REVIEW

OPEN ACCESS Check for updates

At the heart of microbial conversations: endocannabinoids and the microbiome in cardiometabolic risk

Ramsha Nabihah Khan^a, Kristal Maner-Smith^b, Joshua A. Owens^a, Maria Estefania Barbian^c, Rheinallt M. Jones ^b, and Crystal R. Naudin ^b

^aDivision of Gastroenterology and Hepatology, Department of Pediatrics, Children's Healthcare of Atlanta and Emory University, Atlanta, Georgia, USA; ^bEmory Integrated Metabolomics and Lipidomics Core, Emory University, Atlanta, Georgia, USA; ^cDivision of Neonatology, Department of Pediatrics, Children's Healthcare of Atlanta and Emory University, Atlanta, Georgia, USA

ABSTRACT

Cardiometabolic syndrome encompasses intertwined risk factors such as hypertension, dyslipidemia, elevated triglycerides, abdominal obesity, and other maladaptive metabolic and inflammatory aberrations. As the molecular mechanisms linking cardiovascular disease and metabolic disorders are investigated, endocannabinoids have emerged as molecules of interest. The endocannabinoid system (ECS) of biologically active lipids has been implicated in several conditions, including chronic liver disease, osteoporosis, and more recently in cardiovascular diseases. The gut microbiome is a major regulator of inflammatory and metabolic signaling in the host, and if disrupted, has the potential to drive metabolic and cardiovascular diseases. Extensive studies have unraveled the impact of the gut microbiome on host physiology, with recent reports showing that gut microbes exquisitely control the ECS, with significant influences on host metabolic and cardiac health. In this review, we outline how modulation of the gut microbiome affects host metabolism and cardiovascular health via the ECS, and how these findings could be exploited as novel therapeutic targets for various metabolic and cardiac diseases.

The endocannabinoid system

For many, the use of plants and herbal remedies serve as their primary pharmacies. As was meticulously recorded in the world's oldest known pharmacopoeia, the Pen-ts'ao Ching (The Herbal), Cannabis sativa (C. sativa) was used to alleviate an array of conditions, including intestinal constipation and rheumatism. Thus, cannabinoids constitute some of the oldest known medicinal compounds, with records of their earliest usage over 5,000 years ago in China. Cannabinoids are bioactive compounds isolated from C. sativa, known to produce a euphoric effect, and can also act as an analgesic, bronchodilator, appetite stimulant and muscle relaxant¹. Since cannabinoids are primarily synthesized within plants, the term "endocannabinoid" was established for the endogenous molecules within mammals that bind and activate the same G-protein coupled receptors as cannabinoids. These receptors are known as the cannabinoid 1 (CB_1) and cannabinoid 2 (CB_2) receptors. This network comprising the CB₁, and CB₂ receptors, as well as the biologically active endogenous lipids that act on them, is known as the endocannabinoid system (ECS). The ECS has been implicated in several conditions, including chronic liver disease^{2,3} and osteoporosis.^{4,5} Additionally, in recent years, the term 'endocannabinoidome (eCBome)' has also been used to describe this complex signaling network. Particularly in the context of metabolic diseases, as the eCBome encompasses over 100 fatty-acid derived, endocannabinoid-related mediators and their subsequent target proteins and enzymes, many of which play critical roles in metabolic regulation and associated cardiometabolic complications like diabetes mellitus.^{6,7}

 CB_1 and CB_2 activation generate the production of downstream secondary messengers that can modulate the mitogen-activated protein kinases (MAPK), constituents of the nuclear factor NF- κ B pathway, adenylate cyclase and specific ion

ARTICLE HISTORY

Received 20 January 2021 Revised 19 March 2021 Accepted 23 March 2021

KEYWORDS

Endocannabinoid; microbiome; metabolic syndrome; cardiovascular; probiotic; sexual dimorphism



CONTACT Crystal R. Naudin 🔯 crystal.naudin@emory.edu 😰 Division of Gastroenterology, Hepatology, and Nutrition, Department of Pediatrics, Emory University School of Medicine, 615 Michael Street, Atlanta, GA 30322, United States of America.

^{© 2021} The Author(s). Published with license by Taylor & Francis Group, LLC.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

channels.^{8,9} Thus, the ECS can have complex and diverse signaling effects. It is also important to note that endocannabinoids have been found to modulate other receptors besides CB1 and CB2, including intracellular receptors such as transient receptor potential (TRP) channels of vanilloid type-1, -2, -3, -4 or -8 (TRPV1-4, or -8), $^{8,10-13}$ constituents of the transient receptor potential cation channel subfamily V, and the peroxisome proliferatoractivated receptor $-\alpha$ and $-\gamma$ (PPAR- α and PPAR- γ).¹⁰ Other CB receptor agonists, such as N-acylethanolamines (NAE), have also been found to activate other receptors, including G-proteincoupled receptor 55 (GPR55).^{14,15} In the adipose tissue of obese individuals, GPR55 expression is increased and this receptor continues to be studied in order to better understand its metabolic and cardiovascular effects.¹⁶ Notably, AEA can also inhibit T-type Ca²⁺ channels (Ca_v3s), which play versatile roles in regulating various aspects of vascular smooth muscle function, such as pacemaker contractile activity.¹⁷ In addition, endocannabinoids can act intracellularly on mitochondrial CB1 receptors to regulate energy metabolism, with increased in situ endocannabinoids leading to decreased cellular respiration.¹⁸ Furthermore, CB₁ and CB₂ receptors differ in amino acid sequence and tertiary structure and have divergent tissue distribution. CB₁ receptors are the predominant cannabinoid receptor found in the brain and its peripheral tissues, affecting motor activity, memory processing and sensory perception. CB₁ receptors are also expressed in cardiac muscle, vascular smooth muscle cells, vascular endothelium, the kidney, liver hepatocytes, pancreatic β-cells and gastrointestinal tract.^{19,20} In contrast, CB₂ receptors are mostly present on immune cells, and their expression is upregulated in response to the secretion of proinflammatory cytokines.^{21,22} CB₂ receptors are also found in other peripheral organs, like the liver, where they have been found to play critical antifibrinogenic and immunoregulatory roles.^{23,24}

Arachidonoylethanolamide (also known as anandamide; AEA) was among the first endogenous agonists of CB₁ and CB₂ receptors to be identified and has been found to induce many of the same pharmacological effects caused by Δ^9 -THC.²⁵ While AEA is formed and released 'on-demand' by neurons in the brain, AEA can also be synthesized in the gastrointestinal tract as a degradation product of dietary arachidonic acid.²⁶ *In vivo*, AEA can be generated from the membrane phospholipid, *N*-arachidonoyl phophotidylethanolamine (NAPE), via several reaction pathways mediated by *N*-acylphosphatidylethanolamine phospholipase D (NAPE-PLD). NAPE-PLD is also a critical for the synthesis of other endocannabinoids including $2-AG^{27}$ which exhibits similar CB1 and CB2 agonist activity to AEA.²⁸

In addition to the 'true' endocannabinoids AEA and 2-AG, which are formed on-demand (as opposed to being stored in vesicles), other members of the N-acylethanolamines which have structural similarities to the 'true' endocannabinoids can affect the endocannabinoid response, although they do not directly bind to the CB_1 and CB_2 These endocannabinoid analogues receptors. include N-oleoylethanolamine (OEA, 18:1-EA), N-stearoylethanolamine (SEA, 18:0-EA), and N-palmitoylethanolamine (PEA, 16:0-EA), and are from the hydrolysis synthesized N-acylphosphatidylethanolamines.²⁹ Saturated and monounsaturated N-acylethanolamines, like OEA, SEA and PEA do not bind to cannabinoid receptors; however, they play a critical physiological role and are known to activate the peroxisome proliferator-activated receptor α (PPAR- α), as well as other nuclear and extracellular receptors. By activating these receptors, saturated and monounsaturated N-acylethanolamines mediate a variety of effects, including appetite suppression, adipogenesis and inhibiting inflammation,³⁰ and have also been implicated in the regulation of cancer cell proliferation.³¹

Through interactions with CB₁ and CB₂, AEA and 2-AG induce a myriad of bioactivities, known as the cannabinoid tetrad which includes hypothermia, catalepsy, hypo-locomotion and analgesia. Furthermore, activation of the cannabinoid receptors by AEA and 2-AG has been linked to a reduction in intraocular pressure and blood pressure, as well as bradycardia.^{32,33} Throughout various tissue types, 2-AG is detected at hundreds of times more abundant than AEA.³¹ Studies have shown that when fatty acid hydrolase (FAAH), an AEA-degrading enzyme, is inhibited or genetically deficient, the concentration of local AEA increases. This AEA abundance has a more potent influence in driving CB_1 -mediated activities than when AEA levels are low,³⁴ suggesting divergent signaling effects based on the level of AEA present.

The dual role of endocannabinoids and related molecules in inflammation

Many metabolic disorders, including those associated with cardiometabolic syndrome, are characterized by an increased systemic and cardiovascular inflammatory tone. The ECS is a complex endogenous signaling network and serves a critical role in regulating inflammation in a multitude of conditions, including celiac disease,^{35,36} irritable bowel syndrome (IBS)^{37,38} and colorectal cancer.³⁹⁻⁴¹ Furthermore, the ECS has been upregulated in a number of inflammatory, neurodegenerative, and cardiovascular metabolic diseases, a phenomenon that has been suggested as autoprotective to hinder disease progression.⁴² Thus, the inhibition of endocannabinoid degradation to enhance endocannabinoid tone emerged as a potential treatment approach for various diseases. The two leading therapeutic approaches propose to inhibit the cellular uptake of endocannabinoids, or to inhibit the enzymes that catalyze endocannabinoid degradation, such as fatty acid hydrolase (FAAH) or monoacylglycerol lipase (MAGL).²⁵

Endocannabinoids have been shown to help mitigate inflammation primarily through the activation of the CB₂ receptor, which subsequently anti-inflammatory increases cytokines and decreases inflammatory cell levels in a variety of disease processes, including lipopolysaccharide (LPS)-induced pulmonary inflammation,⁴³ trinitrobenzene sulfonic acid (TNBS)-induced colitis,44 experimental hepatitis⁴⁵ and multiple sclerosis.⁴⁶ Intraperitoneal injection of FAAH inhibitor URB597 or genetic knockout of FAAH resulted in decreased inflammation and a decrease in the inflammatory cytokines, tumor necrosis factoralpha (TNF- α) and interleukin-1 beta (IL-1 β).⁴⁷ These anti-inflammatory effects were dependent on CB₂ receptor activity since the responses were blocked with the addition of CB₂ antagonist (SR144528).⁴⁷When AEA levels were enhanced in in vivo models by a selective uptake inhibitor VDM-11, a reduced inflammatory tone in

dinitrobenzene sulfonic acid (DNBS)-induced colitis was observed (Table 1).⁵⁸ However, in this study, FAAH inhibition via AA-5-HT had no effect on either attenuating colitis or lowering inflammatory tone.⁵⁸ Similarly, anti-inflammatory effects were observed when 2-AG levels were enhanced (Figure 1), including a decrease in LPS-induced levels of TNF- α , IL-1 β , IL-6, IL-10 expression in the frontal cortex, as well as a decrease in cell death in hepatic injury models (Table 1).^{48,49} Finally, inducing CB₂ activity in mice reduced atherosclerotic lesions and decreased TNF- α and IL-6 levels (Table 1).⁵⁰

Contrary to their role in responding to and inhibiting inflammation, cannabinoids have also been implicated as drivers of systemic inflammation. CB₁ agonists like AEA have been referred to as 'gate openers,' as they induce gut permeability (Figure 2).⁵⁹ The deletion of NAPE-PLD led to a decrease in N-acylethanolamines, like oleoylethanolamide (OEA), palmitoylethanolamide (PEA) and stearoylethanolamide (SEA) and was linked to increased inflammation in adipose tissue and is associated with increased macrophage infiltration and cytokine production, as well as a reduction in thermogenic browning activity.⁶⁰ Interestingly, AEA levels were unaffected suggesting an alternative. compensatory synthesis pathway.⁶⁰ Additionally, increased levels of 2-AG have been linked to an increase in the recruitment of leukocytes, pro-inflammatory cytokine secretion, and generation of reactive oxygen species (ROS).⁵¹⁻⁵⁴ Many pro-inflammatory effects have been associated with increased 2-AG levels.⁴² For example, mast cells secreted more TNF- α when 2-AG levels were enhanced in vitro (Table 1).⁵⁷ Increased endocannabinoid levels have also been found to drive ROS generation and enhance leukocyte recruitment in the onset of nephropathy and cardiomyopathy.61,62

It is also important to recognize that although AEA and 2-AG have been implicated in many disease states, they do not always exert their immunomodulatory effects through the cannabinoid receptors. For example, AEA and 2-AG can be metabolized to yield other lipid components, and thus serve as a source of arachidonic acid in the biosynthesis of other pro-inflammatory lipids.⁴² The inhibition of FAAH and MAGL increases the levels of endocannabinoids (since they are not

Table 1. The dual	l roles of endocannabinoid signalir	ng in inflammation.			
-	Model	Treatment	Receptor(s)	Effects R	References
	LPS-induced inflammation & nociception	FAAH Inhibitor (URB597)	CB ₂	Decreased TNF-a & IL-1β	47
Anti-Inflammatory Effects	DNBS-induced colitis	AEA reuptake inhibitor (VDM-11)	CB ₁	levels Decreased Inflammation Reduced inflammatory tone	44
	LPS-induced cytokine production in frontal cortex	MAGL Inhibitor (JZL184)	CB ₁ , CB ₂	Decreased TNF-α, IL-1β, IL-6, & IL-10 levels	48,49
	Apoe -/- mice (atherosclerosis)	Low doses of THC which drives endocannabinoid-signaling via CB receptors	CB ₂	Decreased TNF- α & IL-6 levels Decreased atherosclerotic lesions	20
	Human leulkocytes	2-AG	CB ₁ , CB ₂	Increased TNF-α secretion and leukocyte migration	51-53
Pro-Inflammatory Effects	CB ₂ -/- mice	Genetic knockout of CB_2 in mice	Increased CB ₁ activity	Increased M1 (pro-inflammatory) macrophages in liver & adipose tissue	54,55
	FAAH -/- mice	FAAH inhibition & genetic knockout in mice	COX-2, LOX, & p450 oxidation	Increased pro-inflammatory prostaglandins	56
	Mast cells	2-AG	CB_2	Increased TNF-α secretion	57



Figure 1. Anti-inflammatory signaling downstream of CB₂ receptor activation. CB₂ receptor activation by endocannabinoids, AEA and 2-AG, inhibit inflammation through reduced LPS and proinflammatory cytokines. While contradictory to the pro-inflammatory roles of endocannabinoids, in some settings it has been shown that inhibition of FAAH and MAGL (key enzymes that degrade endocannabinoids) can drive inflammation. These effects are CB₂ receptor-dependent, as shown by the loss of anti-inflammatory effects through CB₂ receptor blockade using CB₂ antagonists (i.e. SR144528), whereas blockade of CB₁ has no effect. Created with BioRender.com

being degraded) and leads to increased oxidation of endocannabinoids by cyclooxygenase-2 (COX-2), lipoxygenase (LOX) and p450 that drives inflammation.⁴² Consistent with this notion, increased pro-inflammatory prostaglandins were detected in FAAH knockout mice or conditions with reduced FAAH expression, suggesting increased oxidation of endocannabinoids by COX-2 (Table 1).⁵⁶ Notably, LPS inhibits FAAH and promotes COX-2 expression,⁶³ and these data, when taken together, suggests that endocannabinoids increase gut permeability in a positive, pro-inflammatory feedback loop.

Furthermore, when analyzing the dual roles of endocannabinoids in both driving and reducing inflammatory states, it is important to consider the redundancy in eCBome pathways.⁶⁴ Not only can the same enzymes hydrolyze multiple different endocannabinoid substrates, but hydrolytic or oxidative metabolism of endocannabinoids can lead to the production of metabolites that are also capable of activating the same eCBome receptors.⁶⁴ Thus, the role of eCBome redundancy in activating distinct receptors and altering metabolic enzyme expression may be particularly relevant when considering the conflicting inflammatory phenotypes associated with endocannabinoid signaling.

Deducing the links between the endocannabinoid system, gut microbiome, and gut permeability

The ECS has been shown to mitigate inflammatory responses in gut diseases through host-bacterial interactions in *ex vivo* and *in vivo* models, with studies indicating a mechanism by which the gut microbiota can modulate the expression and activation of CB₁ and CB₂ receptors, as well as downstream inflammation.⁶⁵ Increased permeability of the gut and intestine can be devastating, as leakage



Figure 2. Pro-inflammatory signaling downstream of endocannabinoid receptor activation. Elevated endocannabinoid system tone can drive inflammation. Predominantly CB₁ agonists like AEA can induce gut permeability, causing a 'leaky' gut. This can lead to the onset of metabolic endotoxemia as toxins such as LPS traverse the epithelial barrier to drive inflammatory signaling. 2-AG activation of CB₂ receptors also has pro-inflammatory signaling effects in pathological settings, leading to leukocyte recruitment, pro-inflammatory cytokine production and ROS generation. Created with BioRender.com

of Gram-negative bacterial components can lead to the onset of metabolic endotoxemia (i.e. a rise in plasma LPS levels), which can then exacerbate metabolic disorders like insulin resistance and diabetes.⁶⁶ High-fat diets have been implicated in altering and impairing gut permeability and may catalyze the onset of systemic inflammation through the translocation of toll-like receptor (TLR) bacterial ligands to sub-epithelial compartments.⁶⁷ The ECS has emerged as a potential avenue by which the gut microbiota modulates gut permeability, adipogenesis and plasma LPS levels.^{67,68} The ECS has been implicated in regulating intestinal permeability as endocannabinoids have been shown to enhance the expression of the tight-junction protein, occludin-1, while also decreasing the expression of claudin-1, another tight-junction membrane protein that serves as a paracellular barrier.^{68,69}

Investigations into the mechanisms of how *Lactobacillus rhamnosus GG* (LGG) probiotics tighten the gut barrier demonstrate that these effects are dependent on ROS generation and subsequent activation of Nrf2 cytoprotective

pathways.^{70,71} The effects of LGG may therefore require inhibition of ECS activity, since activation of CB1 or CB2 led to decreased expression or activity of the ROS producing enzyme, cyclooxygenase, and thus suppresses the production of intracellular ROS.⁷² It was also identified that increased intestinal expression of the satiety hormone leptin by LGG probiotic treatments is also ROS dependent,⁷³ and thus may be inhibited by excessive ECS activition.

Notably, recent studies further implicate the role of gut microbiota in ECS signaling, as microbe commensal Bacteroides produces endocannabinoid-like N-acyl amides that structurally resemble OEA.74,75 These bacterialderived endocannabinoid-like molecules also interact with receptors activated the in eCBome, specifically commendamide (N-acyl-3-hydroxypalmitoyl-glycine) was shown to activate G-protein-coupled receptors (GPCRs) G2A/GPR132.74

In genetically obese leptin-deficient mice (ob/ ob) with metabolic endotoxemia arising from impaired gut barrier function, treatment with the CB₁ antagonist SR121716A resulted in the partial rescue of the efficacy of the tight junction proteins, zonula occludens-1 (ZO-1) and occludin, as well as a decrease in plasma levels of LPS.⁶⁶ Moreover, continual administration of the CB1 agonist, HU-210, over an extended period of four weeks led to an increase in bacteriaderived LPS and gut permeability in lean mice, as well as decreased mRNA expression of tight junction molecules, occludin and ZO-1.68 Cotreatment with the CB₁ antagonist, rimonabant, negated the "gate-opening" effects of HU-210, whereas co-treatment with a CB₂ receptor antagonist (SR144528) did not.⁶⁸ Furthermore, inhibition of FAAH and MAGL and the addition of AEA and 2-AG increased gut permeability through a CB₁-mediated mechanism.⁷⁶ Thus, increased AEA levels are strongly linked to increased gut permeability. Additionally, emerging research has indicated that the administration of LPS led to a decrease in FAAH expression, while also enhancing AEA levels in lymphocytes.^{63,77} This human points to a pathway whereby LPS traversing a more permeable gut could potentially increase ECS

tone and thus drive further leakiness of the gut epithelial barrier.

The endocannabinoid system's effects on gut motility

As the eCBome has been shown to have a multitude of effects in modulating energy balance and other metabolic complications, it is critical to consider the effects of endocannabinoids on gut motility. Oral THC administration has been previously shown to inhibit intestinal motility in a murine model⁷⁸ and in humans.⁷⁹ In mice fed a high-fat diet over a long-term period of eight weeks, exhibited increases in intestinal motility found to be associated with a rise in 2-AG levels and a decrease in AEA abundance.⁸⁰ Increased AEA abundance and CB1 overexpression were observed in a mouse model of paralytic ileus (impaired gut motility induced by acetic acid), while a CB₁ antagonist, SR141716A, mitigated this acetic-acid induced decrease in motility.⁸¹ Overall, most studies implicate AEA and CB1 activity in hypomotility, whereby providing further avenues for therapeutic exploration.

The endocannabinoid system: a source of appetite stimulation

In addition to changing metabolic and inflammatory signaling in the body, the endocannabinoid system has also been linked to the regulation of appetite.⁸² Hypothalamic CB₁ receptors continue to be studied for their role in the maintenance of energy balance, and their associations with the appetite-stimulating hormones, ghrelin and glucocorticoids, and the satiety hormone, leptin. Because endocannabinoids are generated de novo from the diet rather than stored in vesicles, their bioavailability also changes in response to restriction of food intake.⁸³ When the endocannabinoids, AEA and 2-AG, were injected into the hypothalamus of rats, a consequent rise in food intake occurs through a CB₁-mediated pathway.⁸⁴ In humans, hedonic eating was associated with increased peripheral levels of both 2-AG and ghrelin, while AEA, PEA, and OEA levels decreased with time following the consumption of both pleasurable and nonpleasurable food.⁸⁵ And more recently close links

have been made between dietary lipid intake and endocannabinoid levels in both males and females.^{86,87} These results may account for the phenomenon commonly referred to as the 'munchies,' in which cannabinoid use is linked with an increase in food intake. Additionally, through interactions with the mesolimbic dopaminergic pathways, endocannabinoids promote the drive to seek high reward foods.⁸⁸ When AEA was administered to the nucleus accumbens shell of rats, the CB₁ receptor was activated, and an increase in the avidity and consumption of sucrose was observed. This behavior is similar to the hedonic response that is observed when Δ^9 -tetrahydrocannabinol increases dopamine release after sucrose exposure.⁸⁹ In both in vitro and in vivo models, endocannabinoids activating the CB₁ receptor were found to augment the neuronal response to sweet taste.⁹⁰ Thus, because the CB₁ receptor is highly expressed in regions of the brain associated with reward behavior and because endocannabinoid levels increase in the limbic system following instances of food deprivation, this data suggests that increased endocannabinoid tone drives the motivation to eat and enjoy foods.^{84,91}

The cephalic phase of gastric secretion occurs before food enters the stomach, and ECS activity has been found to elicit a cephalic phase response that enhances anticipation for meals.^{9,92} In rats, upon high-fat liquid meal feeding, signals via the vagus nerve led to increased intestinal AEA and 2-AG.93 Moreover, this increased AEA and 2-AG was associated with increased food consumption, while a CB₁ antagonist inhibited food intake.⁹³ Furthermore, an enhancement in endocannabinoid tone is linked to an increase in the preferential uptake of palatable (high-calorie) food.⁹⁴ In other studies, sham-feeding emulsions consisting of linoleic acid, oleic acid and unsaturated free fatty acids contributed to an increase in jejunum endocannabinoid levels.⁹⁵ Furthermore, in a two-bottle shamfeeding preference test, rats showed a strong preference for the 18:2 free fatty acid option (highreward diet), rather than the mineral oil, but this effect was not observed when the CB1 receptor antagonists, URB447 and AM6545, were administered.⁹⁵ Thus, the ECS modulates energy balance through the regulation of feeding behavior and associated hormones in various neural circuits associated with reward-related behaviors, such as the hypothalamus.

To exert its appetite-stimulating effect, ghrelin requires efficient CB₁ receptor signaling to activate AMP-activated protein kinase (AMPK),⁹⁶ a critical enzyme that mediates the hormone's function in the hypothalamus.⁹⁷ Ghrelin failed to produce its appetite-stimulating effect in CB₁-knockout mice, and also in mice treated with rimonabant (blockade of CB₁).⁹⁶ In wild-type mice, ghrelin activity led to an increase in the abundance of endocannabinoids in the hypothalamus.⁹⁶ It is also important to note that both cannabinoids and ghrelin catalyze AMPK activity in the heart and hypothalamus, while also causing a reduction in AMPK activity in the liver and adipose tissue.⁹⁸ AMPK is a heterotrimeric enzyme that helps to regulate energy balance and regulate the action of PPAR-y, and is thought to mediate the action of metformin, a diabetes medication that inhibits gluconeogenesis in the liver. Furthermore, enhanced 2-AG and ghrelin levels have been correlated with an increase in food consumption in humans,⁸⁵ while a rise in AEA abundance has been shown to elicit increased ghrelin production and secretion in rodent stomachs,⁹⁹ suggesting an interplay between these two systems.

Glucocorticoids produced in the adrenal glands stimulate gluconeogenesis and thus play an important role in regulating insulin and glucose metabolism. Notably, glucocorticoids also stimulate endocannabinoid-mediated inhibition of the paraventricular nucleus (PVN) neuronal activity and thus inhibit the secretion of hormones from the hypothalamus.¹⁰⁰ Fascinatingly, leptin (the "satiety" hormone) inhibits endocannabinoid-induced suppression of PVN activity.¹⁰¹ These effects may explain why enhancement of ECS tone in the hypothalamus was found to lead to peripheral insulin resistance, as well as a reduction in the efficacy of leptin.¹⁰⁰ Furthermore, CB₁ receptors have been shown to be necessary in glucocorticoid-driven metabolic syndrome, wherein CB1 knockout mice given chronic excess glucocorticoid exposure did not develop metabolic symptoms such as obesity, leptin dysregulation, hepatic steatosis and elevated blood triglycerides (unlike WT mice).¹⁰² Taken together, these data suggest an intricate balance between glucocorticoids and leptin, which is mediated by the ECS.

The complex interplay between the endocannabinoid system and gut microbiome in cardiovascular disease

The ECS continues to emerge as an area of interest, regard particularly in to cardiovascular disease.^{103,104} The connection between cannabinoid use and cardiovascular disease was recently highlighted in a highly publicized statement from the American Heart Association.¹⁰⁵ CB₁ and CB₂ receptors have been identified in human coronary endothelial and smooth muscle cells, and in the mvocardium.^{106,107} The ECS serves a beneficial role in the cardiovascular system, as it decreases oxidative stress, neutrophil infiltration, and inflammation, while also stimulating other protective pathways through CB₁ and CB₂.¹⁰³ CB₂ deficiency appears to escalate atherosclerosis,¹⁰⁸ and the CB₂ receptor has been linked to the proliferation of vascular smooth muscle cells, decreases in oxidative stress and macrophage activation, as well as an overall enhancement of endothelial function.¹⁰⁹ However, clinical administration of peripherally acting CB₁/CB₂ agonists led to adverse cardiovascular side effects.¹¹⁰ It is consistently found that inhibition of FAAH, the main degradation enzyme of AEA, drives both atherosclerosis and myocardial injury.^{111–113} The role of CB₁ receptors in atherosclerosis and CVD is still being investigated and a source of much debate, as evidence of both proand anti-atherosclerotic activity exists.^{107,114}

Inflammation is a particularly potent contributor to various cardiovascular disease etiologies, including coronary artery disease. Increased gut permeability and the subsequent onset of metabolic endotoxemia is caused by increased LPS levels in the plasma, and these conditions often give rise to the chronic low-grade inflammation found in atherosclerosis.^{115–117} Therefore, much attention has recently been devoted to the role of the gut microbiota potentially modulating the endocannato catalyze the onset binoid system of atherosclerosis.^{118,119} Additionally, studies on rat heart models have shown that the administration of the CB₂ receptor antagonist, SR144528, abrogated bacteria-derived LPS levels and heatinduced preconditioning against myocardial ischemia, while the CB₁ receptor antagonist, rimonabant, failed to elicit the same effect.¹¹⁹ These

results suggest that stress from heat shock or LPS may increase endocannabinoid production in inflammatory cells, which then can activate cardiac CB_2 receptors downstream.

Furthermore, increased absorption of endotoxins across the intestinal barrier occurs in dysbiosis and in the disruption of mucosal barrier function. This can trigger CVD pathogenesis, since the injection of endotoxins has been reported to accelerate the onset of atherosclerosis in various animal models.¹²⁰ As discussed previously, CB₁ activation can disrupt barrier function.⁶⁸ LPS leakage into systemic circulation leads to the downstream activation of TNF- α and IL-6 production, and increased circulating levels of pro-inflammatory cytokines. Thus, it is unsurprising that despite the known benefits of the ECS to cardiovascular health, there are several links between the gut microbiome, endocannabinoids and cardiovascular disease.¹²¹⁻¹²⁴

Changes in the gut microbiome and ECS in leptin-deficient mice (db/db mice), which have a mutation in the leptin receptor that confers susceptibility to obesity, insulin resistance and Type 2 diabetes, has been shown to affect apelin regulation in adipose tissue.¹²⁵ Apelin is a recently discovered adipokine that has critical roles in maintaining cardiovascular health, including the regulation of heart contractibility, angiogenesis, blood pressure, cell proliferation and fluid homeostasis.126,127 Interestingly, obese mice also exhibit enhanced apelinergic system tone, in addition to enhanced ECS tone.¹²⁵ The same increase in apelinergic system tone was observed when bacteria-derived LPS levels were also increased. While ECS tone appears to downregulate apelin and apelin G proteincoupled receptor (APJ) expression in lean mice, these effects are not observed in obese mice wherein LPS increases both apelin and ECS signaling. Adipose tissue treated with LPS and HU-210, a cannabinoid receptor agonist, saw an increase in apelin and APJ mRNA expression (Figure 3).¹²⁵ Thus, these data help elucidate the regulatory role of inflammation and the complex crosstalk occurring between the gut microbiome and endocannabinoid system, and the importance of this conversation in the onset of associated cardiometabolic complications.



Figure 3. The role of endocannabinoids in cardiovascular disease pathogenesis. Endocannabinoid system tone is increasingly being linked to cardiovascular diseases. Aberrant or overactive endocannabinoid signaling, resulting from obesity or experimental inhibition of FAAH, has been shown to cause cardiovascular inflammation, atherosclerosis and myocardial injury. The major mechanisms proposed for these effects all result from increased gut permeability, causing elevated levels of circulating LPS and bacterial metabolite TMAO, which drives inflammatory signaling and increased apelinergic signaling. Created with BioRender.com

Sexually dimorphic endocannabinoid signaling effects on the gut-cardiometabolic axis

Many studies of the sexually dimorphic effects of ECS signaling have focused on behavioral or emotional outcomes.¹²⁸ For example, low-dose THC administration increased locomotor activity in female mice but not male mice,¹²⁹ whereas higherdose cannabinoid receptor agonism decreased locomotor activity specifically in females.¹³⁰ However, very little is known about how metabolic and cardiovascular tissues are differentially affected by ECS activation in males versus females. It is not yet well-characterized how microbiome differences between males and females may impact these effects, and since metabolic and cardiovascular pathologies are also highly sexually dimorphic, further studies must be conducted to bridge this significant gap in the knowledge.

In studies of neurological tissues, CB1 receptor densities are generally higher in males in both rodent models and human subjects. This difference is equalized in mice after ovariectomy, suggesting that CB₁ receptor density is controlled by estrogen expression.¹³¹ Similarly, the endocannabinoid 2-AG is expressed differentially in male and female neural tissues and shown to be regulated by the estrous cycle.¹³² Furthermore, endocannabinoids regulate hormone expression and secretion in females.¹³³ Activation of CB₁ and CB₂ receptors in the hypothalamus leads to reduced secretion of reproductive hormones, including luteinizing hormone (LH)¹³⁴ and gonadotropin releasing hormone (GnRH).¹³⁵ This suppression of gonadal hormones by ECS activation results in reduced estrogen and progesterone release from the ovaries. Consistent with this notion, AEA activation of CB₁ receptors expressed in the ovaries has also been shown to cause direct dysfunction of ovarian cells.¹³⁶ Thus, there is bidirectional crosstalk occurring between the ECS and estrogen.

There are also differences in adipose distribution between males and females as endocannabinoids are lipophilic and may be sequestered in adipose tissues and released during lipolysis,¹³⁷ likely contributing to the sex-specific effects of endocannabinoids on metabolism. CB₁ activity in adipose tissue is also associated with ovarian dysfunction in polycystic ovary syndrome (PCOS).¹³⁸ CB₁ activity (but not CB₂) is significantly elevated in adipose tissue of patients with PCOS.¹³⁸ Notably, they also found that this increase correlated with insulin resistance.¹³⁸ Insulin resistance and obesity are key drivers of PCOS, thus increased ECS signaling may represent a link between these conditions in female patients.

Additionally, the ECS has sexually dimorphic effects on gut motility and inflammation. THC administration in three individual dosages was sufficient to slow gastric emptying in women but not men.¹³⁹ THC administration to Rhesus monkeys with simian immunodeficiency virus led to improved survival via inhibition of monocyte and macrophage-driven inflammation in the gut. However, no such effect was observed when female monkeys were tested.^{140,141} The mechanisms responsible for these effects remain largely unclear, and investigating sex-specific effects of ECS signaling on the gut-cardiometabolic axis is an important area for future study.

The oral microbiome and salivary endocannabinoids: potential predictors of cardiometabolic complications

Changes in the oral microbiome and oral ECS tone have both been linked to the onset of cardiovascular disease.^{142,143} Saliva is considered the initial secretory fluid in the digestion process following the ingestion of food and has thus garnered interest as a site for the regulation of food intake and energy balance. The saliva consists of molecules derived from local and systemic sources and is composed of water primarily, as well as mucus, electrolytes, tasting enzymes, gastrointestinal hormones like ghrelin, and bacteria necessary for mucosal protection. Endocannabinoids including AEA and 2-AG, as well as other N-acylethanolamines like palmitoylethanolamide and oleoylethanolamide are also quantifiable in human saliva and are more abundant in obese patients.¹⁴⁴ When analyzing salivary endocannabinoid and associated N-acylethanolamine levels in the fasting saliva and serum of normal-weight patients versus obese patients with insulin resistance, the obese/insulinresistant patients had higher concentrations of circulating and salivary endocannabinoids.¹⁴⁴ Furthermore, bodyweight reduction is also associated with a subsequent decrease in salivary AEA levels.¹⁴⁴ Indeed, it was found that a 5% loss of bodyweight had an impact on salivary endocannabinoid levels, as a decrease in AEA levels was observed, but not in 2-AG and associated N-acylethanolamines [82]. The use of salivary endocannabinoids as surveillance for the onset of cardiometabolic diseases remains promising, as sample collection is simple and noninvasive. It remains to be determined whether salivary endocannabinoids are biomarkers that correlate not only with obesity but also with other cardiometabolic phenotypes.

Prebiotic and probiotic treatments for the therapeutic modulation of the endocannabinoid system

The modulation of the gut microbiome through prebiotics and probiotics is a viable avenue to control downstream ECS tone and which may prove important as novel therapeutic strategies for cardiometabolic diseases. Prebiotics and probiotics are commonly utilized as tools to alter the composition of the gut microbiome. Prebiotics are beneficial non-digestible food ingredients that stimulate microbiota activity, while probiotics are live bacterial cultures that are administered to benefit host health, including favorable effects across multiple organ systems. For example, Lactococcus lactis subspecies cremoris treatments in mice induce cytoprotective and anti-inflammatory activity in a gut colitis model¹⁴⁵ as well as mitigate the deleterious effects of a high fat, high sugar diet on metabolism, in which a lowered expression of hepatic endocannabinoids was noted.¹⁴⁶ In studies examining the role of the CB₂ receptor in the gut, oral administration of *Lactobacillus acidophilus* (*L. acidophilus*) led to an upregulation of both μ -opioid and CB₂ expression in the intestinal epithelium of rats, as well as a consequent decrease in visceral sensitivity.¹⁴⁷ This effect was CB₂-dependent since the blockade of CB₂ receptors prevented this attenuation of visceral sensitivity.147 Thus, this study was the first to establish a microbial effect on host ECS tone.

The study of a mouse model with an intestinespecific knockout of myeloid differentiation primary response protein (MyD88), a key adaptor protein for Toll-like receptors, has offered further insight into how the ECS associates with intestinal dysbiosis and host metabolic health.¹⁴⁸ It was found that mice deficient in MyD88 did not experience insulin resistance and the effects of diet-induced obesity, as they had lower fat mass, enhanced glucose metabolism and regulatory T cell proliferation, and a subsequent increase in energy expenditure.¹⁴⁸ The observed beneficial effects also correlated with a reduction in AEA and 2-AG abundance.¹⁴⁸ Significantly, the observed beneficial effects were transferrable by fecal microbiome transplantation and were therefore microbially mediated.¹⁴⁸ Thus, it appears that enhanced ECS tone and the ensuing gut microbiome dysbiosis result in the divergent metabolic phenotype that causes the plethora of complications associated with many metabolic and cardiovascular disease etiologies.

Certain polyunsaturated fatty acids (PUFAs) have been shown to protect against inflammation and have been recommended as prebiotic supplements to aid in a variety of disease etiologies.^{149,150} Long-chain n-3 PUFAs (n-3 LCPUFAs) have been shown to improve hypertension, insulin resistance, liver steatosis, and heart steatosis. Diets rich in n-3 LCPUFA led to a reduction of fatty liver and inflammation in Zucker rats.¹⁵¹ These effects likely result from the observed decrease in the synthesis of arachidonic acid and its pro-inflammatory metabolites, and increased levels of docosahexaenoic acid and eicosapentaenoic acid.¹⁵² However, the exact mechanisms by which n-3 LCPUFA elicits these changes are yet to be elucidated. These results were consistent with another study where CB₁ blockade with rimonabant led to reduced fatty liver and inflammation in Zucker rats.^{153,154} Thus, it could be speculated that dietary-introduced n-3 LCPUFAs may alleviate metabolic syndrome by reducing the bioavailability of endocannabinoid precursors, thereby also decreasing downstream ECS activation of CB₁.

Blockade of the CB1 receptor (antagonist: SR141716A) in obese leptin-deficient (ob/ob) tightened the gut barrier and reduced plasma LPS, reduced weight gain, adiposity, blood glucose and hepatic inflammation.⁶⁸ In the inverse experiment, it was shown that a chronic 4-week infusion of CB receptor agonist HU-210 using mini-pumps implanted subcutaneously into lean wild-type mice lead to increased plasma LPS.⁶⁸ Altering the configuration of the gut microbiome through prebiotics, probiotics, and a high-fat diet all impact *Cnr1* (the gene encoding the CB₁ receptor) expression in the colon.⁶⁸ Furthermore, prebiotics modulate intestinal levels of endocannabinoids (via increased FAAH) leading to altered gut permeability and adipogenesis in obese leptin-deficient (ob/ *ob*) mice.⁶⁸

Outside of the gut, endocannabinoids directly regulate fat storage and their receptors, CB_1 and

CB₂, are found on adipocytes and hepatocytes where they stimulate fatty acid synthesis. When comparing the adipose tissues of *ob/ob* mice to lean wild-type controls, a significant reduction in FAAH abundance and a significant increase in NAPE-PLD, the primary enzyme involved in AEA synthesis) as well as a corresponding increase in AEA levels, were observed.⁶⁸ It was subsequently found that these changes were modifiable by the administration of prebiotics wherein colonic and adipose CB₁ expression was reduced and FAAH expression was increased. Moreover, these prebiotic-induced changes correlated with decreased adiposity in *ob/ob* mice.⁶⁸

Dietary supplementation of zebrafish with a probiotic mixture called VSL#3 (containing Streptococcus thermophilus, and various Lactobacilli and Bifidobacteria) increased ECS tone. These ECS effects include increased gene expression of endocannabinoid receptors Cnr1, Cnr2 and Trpv1, increased gene expression of abhydrolase domain containing 4 (Abhd4) which is involved in AEA synthesis, and the decreased expression of FAAH and MAGL (enzymes involved in endocannabinoid degradation).⁶⁵ This increase was associated with the induction of TLR signaling to regulate immune function.65 Another study demonstrated that irritable bowel syndrome patients supplementated with the probiotic, L. acidophilus, led to an increase in the expression of Cnr2 mRNA (encoding the CB₂ receptor) in colon epithelial cells.147 Additionally, another study found that A. muciniphila administration prevented fat mass gain, insulin resistance, adipose tissue inflammation and the onset of metabolic endotoxemia in mice.¹⁵⁵ These beneficial metabolic effects were also associated with a consequent increase in the abundance of intestinal 2-AG.¹⁵⁵ Moreover, A. muciniphila increased abundance of two eCBome lipids, palmitoyl-glycerol (1-PG) and 2-palmitoyl-glyerol (2-PG), with PPAR-a agonist activity.¹⁵⁶ Interestingly, prebiotic supplementation of oligofructose in *ob/ob* mice led to decreased *Cnr1* mRNA expression along with decreased AEA synthesis, suggesting an overall reduction in ECS tone. This correlated with a consequent decrease in fat mass and an improvement in gut barrier function.¹⁵⁷ Thus, probiotic and prebiotic supplementation may be a promising approach to regulate

endocannabinoid concentrations and the balance of CB_1 and CB_2 receptors expressed in the gut.

Conclusion and perspectives

As this review has demonstrated, the crosstalk occurring between the gut microbiome and the endocannabinoid system (ECS) appears to be bidirectional. The use of CB₁ receptor antagonists has been explored as a means to treat diet-induced obesity and alleviate associated cardiometabolic complications.^{82,158,159} Furthermore, the administration of certain probiotics, such as A. muciniphila, has been shown to ameliorate visceral sensitivity improve metabolic outcomes through and enhanced CB₂ expression.¹⁵⁵ High-fat diets have also been shown to cause chronic low-grade inflammation and initiate a state of dysbiosis, and many obesity-related and cardiometabolic diseases appear to also be associated with both dysbiosis and enhanced peripheral CB₁ signaling.^{160,161}

Low abundant bioactive lipid species such as endocannabinoids have attracted attention as novel therapeutics. It is now known that endocannabinoids are key players in regulating inflammatory signaling in the host and that endocannabinoid levels influence outcomes in both cardiovascular and metabolic diseases. It was also recently discovered that endocannabinoids can be modulated by gut microbiota and by probiotic treatments. Since probiotics represent a safe and cost-effective therapeutic paradigm, it is critical to determine how to elicit modifications to ECS signaling through the administration of specific probiotics. Several bacterial species, as well as prebiotic or dietary shifts to the microbiome that modulate the endocannabinoid system, have already been identified. Further studies are needed to understand the specific microbial products and signaling effects that contribute to maintaining cardiometabolic homeostasis and drive or prevent disease progression via the endocannabinoid system. The current knowledge of the crosstalk between the gut microbiota and the endocannabinoid system, and its subsequent impact on host cardiometabolic health has been demonstrated in this review article and others. Here, this review also explores these EC-microbiome e1911572-14 👄 R. N. KHAN ET AL.

interactions in the context of the role of salivary endocannabinoids in cardiometabolic diseases, as well as the sexually dimorphic effects associated with ECS signaling. Furthermore, we describe the relevance of these interactions both in cardiovascular and in metabolic diseases, since these are interlinked. Thus, we anticipate that future studies could be aimed at applying existing knowledge of these ECS and host-microbe interactions to develop novel therapeutic approaches to mitigate cardiometabolic risk.

Disclosure statement

No potential conflict of interest was reported by the author(s).

Funding

American Heart Association fellowship 19POST34370006 (C. R.N.); and supported in part by ;National Institutes of Health [R01DK098391, R01CA179424] (R.M.J)

ORCID

Rheinallt M. Jones () http://orcid.org/0000-0002-0415-9487 Crystal R. Naudin () http://orcid.org/0000-0001-6171-2640

References

- 1. Kogan NM, Mechoulam R. Cannabinoids in health and disease. Dialogues Clin Neurosci. 2007;9:413–430.
- Mallat A, Lotersztajn S. Endocannabinoids as novel mediators of liver diseases. J Endocrinol Invest. 2006;29:58-65.
- Mallat A, Teixeira-Clerc F, Deveaux V, Lotersztajn S. Cannabinoid receptors as new targets of antifibrosing strategies during chronic liver diseases. Expert Opin Ther Targets. 2007;11(3):403–409. doi:10.1517/ 14728222.11.3.403.
- Rossi F, Bellini G, Luongo L, Torella M, Mancusi S, De Petrocellis L, Petrosino S, Siniscalco D, Orlando P, Scafuro M, et al. The endovanilloid/endocannabinoid system: a new potential target for osteoporosis therapy. Bone. 2011;48(5):997–1007. doi:10.1016/j.bone.2011.01.001.
- 5. Bab IA. Regulation of skeletal remodeling by the endocannabinoid system. Ann N Y Acad Sci. 2007;1116 (1):414-422. doi:10.1196/annals.1402.014.
- 6. Veilleux A, Di Marzo V, Silvestri C. The expanded endocannabinoid system/endocannabinoidome as a potential target for treating diabetes mellitus. Curr Diab Rep. 2019;19(11):117. doi:10.1007/s11892-019-1248-9.

- Di Marzo V, Silvestri C. Lifestyle and metabolic syndrome: contribution of the endocannabinoidome. Nutrients. 2019;11(8):1956. doi:10.3390/nu11081956.
- Di Marzo V, De Petrocellis L. Why do cannabinoid receptors have more than one endogenous ligand? Philos Trans R Soc Lond B Biol Sci. 2012;367:3216–3228.
- Mazier W, Saucisse N, Gatta-Cherifi B, Cota D. The endocannabinoid system: pivotal orchestrator of obesity and metabolic disease. Trends Endocrinol Metab. 2015;26(10):524–537. doi:10.1016/j.tem.2015.07.007.
- Maccarrone M, Dainese E, Oddi S. Intracellular trafficking of anandamide: new concepts for signaling. Trends Biochem Sci. 2010;35(11):601–608. doi:10.1016/j. tibs.2010.05.008.
- Iannotti FA, Hill CL, Leo A, Alhusaini A, Soubrane C, Mazzarella E, Russo E, Whalley BJ, Di Marzo V, Stephens GJ, et al. Nonpsychotropic plant cannabinoids, cannabidivarin (CBDV) and cannabidiol (CBD), activate and desensitize transient receptor potential vanilloid 1 (TRPV1) channels in vitro: potential for the treatment of neuronal hyperexcitability. ACS Chem Neurosci. 2014;5(11):1131–1141. doi:10.1021/ cn5000524.
- Muller C, Morales P, Reggio PH. Cannabinoid ligands targeting TRP channels. Front Mol Neurosci. 2018;11:487. doi:10.3389/fnmol.2018.00487.
- De Petrocellis L, Orlando P, Moriello AS, Aviello G, Stott C, Izzo AA, Di Marzo V. Cannabinoid actions at TRPV channels: effects on TRPV3 and TRPV4 and their potential relevance to gastrointestinal inflammation. Acta Physiol (Oxf). 2012;204(2):255–266. doi:10.1111/ j.1748-1716.2011.02338.x.
- Ryberg E, Larsson N, Sjogren S, Hjorth S, Hermansson N-O, Leonova J, Elebring T, Nilsson K, Drmota T, Greasley PJ, et al. The orphan receptor GPR55 is a novel cannabinoid receptor. Br J Pharmacol. 2007;152(7):1092–1101. doi:10.1038/sj. bjp.0707460.
- Oka S, Nakajima K, Yamashita A, Kishimoto S, Sugiura T. Identification of GPR55 as a lysophosphatidylinositol receptor. Biochem Biophys Res Commun. 2007;362(4):928–934. doi:10.1016/j. bbrc.2007.08.078.
- 16. Moreno-Navarrete JM, Catalan V, Whyte L, Diaz-Vazquez-Martinez R, Rotellar F, Arteaga A, Guzman R, Gomez-Ambrosi J, Pulido MR, WR, The Russell et al. L-alphalysophosphatidylinositol/GPR55 system and its potential role in human obesity. Diabetes. 2012;61 (2):281-291. doi:10.2337/db11-0649.
- Chemin J, Monteil A, Perez-Reyes E, Nargeot J, Lory P. Direct inhibition of T-type calcium channels by the endogenous cannabinoid anandamide. Embo J. 2001;20(24):7033–7040. doi:10.1093/emboj/20.24.7033.
- Benard G, Massa F, Puente N, Lourenço J, Bellocchio L, Soria-Gómez E, Matias I, Delamarre A, Metna-Laurent

M, Cannich A, et al. Mitochondrial CB1 receptors regulate neuronal energy metabolism. Nat Neurosci. 2012;15(4):558–564. doi:10.1038/nn.3053.

- Joshi N, Onaivi ES. Endocannabinoid system components: overview and tissue distribution. Adv Exp Med Biol. 2019;1162:1–12.
- 20. Maccarrone M, Bab I, Biro T, Cabral GA, Dey SK, Di Marzo V, Konje JC, Kunos G, Mechoulam R, Pacher P, et al. Endocannabinoid signaling at the periphery: 50 years after THC. Trends Pharmacol Sci. 2015;36 (5):277–296. doi:10.1016/j.tips.2015.02.008.
- Lee SF, Newton C, Widen R, Friedman H, Klein TW. Differential expression of cannabinoid CB2 receptor mRNA in mouse immune cell subpopulations and following B cell stimulation. Eur J Pharmacol. 2001;423 (2-3):235-241. doi:10.1016/S0014-2999(01)01122-0.
- 22. Turcotte C, Blanchet MR, Laviolette M, Flamand N. The CB2 receptor and its role as a regulator of inflammation. Cell Mol Life Sci. 2016;73:4449–4470.
- Julien B, Grenard P, Teixeira-Clerc F, Van Nhieu JT, Li L, Karsak M, Zimmer A, Mallat A, Lotersztajn S. Antifibrogenic role of the cannabinoid receptor CB2 in the liver. Gastroenterol. 2005;128(3):742–755. doi:10.1053/j.gastro.2004.12.050.
- Lotersztajn S, Teixeira-Clerc F, Julien B, Deveaux V, Ichigotani Y, Manin S, Tran-Van-Nhieu J, Karsak M, Zimmer A, Mallat A, et al. CB2 receptors as new therapeutic targets for liver diseases. Br J Pharmacol. 2008;153(2):286–289. doi:10.1038/sj.bjp.0707511.
- 25. Borrelli F, Izzo AA. Role of acylethanolamides in the gastrointestinal tract with special reference to food intake and energy balance. Best Pract Res Clin Endocrinol Metab. 2009;23(1):33–49. doi:10.1016/j. beem.2008.10.003.
- Wang J, Ueda N. Biology of endocannabinoid synthesis system. Prostaglandins Other Lipid Mediat. 2009;89(3--4):112-119. doi:10.1016/j.prostaglandins.2008.12.002.
- Basavarajappa BS. Critical enzymes involved in endocannabinoid metabolism. Protein Pept Lett. 2007;14 (3):237–246. doi:10.2174/092986607780090829.
- Mechoulam R, Ben-Shabat S, Hanus L, Ligumsky M, Kaminski NE, Schatz AR, Gopher A, Almog S, Martin BR, Compton DR, et al. Identification of an endogenous 2-monoglyceride, present in canine gut, that binds to cannabinoid receptors. Biochem Pharmacol. 1995;50(1):83–90. doi:10.1016/0006-2952(95)00109-D.
- Muccioli GG. Endocannabinoid biosynthesis and inactivation, from simple to complex. Drug Discov Today. 2010;15 (11–12):474–483. doi:10.1016/j.drudis.2010.03.007.
- Tsuboi K, Uyama T, Okamoto Y, Ueda N. Endocannabinoids and related N-acylethanolamines: biological activities and metabolism. Inflamm Regen. 2018;38(1):28. doi:10.1186/s41232-018-0086-5.
- 31. Sugiura T, Kishimoto S, Oka S, Gokoh M. Biochemistry, pharmacology and physiology of 2-arachidonoylglycerol,

an endogenous cannabinoid receptor ligand. Prog Lipid Res. 2006;45(5):405–446. doi:10.1016/j. plipres.2006.03.003.

- Kunos G, Jarai Z, Batkai S, Goparaju SK, Ishac EJN, Liu J, Wang L, Wagner JA. Endocannabinoids as cardiovascular modulators. Chem Phys Lipids. 2000;108(1--2):159–168. doi:10.1016/S0009-3084(00)00194-8.
- 33. Batkai S, Pacher P, Osei-Hyiaman D, Radaeva S, Liu J, Harvey-White J, Offertaler L, Mackie K, Rudd MA, Bukoski RD, et al. Endocannabinoids acting at cannabinoid-1 receptors regulate cardiovascular function in hypertension. Circulation. 2004;110(14):1996–2002. doi:10.1161/01.CIR.0000143230.23252.D2.
- Kano M, Ohno-Shosaku T, Hashimotodani Y, Uchigashima M, Watanabe M. Endocannabinoidmediated control of synaptic transmission. Physiol Rev. 2009;89(1):309–380. doi:10.1152/physrev.00019.2008.
- 35. D'Argenio G, Petrosino S, Gianfrani C, Valenti M, Scaglione G, Grandone I, Nigam S, Sorrentini I, Mazzarella G, Di Marzo V, et al. Overactivity of the intestinal endocannabinoid system in celiac disease and in methotrexate-treated rats. J Mol Med (Berl). 2007;85 (5):523–530. doi:10.1007/s00109-007-0192-3.
- Burstein SH, Zurier RB. Cannabinoids, endocannabinoids, and related analogs in inflammation. Aaps J. 2009;11(1):109–119. doi:10.1208/s12248-009-9084-5.
- 37. Massa F, Marsicano G, Hermann H, Cannich A, Monory K, Cravatt BF, Ferri G-L, Sibaev A, Storr M, Lutz B, et al. The endogenous cannabinoid system protects against colonic inflammation. J Clin Invest. 2004;113(8):1202–1209. doi:10.1172/ JCI200419465.
- 38. Storr MA, Yuce B, Andrews CN, Sharkey KA. The role of the endocannabinoid system in the pathophysiology and treatment of irritable bowel syndrome. Neurogastroenterol Motil. 2008;20(8):857–868. doi:10.1111/j.1365-2982.2008.01175.x.
- 39. Grill M, Hogenauer C, Blesl A, Haybaeck J, Golob-Schwarzl N, Ferreirós N, Thomas D, Gurke R, Trötzmüller M, Köfeler HC, et al. Members of the endocannabinoid system are distinctly regulated in inflammatory bowel disease and colorectal cancer. Sci Rep. 2019;9(1):2358. doi:10.1038/s41598-019-38865-4.
- Patsos, H. A., D. J. Hicks, R. R. Dobson, A. Greenhough, N. Woodman, J. D. Lane, A. C. Williams, and C. Paraskeva. The endogenous cannabinoid, anandamide, induces cell death in colorectal carcinoma cells: a possible role for cyclooxygenase 2. Gut. 2005;54 (12):1741–1750. doi:10.1136/gut.2005.073403.
- Van Dross RT. Metabolism of anandamide by COX-2 is necessary for endocannabinoid-induced cell death in tumorigenic keratinocytes. Mol Carcinog. 2009;48 (8):724–732. doi:10.1002/mc.20515.
- Turcotte C, Chouinard F, Lefebvre JS, Flamand N. Regulation of inflammation by cannabinoids, the endocannabinoids 2-arachidonoyl-glycerol and

arachidonoyl-ethanolamide, and their metabolites. J Leukoc Biol. 2015;97:1049–1070.

- Berdyshev E, Boichot E, Corbel M, Germain N, Lagente V. Effects of cannabinoid receptor ligands on LPS-induced pulmonary inflammation in mice. Life Sci. 1998;63(8):125–129. doi:10.1016/S0024-3205(98) 00324-5.
- 44. Engel MA, Kellermann CA, Burnat G, Hahn EG, Rau T, Konturek PC. Mice lacking cannabinoid CB1-, CB2-receptors or both receptors show increased susceptibility to trinitrobenzene sulfonic acid (TNBS)-induced colitis. J Physiol Pharmacol. 2010;61:89–97.
- 45. Hegde VL, Hegde S, Cravatt BF, Hofseth LJ, Nagarkatti M, Nagarkatti PS. Attenuation of experimental autoimmune hepatitis by exogenous and endogenous cannabinoids: involvement of regulatory T cells. Mol Pharmacol. 2008;74(1):20–33. doi:10.1124/ mol.108.047035.
- 46. Mestre L, Correa F, Arevalo-Martin A, Molina-Holgado E, Valenti M, Ortar G, Di Marzo V, Guaza C. Pharmacological modulation of the endocannabinoid system in a viral model of multiple sclerosis. J Neurochem. 2005;92(6):1327–1339. doi:10.1111/ j.1471-4159.2004.02979.x.
- 47. Naidu PS, Kinsey SG, Guo TL, Cravatt BF, Lichtman AH. Regulation of inflammatory pain by inhibition of fatty acid amide hydrolase. J Pharmacol Exp Ther. 2010;334(1):182–190. doi:10.1124/ jpet.109.164806.
- 48. Kerr DM, Harhen B, Okine BN, Egan LJ, Finn DP, Roche M. The monoacylglycerol lipase inhibitor JZL184 attenuates LPS-induced increases in cytokine expression in the rat frontal cortex and plasma: differential mechanisms of action. Br J Pharmacol. 2013;169 (4):808–819. doi:10.1111/j.1476-5381.2012.02237.x.
- 49. Cao Z, Mulvihill MM, Mukhopadhyay P, Xu H, Erdélyi K, Hao E, Holovac E, Haskó G, Cravatt BF, Nomura DK, et al. Monoacylglycerol lipase controls endocannabinoid and eicosanoid signaling and hepatic injury in mice. Gastroenterol. 2013;144(4):808–817 e815. doi:10.1053/j.gastro.2012.12.028.
- Steffens S, Veillard NR, Arnaud C, Pelli G, Burger F, Staub C, Zimmer A, Frossard J-L, Mach F. Low dose oral cannabinoid therapy reduces progression of atherosclerosis in mice. Nature. 2005;434(7034):782–786. doi:10.1038/nature03389.
- 51. Gasperi V, Evangelista D, Chiurchiu V, Florenzano F, Savini I, Oddi S, Avigliano L, Catani MV, Maccarrone M. 2-Arachidonoylglycerol modulates human endothelial cell/leukocyte interactions by controlling selectin expression through CB1 and CB2 receptors. Int J Biochem Cell Biol. 2014;51:79–88. doi:10.1016/j.biocel.2014.03.028.
- 52. Siegmund SV, Qian T, De Minicis S, Harvey-White J, Kunos G, Vinod KY, Hungund B, Schwabe RF. The endocannabinoid 2-arachidonoyl glycerol induces

death of hepatic stellate cells via mitochondrial reactive oxygen species. Faseb J. 2007;21(11):2798–2806. doi:10.1096/fj.06-7717com.

- 53. Siegmund SV, Wojtalla A, Schlosser M, Zimmer A, Singer MV. Fatty acid amide hydrolase but not monoacyl glycerol lipase controls cell death induced by the endocannabinoid 2-arachidonoyl glycerol in hepatic cell populations. Biochem Biophys Res Commun. 2013;437(1):48–54. doi:10.1016/j.bbrc.2013.06.033.
- Schmitz K, Mangels N, Haussler A, Ferreiros N, Fleming I, Tegeder I. Pro-inflammatory obesity in aged cannabinoid-2 receptor-deficient mice. Int J Obes (Lond). 2016;40(2):366–379. doi:10.1038/ijo.2015.169.
- 55. Louvet A, Teixeira-Clerc F, Chobert MN, Deveaux V, Pavoine C, Zimmer A, Pecker F, Mallat A, Lotersztajn S. Cannabinoid CB2 receptors protect against alcoholic liver disease by regulating Kupffer cell polarization in mice. Hepatol. 2011;54(4):1217–1226. doi:10.1002/ hep.24524.
- 56. Weber A, Ni J, Ling KH, Acheampong A, Tang-Liu DDS, Burk R, Cravatt BF, Woodward D. Formation of prostamides from anandamide in FAAH knockout mice analyzed by HPLC with tandem mass spectrometry. J Lipid Res. 2004;45(4):757–763. doi:10.1194/jlr. M300475-JLR200.
- Yoo JM, Sok DE, Kim MR. Effect of endocannabinoids on IgE-mediated allergic response in RBL-2H3 cells. Int Immunopharmacol. 2013;17(1):123–131. doi:10.1016/j.intimp.2013.05.013.
- 58. D'Argenio G, Valenti M, Scaglione G, Cosenza V, Sorrentini I, Di Marzo V. Up-regulation of anandamide levels endogenous mechanism as an and pharmacological а strategy to limit colon inflammation. Faseb J. 2006;20(3):568-570. doi:10.1096/fj.05-4943fje.
- 59. Cani PD, Plovier H, Van Hul M, Geurts L, Delzenne NM, Druart C, Everard A. Endocannabinoids-at the crossroads between the gut microbiota and host metabolism. Nat Rev Endocrinol. 2016;12(3):133-143. doi:10.1038/nrendo.2015.211.
- 60. Geurts L, Everard A, Van Hul M, Essaghir A, Duparc T, Matamoros S, Plovier H, Castel J, Denis RGP, Bergiers M, et al. Adipose tissue NAPE-PLD controls fat mass development by altering the browning process and gut microbiota. Nat Commun. 2015;6(1):6495. doi:10.1038/ncomms7495.
- Mukhopadhyay P, Pan H, Rajesh M, Bátkai S, Patel V, Harvey-White J, Mukhopadhyay B, Haskó G, Gao B, Mackie K, et al. CB1 cannabinoid receptors promote oxidative/nitrosative stress, inflammation and cell death in a murine nephropathy model. Br J Pharmacol. 2010;160(3):657–668. doi:10.1111/j.1476-5381.2010.00769.x.
- 62. Mukhopadhyay P, Rajesh M, Batkai S, Patel V, Kashiwaya Y, Liaudet L, Evgenov OV, Mackie K, Haskó G, Pacher P, et al. CB1 cannabinoid receptors promote oxidative stress and cell death in murine

models of doxorubicin-induced cardiomyopathy and in human cardiomyocytes. Cardiovasc Res. 2010;85 (4):773–784. doi:10.1093/cvr/cvp369.

- 63. Maccarrone M, De Petrocellis L, Bari M, Fezza F, Salvati S, Di Marzo V, Finazzi-Agrò A. Lipopolysaccharide downregulates fatty acid amide hydrolase expression and increases anandamide levels in human peripheral lymphocytes. Arch Biochem Biophys. 2001;393(2):321–328. doi:10.1006/ abbi.2001.2500.
- Piscitelli F, Di Marzo V. "Redundancy" of endocannabinoid inactivation: new challenges and opportunities for pain control. ACS Chem Neurosci. 2012;3 (5):356–363. doi:10.1021/cn300015x.
- 65. Gioacchini G, Rossi G, Carnevali O. Host-probiotic interaction: new insight into the role of the endocannabinoid system by in vivo and ex vivo approaches. Sci Rep. 2017;7(1):1261. doi:10.1038/s41598-017-01322-1.
- 66. Cani PD, Amar J, Iglesias MA, Poggi M, Knauf C, Bastelica D, Neyrinck AM, Fava F, Tuohy KM, Chabo C, et al. Metabolic endotoxemia initiates obesity and insulin resistance. Diabetes. 2007;56(7):1761–1772. doi:10.2337/db06-1491.
- Cani PD, Bibiloni R, Knauf C, Waget A, Neyrinck AM, Delzenne NM, Burcelin R. Changes in gut microbiota control metabolic endotoxemia-induced inflammation in high-fat diet-induced obesity and diabetes in mice. Diabetes. 2008;57(6):1470–1481. doi:10.2337/db07-1403.
- Muccioli GG, Naslain D, Backhed F, Reigstad CS, Lambert DM, Delzenne NM, Cani PD. The endocannabinoid system links gut microbiota to adipogenesis. Mol Syst Biol. 2010;6(1):392. doi:10.1038/msb.2010.46.
- Alhamoruni A, Lee AC, Wright KL, Larvin M, O'Sullivan SE. Pharmacological effects of cannabinoids on the Caco-2 cell culture model of intestinal permeability. J Pharmacol Exp Ther. 2010;335 (1):92–102. doi:10.1124/jpet.110.168237.
- 70. Matthews JD, Owens JA, Naudin CR, Saeedi BJ, Alam A, Reedy AR, Hinrichs BH, Sumagin R, Neish AS, Jones RM, et al. Neutrophil-derived reactive oxygen orchestrates epithelial cell signaling events during intestinal repair. Am J Pathol. 2019;189 (11):2221–2232. doi:10.1016/j.ajpath.2019.07.017.
- 71. Jones RM, Desai C, Darby TM, Luo L, Wolfarth A, Scharer C, Ardita C, Reedy A, Keebaugh E, Neish A, et al. Lactobacilli modulate epithelial cytoprotection through the Nrf2 pathway. Cell Rep. 2015;12 (8):1217–1225. doi:10.1016/j.celrep.2015.07.042.
- Kim J, Watkins BA. Cannabinoid receptor antagonists and fatty acids alter endocannabinoid system gene expression and COX activity. J Nutr Biochem. 2014;25 (8):815–823. doi:10.1016/j.jnutbio.2014.03.012.
- 73. Darby TM, Naudin CR, Luo L, Jones RM. Lactobacillus rhamnosus GG-induced expression of leptin in the intestine orchestrates epithelial cell proliferation. Cell

Mol Gastroenterol Hepatol. 2020;9(4):627–639. doi:10.1016/j.jcmgh.2019.12.004.

- 74. Cohen, L. J., D. Esterhazy, S. H. Kim, C. Lemetre, R. R. Aguilar, E. A. Gordon, A. J. Pickard, J. R. Cross, A. B. Emiliano, S. M. Han, et al. Commensal bacteria make GPCR ligands that mimic human signalling molecules. Nat. 2017;549(7670):48–53. doi:10.1038/nature23874.
- 75. Lynch A, Crowley E, Casey E, Cano R, Shanahan R, McGlacken G, Marchesi JR, Clarke DJ. The bacteroidales produce an N-acylated derivative of glycine with both cholesterol-solubilising and hemolytic activity. Sci Rep. 2017;7(1):13270. doi:10.1038/s41598-017-13774-6.
- Izzo, A. A., F. Fezza, R. Capasso, T. Bisogno, L. Pinto, T. Iuvone, G. Esposito, N. Mascolo, V. Di Marzo, and F. Capasso. Cannabinoid CB1-receptor mediated regulation of gastrointestinal motility in mice in a model of intestinal inflammation. Br J Pharmacol. 2001;134 (3):563–570. doi:10.1038/sj.bjp.0704293.
- 77. Liu J, Batkai S, Pacher P, Harvey-White J, Wagner JA, Cravatt BF, Gao B, Kunos G. Lipopolysaccharide induces anandamide synthesis in macrophages via CD14/MAPK/Phosphoinositide 3-Kinase/NF-κB independently of platelet-activating factor. J Biol Chem. 2003;278(45):45034–45039. doi:10.1074/jbc. M306062200.
- Chesher GB, Dahl CJ, Everingham M, Jackson DM, Marchant-Williams H, Starmer GA. The effect of cannabinoids on intestinal motility and their antinociceptive effect in mice. Br J Pharmacol. 1973;49(4):588–594. doi:10.1111/j.1476-5381.1973.tb08534.x.
- 79. McCallum M, Soykan S, Sridhar S, Ricci R, Lange L, Plankey P. Delta-9-tetrahydrocannabinol delays the gastric emptying of solid food in humans: a double-blind, randomized study. Aliment Pharmacol Ther. 1999;13(1):77–80. doi:10.1046/j.1365-2036.1999.00441.x.
- 80. Izzo, A. A., F. Piscitelli, R. Capasso, G. Aviello, B. Romano, F. Borrelli, S. Petrosino, and V. Di Marzo. Peripheral endocannabinoid dysregulation in obesity: relation to intestinal motility and energy processing induced by food deprivation and re-feeding. Br J Pharmacol. 2009;158(5):451–461. doi:10.1111/j.1476-5381.2009.00183.x.
- Mascolo N, Izzo AA, Ligresti A, Costagliola A, Pinto L, Cascio MG, Maffia P, Cecio A, Capasso F, Marzo V, et al. The endocannabinoid system and the molecular basis of paralytic ileus in mice. Faseb J. 2002;16 (14):1973–1975. doi:10.1096/fj.02-0338fje.
- Cota D, Genghini S, Pasquali R, Pagotto U. Antagonizing the cannabinoid receptor type 1: a dual way to fight obesity. J Endocrinol Invest. 2003;26 (10):1041–1044. doi:10.1007/BF03348205.
- 83. Engeli, S., A. C. Lehmann, J. Kaminski, V. Haas, J. Janke, A. A. Zoerner, F. C. Luft, D. Tsikas, and J. Jordan. Influence of dietary fat intake on the endocannabinoid system in lean and obese subjects. Obesity

(Silver Spring). 2014;22(5):E70-76. doi:10.1002/ oby.20728.

- 84. Kirkham TC, Williams CM, Fezza F, Di Marzo V. Endocannabinoid levels in rat limbic forebrain and hypothalamus in relation to fasting, feeding and satiation: stimulation of eating by 2-arachidonoyl glycerol. Br J Pharmacol. 2002;136(4):550–557. doi:10.1038/sj. bjp.0704767.
- 85. Monteleone P, Piscitelli F, Scognamiglio P, Monteleone AM, Canestrelli B, Di Marzo V, Maj M. Hedonic eating is associated with increased peripheral levels of ghrelin and the endocannabinoid 2-arachidonoyl-glycerol in healthy humans: a pilot study. J Clin Endocrinol Metab. 2012;97(6):E917–924. doi:10.1210/jc.2011-3018.
- 86. Yagin NL, Hajjarzadeh S, Aliasgharzadeh S, Aliasgari F, Mahdavi R. The association of dietary patterns with endocannabinoids levels in overweight and obese women. Lipids Health Dis. 2020;19(1):161. doi:10.1186/s12944-020-01341-4.
- 87. Castonguay-Paradis S, Lacroix S, Rochefort G, Parent L, Perron J, Martin C, Lamarche B, Raymond F, Flamand N, Di Marzo V, et al. Dietary fatty acid intake and gut microbiota determine circulating endocannabinoidome signaling beyond the effect of body fat. Sci Rep. 2020;10(1):15975. doi:10.1038/s41598-020-72861-3.
- Monteleone P, Maj M. Dysfunctions of leptin, ghrelin, BDNF and endocannabinoids in eating disorders: beyond the homeostatic control of food intake. Psychoneuroendocrinology. 2013;38(3):312–330. doi:10.1016/j.psyneuen.2012.10.021.
- Mahler SV, Smith KS, Berridge KC. Endocannabinoid hedonic hotspot for sensory pleasure: anandamide in nucleus accumbens shell enhances 'liking' of a sweet reward. Neuropsychopharmacology. 2007;32 (11):2267–2278. doi:10.1038/sj.npp.1301376.
- 90. Soria-Gomez E, Bellocchio L, Reguero L, Lepousez G, Martin C, Bendahmane M, Ruehle S, Remmers F, Desprez T, Matias I, et al. The endocannabinoid system controls food intake via olfactory processes. Nat Neurosci. 2014;17(3):407–415. doi:10.1038/nn.3647.
- 91. Solinas M, Justinova Z, Goldberg SR, Tanda G. Anandamide administration alone and after inhibition of fatty acid amide hydrolase (FAAH) increases dopamine levels in the nucleus accumbens shell in rats. J Neurochem. 2006;98(2):408–419. doi:10.1111/j.1471-4159.2006.03880.x.
- 92. Mennella I, Di Monaco R, Balazy A, Ferracane R, Miele NA, Cavella S, Vitaglione P. Salivary endocannabinoids and N-acylethanolamines upon mastication of a semisolid food: implications in fat taste, appetite and food liking. Food Funct. 2018;9(1):476–484. doi:10.1039/C7FO01772C.
- 93. DiPatrizio NV, Astarita G, Schwartz G, Li X, Piomelli D. Endocannabinoid signal in the gut controls dietary fat

intake. Proc Natl Acad Sci U S A. 2011;108 (31):12904–12908. doi:10.1073/pnas.1104675108.

- 94. Gatta-Cherifi B, Cota D. New insights on the role of the endocannabinoid system in the regulation of energy balance. Int J Obes (Lond). 2016;40(2):210–219. doi:10.1038/ijo.2015.179.
- DiPatrizio NV, Joslin A, Jung KM, Piomelli D. Endocannabinoid signaling in the gut mediates preference for dietary unsaturated fats. Faseb J. 2013;27 (6):2513–2520. doi:10.1096/fj.13-227587.
- 96. Kola B, Farkas I, Christ-Crain M, Wittmann G, Lolli F, Amin F, Harvey-White J, Liposits Z, Kunos G, Grossman AB, et al. The orexigenic effect of ghrelin is mediated through central activation of the endogenous cannabinoid system. PLoS One. 2008;3(3):e1797. doi:10.1371/journal.pone.0001797.
- Hardie DG, Ross FA, Hawley SA. AMPK: a nutrient and energy sensor that maintains energy homeostasis. Nat Rev Mol Cell Biol. 2012;13(4):251–262. doi:10.1038/ nrm3311.
- 98. Kola B, Hubina E, Tucci SA, Kirkham TC, Garcia EA, Mitchell SE, Williams LM, Hawley SA, Hardie DG, Grossman AB, et al. Cannabinoids and ghrelin have both central and peripheral metabolic and cardiac effects via AMP-activated protein kinase. J Biol Chem. 2005;280(26):25196–25201. doi:10.1074/jbc. C500175200.
- Zbucki RL, Sawicki B, Hryniewicz A, Winnicka MM. Cannabinoids enhance gastric X/A-like cells activity. Folia Histochem Cytobiol. 2008;46(2):219–224. doi:10.2478/v10042-008-0033-4.
- 100. Malcher-Lopes R, Di S, Marcheselli VS, Weng F-J, Stuart CT, Bazan NG, Tasker JG. Opposing crosstalk between leptin and glucocorticoids rapidly modulates synaptic excitation via endocannabinoid release. J Neurosci. 2006;26(24):6643–6650. doi:10.1523/ JNEUROSCI.5126-05.2006.
- 101. Di S, Malcher-Lopes R, Halmos KC, Tasker JG. Nongenomic glucocorticoid inhibition via endocannabinoid release in the hypothalamus: a fast feedback mechanism. J Neurosci. 2003;23(12):4850–4857. doi:10.1523/JNEUROSCI.23-12-04850.2003.
- 102. Bowles NP, Karatsoreos IN, Li X, Vemuri VK, Wood J-A, Li Z, Tamashiro KLK, Schwartz GJ, Makriyannis AM, Kunos G, et al. A peripheral endocannabinoid mechanism contributes to glucocorticoid-mediated metabolic syndrome. Proc Natl Acad Sci U S A. 2015;112(1):285–290. doi:10.1073/pnas.1421420112.
- 103. Tuma RF, Steffens S. Targeting the endocannabinod system to limit myocardial and cerebral ischemic and reperfusion injury. Curr Pharm Biotechnol. 2012;13 (1):46–58. doi:10.2174/1389201127988686665.
- 104. Dewald O, D GD. The role for the endocannabinoid system in cardioprotection and myocardial adaptation. Croatia, Intech Open; 2016.

- 105. Page RL 2nd, Allen LA, Kloner RA, et al. Medical Marijuana, recreational cannabis, and cardiovascular health: a scientific statement from the American heart association. Circ. 2020;142(10):e131-e152. doi:10.1161/ CIR.000000000000883.
- 106. Liu J, Gao B, Mirshahi F, Sanyal AJ, Khanolkar AD, Makriyannis A, Kunos G. Functional CB1 cannabinoid receptors in human vascular endothelial cells. Biochem J. 2000;346(3):835–840. doi:10.1042/bj3460835.
- 107. Steffens S, Pacher P. Targeting cannabinoid receptor CB2 in cardiovascular disorders: promises and controversies. Br J Pharmacol. 2012;167(2):313–323. doi:10.1111/j.1476-5381.2012.02042.x.
- 108. Netherland CD, Pickle TG, Bales A, Thewke DP. Cannabinoid receptor type 2 (CB2) deficiency alters atherosclerotic lesion formation in hyperlipidemic Ldlr-null mice. Atherosclerosis. 2010;213(1):102–108. doi:10.1016/j.atherosclerosis.2010.07.060.
- 109. Carbone F, Mach F, Vuilleumier N, Montecucco F. Cannabinoid receptor type 2 activation in atherosclerosis and acute cardiovascular diseases. Curr Med Chem. 2014;21(35):4046–4058. doi:10.2174/ 0929867321666140915141332.
- 110. Kalliomaki J, Annas P, Huizar K, Clarke C, Zettergren A, Karlsten R, Segerdahl M. Evaluation of the analgesic efficacy and psychoactive effects of AZD1940, a novel peripherally acting cannabinoid agonist, in human capsaicin-induced pain and hyperalgesia. Clin Exp Pharmacol Physiol. 2013;40 (3):212–218. doi:10.1111/1440-1681.12051.
- 111. Lenglet S, Thomas A, Soehnlein O, Montecucco F, Burger F, Pelli G, Galan K, Cravatt B, Staub C, Steffens S, et al. Fatty acid amide hydrolase deficiency enhances intraplaque neutrophil recruitment in atherosclerotic mice. Arterioscler Thromb Vasc Biol. 2013;33 (2):215–223. doi:10.1161/ATVBAHA.112.300275.
- 112. Hoyer FF, Khoury M, Slomka H, Kebschull M, Lerner R, Lutz B, Schott H, Lütjohann D, Wojtalla A, Becker A, et al. Inhibition of endocannabinoid-degrading enzyme fatty acid amide hydrolase increases atherosclerotic plaque vulnerability in mice. J Mol Cell Cardiol. 2014;66:126–132. doi:10.1016/j.yjmcc.2013.11.013.
- 113. Mukhopadhyay P, Horvath Β, Rajesh М, Matsumoto S, Saito K, Bátkai S, Patel V, Tanchian G, Gao RY, Cravatt BF, et al. Fatty acid amide hydrolase is а key regulator of endocannabinoid-induced myocardial tissue injury. 2011;50(1):179-195. Free Radic Biol Med. doi:10.1016/j.freeradbiomed.2010.11.002.
- 114. Dol-Gleizes F, Paumelle R, Visentin V, Mares A-M, Desitter P, Hennuyer N, Gilde A, Staels B, Schaeffer P, Bono F, et al. Rimonabant, a selective cannabinoid CB1 receptor antagonist, inhibits atherosclerosis in LDL receptor-deficient mice. Arterioscler Thromb Vasc Biol. 2009;29(1):12–18. doi:10.1161/ ATVBAHA.108.168757.

- 115. Bullon P, Cordero MD, Quiles JL, Morillo JM, Del Carmen Ramirez-tortosa M, Battino M. Mitochondrial dysfunction promoted by Porphyromonas gingivalis lipopolysaccharide as a possible link between cardiovascular disease and periodontitis. Free Radic Biol Med. 2011;50(10):1336–1343. doi:10.1016/j. freeradbiomed.2011.02.018.
- Zeuke S, Ulmer AJ, Kusumoto S, Katus HA, Heine H. TLR4-mediated inflammatory activation of human coronary artery endothelial cells by LPS. Cardiovasc Res. 2002;56(1):126–134. doi:10.1016/S0008-6363(02) 00512-6.
- 117. Sieve I, Ricke-Hoch M, Kasten M, Battmer K, Stapel B, Falk CS, Leisegang MS, Haverich A, Scherr M, Hilfiker-Kleiner D, et al. A positive feedback loop between IL-1β, LPS and NEU1 may promote atherosclerosis by enhancing a pro-inflammatory state in monocytes and macrophages. Vascul Pharmacol. 2018;103-105:16–28. doi:10.1016/j.vph.2018.01.005.
- Lagneux C, Lamontagne D. Involvement of cannabinoids in the cardioprotection induced by lipopolysaccharide. Br J Pharmacol. 2001;132 (4):793-796. doi:10.1038/sj.bjp.0703902.
- 119. Pacher P, Steffens S. The emerging role of the endocannabinoid system in cardiovascular disease. Semin Immunopathol. 2009;31(1):63-77. doi:10.1007/s00281-009-0145-8.
- 120. Westerterp M, Berbee JF, Pires NM, Van Mierlo GJD, Kleemann R, Romijn JA, Havekes LM, Rensen PCN. Apolipoprotein C-I is crucially involved in lipopolysaccharide-induced atherosclerosis development in apolipoprotein E-knockout mice. Circ. 2007;116(19):2173–2181. doi:10.1161/ CIRCULATIONAHA.107.693382.
- 121. Pi-Sunyer FX, Aronne LJ, Heshmati HM, Devin J, Rosenstock J, RIO-North America Study Group FT, Group RI-NAS. Effect of rimonabant, a cannabinoid-1 receptor blocker, on weight and cardiometabolic risk factors in overweight or obese patients: RIO-North America: a randomized controlled trial. JAMA. 2006;295(7):761–775. doi:10.1001/jama.295.7.761.
- 122. Popa C, Netea MG, Van Riel PL, Van Der Meer JW, Stalenhoef AF. The role of TNF-α in chronic inflammatory conditions, intermediary metabolism, and cardiovascular risk. J Lipid Res. 2007;48(4):751–762. doi:10.1194/jlr.R600021-JLR200.
- 123. Libby P. Inflammation in atherosclerosis. Arterioscler Thromb Vasc Biol. 2012;32(9):2045–2051. doi:10.1161/ ATVBAHA.108.179705.
- 124. Moludi J, Alizadeh M, Lotfi Yagin N, Pasdar Y, Nachvak SM, Abdollahzad H, Sadeghpour Tabaei A. New insights on atherosclerosis: a cross-talk between endocannabinoid systems with gut microbiota. J Cardiovasc Thorac Res. 2018;10(3):129–137. doi:10.15171/jcvtr.2018.21.
- 125. Geurts L, Lazarevic V, Derrien M, Everard A, Van Roye M, Knauf C, Valet P, Girard M, Muccioli GG,

François P, et al. Altered gut microbiota and endocannabinoid system tone in obese and diabetic leptin-resistant mice: impact on apelin regulation in adipose tissue. Front Microbiol. 2011;2:149. doi:10.3389/fmicb.2011.00149.

- 126. Scimia MC, Hurtado C, Ray S, Metzler S, Wei K, Wang J, Woods CE, Purcell NH, Catalucci D, Akasaka T, et al. APJ acts as a dual receptor in cardiac hypertrophy. Nat. 2012;488(7411):394–398. doi:10.1038/nature11263.
- 127. Maguire JJ, Kleinz MJ, Pitkin SL, Davenport AP. [Pyr1] apelin-13 identified as the predominant apelin isoform in the human heart: vasoactive mechanisms and inotropic action in disease. Hypertens. 2009;54(3):598–604. doi:10.1161/HYPERTENSIONAHA.109.134619.
- 128. Anderson BM, Rizzo M, Block RI, Pearlson GD, O'Leary DS. Sex differences in the effects of marijuana on simulated driving performance. J Psychoact Drugs. 2010;42(1):19–30. doi:10.1080/02791072.2010.10399782.
- 129. Wiley JL. Sex-dependent effects of Δ9-tetrahydrocannabinol on locomotor activity in mice. Neurosci Lett. 2003;352(2):77–80. doi:10.1016/j. neulet.2003.08.050.
- Tseng AH, Craft RM. Sex differences in antinociceptive and motoric effects of cannabinoids. Eur J Pharmacol. 2001;430(1):41–47. doi:10.1016/S0014-2999(01)01267-5.
- Riebe CJ, Hill MN, Lee TT, Hillard CJ, Gorzalka BB. Estrogenic regulation of limbic cannabinoid receptor binding. Psychoneuroendocrinol. 2010;35 (8):1265–1269. doi:10.1016/j.psyneuen.2010.02.008.
- 132. Krebs-Kraft DL, Hill MN, Hillard CJ, McCarthy MM. Sex difference in cell proliferation in developing rat amygdala mediated by endocannabinoids has implications for social behavior. Proc Natl Acad Sci U S A. 2010;107(47):20535–20540. doi:10.1073/ pnas.1005003107.
- 133. Walker OS, Holloway AC, Raha S. The role of the endocannabinoid system in female reproductive tissues. J Ovarian Res. 2019;12(1):3. doi:10.1186/ s13048-018-0478-9.
- 134. Bloch E, Thysen B, Morrill GA, Gardner E, Fujimoto G. Effects of cannabinoids on reproduction and development. Vitam Horm. 1978;36:203–258.
- 135. Gammon CM, Freeman GM Jr., Xie W, Petersen SL, Wetsel WC. Regulation of gonadotropin-releasing hormone secretion by cannabinoids. Endocrinol. 2005;146 (10):4491–4499. doi:10.1210/en.2004-1672.
- 136. Adashi EY, Jones PB, Hsueh AJ. Direct antigonadal activity of cannabinoids: suppression of rat granulosa cell functions. Am J Physiol. 1983;244(2):E177–185. doi:10.1152/ajpendo.1983.244.2.E177.
- 137. Gunasekaran N, Long LE, Dawson BL, Hansen GH, Richardson DP, Li KM, Arnold JC, McGregor IS. Reintoxication: the release of fat-stored Δ 9-tetrahydrocannabinol (THC) into blood is enhanced by food deprivation or ACTH exposure. Br

J Pharmacol. 2009;158(5):1330–1337. doi:10.1111/ j.1476-5381.2009.00399.x.

- 138. Juan CC, Chen KH, Wang PH, Hwang JL, Seow KM. Endocannabinoid system activation may be associated with insulin resistance in women with polycystic ovary syndrome. Fertil Steril. 2015;104(1):200–206. doi:10.1016/j.fertnstert.2015.03.027.
- 139. Esfandyari T, Camilleri M, Ferber I, Burton D, Baxter K, Zinsmeister AR. Effect of a cannabinoid agonist on gastrointestinal transit and postprandial satiation in healthy human subjects: a randomized, placebo-controlled study. Neurogastroenterol Motil. 2006;18(9):831–838. doi:10.1111/j.1365-2982.2006.00834.x.
- 140. Molina PE, Amedee AM, LeCapitaine NJ, Zabaleta J, Mohan M, Winsauer PJ, Vande Stouwe C, McGoey RR, Auten MW, LaMotte L, et al. Modulation of gut-specific mechanisms by chronic δ9-tetrahydrocannabinol administration in male rhesus macaques infected with simian immunodeficiency virus: a systems biology analysis. AIDS Res Hum Retroviruses. 2014;30(6):567–578. doi:10.1089/aid.2013.0182.
- 141. Amedee AM, Nichols WA, LeCapitaine NJ, Stouwe CV, Birke LL, Lacour N, Winsauer PJ, Molina PE. Chronic Δ9-tetrahydrocannabinol administration may not attenuate simian immunodeficiency virus disease progression in female rhesus macaques. AIDS Res Hum Retroviruses. 2014;30(12):1216–1225. doi:10.1089/ aid.2014.0108.
- 142. Nakajima Y, Furuichi Y, Biswas KK, Hashiguchi T, Kawahara K-I, Yamaji K, Uchimura T, Izumi Y, Maruyama I. Endocannabinoid, anandamide in gingival tissue regulates the periodontal inflammation through NF-κB pathway inhibition. FEBS Lett. 2006;580 (2):613–619. doi:10.1016/j.febslet.2005.12.079.
- Leishman SJ, Do HL, Ford PJ. Cardiovascular disease and the role of oral bacteria. J Oral Microbiol. 2010;2 (1):5781. doi:10.3402/jom.v2i0.5781.
- 144. Matias I, Gatta-Cherifi B, Tabarin A, Clark S, Leste-Lasserre T, Marsicano G, Piazza PV, Cota D. Endocannabinoids measurement in human saliva as potential biomarker of obesity. PLoS One. 2012;7(7): e42399. doi:10.1371/journal.pone.0042399.
- 145. Darby TM, Owens JA, Saeedi BJ, Luo L, Matthews JD, Robinson BS, Naudin CR, Jones RM. Lactococcus lactis subsp. cremoris is an efficacious beneficial bacterium that limits tissue injury in the intestine. iScience. 2019;12:356–367. doi:10.1016/j.isci.2019.01.030.
- 146. Naudin CR, Maner-Smith K, Owens JA, Wynn GM, Robinson BS, Matthews JD, Reedy AR, Luo L, Wolfarth AA, Darby TM, et al. Lactococcus lactis subspecies cremoris elicits protection against metabolic changes induced by a Western-style diet. Gastroenterol. 2020;159(2):639–651 e635. doi:10.1053/ j.gastro.2020.03.010.
- 147. Rousseaux C, Thuru X, Gelot A, Barnich N, Neut C, Dubuquoy L, Dubuquoy C, Merour E, Geboes K,

Chamaillard M, et al. Lactobacillus acidophilus modulates intestinal pain and induces opioid and cannabinoid receptors. Nat Med. 2007;13(1):35–37. doi:10.1038/nm1521.

- 148. Everard A, Geurts L, Caesar R, Van Hul M, Matamoros S, Duparc T, Denis RGP, Cochez P, Pierard F, Castel J, et al. Intestinal epithelial MyD88 is a sensor switching host metabolism towards obesity according to nutritional status. Nat Commun. 2014;5 (1):5648. doi:10.1038/ncomms6648.
- 149. Todoric J, Loffler M, Huber J, Bilban M, Reimers M, Kadl A, Zeyda M, Waldhäusl W, Stulnig TM. Adipose tissue inflammation induced by high-fat diet in obese diabetic mice is prevented by n-3 polyunsaturated fatty acids. Diabetologia. 2006;49(9):2109–2119. doi:10.1007/ s00125-006-0300-x.
- 150. Ajabnoor SM, Thorpe G, Abdelhamid A, Hooper L. Long-term effects of increasing omega-3, omega-6 and total polyunsaturated fats on inflammatory bowel disease and markers of inflammation: a systematic review and meta-analysis of randomized controlled trials. Eur J Nutr. 2020. doi:10.1007/s00394-020-02413-y.
- 151. Batetta B, Griinari M, Carta G, Murru E, Ligresti A, Cordeddu L, Giordano E, Sanna F, Bisogno T, Uda S, et al. Endocannabinoids may mediate the ability of (n-3) fatty acids to reduce ectopic fat and inflammatory mediators in obese Zucker rats. J Nutr. 2009;139 (8):1495–1501. doi:10.3945/jn.109.104844.
- 152. Matias I, Carta G, Murru E, Petrosino S, Banni S, Di Marzo V. Effect of polyunsaturated fatty acids on endocannabinoid and N-acyl-ethanolamine levels in mouse adipocytes. Biochim Biophys Acta. 2008;1781(1-2):52--60. doi:10.1016/j.bbalip.2007.11.001.
- 153. Serrano A, Del Arco I, Javier Pavon F, Macias M, Perez-Valero V, Rodriguez De Fonseca F. The cannabinoid CB1 receptor antagonist SR141716A (Rimonabant) enhances the metabolic benefits of long-term treatment with oleoylethanolamide in Zucker rats. Neuropharmacology. 2008;54(1):226–234. doi:10.1016/ j.neuropharm.2007.03.007.
- 154. Gary-Bobo M, Elachouri G, Gallas JF, Janiak P, Marini P, Ravinet-Trillou C, Chabbert M, Cruccioli N, Pfersdorff C, Roque C, et al. Rimonabant reduces

obesity-associated hepatic steatosis and features of metabolic syndrome in obese Zucker fa/fa rats. Hepatology. 2007;46(1):122–129. doi:10.1002/ hep.21641.

- 155. Everard A, Belzer C, Geurts L, Ouwerkerk JP, Druart C, Bindels LB, Guiot Y, Derrien M, Muccioli GG, Delzenne NM, et al. Cross-talk between Akkermansia muciniphila and intestinal epithelium controls diet-induced obesity. Proc Natl Acad Sci U S A. 2013;110(22):9066–9071. doi:10.1073/pnas.1219451110.
- 156. Depommier C, Vitale RM, Iannotti FA, Silvestri C, Flamand N, Druart C, Everard A, Pelicaen R, Maiter D, Thissen J-P, et al. Beneficial effects of Akkermansia muciniphila are not associated with major changes in the circulating endocannabinoidome but linked to higher mono-palmitoyl-glycerol levels as new PPARα agonists. Cells. 2021;10(1):185. doi:10.3390/cells10010185.
- 157. Cani PD, Possemiers S, Van De Wiele T, Guiot Y, Everard A, Rottier O, Geurts L, Naslain D, Neyrinck A, Lambert DM, et al. Changes in gut microbiota control inflammation in obese mice through a mechanism involving GLP-2-driven improvement of gut permeability. Gut. 2009;58(8):1091–1103. doi:10.1136/gut.2008.165886.
- 158. Matias I, Gonthier MP, Orlando P, Martiadis V, De Petrocellis L, Cervino C, Petrosino S, Hoareau L, Festy F, Pasquali R, et al. Regulation, function, and dysregulation of endocannabinoids in models of Adipose and β -pancreatic cells and in obesity and hyperglycemia. J Clin Endocrinol Metab. 2006;91 (8):3171–3180. doi:10.1210/jc.2005-2679.
- 159. Di Marzo V, Despres JP. CB1 antagonists for obesitywhat lessons have we learned from rimonabant? Nat Rev Endocrinol. 2009;5(11):633-638. doi:10.1038/ nrendo.2009.197.
- Di Marzo V. New approaches and challenges to targeting the endocannabinoid system. Nat Rev Drug Discov. 2018;17:623–639.
- 161. Pacher P, Steffens S, Hasko G, Schindler TH, Kunos G. Cardiovascular effects of marijuana and synthetic cannabinoids: the good, the bad, and the ugly. Nat Rev Cardiol. 2018;15:151–166.