewski M, Balsewicz J, Nowak M, Maciejewska N, Cyranka-Czaja A, Zalewska-Piątek B, et al. (2015) Characterization of a Single-Stranded DNA-Binding-Like Protein from *Nanoarchaeum equitans*—A Nucleic Acid Binding Protein with Broad Substrate Specificity. PLoS ONE 10(5): e0126563. doi:10.1371/journal.pone.0126563

Academic Editor: Sergey Korolev, Saint Louis University, UNITED STATES

Received: February 9, 2012

Accepted: April 3, 2015

Published: May 14, 2015

Copyright: © 2015 Olszewski et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This study was partially supported by the Polish State Committee for Scientific Research grants N302 439439 to J.K. and N302 116636 to M.O. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript. No additional external funding received for this study.

Competing Interests: The authors have declared that no competing interests exist.

Abstract

Background

SSB (single-stranded DNA-binding) proteins play an essential role in all living cells and viruses, as they are involved in processes connected with ssDNA metabolism. There has recently been an increasing interest in SSBs, since they can be applied in molecular biology techniques and analytical methods. *Nanoarchaeum equitans*, the only known representative of *Archaea* phylum Nanoarchaeota, is a hyperthermophilic, nanosized, obligatory parasite/symbiont of *Ignicoccus hospitalis*.

Results

This paper reports on the *ssb*-like gene cloning, gene expression and characterization of a novel nucleic acid binding protein from *Nanoarchaeum equitans* archaeon (*NeqSSB*-like protein). This protein consists of 243 amino acid residues and one OB fold per monomer. It is biologically active as a monomer like as SSBs from some viruses. The *NeqSSB*-like protein displays a low sequence similarity to the *Escherichia coli* SSB, namely 10% identity and 29% similarity, and is the most similar to the *Sulfolobus solfataricus* SSB (14% identity and 32% similarity). The *NeqSSB*-like protein binds to ssDNA, although it can also bind mRNA and, surprisingly, various dsDNA forms, with no structure-dependent preferences as evidenced by gel mobility shift assays. The size of the ssDNA binding site, which was estimated using fluorescence spectroscopy, is 7±1 nt. No salt-dependent binding mode transition was observed. *NeqSSB*-like protein probably utilizes a different model for ssDNA binding than the SSB proteins studied so far. This protein is highly thermostable; the half-life of the ssDNA binding activity is 5 min at 100°C and melting temperature (T_m) is 100.2°C as shown by differential scanning calorimetry (DSC) analysis.

Conclusion

NeqSSB-like protein is a novel highly thermostable protein which possesses a unique broad substrate specificity and is able to bind all types of nucleic acids.

Background

Nanoarchaeum equitans is the only known representative of *Archaea* phylum Nanoarchaeota. Isolated from a submarine hot vent near Kolbeinsey island, north of Iceland, *Nanoarchaeum equitans* is a hyperthermophilic, obligate parasite/symbiont of craenarchaeon *Ignicoccus hospitalis*. The growth of the coculture of these microbes occurs between 70 and 98°C and optimally at 90°C, under strict anaerobic conditions. With a diameter of only 400 nm, *Nanoarchaeum equitans* stands out as one of the tiniest known living organisms [1]. Moreover, next to that of *Candidatus Carsonella ruddii* [2], it has the smallest-ever sequenced genome, which is only 490,885 base pairs long. It is also one of the most compact genomes. Computational predictions suggest that approximately 95% of the DNA encodes proteins or stable RNA. *Nanoarchaeum equitans* lacks genes for most vital metabolic pathways, including lipid, cofactor, amino acid and nucleotide biosynthesis. However, contrary to most known organisms with reduced genomes, it has a full set of the enzymes involved in DNA replication, repair and recombination, one of which is a single-stranded DNA binding like protein [3].

Single-stranded DNA binding proteins are vital elements of living cells and are present in all life domains and in viruses. By means of sequence independent interaction with ssDNA, these proteins prevent strand pairing, secondary structure formation and nuclease degradation [4]. In this way, SSB proteins play a part in every process involving ssDNA metabolism, such as, for instance, replication, recombination and repair [5–8]. SSB proteins can be identified by the presence of a highly conserved DNA binding domain known as the OB fold, which is to say, the oligonucleotide/oligosaccharide/oligopeptide binding fold [9], typically consisting of approximately 100 amino acid residues. However, the subunit composition varies over life domains. The bacterial SSBs characterized to date [10–13], with the exception of those from *Deinococcus-Thermus* [14–16], form homotetramers, while most of eukaryotic SSBs, known as replication protein A (RPAs), usually act in solution as heterotrimers [17]. Current knowledge in respect of the archaeal SSB proteins members of Craenarchaeota phylum posits that they resemble the bacterial-type SSBs in domain organization [18–20], whereas Euryarchaeota have eukaryotic-like RPAs [21,22]. At present, no SSB proteins in the other three *Archaea* phyla, namely Korarchaeota, Thaumarchaeota and Nanoarchaeota, have been reported.

To date, four nanoarchaeal proteins have been published, namely reverse gyrase [23], tRNA splicing endonuclease [24], neelaredoxin [25] and family B DNA polymerase [26]. Interestingly, all of them possess unusual features [23–27]. The sequence analysis of the *Nanoarchaeum equitans* SSB protein indicates that it may also fit the trend. The aim of this study was to clone and overexpress a *Nanoarchaeum equitans* Kin-4m *ssb*-like gene in *E. coli*, purify the gene product and study the biochemical properties and protein-DNA interactions of *NeqSSB*-like protein.

Results

NeqSSB-like protein sequence analysis

The sequence analysis of the *Nanoarchaeum equitans* Kin-4M genome [GenBank: AE017199] indicated the presence of a single *ssb*-like gene. On the basis of this nucleotide sequence, the

NeqSSB-like protein contains 243 amino acid residues including the N-terminal methionine and has a predicted molecular weight of 27.82 kDa. The RPS-BLAST search revealed one putative DNA binding sequence situated in the region between amino acid residues 71 and 145. Both, this location and the domain length, is unusual, since the OB fold can typically be found near the N-terminus and consists of about 100 amino acid residues. Similarly to *SsoSSB*, *NeqSSB*-like lacks the conserved C-terminal DIPF sequence, although the C-terminal domain does, indeed, have an acidic character. Fig 1A shows the multiple amino acid alignment of the *Nanoarchaeum equitans*, *Sulfolobus solfataricus*, *Sulfolobus acidocaldarius*, *Desulfurococcus kamchatkensis*, *Staphylothermus marinus*, *Escherichia coli*, *Thermotoga maritima* and *Thermoanaerobacter tengcongensis* SSB proteins, all of which contain one OB fold domain. Fig 1B shows the multiple amino acid alignment of *Nanoarchaeum equitans* and craeonarchaeal SSBs from *Sulfolobus solfataricus*, *Sulfolobus acidocaldarius*, *Desulfurococcus kamchatkensis* and *Staphylothermus marinus*. The identity and similarity between *NeqSSB*-like and *EcoSSB* is 10% and 29% respectively, whereas the alignment between *NeqSSB*-like and *SsoSSB* displays 14% identity and 32% similarity, respectively. These low scores may result from the large difference in the length of amino acid sequences, because sequence of *NeqSSB*-like protein is much longer than the others (from 62 to nearly 100 amino acid residues). The multiple amino acid alignment also indicates that the *NeqSSB*-like W108 residue probably is involved in ssDNA binding, as it is conserved in the *Nanoarchaeum equitans* and craeonarchaeal SSB proteins. It corresponds to the base-stacking residue W56 in *SsoSSB* which has already been identified [28].

The cloning, expression and purification of *NeqSSB*-like

Using *E. coli* TOP10F' cells carrying the recombinant plasmid pBAD/*NeqSSB*-likeHT, an efficient expression platform under the strict control of a P_{BAD} promoter was constructed. *NeqSSB*-likeHT was produced in a soluble form in cytosol. The SDS-PAGE analysis indicated the presence of a expressed protein with a molecular mass of 28 kDa. The *E. coli* overexpression system used in this study allowed 7.5 mg *NeqSSB*-like protein per 1 l of induced culture to be produced. The purity of the protein after all purification steps, namely Ni^{2+} -affinity chromatography, TEV cleavage procedure and ssDNA-cellulose affinity chromatography, was approximately 98%, as evaluated by SDS-PAGE (Fig 2).

The oligomerization status of *NeqSSB*-like

The SDS-PAGE analysis of the purified *NeqSSB*-like revealed a single major band with a molecular mass of approximately 28 kDa. The analysis of a purified protein, using analytical gel filtration chromatography at a concentration range between 12 and 236 μ M and a different concentration of NaCl (10–300 mM), revealed a homogeneous protein state population (Fig 3). There was one major peak with a molecular mass of 26.36 kDa, as calculated using a regression curve equation (Fig 3). The native molecular mass of the peak represents 0.94 of the monomer mass. Thus *NeqSSB*-like exists in solution as monomer.

We also used centrifugation in order to analyze the oligomeric state of *NeqSSB*-like in 15 to 30% (w/v) glycerol gradients. To prevent nonspecific aggregation of the protein during the experiments, NaCl at a final concentration of 0.5 M was added to the solutions used for the glycerol gradients. *NeqSSB*-like protein (27.82 kDa) sediments in the position corresponding to a monomeric state (Fig 4). The centrifugation in the glycerol gradients of *NeqSSB*-like was carried out three times and the same sedimentation behaviors were observed in all the independent tests. In addition, the same result was obtained when 50 μ l of a 12 μ M *NeqSSB*-like protein was examined during this experiment (data not shown), which indicates that the oligomerization state of *NeqSSB*-like is independent of concentration. Thus the sedimentation

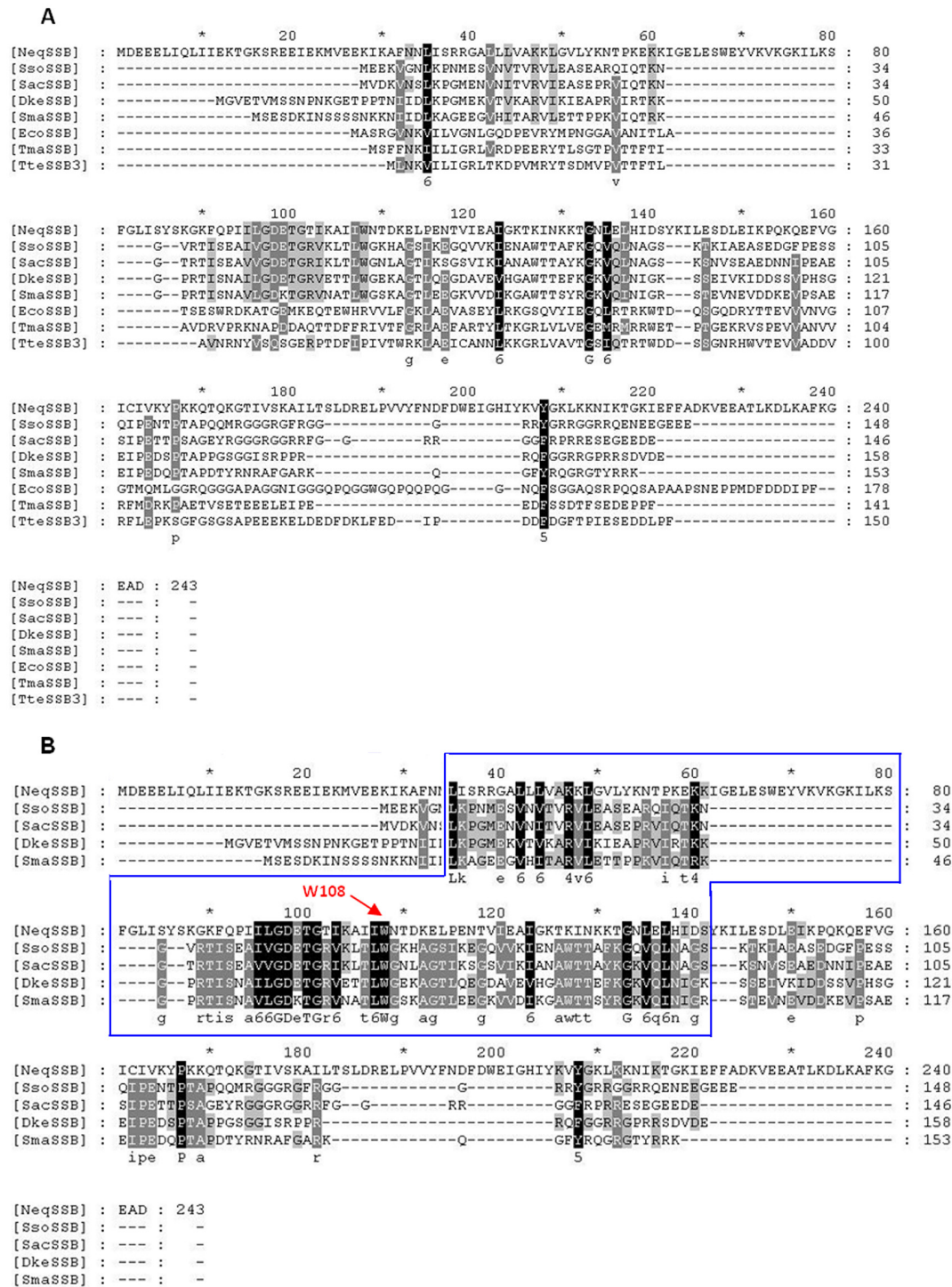


Fig 1. The multiple amino acid alignment. **A** The multiple amino acid alignment of nucleic acid binding protein from *Nanoarchaeum equitans*, bacterial and craeonarchaeal SSB proteins. **B** The multiple amino acid alignment of nucleic acid binding protein from *Nanoarchaeum equitans* and craeonarchaeal SSB proteins. The alignments were performed by dividing the amino acids into six similarity groups: group 1 V, L, I, M, group 2 W, F, Y, group 3 E, D, group 4 K, R, group 5 Q, D, and group 6 S, T. The capital letters represent single amino acid codes. White fonts on black boxes represent 100% similarity, white fonts on grey boxes denote <80% similarity, and black fonts on grey boxes show <60% similarity. Abbreviations: NeqSSB—nucleic acid binding protein from *Nanoarchaeum equitans* Kin-4M, SsoSSB *Sulfolobus solfataricus* strain P2, SacSSB *Sulfolobus acidocaldarius* DSM 639, SmaSSB *Staphylothermus marinus* F1, DkeSSB *Desulfurococcus kamchatkensis* 1221n, EcoSSB *Escherichia coli* K12, TmaSSB *Thermotoga maritima* strain MSB8, and TteSSB3 *Thermoanaerobacter tengcongensis* MB4. The W108 residue, important in base-stacking interactions is indicated in the panel B. The blue box indicates OB-fold region.

doi:10.1371/journal.pone.0126563.g001

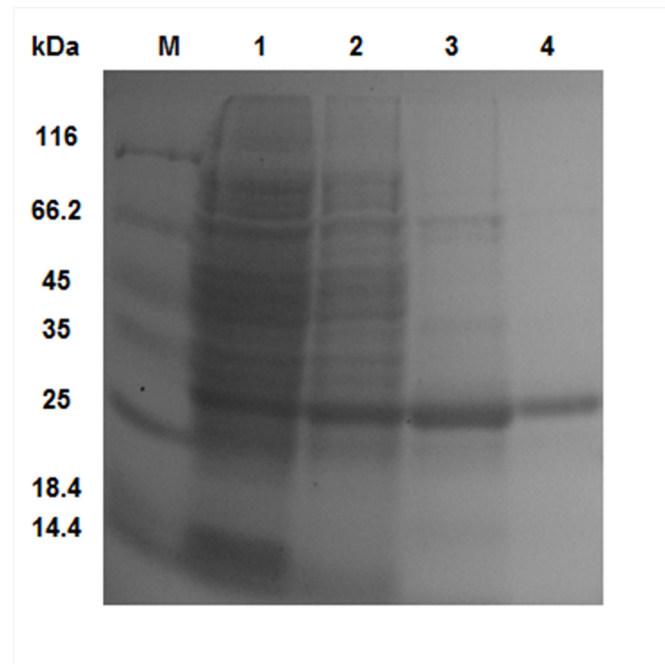


Fig 2. The expression and purification of *NeqSSB*-like from *E. coli* TOP10F'+pBAD/*NeqSSBHT*. The proteins were analyzed on a 12% polyacrylamide gel. Lane M: Unstained Protein Weight Marker (Fermentas, Lithuania), with the molecular mass of proteins marked. Lane 1: soluble protein cell extracts after arabinose induction of protein expression (10 μ l). Lane 2: *NeqSSB*-like after the Ni²⁺-affinity chromatography step (10 μ l). Lane 3: *NeqSSB*-like after His-tag cleavage with TEV protease (10 μ l). Lane 4: *NeqSSB*-like after chromatography on an ssDNA-cellulose column (10 μ l).

doi:10.1371/journal.pone.0126563.g002

pattern of *NeqSSB*-like and standard proteins in the glycerol gradients suggests that the *NeqSSB*-like only forms the monomeric structure.

ssDNA binding properties

Initial studies on ssDNA binding were carried out using a fixed quantity of oligo (dT)_n (n = 35, 76, 120). The oligos were incubated with various concentrations of *NeqSSB*-like and the resulting complexes were analyzed, using an agarose gel electrophoresis (Fig 5). The binding reaction started when the *NeqSSB*-like concentration in the test tube was 20 pmol. When an increasing protein amount binds to the oligo (dT)_n, their mobility in the gel is progressively reduced, resulting in at least five visible complexes, until an end-point, where all available ssDNA stay in the gel well, is reached. These observations do not depend on the length of the oligonucleotide used. In an assay with (dT)₇₆ and (dT)₁₂₀, sharp bands were obtained, whereas the picture was somewhat smeary with (dT)₃₅. This would suggest that concentration-dependent oligomerization on DNA substrate is occurring.

The binding of *NeqSSB*-like to circular M13 ssDNA (6407 bp) was also examined. A fixed quantity of M13 ssDNA was incubated with an increasing amount of *NeqSSB*-like and the resulting complexes were analyzed, using agarose gel electrophoresis. A progressive decrease of M13 ssDNA mobility, along with a quenching of ethidium bromide fluorescence, owing probably to the relaxation of the DNA secondary structures, was observed when increasing the amount of protein added (Fig 6).

To further explore the ssDNA binding properties of *NeqSSB*-like, fluorescence spectroscopy was used (Fig 7). All the SSBs studied to date show a dramatic decrease in naturally-occurring

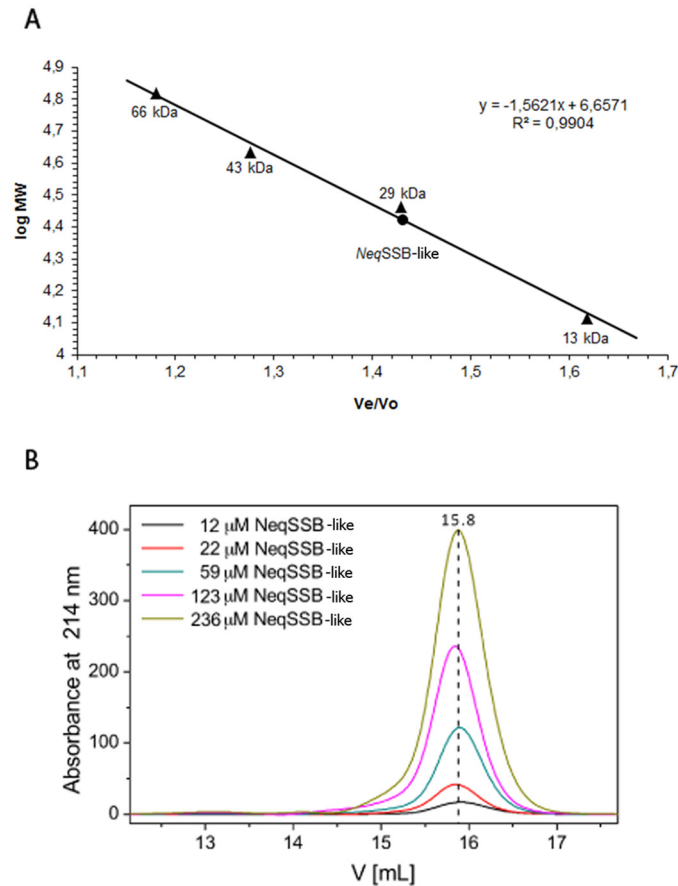


Fig 3. Results of the analytical gel filtration of *NeqSSB-like* on the Superdex 75 10/300 GL column. A The standard linear regression curve was generated by plotting the log of the molecular mass of calibration proteins against V_e/V_0 value, namely elution volume divided by void volume. The calibration proteins represented by black triangles include bovine albumin (66 kDa), ovalbumin (43 kDa), carbon anhydrase (29 kDa) and ribonuclease A (13 kDa). *NeqSSB-like* is represented by the black circle. The regression curve equation and coefficient of determination are shown. **B** Effects of *NeqSSB-like* protein concentrations on the elution profiles of gel filtration. The entire range of *NeqSSB-like* concentrations (12–236 μM) show an elution volume of 15.8 ml corresponding to the monomeric protein.

doi:10.1371/journal.pone.0126563.g003

tryptophan fluorescence when binding to ssDNA. With an excitation wavelength of 295 nm, the emission spectrum of SSB proteins at 25°C had a maximum at 348 nm, consistent with tryptophan. On the addition of a saturating quantity of ssDNA, the intrinsic fluorescence at 348 nm was quenched by 69% in 100 mM and 500 mM NaCl containing buffers and by 63% in a 2 mM NaCl containing buffer. The estimated binding site in the presence of 2, 100 and 500 mM NaCl was determined as 6 ± 1 , 7 ± 1 and 7 ± 1 nt, respectively. Therefore, practically no binding mode transition was observed when changing the ionic strength from low to high salt.

Taking into account all the data collected from the *NeqSSB-like* ssDNA binding studies, a cooperative binding model resembling that of DnaA protein complexes is proposed [29]. This would explain the 7 nt binding site and oligonucleotide length-independent number of shifted complexes. The large *NeqSSB-like* complexes bound to ssDNA of various lengths produce differences undistinguishable at the resolution offered by agarose gel electrophoresis. However, the gel smears in the assay with (dT)₃₅ may indicate the most cooperative and unorganized binding mode, where only one protein molecule per oligo is bound directly to ssDNA and the others attach to each other cooperatively, forming complexes varying in size. The correctness

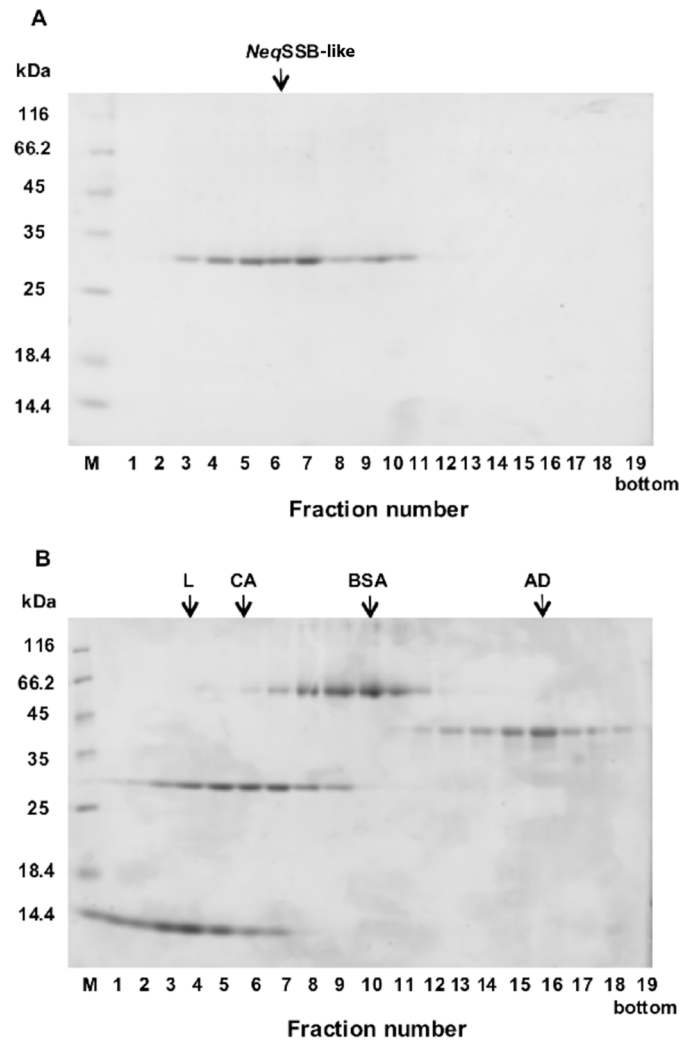


Fig 4. Sedimentation analysis of *Neq*SSB-like (A) and standard proteins (B). The proteins were analyzed on a 12% polyacrylamide gel. Lane M: Unstained Protein Weight Marker (Fermentas, Lithuania), with the molecular mass of proteins marked. Lane 1–19: fraction number. 50 μ l of 150 μ M *Neq*SSB-like and the corresponding amounts of standard proteins were centrifuged in linear 15 to 30% (w/v) glycerol gradients, as described in the “Methods”. The fractions with proteins were analyzed by SDS-PAGE. The fractions at which the maximal amount of protein appears are shown by arrows in each panel. The standard proteins used are: L, lysozyme (14 kDa); CA, carbonic anhydrase (29 kDa); BSA, bovine serum albumin (66 kDa) and AD, alcohol dehydrogenase (150 kDa). The oligomerization state estimation of *Neq*SSB-like was made with these proteins.

doi:10.1371/journal.pone.0126563.g004

of this hypothesis is to be further examined by crystallographic studies or high resolution AFM imaging.

Other nucleic acid binding properties and binding preferences

Taking into consideration the low sequence similarity of *Neq*SSB-like to the SSBs studied so far and the extraordinary nature of *Nanoarchaeum equitans*, protein interactions with other nucleic acids were tested. mRNA and dsDNA samples were incubated with increasing amount of *Neq*SSB-like and the resulting complexes were analyzed with agarose gel electrophoresis. The tests showed the capability of mRNA binding, and surprisingly, dsDNA (Fig 8). As with the

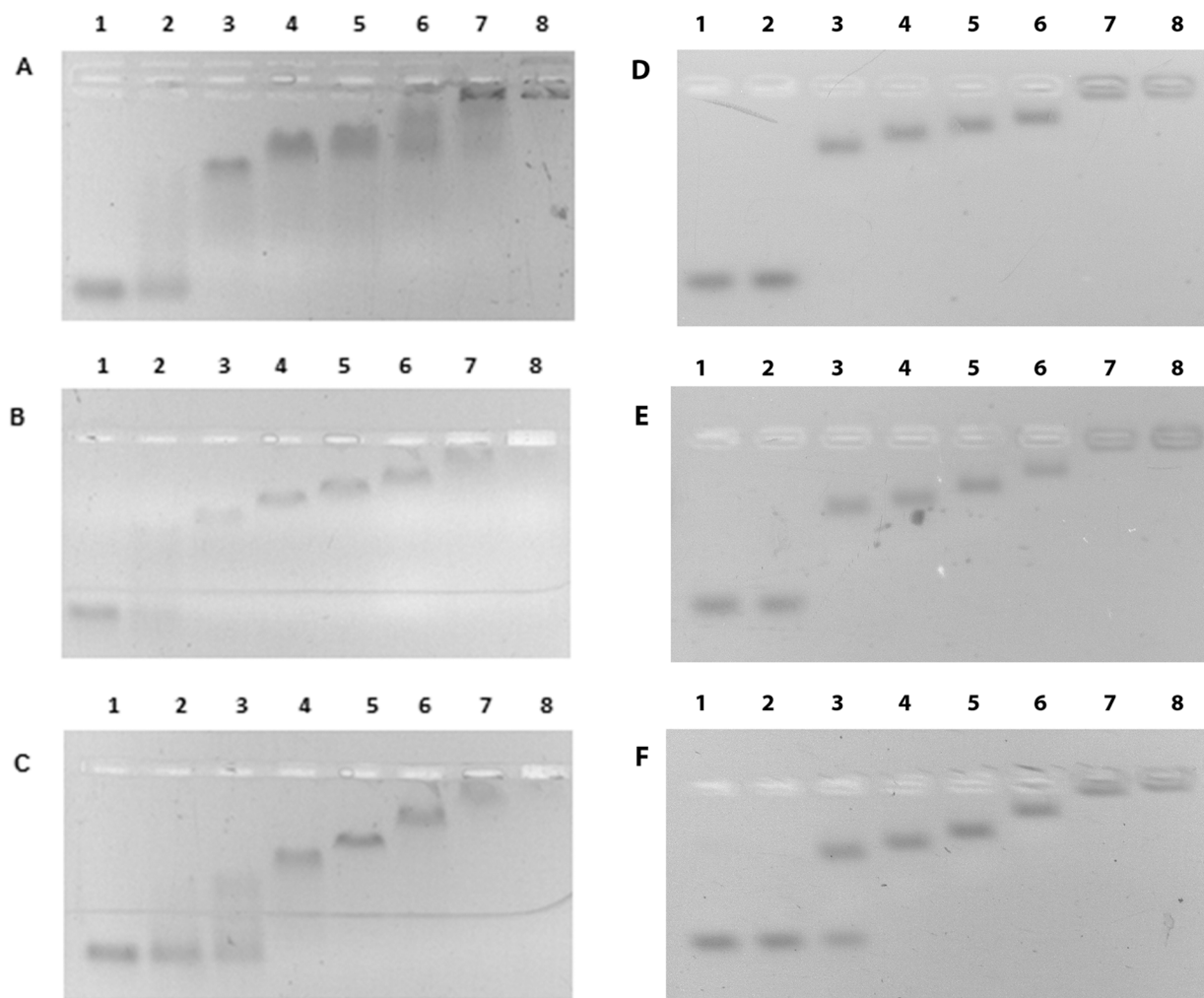


Fig 5. Binding of *Neq*SSB-like to a fixed quantity (10 pmol) of oligonucleotides. A (dT)₃₅ B (dT)₇₆ C (dT)₁₂₀. Lanes 1–8 contain 0, 10, 20, 40, 80, 160, 320 and 640 pmoles of *Neq*SSB-like, respectively.

doi:10.1371/journal.pone.0126563.g005

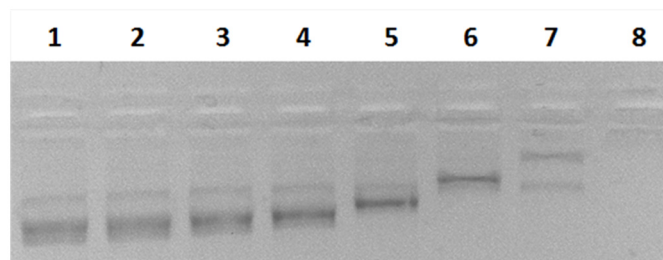


Fig 6. Binding of *Neq*SSB-like to M13 ssDNA. Lanes 1–8 contain (0.07 pmol) of M13 ssDNA and 0, 3.5, 7, 14, 28, 56, 112 and 224 pmoles of *Neq*SSB-like, respectively.

doi:10.1371/journal.pone.0126563.g006

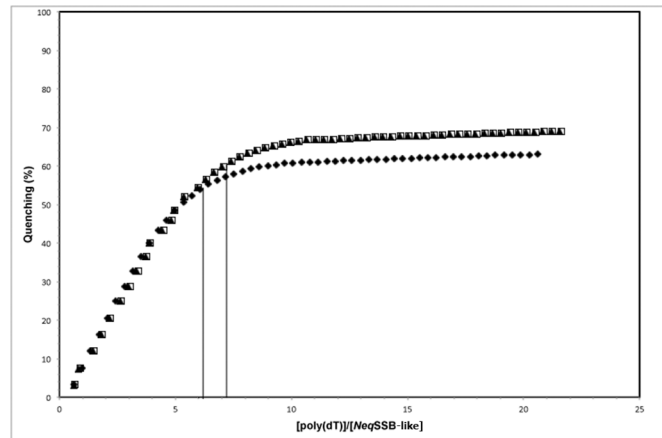


Fig 7. The inverse fluorescence titration of *NeqSSB*-like with poly(dT). 2 nM samples were titrated with a saturating quantity of poly(dT) in 2 mM, represented by the fill diamonds, 100 mM, shown as fill triangles and 500 mM NaCl, given as open squares, in a binding buffer. The vertical lines indicate the binding site size calculated for each assay.

doi:10.1371/journal.pone.0126563.g007

ssDNA studies, the mobility of mRNA and dsDNA is progressively reduced when increasing the amount of *NeqSSB*-like until the whole sample stays in gel well. No preference for a particular type of dsDNA topology was observed.

These experiments gave rise to the question as to whether *NeqSSB*-like can be classified as an SSB protein. To solve this, *NeqSSB*-like was incubated with both ssDNA and dsDNA and the resulting complexes were separated, using agarose gel electrophoresis (Fig 9). The results showed that, with a low protein concentration, ssDNA is bound preferentially and that after saturation, an affinity for dsDNA is observed. Summing up, the results obtained generate speculations about the pleiotropic character of *NeqSSB*-like protein. *NeqSSB*-like may be involved in multiple metabolic pathways with both ds and ssDNA and RNA. Whether this unusual, broad substrate specificity is of ancestral character, or is a consequence of genome reduction remains unknown.

Kinetic analysis of *NeqSSB*-like interaction with single- and double stranded DNA

In order to conduct further study on the interaction mode of *NeqSSB*-like with single- and double-stranded DNA (ss- and dsDNA 60-mer), the SPR studies were conceived (Fig 10). They confirmed the unique nature of protein in question, revealing an affinity towards ss- and dsDNA which is unusual for SSB proteins. Using the same ssDNA 60-mer, typical dissociation constants (K_d) for SSB proteins such as *EcoSSB* or *MtuSSB* have been determined at the level of nanomolar values [30]. The study also included a control measurement of binding recombinant *EcoSSB* to ssDNA (Table 1). The interaction of *NeqSSB*-like with ssDNA is significantly lower, as is reflected in the K_d value of micromolar range (Table 1). Surprisingly, the difference in the preference of *NeqSSB*-like binding to the ssDNA versus the dsDNA has not been meaningfully shown by SPR studies. Though the affinity of *NeqSSB*-like to ssDNA is higher, the values of the kinetic constants are within the same order of magnitude. The dissociation rate for ssDNA is slower, which is reflected in the shape of the sensograms in the dissociation phase and calculated k_{off} values. When compared to other SSB proteins, the main difference in the mode of binding to ssDNA is observed in the dissociation phase; typical values of dissociation rate fall within the 10^{-4} 1/s range [30, 31]. This can easily be explained by the oligomeric state

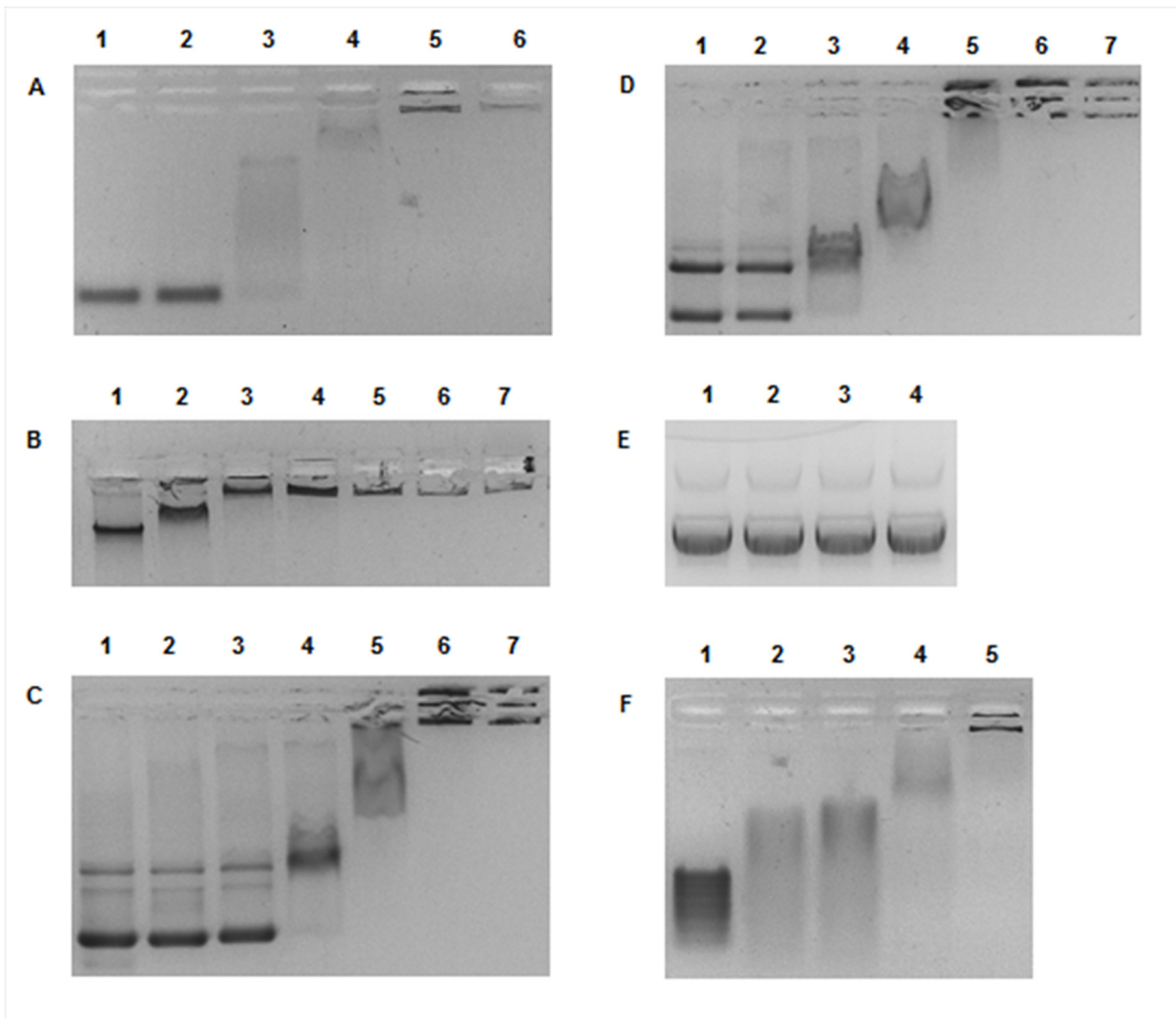


Fig 8. *Neq*SSB-like dsDNA and mRNA binding properties. **A** Binding to 2.5 pmol of 100 bp PCR product. Lanes 1–6 contain 0, 10, 20, 40, 80 and 160 pmoles of *Neq*SSB-like, respectively. **B** Binding to 0.132 pmol of *Escherichia coli* genomic DNA. Lanes 1–7 contain 0, 10, 20, 40, 80, 160 and 320 pmoles of *Neq*SSB-like, respectively. **C** Binding to 0.2 pmol of pDONR201 plasmid DNA (4470 bp). Lanes 1–7 contain 0, 10, 20, 40, 80, 160 and 320 pmoles of *Neq*SSB-like, respectively. **D** Binding to 0.1 pmol of pDONR201 plasmid DNA + 0.05 pmol of linearized pDONR201 plasmid DNA. Lanes 1–7 contain 0, 10, 20, 40, 80, 160 and 320 pmoles of *Neq*SSB-like, respectively. **E** Control binding reaction with 0.2 pmol of pDONR201 plasmid DNA. Lanes 1–4 contain 0, 10, 20 and 40 pmoles of *Taq*SSB, respectively. **F** Binding to 980 ng of mRNA. Lanes 1–5 contain 0, 10, 20, 40, 80 pmoles of *Neq*SSB-like, respectively.

doi:10.1371/journal.pone.0126563.g008

of *Neq*SSB-like, which was determined in this study as monomeric. For other SSB proteins, which are oligomers, most often tetramers, the slow dissociation rate observed by the SPR studies is caused by the avidity effect of multiple binding sites in the oligomer structure.

Thermostability

The half-lives of *Neq*SSB-like ssDNA-binding activity at different temperatures, as determined by a gel mobility shift assay of (dT)₃₅ in 2% agarose gel, are 60 min at 85°C, 30 min at 90°C, 15 min at 95°C and 5 min at 100°C (Fig 11). As regards *Taq*SSB, which was tested as a control protein, the numbers were 10 min at 85°C, 3 min at 90°C, 1 min at 95°C and 0 min at 100°C, as has previously been shown by Dąbrowski *et al.* [14].

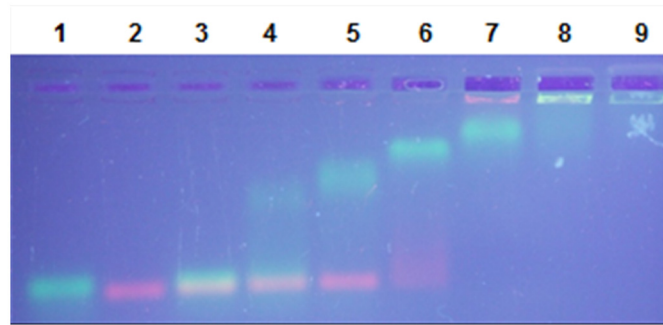


Fig 9. *Neq*SSB-like binding preferences. The reactions contained a fixed quantity of the sample DNA: 10 pmol of (dT)₇₆ and 2.5 pmol of 100 bp PCR product. Lane 1: (dT)₇₆ with 0 pmol of *Neq*SSB-like. Lane 2: 100 bp with 0 pmol of *Neq*SSB-like. Lane 3: (dT)₇₆ and 100 bp PCR product with 0 pmol of *Neq*SSB-like. Lanes 4–9 contain 10, 20, 40, 80, 160 and 320 pmoles of *Neq*SSB-like, respectively.

doi:10.1371/journal.pone.0126563.g009

The Differential Scanning Calorimetry (DSC) analysis of *Neq*SSB-like in Power = f(T) coordinate system showed that thermal unfolding was an irreversible process, as seen on a rescan thermogram (Fig 12). Therefore *Neq*SSB-like does not possess thermodynamic stability. The melting temperature (T_m) was determined as being 100.2°C, which supports the results obtained for the indirect method. The thermogram analysis indicated no signs of heavy protein aggregation after heat denaturation. Furthermore, the combined results obtained from both methods suggest that the loss of ssDNA binding activity is connected with an irreversible thermal unfolding of *Neq*SSB-like.

*Neq*SSB-like cannot replace *Eco*SSB *in vivo*

In the complementation experiments, we attempted to replace the resident plasmid (pRPZ146, ori ColEI, Tc^R, harbouring a wild-type *ssb* gene) from *E. coli* RDP268(DE3) ($\Delta ssb::Kan$) with the plasmids harbouring test *ssb* genes (pET23D(+)*Neq*SSB-like or pET23-D(+)*Eco*SSB, ori ColEI, Amp^R). As SSB is an essential protein, success in replacement of the original Tc^R plasmid by the incoming Amp^R plasmid, resulting in a Tc^S, Amp^R phenotype, shows that the test SSB complements the Δssb strain of *E. coli*. The pET23D(+)*Eco*SSB was used as a control to assess the efficacy of the complementation assay. After transformation of modified *E. coli* cells using pET23D(+)*Neq*SSB-like or pET23-D(+)*Eco*SSB, which encodes resistance against ampicillin, and subsequent inoculations, we could isolate clones that showed resistance to ampicillin and kanamycin but not to tetracycline. These clones must have lost the pRPZ146 plasmid encoding for *Eco*SSB. Additionally, an analysis of plasmid DNA was carried out after plasmid isolation from tested strains. Unfortunately, despite the implementation of a number of passages to replace the resident plasmid pRPZ146 from *E. coli* RDP268(DE3) with the plasmids pET23D(+)*Neq*SSB-like is not achieved.

Discussion

The research detailed in this work encompassed the cloning, purification and initial characterization of the novel nucleic acid binding protein from the hyperthermophilic archaeon *Nanoarchaeum equitans*. From the sequence analysis obtained, it can be ascertained that *Neq*SSB-like possesses a non-canonical OB fold domain. Using multiple amino acid alignment, the W108 residue was identified as being important in base-stacking interactions. It corresponds to W56 in *Sso*SSB, which has been characterized during thermodynamic studies of ssDNA binding as the greatest DNA binding energy contributor [28].

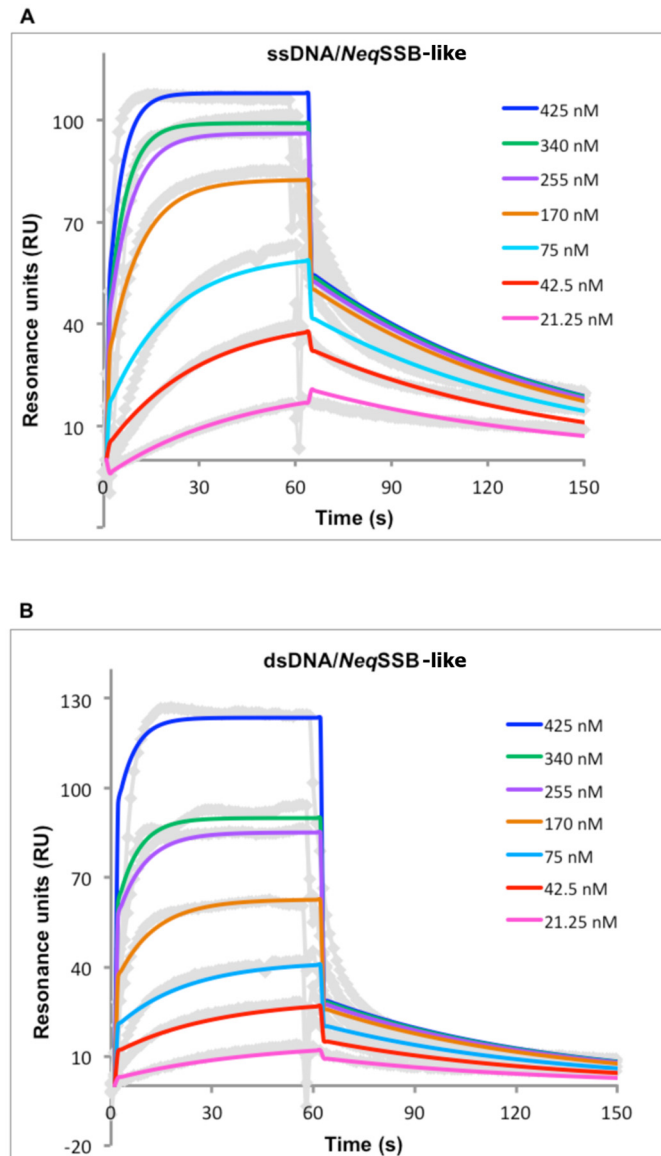


Fig 10. Interaction analysis of *NeqSSB*-like (A) with ssDNA and dsDNA (B). Different concentrations of the protein were injected with a flow rate of 30 μ l/min on a streptavidin chip coated with ssDNA 60-mer and dsDNA 60-mer on separate flow channels. A flow cell with streptavidin was used as a reference. After each injection the chip was regenerated with 0.01% SDS. The different colors of the sensograms represent the concentrations of the *NeqSSB*-like injected. Solid lines state for fitted curves in accordance with the Langmuir model and using BiaEval 3.0 software.

doi:10.1371/journal.pone.0126563.g010

The gel filtration and sedimentation experiments showed only the monomeric state of *NeqSSB*-like protein that is independent of salt and protein concentration. The observations are the first to date in respect of non-viral SSB proteins. In comparison, the *SsoSSB* protein can form monomers and tetramers in solution [18, 19].

Non-canonical structural properties appear to be important in protein universality and multifunctionality. *NeqSSB*-like possesses the ability to bind all nucleic acid types, with a clear preference for single-stranded forms. On the basis of this study a cooperative model is proposed. Only after solving the *NeqSSB*-like apo and holo crystal structures in order to gain insights into

Table 1. Binding parameters for the *Neq*SSB-like interaction with single- and double-stranded DNA.

	K_D [M]	k_{on} [1/Ms]	k_{off} [1/s]
<i>Neq</i> SSB-like versus ssDNA	$2.66 \times 10^{-6} \pm 0.64$	$7.91 \times 10^3 \pm 1.79$	$2.39 \times 10^{-2} \pm 0.46$
<i>Neq</i> SSB-like versus dsDNA	$5.26 \times 10^{-6} \pm 1.42$	$8.30 \times 10^3 \pm 2.2$	$4.89 \times 10^{-2} \pm 1.7$
<i>Eco</i> SSB versus ssDNA	$5.36 \times 10^{-9} \pm 1.08$	$3.45 \times 10^4 \pm 0.83$	$1.85 \times 10^{-4} \pm 0.23$

Binding reactions were performed as described in the “Methods” section. The kinetic data of binding *Neq*SSB-like protein (analyte) to the 60-mer ss- and dsDNA (ligand) were fitted using the Langmuir binding model. The parameters calculated for *Eco*SSB are based on the control experiment conceived within the study. The data shown are the means of at least three independent measurements.

doi:10.1371/journal.pone.0126563.t001

the above-mentioned structural properties will it be possible to present data confirming the suggested binding model.

A salt dependent binding size transition is a common observation in the studies of most of the known SSBs, such as, for example, in *Escherichia coli* SSB [6] and *Deinococcus-Thermus*-type SSBs [14–16]. The most extensively studied *Eco*SSB has at least two different DNA binding modes. In high salt concentrations, 65 nucleotides bind per functional protein tetramer, with fluorescence quenching of almost 90%, whereas, in low salt concentrations, 35 nucleotides saturate the protein and quench its intrinsic fluorescence by 53%. The exceptions to this rule were the *Thermotoga maritima* and *Thermotoga neapolitana* SSBs [13]. *Neq*SSB-like also lacks this distinctive property. The spectrofluorimetric studies indicated a salt-independent binding site of approximately 7 nucleotides. In comparison, *Sulfolobus solfataricus* SSB binds 4–5 nt ssDNA per monomer or 20–25 nt per tetramer [18].

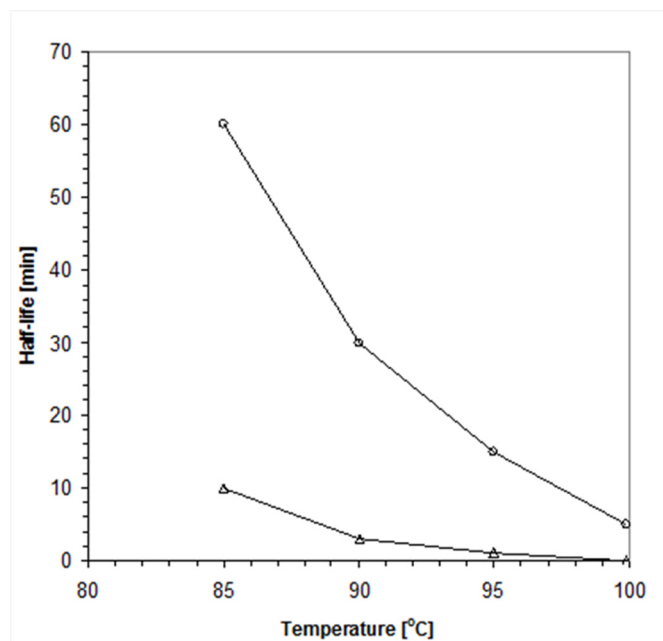


Fig 11. The determination of ssDNA-binding activity half-life, using gel mobility shift assays, for *Neq*SSB-like and *Taq*SSB. *Neq*SSB-like represented by the open circles and *Taq*SSB, shown as open triangles.

doi:10.1371/journal.pone.0126563.g011

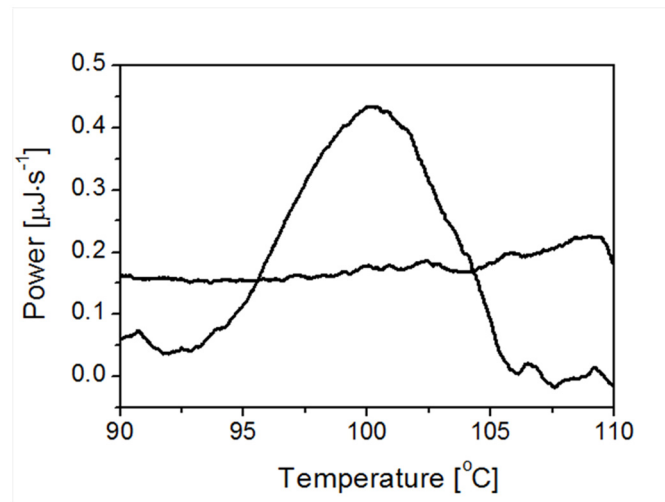


Fig 12. NeqSSB-like thermogram. A sample containing 1 mg/ml of purified protein was analyzed in a 20 mM phosphate buffer pH 7.5 containing 150 mM NaCl. NeqSSB-like melting temperature is shown.

doi:10.1371/journal.pone.0126563.g012

The interaction studies by means of SPR assays demonstrated the extraordinary nature of the NeqSSB-like in question in respect of both the strength and preference of binding DNA molecules. As the gel-shift assays showed, NeqSSB-like can interact with ssDNA, as well as dsDNA, with a slight preference for the first. The values of the kinetic constants are much lower than for the SSB proteins previously studied, though this is also caused by the monomeric structure of the protein, which does not alter the interaction studies by avidity effect, as with other multimeric SSB proteins.

NeqSSB-like possesses a fairly high thermostability and its ssDNA binding activity is preserved for 5 minutes, even at a temperature of 100°C. Furthermore, the results obtained from differential scanning microcalorimetry (DSC) demonstrated the melting temperature (T_m) of NeqSSB-like to be 100.2°C. These numbers do not match the SSBs known to be the most thermostable, namely TmaSSB (T_m of 109.3°C) and TneSSB (T_m of 112.5°C) [13], but are significantly higher than those observed for TaqSSB (T_m of 86.3°C) [14]. EcoSSB is essential for the survival of the *E. coli* cell [32]. Using an *E. coli* *ssb* mutant strain, we could show that only NeqSSB-like can take over the function of EcoSSB *in vivo*. We concluded that NeqSSB-like is not a single-stranded DNA binding protein involved in routine DNA functions (analogous to the *E. coli* SSB protein) or it works incorrectly in the mesophilic *E. coli* host because of its about 1000 folds lower activity for ssDNA binding than EcoSSB.

Conclusions

This paper reports on the purification and characterization of the *Nanoarchaeum equitans* SSB protein. Its relation to other known members of this protein class is also presented.

The molecular mass of NeqSSB-like, deduced from its 243 amino acid sequence, is 27.82 kDa.

The results of the study show its unique ability to bind different types of nucleic acids including, ssDNA, dsDNA and mRNA, with a confirmed preference to single-stranded substrates. Therefore the *in vivo* multifunctional biological role of NeqSSB-like is suggested.

The high thermostability of NeqSSB-like, with a half-life of 5 min at 100°C and a DSC-determined T_m of 100.2°C, together with its unique binding properties, may offer an attractive tool for thermal nucleic acid amplification techniques.

Methods

Cloning of the *ssb*-like gene from *Nanoarchaeum equitans* Kin4-M

Nanoarchaeum equitans Kin4-M genome DNA was obtained from the Institute for Microbiology at the University of Regensburg, courtesy of Dr Harald Huber. To confirm the presence of *ssb*-like gene, primers complementary to the flanking sequences, which encode the hypothetical protein (NEQ200) and aspartyl-tRNA (NEQ_t17), were designed and synthesized. The forward primer was 5' ATATTAAAGCAAACCATTAGACAATTAAAGC (31 nt) and the reverse primer was 5' TTATTCTCTAAAAGGCTATATAATAGTGG (29 nt). The PCR reaction solution consisted of 0.2 µg of *Nanoarchaeum equitans* Kin4-M genome DNA, 1 µl (10 µM) of each primer, 2.5 µl (10 mM) dNTPs, 2 µl (25 mM) MgCl₂, 2.5 µl of 10 x Hot Start Buffer (200 mM Tris-HCl pH 8.3, 200 mM KCl, 50 mM (NH₄)₂SO₄), and 2 U of Maxima Hot Start *Taq* DNA Polymerase (Fermentas, Lithuania). 40 cycles were performed, using the Veriti 96 Well Thermal Cycler (Applied Biosystems, USA), with a temperature profile of 60 s at 94°C, 60 s at 56°C and 60 s at 72°C. Specific, approximately 940 bp products were obtained, then purified, using the Agarose-Out DNA Purification Kit (EURx, Poland) and sequenced in order to confirm the presence of the *ssb*-like gene. The respective data are not shown.

Based on the *ssb*-like gene sequence thus confirmed, specific PCR primers were designed and synthesized. The forward primer was NeqSSB-HT-Nco 5' ggaggaccatggctcaccatcatcatcatcatgagaacctgtacttccagggtGATGAAGAGGAACTAATACAAC (75 nt, containing the *Nco*I recognition site, underlined, and the His-Tag domain and TEV cleavage site, given in lower case, italics) and the reverse primer was NeqSSB-HT-Eco 5' ttagcgaattcTCAATCGGCCTCTCCTTTAAAG (34 nt, containing the *Eco*RI recognition site, underlined). The sections of both primer sequences given in bold are complementary to the nucleotide sequence of the *ssb*-like gene of *Nanoarchaeum equitans* Kin4-M.

The PCR reaction conditions were the same as those already described, with the exception of the temperature profile, which was 60 s at 94°C, 120 s at 51°C and 60 s at 72°C. The PCR products were analyzed, by means of electrophoresis on 2% agarose gel stained with ethidium bromide, at a final concentration of 0.5 µg/ml. Specific, approximately 800 bp products were obtained and isolated from the gel, using the Agarose-Out DNA Purification Kit (EURx, Poland). The PCR product was digested with *Nco*I + *Eco*RI (Fermentas, Lithuania), then purified, using the PCR / DNA Clean-Up Purification Kit (EURx, Poland) and ligated into pBAD/*myc*-His A (Invitrogen, USA) between the *Nco*I and *Eco*RI sites. The *E. coli* TOP10F' cells were transformed with the ligation mixture and 23 colonies were examined for the presence of the *ssb*-like gene from *Nanoarchaeum equitans*, using a gel retardation assay and restriction analysis. One clone was selected and sequenced to confirm the presence of the *ssb*-like gene.

Protein sequence analysis

The amino acid sequence of NeqSSB-like was analyzed using standard protein—protein BLAST and RPS-BLAST. Multiple sequence alignment was generated in ClustalX using PAM 500 score matrix. The results were prepared, using the GeneDoc editor program (copyright Karl Nicholas).

Expression and purification of NeqSSB-like

E. coli TOP10F' cells carrying pBAD/NeqSSBHT were grown at 37°C in Luria-Bertani medium, supplemented with 100 µg/ml of ampicillin and 12.5 µg/ml of tetracycline to an OD₆₀₀ of 0.4 and were induced by incubation in the presence of arabinose, at a final concentration of 0.02%, for 24 h. The cells were then harvested by centrifugation at 4612xg for 10 min and the pellets

were resuspended in 20 ml of buffer A (20 mM Tris-HCl pH 7.9, 0.5 M NaCl, 0.1% Triton X-100 and 5 mM imidazole). The samples were sonicated three times, for 30 s at 4°C, and centrifuged at 10000xg for 15 min. The *Neq*SSB-like was next purified in a three-step procedure. First, the *Neq*SSBHT-like fusion protein was purified, using Ni²⁺-affinity chromatography. The supernatant containing *Neq*SSBHT-like was loaded directly onto an His•Bind Column (Novagen, USA), which has previously been prepared and equilibrated with 30 ml of A buffer. The *Neq*SSBHT-like was eluted with a 5–500 mM imidazole gradient and the elution fraction was then dialyzed against 50 mM Tris-HCl pH 8, 150 mM NaCl and 0.5 mM EDTA. In order to obtain wild-type protein, the His-Tag was then cleaved, using AcTEV Protease (Invitrogen, USA) in accordance with the protocol provided by the supplier. The post-cleavage mixture was dialyzed against the A buffer, loaded onto the prepared and equilibrated column again and flow-through fraction was collected. This fraction containing SSB was first dialyzed against DD buffer (50 mM potassium phosphate buffer pH 7.5, 150 mM NaCl and 1 mM EDTA) and, after rapid dilution to a NaCl concentration of 50 mM, was then loaded onto an ssDNA-cellulose column (5 ml, USB, USA). The *Neq*SSB-like was eluted with 500 mM NaCl and 50% ethylene glycol and the elution fraction was again dialyzed against the DD buffer and concentrated to 1 mg/ml, using the Amicon Ultra-15 Filter Device MWCO 10000 (Millipore, USA). The Ultra-free-MC Centrifugal Filter Devices (Millipore, USA) with microporous membranes and pore sizes of 0.1 μm, were used to remove the particles, precipitates and huge aggregates. The purity of the *Neq*SSB-like was estimated using SDS-PAGE.

Estimation of the native molecular mass

The molecular mass of *Neq*SSB-like was estimated by means of analytical gel filtration chromatography carried out on a Superdex 75 10/300 GL column (Amersham Biosciences, USA) equilibrated with 20 mM Tris-HCl pH 7.5, 10 mM EDTA and using different concentrations of salt (10, 75, 150 and 300 mM NaCl). The samples containing various concentrations of purified protein (12–236 μM) were eluted with the same buffer, at a flow rate of 0.5 ml/min. The elution profile was monitored by recording the absorbance at 280 nm. The molecular weight of *Neq*SSB-like was determined by comparing the elution pattern with those of standard proteins, namely bovine albumin (66 kDa), ovalbumin (43 kDa), carbon anhydrase (29 kDa) and ribonuclease A (13 kDa). The void column volume was determined by monitoring the elution profile of blue dextran (2000 kDa).

Sedimentation experiments

Linear 15 to 30% (w/v) glycerol gradients containing loading buffer (50 mM Tris-HCl, pH 7.5, 0.5 M NaCl, 1 mM EDTA and 5 mM β-mercaptoethanol), were prepared in 5 ml Beckman centrifuge tubes. 50 μl of a 12 μM or 150 μM *Neq*SSB-like protein in loading buffer, and the corresponding amounts of standard proteins (lysozyme 14.3 kDa, carbon anhydrase 29 kDa, bovine albumin 66 kDa, alcohol dehydrogenase 150 kDa), were layered over 3.5 ml of the glycerol gradient and were centrifuged in individual tubes. Gradients were centrifuged at 4°C in a Beckman SW 60 rotor at 46,000 rpm for 24 h; fractions were collected from the top. The proteins present in the fractions were separated by SDS-PAGE and after densitometric analysis of the gels, the molecular mass of each protein versus the number of the fraction at which its maximal amount appears was represented.

Gel mobility shift assays: binding to ssDNA

Studies on ssDNA binding were performed using a fixed quantity (10 pmol) of 5'-end fluorescein-labelled oligonucleotides (dT)₃₅, (dT)₇₆ and (dT)₁₂₀ or M13 phage ssDNA (0.07 pmol).

The oligos were incubated with 10, 20, 40, 80, 160, 320 and 640 pmol of *Neq*SSB-like and the M13 ssDNA was incubated with 3.5, 7, 14, 28, 56, 112 and 224 pmol of *Neq*SSB-like for 10 minutes at 25°C in a binding buffer (20 mM Tris-HCl pH 7.5, 100 mM NaCl and 1 mM EDTA) to a final reaction volume of 20 µl. Subsequently the reaction products with oligos and the reaction products with M13 ssDNA were loaded onto 2% agarose gel, without ethidium bromide and with ethidium bromide, at a final concentration of 0.5 µg/ml, respectively and separated by electrophoresis in a TAE buffer (40 mM Tris acetate pH 7.5 and 1 mM EDTA). The bands corresponding to the unbound ssDNA and various *Neq*SSB-like-ssDNA complexes were visualized under UV light and photographed.

Gel mobility shift assays: binding to dsDNA

Tests on *Neq*SSB-like dsDNA binding properties were carried out with various DNA types. Specific 100 bp PCR product was obtained using the forward primer M13/pUCf 5' CCCAGT CACGACGTTGTAAAACG (23 nt) and the reverse primer pUC100r 5' catgattacgc caagcttgcatgc (24 nt). The PCR reaction solution consisted of 0.1 µg pUC19 DNA (Invitrogen, USA), 4 µl (10 µM) of each primer, 2.5 µl (10 mM) dNTPs, 2 µl (25 mM) MgCl₂ and 2.5 µl of 10 x Hot Start Buffer (200 mM Tris-HCl pH 8.3, 200 mM KCl and 50 mM (NH₄)₂SO₄), and 2 U of Maxima Hot Start *Taq* DNA Polymerase (Fermentas, Lithuania). 30 cycles were performed, using the Veriti 96 Well Thermal Cycler (Applied Biosystems, USA), with a temperature profile of 60 s at 94°C, 60 s at 57°C and 60 s at 72°C. pDONR201 (Invitrogen, USA) was used as the supercoiled DNA sample, pDONR201, linearized with *Nco*I, as the long, 4470 bp linear DNA sample and *Escherichia coli* genome DNA as the genomic DNA sample. To check the affinity for dsDNA, protein samples were incubated for 10 min at 25°C with fixed quantities of DNA, namely, 2.5 pmol of the 100 bp PCR product with 10, 20, 40, 80 and 160 pmol of *Neq*SSB-like, 0.132 fmol of *Escherichia coli* genome DNA, 0.2 pmol of pDONR201 DNA and mixture of 0.1 pmol pDONR201 plus 0.05 pmol pDONR201/*Nco*I with 10, 20, 40, 80, 160 and 320 pmol of *Neq*SSB-like. A control experiment with 10, 20, 40 pmol of *Taq*SSB protein and 0.2 pmol of pDONR201 DNA was also conducted. All the reaction products were separated, by means of electrophoresis in a TAE buffer, on 1% or 2% agarose gel, stained with ethidium bromide at a final concentration of 0.5 µg/ml, for the plasmid and genomic DNA and the 100 bp of product, respectively. The bands corresponding to unbound DNA and protein-DNA complexes were visualized under UV light and photographed.

Gel mobility shift assays: binding to mRNA

To verify *Neq*SSB-like mRNA binding properties, 10, 20, 40 and 80 pmol of protein were incubated for 10 minutes at 25°C with 2 µl, corresponding to 980 ng of mRNA, of RiboRuler Low Range RNA Ladder, ready-to-use, 100–1000 bases (Fermentas, Lithuania). Reactions were carried out in a binding buffer (20 mM Tris-HCl pH 7.5, 100 mM NaCl and 1 mM EDTA) to a final volume of 10 µl. The products were then loaded onto 2% agarose gel stained with ethidium bromide, at a final concentration of 0.5 µg/ml, and separated by means of electrophoresis in a TAE buffer. The bands obtained were visualized under UV light and photographed.

Binding preference studies

Studies into whether *Neq*SSB-like binds preferentially to ssDNA or to dsDNA were carried out using fixed quantities of single stranded oligo (dT)₇₆ (10 pmol) and the double stranded 100 bp PCR product (2.5 pmol). Nucleic acid samples were incubated for 10 min at 25°C with 10, 20, 40, 80, 160, and 320 pmol of *Neq*SSB-like in a binding buffer (20 mM Tris-HCl pH 7.5, 100 mM NaCl and 1 mM EDTA) to a final reaction volume of 20 µl. The reaction products were

loaded onto 2% agarose gel, stained with ethidium bromide, at a final concentration of 0.5 µg/ml and separated by means of electrophoresis in a TAE buffer. The bands obtained were visualized under UV light and photographed.

Fluorescence titration

Fluorescence spectroscopy measurements were used to determine the length of binding site and binding properties of *Neq*SSB-like precisely. The fluorescence titrations were made using a Perkin-Elmer LS-5B luminescence spectrometer, as described previously by Curth *et al.* [34]. The binding reactions were assembled in 2 ml of binding buffer (20 mM Tris-HCl pH 7.5 and 1 mM EDTA) containing 2, 100 and 500 mM NaCl and were incubated at 25°C. A constant quantity of *Neq*SSB-like (2 nM) was incubated in the appropriate buffer at 25°C with the amount of poly(dT) (Midland Certified Reagent, Midland, TX) being increased from 0 to 6 nM. The excitation and emission wavelengths were 295 and 348 nm, respectively. The binding curves obtained were analyzed using the model described by Schwarz and Watanabe [35], with n as the binding site size, $\omega \cdot K$ as the cooperative binding affinity and the fluorescence quench Q_f as the parameter. Q_f is defined as $(1 - F_{\text{bound}})/F_{\text{free}}$, where F_{bound} stands for the fluorescence intensity measured for nucleic acid bound protein and F_{free} stands for the fluorescence intensity measured for unbound protein.

Surface Plasmon Resonance (SPR) measurements

Biacore 3000 was used at a temperature of 25°C for the SPR measurements. The streptavidin coated chip (SA chip, GE Healthcare, USA) was first washed with 10 mM NaOH, 1 M NaCl to remove loose streptavidin. Biotinylated ssDNA 60-mer (5' – AATTCTGGGTGTGTGGGTGTGTGGGTGTGTGGGTGTGGTCAAGTTGACTACGTATACATC biotin-3') and dsDNA 60-mer (ssDNA 60-mer with complementary ssDNA) were bound at a level of 500 RU at two different channels, leaving flow cell number 1 as reference. The dsDNA 60-mer was prepared by hybridization of biotinylated ssDNA 60-mer with complementary ssDNA fragment and further agarose gel extraction in order to remove the unpaired ssDNA fragments). Different concentrations of *Neq*SSB-like were injected on the chip, with a flow rate of 30 µl/min, in a HBS buffer (10 mM Hepes, pH 7.4, 150 mM NaCl and 0.005% P20). The analyses of ssDNA and dsDNA interactions with *Neq*SSB-like were conducted in parallel, injecting the protein on the respective flow cells at the same time. Between injections, the flow cells were regenerated with 0.01% SDS. As a control experiment, the interaction between recombinant *Eco*SSB and ssDNA 60-mer was also studied in line with the protocol described for *Neq*SSB-like. The binding parameters for all settings were calculated with BiaEval 3.0 software.

Thermostability

The *Neq*SSB-like thermostability was determined using both the direct and indirect methods. For the indirect method, a fixed quantity (10 pmol) of 5'-end fluorescein-labelled oligonucleotide (dT)₃₅ was added to 20 pmol of *Neq*SSB-like or *Taq*SSB, the control protein, preincubated at, 85, 90, 95 and 100°C for 1, 3, 5, 10, 15, 30 and 60 min in a binding buffer (20 mM Tris-HCl pH 7.5, 100 mM NaCl and 1 mM EDTA) to a final reaction volume of 20 µl. After 10 min incubation at 25°C, the SSB-ssDNA complexes were separated from the unbound DNA by means of agarose gel electrophoresis in a TAE buffer on 2% gel.

For the direct method, Differential Scanning Calorimetry (DSC) was used. The measurements were performed using a NanoDSC microcalorimeter (Calorimetry Science Corporation, USA). The *Neq*SSB-like sample, concentrated to 1 mg/ml in 50 mM of potassium phosphate

buffer pH 7.5 and 150 mM NaCl was analyzed, by calorimetric scan, between 15 and 120°C, with a scan rate of 1°C/min. The reversibility of the heat unfolding transition was tested by heating and cooling the same sample with a scan rate of 1°C/min. The results were analyzed with the NanoAnalyze Software V 1.1 (TA Instruments, USA).

Complementation assay

The *E. coli* RDP268(DE3) strain harboring pRPZ146 (ColEI *ori*, Tc^R) plasmid coding wild-type *EcoSSB* protein [32] was used in complementation assay. In this strain expression of target genes cloned in the pET vectors under the control of the T7 promoter is possible and the chromosomal *ssb* gene is replaced by a kanamycin resistance ($\Delta ssb::Kan^R$). The pRPZ146 is essential for the survival of the cells and can be replaced by another plasmid, only if it contains a gene whose product can take over *EcoSSB* function *in vivo* [33]. The *E. coli* RDP268(DE3)/pRPZ146 cells transformed with pET23D(+)/NeqSSB-like plasmid (data not shown) were grown in the presence of ampicillin (100 µg/ml) and kanamycin (25 µg/ml)—present during all the steps. After six consecutive overnight subculturings in 3 ml of 2YT (1.6% tryptone, 1% yeast extract and 0.5% NaCl with or without 1 mM IPTG) containing the same antibiotics, the colonies were patched on 2YT agar plates containing ampicillin alone (100 µg/ml) and on plates containing both tetracycline (25 µg/ml) and ampicillin (100 µg/ml). At the same time, plasmids were isolated from the cultures to estimate the size of plasmids present inside the cells (parallel to inoculating the plates). An Amp^R and a Tc^S phenotype shows that the incoming plasmid harbors a gene that complements *EcoSSB*.

Acknowledgments

We kindly thank Dr. Harald Huber (Institute for Microbiology, University of Regensburg, Germany) for the *Nanoarchaeum equitans* Kin4-M genome DNA used in this study.

Author Contributions

Conceived and designed the experiments: MO. Performed the experiments: MO JB MN NM ACC BZP RP. Analyzed the data: MO JB JK. Contributed reagents/materials/analysis tools: MO. Wrote the paper: MO JB JK.

References

1. Huber H, Hohn MJ, Rachel R, Fuchs T, Wimmer VC, Stetter KO. A new phylum of *Archaea* represented by a nanosized hyperthermophilic symbiont. *Nature*. 2002 May 2; 417(6884):63–7. PMID: [11986665](#)
2. Nakabachi A, Yamashita A, Toh H, Ishikawa H, Dunbar HE, Moran NA, et al. The 160-kilobase genome of the bacterial endosymbiont *Carsonella*. *Science*. 2006 Oct 13; 314(5797):267. PMID: [17038615](#)
3. Waters E, Hohn MJ, Ahel I, Graham DE, Adams MD, Barnstead M, et al. The genome of *Nanoarchaeum equitans*: insights into early archaeal evolution and derived parasitism. *Proc Natl Acad Sci U S A*. 2003 Oct 28; 100(22):12984–8. PMID: [14566062](#)
4. Greipel J, Urbanke C, Maass G. The single-stranded DNA binding protein of *Escherichia coli*. Physico-chemical properties and biological functions. In: Saenger W, Heinemann U, editors. *Protein-Nucleic Acid Interaction*. London: Macmillan; 1989. p. 61–86.
5. Alani E, Thresher R, Griffith JD, Kolodner RD. Characterization of DNA-binding and strand-exchange stimulation properties of γ -RPA, a yeast single-strand-DNA-binding protein. *J Mol Biol*. 1992 Sep 5; 227(1):54–71. PMID: [1522601](#)
6. Lohman TM, Overman LB. Two binding modes in *Escherichia coli* single strand binding protein-single stranded DNA complexes. Modulation by NaCl concentration. *J Biol Chem*. 1985 Mar 25; 260(6):3594–603. PMID: [3882711](#)
7. Meyer RR, Laine PS. The single-stranded DNA-binding protein of *Escherichia coli*. *Microbiol Rev*. 1990 Dec; 54(4):342–80. PMID: [2087220](#)

8. Shereda RD, Kozlov AG, Lohman TM, Cox MM, Keck JL. SSB as an organizer/mobilizer of genome maintenance complexes. *Crit Rev Biochem Mol Biol*. 2008 Sep-Oct; 43(5):289–318. doi: [10.1080/10409230802341296](https://doi.org/10.1080/10409230802341296) PMID: [18937104](https://pubmed.ncbi.nlm.nih.gov/18937104/)
9. Murzin AG. OB(oligonucleotide/oligosaccharide binding)-fold: common structural and functional solution for non-homologous sequences. *EMBO J*. 1993 Mar; 12(3):861–7. PMID: [8458342](https://pubmed.ncbi.nlm.nih.gov/8458342/)
10. Weiner J, Bertsch L, Kornberg A. The deoxyribonucleic acid unwinding protein of *Escherichia coli*. Properties and functions in replication. *J Biol Chem*. 1975 Mar 25; 250(6):1972–80. PMID: [1090613](https://pubmed.ncbi.nlm.nih.gov/1090613/)
11. Purnapatre K, Varshney U. Cloning, over-expression and biochemical characterization of the single-stranded DNA binding protein from *Mycobacterium tuberculosis*. *Eur J Biochem*. 1999 Sep; 264(2):591–8. PMID: [10491108](https://pubmed.ncbi.nlm.nih.gov/10491108/)
12. Olszewski M, Mickiewicz M, Kur J. Two highly thermostable paralogous single-stranded DNA-binding proteins from *Thermoanaerobacter tengcongensis*. *Arch Microbiol*. 2008 Jul; 190(1):79–87. doi: [10.1007/s00203-008-0366-6](https://doi.org/10.1007/s00203-008-0366-6) PMID: [18392610](https://pubmed.ncbi.nlm.nih.gov/18392610/)
13. Olszewski M, Grot A, Wojciechowski M, Nowak M, Mickiewicz M, Kur J. Characterization of exceptionally thermostable single-stranded DNA-binding proteins from *Thermotoga maritima* and *Thermotoga neapolitana*. *BMC Microbiol*. 2010 Oct 15; 10:260. doi: [10.1186/1471-2180-10-260](https://doi.org/10.1186/1471-2180-10-260) PMID: [20950419](https://pubmed.ncbi.nlm.nih.gov/20950419/)
14. Dąbrowski S, Olszewski M, Piątek R, Brillowska-Dąbrowska A, Konopa G, Kur J. Identification and characterization of single-stranded-DNA-binding proteins from *Thermus thermophilus* and *Thermus aquaticus*—new arrangement of binding domains. *Microbiology*. 2002 Oct; 148(Pt 10):3307–15. PMID: [12368464](https://pubmed.ncbi.nlm.nih.gov/12368464/)
15. Eggington JM, Haruta N, Wood EA, Cox MM. The single-stranded DNA-binding protein of *Deinococcus radiodurans*. *BMC Microbiol*. 2004 Jan 12; 4:2. PMID: [14718065](https://pubmed.ncbi.nlm.nih.gov/14718065/)
16. Filipkowski P, Duraj-Thatte A, Kur J. Novel thermostable single-stranded DNA-binding protein (SSB) from *Deinococcus geothermalis*. *Arch Microbiol*. 2006 Aug; 186(2):129–37. PMID: [16802171](https://pubmed.ncbi.nlm.nih.gov/16802171/)
17. Wold MS. Replication protein A: a heterotrimeric, single-stranded DNA-binding protein required for eukaryotic DNA metabolism. *Annu Rev Biochem*. 1997; 66:61–92. PMID: [9242902](https://pubmed.ncbi.nlm.nih.gov/9242902/)
18. Wadsworth RI, White MF. Identification and properties of the crenarchaeal single-stranded DNA binding protein from *Sulfolobus solfataricus*. *Nucleic Acids Res*. 2001 Feb 15; 29(4):914–20. PMID: [11160923](https://pubmed.ncbi.nlm.nih.gov/11160923/)
19. Haseltine CA, Kowalczykowski SC. A distinctive single-strand DNA-binding protein from the Archaeon *Sulfolobus solfataricus*. *Mol Microbiol*. 2002 Mar; 43(6):1505–15. PMID: [11971263](https://pubmed.ncbi.nlm.nih.gov/11971263/)
20. Rolfsmeier ML, Haseltine CA. The single-stranded DNA binding protein of *Sulfolobus solfataricus* acts in the presynaptic step of homologous recombination. *J Mol Biol*. 2010 Mar 19; 397(1):31–45. doi: [10.1016/j.jmb.2010.01.004](https://doi.org/10.1016/j.jmb.2010.01.004) PMID: [20080104](https://pubmed.ncbi.nlm.nih.gov/20080104/)
21. Kelly TJ, Simanek P, Brush GS. Identification and characterization of a single-stranded DNA-binding protein from the archaeon *Methanococcus jannaschii*. *Proc Natl Acad Sci U S A*. 1998 Dec 8; 95(25):14634–9. PMID: [9843941](https://pubmed.ncbi.nlm.nih.gov/9843941/)
22. Komori K, Ishino Y. Replication protein A in *Pyrococcus furiosus* is involved in homologous DNA recombination. *J Biol Chem*. 2001 Jul 13; 276(28):25654–60. PMID: [11342551](https://pubmed.ncbi.nlm.nih.gov/11342551/)
23. Capp C, Qian Y, Sage H, Huber H, Hsieh TS. Separate and combined biochemical activities of the subunits of a naturally split reverse gyrase. *J Biol Chem*. 2010 Dec 17; 285(51):39637–45. doi: [10.1074/jbc.M110.173989](https://doi.org/10.1074/jbc.M110.173989) PMID: [20929866](https://pubmed.ncbi.nlm.nih.gov/20929866/)
24. Randau L, Calvin K, Hall M, Yuan J, Podar M, Li H, et al. The heteromeric *Nanoarchaeum equitans* splicing endonuclease cleaves noncanonical bulge-helix-bulge motifs of joined tRNA halves. *Proc Natl Acad Sci U S A*. 2005 Dec 13; 102(50):17934–9. PMID: [16330750](https://pubmed.ncbi.nlm.nih.gov/16330750/)
25. Rodrigues JV, Victor BL, Huber H, Saraiva LM, Soares CM, Cabelli DE, et al. Superoxide reduction by *Nanoarchaeum equitans* neelaredoxin, an enzyme lacking the highly conserved glutamate iron ligand. *J Biol Inorg Chem*. 2008 Feb; 13(2):219–28. PMID: [17968598](https://pubmed.ncbi.nlm.nih.gov/17968598/)
26. Choi JJ, Nam KH, Min B, Kim SJ, Soll D, Kwon ST. Protein trans-splicing and characterization of a split family B-type DNA polymerase from the hyperthermophilic archaeal parasite *Nanoarchaeum equitans*. *J Mol Biol*. 2006 Mar 10; 356(5):1093–106. PMID: [16412462](https://pubmed.ncbi.nlm.nih.gov/16412462/)
27. Choi JJ, Song JG, Nam KH, Lee JI, Bae H, Kim GA, et al. Unique substrate spectrum and PCR application of *Nanoarchaeum equitans* family B DNA polymerase. *Appl Environ Microbiol*. 2008 Nov; 74(21):6563–9. doi: [10.1128/AEM.00624-08](https://doi.org/10.1128/AEM.00624-08) PMID: [18791030](https://pubmed.ncbi.nlm.nih.gov/18791030/)
28. Kerr ID, Wadsworth RI, Cubeddu L, Blankenfeldt W, Naismith JH, White MF. Insights into ssDNA recognition by the OB fold from a structural and thermodynamic study of *Sulfolobus* SSB protein. *EMBO J*. 2003 Jun 2; 22(11):2561–70. PMID: [12773373](https://pubmed.ncbi.nlm.nih.gov/12773373/)
29. Fuller RS, Funnell BE, Kornberg A. The dnaA protein complex with the *E. coli* chromosomal replication origin (oriC) and other DNA sites. *Cell*. 1984 Oct; 38(3):889–900. PMID: [6091903](https://pubmed.ncbi.nlm.nih.gov/6091903/)

30. Reddy M, Guhan N, Muniyappa K. Characterization of single-stranded DNA-binding proteins from Mycobacteria. The carboxyl-terminal of domain of SSB is essential for stable association with its cognate RecA protein. *J Biol Chem*. 2001 Dec 7; 276(49):45959–68. PMID: [11577073](#)
31. Ehn M, Nilsson P, Uhlén M, Hober S. Overexpression, rapid isolation, and biochemical characterization of *Escherichia coli* single-stranded DNA-binding protein. *Protein Expr Purif*. 2001 Jun; 22(1):120–7. PMID: [11388809](#)
32. Porter RD, Black S, Pannuri S, Carlson A. Use of the *Escherichia coli* SSB gene to prevent bioreactor takeover by plasmidless cells. *Biotechnology (N Y)*. 1990 Jan; 8(1):47–51. PMID: [1366512](#)
33. Filipkowski P, Kur J. Identification and properties of the *Deinococcus grandis* and *Deinococcus proteolyticus* single-stranded DNA binding proteins (SSB). *Acta Biochim Pol*. 2007; 54(1):79–87. PMID: [17325748](#)
34. Curth U, Greipel J, Urbanke C, Maass G. Multiple binding modes of the single-stranded DNA binding protein from *Escherichia coli* as detected by tryptophan fluorescence and site-directed mutagenesis. *Biochemistry*. 1993 Mar 16; 32(10):2585–91. PMID: [8448116](#)
35. Schwarz G, Watanabe F. Thermodynamics and kinetics of co-operative protein-nucleic acid binding. I. General aspects of analysis of data. *J Mol Biol*. 1983 Jan 25; 163(3):467–84. PMID: [6834432](#)