

Crystal Ball

An underappreciated DIET for anaerobic petroleum hydrocarbon-degrading microbial communities

Federico Aulenta,^{1*}  Matteo Tucci,¹ Carolina Cruz Viggli,¹ Jan Dolfig,² Ian M. Head³ and Amelia-Elena Rotaru⁴

¹Water Research Institute (IRSA), National Research Council (CNR), Monterotondo, RM, Italy.

²School of Engineering, Newcastle University, Newcastle upon Tyne, UK.

³School of Natural and Environmental Sciences, Newcastle University, Newcastle upon Tyne, UK.

⁴Department of Biology, University of Southern Denmark, Odense, Denmark.

The discovery of direct interspecies electron transfer

The discovery of direct interspecies electron transfer (DIET) between microbial cells revealed a new and exciting possibility for microbial cooperation in energy-limited anaerobic ecosystems (Summers *et al.*, 2010). The first observation of a DIET association was between two *Geobacter* species, *Geobacter metallireducens* and *Geobacter sulfurreducens*, grown on a medium provided with an electron donor (ethanol) for the first species, and an electron acceptor (fumarate) for the other species. Together the two *Geobacter* formed electrically conductive aggregates. In these aggregates, electrons from the oxidation of the electron donor by *G. metallireducens* were passed to *G. sulfurreducens*, which could use them to reduce fumarate to succinate. The electrical wiring between the two microorganisms was ensured by *c*-type cytochromes and conductive type IV pili (Summers *et al.*, 2010). Pili and *c*-type cytochromes were only required for DIET associations and unnecessary for associations between *Geobacter* and partners capable of formate or H₂ transfer (Rotaru *et al.*, 2012).

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*For correspondence. E-mail aulenta@irsa.cnr.it; Tel. +39-0690672751; Fax +39-0690672787.

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This disruptive discovery shook up a consolidated paradigm according to which the exchange of electrons (as well as of nutrients, carbon substrates and information) among microbes, proceeds via molecular diffusion of soluble (redox-active) molecules such as H₂ or formate (Stams and Plugge, 2009; Schink and Stams, 2013). Theoretical analyses indicated that DIET (governed by electrical conductivity) could be substantially faster and more effective than interspecies H₂ transfer, governed by Fick's law of molecular diffusion (Cruz Viggli *et al.*, 2014; Cheng and Call, 2016). This leads to the intriguing hypothesis that direct electron transfer between microbial species could be much more widespread in natural environments than previously recognized.

Following the proof-of-concept study reported by Summers *et al.*, (2010), DIET between a range of microorganisms was explored, most notably between *Geobacter* spp. and methanogens (Rotaru *et al.*, 2014; Yee *et al.*, 2019; Yee and Rotaru, 2020). Interestingly, several studies showed that DIET interactions could be stimulated by conductive materials (Lovley, 2017). For example, even the addition of relatively small amounts (<50 mg Fe l⁻¹) of nano-sized magnetite nanoparticles could trigger DIET between *Geobacter* species, with a negligible lag phase (You *et al.*, 2019). This phenomenon was also observed in syntrophic methanogenic communities (Dubé and Guiot, 2015) and in co-cultures of *Geobacter sulfurreducens* and *Thiobacillus denitrificans* (Kato *et al.*, 2012). It was shown that conductive magnetite particles attached to microbes and served as abiotic electron conduits replacing *c*-type cytochromes, thus connecting redox reactions catalysed by different species of microorganisms (Liu *et al.*, 2015). More recently, a wide range of conductive or semi-conductive minerals and materials such as pyrite, various types of chars and graphite were found to serve as substitutes for biological connectors to promote and facilitate electron transfer between species (Cheng and Call, 2016).

Practical implications of DIET in engineered bioprocesses

In the past few years, DIET has been the focus of intense research activities and remarkable efforts have

been made to translate such fundamental scientific discoveries into practical strategies to improve engineered bioprocesses. In particular, much attention and research efforts have been dedicated to exploring the use of conductive materials to accelerate and stabilize the anaerobic digestion (AD) of waste organic substrates (Dubé and Guiot, 2015). Indeed, a large number of publications indicated that the addition of electrically conductive particles to anaerobic digestors has the potential to accelerate some of the rate-limiting steps in the methanogenic conversion of organic substrates, such as the syntrophic oxidation of volatile fatty acids or alcohols (Cruz Viggli *et al.*, 2014). The acceleration of syntrophic metabolism by conductive particles is most likely because conductive materials promote higher rates of electron transfer between substrate-oxidizing *Bacteria* and methanogenic *Archaea* (Morita *et al.*, 2011; Liu *et al.*, 2012; Martins *et al.*, 2018; Park *et al.*, 2018). In general, in anaerobic digestors amended with electrically conductive particles, the rate of production and yield of methane were typically higher than in control digestors. Amended digestors were also found to be less susceptible to environmental stresses and/or abrupt changes in operating conditions, such as the applied organic loading rate (Baek *et al.*, 2015; Dubé and Guiot, 2015).

The benefits of improving conventional AD processes are clear and are, in part, justified by the rapidly expanding AD market which is being promoted by an increased focus on decarbonizing the energy sector (Edwards *et al.*, 2015). Nevertheless, there are many other opportunities and applications for DIET on the horizon. One broad area, which remains largely unexplored, is bioremediation of soil, sediment and groundwater. Here, we point out the role of DIET on the anaerobic biodegradation of petroleum hydrocarbons, driven by naturally occurring or specifically added electro-conductive minerals or materials. We anticipate that harnessing DIET could contribute to the development of new, more-effective bioremediation technologies.

Syntrophic anaerobic biodegradation of hydrocarbon contaminants

Many subsurface environments, such as soils, sediments and aquifers, become rapidly anaerobic after accidental contamination with petroleum hydrocarbons. This is because hydrocarbons serve as electron donors in the metabolism of fast-growing aerobic microbial communities, thereby leading to a rapid depletion of oxygen, which is only slowly replenished from surrounding oxic environments. Albeit slower than its aerobic counterpart, the biodegradation of a broad variety of aliphatic and aromatic hydrocarbons under anaerobic conditions has been widely documented (Widdel and Rabus, 2001;

Gieg *et al.*, 2014; Rabus *et al.*, 2016). In this context, we expect anaerobic hydrocarbon biodegradation to become an increasingly relevant process in the managed attenuation of contaminated subsurface environments.

Under methanogenic conditions, the biodegradation of hydrocarbons requires the syntrophic cooperation of *Bacteria* and *Archaea* (Gieg *et al.*, 2014). Following an activation step, typically proceeding via fumarate addition, carboxylation or hydroxylation (Heider, 2007; Meckenstock and Mouttaki, 2011), *Bacteria* catalyse the oxidation of hydrocarbons to CO₂/H₂, acetate or a mixture thereof (Fig. 1A). Such reactions are, however, largely endergonic under standard biochemical conditions (e.g. +471 kJ mol⁻¹ for hexadecane oxidation to acetate and H₂), which in turn forces *Bacteria* to rely on the activity of *Archaea* to rapidly convert the produced H₂ and acetate into methane via exergonic reactions (e.g. -36 kJ mol⁻¹ for acetoclastic and -131 kJ mol⁻¹ for hydrogenotrophic methanogenesis), thereby making the overall reaction energetically favourable (Dolfing *et al.*, 2008). Interestingly, an ever-increasing number of studies have suggested that syntrophic processes also play a relevant role during biodegradation of hydrocarbons under nitrate-, iron- or sulphate-reducing conditions (Gieg *et al.*, 2014; Fig. 1A). Even the anaerobic oxidation of methane, a primary mechanism for methane removal in ocean sediments, has been found to be driven by the syntrophic partnership between anaerobic methanotrophic (ANME) *Archaea* and deltaproteobacterial sulphate-reducing bacteria (SRB; Boetius *et al.*, 2000; Skennerton *et al.*, 2017). The occurrence of all the above described syntrophic microbial relationships has been, however, typically inferred from results of phylogenetic analyses, in some cases corroborated by thermodynamic calculations. Similarly, the electron transfer mechanisms actually involved in the biodegradation processes have been rarely experimentally verified, primarily due to intrinsic methodological difficulties in tracking the involved electron carriers (i.e. H₂, formate, acetate) transiently accumulating only at extremely low concentration levels and the lack of alternative investigation techniques.

Hints of DIET-driven hydrocarbon degradation and urgent research needs

Today, DIET is considered a well-established metabolic route in the context of anaerobic digestion (Baek *et al.*, 2015; Dubé and Guiot, 2015); however, the relevance of this relatively new mechanism in the field of biogeochemistry and/or bioremediation is only marginally explored, in spite of the fact that a few highly relevant publications have clearly pinpointed its likely significance. As an example, recent studies have provided

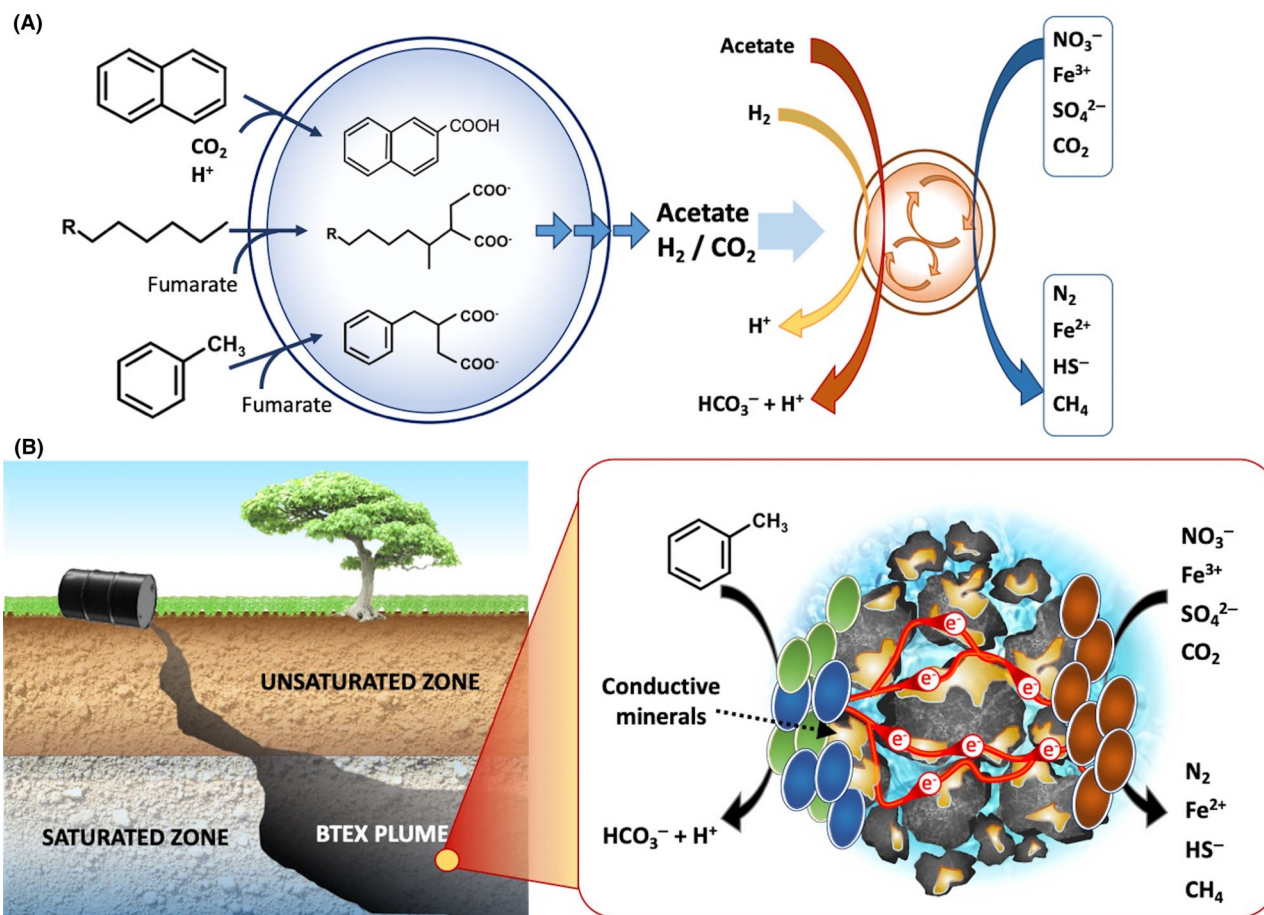


Fig. 1. (A) Syntrophic anaerobic degradation of model hydrocarbons under different terminal electron-accepting conditions. (B) *In situ* syntrophic degradation of toluene driven by direct interspecies electron transfer mediated by conductive minerals.

multiple lines of evidence that the syntrophic oxidation of methane (as well as of low molecular weight alkanes, such as butane) by anaerobic methanotrophic *Archaea* (ANME) coupled to the reduction of sulphate by deltaproteobacteria is actually powered by a direct transfer of electrons among the syntrophic partners, occurring via electrical connections based on multihem cytochromes and/or type IV pili (McGlynn *et al.*, 2015; Wegener *et al.*, 2015; Chen *et al.*, 2019). Activated carbon (AC), an electron conductive material, was found to strongly stimulate (+96%) the biodegradation of naphthalene, a common polycyclic aromatic hydrocarbon (PAH), under anaerobic conditions (Bonaglia *et al.*, 2020). By contrast, no stimulatory effects were observed under aerobic conditions, thereby suggesting that AC particles promoted direct interspecies electron transfer (DIET) between microorganisms involved in syntrophic PAH degradation pathways. Rotaru and colleagues (Rotaru *et al.*, 2018, 2019) demonstrated that enrichment of microorganisms capable of syntrophic (methanogenic) degradation processes is strictly dependent on the

presence of naturally occurring, semi-conductive iron minerals (e.g. Fe-oxides and Fe-sulphides) in sediments from lake La Cruz (Spain) and in coastal sediments of the Baltic Sea. Nielsen and colleagues (Nielsen *et al.*, 2010) reported the occurrence of electric currents in marine sediments of Aarhus Bay (DK) which coupled spatially separated redox processes. The authors suggested that semi-conductive pyrite (i.e. FeS₂) minerals, highly abundant (up to 30 μmol g⁻¹) in such sediments, could be, at least partially, responsible for conducting such electric currents, with these latter being generated by the microbial oxidation of reduced sedimentary organic and inorganic compounds. Moreover, electrically conducting “cable bacteria” similar to those described by Nielsen *et al.* (2010) may have a role in recycling electron acceptors such as sulphate in hydrocarbon contaminated anoxic environments (Müller *et al.*, 2016) and thus sustaining anaerobic hydrocarbon degradation.

Electricity spontaneously and widely generated in deep-sea hydrothermal fields from the microbially driven oxidation of hydrothermal fluids (containing metal ions

such as Fe^{2+} , Cu^{2+} , and reduced gases such as H_2S , H_2 and CH_4) and propagated via sulphide-rich mineral deposits (Yamamoto *et al.*, 2017).

In principle, the enhancement of DIET could also represent a novel bioremediation approach, which might involve the distribution of a colloidal suspension of electrically conductive nanoparticles into the contaminated matrix, so as to create *in situ* a large, electrically conductive and biologically reactive treatment zone (Fig. 1B). An alternative approach may involve devising strategies to stimulate the (biological) formation of conductive particles (e.g. magnetite is an end-product of the metabolism of iron-reducing bacteria; iron sulphides can be formed from the reaction of biologically produced sulphide with iron ores) directly *in situ*. The addition of ethanol, or substrates releasing ethanol upon fermentation, is another interesting strategy which could also be devised to stimulate hydrocarbons biodegradation in subsurface environments, possibly in combination with those previously described. Indeed, recent studies demonstrated that ethanol addition as a co-substrate during anaerobic degradation of organic substrates triggers DIET in mixed microbial cultures, with this syntrophic/cooperative pathway stably persisting even after ethanol supplementation ceased (Zhao *et al.*, 2016, 2017; Zhao and Zhang, 2019). Moreover, the stimulation of methanogenic degradation of organic waste and coal by addition of saturating amounts of the phenazine dye, neutral red, such that filaments of electrically conducting neutral red crystals form, presents an intriguing opportunity for managing direct interspecies electron transfer in natural and engineered environments (Beckmann *et al.*, 2016). A similar approach, but involving the use of carbon nanotubes, has also proved effective in triggering DIET-based syntrophic degradation processes (Salvador *et al.*, 2017). Clearly, a critical step towards a deeper comprehension and effective management of DIET-based biotechnological approaches will necessarily be the development and *ad hoc* application of a broad array of analytical, electrochemical, microscopy and biomolecular techniques specifically designed to identify, monitor and possibly manipulate such tiny electric currents and related chemical reactions, as well as to identify the abundance and expression of genes specifically involved in extracellular electron transfer processes.

In conclusion, it is apparent that a tangible potential exists that hydrocarbon-degrading microbial communities inhabiting (anthropogenically or naturally) contaminated subsurface environments such as soil, sediments and aquifers can team-up with a variety of conductive and semi-conductive particles, abundantly and ubiquitously present in such ecosystems (or *ad hoc* supplemented), to couple, even spatially distant, biogeochemical

reactions and accordingly carry out specific syntrophic and/or cooperative metabolisms of major environmental relevance.

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References

- Baek, G., Kim, J., Cho, K., Bae, H., and Lee, C. (2015) The biostimulation of anaerobic digestion with (semi)conductive ferric oxides: their potential for enhanced biomethanation. *Appl Microbiol Biotechnol* **99**: 10355–10366.
- Beckmann, S., Welte, C., Li, X., Oo, Y.M., Kroeninger, L., Heo, Y., *et al.* (2016) Novel phenazine crystals enable direct electron transfer to methanogens in anaerobic digestion by redox potential modulation. *Energy Environ Sci* **9**: 644–655.
- Boetius, A., Ravensschlag, K., Schubert, C.J., Rickert, D., Widdel, F., Gieseke, A., *et al.* (2000) A marine microbial consortium apparently mediating anaerobic oxidation of methane. *Nature* **407**: 623–626.
- Bonaglia, S., Broman, E., Brindefalk, B., Hedlund, E., Hjorth, T., Rolff, C., *et al.* (2020) Activated carbon stimulates microbial diversity and PAH biodegradation under anaerobic conditions in oil-polluted sediments. *Chemosphere* **248**: 126023.
- Chen, S.-C., Musat, N., Lechtenfeld, O.J., Paschke, H., Schmidt, M., Said, N., *et al.* (2019) Anaerobic oxidation of ethane by archaea from a marine hydrocarbon seep. *Nature* **568**: 108–111.
- Cheng, Q., and Call, D.F. (2016) Hardwiring microbes via direct interspecies electron transfer: mechanisms and applications. *Environ Sci Process Impacts* **18**: 968–980.
- Cruz Viggli, C., Rossetti, S., Fazi, S., Paiano, P., Majone, M., Aulenta, F., *et al.* (2014) Magnetite particles triggering a faster and more robust syntrophic pathway of methanogenic propionate degradation. *Environ Sci Technol* **48**: 7536–7543.
- Dolfing, J., Larter, S.R., and Head, I.M. (2008) Thermodynamic constraints on methanogenic crude oil biodegradation. *ISME J* **2**: 442–452.
- Dubé, C.-D., and Guiot, S.R. (2015) Direct interspecies electron transfer in anaerobic digestion: a review. In Guebitz, G., Bauer, A., Bochmann, G., Gronauer, A. & Weiss, S. (Eds.), *Biogas Science and Technology. Advances in Biochemical Engineering/Biotechnology*. Cham, Switzerland: Springer, **151**: 101–115. https://doi.org/10.1007/978-3-319-21993-6_4
- Edwards, J., Othman, M., and Burn, S. (2015) A review of policy drivers and barriers for the use of anaerobic digestion in Europe, the United States and Australia. *Renew Sustain Energy Rev* **52**: 815–828.
- Gieg, L.M., Fowler, S.J., and Berdugo-Clavijo, C. (2014) Syntrophic biodegradation of hydrocarbon contaminants. *Curr Opin Biotechnol* **27**: 21–29.

- Heider, J. (2007) Adding handles to unhandy substrates: anaerobic hydrocarbon activation mechanisms. *Curr Opin Chem Biol* **11**: 188–194.
- Kato, S., Hashimoto, K., and Watanabe, K. (2012) Microbial interspecies electron transfer via electric currents through conductive minerals. *Proc Natl Acad Sci USA* **109**: 10042–10046.
- Liu, F., Rotaru, A.-E., Shrestha, P.M., Malvankar, N.S., Nevin, K.P., and Lovley, D.R. (2012) Promoting direct interspecies electron transfer with activated carbon. *Energy Environ Sci* **5**: 8982.
- Liu, F., Rotaru, A.E., Shrestha, P.M., Malvankar, N.S., Nevin, K.P., and Lovley, D.R. (2015) Magnetite compensates for the lack of a pilin-associated c-type cytochrome in extracellular electron exchange. *Environ Microbiol* **17**: 648–655.
- Lovley, D.R. (2017) Syntrophy goes electric: direct interspecies electron transfer. *Annu Rev Microbiol* **71**: 643–664.
- Martins, G., Salvador, A.F., Pereira, L., and Alves, M.M. (2018) Methane production and conductive materials: a critical review. *Environ Sci Technol* **52**: 10241–10253.
- McGlynn, S.E., Chadwick, G.L., Kempes, C.P., and Orphan, V.J. (2015) Single cell activity reveals direct electron transfer in methanotrophic consortia. *Nature* **526**: 531–535.
- Meckenstock, R.U., and Mouttaki, H. (2011) Anaerobic degradation of non-substituted aromatic hydrocarbons. *Curr Opin Biotechnol* **22**: 406–414.
- Morita, M., Malvankar, N., Franks, A., Summers, Z., Gileteaux, L., Rotaru, A., et al. (2011) Potential for direct interspecies electron transfer in methanogenic wastewater digester aggregates. *MBio* **2**(4), e00159-11.
- Müller, H., Bosch, J., Griebler, C., Damgaard, L.R., Nielsen, L.P., Lueders, T., and Meckenstock, R.U. (2016) Long-distance electron transfer by cable bacteria in aquifer sediments. *ISME J* **10**: 2010–2019.
- Nielsen, L.P., Risgaard-Petersen, N., Fossing, H., Christensen, P.B., and Sayama, M. (2010) Electric currents couple spatially separated biogeochemical processes in marine sediment. *Nature* **463**: 1071–1074.
- Park, J.-H., Kang, H.-J., Park, K.-H., and Park, H.-D. (2018) Direct interspecies electron transfer via conductive materials: A perspective for anaerobic digestion applications. *Bioresour Technol* **254**: 300–311.
- Rabus, R., Boll, M., Heider, J., Meckenstock, R.U., Buckel, W., Einsle, O., et al. (2016) Anaerobic microbial degradation of hydrocarbons: from enzymatic reactions to the environment. *J Mol Microbiol Biotechnol* **26**: 5–28.
- Rotaru, A.E., Shrestha, P.M., Liu, F., Ueki, T., Nevin, K., Summers, Z.M., and Lovley, D.R. (2012) Interspecies electron transfer via hydrogen and formate rather than direct electrical connections in cocultures of *Pelobacter carbinolicus* and *Geobacter sulfurreducens*. *Appl Environ Microbiol* **78**: 7645–7651.
- Rotaru, A.-E., Shrestha, P.M., Liu, F., Shrestha, M., Shrestha, D., Embree, M., et al. (2014) A new model for electron flow during anaerobic digestion: direct interspecies electron transfer to *Methanosaeta* for the reduction of carbon dioxide to methane. *Energy Environ Sci* **7**: 408–415.
- Rotaru, A.-E., Calabrese, F., Stryhanyuk, H., Musat, F., Shrestha, P.M., Weber, H.S., et al. (2018) Conductive particles enable syntrophic acetate oxidation between *Geobacter* and *Methanosarcina* from coastal sediments. *MBio* **9**(3), e00226-18.
- Rotaru, A.E., Posth, N.R., Löscher, C.R., Miracle, M.R., Vicente, E., Cox, R.P., et al. (2019) Interspecies interactions mediated by conductive minerals in the sediments of the iron rich meromictic Lake La Cruz, Spain. *Limnetica* **38**: 31–40.
- Salvador, A.F., Martins, G., Melle-Franco, M., Serpa, R., Stams, A.J.M., Cavaleiro, A.J., et al. (2017) Carbon nanotubes accelerate methane production in pure cultures of methanogens and in a syntrophic coculture. *Environ Microbiol* **19**: 2727–2739.
- Schink, B., and Stams, A.J.M. (2013) Syntrophism among prokaryotes. Dworkin, M., Falkow, S., Rosenberg, E., Schleifer, K.H. & Stackebrandt, E., In *The Prokaryotes*. New York (NY): Springer, pp. 471–493.
- Skenneron, C.T., Chourey, K., Iyer, R., Hettich, R.L., Tyson, G.W., and Orphan, V.J. (2017) Methane-fueled syntrophy through extracellular electron transfer: uncovering the genomic traits conserved within diverse bacterial partners of anaerobic methanotrophic archaea. *MBio* **8**: 1–14.
- Stams, A.J.M., and Plugge, C.M. (2009) Electron transfer in syntrophic communities of anaerobic bacteria and archaea. *Nat Rev Microbiol* **7**: 568–577.
- Summers, Z.M., Fogarty, H.E., Leang, C., Franks, A.E., Malvankar, N.S., and Lovley, D.R. (2010) Direct exchange of electrons within aggregates of an evolved syntrophic coculture of anaerobic bacteria. *Science* **330**: 1413–1415.
- Wegener, G., Krukenberg, V., Riedel, D., Tegetmeyer, H.E., and Boetius, A. (2015) Intercellular wiring enables electron transfer between methanotrophic archaea and bacteria. *Nature* **526**: 587–590.
- Widdel, F., and Rabus, R. (2001) Anaerobic biodegradation of saturated and aromatic hydrocarbons. *Curr Opin Biotechnol* **12**: 259–276.
- Yamamoto, M., Nakamura, R., Kasaya, T., Kumagai, H., Suzuki, K., and Takai, K. (2017) Spontaneous and widespread electricity generation in natural deep-sea hydrothermal fields. *Angew Chemie Int Ed* **56**: 5725–5728.
- Yee, M.O., and Rotaru, A.-E. (2020) Extracellular electron uptake in *Methanosarcinales* is independent of multiheme c-type cytochromes. *Sci Rep* **10**: 372.
- Yee, M.O., Snoeyenbos-West, O.L., Thamdrup, B., Ottosen, L.D.M., and Rotaru, A.-E. (2019) Extracellular electron uptake by two *Methanosarcina* species. *Front Energy Res* **7**: 29.
- You, Y., Zheng, S., Zang, H., Liu, Feng, Liu, Fanghua, and Liu, J. (2019) Stimulatory effect of magnetite on the syntrophic metabolism of *Geobacter* co-cultures: Influences of surface coating. *Geochim Cosmochim Acta* **256**: 82–96.
- Zhao, Z., and Zhang, Y. (2019) Application of ethanol-type fermentation in establishment of direct interspecies electron transfer: a practical engineering case study. *Renew Energy* **136**: 846–855.

Zhao, Z., Zhang, Y., Yu, Q., Dang, Y., Li, Y., and Quan, X. (2016) Communities stimulated with ethanol to perform direct interspecies electron transfer for syntrophic metabolism of propionate and butyrate. *Water Res* **102**: 475–484.

Zhao, Z., Li, Y., Quan, X., and Zhang, Y. (2017) New application of ethanol-type fermentation: stimulating methanogenic communities with ethanol to perform direct interspecies electron transfer. *ACS Sustain Chem Eng* **5**: 9441–9453.