



Seasonal Variation of Microbial Diversity of Coastal Sediment in Tongyeong, South Korea, Using 16S rRNA Gene Amplicon Sequencing

 Nur Indradewi Oktavetri,^{a,b} Jong-Oh Kim,^c  Kyunghoi Kim^a

^aDepartment of Ocean Engineering, Pukyong National University, Busan, Republic of Korea

^bEnvironmental Engineering, Faculty of Science and Technology, Universitas Airlangga, Surabaya, Indonesia

^cInstitute of Marine Biotechnology, Pukyong National University, Busan, Republic of Korea

ABSTRACT Benthic microbial diversity in Tongyeong, South Korea, was analyzed using next-generation sequencing of the 16S rRNA genes, to reveal the effects of seasonal variations on the microbial community in sediment. *Proteobacteria* was the dominant phylum, with a relative abundance of 61.5 to 68.1%.

Predicting the distribution and abundance of bacteria as a response to ecosystem changes is important (1). Suh et al. (2) reported that the changing of the seasons affects the abundance of microbes in the water mass in the South Sea of South Korea. Microbial classes in the water masses in winter and spring were different than those in summer and autumn (2).

The conditions in the semienclosed bay at Tongyeong, South Korea, are interesting to study; the water is shallow, and the water exchange is slow (3). Seasonal effects on the semi-enclosed bay cause problems such as red tide and hypoxia (4). Although environmental problems in Tongyeong Bay are serious, studies on the microbial diversity in sediment are lacking. An exploration of the taxonomic diversity with high-throughput pyrosequencing techniques showed the distribution and abundance of marine bacteria (2). Thus, we investigated the microbial diversity of Tongyeong Bay sediment through the sequencing of 16S rRNA genes by using high-throughput sequencing.

Sampling was conducted in April, August, October, and December 2019, to represent seasonal data for spring, summer, autumn, and winter, respectively. Sediments were collected at a 20-cm depth from the surface of the sediment in Tongyeong Bay (34°47.4420'N, 128°25.5700'E) using a grab sampler. The sediment samples were immediately transferred to the laboratory using an ice box at 4°C and were stored at −20°C until DNA extraction. Samples were homogenized, and total genomic DNA was extracted using the DNeasy PowerMax soil kit (Qiagen). Library preparation was performed with the Herculase II Fusion DNA polymerase and Nextera XT index kit v2, using Bakt_341F and Bakt_805R primers (5, 6) to amplify the V3 to V4 region of the 16S rRNA gene. The prepared libraries were sequenced using the Illumina MiSeq platform at Macrogen, Inc. (South Korea). Raw reads were paired-end

TABLE 1 Summary description of the bioinformatic process

| Month | No. of input reads | No. of filtered reads | No. of denoised forward reads | No. of denoised reverse reads | No. of merged reads | No. of nonchimeric reads |
|----------|--------------------|-----------------------|-------------------------------|-------------------------------|---------------------|--------------------------|
| April | 174,306 | 95,893 | 71,036 | 68,274 | 22,055 | 11,257 |
| August | 182,753 | 112,555 | 86,452 | 88,083 | 33,611 | 16,633 |
| October | 116,328 | 97,399 | 70,867 | 78,484 | 29,823 | 18,482 |
| December | 112,631 | 90,984 | 69,757 | 74,367 | 29,917 | 18,106 |

Citation Oktavetri NI, Kim J-O, Kim K. 2021. Seasonal variation of microbial diversity of coastal sediment in Tongyeong, South Korea, using 16S rRNA gene amplicon sequencing. *Microbiol Resour Announc* 10:e00446-21. <https://doi.org/10.1128/MRA.00446-21>.

Editor Frank J. Stewart, Montana State University

Copyright © 2021 Oktavetri et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Kyunghoi Kim, hoikim@pknu.ac.kr.

Received 6 May 2021

Accepted 8 June 2021

Published 8 July 2021

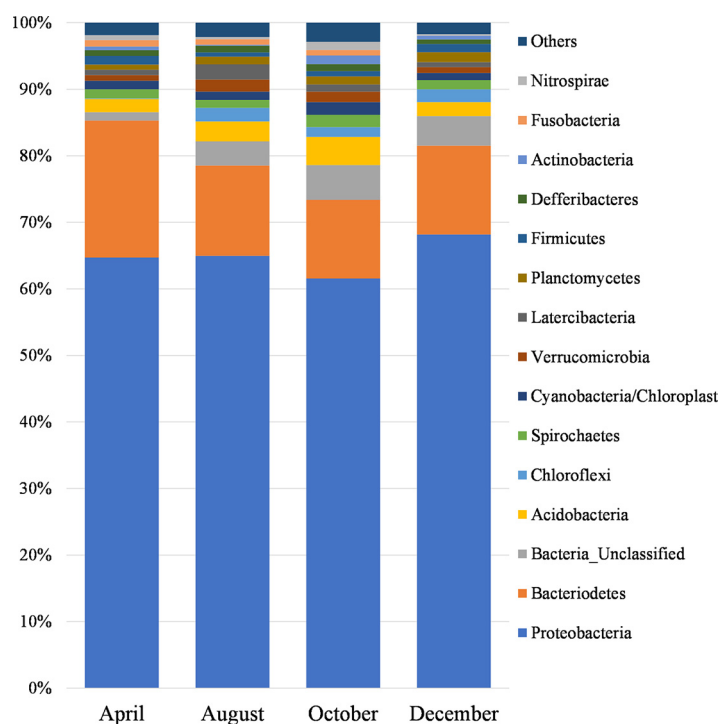


FIG 1 Top 15 microbial phyla in relative abundance in different months. Each color represents a different phylum.

merged using FLASH v1.2.11 (7). Denoising strategies were applied to obtain amplicon sequence variants (ASVs) by Divisive Amplicon Denoising Algorithm 2 (DADA2) v1.16.1 (8) in R v4.0. Before the denoising analysis, both primer sequences were removed using Cutadapt (9). The standard processing steps in the DADA2 workflow were performed, including quality filtering [maxEE = c(2,5)], dereplication, learning the data set-specific error model, ASV inference, and chimera removal. The naive Bayesian classifier (10) method was implemented for taxonomic assignment using the Ribosomal Database Project (RDP) training set 18 database (11).

A description of the bioinformatic process is presented in Table 1. Analysis of the microbial diversity in Tongyeong Bay showed that *Proteobacteria* was the most abundant phylum, with a relative abundance of 61.5 to 68.1%, followed by *Bacteroidetes* (11.8 to 20.6%), unclassified bacteria (1.3 to 5.2%), and *Acidobacteria* (2.0 to 4.3%). (Fig. 1). *Proteobacteria* contributed more than one-half of the biomass of bacterial phyla in most surface marine sediments (12). *Proteobacteria* played a role in nutrient and organic matter decomposition in eutrophicated (13, 14) and polluted marine fish farm (15) sediments. Our results provide useful information on the microbial community and may facilitate environmental remediation for Tongyeong Bay.

Data availability. The 16S rRNA gene amplicon sequences obtained in this study have been deposited in the NCBI Sequence Read Archive (SRA) under the accession number [PRJNA712535](https://www.ncbi.nlm.nih.gov/sra/PRJNA712535).

ACKNOWLEDGMENT

This research was supported by a research grant from Pukyong National University (2019).

REFERENCES

1. Chapin FS, Zavaleta ES, Eviner VT, Naylor RL, Vitousek PM, Reynolds HL, Hooper DU, Lavorel S, Sala OE, Hobbie SE, Mack MC, Diaz S. 2000. Consequences of changing biodiversity. *Nature* 405:234–242. <https://doi.org/10.1038/35012241>.
2. Suh S-S, Park M, Hwang J, Kil E-J, Jung SW, Lee S, Lee T-K. 2015. Seasonal dynamics of marine microbial community in the South Sea of Korea. *PLoS One* 10:e0131633. <https://doi.org/10.1371/journal.pone.0131633>.

3. Kim S-H, Kim HC, Choi S-H, Lee W-C, Jung R-H, Hyun J-H, Kim SH, Lee JS. 2020. Benthic respiration and nutrient release associated with net cage fish and longline oyster aquaculture in the Geoje-Tongyeong coastal waters in Korea. *Estuaries Coasts* 43:589–601. <https://doi.org/10.1007/s12237-019-00567-5>.
4. Tanaka Y. 2013. A study on the seasonal variation of water exchange in a semi-enclosed bay. *J Jpn Soc Civil Eng* 69:1061–1065. https://doi.org/10.2208/kaigan.69.i_1061.
5. Herlemann DPR, Labrenz M, Jürgens K, Bertilsson S, Waniek JJ, Andersson AF. 2011. Transitions in bacterial communities along the 2000 km salinity gradient of the Baltic Sea. *ISME J* 5:1571–1579. <https://doi.org/10.1038/ismej.2011.41>.
6. Lee J, Jeong I, Kim J-O, Kim K. 2021. Microbial diversity analysis of sediment from Yeosu New Harbor of South Korea using 16S rRNA gene amplicon sequencing. *Microbiol Resour Announc* 10:e01229-20. <https://doi.org/10.1128/MRA.01229-20>.
7. Magoč T, Salzberg SL. 2011. FLASH: fast length adjustment of short reads to improve genome assemblies. *Bioinformatics* 27:2957–2963. <https://doi.org/10.1093/bioinformatics/btr507>.
8. Callahan BJ, McMurdie PJ, Rosen MJ, Han AW, Johnson AJA, Holmes SP. 2016. DADA2: high-resolution sample inference from Illumina amplicon data. *Nat Methods* 13:581–583. <https://doi.org/10.1038/nmeth.3869>.
9. Martin M. 2011. Cutadapt removes adapter sequences from high-throughput sequencing reads. *EMBnet J* 17:10–12. <https://doi.org/10.14806/ej.17.1.200>.
10. Wang Q, Garrity GM, Tiedje JM, Cole JR. 2007. Naïve Bayesian classifier for rapid assignment of rRNA sequences into the new bacterial taxonomy. *Appl Environ Microbiol* 73:5261–5267. <https://doi.org/10.1128/AEM.00062-07>.
11. Cole JR, Wang Q, Fish JA, Chai B, McGarrell DM, Sun Y, Brown CT, Porras-Alfaro A, Kuske CR, Tiedje JM. 2014. Ribosomal Database Project: data and tools for high throughput rRNA analysis. *Nucleic Acids Res* 42:D633–D642. <https://doi.org/10.1093/nar/gkt1244>.
12. Bowman JP, McCammon SA, Dann AL. 2005. Biogeographic and quantitative analyses of abundant uncultivated γ -proteobacterial clades from marine sediment. *Microb Ecol* 49:451–460. <https://doi.org/10.1007/s00248-004-0070-2>.
13. Huang W, Chen X, Jiang X, Zheng B. 2017. Characterization of sediment bacterial communities in plain lakes with different trophic statuses. *Microbiologyopen* 6:e00503. <https://doi.org/10.1002/mbo3.503>.
14. Bai Y, Shi Q, Wen D, Li Z, Jefferson WA, Feng C, Tang X. 2012. Bacterial communities in the sediments of Dianchi Lake, a partitioned eutrophic waterbody in China. *PLoS One* 7:e37796. <https://doi.org/10.1371/journal.pone.0037796>.
15. McCaig AE, Phillips CJ, Stephen JR, Kowalchuk GA, Harvey SM, Herbert RA, Embley TM, Prosser JI. 1999. Nitrogen cycling and community structure of proteobacterial β -subgroup ammonia-oxidizing bacteria within polluted marine fish farm sediments. *Appl Environ Microbiol* 65:213–220. <https://doi.org/10.1128/AEM.65.1.213-220.1999>.