

# Downregulation of miR-200a Protects Mouse Leydig Cells Against Triptolide by Triggering Autophagy

This article was published in the following Dove Press journal:  
*Drug Design, Development and Therapy*

Hui Miao  
Congxiu Miao  
Jing Han  
Na Li

Department of Reproduction and Genetics, Heping Hospital Affiliated to Changzhi Medical College, Changzhi, Shanxi 046000, People's Republic of China

**Background:** MicroRNAs play important roles in testicular development and spermatogenesis. Previous research has indicated that the level of miR-200a was significantly upregulated in patients with different spermatogenic impairments. However, the mechanism by which miR-200a regulated spermatogenic impairments remains unclear.

**Methods:** Leydig cells were treated with triptolide (TP) to mimic spermatogenic impairments. CCK-8 and flow cytometry were used to detect the proliferation and apoptosis in Leydig cells, respectively. In addition, Western blot assay was used to examine ATG7, ATG5, p62 protein levels in MLTC-1 cells.

**Results:** TP dose-dependently upregulated the expression of miR-200a in MLTC-1 cells. In addition, TP inhibited the proliferation of MLTC-1 cells via inducing apoptosis and oxidative stress; however, these phenomena were notably reversed by miR-200a antagomir. Furthermore, luciferase reporter assay identified that ATG7 was the direct binding target of miR-200a. TP treatment markedly inhibited the activation of autophagy in MLTC-1 cells via inhibition of ATG7. Conversely, downregulation of miR-200a significantly induced autophagy in TP-treated MLTC-1 cells by activation of ATG7. Meanwhile, the cell protective effects of miR-200a against TP were reversed by autophagy inhibitor 3MA, indicating that autophagy plays an important role.

**Conclusion:** These results indicated that downregulation of miR-200a could protect MLTC-1 cells against TP by inducing autophagy. Therefore, miR-200a might serve as a new therapeutic target for the treatment of male hypogonadism.

**Keywords:** hypogonadism, triptolide, microRNA-200a, autophagy, apoptosis

## Introduction

Hypogonadism is a condition characterized by testosterone deficiency and impairment of spermatogenesis.<sup>1</sup> Hypogonadism affects approximately 6% of males.<sup>2</sup> Male reproduction ability mainly relies on androgens, which are largely secreted by testicular Leydig cells.<sup>3</sup> Testosterone is a steroid hormone, which is mainly produced by Leydig cells.<sup>4</sup> Moreover, hypogonadism often leads to some comorbidities, such as obesity, insulin resistance (IR), type-2 diabetes mellitus (T2DM), and skeletal muscle wasting.<sup>5</sup> Meanwhile, the number of Leydig cells in the adult human testis will reduce with increasing age.<sup>6</sup> Therefore, improving the treatment of hypogonadism contributes to maintaining the sexual function of males.

Triptolide (TP) was isolated from *Tripterygium wilfordii* Hook F, which is a diterpenoid triepoxide.<sup>7</sup> There is evidence that TP exhibits anti-tumor,

Correspondence: Congxiu Miao  
Department of Reproduction and Genetics, Heping Hospital Affiliated to Changzhi Medical College, 110 South Yan'an Road, Changzhi, Shanxi 046000, People's Republic of China  
Email miaocongxiu321@126.com

immunosuppressive and anti-inflammatory properties,<sup>7,8</sup> and also that impairment of spermatogenesis is associated with longer treatment duration with TP.<sup>9</sup> In addition, Lue et al. found that TP could induce infertility in the adult rats, indicating that TP could increase the deformity rate of sperm.<sup>10</sup>

MicroRNAs (miRNAs) are a class of endogenous non-coding RNA molecules with 18–22 nucleotides.<sup>11</sup> MiRNAs can directly bind to the 3'UTR region of its target genes, and regulate the expression of targeted genes.<sup>12</sup> In addition, miRNAs participate in various pathological processes including cell differentiation, cell survival, apoptosis, and protein metabolism.<sup>13</sup> Evidence has shown that some miRNAs are abundantly expressed in various type of male germ cells.<sup>14</sup> Abu-Halima et al. found that the level of miR-200a was notably upregulated in patients with different spermatogenic impairments.<sup>15</sup> However, the mechanism by which miR-200a regulated spermatogenic impairments remains unclear. Hence, this study aimed to investigate the role of miR-200a in spermatogenic impairments using an in vitro cell model.

## Materials and Methods

### Cell Culture and Cell Transfection

The mouse leydig MLTC-1 and TM3 cell lines were purchased from American Type Culture Collection (ATCC, Rockville, MD, USA). Cells were cultured in complete media (DMEM medium containing 10% fetal bovine serum and 1% penicillin/streptomycin) and incubated in an incubator with 5% CO<sub>2</sub> at 37°C. The passage number of cell lines was 3.

MiR-200a agomir and miR-200a antagomir were obtained from GenePharma (Shanghai, China). MLTC-1 or TM3 cells were plated into 6-well plates at a density of 4 x 10<sup>5</sup> cells per well overnight at 37°C. Then, cells were transfected with miR-200a agomir or miR-200a antagomir for 24 h at 37°C using Lipofectamine 2000 reagent (Thermo Fisher Scientific, Waltham, MA, USA) according to the manufacturer's instructions.

### Real-Time Quantitative Polymerase Chain Reaction (RT-qPCR)

Total RNA was extracted using RNAiso Plus (Takara Biotechnology, Co. Ltd, Dalian, China) according to the provided protocol. Then, the PrimeScript™ RT Master Mix Kit (Takara Biotechnology) was used to synthesize complementary DNA. After that, RT-qPCR was performed using a SYBR Green™ Premix Ex Taq™ (Takara

Biotechnology). The primers used were as follows: U6: forward, 5'-CTCGCTTCGGCAGCACAT-3'; reverse, 5'-AACGCTTCACGAATTTGCGT-3'. MiR-200a: forward, 5'-GGAGTTCGTATCGGCTGCGATG-3'; reverse, 5'-CGACCGTGTAAATCGTCGTTGC-3'. The level of miR-200a was normalized to that of U6.

### Cell Counting Kit-8 (CCK-8)

TP (purity: 99.79%) was purchased from MedChem Express (Monmouth Junction, NJ, USA). TP was dissolved in DMSO (10 mM) first; the store solution of TP was diluted to the indicated concentration with DMEM. MLTC-1 or TM3 cells (5 x 10<sup>4</sup> cells per well) were plated onto 96-well plates and incubated overnight at 37°C. After that, cells were treated with TP (30, 90, 120 or 150 nM) for 24 h. Later on, cells were incubated with 10 µL CCK8 reagent (Beyotime, Shanghai, China) at 37°C for another 2 h. The absorbance was measured at a wavelength of 450 nm using a microplate reader (Bio-Rad Laboratories, Inc., Hercules, CA, USA). The dose of TP was calculated based on previous studies.<sup>16,17</sup>

### Flow Cytometric Analysis

Apoptosis in MLTC-1 and in TM3 cells was detected by an Annexin V-FITC/PI Apoptosis Detection Kit (Sigma-Aldrich, St. Louis, MO, USA). Cells were washed three times with pre-cold PBS, and then were resuspended in 500 µL of binding buffer. After that, cells were stained with 5 µL of propidium iodide (PI) and 5 µL of Annexin V-FITC at room temperature for 15 min in the dark. Subsequently, the number of annexin V-FITC-positive cells was determined with a FACS flow cytometer (BD Biosciences, Franklin Lakes, NJ, USA).

### Trypan Blue Exclusion Assay

For evaluation of cell death, MLTC-1 cells were incubated with 0.25% trypan blue solution (Gibco, Paisley, UK). Subsequently, viable cells were counted using a hemacytometer (Neubauer improved, Marienfeld, Germany).

### Dual-Luciferase Reporter Assay

The fragment of ATG7 3'-UTR containing the binding site of miR-200a was inserted into the pmirGLO dual-Luciferase vector, named wild-type (WT)-ATG7. The vector pmirGLO-200a-MT (mutant) was inserted by the ATG7-MT, which contains point mutations of the miR-200a seed region binding site. After that, MLTC-1 cells were co-transfected with WT-ATG7 or MT-ATG7 plasmid,

with miR-200a agomir respectively using Lipofectamine 2000. After 48 h of incubation, the luciferase activity in cell lysate was detected using the Dual Luciferase Reporter Assay System (Promega, Madison, WI, USA) according to the manufacturer's protocol.

## Immunofluorescence Assay

MLTC-1 cells were fixed with PBS containing 4% paraformaldehyde for 1 h, and then permeabilized with 0.1% Triton X-100 for 30 min. After that, cells were blocked with 1% bovine serum albumin for 2 h, and then incubated overnight with an antibody against LC3B (1:100; 10 µg/mL, Abcam Cambridge, MA, USA) at 4°C. Later on, cells were incubated with horseradish peroxidase-conjugated secondary antibody (1:1000, Abcam) at room temperature for 1 h, and then observed under a confocal microscopy (Olympus CX23 Tokyo, Japan). The nucleus was stained with DAPI for 5 min. Autophagy inhibitor 3-Methyladenine (3MA) was purchased from Sigma Aldrich (St. Louis, MO, USA).

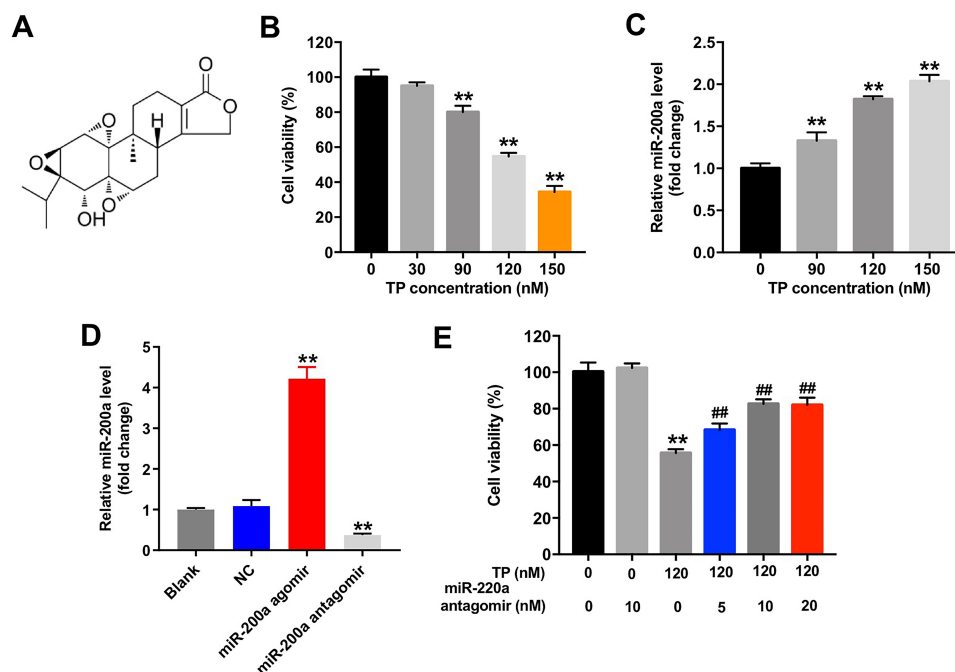
## ROS Analysis

Intracellular ROS levels of MLTC-1 cells were determined by staining of cells with 2',7'-dichlorodihydrofluorescein

diacetate (DCFH-DA; Sigma Aldrich). The fluorescence was detected by flow cytometry as described in previous studies.<sup>18,19</sup>

## Western Blot Assay

Total proteins were quantified using a BCA protein assay kit (Beyotime Institute of Biotechnology, Beijing, China). Protein lysates (30 µg loading) were separated by 10% sodium dodecyl sulfate-polyacrylamide gel electrophoresis. After that, the gels were transferred onto polyvinylidene difluoride (PVDF) membrane (Millipore, Billerica, MA, USA), followed by blocking with 5% skimmed milk in TBST for 1 h at room temperature. Later on, the membrane was incubated in primary antibodies against ATG7 (1:1000), p-ERK (1:1000), ERK (1:1000), ATG5 (1:1000), p62 (1:1000), Bax (1:1000), Bcl-2 (1:1000), active caspase 3 (1:1000), active caspase 9 (1:1000), LC3B (1:1000) and β-actin (1:1000) at 4°C overnight, followed by incubating with a secondary antibody (1:5000). Subsequently, signals were visualized using a KeyGEN Enhanced ECL detection kit (KeyGEN, Nanjing, China). All antibodies were obtained from Abcam (Cambridge, MA, USA). The concentration of primary antibodies used in Western blot was 1 µg/mL.



**Figure 1** Downregulation of miR-200a attenuated TP-induced cytotoxicity in MLTC-1 cells. **(A)** The chemical structure of triptolide. **(B)** MLTC-1 cells were treated with TP (30, 90, 120, or 150 nM) for 24 h. CCK-8 assay was used to detect cell viability. **(C)** MLTC-1 cells were treated with TP (90, 120, or 150 nM) for 24 h. RT-qPCR was used to determine the level of miR-200a in cells. **(D)** MiR-200a level in MLTC-1 cells transfected with miR-200a agomir or miR-200a antagonist was analyzed by qRT-PCR. **(E)** MLTC-1 cells were transfected with miR-200a antagonist (5, 10, or 20 nM) first, and then exposed to 120 nM TP for 24 h. CCK-8 assay was used to detect the cell viability. These experiments were repeated three times. \*\*P<0.01 compared with control group; ##P<0.01 compared with TP (120 nM) group.

**Abbreviations:** TP, triptolide; CCK-8, Cell Counting Kit-8; NC, negative control.

## ELISA Assay

ELISA kits (Nanjing Jiancheng Bioengineering Institute, Jiangsu, China) were used to detect the levels of MDA, SOD, GSH in cell culture supernatants.

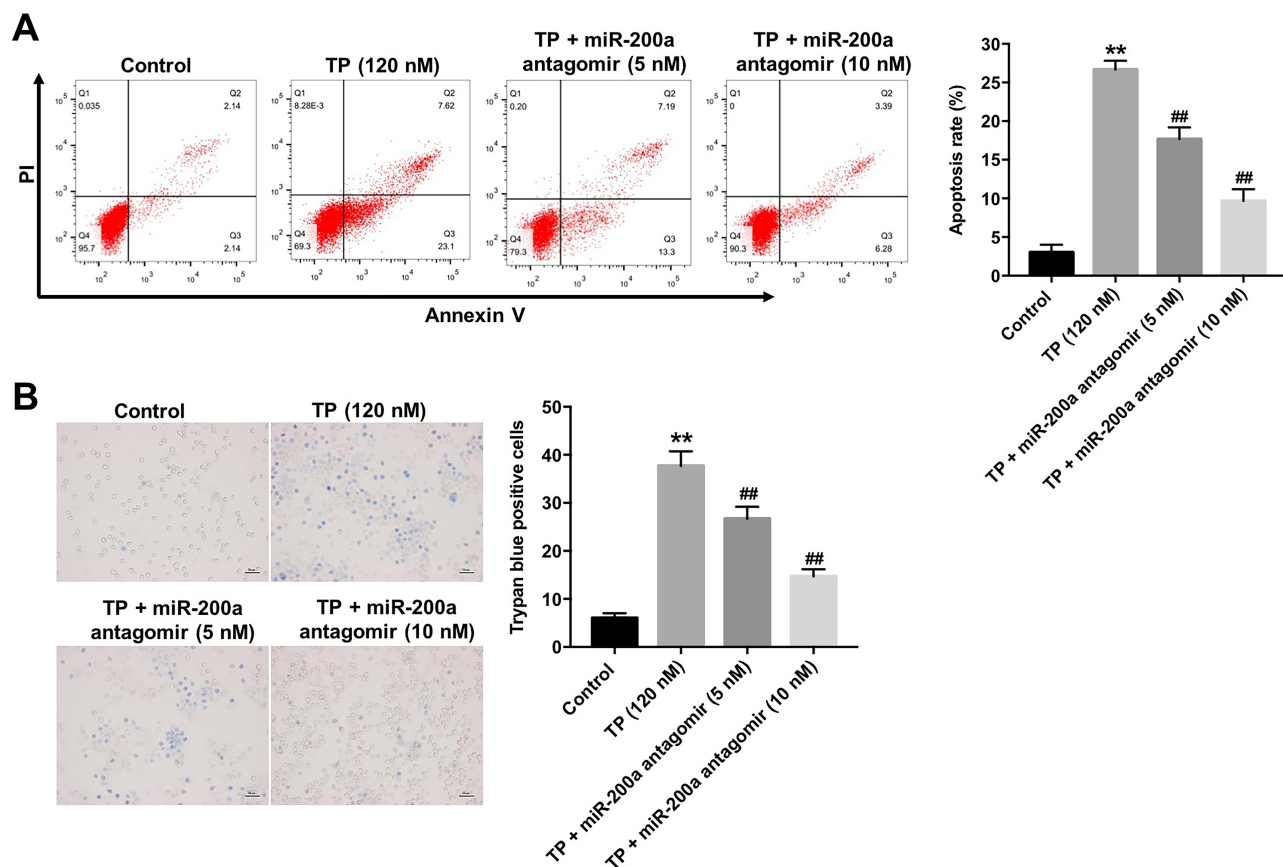
## Statistical Analysis

All data were repeated in triplicate. Data are presented as the mean  $\pm$  standard deviation (S.D.). All statistical analyses were performed using GraphPad Prism software (version 7.0, La Jolla, CA, USA). The results of CCK-8, RT-qPCR, trypan blue exclusion, immunofluorescence, Western blot, flow cytometry, ROS detection and ELISA assays were analyzed by one-way analysis of variance (ANOVA) followed by Tukey's tests (multiple comparisons). In addition, the result of dual-luciferase reporter assay was analyzed by two-way ANOVA followed by Tukey's tests (multiple comparisons). \* $P < 0.05$  was considered to be statistically significant.

## Results

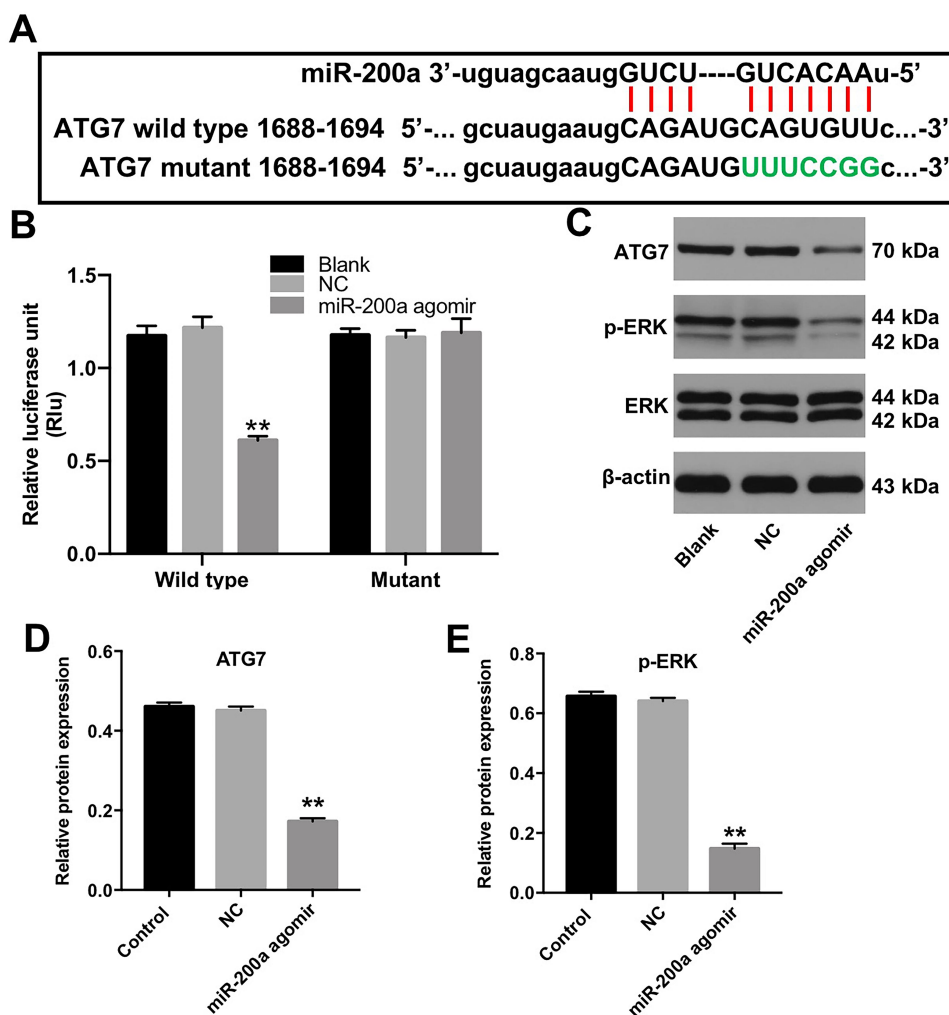
### Downregulation of miR-200a Attenuated TP-Induced Cytotoxicity in Leydig Cells

To assess the effect of TP on the proliferation of MLTC-1 cells, CCK-8 assay was used. The chemical structure of TP is shown in Figure 1A and CCK-8 data indicated TP inhibited the proliferation of MLTC-1 cells in a dose dependent manner (Figure 1B). In addition, the level of miR-200a was significantly upregulated in TP-treated cells (Figure 1C). As expected, miR-200a agomir markedly increased the level of miR-200a in MLTC-1 cells, while miR-200a antagomir exhibited a completely opposing effect (Figure 1D). Furthermore, downregulation of miR-200a attenuated the cytotoxicity of TP on MLTC-1 and TM-3 cells (Figure 1E and Supplementary Figure 1A). These data indicated that downregulation of miR-200a could attenuate TP-induced cytotoxicity in Leydig cells.



**Figure 2** Downregulation of miR-200a attenuated TP-induced cell death in MLTC-1 cells. MLTC-1 cells were transfected with miR-200a antagomir (0, 5, or 10 nM) first, and then exposed to 120 nM TP for 24 h. (A) Apoptotic cells were measured with Annexin V and PI double staining. (B) Trypan blue staining assay was used to detect cell death. These experiments were repeated three times. \*\* $P < 0.01$  compared with control group; ## $P < 0.01$  compared with TP (120 nM) group.

**Abbreviations:** TP, triptolide; PI, propidium iodide.



**Figure 3** ATG7 was a direct binding target of miR-200a. **(A)** The putative binding sites of miR-200a on ATG7 and the mutated sequences are shown. **(B)** Luciferase activity in MLTC-1 cells following co-transfection with ATG7-WT/MT 3'-UTR plasmid and miR-200a agomir was measured using dual luciferase reporter assay. **(C)** Western analysis of ATG7 and p-ERK protein levels in MLTC-1 cells. **(D and E)** The relative expressions of ATG7 and p-ERK in MLTC-1 cells were quantified by normalizing to  $\beta$ -actin and ERK. These experiments were repeated three times. \*\* $P < 0.01$  compared with the NC group.

**Abbreviation:** NC, negative control.

## Downregulation of miR-200a Attenuated TP-Induced Apoptosis of Leydig Cells

To further explore the protective effect of miR-200a antagomir in TP-treated Leydig cells, flow cytometry was applied. As indicated in [Figure 2A](#) and [supplementary Figure 1B](#), TP significantly induced apoptosis of Leydig cells, which was notably reversed after transfection with 10 nM miR-200a antagomir. In addition, trypan blue staining assay indicated that downregulation of miR-200a markedly blocked TP-induced cell death in Leydig cells ([Figure 2B](#) and [supplementary Figure 1C](#)). These data suggested that downregulation of miR-200a could attenuate TP-induced cell death in Leydig cells.

## ATG7 Was a Direct Binding Target of miR-200a

The online bioinformatics tools miRDB (<http://www.mirdb.org>) and microRNA (<http://www.microrna.org/microrna/home.do>) were used to identify the target genes of miR-200a. As shown in [Figure 3A](#), miR-200a had a complementary sequence to ATG7. In addition, dual-luciferase reporter assay indicated that miR-200a agomir suppressed the luciferase activity of ATG7-WT in MLTC-1 cells, but it did not affect the luciferase activity of ATG7-MT ([Figure 3B](#)). In addition, miR-200a agomir notably decreased the expressions of ATG7 and phosphorylation of ERK in MLTC-1 cells ([Figure 3C-E](#)). All these results illustrated that ATG7 was a direct binding target of miR-200a.

## Downregulation of miR-200a Triggered Autophagy in TP-Treated MLTC-1 Cells

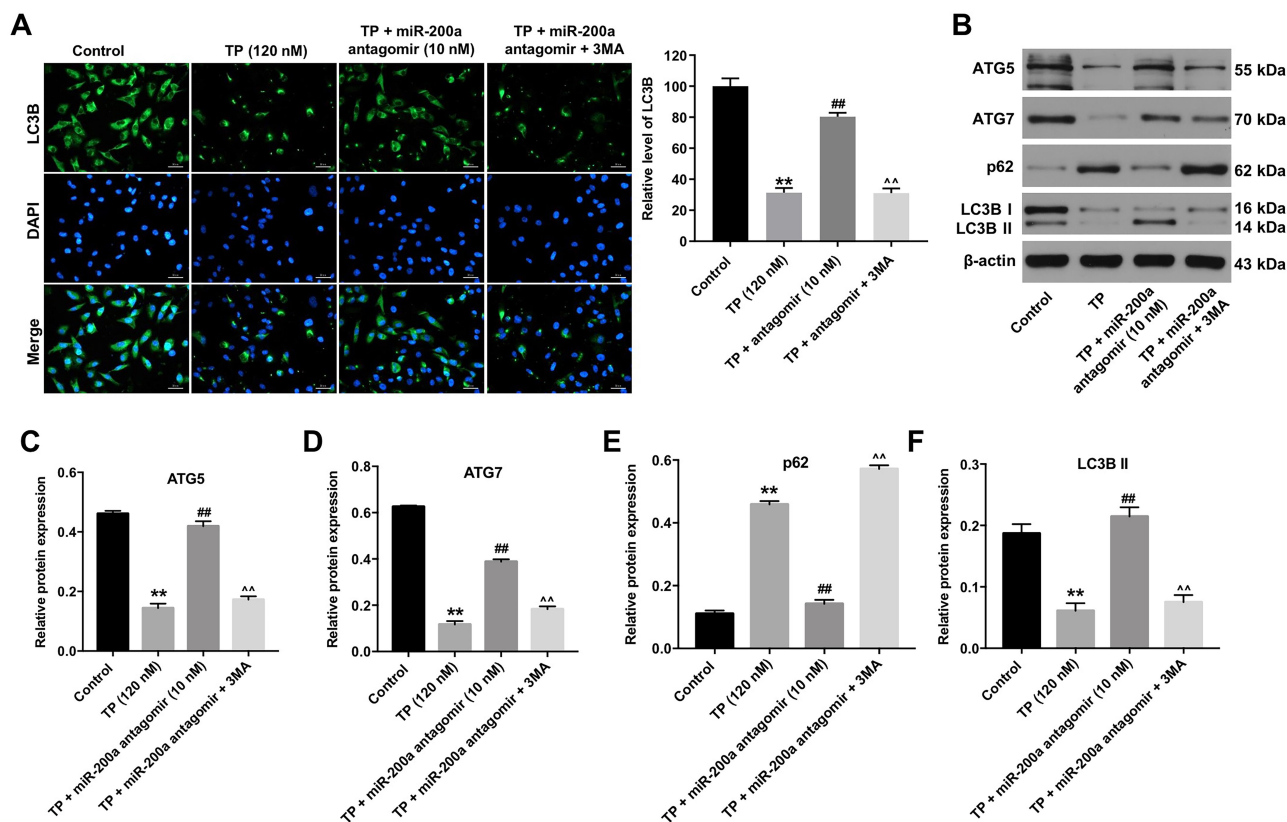
To investigate MLTC-1 cell autophagy, immunofluorescent assay was used. As shown in Figure 4A, TP significantly inhibited the formation of LC3B puncta in MLTC-1 cells, indicating that TP could inhibit autophagy. In contrast, downregulation of miR-200a increased the accumulation of LC3B in TP-treated MLTC-1 cells, compared with TP treatment group; however, that effect was abolished in the presence of 3MA (Figure 4A). In addition, Western blot data indicated that TP decreased the expressions of ATG7, ATG5 and LC3B II and increased the level of p62 in MLTC-cells, while these phenomena were reversed by miR-200a antagonist (Figure 4B–F). However, the inductive effect of miR-200a antagonist on autophagy in TP-treated MLTC-1 cells was reversed by 3MA (Figure 4B–F). These data

suggested that downregulation of miR-200a could trigger autophagy in TP-treated MLTC-1 cells.

## Downregulation of miR-200a Attenuated TP-Induced Oxidative Stress in MLTC-1 Cells via Inducing Autophagy

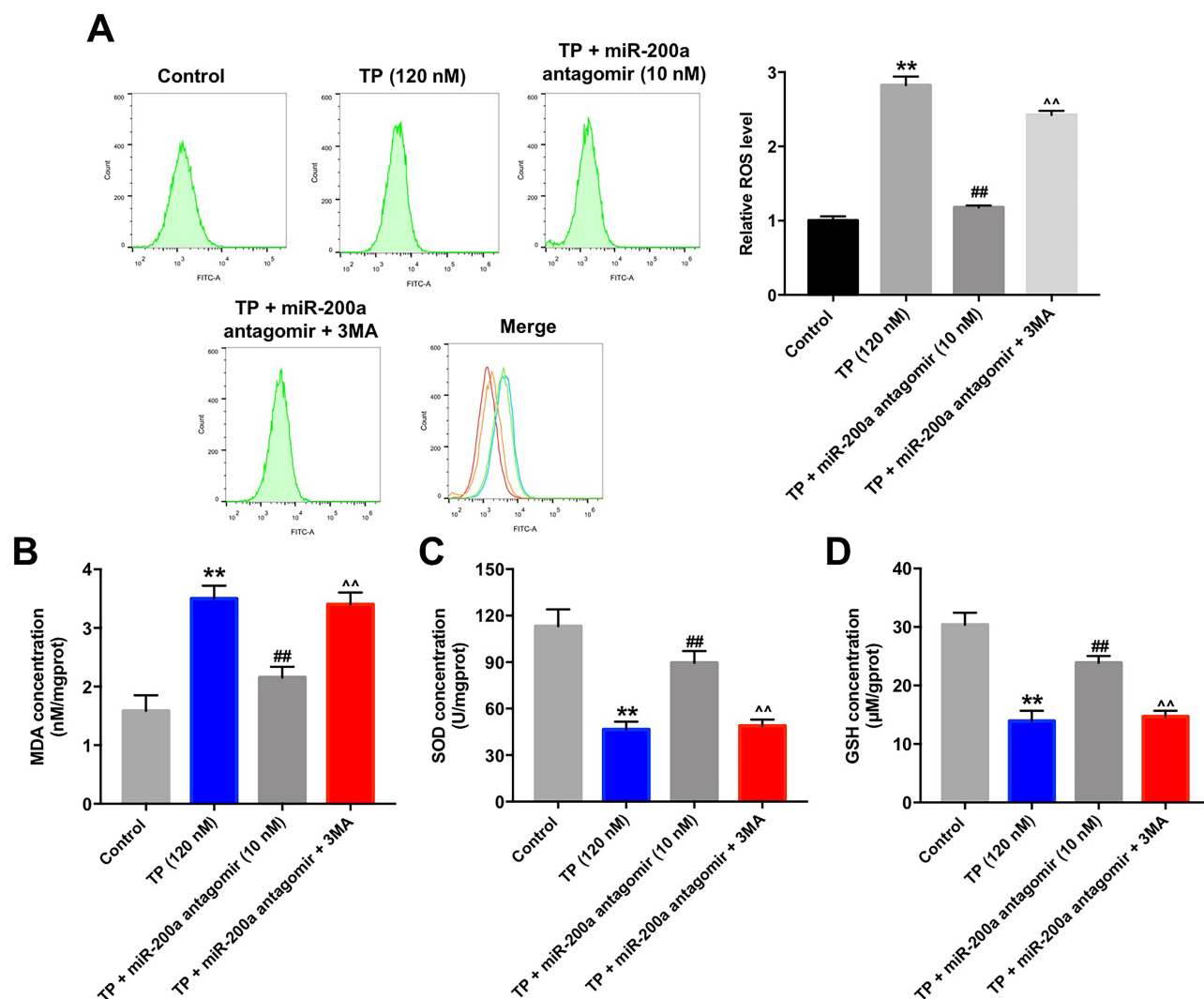
We next explored the effect of miR-200a antagonist on oxidative stress in TP-treated MLTC-1 cells by detecting the relevant biomarkers. As shown in Figure 5A–D, TP markedly increased the levels of ROS and MDA, and decreased the activities of SOD and GSH in MLTC-1 cells; however, these TP-induced changes were reversed by miR-200a antagonist.

We next investigated whether cell autophagy was involved in these processes. As indicated in Figure 5A–D, the inhibitory effects of miR-200a antagonist on oxidative stress in TP-treated MLTC-1 cells were significantly



**Figure 4** Downregulation of miR-200a triggered autophagy in TP-treated MLTC-1 cells. MLTC-1 cells were transfected with 10 nM miR-200a antagonist first, and then exposed to 120 nM TP for 24 h. Cells were treated with 5 mM 3MA prior to 12 h of the miR-200a antagonist treatment. (A) LC3B puncta were observed by immunofluorescence using a confocal microscope. (B) Western analysis of ATG7, ATG5, p62 and LC3B protein levels in MLTC-1 cells. (C–F) The relative expressions of ATG7, ATG5, p62 and LC3B in MLTC-1 cells were quantified by normalizing to β-actin. These experiments were repeated three times. \*\*P<0.01 compared with control group; ##P<0.01 compared with TP (120 nM) group; ^^P<0.01 compared with TP + miR-200a antagonist group.

**Abbreviations:** TP, triptolide; 3MA, 3-methyladenine.



**Figure 5** Downregulation of miR-200a attenuated TP-induced oxidative stress in MLTC-1 cells via inducing autophagy. MLTC-1 cells were transfected with 10 nM miR-200a antagonist first, and then exposed to 120 nM TP for 24 h. Cells were treated with 5 mM 3MA prior to 12 h of the miR-200a antagonist treatment. **(A)** Intracellular ROS generation was assessed by flow cytometry. **(B–D)** Levels of MDA, SOD, and GSH in cells were detected with ELISA. These experiments were repeated three times. \*\* $P < 0.01$  compared with control group; ## $P < 0.01$  compared with TP (120 nM) group; ^^ $P < 0.01$  compared with TP + miR-200a antagonist group.

**Abbreviations:** TP, triptolide; 3MA, 3-methyladenine.

abolished by 3MA. These data indicated that downregulation of miR-200a could attenuate TP-induced oxidative stress in MLTC-1 cells via inducing autophagy.

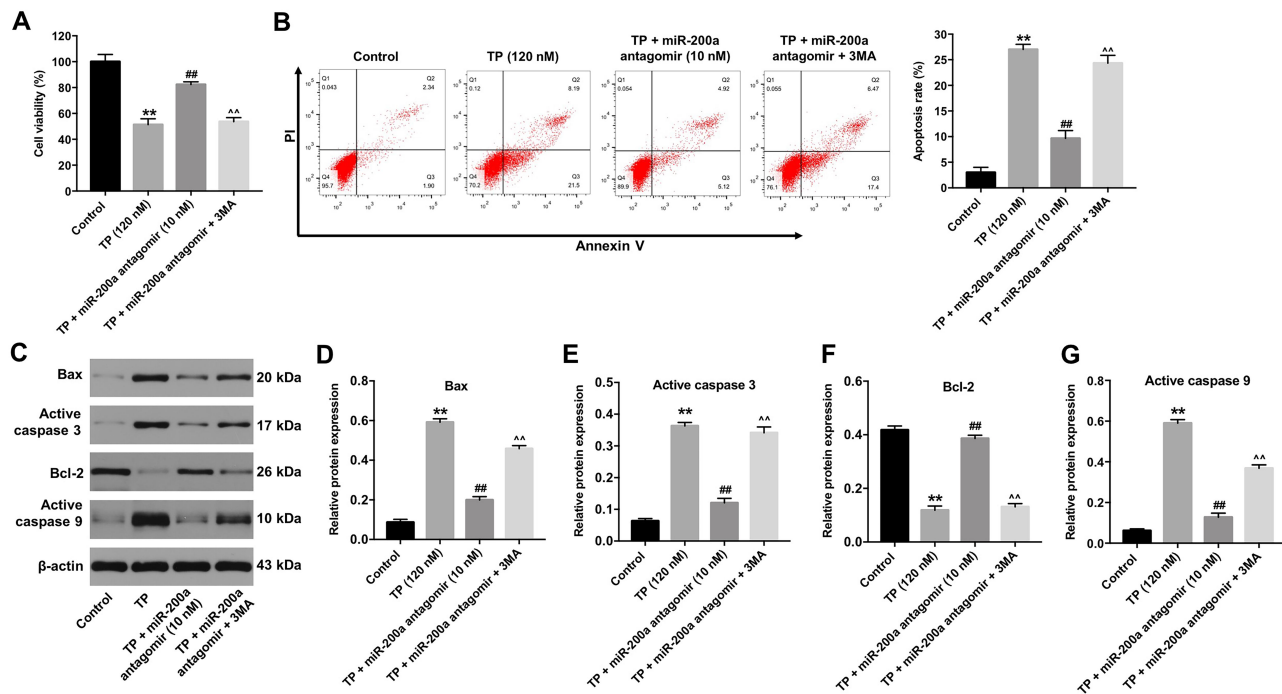
## Downregulation of miR-200a Attenuated TP-Induced Apoptosis in MLTC-1 Cells via Inducing Autophagy

Since miR-200a antagonist could inhibit apoptosis and induce autophagy in TP-treated MLTC-1 cells, we sought to investigate the interaction between autophagy and apoptosis. As shown in Figure 6A, treatment with 3MA significantly abolished the protective effect of miR-200a antagonist in TP-treated MLTC-1 cells. In addition, the inhibitory effect

of miR-200a antagonist on apoptosis in TP-treated MLTC-1 cells was abolished by the treatment with 3MA (Figure 6B). Western blot data showed the effect of miR-200a antagonist on the expressions of Bax, active caspase 3, active caspase 9, and Bcl-2 was reversed by 3MA as well (Figure 6C–G). All these results revealed that downregulation of miR-200a could attenuate TP-induced apoptosis in MLTC-1 cells via inducing autophagy.

## Discussion

Evidence has shown that TP could result in degenerative changes in seminiferous tubules and epididymides and lead to a significant reduction in the weights of testis and epididymis.<sup>20</sup> In addition, TP could induce reproductive



**Figure 6** Downregulation of miR-200a attenuated TP-induced apoptosis in MLTC-1 cells via inducing autophagy. MLTC-1 cells were transfected with 10 nM miR-200a antagonist first, and then exposed to 120 nM TP for 24 h. Cells were treated with 5 mM 3MA prior to 12 h of the miR-200a antagonist treatment. **(A)** CCK-8 assay was used to detect cell viability. **(B)** Apoptotic cells were measured with Annexin V and PI double staining. **(C)** Western analysis of Bax, active caspase 3, Bcl-2 and active caspase 9 protein levels in MLTC-1 cells. **(D–G)** The relative expressions of Bax, active caspase 3, Bcl-2 and active caspase 9 in MLTC-1 cells were quantified by normalizing to  $\beta$ -actin. These experiments were repeated three times. \*\* $P < 0.01$  compared with control group; ### $P < 0.01$  compared with TP (120 nM) group; ^^ $P < 0.01$  compared with TP + miR-200a antagonist group.

**Abbreviations:** TP, triptolide; 3MA, 3-methyladenine; CCK-8, Cell Counting Kit-8; PI, propidium iodide.

toxicity in Leydig cells via triggering apoptosis and accumulation of ROS.<sup>21</sup> Excessive amounts of ROS can result in lasting oxidative stress and can induce cell apoptosis.<sup>22,23</sup> In addition, oxidative stress could affect normal spermatozoa production by impairing the steroidogenic capacity of the testis.<sup>22</sup> As we know, MDA is used as an important biomarker for examining oxidative stress, while SOD and GSH are important antioxidants.<sup>24</sup> In this study, we found that TP significantly increased the levels of ROS and MDA, and decreased the activities of SOD and GSH in MLTC-1 cells. Meanwhile, TP markedly increased the expressions of Bax and active caspase 3 in MLTC-1 cells. These data indicated that TP significantly induced apoptosis and oxidative stress in MLTC-1 cells, which were consistent with previous studies.<sup>21,25</sup> Xiong et al. found that ectopic expression of miR-200a significantly inhibited motility traits of sperm.<sup>26</sup> Our data found that downregulation of miR-200a inhibited apoptosis and oxidative stress in TP-treated MLTC-1 cells, indicating that miR-200a may play a critical role in the development of hypogonadism.

MiRNAs have been found to exert their functions by suppressing the expression of their target genes.<sup>27</sup> Our data

identify ATG7 as a binding target of miR-200a. ATG7 is an important autophagy-related protein, which could increase autophagy.<sup>28</sup> Autophagy, as a self-digestion process, can maintain intracellular homeostasis and contribute to cell survival.<sup>29</sup> In this study, miR-200a agomir significantly downregulated the expressions of ATG7 and phospho-ERK. Gao et al. found that dysfunction of autophagy could reduce testosterone production in some patients with oligospermia.<sup>30</sup> In addition, inhibition of autophagy could induce oxidative stress and reduce testosterone levels.<sup>31</sup> In the current study, TP significantly inhibited autophagy in MLTC-1 cells via downregulation of ATG7, ATG5 and LC3B, which was consistent with previous research.<sup>32</sup> Conversely, downregulation of miR-200a increased the expressions of LC3B, ATG7 and ATG5, but decreased the expression of p62 in TP-treated Leydig cells, thereby promoting cell autophagy. However, the effects of miR-200a antagonist were abolished by 3MA. All these data indicated that miR-200a antagonist could activate the autophagy in TP-treated MLTC-1 cells by targeting ATG7. Nevertheless, it is possible that miR-200a regulated other targets in this setting and more investigations are needed in future.



The cytoprotective function of autophagy can inhibit cell apoptosis and reduce ROS accumulation.<sup>33</sup> Our data indicated that downregulation of miR-200a could protect Leydig cells from TP-induced apoptosis and oxidative stress. However, the inhibitory effects of miR-200a antagonist on apoptosis and oxidative stress in TP-treated MLTC-1 cells were reversed by the treatment with 3MA. These data indicated that downregulation of miR-200a could ameliorate apoptosis and oxidative stress in TP-treated MLTC-1 cells via triggering autophagy.

Male hypogonadism is the result of deficiency of the hormone testosterone.<sup>34</sup> The Leydig cells are responsible for producing testosterone, and testosterone is critical for spermatogenesis.<sup>35</sup> Thus, in the future, it is important to investigate whether miR-200a could affect the level of hormone testosterone in TP-treated MLTC-1 cells.

## Conclusion

Our study demonstrates that downregulation of miR-200a protects against TP-induced reproductive toxicity via targeting ATG7, suggesting that miR-200a might serve as new therapeutic target for the treatment of male hypogonadism.

## Funding

This work was supported by Shanxi Health Commission (No. 2018125).

## Disclosure

The authors declare no competing financial interests.

## References

- Pivonello R, Menafrà D, Riccio E, et al. Metabolic disorders and male hypogonadotropic hypogonadism. *Front Endocrinol (Lausanne)*. 2019;10:345. doi:10.3389/fendo.2019.00345
- Leisegang K, Henkel R. The in vitro modulation of steroidogenesis by inflammatory cytokines and insulin in TM3 Leydig cells. *Reprod Biol Endocrinol*. 2018;16(1):26. doi:10.1186/s12958-018-0341-2
- Kristensen DM, Desdoits-Lethimonier C, Mackey AL, et al. Ibuprofen alters human testicular physiology to produce a state of compensated hypogonadism. *Proc Natl Acad Sci U S A*. 2018;115(4):E715–e724. doi:10.1073/pnas.1715035115
- Dohle GR, Smit M, Weber RF. Androgens and male fertility. *World J Urol*. 2003;21(5):341–345. doi:10.1007/s00345-003-0365-9
- Corona G, Bianchini S, Sforza A, Vignozzi L, Maggi M. Hypogonadism as a possible link between metabolic diseases and erectile dysfunction in aging men. *Hormones (Athens)*. 2015;14(4):569–578.
- O'Hara L, McInnes K, Simitsidellis I, et al. Autocrine androgen action is essential for Leydig cell maturation and function, and protects against late-onset Leydig cell apoptosis in both mice and men. *FASEB J*. 2015;29(3):894–910. doi:10.1096/fj.14-255729

- Brinker AM, Raskin I. Determination of triptolide in root extracts of *Tripterygium wilfordii* by solid-phase extraction and reverse-phase high-performance liquid chromatography. *J Chromatogr A*. 2005;1070(1–2):65–70. doi:10.1016/j.chroma.2005.02.083
- Xue M, Cheng Y, Han F, et al. Triptolide attenuates renal tubular epithelial-mesenchymal transition via the MiR-188-5p-mediated PI3K/AKT pathway in diabetic kidney disease. *Int J Biol Sci*. 2018;14(11):1545–1557. doi:10.7150/ijbs.24032
- Huynh PN, Hikim AP, Wang C, et al. Long-term effects of triptolide on spermatogenesis, epididymal sperm function, and fertility in male rats. *J Androl*. 2000;21;5:689–699.
- Lue Y, Sinha Hikim AP, Wang C, et al. Triptolide: a potential male contraceptive. *J Androl*. 1998;19;4:479–486.
- Huang R, Li J, Pan F, Zhang B, Yao Y. The activation of GPER inhibits cells proliferation, invasion and EMT of triple-negative breast cancer via CD151/miR-199a-3p bio-axis. *Am J Transl Res*. 2020;12(1):32–44.
- Wang Y, Deng X, Dai Y, Niu X, Zhou M. miR-27a downregulation promotes squamous cell carcinoma progression via targeting EGFR. *Front Oncol*. 2019;9:1565. doi:10.3389/fonc.2019.01565
- Wei J, Yang L, Wu YN, Xu J. Serum miR-1290 and miR-1246 as potential diagnostic biomarkers of human pancreatic cancer. *J Cancer*. 2020;11(6):1325–1333. doi:10.7150/jca.38048
- Li L, Zhu Y, Chen T, et al. MiR-125b-2 knockout in testis is associated with targeting to the PAP gene, mitochondrial copy number, and impaired sperm quality. *Int J Mol Sci*. 2019;20(1).
- Abu-Halima M, Hammadeh M, Schmitt J, et al. Altered microRNA expression profiles of human spermatozoa in patients with different spermatogenic impairments. *Fertil Steril*. 2013;99(5):1249–1255. e1216. doi:10.1016/j.fertnstert.2012.11.054
- Yan X, Ke XX, Zhao H, et al. Triptolide inhibits cell proliferation and tumorigenicity of human neuroblastoma cells. *Mol Med Rep*. 2015;11(2):791–796. doi:10.3892/mmr.2014.2814
- Liang H, Zhang S, Li Z. Ginsenoside Rg3 protects mouse leydig cells against triptolide by downregulation of miR-26a. *Drug Des Devel Ther*. 2019;13:2057–2066. doi:10.2147/DDDT.S208328
- Passos JF, Nelson G, Wang C, et al. Feedback between p21 and reactive oxygen production is necessary for cell senescence. *Mol Syst Biol*. 2010;6:347. doi:10.1038/msb.2010.5
- Tang L, Li J, Fu W, Wu W, Xu J. Suppression of FADS1 induces ROS generation, cell cycle arrest, and apoptosis in melanocytes: implications for vitiligo. *Aging (Albany NY)*. 2019;11(24):11829–11843. doi:10.18632/aging.102452
- Ni B, Jiang Z, Huang X, et al. Male reproductive toxicity and toxicokinetics of triptolide in rats. *Arzneimittelforschung*. 2008;58(12):673–680.
- Hu J, Yu Q, Zhao F, et al. Protection of quercetin against triptolide-induced apoptosis by suppressing oxidative stress in rat Leydig cells. *Chem Biol Interact*. 2015;240:38–46. doi:10.1016/j.cbi.2015.08.004
- Sedha S, Kumar S, Shukla S. Role of oxidative stress in male reproductive dysfunctions with reference to phthalate compounds. *Urol J*. 2015;12(5):2304–2316.
- Gao S, Wang Z, Zhang C, Jia L, Zhang Y. Oral exposure to atrazine induces oxidative stress and calcium homeostasis disruption in spleen of mice. *Oxid Med Cell Longev*. 2016;2016:7978219. doi:10.1155/2016/7978219
- Chen Y, Feng X, Hu X, et al. Dexmedetomidine ameliorates acute stress-induced kidney injury by attenuating oxidative stress and apoptosis through inhibition of the ROS/JNK signaling pathway. *Oxid Med Cell Longev*. 2018;2018:4035310.
- Wang H, Chen L, Ye XY. [Triptolide induces oxidative stress and apoptosis and activates PIK3/Akt signaling pathway in TM4 sertoli cells]. *Beijing Da Xue Xue Bao Yi Xue Ban*. 2018;50(4):607–612. Chinese.

26. Xiong S, Ma W, Jing J, et al. An miR-200 cluster on chromosome 23 regulates sperm motility in zebrafish. *Endocrinology*. 2018;159(5):1982–1991. doi:10.1210/en.2018-00015
27. Li P, Chen Y, Juma CA, et al. Differential inhibition of target gene expression by human microRNAs. *Cells*. 2019;8(8):791. doi:10.3390/cells8080791
28. Zheng W, Xie W, Yin D, et al. ATG5 and ATG7 induced autophagy interplays with UPR via PERK signaling. *Cell Commun Signal*. 2019;17(1):42. doi:10.1186/s12964-019-0353-3
29. Codogno P, Meijer AJ. Autophagy and signaling: their role in cell survival and cell death. *Cell Death Differ*. 2005;12(Suppl 2):1509–1518. doi:10.1038/sj.cdd.4401751
30. Gao F, Li G, Liu C, et al. Autophagy regulates testosterone synthesis by facilitating cholesterol uptake in Leydig cells. *J Cell Biol*. 2018;217(6):2103–2119. doi:10.1083/jcb.201710078
31. Yahyavy S, Valizadeh A, Saki G, Khorsandi L. Taurine induces autophagy and inhibits oxidative stress in mice Leydig cells. *JBRA Assist Reprod*. 2020. doi:10.5935/1518-0557.20190079
32. Kim KH, Lee MS. Autophagy—a key player in cellular and body metabolism. *Nat Rev Endocrinol*. 2014;10(6):322–337. doi:10.1038/nrendo.2014.35
33. Kaminsky VO, Zhivotovsky B. Free radicals in cross talk between autophagy and apoptosis. *Antioxid Redox Signal*. 2014;21(1):86–102. doi:10.1089/ars.2013.5746
34. Çatakoğlu AB, Kendirci M. Testosterone replacement therapy and cardiovascular events. *Turk Kardiyol Dern Ars*. 2017;45(7):664–672.
35. Wang Y, Chen F, Ye L, Zirkin B, Chen H. Steroidogenesis in Leydig cells: effects of aging and environmental factors. *Reproduction*. 2017;154(4):R111–r122. doi:10.1530/REP-17-0064

## Drug Design, Development and Therapy

Dovepress

### Publish your work in this journal

Drug Design, Development and Therapy is an international, peer-reviewed open-access journal that spans the spectrum of drug design and development through to clinical applications. Clinical outcomes, patient safety, and programs for the development and effective, safe, and sustained use of medicines are a feature of the journal, which has also

been accepted for indexing on PubMed Central. The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.

Submit your manuscript here: <https://www.dovepress.com/drug-design-development-and-therapy-journal>