Commentary

For reprint orders, please contact: reprints@futuremedicine.com

Potential alteration of COVID-19 by beta-mercaptoethanol

Robert E Click*,1

¹Altick Associates, 2000 Maxwell Drive, Hudson, WI 54016, USA *Author for correspondence: mbclicker@outlook.com

******Based on many group's attitude to disregard preventative suggestions of infectious disease experts, along with the relaxation of government regulations, mini-herd immunity has essentially already begun.*****

First draft submitted: 4 June 2020; Accepted for publication: 21 August 2020; Published online: 21 September 2020

Keywords: covid-19 • herd immunity • beta-mercaptoethanol (BME) • T- and B-cell enhanced functions • suppressor cell inactivation

When and how the COVID-19 pandemic caused by SARS-CoV-2 ends is the topic of substantial debate. Potential options to return societies back to a somewhat normal lifestyle appear to depend upon achieving one of the following: develop a vaccine; discover a drug that will disrupt the viruses' survival/reproductive processes; and a combination of the other two. Development of any of these may require a considerable amount of time without any assurances. Hardly an inevitable situation.

While following the chatter on TV and internet, an interview with Harvard's Professor Lipsitch, on the concept of 'herd immunity' was very intriguing. Almost immediately I pondered: Why is this concept not more seriously considered as a means to combat COVID-19? Based on many group's attitude to disregard preventative suggestions of infectious disease experts, along with the relaxation of government regulations, mini-herd immunity has essentially already begun. From an outsider's perspective, such an approach: would take considerably less time to implement than that projected to develop a vaccine; may not require isolation of virus or viral-constituents; and if the severity between initial infection and pathological consequences were minimized/curtailed, the death rate would likely be considerably less. It could be argued that now is a perfect opportunity to explore the means of curtailing the severity of accompanying pathological events.

A search of the literature for a magic, antiseverity protocol resulted in multiple articles that described means to alter various aspects of viral diseases. One especially interesting group is composed of antioxidant, low-molecular-weight (LMW) thiols; the most thoroughly studied for human use is *N*-acetylcysteine (NAC) and to a much lesser extent, beta-mercaptoethanol (BME). In one study, the proliferation of *in vitro*, nonstimulated, T-cell colonies present in PBMC of 'normal' (186%) and AIDS (234%) individuals was equally enhanced in the presence of NAC; results that contrasted to the greater increase in colonies from AIDS (401%) than normal (168%) individuals when BME replaced NAC in the culture medium. The difference in growth of AIDS cells was attributed to BME more effectively inactivating suppressor cells because plasma of AIDS patients equally inhibited growth of control T cells in the presence (48%) or absence (52%) of BME [1]. In a separate study [2], pretreatment of anergic CD4⁺ cells of HIV-infected patients with a NAC/BME admixture enhanced the CD3-mAb stimulated responses of 'high', but not 'low', responders. Additional investigations by others indicated that: NAC therapy of HIV-infected individuals improved their survival, which was associated with reversal of T-cell glutathione deficiencies [3]; modulation of cell surface redox events suppressed the cellular entry of HIV [4]; intracellular HIV replication was suppressed by NAC [5,6]; and inhibitory factors produced by AIDS T cells that specifically blocked B-cell antibody production *in vitro* were inactivated by BME [7].

It was somewhat surprising that a survey in April 2020 on behalf of the Oxford COVID-19 Evidence Service Team, failed to find any evidence that indicated NAC possessed COVID-19 specific benefits [8]. However, it is easy to argue that it is premature for data from any such study (if it exists) to be published. Indeed, since submission of



Future

CROBIOLOGY

the present manuscript, a perspective was published on why NAC should be considered as a therapeutic option for COVID-19 [9]. Other recent information indicates that among 827 COVID-19 patients, 76% had low total, 76% low CD4⁺ and 72% low CD8⁺ T-cell counts [10]. Further, the CD4⁺ and CD8⁺ T-cell counts were lower in ICU than non-ICU patients suggesting that their T cells were exhausted. The authors concluded [10]: "Taken together, new therapeutic measures are needed for treatment of COVID-19 ICU patients." Considering that both intracellular and cell surface thiols of adaptive and innate immune cells are modifiable by external LMW thiols [11,12], new therapeutic areas of interest may include: alteration of undetectable antigenic epitopes; increase insufficient effector cells (precursor or nonexhausted) to overcome suppressor cell functions; strengthen TCR-peptide/MHC interactions; modify poor costimulatory signals; Ir-genes; and others [13-16]. The present report will summarize data that indicates BME is a better thiol choice than NAC, because it is more potent, has broader pleotropic effects, requires up to a 100-fold lower dose, is recycled without being modified or biologically incorporated, and lacks some of the adverse insults of NAC [17,18]. These attributes suggest that it is time to assess whether BME has any benefits for human diseases irrespective of its designation as a 'poison'. The benefits of daily consumption of mg of BME for entire lives of many different strains of mice by three different research groups [19-24] suggest that functional levels of BME are not poisonous and (as yet) have not been shown to cause any serious side effects. The present COVID-19 pandemic offers an ideal opportunity to test BME for safety/benefits in humans as dosage, formulation and packaging are established. Furthermore, considering the reality of a COVID-19 vaccine, one need only hope that the experience with AIDS 'vaccines' will not be encountered with COVID-19. Such a possibility should encourage multiple, including unconventional, investigations.

BME alterations of immune functions

Until recently, modes which alter immune processes have not been very successful as therapeutic cancer treatments. With the discovery of immune checkpoint inhibitors, this lull seems to be evaporating. To gain an appreciation of BME benefits, a short history should be enlightening as it relates to the present-day pandemic.

In vitro

Numerous investigations on adaptive immunity followed the initial reports in the '70s that both humoral [25–27] and cell mediated [28–31] responses *in vitro* were enhanced multifold by LMW thiols, with BME and dithiothreitol (DTT) being the most potent. Similar antibody enhancements were described soon thereafter for cysteamine [32] and α -thioglycerol (α -TG) [33]. In some cases, BME enhanced primary antibody synthesis >100-fold [25,26] and proliferation/differentiation of primary and secondary alloantigen specific cytotoxic T lymphocytes >50-fold [34,35]. Over the ensuing years, alterations of *in vitro* processes by BME were confirmed and extended by hundreds of investigators. The magnitude of enhancements by, plus dosages of, these thiols differed enormously regardless of whether they were added to serum-free culture medium or medium supplemented with homologous or heterologous sera. The most potent, BME, cysteamine, DTT and a-TG, are unique in that they are **not** endogenous bioconstituents. Further, the reduced and oxidized forms of BME, cysteamine and DTT increased the number of sRBC antibody producing B cells equally [32]; the mechanism of DTT, however, differed from that of BME and cysteamine in that enhancement was via T-cell activation of B cells rather than direct activation of B cells. This mechanism was similar to that of lipoic acid [36] and NAC [37] in that none facilitated uptake of extracellular cystine.

In addition to the importance of adaptive immunity, innate type I and II NK cells are also major participants in viral infections and cancer [38]. NK cells can be activated by a number of receptors, including NKG2D, a disulfide-linked homodimer that recognizes a number of ligands [39]. It is not surprising, therefore that cell surface thiols covalently coupled with the nonpenetrating reagent, monobromotrimethyl-ammoniobimane, prevented a line of cloned, murine, NK cells to bind to and lyse target cells [11]. Subsequently, this was extended to include a thiol role for lytic processes of human NK cells [40]. Further, an *in vitro* requirement for BME was found for induction of lymphokine-activated killer cell activity of rat cultures [41] and for enriched human NK cells [42]. In the latter, both thiol deprivation and H_2O_2 treatment resulted in oxidative stress that increased intracellular ROS levels. This altered internalization of IL-2 and maturation of the lytic mechanism by inhibition of *FasL* induction. Based on these *in vitro* results the authors hypothesized that thiol-reducing compounds may be a physiological **requirement** for maintenance of optimal NK functions under *in vivo* oxidative conditions [42–44]. A further effect of BME was its requirement for NK cell generation in nonconditioned, long-term bone marrow cultures directly from enriched NK precursors present in rat marrow [45].

In vivo

As might be expected from the extensive literature on LMW thiol alterations of immune responses *in vitro*, investigations on *in vivo* disease processes soon followed. One of the first areas to be investigated was age-associated, declines of both primary and secondary antibody synthesis, which were slowed/reversed by *in vitro* [46–51] or *in vivo* [19,20] 'BME-therapy'. The declines of these adaptive immune responses were caused by increases in nonadherent suppressive T cells [46,47]. BME [52] and glutathione [53] also reversed *in vitro*, the age associated, declines of immune functions of rats. Makinodan, collaborators and others reported that innate immune functions of mice also declined with age; declines postulated to be a consequence of multiple factors that included: the absolute loss of NK cells due to reduced generation in the bone marrow; a decreased capacity of NK cells to bind to target cells; and increased adherent cell suppressor functions [54–56] – the latter cells differed from those that suppressed antibody synthesis [46,47]. Further, NK functions as well as immune responses of old mice were enhanced by other thiols added to their feed [57,58], including NAC and thioproline (BME was not tested). Stimulation by more classic antioxidants, alpha-tocopherol (vitamin E) and ascorbic acid, was less than those by the two sulfur containing antioxidants [57].

Role of cysteine

Finally, many functions altered by exogenous LMW thiols (but not lipoic acid or NAC) are directly dependent upon the availability of cysteine/cystine. This dependence was first noted when BME enhancement of primary antibody formation to sRBC did not occur when cystine was omitted from the culture medium [59]. There was a specific LMW thiol rank-order in uptake of (³⁵S)-cystine by splenocytes – the largest was by BME, followed by DTT and lastly by cysteamine [60]. Not surprising, the increase in the number of specific IgM producing B cells followed the same rank order. A steady state of replenishment of intracellular cysteine by BME was reached within 24 h by the two–fivefold enhanced rate of uptake [59]; a consequence of nonenzymatic formation of a mixed disulfide [60] taken up mainly via the L system, a transporter system for neutral alpha-amino acids such as leucine. The intracellular, mixed disulfide underwent rapid reduction to cysteine and BME; the latter escaped into the extracellular fluids where it was available to continue the cycle.

Depletion of intracellular cysteine was not unexpected since lymphocytes lack cystathionase, which converts methionine to cysteine and they lack an intact xc-transporter system to directly acquire cystine, the prominent bioform, from liquid surroundings. Under such constraints, cysteine replacement is dependent upon the transport by the alanine, serine, cysteine (ASC) system of minuscule amounts not yet oxidized to the more stable disulfide form, cystine. These inadequacies are in part compensated for by uptake of cystine by APC/dendritic cells (both possess an intact xc-transporter system) which is reduced and exported as cysteine. Unfortunately, myeloid-derived suppressor cells further exacerbate availability by sequestering cystine via their xc-transporter system, **but** because they lack the ASC transporter, do not export cysteine [61,62]. Consequently, the lowered amount of cystine available for conversion to cysteine by APC/dendritic cells results in less extracellular cysteine in the surrounding milieu [37,61–66]. Failure to adequately replenish intracellular cysteine, the rate limiting component for GSH synthesis during lymphocytic stimulation, also resulted in depletion of glutathione (GSH). The consequence was lymphocytes became unable to proliferate (exhaustion?) to further stimulation. More recent findings added a significant modification, namely neither depleted glutathione nor cysteine alone is the cause of reduced T-cell proliferation, but they become rate limiting only in the absence of endogenous LMW thiols [67].

Summary

Results presented herein demonstrating BME alters immune functions at very low concentrations, suggest it may safely curtail the severity of COVID-19 disease and prevent deaths by enhancement of antiviral, immune responses while also blocking suppressor cell activities; a hypothesis that should be easy to assess in a highly populated herd immune environment, such as a meat processing company or a nursing home, that would satisfy appropriate statistical validation. Input is encouraged from anyone with clinical trial expertise wishing to assess benefits of BME on COVID-19 disease.

Author contributions

RE Click was the sole contributor to this article.

Financial & competing interests disclosure

The author has no relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript. This includes employment, consultancies, honoraria, stock ownership or options, expert testimony, grants or patents received or pending, or royalties.

No writing assistance was utilized in the production of this manuscript.

References

- Wu J, Levy EM, Black PH. 2-Mercaptoethanol and N-acetylcysteine enhance T cell colony formation in AIDS and ARC. *Clin. Exp. Immunol.* 77(1), 7–10 (1989).
- Cayota A, Vuillier F, Gonzalez G, Dighiero G. In vitro antioxidant treatment recovers proliferative responses of anergic CD4+ lymphocytes from human immunodeficiency virus-infected individuals. Blood 87(11), 4746–4753 (1996).
- Herzenberg LA, De Rosa SC, Dubs JG et al. Glutathione deficiency is associated with impaired survival in HIV disease. Proc. Natl Acad. Sci. USA 94(5), 1967–1972 (1997).
- 4. Matthias LJ, Hogg PJ. Redox control on the cell surface: implications for HIV-1 entry. Antioxid. Redox Signal. 5(1), 133–138 (2003).
- Ho WZ, Douglas SD. Glutathione and N-acetylcysteine suppression of human immunodeficiency virus replication in human monocyte/macrophages in vitro. AIDS Res. Hum. Retroviruses 8(7), 1249–1253 (1992).
- 6. Staal FJ, Roederer M, Raju PA *et al.* Antioxidants inhibit stimulation of HIV transcription. *AIDS Res. Hum. Retroviruses* 9(4), 299–306 (1993).
- 7. Laurence J, Gottlieb AB, Kunkel HG. Soluble suppressor factors in patients with acquired immune deficiency syndrome and its prodrome. Elaboration *in vitro* by T lymphocyte-adherent cell interactions. *J. Clin. Invest.* 72(6), 2072–2081 (1983).
- 8. Van Hecke O, Lee J. Oxford COVID-19 evidence service. Nacetylcysteine: a rapid review of the evidence for effectiveness in treating COVID-19. http://www.cebm.net/covid19/n-acetylcysteine-a-rapid-review-of-the-evidence-for effectiveness-in-treating-covid-19/
- 9. Poe FL, Corn J. N-Acetylcysteine: a potential therapeutic agent for SARS-CoV-2. Med. Hypotheses 143(5), 109862 (2020).
- 10. Diao B, Wang C, Tan Y et al. Reduction and functional exhaustion of T cells in patients with coronavirus disease (COVID-19). Front. Immunol. 11, 827 (2020).
- 11. Ristow SS, Starkey JR, Stanford DR, Davis WC, Brooks CG. Cell surface thiols, but not intracellular glutathione, are essential for cytolysis by a cloned murine natural killer cell line. *Immunol. Invest.* 14(5), 401–414 (1985).
- 12. Redelman D, Hudig D. The mechanism of cell-mediated cytotoxicity. I. killing by murine cytotoxic T lymphocytes requires cell surface thiols and activated proteases. *J. Immunol.* 124(2), 870–878 (1980).
- 13. Macphail S, Stutman O. Suppressor T cells in a primary *in vitro* response to non-major histocompatibility alloantigens. *J. Exp. Med.* 156(5), 1398–1414 (1982).
- 14. Corse E, Gottschalk RA, Allison JP. Strength of TCR-peptide/ MHC interactions and *in vivo* T-cell responses. *J. Immunol.* 186(9), 5039–5045 (2011).
- 15. Kahan SM, Zajac AJ. Immune exhaustion: past lessons and new insights from lymphocytic choriomeningitis virus. *Viruses* 11(2), 156 (2019).
- 16. Yi JY, Cox MA, Zajac AJ. T-cell exhaustion: characteristics causes and conversion. Immunology 129(4), 474-481 (2010).
- 17. Volkan I Sayin VI, Ibrahim MX et al. Antioxidants accelerate lung cancer progression in mice. Sci. Transl. Med. 6, 221 (2014).
- 18. Wiel C, Le Gal K, Ibrahim MX *et al.* BACH1 stabilization by antioxidants stimulates lung cancer metastasis. *Cell* 178(2), 330–345 (2019).
- 19. Makinodan T, Albright JW. Restoration of impaired immune functions in aging animals. III. Effect of mercaptoethanol in enhancing the reduced primary antibody responsiveness *in vivo. Mech. Ageing Dev.* 11(1), 1–8 (1979).
- 20. Heidrick ML, Hendricks LC, Cook DE. Effect of dietary 2-mercaptoethanol on the life span, immune system, tumor incidence and lipid peroxidation damage in spleen lymphocytes of aging BC3F1 mice. *Mech. Ageing Dev.* 27(3), 341–358 (1984).
- 21. Click RE. Obesity, longevity, quality of life: alteration by dietary 2-mercaptoethanol. Virulence 1(6), 509-515 (2010).
- 22. Click RE. Alteration of radiation-sensitive processes associated with cancer and longevity by dietary 2-mercaptoethanol. J. Cancer Res. Ther. 10(1), 127–132 (2014).
- 23. Fischer HD, Wustmann C, Rudolph E *et al.*.. Effect of 2-mercaptoethanol on posthypoxic and age related biochemical and behavioral changes in mice and rats. *Biomed. Biochim. Acta* 49(10), 1085–1090 (1990).
- 24. Fehér E, Pénzes L. Effect of an antioxidant compound (2-mercaptoethanol) on the nerve terminals of the aging small intestine. *Exp. Gerontol.* 25(2), 135–140 (1990).
- 25. Click RE, Benck L, Alter BJ. Enhancement of antibody synthesis in vitro by mercaptoethanol. Cell. Immunol. 3(1), 156-160 (1972).
- 26. Click RE, Benck L, Alter BJ. Immune responses *in vitro*. I. Culture conditions for antibody synthesis.Cell. Immunol. 3(2), 264–276 (1972).

- Click RE, Benck L, Alter BJ, Lovchik JC. Immune responses in vitro. VI. Genetic control of the in vivo-in vitro discrepancies in 19S antibody synthesis. J. Exp. Med. 136(5), 1241–1257 (1972).
- Heber-Katz E, Click RE. Immune responses *in vitro* V. role of mercaptoethanol in the mixed-leukocyte reaction. *Cell. Immunol.* 5(3), 410–418 (1972).
- Heber-Katz E, Peck AB, Click RE. Immune responses in vitro. II. Mixed leukocyte interaction in a protein-free medium. Eur. J. Immunol. 3(7), 379–385 (1973).
- Peck AB, Katz-Heber E, Click RE. Immune responses in vitro. IV. A comparison of the protein-free and mouse serum supplemented mouse mixed lymphocyte interaction assays. *Eur. J. Immunol.* 3(8), 516–519 (1973).
- 31. Peck AB, Click RE. Immune responses in vitro. III. Enhancement of the mouse mixed lymphocyte interaction by isologous and homologous sera. *Eur. J. Immunol.* 3(7), 385–392 (1973).
- 32. Ohmori H, Yamauchi T, Yamamoto I. Augmentation of in vitro antibody response by disulfide compounds I. Comparison between intermolecular and intramolecular disulfides. *Jpn. J. Pharmacol.* 37(1), 13–19 (1985).
- Goodman MG, Weigle WO. Nonspecific activation of murine lymphocytes. I. proliferation and polyclonal activation induced by 2-mercaptoethanol and α-thioglycerol. J. Exp. Med. 145(3), 473–489 (1977).
- 34. Cerottini J-C, Enger HD, MacDonald HR, Brunner KT. Generation of cytotoxic T lymphocytes *in vitro*. I. Response of normal and immune mouse spleen cells in mixed leukocyte cultures. *J. Exp. Med.* 140(3), 703–717 (1974).
- Engers HD, MacDonald HR, Cerottini JC, Brunner KT. Effect of delayed addition of 2-mercaptoethanol on the generation of mouse cytotoxic T lymphocytes in mixed leukocyte cultures. *Eur. J. Immunol.* 5(3), 223–225 (1975).
- 36. Ohmori H, Yamauchi T, Yamamoto I. Augmentation of the antibody response by lipoic acid in mice. I. Analysis of the mode of action in *in vitro* culture system. *Jpn J. Pharmacol.* 42(1), 135–140 (1986).
- 37. Sato H, Shiiya A, Kimata M *et al.* Redox imbalance in cystine/glutamate transporter-deficient mice. *J. Biol. Chem.* 280(45), 37423–37429 (2005).
- 38. Smyth MJ, Thia KY, Street SE *et al.* Differential tumor surveillance by natural killer (NK) and NKT cells. *J. Exp. Med.* 191(4), 661–668 (2000).
- 39. Macho-Fernandez E, Brigl M. The extended family of CD1d-Restricted NKT cells: sifting through a mixed bag of TCRs, antigens and functions. *Front. Immunol.* 6, 362 (2015).
- 40. Stacey NH, Craig GK. Role of thiols in human peripheral blood natural killer and killer lymphocyte activities. *Experientia* 45(2), 180–181 (1989).
- Kuppen PJ, Eggermont AM, Marinelli A, de Heer E, van de Velde CJ, Fleuren GJ. Induction of lymphokine-activated killer activity in rat splenocyte cultures: the importance of 2-mercaptoethanol and indomethacin. *Cancer Immunol. Immunother.* 33(1), 28–32 (1991).
- 42. Yamauchi A, Bloom ET. Requirement of thiol compounds as reducing agents for IL-2-mediated induction of LAK activity and proliferation of human NK cells. *J. Immunol.* 151(10), 5535–5544 (1993).
- Furuke K, Shiraishi M, Mostowski HS, Bloom ET. Fas ligand induction in human NK cells is regulated by redox through a calcineurin-nuclear factors of activated T-cell dependent pathway. *J. Immunol.* 162(4), 1988–1993 (1999).
- 44. Delneste Y, Jeannin P, Sebille E, Aubry JP, Bonnefoy JY. Thiols prevent Fas (CD95)-mediated T cell apoptosis by downregulating membrane Fas expression. *Eur. J. Immunol.* 26(12), 2981–2988 (1996).
- 45. van den Brink RM, Boggs SS, Herberman RB, Hiserodt JC. The generation of natural killer (NK) cells from NK precursor cells in rat long term bone marrow cultures. *J. Exp. Med.* 172(1), 303–313 (1990).
- 46. Heidrick ML, Makinodan T. Presence of impairment of humoral immunity in nonadherent spleen cells of old mice. J. Immunol. 111(5), 1502–1506 (1973).
- Segre D, Segre M. Humoral immunity in aged mice. II. Increased suppressor T-cell activity in immunologically deficient old mice. J. Immunol. 116(3), 735–738 (1976).
- Makinodan T, Albright JW, Good PI, Peter CP, Heidrick ML. Reduced humoral immune activity in long-lived old mice: an approach to elucidating its mechanisms. *Immunology* 31(6), 903–911 (1976).
- 49. Makinodan T, Albright JW. Restoration of impaired immune functions in aging animals. II. Effect of mercaptoethanol in enhancing the reduced primary antibody responsiveness *in vitro*. *Mech. Ageing Dev.* 10(5), 325–340 (1979).
- Heidrick ML, Albright JW, Makinodan T. Restoration of impaired immune functions in aging animals. IV. Action of 2-mercaptoethanol in enhancing age-reduced immune responsiveness. *Mech. Ageing Dev.* 13(4), 367–378 (1980).
- 51. Fong TC, Makinodan T. Preferential enhancement by 2-mercaptoethanol of IL-2 responsiveness of T blast cells from old over young mice is associated with potentiated protein kinase C translocation. *Immunol. Lett.* 20(2), 149–154 (1989).
- Nauss KM, Connor AM, Newberne PM. Alterations in immune function in rats caused by dietary lipotrope deficiency: effect of culture medium, 2-mercaptoethanol and mitogen dose on the *in vitro* lymphocyte transformation response. J. Nutr. 112(12), 2342–2352 (1982).
- 53. Franklin RA, Li YM, Arkins S, Kelley KW. Glutathione augments *in vitro* proliferative responses of lymphocytes to concanavalin A to a greater degree in old than in young rats. *J. Nutr.* 120(12), 1710–1717 (1990).

- 54. Irimajiri N, Bloom ET, Makinodan T. Suppression of murine natural killer cell activity by adherent cells from aging mice. *Mech. Ageing Dev.* 31(2), 155–162 (1985).
- Dussault I, Miller SC. Decline in natural killer cell-mediated immunosurveillance in aging mice–a consequence of reduced cell production and tumor binding capacity. *Mech. Ageing Dev.* 75(2), 115–129 (1994).
- Mikael N, Mirza NM, Zaharian BI *et al.* Genetic control of the decline of natural killer cell activity in aging mice. *Growth Dev. Aging* 58(1), 3–12 (1994).
- 57. Ferrández MD, Correa R, Del Rio M, De la Fuente M. Effects *in vitro* of several antioxidants on the natural killer function of aging mice. *Exp. Gerontol.* 34(5), 675–685 (1999).
- De La Fuente M, Miquel J, Catalán MP, Víctor VN, Guayerbas N. The amount of thiolic antioxidant ingestion needed to improve several immune functions is higher in aged than in adult mice. *Free Radical Res.* 36(2), 119–126 (2002).
- Ohmori H, Yamamoto I. Mechanism of augmentation of the antibody response *in vitro* by 2-mercaptoethanol in murine lymphocytes. I.
 2-Mercaptoethanol-induced stimulation of the uptake of cystine, an essential amino acid. *J. Exp. Med.* 155(5), 1277–1290 (1982).
- 60. Ohmori H, Yamamoto I. Mechanism of augmentation of the antibody response *in vitro* by 2-mercaptoethanol in murine lymphocytes. II. A major role of the mixed disulfide between 2-mercaptoethanol and cysteine. *Cell. Immunol.* 79(1), 173–185 (1983).
- Srivastava MK, Sinha P, Clements VK, Rodriguez P, Ostrand-Rosenberg S. Myeloid-derived suppressor cells inhibit T-cell activation by depleting cysteine and cysteine. *Cancer Res.* 70(1), 68–77 (2010).
- 62. Ghosh T, Barik S, Bhuniya A *et al.* Tumor-associated mesenchymal stem cells inhibit naïve T cell expansion by blocking cysteine export from dendritic cells. *Int. J. Cancer* 139(9), 2068–2081 (2016).
- 63. Yan Z, Garg SK, Kipnis J, Banerjee R. Extracellular redox modulation by regulatory T cells. Nat. Chem. Biol. 5(10), 721–723 (2009).
- 64. Edinger AL, Thompson CB. Antigen presenting cells control T cell proliferation by regulating amino acid availability. *Proc. Natl Acad. Sci. USA* 99(3), 1107–1109 (2002).
- 65. Ishii T, Sugita Y, Bannai S. Regulation of glutathione levels in mouse spleen lymphocytes by transport of cysteine. J. Cell. Physiol. 133(2), 330–336 (1987).
- Angelini G, Gardella S, Ardy M *et al.* Antigen-presenting dendritic cells provide the reducing extracellular microenvironment required for T lymphocyte activation. *Proc. Natl Acad. Sci. USA* 99(3), 1491–1496 (2002).
- Hadzic T, Li L, Cheng N, Walsh SA, Spitz DR, Knudson CM. The role of low molecular weight thiols in T lymphocyte proliferation and IL-2 secretion. J. Immunol. 176(9), 7965–7972 (2005).