

REVIEW

Application of immune checkpoint targets in the anti-tumor novel drugs and traditional Chinese medicine development



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Abstract Immune checkpoints are the crucial regulators of immune system and play essential roles in maintaining self-tolerance, preventing autoimmune responses, and minimizing tissue damage by regulating the duration and intensity of the immune response. Furthermore, immune checkpoints are usually overexpressed in cancer cells or noninvasive cells in tumor tissues and are capable of suppressing the antitumor response. Based on substantial physiological analyses as well as preclinical and clinical studies, checkpoint molecules have been evaluated as potential therapeutic targets for the treatment of multiple types of cancers. In the last few years, extensive evidence has supported the immunoregulatory effects of traditional Chinese medicines (TCMs). The main advantage of TCMs and natural medicine is that they usually contain multiple active components, which can act on multiple targets at the same time, resulting

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in additive or synergistic effects. The strong immune regulation function of traditional Chinese medicine on immune checkpoints has also been of great interest. For example, *Astragalus membranaceus* polysaccharides can induce anti-PD-1 antibody responses in animals, and these antibodies can overcome the exhaustion of immune cells under tumor immune evasion. Furthermore, many other TCM molecules could also be novel and effective drug candidates for the treatment of cancers. Therefore, it is essential to assess the application of immune checkpoints in the development of new drugs and TCMs. In this review, we focus on research progress in the field of immune checkpoints based on three topics: (1) immune checkpoint targets and pathways, (2) development of novel immune checkpoint-based drugs, and (3) application of immune checkpoints in the development of TCMs.

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1. Introduction

In cancer and chronic viral infections, T cells are exposed to persistent antigen stimulation, leading to the expression of multiple inhibitory receptors, also called “immune checkpoints”. Immune checkpoints are the regulators of immune system and are involved in self-tolerance, which prevents the immune system from attacking cells indiscriminately. The concept of immune checkpoint molecules was proposed many years ago. The immune function of the human body is activated after stimulation; however, overactivation is prevented through immune checkpoint molecule applying a “brake” in order to maintain normal activation of the immune system¹.

CD8⁺ cytotoxic T lymphocytes (CTLs) and CD4⁺ T-helper (Th) 1, Th2, and Th17 cells are all subtypes of effector T cells that induce protective immunity in response to causative agents, parasites, and tumors. These T cells can also promote inflammation and participate in other immunopathology and/or autoimmunity processes. Moreover, various regulatory cells [regulatory T cells (Tregs), myeloid-derived suppressor cells (MDSCs), and M2-type macrophages] and secreted cytokines [interleukin (IL)-2, IL-10, transforming growth factor (TGF)- β , interferon (IFN)- γ] in the innate and adaptive immune systems mediate immune regulation and facilitate immune integration to affect cancer and acute and chronic infections; therefore, abnormal integration is an essential therapeutic target in clinical diagnosis and treatment². Abnormal expression and function of immune checkpoint molecules are important causes of many diseases. For example, overexpression or hyperactivation of immune checkpoint molecules can block immune function, increasing the risk of cancer. Conversely, if immune suppression of checkpoint molecules is poor or the immune checkpoint are regulated by targeted checkpoint molecule inhibitors, the body's immune function is enhanced. Currently approved checkpoint inhibitors block cytotoxic T lymphocyte antigen-4 (CTLA4), programmed cell death 1 protein (PD-1), and PD-1 ligand (PD-L1)¹. James P. Allison and Tasuku Honjo won the Tang Prize in Biopharmaceutical Science and the Nobel Prize in Physiology or Medicine in 2018 for research related to these discoveries^{3,4}.

In this review, we provide a discussion of the applications of immune checkpoints in the development of novel drugs and TCMs based on the following three topics: (1) immune checkpoint targets and pathways; (2) development of novel immune

checkpoint-based drugs; and (3) development of immune checkpoint-related TCMs.

2. Checkpoint targets and pathways with different effects

2.1. Stimulatory checkpoint molecules

The activation of stimulatory immune checkpoints can augment the impact of the immune response and immune subclasses of the tumor microenvironment. For example, enhancement of the activation, proliferation, and activities of CD8⁺ T cells, CD4⁺ T cells, natural killer (NK) cells, and macrophages can lead to increased production of inflammatory agents. Moreover, these activities inhibit the proliferation and function of MDSCs and Tregs. Five stimulatory checkpoint molecules, *i.e.*, CD27, CD40, OX40, glucocorticoid-induced tumor necrosis factor (TNF) receptor (TNFR)-related protein (GITR), and CD137, are members of the TNFR superfamily, whereas two other stimulatory checkpoint molecules, *i.e.*, CD28 and inducible T-cell costimulatory (ICOS), belong to the B7-CD28 superfamily (Table 1^{5–49}).

2.2. Inhibitory checkpoint molecules

In the last decade, inhibitory immune checkpoints have emerged as pivotal molecules preventing tumor attack by the immune system. Cancer cells exploit immune-inhibitory molecules to contrast immune interventions and induce tumor tolerance. Additionally, inhibitory checkpoint molecules are targets for cancer immunotherapy because of their potential for use in multiple types of cancers. Molecular agents that target these checkpoints represent a new frontier for cancer treatment. Immune checkpoint-targeted therapy has been shown to be helpful for the treatment of selected and even histologically different types of cancer. Unlike most currently approved antibodies that are used in cancer therapy, antibodies for blocking immune checkpoints do not target tumor cells directly; instead, they target lymphocyte receptors or their ligands to enhance endogenous antitumor activity. ThemoAb-based immunotherapy, which targets PD-1, PD-L1, or CTLA4, is the most frequently used antibody therapy. PD-1, a transmembrane receptor found on T cells from the immunoglobulin B7-CD28 family, consists of an

Table 1 Checkpoint targets with different effects and its pathways.

Checkpoint molecule	Molecule expression	Application
Stimulatory checkpoint molecules		
CD27	(1) The molecule supports antigen-specific expansion of naïve T cells and is vital for the generation of T cell memory ⁵ ; (2) A memory marker of B cells ⁶ , the activity is governed by the transient availability of its ligand, CD70, on lymphocytes and dendritic cells ⁷ ; (3) CD27 co-stimulation is known to suppresses Th17 effector cell function ⁸ .	Celldex Therapeutics is working on CDX-1127, an agonistic anti-CD27 monoclonal antibody which in animal models has been shown to be effective in the context of T cell receptor stimulation ^{9,10}
CD28	(1) The molecule is constitutively expressed on almost all human CD4 ⁺ T cells and on around half of all CD8 T cell ¹¹ ; (2) Binding with its two ligands are CD80 and CD86, expressed on dendritic cells, prompts T cell expansion ¹¹ .	CD28 was the target of the TGN1412 (superagonist) which caused severe inflammatory reactions in the first-in-man study ¹¹
CD40	(1) The molecule, found on a variety of immune system cells including antigen presenting cells has CD40L, as CD154 and transiently expressed on the surface of activated CD4 ⁺ T cells, as its ligand ¹² ; (2) CD40 signaling is known to “license” dendritic cells to mature and thereby trigger T-cell activation and differentiation ¹² .	VLST in-licensed an anti-CD40 agonist monoclonal antibody in 2012 ¹³
CD122	The molecule, which is the interleukin-2 receptor β subunit, is known to increase proliferation of CD8 ⁺ effector T cells ¹⁴ .	Therapeutics is working on NKTR-214, a CD122-biased immunestimulatory cytokine phase I results announced in 2016 ^{15,16}
CD137	The molecule (4-1BB) is bound by CD137 ligand, the result is T-cell proliferation. CD137-mediated signaling is known to protect T cells, and in particular, CD8 ⁺ T cells from activation-induced cell death ¹⁷ .	It has developed an engineered lipocalin that is bi-specific for CD137 and HER2 ¹⁸
OX40	CD134 has OX40L, or CD252, as its ligand. OX40 promotes the expansion of effector and memory T cells, it is noted for its ability to suppress the differentiation and activity of T-regulatory cells, and for its regulation of cytokine production ¹⁹ .	OX40 as a drug target primarily lies in the fact that, being transiently expressed after T-cell receptor engagement, it is to upregulate on the most antigen-activated T cells within inflammatory lesions ²⁰ . Anti-OX40 antibodies have been used in advanced cancer ²¹ . Three drugs have in development for targeting therapy ²²
GITR	The molecule is glucocorticoid-induced TNFR family related gene, prompts T cell expansion, including Treg-expansion. The ligand for GITR is mainly expressed on antigen presenting cells. Antibodies to GITR shown to promote an anti-tumor response through loss of Treg lineage stability ^{23–25} .	Transgenic (TG) therapeutics is working on anti-GITR antibodies ²⁶
ICOS	CD278 is expressed on activated T cells. Its ligand is ICOSL, expressed mainly on B cells and dendritic cells. The molecule seems to be important in T cell effector function ²⁷ .	Therapeutics is developing an ICOS agonist ²⁷
Inhibitory checkpoint molecules		
A2AR	Adenosine A2A receptor (A2AR) is regarded as an important checkpoint in cancer therapy because adenosine in the immune microenvironment, leading to the activation of the A2a receptor.	A2AR is negative immune feedback loop and the tumor microenvironment has relatively high concentrations of adenosine ²⁸
B7-H3	B7-H3 (CD276) is originally understood to be a co-stimulatory molecule ²⁹ but is now regarded as co-inhibitory ³⁰ .	MGA271 of Macro-Genics is an Fc-optimized monoclonal antibody that targets B7-H3. B7-H3 receptors have not yet been identified ^{31,32}

(continued on next page)

Table 1 (continued)

Checkpoint molecule	Molecule expression	Application
B7-H4	B7-H4 (VTCN1) is expressed by tumor cells and tumor-associated macrophages ³³ .	B7-H4 plays a role in tumor escape ³³
BTLA	CD272, short for B and T lymphocyte attenuator has HVEM (herpesvirus entry mediator) as its ligand. Surface expression of BTLA is downregulated during differentiation of human CD8 ⁺ T cells from the naive to effector cell phenotype ³⁴ .	BTLA is downregulated during differentiation of human CD8 ⁺ T cells from the naive to effector cell phenotype ³⁴ .
CTLA-4	CTLA-4 is known as CD152, is the target of melanoma drug, expression of CTLA-4 on Treg cells serves to control T cell proliferation ^{35,36} .	Yervoy is gained FDA approval ^{35,36}
IDO	IDO is a tryptophan catabolic enzyme with immune-inhibitory properties. Another important molecule is TDO, tryptophan 2,3-dioxygenase ^{37,38} .	IDO is known to suppress T and NK cells, generate and activate Treg and myeloid-derived suppressor cells, and promote tumor angiogenesis ^{37,38}
KIR	KIR is a receptor for MHC class I molecules on natural Killer cells.	Lirilumab, a monoclonal antibody to KIR, is developing
LAG3	LAG3 works to suppress an immune response by action to Treg and direct effects on CD8 ⁺ T cells ^{36,38,39} .	The phase I clinical study of anti-LAG3 (BMS-986016) is carried out ⁴⁰
Nicotinamide adenine dinucleotide phosphate NADPH oxidase isoform 2 (NOX2)	NOX2 is an enzyme of myeloid cells that generates immunosuppressive reactive oxygen species. Genetic and pharmacological inhibition of NOX2 in myeloid cells improves anti-tumor functions of adjacent NK cells and T cells and also triggers autoimmunity in humans and experimental animals ^{36,41} .	Ceplene has gained approval for use in acute myeloid leukemia within the EU ^{36,41}
PD-1	PD-1 receptor, has two ligands, PD-L1 and PD-L2. This checkpoint is the target of melanoma drug Keytruda ^{36,42} .	An advantage of targeting PD-1 can restore immune function in the tumor microenvironment ^{36,42}
TIM-3	TIM-3, expresses on activated human CD4 ⁺ T cells and regulates Th1 and Th17 cytokines ⁴³ .	TIM-3 acts as a negative regulator of Th1/Tc1 function by triggering cell death upon interaction with ligand (galectin-9) ^{36,44}
VISTA	VISTA is primarily expressed on hematopoietic cells ⁴⁵ .	The consistent expression of VISTA on leukocytes within tumors may allow VISTA blockade to be effective across a broad range of solid tumors ^{36,46}
Sialic acid-binding immunoglobulin-type lectin (SIGLEC)	SIGLEC7 is designated as CD328 and SIGLEC9 is designated as CD329, and are proteins found on the surface of various immune cells, including natural killer cells and macrophages or neutrophils, macrophages, dendritic cells and activated T-cells ⁴⁷ .	SIGLECs 7 and 9 suppress the immune function of these cells by binding to terminal sialic acid on glycans that cover the surface of cells ^{48,49}
Immune checkpoint target		Immune checkpoint pathway
Co-Stimulatory immune checkpoint targets and pathways		CD266 & CD155 CD40 & CD40L OX40 & OX40L HVEM & LIGHT CD28 & CD80 (CD86) GITR & GITR ligand CD27 & CD70 4-1BB & 4-1BBL ICOS & ICOS ligand PD1, PD-L1 CTLA-4, CD80 (CD86) B7-H3/CD276 B7-H4/B7S1/B7x HVEM/TNFRSF14, BTLA HVEM/TNFRSF14, CD160
Co-inhibitory immune checkpoint targets and pathways		CD266 & CD155 CD40 & CD40L OX40 & OX40L HVEM & LIGHT CD28 & CD80 (CD86) GITR & GITR ligand CD27 & CD70 4-1BB & 4-1BBL ICOS & ICOS ligand PD1, PD-L1 CTLA-4, CD80 (CD86) B7-H3/CD276 B7-H4/B7S1/B7x HVEM/BT LA HVEM/CD160

Table 1 (continued)

Immune checkpoint target	Immune checkpoint pathway
LAG3/CD223	LAG3/CD223
Galectin-9/LGALS9, TIM-3/HAVCR2	Galectin-9, TIM-3
Indoleamine2,3-dioxygenase/IDO	Indoleamine2,3-dioxygenase/IDO
VISTA/B7-H5/GI24	VISTA/B7-H5/GI24
CEACAM1/CD66a	CEACAM1/CD66a
SIRPalpha/CD172a, CD47	SIRPalpha, CD47
2B4/CD244, CD48/SLAMF2	2B4, CD48
TIGIT/VSTM3, CD155/PVR	TIGIT, CD155

extracellular segment, a hydrophobic transmembrane region, and an intracellular segment⁵⁰. After PD-1 binds to its ligand, the immunoreceptor tyrosine motif of the PD-1 intracellular domain is phosphorylated, and tyrosine phosphatase is recruited to the intracellular region. These phosphatases dephosphorylate key proteins in the T-cell antigen receptor (TCR) signaling pathway; inhibit downstream phosphatidylinositol 3-kinase (PI3K)/AKT/mammalian target of rapamycin (mTOR), RAS/mitogen-activated protein kinase kinase (MEK)/extracellular signal-regulated kinase (ERK), and other signaling pathways; and block the proliferation and differentiation of T cells and the production of cytokines⁵¹. To date, many new immune checkpoints have been discovered and developed as potential therapeutic targets (Table 1).

2.3. Immune checkpoint pathways

Under normal conditions, immune checkpoint molecules maintain self-tolerance and prevent immunopathology; however, their sustained expression deteriorates T-cell function. Recent advances in cancer immunotherapy involve blockade of CTLA4 (also known as CD279) and PD-1 in order to reverse T-cell exhaustion and reinvigorate immunity in metastatic melanoma and lung cancer. First, T-cell responses rely on TCR-dependent recognition of MHC/peptide complexes on antigen-presenting cells (APCs). Further engagement of costimulatory and co-inhibitory molecules guarantees the onset and limitation of T-cell activities, supporting the functions of immune checkpoints. Initial studies focused on relieving the immunosuppressive brake applied by the co-inhibitory CTLA-4 and PD-1 receptors⁵². CTLA-4 is expressed by activated Tregs and exhibits competitive binding with stimulatory CD28 ligands (CD80/CD86). PD-1 is expressed by activated and exhausted T cells, and binding to its ligands PD-L1 and PDL-2 directly inhibits TCR signaling through SHP2-mediated dephosphorylation of proximal signaling elements. Notably, recent findings have identified CD28 as a convergent regulatory target for both CTLA-4 and PD-1 and have demonstrated the regulation of intratumoral T-cell trafficking by PD-1. A schematic of these characteristics of the immune checkpoint pathway is presented in Fig. 1.

For therapeutic vaccines against chronic infections, *e.g.*, human immunodeficiency virus (HIV), human papilloma virus (HPV), hepatitis B virus, and hepatitis C virus, adjunct checkpoint blockade strategies are required, including blockade of other inhibitory receptors (*e.g.*, T-cell immunoreceptor) with immunoglobulin (Ig) and immunoreceptor tyrosine-based inhibitory motif-domains, *e.g.*, T-cell Ig and mucin domain 3 (TIM-3), lymphocyte activation gene 3 (LAG3), and V-domain Ig-containing suppressor of T-cell activation (VISTA). Different chronic viral infections and cancers are likely to influence the level, composition, and pattern

of inhibitory receptors expressed by responding T cells, thereby affecting checkpoint antibody blockade strategies. Recent advances have identified co-inhibitory receptors and new antibody-blockade therapeutic targets for T-cell exhaustion in chronic viral infections and cancer. Therefore, understanding the mechanisms of T-cell exhaustion in response to infections and cancer and the characteristics of T-cell responses will contribute to further improvement of immune checkpoint blockade strategies.

Suppression of co-inhibitory receptors shows unprecedented efficacy in the treatment of some tumors, and characterization of immunotherapy targets may increase the number of patients who can be assisted by these drugs. Immunotherapy research programs are now exploring a wide range of both co-inhibitory (*e.g.*, LAG3/TIGIT/TIM-3) and costimulatory (*e.g.*, GITR/4-1BB/OX40) receptors as individual or combination therapies. The complex immune system is regulated by a broad network of co-inhibitory and costimulatory receptors that control the type, scale, and duration of immune responses. Thus, these receptors are now recognized as promising immunotherapeutic targets for the treatment of cancers and autoimmune diseases. Table 1 lists the costimulatory and co-inhibitory immune checkpoint targets and pathways.

3. Immune checkpoints in the development of novel drugs

Inflammatory immune responses are often difficult to treat specifically because of their highly complex multitarget networks. There are two well-known immune checkpoint receptors that have been actively studied. One of the most widely studied anticancer targets is the important immune checkpoint molecule PD-1 and its ligand PD-L1. Another important checkpoint molecule is CTLA-4. Antibodies targeting these molecules are currently the most effective treatments available. Corresponding antibodies can inhibit the functions of the receptors and enhance the immunity of antitumor cells. Moreover, multiple additional immune checkpoints are being studied as promising targets for anticancer therapy.

Novel immune checkpoint molecules and drugs that regulate the expression or function of these molecules may have promising clinical applications. Drugs or drug candidates that inhibit or block inhibitory checkpoint molecules are sometimes known as checkpoint inhibitors or the immune checkpoint blockade¹.

Checkpoint inhibitors have been evaluated by various pharmaceutical companies as potential anticancer agents⁵³. The U.S. Food and Drug Administration (FDA) has approved two anti-CTLA-4 antibodies, *i.e.*, ipilimumab (Yervoy) developed by Bristol-Myers-Squibb⁵⁴ and tremelimumab developed by Pfizer⁵⁵, for the treatment of melanoma and mesothelioma, respectively. Immunotherapies targeting PD-1/PD-L1 stimulates the body's antitumor immune function by blocking the PD-1/PD-L1 signal pathway (Table 2). FDA has approved four PD-1/PD-L1

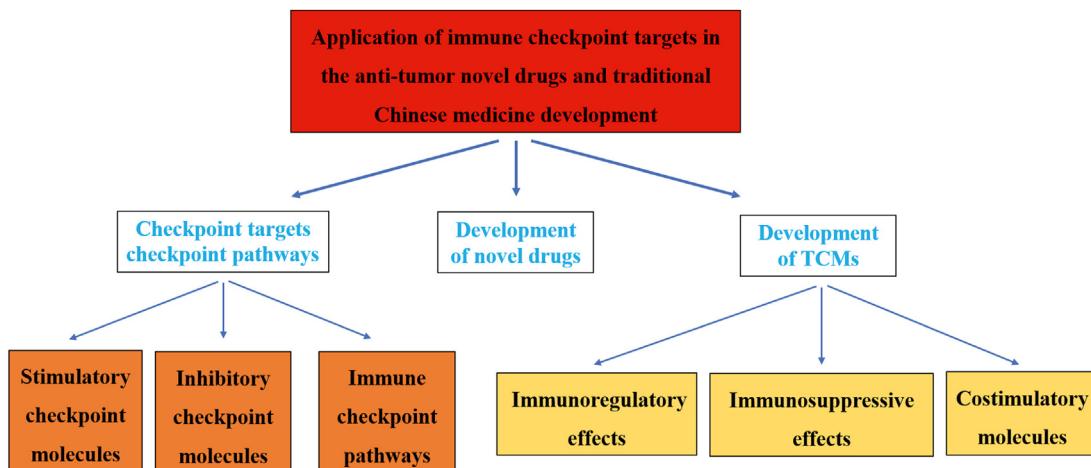


Figure 1 The mechanism of immunotherapy regulation based on immune checkpoint pathways (PD-1 and PD-L1).

antibodies, *i.e.*, nivolumab (Opdivo) developed by Bristol-Myers-Squibb⁵⁶, pembrolizumab (Keytruda) by MSD⁵⁷, atezolizumab (Tecentriq) by Roche⁵⁸, and avelumab (Bavencio) by Merck and Pfizer⁵⁹. Clinical trials for advanced melanoma, non-small cell lung cancer, kidney cancer, Hodgkin's lymphoma, head and neck squamous cell carcinoma, bladder cancer, and metastatic Meckel's cell cancer are ongoing for a variety of other tumors, including liver cancer and bowel cancer. In the European Union, nivolumab⁶⁰ has also been approved for the treatment of locally advanced or metastatic squamous non-small cell lung cancer with prior chemotherapy, and pembrolizumab has been approved for the treatment of previously and untreated unresectable or metastatic melanoma and advanced non-small cell lung cancer. In addition to the above two main immune checkpoints, immunotherapies targeting other molecules, such as TIM-3, LAG3, killer-cell Ig-like receptor (KIR), GITR, VISTA, indoleamine-pyrrole 2,3-dioxygenase (IDO), 4-1BB, and tryptophan 2,3-dioxygenase, are also being explored. For example, LAG-3 is an important immune checkpoint *in vivo* and plays a balanced regulatory role in the human immune system. LAG-3 negatively regulates T lymphocytes by binding to the extracellular domain of the ligand, thus avoiding autoimmunity caused by excessive activation of T cells. Currently, there are no drugs to target LAG-3 in the global market, but 12 drugs are in clinical research. Among them, IMP321 developed by Prima BioMed/Immutep has the fastest clinical research progress and is in stage IV treatment of breast cancer⁶¹. VISTA is a member of the B7 family. Unlike other negative checkpoint molecules, it is constitutively expressed on naïve T cells. The lack of VISTA leads to a breakdown of self-tolerance and the development of inflammatory T cell self-reactive responses. CA-170, an orally delivered dual inhibitor of VISTA and PD-L1, has shown to have clinical efficacy in phase I and II clinical trials from different advanced solid tumor types. However, further data are needed to determine whether this drug can become a new therapeutic option of cancer patients expressing VISTA⁶². TIM-3 antibodies include TSR-022 from Tesaro for the treatment of advanced or metastatic solid tumors alone or in combination with PD-1 antibody and TSR-022 from Novartis for treatment alone or in combination with PD-1 antibody PDR001 MBG-453 to target advanced malignant tumors currently in clinical trials⁶³.

In the clinical setting, some anti-PD-1 therapies are not effective in certain patients. This has led to the development of related detection technology. In addition to cancer treatment, anti-PD-1 immune checkpoint therapy is also effective in the treatment of other diseases. For example, chronic inflammation represents a central component in the pathogenesis of Alzheimer's disease; however, animal model data do not support further evaluation of PD-1 checkpoint inhibition as a therapeutic modality for Alzheimer's disease⁶⁴. Therefore, further studies are needed to assess the application of anti-PD-1 therapies in the treatment of other diseases.

Understanding the key steps in the regulation of T-cell responses has led to the groundbreaking development of immune checkpoint blocking monoclonal antibodies (mAbs) to fight cancer. The first FDA-approved mAbs have resulted in unprecedented remission in melanoma and non-small cell lung cancer, although response rates vary dramatically (10%–90%), and significant toxicity has been noted^{65,66}. This revolution in cancer therapy is now the basis for new immune checkpoint-based curative strategies.

Therapeutic immune checkpoint-blocking mAbs have dual activities inherent to their structure; the variable regions bind to immune checkpoint-epitopes, whereas the “fragment crystallizable” (Fc) region mediates targeted cell death through selective interaction with the complement molecule C1q (called complement-dependent cytotoxicity) and the Fc receptors on innate effector cells (called antibody-dependent cellular cytotoxicity or antibody-dependent cellular phagocytosis)⁶⁷. To date, two types of approved immune checkpoint-mAbs, *i.e.*, IgG1s and IgG4s, have been developed to protect or kill target cells, depending on the specific need⁶⁸. The anti-CTLA-4 antibody ipilimumab and the anti-PDL1 antibody tezozolimab are IgG1s that are expected to cause preferential Treg and tumor cell depletion, respectively^{52,65}. In contrast, the anti-PD1 antibodies nivolumab and pembrolizumab are modified IgG4s with low effector functions and mainly function by blocking PD-1 interaction with its ligands^{65,68}. Taken together, these immune checkpoint-mAbs allow better activation of effector T cells, and the combination of anti-CTLA4 and anti-PD-1/PD-L1 antibodies improves survival^{52,65}.

Table 2 Research progress of new drugs to target PD-1.

Medicine	Research institute	Application	Clinical phase
Nivolum (Opdivo)	Ono Pharmaceutical and Bristol-Myers-Squibb	Non-small cell lung cancer, malignant melanoma, renal cell carcinoma, head and neck squamous cell carcinoma, urothelium carcinoma, colorectal cancer, liver cancer, classical hodgkin lymphomas	2014-06 (Marketing)
Pembrolizumab (Keytruda)	Merck	Non-small cell lung cancer, malignant melanoma, renal cell carcinoma, head and neck squamous cell carcinoma, urothelium carcinoma, colorectal cancer, liver cancer, classical hodgkin lymphomas	2014-09 (Marketing)
Cemiplimab (Libtayo)	Sanofi and Regeneron	Metastatic squamous cell carcinoma of the skin	2018-09 (Marketing)
Toripalimab	Shanghai Junshi Biosciences (China), Suzhou Zhonghe Biomedical Technology (China)	Hepatocellular carcinoma, melanoma	2018-12 (Marketing)
Sintilimab	Innovent (China), Eli Lilly and Company	Hodgkin lymphomas	2018-12 (Marketing)
Camrelizumab	Jiangsu Hengrui Medicine (China)	Esophageal squamous cell carcinoma, advanced solid-tumor, hepatocellular carcinoma, hodgkin lymphomas	2019-05 (Marketing)
Tislelizumab	Beigene (China), Celgene, Boehringer-Ingelheim	Urothelium carcinoma, hodgkin lymphomas	2019-12 (Marketing)
CX-188	CytomX Therapeutics	Solid tumors	Phase I clinical trial
Sym-021	Symphegen	Solid tumors, lymphomas	Phase I clinical trial
STW204	Stainwei Biotech (China)	Solid tumors	Phase I clinical trial
Millamolecule	Bristol-Myers-Squibb	Immune diseases	Phase I clinical trial
XmAb20717	Xencor	Solid tumors	Phase I clinical trial
CC-90006	AnaptysBio, Celgene	Autoimmune diseases, psoriasis	Phase I clinical trial
YBL-006	Y Biologics	Advanced solid-tumor	Phase I clinical trial
ONO-4685	Merus, Ono Pharmaceutical	Autoimmune diseases	Phase I clinical trial
MGD-019 (MacroGenics)	MacroGenics	Solid tumors	Phase I clinical trial
SL-279252	Shattuck Labs	Gastric adenocarcinoma, non-small cell lung cancer, diffuse large b cell lymphoma, renal cell carcinoma, urothelium carcinoma, solid tumors, head and neck squamous cell carcinoma, hodgkin lymphomas, adeno-carcinoma of esophagogastric junction, melanoma	Phase I clinical trial
RG-6084	Roche	Hepatitis B	Phase I clinical trial
AMP-224	Amplimmune, MedImmune, National Cancer Institute, GlaxoSmithKline	Solid tumors, colorectal cancer	Phase I clinical trial
PD-1 knockout engineered T cells	Cell Biotech	Renal cell carcinoma, prostatic cancer bladder cancer	Phase I clinical trial
IMU-201	Imugene	Non-small cell lung cancer	Phase I clinical trial
MEDI-5752	MedImmune	Solid tumors	Phase I clinical trial
JTX-4014	Jounce Therapeutics	Cancer	Phase I clinical trial
RO-7247669	Roche	Solid tumors	Phase I clinical trial
Budigalimab	Abbvie	Non-small cell lung cancer, solid tumors, colorectal cancer, ovarian cancer	Phase I clinical trial
LY-3434172	Eli Lilly and Company	Solid tumors	Phase I clinical trial
INC8-086550	Incyte	Solid tumors	Phase I clinical trial
MEDI-0680	MedImmune	B cell lymphoma	Phase II clinical trial
EDP-1503	The University of Chicago, Evelo Biosciences, Merck Sharp & Dohme	Melanoma	Phase II clinical trial

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Table 2 (continued)

Medicine	Research institute	Application	Clinical phase
BCD-217	Biocad	Melanoma	Phase II clinical trial
Balistilimab	Agenus, Ludwig Institute for Cancer Research	Solid tumors, cervical cancer	Phase II clinical trial
BI-754091	Boehringer-Ingelheim	Solid tumors	Phase II clinical trial
TY-101	Tayu BioTech	Solid tumors, lymphomas	Phase II clinical trial
GS-4224	Gilead Sciences	Solid tumors	Phase II clinical trial
CA-170	Aurigenex, Curis	Cancer	Phase II clinical trial
MGD-013	MacroGenics, ZaiLab (China)	Gastric cancer, esophageal cancer	Phase III clinical trial
Sasanlimab	Pfizer	Bladder cancer	Phase III clinical trial
Retifanlimab	MacroGenics, Incyte, ZaiLab (China)	Gastric cancer, esophageal cancer	Phase III clinical trial
Spartalizumab	Novartis	Melanoma	Phase III clinical trial
Cetrelimab	Johnson & Johnson	Multiple myeloma	Phase III clinical trial
Nivolumab/Relatlimab	Bristol-Myers-Squibb	Melanoma	BLA
Dostarlimab	AnaptysBio, Tesaro, GlaxoSmithKline Biosciences	Fallopian tube cancer, endometrial cancer, peritoneal cancer, ovarian cancer	BLA
Zimberelimab	Gloria Pharmaceuticals (China), WuXi AppTec (China), Arcus Biosciences	Triple negative breast cancer, gastric cancer, solid tumors, liver cancer, hodgkin lymphomas	BLA

Immune checkpoint immunotherapy is used to improve mAb response rates in many cancers through various approaches. The first approach is to modulate the functions of mAbs through Fc engineering⁶⁹. For example, tezolizumab is a nonglycosylated IgG1 that retains its blocking activity but lacks cytotoxic functions⁵². New anti-CLTA-4 mAbs with increased or dampened effector functions have also been developed and are currently being evaluated in clinical trials⁶⁴. Other strategies targeting more co-inhibitory molecules on T cells (*e.g.*, LAG3, TIM-3, TIGIT, and VISTA) have entered clinical trials, although their biological roles are not fully understood^{52,70}. Additionally, next-generation therapeutic mAbs also include agonist agents targeting costimulatory molecules (*e.g.*, OX40, ICOS, GITR, 4-1BB, and CD40) on T cells to potentiate effector responses^{52,70}. Importantly, a combination of the above-mentioned approaches may be quite effective⁷⁰.

In cancer, immune checkpoints are prematurely activated at a stage when cancer cells are not completely eradicated, resulting in the escape of tumor cells from immune rejection. The first checkpoint inhibitor to be tested in the clinic was directed against CTLA-4 and showed impressive clinical results, including long-term survival in approximately 20% of patients with metastatic melanoma⁷¹. Second-generation checkpoint inhibitors (anti-PD-1 antibodies) show increased efficacy, not only in patients with melanoma but also in patients with advanced urothelial carcinoma, head and neck squamous cell carcinoma, renal cell carcinoma, and non-small cell lung cancer^{72–75}. Moreover, a study by Zhu et al.⁷⁶ highlighted another mechanism that could account for reduced T-cell infiltration in tumors. Indeed, sufficient T-cell infiltration in tumor tissues is often a prerequisite for the response to immunotherapy. Interfering with the Fas/Fas ligand (Fas–FasL) pathway in the tumor microenvironment could increase the efficacy of cancer immunotherapy.

The clinical success of checkpoint inhibitors has supported the potential efficacy of cancer immunotherapies^{63,77–79}. Future research challenges include the following: (1) increasing the efficacy of checkpoint blockers and the combination of checkpoint inhibitors to achieve good and lasting results in patients who cannot be treated with blockers alone; (2) studying the responses and resistance mechanisms of immune checkpoint co-inhibitors in order to understand the similarities and differences in co-inhibitory pathways and the synergistic mechanisms of combined co-inhibitory pathways with scientific evidence and to optimize the design of immune checkpoint combined therapy; (3) studying the mechanisms of durability of the immune checkpoint blockade and the time required for treatment; (4) evaluating biomarkers to predict the response to immune checkpoint blockade, help patients with stratified treatment, and predict the responses of patients to immune blockade alone therapy and assessing the necessity for combination therapy or other treatments; and (5) developing effective combination therapies of immune checkpoint blockades in order to enhance efficacy and reduce side effects, performing research on reducing tumor evasive immunity, and understanding the necessity for immunosuppression in the tumor microenvironment.

The future of antitumor immunotherapy relies on the induction of responses at multiple levels, including harnessing of other effector cells (*e.g.*, NK cells and neutrophils) in addition to T cells. As an example, the anti-NKG2A antibody monalizumab blocks inhibitory signaling in NK cells and subsets of cytotoxic T cells and further potentiates the effects of other therapeutic mAbs⁸⁰. The identification of reliable predictive biomarkers and

the combination of immune checkpoint therapies with other immunotherapies (such as adoptive T-cell therapy, oncolytic viruses, agonists for pattern recognition receptors), and radiotherapies/chemotherapies may lead to maximization of response rates⁸¹.

4. Immune checkpoints in the development of TCMs

4.1. Immunoregulatory effects of TCMs

Recent research progress on the immunomodulatory effects of TCMs on immune organs, immune cells, and immune molecules and the inhibitory effects of TCMs on inflammatory reactions, hypersensitivity reactions, autoimmune diseases, and rejection reactions has highlighted the potential use of TCMs as immune checkpoint modulators in cancer. With increasing need for scientific rigor, these traditional medicine therapies have been widely studied in recent years. Given the recent successes of immunotherapies and checkpoint blockades, there is a renewed interest in identifying novel drugs, including TCMs, with immunomodulatory effects.

TCMs can affect various immune molecules, including those that regulate human immune function and participate in various inflammatory reactions⁸². For example, Lemmon et al.⁸³ studied the effects of high-molecular-weight polysaccharides from American ginseng on human immune cells and found that American ginseng could upregulate various cytokines, including interferon- γ (IFN- γ), IL-6, and IL-23a, and downregulate other cytokines, including TNF- β and IL-13. Nakada et al.⁸⁴ treated two mouse strains (C57BL/6 and BALB/c) with the same amount of ginseng nourishing decoction to observe changes in cytokine levels in the spleen; they showed that the amount of IL-4 in spleen cells in C57BL/6 mice increased significantly, whereas the concentration of IFN- γ decreased slightly. In contrast, IFN- γ secretion increased significantly in spleen cells from BALB/c mice. These findings demonstrated that ginseng nourishing decoction could maintain the stability of the immune system by regulating the secretion of cytokines in mouse spleen cells. TCM immune-enhancers have different effects on NK cells, mononuclear macrophages, red blood cells, hematopoietic stem cells, and other immune cells. Many TCMs, such as *Astragalus membranaceus*, *Codonopsis*, *Ginseng*, *Atractylodes macrocephala*, *Poria cocos*, *Angelica sinensis*, and *Acanthopanax*, have the effect of promoting antibody generation^{85,86}. Flavonoid ingredients in *Astragalus* seeds have been shown to have immunoregulatory effects on NK cells *in vitro*⁸⁷. Moreover, by enhancing IFN- γ levels and increasing the expression of CD25 and CD69, flavonoids can significantly promote the proliferation and cytotoxicity of NK-92 cells.

Fujiwara et al.⁸⁸ demonstrated that MDSCs isolated from the spleens of tumor-bearing mice prevent the proliferation of CD4- and CD8-positive T cells. Additional studies have indicated that immature MDSCs are important drug development targets in the treatment for tumors and chronic inflammation. Chemical phytochemicals of different herbal categories, including flavonoids, terpenoids, retinoids, curcuminoids, and β -glucans, possess MDSC-dependent antitumor and anti-inflammatory properties in both *in vitro* and *in vivo* experiments. A randomized phase II study investigated the immunological efficacy of the herbal medicines Houchi-ekki-to (a spray-dried powder extract composed of equal volumes of *Cinnamomum Cortex*, *Hoelen*, *Moutan Cortex*, *Paeoniae Radix*, and *Persicae Semen*) and Keishi-bukuryo-gan (a

spray-dried powder extract made from 10 medical herbs) in combination with a personalized peptide vaccination (PPV) for curing castration-resistant prostate cancer; in this study, Tregs were defined as CD4 $^{+}$ CD25 $^{+}$ FOXP3 $^{+}$ cells in lymphocytes, and MDSCs were identified as CD33 $^{+}$ 11b $^{+}$ cells from lineage markers (CD3, CD19, CD56, and CD16) and HLA-DR cells. The monocytic subset was identified as CD14 $^{+}$ by Noriko et al.⁸⁹; they found that the frequencies of Mo-MDSCs and levels of IL-6 in the PPV-alone group were significantly increased. The study results also suggested that the combined use of herbal medicines had the potential to prevent immunosuppression induced by Mo-MDSCs or IL-6 during immunotherapy.

4.2. Immunosuppressive effects of TCMs

TCMs inhibit hypersensitivity, which is an abnormal and excessive immune response that interacts with antigenic substances under certain conditions to produce sensitized lymphocytes; these responses, if combined with re-entering antigens, can lead to disruption of the physiological functions of the body and damage to tissues⁹⁰. Li et al.⁹¹ studied the regulatory and immunosuppressive mechanisms of artemisinin by establishing a model of delayed hypersensitivity in mice; they found that artemisinin could reduce the degree of ear swelling in model mice, indicating that artemisinin could inhibit the effects of the delayed hypersensitivity reaction.

Autoimmune diseases, including rheumatoid arthritis, systemic lupus erythematosus, multiple sclerosis, and autoimmune liver, are diseases caused by the immune response to the body's own components. Tripterygium has been widely used in the treatment of autoimmune diseases and inflammatory neurodegenerative diseases⁹².

Most patients with cancer exhibit unfavorable responses to PD-1/PDL-1 inhibitors; however, PD-1/PDL-1 inhibitors can show good antitumor effects and are approved by the FDA for use in various tumor cases. Thus, increasing evidence suggests that TCMs and combination therapies may be applied to overcome this limitation (Table 3)^{93–105}. Investigators have shown that primoglucoyslcamifugin (POG) extracted from the *Saposhnikovia* root is an effective polymorphonuclear (PMN)-MDSC inhibitor. Indeed, POG can prevent the proliferation, metabolism, and immunosuppressive ability of PMN-MDSCs; enhance the tumor immunosuppressive microenvironment; and exert synergistic effects with PD-1 inhibitors in B16F10 and 4T1 mouse tumor models⁹³. Additionally, Chen et al.⁹⁴ showed that total glucosides of paeony (TGP) increase the expression of PD-1/PD-L1 in peripheral blood mononuclear cells by modulating regulatory T cells/Th cells¹⁷. Other natural plants, such as *Ganoderma lucidum*⁹⁵, *Astragalus* polysaccharides (APS)⁹⁶, and icariin⁹⁷, also mediate the immunomodulatory effects of PD-1, resulting in regulation of immune cytokine and chemokine expression. Classical TCM formulas^{98–100}, e.g., Gegen Qinlian decoction (GQD), combined with anti-mouse PD-1 antibodies potently inhibited the growth of CT26 tumors and significantly enriched gut microbiota. Two metabolic signaling pathways, *i.e.*, glycerophospholipid metabolism and sphingolipid metabolism, were also altered by this treatment. Furthermore, association strategies significantly increased the proportion of CD8 $^{+}$ T cells in peripheral blood and tumor tissues, enhanced IFN- γ expression, downregulated PD-1, and increased IL-2 levels. These data suggested that application of the formula GQD based on PD-1 blockade may be a novel therapeutic strategy for patients with cancer⁹⁸.

Table 3 Immune mechanism of TCMs as PD-1/PDL-1 inhibitors.

TCM	Immune mechanism
Saposhnikovia root extract Prim-O-glucosylcimifugin (POG) ⁹³	Inhibited the proliferation, metabolism and immunosuppressive ability of PMN-MDSCs, improved the tumor immunosuppressive microenvironment Created a synergistic effect with PD-1 inhibitors
Total glucosides of paeony (TGP) ⁹⁴	Increased the expression of PD-1/PD-L1 in the peripheral blood mononuclear cells Downregulated Treg cells/T helper 17 cells
Ganoderma lucidum ⁹⁵	Mediated the immunomodulation effect of PD-1 increased the expression of CCL5 chemokine in the cultured B-lymphocytes
Astragalus polysaccharides (APS) extracted from astragalus membranaceus ⁹⁶	Inhibited MOG ₃₅₋₅₅ -specific T cell proliferation and reduced the immune cytokines expression of IFN-γ, TNF-α, IL-2, IL-17, IL-4 and IL-10 Up-regulated the expression of PD-1 and PD-L1
Icaritin isolated from <i>Epimedium</i> ⁹⁷	Effectively decrease tumor burden in a T-cell dependent manner and increased CD8 T-cell infiltration and increased effector memory T-cell frequency Reduced frequency of CD11b ⁺ Gr1 ⁺ MDSCs infiltration and downregulation of PD-L1 expression and the expression of PD-L1 on neutrophils
Gegen Qinlian decoction ⁹⁸	The combination of GQD and anti-mouse PD-1 could potently inhibit the growth of CT26 tumors in a xenograft model Enriched gut microbiota and altered metabolic signaling pathways Increased the proportion of CD8 ⁺ T cells in peripheral blood and tumor tissues, increased the expression of IFN-γ, downregulated PD-1 and increased IL-2 levels
Ginseng and Astragalus granules ⁹⁹	Increased the level of insulin and reduced the level of blood glucose Increased both CD4 ⁺ FoxP3 ⁺ and CD8 ⁺ CD122 ⁺ PD-1 ⁺ Treg numbers in both spleens and lymph nodes of NOD mice Reversed a decline in CD4 ⁺ FoxP3 ⁺ Tregs The percentage of effector/memory CD8 ⁺ T cells (CD44 ^{high} CD62L ^{low}) was significantly reduced and attenuated cellular infiltration and lowered CD3 ⁺ T cell numbers around and in islets
SORICM02 (Curcumae Rhizoma, Radix Paeoniae Rubra, Rhizoma Smilacis Glabrae, Mume Fructus, and Sarcandrae Herba) ¹⁰⁰	Significantly inhibited murine skin allograft rejection and reduced graft-infiltration of CD3 ⁺ T cells increased CD8 ⁺ CD122 ⁺ PD-1 ⁺ Treg frequency with CD4 ⁺ FoxP3 ⁺ Tregs remaining unchanged Hindered CD11c ⁺ DC maturation post transplantation Induced CD8 ⁺ CD122 ⁺ PD-1 ⁺ Tregs Suppressed T cell proliferation <i>in vitro</i> and inhibited both mTOR and NF-κB signaling pathways increased IL-10 production, reduced IFN-γ level
PG2 isolated from <i>Astragalus membranaceus</i> ¹⁰¹	Inhibited the expression of PD-L1 on the cell surface by the protein kinase B (AKT)/mammalian target of rapamycin (mTOR)/ribosomal protein S6 kinase beta-1 (p70S6K) pathway
Indirubin, an active ingredient of <i>Indigofera tinctoria</i> L. ¹⁰²	Repaired the expression of PD1 and phosphatase and tensin homolog (PTEN) on the CD4 ⁺ T cells of ITP patients
PSORI-CM02 consisting of five herbs (Curcumae Rhizoma, Radix Paeoniae Rubra, Rhizoma Smilacis Glabrae, Mume Fructus, and Sarcandrae Herba) ¹⁰³	Increased CD8 ⁺ CD122 ⁺ PD-1 ⁺ Treg frequency <i>in vivo</i> Inhibited mTOR and NF-κB signaling pathways <i>in vitro</i>
Damingyin, an herb formulation ¹⁰⁴	Improved visual acuity of patients with proliferative diabetic retinopathy by inhibiting the expression of PD1/PD-L1, then repaired the function of monocytes
Yin Zi Huang, an herbal medicine ¹⁰⁵	Inhibited the expression of CD25, CD69, PD-1, and ICOS by stimulated CD4 ⁺ T cells in mouse

Table 4 Regulation the cancer immunity of some compounds/herbs from TCMs.

Compound/herb	Immune cell	Cancer type	Regulation mechanism
Astragaloside IV extracted from <i>Astragalus</i> ¹²⁶ <i>Rhodiola</i> ¹²⁷	M2-type macrophage	A549 and H1299 cells, Lewis lung cancer	Reduced the growth, invasion, migration, and angiogenesis of lung cancer by blocking the macrophages polarization partially through the AMPK signaling pathway
	Granulocyte-macrophage, lymphocytes	Breast cancer	Enhanced the levels of IL-2, IL-4 and granulocyte-macrophage colony-stimulating factor; increased the proliferation of lymphocytes and white blood cell (WBC) count
<i>Crassocephalum crepidioides</i> ¹²⁸	RAW264.7 macrophages	S-180 cells	Decreased tumor growth, induced NF-κB signaling pathway for releasing nitric oxide (NO)
Soyasapogenols extracted from soybean ¹²⁹	M2-type and M1-type macrophages	U373, SaOS2 and LM8 tumor cells	Inhibited the M2 polarization and increased the secretion of IL-12, suppressed the activation of STAT3 and the proliferation of these tumor cells
Soyasapogenols B ¹²⁹	Macrophage	LM8 tumor cells	Delayed subcutaneous tumor development and lung metastasis, induced anti-tumor immune response
A standardized herbal extract ¹³⁰	Macrophage	Low-risk prostate cancer	Stimulated phagocytosis and the expression of inflammatory mediators (C4b, CXCL3, lymphotoxin, NOS2, TLRI, TNF14), suppressed the tumor size, increased splenocyte cytotoxicity and numbers of CD8 T cells, macrophages, and dendritic cells in the spleens
<i>Astragalus</i> polysaccharides and <i>Codonopsis</i> polysaccharides ¹³¹	Dendritic cells (DCs), CD4 ⁺ and CD8 ⁺ T cell	4T1 breast cancer	Enhanced mouse CD4 ⁺ and CD8 ⁺ T-cell proliferation and anti-4T1 metastasis activity, increased expression of CD40, CD80 and CD86, activated immune cells by mediating the expression of cytokines/chemokines
Lupanol ¹³²	NK cells	BGC823, N87 and HGC27 of human gastric cancer cells	Promoted the proliferation of NK cells, inhibit the proliferation of BGC823, N87 and HGC27, and increased the killing effect of NK cells on gastric cancer cells, via the activated of PI3K/AKT and WNT/B-catenin signaling pathways for increasing PFP, IFN-γ, and CD107a expression
Icaruin or 3,5,7-trihydroxy-4'-methoxy-8- (3-hydroxy-3-methylbutyl)-flavone ¹³³	MDSCs	4T1-Neu tumors	Reduced MRP8/MRP14 and toll-like receptor 4 (TLR4) expression, inhibited tumor growth in 4T1-Neu tumor-bearing mice and decreased MDSC numbers in the spleen, restarted IFN-γ production, decreased the production of NO and reactive oxygen species (ROS) in MDSC, decreased the expression of S100A8/9 and inhibition of activation of STAT3 and AKT
Glycoprotein ZPDC extracted from Chinese prickly ash ¹³⁴	NK cells	Liver cancer cells	Induced secretion of perforin and granzyme B, NK cells activity and apoptosis-related factors (BID, cytochrome c, and caspase-3) in liver tissues
Shugan Jianpi formula ¹³⁵	CD8 ⁺ T cells and MDSCs	Breast cancer	Reduced CD8 ⁺ T lymphocyte apoptosis and tumor cell activity, increased immune surveillance capability, and inhibited MDSC proliferation
Shikonin ¹³⁶	DCs	B16 tumor cells	Increased the population of skin DCs migrating into the draining lymph nodes and cytotoxic T lymphocyte activities in B16 tumor model
Ginsenoside or ginsenoside Rg3 ^{137,138}	DCs and naive T cells	H22 tumor	Enhanced CD1a, CD80, CD83, CD86 and HLA-DR expression and T cell stimulatory capacity, decreased endocytic activity, Naïve T cells turned into typical Th1 cells Inhibited the growth of H22 tumors, enhanced the cellular immunity of H22-bearing mice
Tetramethylpyrazine phosphate ¹³⁹	Macrophages and Th2 cells	Lung cancer	Enhance the expression levels of IFN-γ, IL-2 and T-bet, but reduced type 2 cytokines (IL-4, IL-6, IL-10) and GATA3
Polysaccharide extracted from <i>Glycyrrhizia Radix</i> ¹⁴⁰	Treg cells	H22 tumor	Decreased the population of Treg cells and lymph node FOXP3 and IL-10 expression, decreased IL-10 and TGF-β level and increased IL-2 and IL-12p70 level
<i>Prunella vulgare</i> ¹⁴¹	B cells	TPC-1 and FTC-133 cell lines	Induced apoptosis in TPC-1 and FTC-133 cell lines, increased BCL-2-associated X protein and caspase-3 expression, and downregulated B-cell lymphoma-2 expression in TPC-1 and FTC-133
<i>Ganoderma lucidum</i> polysaccharides ¹⁴²	Teff/Treg/Th2 cells	Hepatocellular carcinoma	Suppressed tumor growth in hepatoma-bearing mice associated with an increase of the ratio of Teffs/Tregs, increased IL-2 secretion for eliminating Treg suppression of Teff proliferation, inhibited NOTCH1 and FOXP3 expression

4.3. Stimulatory effects of TCMs on costimulatory molecules

TCMs are complex systems with multiple components and targets, resulting in multitarget effects in the body; these multitarget effects could provide new directions for the treatment of immune diseases. Compared with mAbs, TCMs are less expensive, simpler to obtain, induce fewer adverse reactions, and are abundantly available. However, most studies of TCMs for the treatment of immune diseases are still in the basic research phase, and few human trials have been conducted. Accordingly, additional, higher-quality clinical trials are needed to evaluate the immunoregulatory mechanisms of TCMs in the human body. Identifying the active components and drug targets in TCMs could provide important clues for the control and treatment of immune diseases.

Activation of immune cells is an important part of the immune response. Previous studies have shown that the activation of T cells requires multiple signals; that is, the TCR recognizes the MHC-antigen peptide complex on the surface of the APC and then generates the first signal. Other molecular interactions between the two provide a second signal. If only the first signal is detected, T-cell response incompetence or even programmed death could occur. Thus, costimulatory molecules are essential for inducing effective immune responses^{106,107}.

Costimulatory molecules can be divided into positive and negative types. The former produces costimulatory signals that initiate or sustain the immune response, whereas the latter is mainly involved in termination of the immune response in a timely and effective manner to restrict the immune response. Negative type costimulatory molecules include CTLA-4-b7-1/b7-2 and FasL. In the later stages of the initial immune response, CTLA-4 is induced and binds with the corresponding ligand B7 to downregulate the immune response. In addition, these costimulatory signals regulate body functions in a bidirectional and holistic manner. These findings are consistent with the immunomodulatory mechanisms of TCMs and costimulatory molecules. Indeed, elucidation of the regulatory effects of TCMs on costimulatory molecules is required to understand the immunomodulatory roles of TCMs.

Many TCMs or active components have regulatory or even bidirectional regulatory effects on adhesion molecules, such as Ig^{108–114} and intercellular adhesion molecule 1 (ICAM-1)^{115–117}. Peony-glucosides, ginsenosides, *Astragalus membranaceus*, and Chailing decoction¹¹⁸ all have bidirectional regulatory effects on TNF. Some costimulatory molecules are members of the Ig or TNF/TNFR superfamily, whereas some are adhesion molecules, such as ICAM-1; thus, TCMs can regulate the expression of costimulatory molecules, particularly induced costimulatory molecules. Indeed, TCMs have regulatory effects on many cytokines, such as IL-1, IL-4, IL-5, IL-10, IL-12, IFN- γ , and TGF- β , and can also indirectly regulate the expression of costimulatory molecules by mediating cytokine expression. For example, taxifolin, a natural catechol-type flavonol element (also known as dihydroquercetin) has strong antioxidant and antiglycation activities and has been reported to clear cerebrovascular amyloid- β deposits. *In vivo*, taxifolin suppresses inflammation (*via* IL-1 β , IL-6, IL-10, TNF- α/β , TGF- β , and vascular endothelial growth factor) and alleviates the accumulation of triggering receptor expressed on myeloid cell 2-expressing cells in the brain. Moreover, intracerebral production of amyloid- β can be inhibited by suppressing the ApoE/ERK1/2/amyloid- β precursor protein axis¹¹⁹. Vascular amyloid- β and 40-residue amyloid- β protein (amyloid- $\beta_{1–40}$) deposits were decreased in taxifolin-treated Tg-

SwDI mice¹²⁰. *In vitro* studies have demonstrated that taxifolin has inhibitory effects on the aggregation of the 42-residue amyloid- β protein (amyloid- $\beta_{1–42}$), which is involved in the pathogenesis of Alzheimer's disease¹²¹. Therefore, the biological component taxifolin may be a potential target for clinical applications to prevent or treat cerebral amyloid angiopathy.

Increasing researches have shown that cancers and tumor cells are associated with abnormal expression of costimulatory molecules. Low expression or even absence of co-stimulatory molecules is an important reason for immune escape of malignant tumors, such as breast cancer¹²², ovarian cancer¹²³ and hematological malignancies¹²⁴. TCMs has also been reported to inhibit angiogenesis and tumor growth of Lewis lung or prostate cancer¹²⁵. For example, Xu et al.¹²⁶ found that astragaloside IV extracted from *Astragalus* significantly restrained the reductive expression of CD206 and M2-related genes in the M2-type macrophage that polarized by IL-13 and IL-4, exhibited the inhibition of the invasion, migration and angiogenesis of A549 and H1299 cancer cells induced by M2-type macrophages, and suppressed AMPK α in M2-type macrophages activation. In addition, the *in vivo* experiments also showed that astragaloside IV could inhibit the tumor growth rate and cut down the metastasis of Lewis lung cancer. As shown in Table 4^{126–142}, there are some compounds or herbs from TCMs which can regulate the cancer immunity, especially in the tumor microenvironment. What's more, TCMs can exert the regulation of cancer immunity in clinic. Schwartzberg et al.¹⁴³ discovered that a neutral oil extracted and isolated from *Coix* seed had a statistically significant improvement in progression-free survival (PFS) in pancreatic cancer patients, and showed anti-neoplastic activity. *Rhodiola* can improve the immunity in patients receiving postoperative chemotherapy for breast cancer and reduce the occurrence of chemotherapy-related oral ulcer. *Rhodiola* therefore has the potential to be used in conjunction with chemotherapy to reduce the incidence of oral ulcers¹²⁷. TCMs has played, and still plays, an integral role in immunity of human health care all over the world. A “hot topic” has also been aroused in anti-cancer research by natural products derived from TCMs. Despite the unique immunity regulatory features of many compounds originated from TCMs, their clinical applications are disproportionately limited, and some TCMs are indeed highly toxic, particularly, their toxicity increases dramatically as herbs absorb a growing number of toxic substances in contaminated areas. Further, some herbal formulations contain too many herbs, which may cause additional toxicity. We believe that as the demonstration for immune regulation and toxicology research continue to be explored, which improve the comprehension about the mechanistic actions and clinical potential use of these compounds in the near future. TCMs will also serve as a huge community from which many promising compounds will be more extensively studied their applications in antitumor therapy for clinical use.

5. Conclusions

Immune checkpoints are critical regulators of T-cell responses in tissues. Accordingly, decreasing the function of immune checkpoints by antibody blockade in cancer or chronic disorders can increase the availability of ligands to permit the activation of potentially self-reactive T cells, thereby altering cellular homeostasis. Moreover, overexpression of numerous immune checkpoints on cells can stimulate or inhibit T-cell responses. The use of

immune checkpoint inhibitors may help to reverse immunosuppression, which is commonly observed in chronic infections and can promote immune responses. Importantly, combining immune checkpoint blockade with therapeutic vaccination can enhance vaccine-induced immune responses by depleting Tregs or blocking the production or function of immunosuppressive cytokines. Although blocking Tregs or immune checkpoints is unlikely to enhance the efficacy of routine prophylactic vaccination, it may have the potential to increase the efficacy of therapeutic vaccines against many chronic infections, such as malaria, tuberculosis, and HIV. Thus, further studies of immune checkpoints could facilitate our understanding of the mechanisms through which TCMs and novel drugs regulate immune function.

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Author contributions

He Huang and Changxiao Liu: proposition proposal, designed and final revision; Yuli Wang: organizational framework and construction, paper drafting; Xingyan Zhang, Wenjing Zhao and Huling Li: revision, analysis, final revision; Yuyan Wang, Lixing Zhang, Li Xinping: collected data and provided materials; Tiejun Zhang and Hongbing Zhang: revision.

Conflicts of interest

The authors report no conflicts of interest.

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